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# Defect engineering of polyethylene-like polyphosphoesters: solid-state NMR characterization and surface chemistry of anisotropic polymer nanoplatelets†

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Anisotropic materials with very high aspect ratios, such as nanoplatelets, are an exciting class of materials due to their unique properties based on their unilamellar geometry. Controlling their size and surface-functionality is challenging and has only been achieved in some cases for synthetic polymers. We present a general approach to prepare polymer-nanoplatelets with control over the lateral size and basal functionality, by simple polycondensation of precisely spaced and functional phosphate groups in polyethylene-like polymers. Because of the relatively large size of the phosphate groups, they are expelled from the bulk crystal to the basal surface. This allows to control the chain-folding during crystallization, which we analyzed via solid-state NMR and TEM. Furthermore, we present chemistry “on the surface” of the platelets: the pendant ester group at the phosphate offers the possibility to introduce functional groups accessible for further chemical modification on the crystal surface. This is shown by introducing a 2-acetylthioethyl ester group and subsequently cleaving this 2-acetylthioethyl ester group to the free P–OH. Together, these results render polyethylene-like polyphosphoesters a versatile platform for soft-matter nanoplatelets as functional colloids.

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## Introduction

Due to the pioneering work of Karl Ziegler and Giulio Natta, the synthesis of polyethylene and propylene at mild temperatures and pressure became possible, which has led to more than hundred million tons of polyethylene (PE) and polypropylene (PP) produced per year.<sup>1–3</sup> The application of these polymers is determined by their bulk properties like melting or glass transition temperature. Today, PE and PP are known as commodity material with various applications.<sup>4–7</sup>

In contrast to these bulk materials, planar materials with very high aspect ratios are an exciting material class due to their unique properties based on their unilamellar geometry, allowing anisotropic material properties. Examples for such materials are graphene or inorganic platelets, which are often

used to reinforce bulk polymers.<sup>8</sup> To date, only very few reports on chemically functional platelet-shaped nanomaterials based on commodity plastics like polyethylene were reported.<sup>9–11</sup>

We utilized phosphorus chemistry and simple polycondensation reactions to introduce precisely spaced phosphoester defects in a polymer chain to achieve polyethylene-like polyphosphoesters (PPEs).<sup>12</sup> PPEs have been a focus of our research in recent years ranging from biomedical to material science applications.<sup>13,14</sup> For the PE-like PPEs, the phosphates can introduce additional functionality but act as defects for the crystallization, as they are too large to be incorporated into the lamellar crystal. This led to a layered morphology of crystallites in the bulk.<sup>12,15</sup> Besides, the phosphate units also confine the thickness of the polymer lamellae, which resulted in the formation of anisotropic polymer platelets, when crystallized from dilute solution<sup>15</sup> or at the air–water interface.<sup>16</sup> In addition, the pendant ester group at the phosphate gives the possibility for the introduction of functional groups which are accessible for further chemical modification on the crystal surface after crystallization from solution. Ramakrishnan's group followed a related concept for the preparation of liquid crystals from bulk by preparing polyethylene-like polyesters,

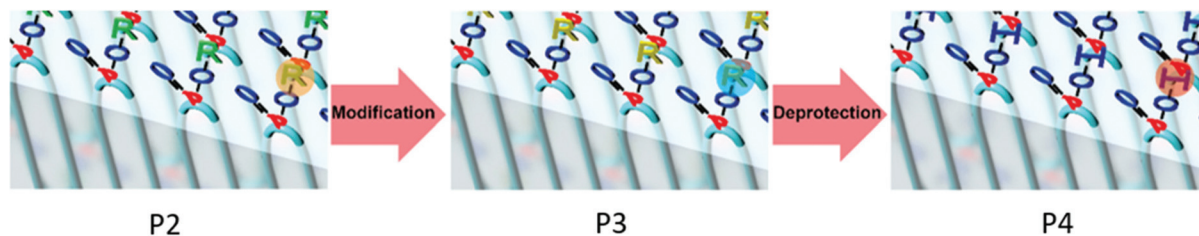
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**Scheme 2** Schematic representation of the multi-step reaction, which was conducted “on surface” of the polymer platelets: 1. step: modification of the Br-group to a thioacetate-group. 2. step: cleavage of the 2-acetylthioethyl protective group.

the  $^{31}\text{P}$  sites relative to the orthorhombic/pseudo-hexagonal lattice of the aliphatic chains can be derived (Fig. 2 top) and compared to structural information gathered by solid-state NMR.

Commonly used solid-state NMR methods like  $^{31}\text{P}$  magic angle spinning (MAS) NMR or  $^{31}\text{P}\{^1\text{H}\}$  CP-MAS NMR may probe the local chemical environment of the  $^{31}\text{P}$  sites in the solid-state, however, they will not provide any information on spatial proximities of the chemically isolated  $^{31}\text{P}$  sites in **P1**. Utilizing double-quantum (DQ) MAS-NMR methods, it is possible to probe molecular proximities in the range of 0.5 nm and thus gaining information regarding conformations and packing.<sup>35</sup> In addition to  $^{31}\text{P}$ - $^{31}\text{P}$  double-quantum single-quantum (DQ-SQ) correlation experiments, probing directly spatial proximities between chemically distinct  $^{31}\text{P}$  sites, this method can probe as well  $^{31}\text{P}$ - $^{31}\text{P}$  DQ dipolar coupling between chemically equivalent  $^{31}\text{P}$  sites based on measurements of build-up curves. These build-up curves (Fig. 1c) are sensitive to P-P inter-nuclear distances, as the nuclear dipolar

coupling constant  $D_{ij} = \frac{\mu_0 \hbar^2 \gamma_i \gamma_j}{4\pi^2 r_{ij}^3}$  between two spins  $i$  and  $j$  is proportional to the inverse cube of the inter-nuclear distance  $r_{ij}$ , and the strongest  $^{31}\text{P}$  dipolar coupling, thus the closest spatial proximity, determines predominantly the initial  $t^2$ -proportional rising of the double quantum build-up curve (with  $\mu_0$  the vacuum permittivity;  $\hbar$  the Planck constant;  $\gamma_i$  and  $\gamma_j$  are the nuclear gyro-magnetic ratios of the two spins involved in the dipolar coupling). Moreover, the long term behavior of DQ build up curves provides qualitative information on the distribution of the  $^{31}\text{P}$  dipolar couplings and spatial proximities. In crystalline arrangements with well-defined distinct distances between neighboring  $^{31}\text{P}$  sites, oscillatory behavior of the DQ build-up curves is observed (see Fig. 6b, in the publication by Saalwächter *et al.*<sup>36</sup>). For very long excitation times, these oscillations vanish in powder samples due to the orientation dependence of the DQ excitation efficiency. In less controlled assemblies with broad distributions of distances and thus dipolar couplings, the oscillatory behavior of the DQ build-up curve vanishes due to destructive interferences of the different oscillation frequencies of discrete dipolar coupling values completely. In these cases, a second moment description of the dipolar coupling assuming a dipolar interaction of a single site with continuously distributed surrounding  $^{31}\text{P}$  sites is

more appropriate and provides a better description of the DQ build-up behavior. The smooth DQ build-up curve with its quadratic initial behavior obtained for the local packing of the  $^{31}\text{P}$  sites in crystallized **P1** samples shown in Fig. 1 indicates that the second moment analysis of the local packing should be appropriate (*i.e.* indicating a crystalline, planar arrangement of P on the crystal surface). Please note, that the fluctuations occurring in the DQ build-up curve for longer DQ excitation times result from increasing relative experimental uncertainties with increasing DQ excitation times and thus do not indicate the presence of discrete dipolar coupling values resulting from a crystalline lattice (which should vanish for long DQ excitation times) like the oscillations mentioned above.

In Fig. 2, we have drawn three possibilities, how the polymer chains may propagate along with the unit cell of the crystallites, with the phosphate groups positioned between the lattice positions of the alkyl chain in an alternating pattern as illustrated in Fig. 2. For clarity, only the phosphorus atoms of the top surface are shown. In Model 1 (Fig. 2a) the polymer chain propagates diagonal to the unit cell along the (110) direction of the crystal. In Model 2 (Fig. 2b), the polymer chain propagates along the  $b$  axis of the unit cell, but the neighboring polymer chain is shifted by  $b/2$ . In Model 3 (Fig. 2c), the polymer chains propagate along the  $b$  axis, but the shift alternates between  $+$  and  $-b/2$ .

From the different models, the distance  $r_{ij}$  between two phosphate groups in close spatial proximity is known and thus can be used to calculate the  $^{31}\text{P}$  dipolar second moment ( $M_2$ ) ( $M_2 = \frac{3}{5} \gamma^4 \hbar^2 I(I+1) \sum_j r_{ij}^{-6}$ )<sup>37</sup> and compare it to the  $M_2$  obtained by the experimental DQ build-up curves of 0.197  $\text{kHz}^2$  (for details on data acquisition, processing and analysis see Solid-state NMR Experimental section). For the calculation of  $M_2$ , five unit cells in each direction were taken into account. Note that the integral contribution of more remote  $^{31}\text{P}$  sites to the value of  $M_2$  is less than 0.1%, as their contribution to the second moment decreases with the 6<sup>th</sup> power of the distance. For all 3 models, the calculation yields a  $M_2$  of  $\sim 0.14 \text{ kHz}^2$  with only minor variations, which is significantly lower than the  $M_2$  value obtained from the experimental DQ build-up curves. However, the crystals were measured in a dry state, so that the stacking of multiple crystalline layers is possible.



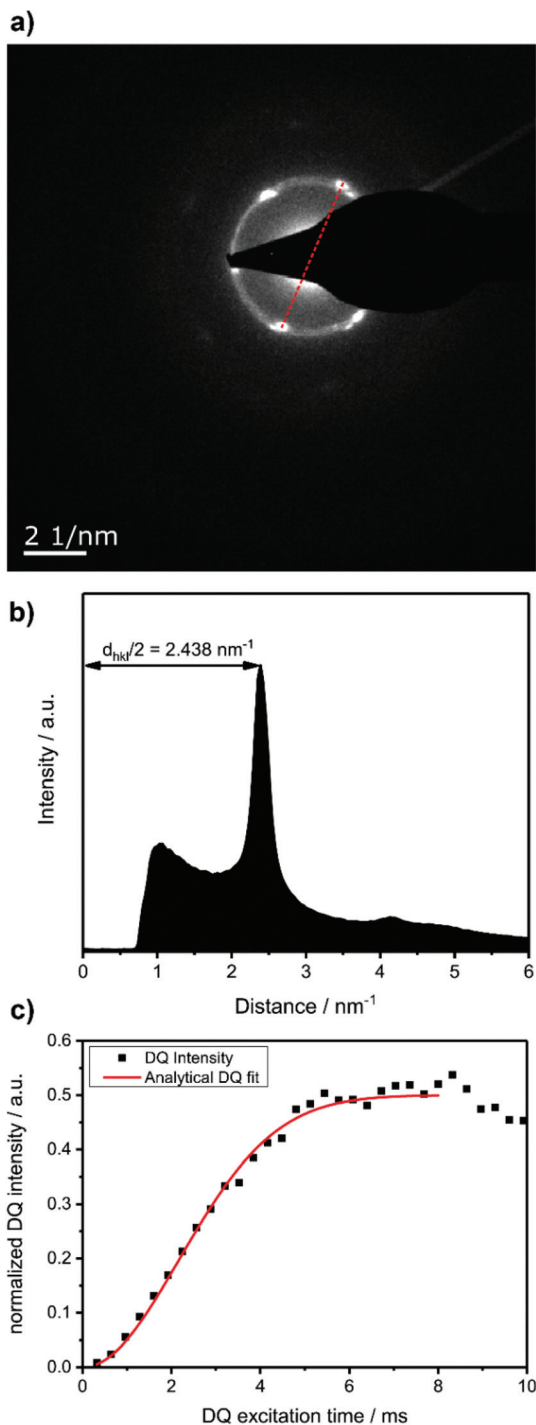


Fig. 1 (a) Diffraction pattern of a single crystal **P1** showing the (110) and (200) plane. (b) Radial intensity profile of the diffraction pattern. (c) Double quantum build-up curve of solution crystallized **P1**, recorded at 25 kHz MAS,  $T = 25\text{ }^{\circ}\text{C}$ , and 202 MHz  $^{31}\text{P}$  Larmor frequency.

Considering the adjacent-reentry model, the phosphate groups are located on the crystal surface and the interactions of phosphate groups in the  $c$  direction with neighboring crystalline layers have also to be taken into account. It should be noted that the interaction between the two phosphate planes within

a single platelet can be neglected because the distance between them is by far too large. Shifting the parallel surfaces of the two single crystals in  $x$ ,  $y$ , and  $z$  direction relative to each other the spatial distribution of the second moment  $M_2$  is computed for the different models (for illustration see Fig. S16 in the ESI†). From the computed three dimensional arrays of  $M_2$  values model dependent bended iso-surface on which  $M_2$  equals the experimentally obtained value 0.197 kHz<sup>2</sup> (Fig. 2 bottom). These iso-surfaces predict distances of 4.0–5.2 Å between the phosphorus atoms in the  $z$ -direction, which is in a reasonable range.

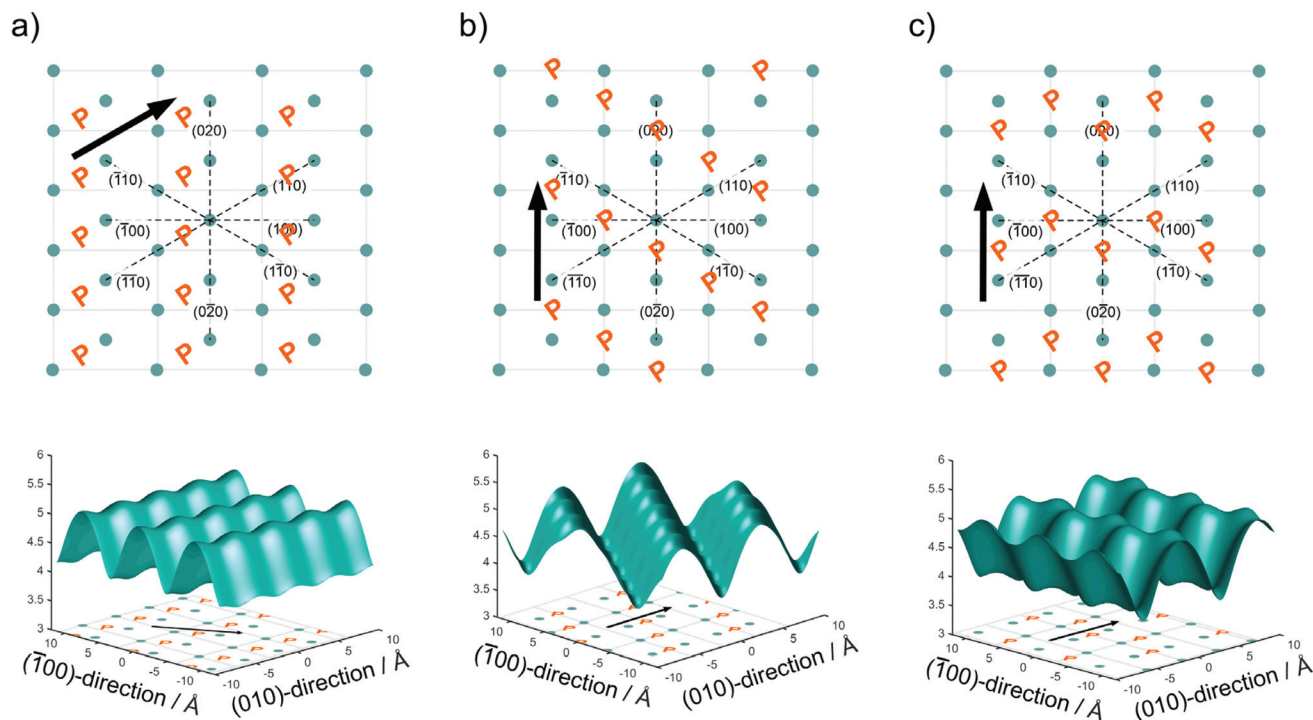
For these predicted distances, the phosphorous attached ethoxy chains between the crystal surfaces have a density between 1012 and 1320 kg m<sup>-3</sup>, which compares very well to the density of PEG (~1128 kg m<sup>-3</sup>). It should be pointed out, that the lowest density value of 1012 kg m<sup>-3</sup> results from an unfavorable geometry, where the  $^{31}\text{P}$  atoms of neighboring layers would sit directly on top of each other causing maximal steric interaction between the attached ethoxy groups.

In contrast to pure polyethylene, the situation of local densities in the crystalline and the non-crystalline part of the material in the precise phosphorylated PE chains is significantly different. The phosphate groups, expelled from the solution crystallized PE, is difficult to compare with a non-crystalline PE chain. The local structure is not necessarily non-crystalline with a lower density as the phosphate group has completely different possibilities to organize locally. An overall tilting of the chains would lead to a slightly increased in plane distance of the  $^{31}\text{P}$  sites. The resulting lower  $M_2$  value for a single layer would still require the presence of the double layer in order to explain the results, however, with a slightly reduced inter layer distance. Keeping the density of the attached ethoxy groups in mind, the distance between two adjacent “ $^{31}\text{P}$  layers” do not vary significantly.

### Surface modification of polymer platelets

The chemical accessibility of the pendant chains in the phosphates at the surface of the polymer platelets was studied using a multi-step reaction, which was conducted “on surface” (Scheme 2). The polymer platelets were prepared by solution crystallization of **P2**, which was prepared as followed: **M2** was polymerized *via* ADMET (**P2a**,  $M_n = 20\,500\text{ g mol}^{-1}$ ,  $M_w/M_n = 2.07$ ), the purified polymer was then hydrogenated using a Pd/C catalyst to give **P2** ( $M_n = 15\,900$ ,  $M_w/M_n = 1.67$ ). To prepare the dispersion of the polymer platelets of **P2**, the polymer was dissolved in ethyl acetate at 60 °C at a concentration of 1 mg mL<sup>-1</sup>. Then the solution was cooled in a temperature bath, which was set to 20 °C. A part of the cooled solution was transferred to a separate vial as a reference for later TEM images. The first reaction step “on surface” was the nucleophilic substitution of the bromides in the polymer platelets of **P2** by potassium thioacetate to **P3**. Afterwards, a small amount of sample was taken and the solvent was removed at reduced pressure. The dried polymer platelets were then dissolved in deuterated chloroform and  $^1\text{H}$  NMR spectroscopy proved the characteristic resonances at 3.17 ppm





**Fig. 2** Arrangement of the P atoms on the crystal surface depending on the direction of the chain folding along the crystal lattice, indicated by the black arrow (a) polymer chain propagating along the (110) direction of the crystal lattice (Model 1). (b) Polymer chain propagating along the *b* axis of the crystal shifted by *b*/2 (Model 2). (c) Polymer chain propagating along the *b* axis of the crystal shifted by *b*/2 alternating (Model 3). (bottom) Plane on which  $M_2$  equals the experimentally obtained value of 0.197 kHz<sup>2</sup>.

(methylene group next to the thioacetate group) and 2.35 ppm (methyl group of the thioacetate group) for **P3** (Fig. S11†) for a successful modification. When the same reaction was conducted in solution with **P2**, the NMR spectra of the product proved the same resonances for **P3** (Fig. S9†). The <sup>31</sup>P NMR spectrum shows a single signal at −1.13 ppm (Fig. S10†) confirming only one phosphorus species present. The TEM images in Fig. S12 and S13† prove the unchanged lateral dimensions of the polymer platelets of **P2** and **P3** before and after the “on surface” reaction. In DSC measurements, **P3** shows a  $T_g$  of −49 °C and a  $T_m$  of 58 °C (Fig. 3). The second reaction step was the hydrolysis of the 2-acetylthioethyl protective group, which was conducted “on surface” as well. The 2-acetylthioethyl ester group is a protective group for phosphoric acid (P–OH-groups) and can be cleaved under acidic or basic conditions to release P–OH functionality, which should alter the surface properties of the polymer platelets.<sup>38</sup>

The dispersion was cooled to 0 °C for 15 min before starting stirring and adding the hydrazine in ethanol. After *ca.* 45 min, aggregation of the crystals was visible. The solvent was allowed to evaporate while keeping the temperature at 0 °C. The product (**P4**) was not soluble in any solvent, as expected for a polyphosphodiester.<sup>31</sup>

Fig. 4 shows the TEM images of the crystal dispersion before (**P3**) (Fig. 4a and b) and after (**poly6**) (Fig. 4c and d) the addition of hydrazine. Before the treatment, the crystals exhibited a size of around 128 ± 40 nm and are crystalline as it is

apparent from the diffraction pattern. After the addition of the hydrazine, the crystal size remained at around 132 ± 46 nm and the diffraction pattern still proves their crystallinity. The P–OH groups on the crystal surface induced hydrogen bonds, which lead to the stacking of the crystals, with P–OH as an H-bond donor and P=O as an H-bond acceptor.<sup>31</sup> Fig. 4c shows a TEM image of the crystals stacking on top of each other.

IR spectroscopy proved the successful cleavage of the thioacetyl-protective group and the formation of the P–OH groups as shown in Fig. 5. The C=O stretching frequency at 1694 cm<sup>−1</sup> and the C(O)–S stretching frequency at 625 cm<sup>−1</sup>, which are characteristic for thioacetates,<sup>39</sup> vanished and a new resonance at 1200 cm<sup>−1</sup> appeared, which is characteristic for P=O stretching in acidic phosphates.<sup>40</sup>

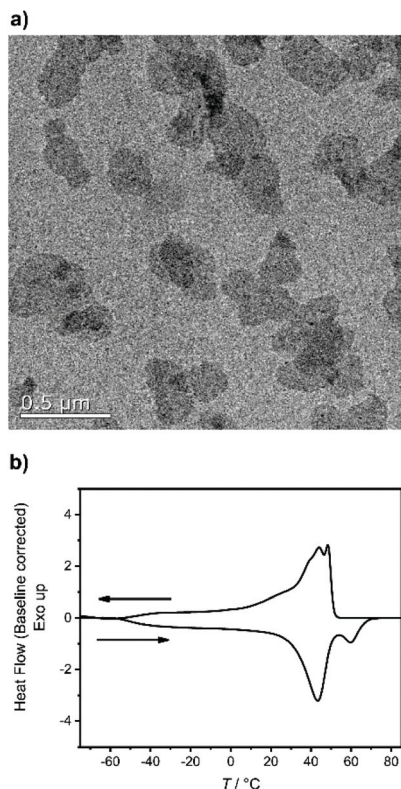
## Experimental section

### Materials and methods

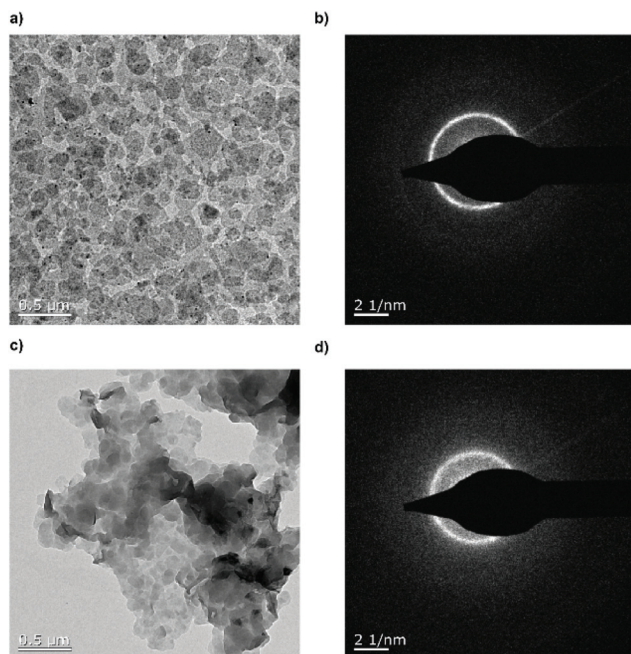
All chemicals were purchased from commercial suppliers as reagent grade and used without further purification.

**NMR.** <sup>1</sup>H and <sup>31</sup>P nuclear magnetic resonance (NMR) spectra were recorded on Bruker AV 300 at 300 MHz (<sup>1</sup>H) and 121.50 MHz (<sup>31</sup>P) or Bruker AV 700 spectrometers at 700 MHz (<sup>1</sup>H). The temperature of measurement is indicated in the corresponding figure captions. Chemical shifts are reported in

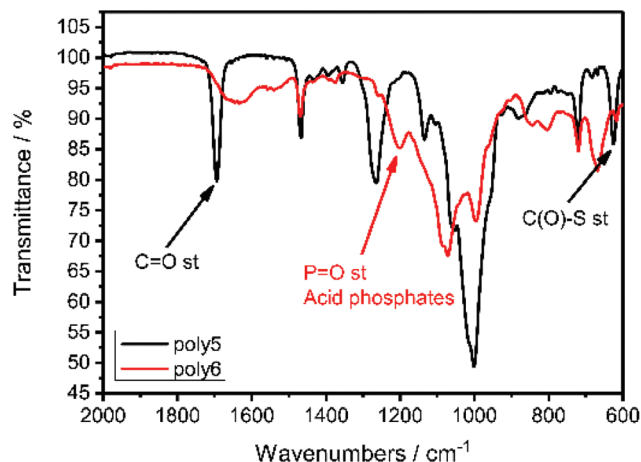




**Fig. 3** (a) Transmission electron microscope micrograph of the single P3 crystals ( $0.1 \text{ mg mL}^{-1}$ ). (b) Differential scanning calorimetry (baseline corrected) of P3 showing the second heating and cooling curve with a  $T_g$  at  $-49 \text{ }^\circ\text{C}$  and  $T_m$  at  $58 \text{ }^\circ\text{C}$  determined in the second heating curve.



**Fig. 4** (a) EM micrograph of  $1 \text{ mg mL}^{-1}$  P3. (b) EM diffraction pattern of  $1 \text{ mg mL}^{-1}$  P3. (c) Transmission electron microscope image of  $1 \text{ mg mL}^{-1}$  P3. (d) Transmission electron microscope diffraction pattern of  $1 \text{ mg mL}^{-1}$  P4.



**Fig. 5** FTIR spectrum of P3 and P4 (after two step reaction) highlighting the important frequencies.

ppm using the residual non-deuterated solvent signals as an internal reference.

**DSC.** The thermal properties of the synthesized polymers have been measured by differential scanning calorimetry (DSC) on a Mettler Toledo DSC 823 calorimeter. Three scanning cycles of heating/cooling were performed in a nitrogen atmosphere ( $30 \text{ mL min}^{-1}$ ) with a heating and cooling rate of  $10 \text{ }^\circ\text{C min}^{-1}$ .

**SEC.** Size exclusion chromatography (SEC) measurements were performed in THF on an Agilent Technologies 1260 instrument consisting of an autosampler, pump and column oven. The column set consists of 3 columns: SDV  $10^6 \text{ \AA}$ , SDV  $10^4 \text{ \AA}$  and SDV  $500 \text{ \AA}$  (PSS Standards Service GmbH, Mainz, Germany), all of  $300 \times 8 \text{ mm}$  and  $10 \text{ }\mu\text{m}$  average particle size were used at a flow rate of  $1.0 \text{ mL min}^{-1}$  and a column temperature of  $30 \text{ }^\circ\text{C}$ . The injection volume was  $100 \text{ }\mu\text{L}$ . Detection was accomplished with an RI detector (Agilent Technologies). The data acquisition and evaluation were performed using PSS WINGPC UniChrom (PSS Polymer Standards Service GmbH, Mainz, Germany). Calibration was carried out by using polystyrene provided by PSS Polymer Standards Service GmbH (Mainz, Germany).

**Transmission electron microscopy (TEM).** The crystal morphology, thickness, and crystal structure were determined using an FEI Tecnai F20 transmission electron microscope operated at an acceleration voltage of  $200 \text{ kV}$ .

The crystallization behavior of these polymers was studied by drop-cast TEM measurement. A TEM specimen of solution-grown polymer crystals was prepared from a  $1 \text{ mg mL}^{-1}$  or  $0.1 \text{ mg mL}^{-1}$  dispersion in ethyl acetate. The solution was heated and cooled according to the different temperature profiles. One drop of each dispersion was applied to a carbon-coated TEM grid, excess liquid was blotted off with a filter paper and the specimen was allowed to dry under ambient conditions.

**Solid-state NMR.** The solid-state NMR measurements have been performed on a Bruker Avance III console operating at



500.2 MHz  $^1\text{H}$  Larmor frequency using a commercial double-resonance MAS NMR probe supporting zirconia rotors with 2.5 mm outer diameter at 25 kHz MAS spinning frequency and 100 kHz rf nutation frequency on the variable x-channel of the probe tuned to the  $^{31}\text{P}$  Larmor frequency of 202 MHz. The BABA-xy16 sequence<sup>36</sup> has been applied for DQ excitation and reconversion. For data analysis the normalization and fitting procedures described for static  $^1\text{H}$  DQ applications by Saalwächter in detail<sup>41</sup> have been applied to our  $^{31}\text{P}$  DQ data, acquired under fast MAS conditions. Combining the analytical expression for DQ build-up depending on the dipolar second moment  $M_2$  (Saalwächter *et al.*,<sup>41</sup> eqn (15)) and the DQ excitation efficiency of the BABA-xy16 pulse sequence (Saalwächter *et al.*,<sup>36</sup> eqn (11)) we get

$$I_{\text{DQ}}(\tau_{\text{DQ}}) = \frac{1}{2} \left( 1 - \exp \left\{ \frac{16}{3\pi^2} M_2 \tau_{\text{DQ}}^2 \right\} \right)$$

for the analytical description of the  $^{31}\text{P}$  DQ build-up, where  $\tau_{\text{DQ}}$  is the experimental DQ excitation time and the dipolar  $^{31}\text{P}$  second Moment  $M_2$  the only free parameter.<sup>36</sup>

## Syntheses

**2-Bromoethyl di(undec-10-en-1-yl) phosphate (M2).** A three-necked round bottom flask, equipped with a dropping funnel, was charged with 2-bromoethyl phosphorodichloridate (11.87 g, 40.1 mmol, 1 eq.), dissolved in dry toluene (100 mL) under an argon atmosphere. The solution was cooled with a water bath. Triethylamine (14.3 mL, 103.1 mmol, 2.1 eq.), and 10-undecen-1-ol (20.7 mL, 103.1 mmol, 2.1 eq.) were dissolved in dry toluene (60 mL) and added slowly *via* the dropping funnel. After the addition, the reaction was stirred overnight at room temperature. The crude mixture was concentrated at reduced pressure, dissolved in diethyl ether, and filtered. The organic phase was washed with  $\text{NaHCO}_3$  solution, with 10% aqueous hydrochloric acid solution, and with brine. The organic layer was dried over magnesium sulfate, filtered, concentrated at reduced pressure, and purified by flash chromatography over neutral alumina using dichloromethane as eluent to give a clear yellowish liquid.

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*,  $\delta$ ) 5.82 (ddt,  $J = 17.0, 10.3, 6.7$  Hz, 2H), 4.97 (dd,  $J = 18.3, 13.6$  Hz, 4H), 4.31 (q,  $J = 6.8$  Hz, 2H), 4.06 (p,  $J = 6.4, 6.0$  Hz, 4H), 3.55 (t,  $J = 6.3$  Hz, 2H), 2.05 (q,  $J = 7.0$  Hz, 4H), 1.70 (p,  $J = 6.7$  Hz, 4H), 1.30 (s, 24H).  $^{31}\text{P}$  {H} NMR (121 MHz, Chloroform-*d*,  $\delta$ ) -1.25 (s, 1P).

**P2a: M2** (500 mg, 0.98 mmol) was placed in a 25 mL Schlenk tube equipped with a stir bar and the system was degassed by three consecutive Argon/vacuum cycles. 8 mg of Grubbs catalyst 1<sup>st</sup> generation (0.1 mol%) were added and polymerization was carried out at 40 °C at reduced pressure (0.1 mbar). Increasing viscosity of the solution indicated ongoing polymerization. After 18 h, the second portion of Grubbs catalyst 1<sup>st</sup> generation was added and the polymerization was continued for an additional 24 h. The solution was allowed to cool down to room temperature, then 2 mL  $\text{CH}_2\text{Cl}_2$  were added to dissolve the polymer and 200  $\mu\text{L}$  ethyl vinyl

ether were added to quench the catalyst. The mixture was kept stirring for 2 h while its color changed from purple to orange. Further  $\text{CH}_2\text{Cl}_2$  was added to dilute the solution and the polymer was precipitated from methanol. After centrifugation, the product was isolated and dried under vacuum to yield a dark yellow, honey-like polymer (67% yield).

SEC (PS standard, THF as the eluent):  $M_n = 20\,500$  g mol<sup>-1</sup>,  $M_w = 42\,500$  g mol<sup>-1</sup>,  $M_w/M_n = 2.07$ .

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta = 5.36$  (b, 2H,  $-\text{CH}_2-\text{CH}=\text{CH}-$ ), 4.29 (q,  $J = 6.9$  Hz, 2H,  $-\text{O}-\text{CH}_2-\text{CH}_2-\text{Br}$ ), 4.05 (q,  $J = 6.8$  Hz, 4H,  $-\text{OPO}_3-\text{CH}_2-$ ), 3.53 (t,  $J = 6.3$  Hz, 2H,  $-\text{O}-\text{CH}_2-\text{CH}_2-\text{Br}$ ), 1.98 (m, 4H,  $-\text{CH}_2-\text{CH}=\text{CH}-$ ), 1.68 (m, 4H,  $-\text{OPO}_3-\text{CH}_2-\text{CH}_2-$ ), 1.41–1.24 (m, 24H, alkyl).  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta = 130.32, 68.15$  (d,  $J = 6.2$  Hz), 66.47 (d,  $J = 5.3$  Hz), 32.62, 30.26 (d,  $J = 6.8$  Hz), 29.78–29.16 (m), 27.23, 25.42.  $^{31}\text{P}$  NMR (121 MHz,  $\text{CDCl}_3$ )  $\delta = -1.24$ .

**P2: P2a** (260 mg, 0.54 mmol) was dissolved in 10 mL dry toluene in a glass vessel equipped with a stirring bar. Argon was bubbled through the solution for 10 min to remove oxygen. 52 mg of 10 wt% Pd/C was added, and the glass vessel was charged in a 250 mL ROTH autoclave. Hydrogenation was performed at 50 °C and 50 bar of  $\text{H}_2$  for 19 h. Pd/C was removed by filtration and the polymer was obtained after solvent evaporation as an off-white, hard material (quantitative yield).

SEC (PS standard, THF as the eluent):  $M_n = 15\,900$  g mol<sup>-1</sup>,  $M_w = 26\,500$  g mol<sup>-1</sup>,  $M_w/M_n = 1.67$ .

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ):  $\delta = 4.29$  (q,  $J = 8.4, 6.3$  Hz, 2H,  $-\text{O}-\text{CH}_2-\text{CH}_2-\text{Br}$ ), 4.04 (q,  $J = 6.4$  Hz, 4H,  $-\text{OPO}_3-\text{CH}_2-$ ), 3.53 (t,  $J = 6.3$  Hz, 2H,  $-\text{O}-\text{CH}_2-\text{CH}_2-\text{Br}$ ), 1.68 (p,  $J = 6.7$  Hz, 4H,  $-\text{OPO}_3-\text{CH}_2-\text{CH}_2-$ ), 1.25 (s, 32H, alkyl), 0.87.  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta = 68.16$  (d,  $J = 6.2$  Hz), 66.47 (d,  $J = 5.3$  Hz), 30.26 (d,  $J = 6.8$  Hz), 29.76–29.44 (m), 29.15, 25.43.  $^{31}\text{P}$  NMR (121 MHz,  $\text{CDCl}_3$ )  $\delta = -1.25$ .

**P3:** A Schlenk tube was charged with **P2** (48.1 mg, 93.7  $\mu\text{mol}$ , 1 eq.) and potassium thioacetate (12.8 mg, 112.4  $\mu\text{mol}$ , 1.2 eq.), dissolved in dry THF (3 mL) under an argon atmosphere. The reaction was stirred overnight at 40 °C. The crude mixture was filtered, concentrated at reduced pressure, and dissolved in dichloromethane. The organic phase was washed with distilled water. The organic layer was dried over magnesium sulfate, filtered, and concentrated at reduced pressure to give a yellowish powder.

$^1\text{H}$  NMR (250 MHz, Chloroform-*d*,  $\delta$ ) 4.07 (m, 6H), 3.17 (t,  $J = 6.5$  Hz, 2H), 2.35 (s, 3H), 1.67 (t,  $J = 7.0$  Hz, 4H), 1.25 (m, 32H).  $^{31}\text{P}$  {H} NMR (202 MHz, Chloroform-*d*,  $\delta$ ) -1.13 (s, 1P).

**P4:** A vial was charged with **P3** (30 mg, 59.0  $\mu\text{mol}$ , 1 eq.) and ethyl acetate (30 mL). The dispersion was heated to 60 °C and allowed to cool to R.T. for 4–5 h. Then the crystal dispersion was cooled to 0 °C and hold at this temperature for 15 min. The stirring was turned on and 1 M hydrazine solution in ethanol (2 mL, 2 mmol, 40 eq.) was added while keeping the temperature at 0 °C. After 24 h the solvent was evaporated with a flow of nitrogen while still keeping the temperature at 0 °C.





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