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Sustainability spotlight

The persistence of PFAS in the environment and their bioaccumulation in humans has prompted their regulation, improved detection and analysis, remediation and destruction, as well as their replacement by safer alternatives. The tutorial review has highlighted areas where chemists and collaborators can help to manage problems associated with PFAS. Specifically, in alignment with the UN sustainability goals, remediation of PFAS from water (Goal 6, clean water and sanitation), finding safe alternatives to replace PFAS in consumer products (Goal 3, good health and well-being) and regulators limiting the manufacture and use of PFAS which in turn, lowers the concentrations we are exposed to (Goal 12, responsible consumption and production).

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ARTICLE

Addressing the Persistence of Per- and Poly-fluoroalkyl substances (PFAS): Current Challenges and Potential Solutions

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The combined stability, mobility, and bioaccumulation of per- and poly-fluoroalkyl substances (PFAS) has prompted a global environmental crisis. PFAS have unique properties owing to their strong, hydrophobic C-F bonds, which result in their resistance to water, oil, chemicals, and heat. Applications of PFAS include their use as water-, grease-, and fire-proof coatings, emulsifiers, and surfactants, spanning most manufacturing sectors. The continued regulation of specific PFAS provides significant research opportunities for chemists and their collaborators across environmental, social, engineering, and materials sciences. Solutions in the areas of detection and analysis, immobilisation and destruction, and the creation of viable and safe alternatives are urgently needed. In this tutorial review, PFAS and their associated challenges are described, followed by a summary of existing solutions and future research opportunities.

1. Overview of PFAS

1.1. Background and synthesis

The specific definition of per- and polyfluoroalkyl substances (PFAS) has evolved over time from "highly fluorinated substances" to a more concise definition that reflects the chemical structure of the compound.[1] Accordingly, PFAS are organofluorine compounds that contain at least one CF_3 or CF_2 moiety in their structure (red, Figure 1).[2, 3] Many of these substances are amphiphilic in nature as they also contain a hydrophilic head (blue, Figure 1a). They have important applications in everyday products such as non-stick protective coatings, carpets, clothing, lubricants and as fire suppressors in aqueous film-forming foams (AFFFs).[4] PFAS are characterised by strong C-F bonds and thermal and chemical stability, making PFAS resistant to environmental degradation.[5] Their inertness can be explained by the electronegative fluorine atoms clustering around the carbon atom, shielding the atoms from chemical reactions. As a result of their widespread use, environmental persistence, high mobility, bioaccumulation, and toxicity, PFAS are now regarded as high-risk environmental chemicals,[6, 7] and are widely referred to as "*forever chemicals.*"

The accidental discovery of poly(tetrafluoroethylene) (PTFE) ($[CF_2]_n$, n = length of polymer chain; Figure 1b) on April 6, 1938, at DuPont by Roy J. Plunkett was the first ever PFAS produced.[8] At DuPont, tetrafluoroethylene (C_2F_4), made by zinc dichlorination of ClF_2CCF_2Cl, was stored in cylinders where it polymerised slowly over time to form a white solid, PTFE. After controlled polymerisation and polymer fabrication techniques, DuPont commercialised PTFE under the trade name Teflon® in 1948.[8]

PFAS are synthesised through two main processes: free radical electrochemical fluorination (ECF, also known as Simons Electrochemical Fluorination)[9] and fluorotelomerisation (Scheme 1).[3] In the ECF process, an organic compound (e.g., sulfonyl halide, $C_xH_ySO_2X$, X = F or Cl,[10] Scheme 1a) is dissolved in anhydrous HF for fluorination at the anode, usually nickel. As all of the C-H bonds are replaced with C-F bonds and the C-C multiple bonds are saturated with F atoms, the resulting acid, H^+, is concomitantly reduced to hydrogen gas at the cathode.[3, 11] ECF was a dominant synthetic process for PFAS until the 1990s and usually results in a mixture of linear (70%) and branched (30%) PFAS.[3, 12] The yields of the products decrease with an increase in chain-length.[9] Using ECF, 3M (formerly known as Minnesota Mining and Manufacturing Company) commercially synthesised perfluorooctanesulfonic acid (PFOS)[13] from the perfluorooctanesulfonyl fluoride (POSF) precursor, as well as perfluorooctanoic acid (PFOA),[14] to be widely applied in aqueous film forming foams (AFFF), treated textiles, and cookware coatings.

A typical fluorotelomerisation process involves the reaction of perfluoroethyl iodide (C_2F_5I), the starting transfer agent or talogen,[15, 9] with the unsaturated tetrafluoroethylene (C_2F_4) (taxogen) to yield a mixture of even-numbered carbon linear perfluoroalkyl iodides (Scheme 1b, Telomer A). Telomer A is then reacted with ethylene (C_2H_4) to form fluorotelomer iodide (Scheme 1b, Telomer B). Telomer B further reacts with water or

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oleum to make fluorotelomer alcohols (FTOHs). Telomer A, Telomer B and FTOHs are the basic raw materials used to manufacture fluorotelomer-based surfactants (non-polymer) and polymer products.[3]

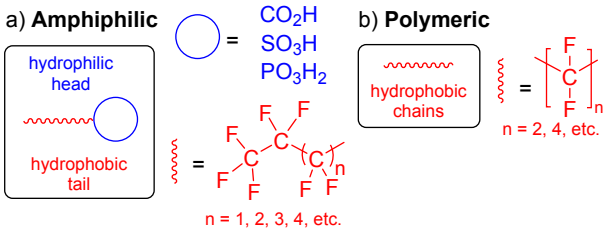
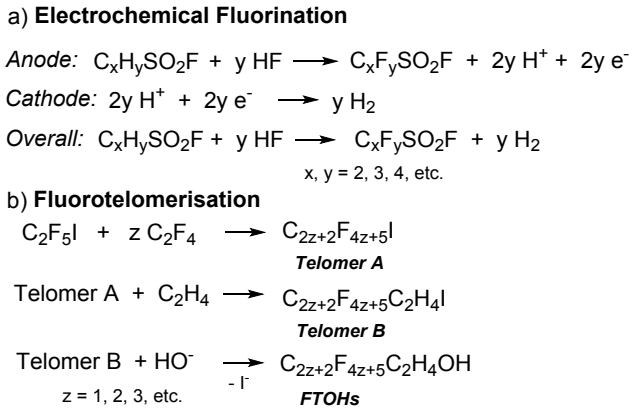


Figure 1. Generic structure of a) amphiphilic (non-polymeric), and b) polymeric PFAS



Scheme 1. Synthetic routes to PFAS

1.2. Classes

The functional groups and the length of the fluorocarbon chain dictate the class and subclass of a specific PFAS (Figure 2).[3] Polymeric PFAS have a longer chain backbone consisting of several repeat units (i.e., >10). There are three subclasses of polymeric PFAS: fluoropolymers, side-chain fluorinated polymers, and perfluoropolyethers (Figure 2). Non-polymeric PFAS typically contain a backbone with 2-10 fluorocarbon repeat units and fall into either perfluoroalkyl (fully fluorinated carbon chain) or polyfluoroalkyl (partially fluorinated carbon chain) subclasses. Further classification is based on the different hydrophilic head functional groups attached to the fluorocarbon backbones, and most commonly are carboxylic (CA), sulfonic (SA), and phosphonic acids (PA). Ultrashort chain PFAS are classified based on the number of units (i.e., 1-3).[16, 17] Typical examples are perfluoroethane sulfonate (PFEtS, Et = ethyl = 2 units, $CF_3CF_2SO_3^-$), and perfluoropropanoic acid (PFPrA, Pr = propyl = 3 units, note that in the case of CAs the C in the COOH head group counts as 1 unit, $CF_3CF_2CO_2H$). By contrast, short-chain PFAS typically contain 4-6 perfluorocarbon units,[18] and long-chain PFAS contain ≥ 6 or 7 perfluorocarbon units.[19] While many PFAS are open-chain molecules (linear, straight or branched), cyclic PFAS also exist. Examples include perfluoromethylcyclohexane sulfonate (PFMeCHS, $CF_3C([CF]_2)_5SO_3H$) and perfluoroethylcyclohexane sulfonate (PFEtCHS, $CF_3CF_2C([CF]_2)_5SO_3H$).[20, 21]

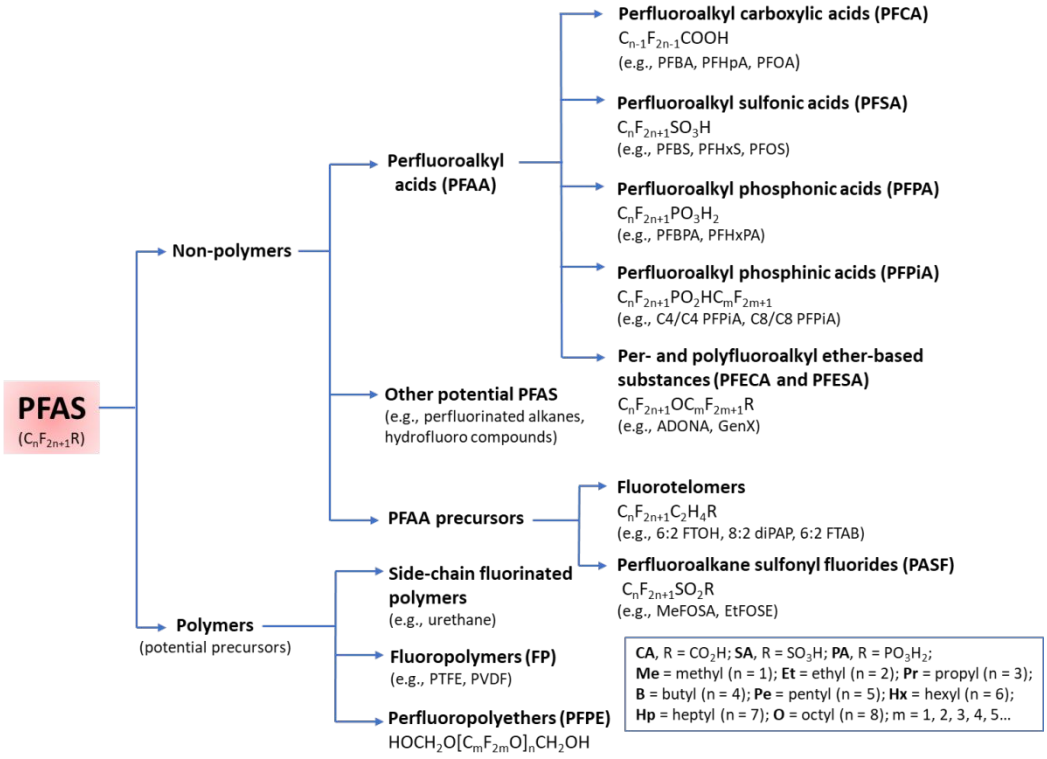


Figure 2. A summary of PFAS classes and subclasses. Adapted with permission from Z. Wang, J. C. DeWitt, C. P. Higgins and I. T. Cousins, *Environ. Sci. Technol.*, 2017, **51**, 2508-2518. Copyright 2017 American Chemical Society.[22]

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1.3. Uses

PFAS possess important physical and chemical properties desired by chemical manufacturing industries, such as high thermal and chemical stability. The strong C-F bonds in PFAS (e.g., 485 kJ/mol *c.f.* 346 kJ/mol for C-C) requires temperatures >1000°C to degrade and help to impart these features.[8, 23] Their complementary hydrophobic and hydrophilic structure (red tail and blue head, Figure 1) is ideal for generating surfactants, and their hydrophobicity and lipophobicity are useful in water-proof[24] and grease-proof[25] coatings. Hence, PFAS are used in numerous sectors for the manufacture of everyday items (Table 1). Recent regulations (see section 2.0), have prompted the development of safer alternatives for both essential (e.g., selected medical applications, occupational clothing, Nafion™ membranes) and non-essential (e.g., food packaging, personal care products) uses of PFAS (Table 1 and ensuing discussion in section 5.0).[26, 27]

Table 1. Selected examples of uses of PFAS in different sectors and potential alternatives

Sector	Desirable Properties	Uses	Potential Alternatives
Food[28, 29]	• grease/oil-repellent • thermally stable	• fast-food, food, and microwavable wrappers, containers, trays, and bowls • coatings for non-stick cookware • supercritical fluids used in ceramic powders	• physical: cellulose based, clay, bamboo, wheat straw, aluminium • chemical: silicones, synthetic biopolymers, bio-waxes
Textiles[30]	• water/oil/stain-repellent • thermally and chemically stable	• occupational protective and durable outdoor clothing • coatings for carpets and furniture	• silicones, hydrocarbon-based, dendrimer chemistry, inorganic nanoparticles
Personal care products[31, 32]	• film forming • stabilising • surfactant • emulsifier	• sunscreen • hair, face, and body products	• silicones, synthetic waxes, bio-based oils, fats
Firefighting[27]	• surfactant • fire-suppressant	• Aqueous film-forming foams (AFFFs)	• class B F3 foams: hydrocarbons, detergents, siloxanes, proteins
Oil and gas mining[28, 33]	• surfactant	• enhance oil recovery • mine floating • well simulation additives • solution for hydrostatic blockage • tracers in geological communication	• silicone/siloxane-based antifoaming agents • halogenated and radioactive tracers • fluoropolymer material alternatives: metal alloys, ceramic-based, epoxy-based, nylon
Electronics[28]	• wetting agents • water/oil- repellent	• low-foaming noncorrosive components in solders • coating surfaces or casing	• silicone based materials, bio-based polymers, fluorine-free coatings, glass-reinforced composites
Agriculture[28]	• surfactant	• pesticide products • coatings in fertilisers	• non-fluorinated surfactants
Metal plating and finishing processes[34]	• surfactant • fume suppressant • corrosion inhibitor • wetting agents • improved bath stability	• electro plating and electroless plating • metal treatment (cleaning, etching, etc.)	• non-fluorinated surfactants and fume suppressants • mechanical controls

1.4. Environmental concerns

1.4.1. Sources

Historically, industrial and manufacturing processes have been the major sources of PFAS in the environment. 3M and DuPont are two notable companies involved in the commercial manufacturing of PFAS since 1947. From 1970 to 2002, an estimated 96 thousand tonnes of POSF (a precursor for PFOS) was manufactured and used in commercial consumer products globally.[13] From this, an estimated 450-2700 tonnes of PFOS were emitted into the environment.[13] Similarly, the estimated total global production of perfluorocarboxylates (PFCA, including trifluoroacetic acid and PFOA), ammonium perfluorooctanoate and ammonium perfluorononanoate was 4400-8000 tonnes between 1951 to 2004, with an estimated 470-900 tonnes of PFCA emitted into the environment during the same period.[14]

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One of the most significant contributors to PFAS in the environment is from fire-suppressants in the form of aqueous film-forming foams (AFFFs).[35, 36] It is estimated that the US military was the source of more than 70% of AFFFs used in the US. This resulted in the release of elevated concentrations of PFAS into the environment through AFFFs, mainly on military bases, at training sites, or at municipal airports.[36] PFAS are mobile in the environment and are transported away from these sites contaminating soil, groundwater and surface water resources.

Wastewater treatment plant (WWTP) effluent is another significant contributor of PFAS in the environment due to PFAS-containing product usage, industrial discharge, and their incomplete removal during water treatment. In fact, recent studies have revealed an increase in PFAS concentrations in the liquid phase after secondary treatment, as PFAS precursors can biotransform to form PFAS. Additionally, due to their hydrophobicity, a significant portion of PFAS partitions to the solid phase in WWTPs.[37] Indeed, reports have indicated that PFAS are present at detectable levels in biosolids used in agricultural soils in Australia.[38] Plants and crops grown on biosolid-applied soils uptake PFAS and could serve as a significant exposure route of PFAS to humans and animals.[39] Apart from the use of biosolids in agricultural soils, it used to be common practice to bury PFAS-contaminated products in landfills, which directly results in contamination of groundwater sources through leachate,[40-42] and the pollution of air around landfills through landfill off-gassing.[41]

In addition to AFFFs and WWTP effluent, incineration of materials containing PFAS is another major contributor of PFAS in the environment. As of one of the main destruction methods of PFAS, incineration requires high temperatures (> 1000°C) to achieve chemical breakdown.[35, 43] Specifically, incineration temperatures of up to 1200°C are required to destroy waste containing PFOS, while over 1000°C is required to breakdown PFAS adsorbed on spent activated carbon, and ca. 1100°C is required to destroy PFAS in the gas-phase.[44] According to the US-EPA, CF_4, which is the most difficult fluorinated organic compound to decompose, requires temperatures over 1400°C.[45] Despite this energy intensive process, incomplete destruction can occur, leaving small molecule PFAS and fluorine-containing by-products in the vicinity of the incineration facilities. A study of the concentration and distribution of PFAS in surface water and soil samples around a PFAS incinerator facility in the US found significant measurable amounts of PFOS (up to 8.3 µg/kg), PFOA (up to 1.3 µg/kg) and hexafluoropropylene oxide (GenX) (up to 1.5 µg/kg) in soil samples and up to 19 ng/L and 11 ng/L of PFOS and PFOA, respectively, in surface water.[46] Moreover, in southern China, a study of three municipal solid waste incineration plants found a high concentration of PFAS (up to 0.7 µg/mL) in leachate, with variability attributed to the type of waste.[47]

1.4.2. Mobility

Anthropogenic activities have allowed the release of PFAS into the environment and their infiltration into the food web.[48] PFAS have been detected thousands of kilometres away from where they are created or used: from the top of Mt. Everest[49] to the high Arctic Svalbard ice core[50] and in wildlife globally.[48, 51] The water cycle is a major way to move PFAS from one region to another.

PFAS in the atmosphere originate from the industrial stack and fugitive emissions from manufacturers and allow distribution far away from their direct sources.[52] For example, a study on the non-targeted screening of PFAS in China found at least 34 emerging PFAS in airborne particulate matter.[53] Since precipitation (rainfall or snowfall) is an effective scavenger of gas-phase and particle-bound pollutants from the atmosphere, atmospheric PFAS are deposited on soil, surface water, and vegetation.[52, 54] Moreover, melting glaciers serve as secondary sources of PFAS for the receiving lakes where PFAS are accumulated in lake sediments.[54] More importantly, emerging evidence suggests PFAS accumulate at the air-water interface (AWI), through an interaction of the hydrophilic head with the surface of the water and the hydrophobic tail with the air. As sea spray aerosols (SSA) form from wind and waves, the aerosols become enriched with a surface microlayer containing PFAS.[55 56] This is problematic as PFAS concentrations have been reported to be >100 000 times higher in SSA when compared with seawater[57] and are stable, mobile, long-term sources of PFAS.[58]

1.4.3. Persistence, bioaccumulation, and toxicity

The presence of *multiple* C-F bonds on geminal carbons in PFAS provides additional strength to their structures.[59] Hence, PFAS can resist adverse environmental conditions, including high temperatures. In addition, fluorine has a strong electron-withdrawing effect, which results in the formation of strong and very difficult to break C-F bonds in PFAS.[59] As a result, PFAS are persistent in the environment.

PFAS concentration, the length of the C-F chain, the presence of specific functional groups, the biotransformation of PFAS precursors, and the presence of organic matter are some of the factors that can affect the accumulation of PFAS in an organism.[60] Analysis of the bioaccumulation factors (BAFs) for 27 genera of agricultural crops from 24 studies revealed that 45 PFAS accumulated less in reproductive and storage organs than in vegetative organs (e.g., leaves, shoot buds, root, and stem).[61] This is likely because the vegetative organs are mainly responsible for the transport of plant nutrients, thereby accumulating PFAS in the process. A study of bioaccumulation of PFAS by benthic macroinvertebrates (worms, snails, and mussels) found that BAFs in worms (412.84 L/kg) were higher than in snails (8.08 L/kg) and mussels (27.12 L/kg).[62] In a human study of autopsy tissues, perfluorobutanoic acid was found to have accumulated in lung tissues (median value: 807 ng/g) and in kidneys (median value: 263 ng/g). Similarly, perfluorohexanoic acid was found to have accumulated in the

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liver tissues (median value: 68.3 ng/g) and in the brain tissues (median value: 141 ng/g). In addition, PFOA was found to have accumulated in the bone (median value: 20.9 ng/g).[63] Other reports have suggested that protein-PFAS interactions in humans could be the major molecular mechanisms responsible for the bioaccumulation of PFAS in specific human tissues. For example, the liver-type-fatty acid binding proteins in the liver and serum albumin have high binding affinity to PFAS, hence their accumulation in these tissues.[64]

Bioaccumulation of PFAS in human body tissues has been linked to some adverse health conditions.[63, 65] Toxicological studies in animals (mice) have correlated high concentrations of PFAS to endocrine disruption, delay in physical development, cancer and neonatal mortality.[65-67] A study in zebrafish (*Danio rerio*) exposed to PFAS found abnormal ventroflexion of the tail and failed swim bladder inflation.[68] In humans, PFAS have been reported to activate receptors associated with carcinogenesis, e.g. peroxisome proliferator-activated receptors (PPARs), due to their structural resemblance to fatty acids and can disrupt lipid metabolism, resulting in dyslipidemia.[65, 69] The binding of PFAS to PPAR has been linked to poor fetal growth and immune function.[65]

2.0 Policies, Management, and Regulation

The persistence of PFAS in the environment has prompted government agencies to adopt international agreements that aim to restrict the use of these chemicals (Stockholm Convention on Persistent Organic Pollutants) and to issue regulatory frameworks on the use of PFAS (Figure 3). For example, in 2006 the US-EPA issued a PFOA stewardship program aimed at a 95% reduction in emission of PFOA and related chemicals by 2015, including a health advisory level of 70 ng/L for both PFOA and PFOS in 2016.[70] In 2018, the US Agency for Toxic Substances and Disease Registry issued provisional daily oral minimal intake risk levels for PFAS as a guideline for the public. Similar regulatory frameworks have been issued in other countries.

The US-EPA has also provided guidelines for the remediation and destruction of PFAS contamination and has recently dramatically reduced the drinking water health advisory levels.[71] An enforceable drinking water maximum contaminant level (MCL) of 4 ng/L for PFOS and PFOA was proposed by the US-EPA in March 2023. Similarly, the US-EPA set a chronic oral reference dose of 3 ng/kg daily limit for the PFOA replacement, (GenX).[35] In July 2022, the US House of Representatives passed a bill to limit PFAS in wastewater discharge from organic chemical manufacturers and mandated the manufacturers to provide supply production, use, and other data to the EPA. The law is expected to come into effect in 2024.[72] The European Union (EU) stated that it was necessary to introduce a "strategy that addressed all PFAS through regulatory and non-regulatory interventions", at the latest by 2025 and to be in effect by 2030, with the ultimate aim of phasing out PFAS at the EU-level.[73] Under the EU's Persistent Organic Pollutants Regulation, PFOS was restricted for use in the EU, according to European Chemicals Agency (ECHA). In 2023, the ECHA also introduced a proposal to restrict the use of around 10,000 PFAS in materials with the aim of reducing PFAS emissions into the environment, thereby making products and processes safer for people.[74] In addition, to ensure global elimination from products, PFOS, and its derivatives were included in the international Stockholm Convention in 2009, while PFOA and its derivatives were banned in 2020. Similarly, perfluorohexane sulfonate (PFHxS), which was a replacement for PFOS, has been included in the Stockholm Convention, with a global ban expected to follow.[75]

Regulations on PFAS are country and region dependent (Figure 3). For example, in 2014, Norway prohibited PFOA in solid and liquid consumer products, including carpets and textiles and set regulatory limits of 10 mg/kg PFOA in liquid mixtures, 1000 mg/kg PFOA in solid products and 1 μg/m^2 PFOA in textile materials. Food contact materials including paper and cardboard with any level of fluorine content were banned by Denmark in 2020.[76] While in the USA, the use of AFFF will be prohibited by Oct 2024[77] and NZ is one of the first to ban the use of PFAS in all cosmetics, from Dec 2026.[78] These initiatives, while location specific, are crucial to generate international momentum towards further regulating PFAS in specific applications as well as across sectors.[79, 80]

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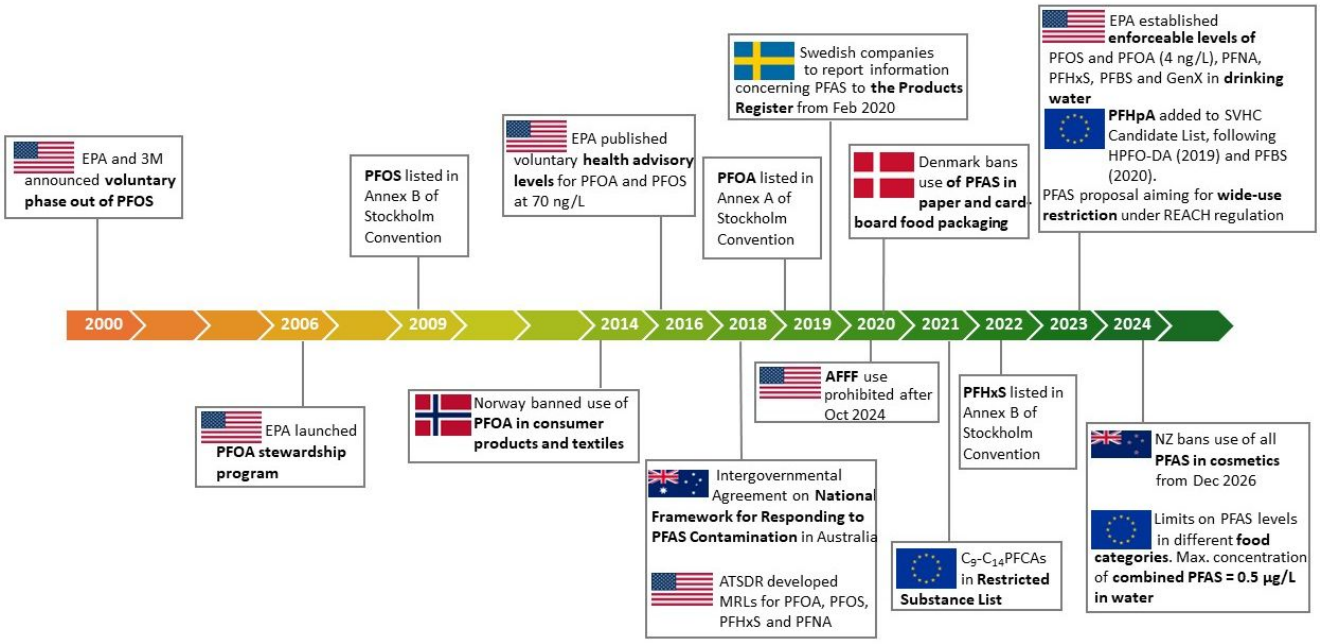

Figure 3. Examples of the evolving PFAS regulatory landscape. Adapted from C. Schiavone and C. Portesi, *Applied Sciences*, 2023, **13**, 6696. Copyright 2023 MDPI.[79]

3.0 Detection and Analysis

Detecting and analysing PFAS presents formidable challenges. Current analytical techniques have mainly centred on a limited subset of PFAS, perfluoroalkyl acids (PFAAs).[81] The complex landscape of proprietary or mixed PFAS complicates the situation, as does the potential for a single PFAS parent to generate a mixture of intermediate transformation products. Targeted liquid chromatography coupled with high-resolution mass spectrometry (LC-HRMS) or tandem mass spectrometry (LC-MS/MS) captures many known PFAS, while non-targeted HRMS identifies additional suspected or previously uncharacterised PFAS (Figure 4).[82] Surrogate techniques through bulk organofluorine measurements, such as total fluorine and total organic fluorine (TOF) analysis, provide complementary information about the unidentified fraction of PFAS in environmental samples.[83, 84] This multifaceted approach,[83] while not without challenges, is essential for unravelling the complex landscape of PFAS contamination, identifying precursors, and ultimately contributing to informed environmental management and health risk assessments.

Figure 4. A generic summary of the analytical methods for PFAS detection and analysis.

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3.1. Targeted methods

Targeted analysis of PFAS screens analytes against a library of known analytical standards, typically <50 PFAS. LC-HRMS or LC-MS/MS are the most conventional, sensitive, and selective techniques for analysing PFAS from environmental matrices due to the presence of ionisable groups (e.g., COO^- or SO_3^-). Mass spectrometry has an extremely low detection limit (in the ng/L); however, it is cumbersome (usually requiring solid-phase extraction processes), expensive (ca. $200-300 USD per sample), time-consuming, and requires highly trained personnel to operate, which prohibits using this method for routine measurements.[85]

As a result, complementary surrogate analytical methods aimed at screening and identifying unknown PFAS through their signature molecular masses (i.e., HRMS), or characteristic C-F bonds (i.e., TOF), are developing.[86] With all of these analyses in hand, the list of PFAS and their transformation products continues to increase. A database of PFAS according to US-EPA has a list of over 12034 PFAS compounds (as of August 2021).[87]

3.2. Surrogate and non-targeted methods

In the realm of PFAS analysis, both surrogate techniques and advanced analytical methods play pivotal roles. The Total Oxidizable Precursor Assay (TOPA) selectively targets compounds that can be oxidised to form specific PFAAs, offering insights into PFAS precursor content by comparing PFAA concentrations before and after oxidation.[88] Total fluorine analysis, achieved through techniques like combustion ion chromatography (CIC), quantifies the overall fluorine content in a sample, including both organic and inorganic fluorine species.[89] Additionally, Fluorine-19 Nuclear Magnetic Resonance Spectroscopy (^{19}F NMR) is a versatile tool for characterising organofluorine compounds and quantifying TOF,[90] with a more recent adaptation focusing on the selective measurement of PFAS-related compounds through chemical shift monitoring.[91] Meanwhile, Particle-Induced Gamma Ray Emission (PIGE) spectroscopy provides a unique surface analysis technique for elemental fluorine quantification through proton bombardment and gamma-ray emission.[81, 92] Other methods include, inductively coupled plasma/mass spectrometry (ICP-MS/MS), used to detect fluorine in unknown organofluorine compounds,[93] and surface-enhanced Raman spectroscopy (SERS), a sensitive technique able to identify the unique molecular signatures in PFAS after interacting with a substrate (e.g., Au nanoparticles).[94]

It is no surprise that the number of analytical methods to detect and analyse PFAS in environmental samples, such as drinking water, soils, sediments, biota, and biosolids is expanding. Importantly, this includes adapting existing technologies or developing new technologies towards field deployable sensing to allow for real-time on-site monitoring of PFAS.

3.3. Real-time on-site monitoring

A cheap, compact, and effective way to detect low levels of PFAS in the field is desirable. Despite its cost, mass spectrometry is an attractive method of on-site PFAS detection. Iodide time-of-flight chemical ionization mass spectrometry (iodide-ToF-CIMS), in particular, has shown promise for the more challenging to detect gas and aerosol phase PFAS in indoor air,[95] or atmosphere in a semi-continuous online fashion.[96, 97] A significant benefit of this technique is that it avoids the need to perform the aforementioned cumbersome solid-phase extraction processes.

In addition to investigating portable solutions for instrumentation,[98] sensors are being developed for rapid PFAS identification, typically using optical or electrochemical detectors despite the fact that PFAS are not optically or electrochemically active.[85, 99]

Colourimetric strategies use nanoparticles, which show a change in surface properties[100-102] or redox dyes, which change intensity[103, 104] upon interacting with PFAS. Sensors based on fluorescence or luminescence use fluorescent dyes with molecularly imprinted polymers (MIPs) containing pockets designed to capture specific PFAS, appended to nanoparticles[105-108] or luminescent metal-organic frameworks (MOFs) containing a chromophore ligand.[109] When PFAS, in concentrations as low as µg/L, enter the cavities, the fluorescence or luminescence is quenched or modified.

Electrochemical methods use redox probes, such as ferrocene carboxylic acids, which see a change in electron transfer resistance when PFAS molecules enter the cavities of MIPs, such as poly(*o*-phenylenediamine).[110-114] A recent report of a sensor operating via PFAS interrupted energy transfer between fluorinated poly(*p*-phenylene ethynylene) and a fluorinated squaraine dye demonstrated PFAS detection in water with concentrations in the µg/L range.[115] Detection by electrochemiluminescence, achieved through electrochemically imprinting ultra-thin graphitic carbon nitride nanosheets, generated a sensitive sensor selective for PFOA.[116] It has been shown that switching to an impedimetric detection method by employing nanoporous MOF geometry with interdigitated microelectrodes dramatically increased sensitivity with detection limits in the ng/L range.[117] Photochemical sensors, comprised of BiOI nanoflake arrays on screen-printed electrodes containing grafted MIP in the form of a disposable sensing strip, have been developed to analyse µg/L concentrations of PFOSF in real water samples.[118]

In another study, researchers have leveraged the amphiphilic properties of PFAS to develop a water-based sensor capable of measuring surface and interfacial tensions. The tensions exhibit an increase in emission intensity, based on the differential behaviour at the interfaces of complex droplets, when PFAS surfactants in mg/L concentrations are present.[119]

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Despite being very promising, these techniques for detecting and analysing PFAS require more validation to demonstrate their applicability to real samples with low concentrations and matrix effects.

3.4 Opportunities for contributions from chemists

In order to address the current limitations in effectively detecting and analysing PFAS, and to remain current with the decreasing allowable concentrations of PFAS, particularly in drinking water, analytical instrumentation will continue to become smaller, cheaper, and more effective over time. As this occurs, there are significant opportunities for chemists to contribute to the improved detection (lower limits) and analysis of PFAS (managing the increasing library of PFAS and their derivatives). Examples of these include:

1. Increased understanding of PFAS exposure sources, including consideration of potential contamination from sampling, use of laboratory consumables, and instrumentation parts[120]
2. Development of the lab-scale PFAS detection methods into practical commercial-scale real-time on-site PFAS monitoring in all media with very low detection limits (from mg/L to µg/L to ng/L)
3. Improved analysis of environmentally induced PFAS by-products alongside an increased understanding of their thermal, oxidative, mechanical, and biological degradation mechanisms

4.0 Separation and Destruction Technologies

The major sites for PFAS point source contamination are PFAS manufacturing plants, consumer product manufacturing plants, firefighting foam discharge locations (e.g., military training areas), large-scale cleaning facilities, wastewater treatment facilities, waste disposal facilities, and landfills.[121] All of these result in the widespread deposition of PFAS in our soils and in our waterways, enabling them to enter the food chain.

Current methods used commercially to remediate PFAS include sorptive removal from aqueous solution, burying waste in lined landfills (with or without stabilisation), or incineration for contaminated spent sorbents, soils, and products.[122] Unfortunately, all these techniques have important limitations and they rarely apply across all PFAS. For instance, incineration suffers from high energy consumption, products of incomplete combustion, and cannot be used for liquid samples.[123-126] The destruction of PFAS is complicated by the strength of the C-F bond (which increases with an increase in number of fluorine bonds on C), the different types (i.e., variety of tail lengths, head groups) and concentrations, as well as where they exist in the environment (i.e., waters, solids, air).[127, 128]

This presents an opportunity to develop more cost-effective, selective, and efficient technologies. Ideally, a treatment train or tandem approach to managing PFAS in the environment can be employed to achieve both their separation from complex media and their complete destruction. The desired characteristics of an effective technology for managing PFAS include its ability to remove different types of PFAS, including long-chain, (ultra) short-chain, and more complex derivatives, and stage of development (maturity) as well as practicality across different media (e.g. water, solids) (Table 2). From this summary, it is clear that there is significant room for improvement (yellow and red) across all technologies.

4.1 Separation technologies

Separation of PFAS, from aqueous solution by immobilisation, or soil and sediments by mobilisation, involves the use of natural (e.g., clay, plants),[129, 130] semi-natural (e.g., activated carbon)[131-133] or synthetic (e.g., imprinted polymer)[134, 135] materials as sorbents or resins. These technologies rely on either adsorption, or absorption.

4.1.1. Ion exchange and adsorption

Ion exchange is a reversible separation process in which similarly charged ions in the liquid phase (e.g., PFAS) are exchanged between an immobile solid phase, resulting in no significant change to the solid phase.[136-138,139, 140] This interaction has been verified by varying the pH of the PFAS solution or by observing the release of charged equivalent ions from the solid phase, such as chloride ions.[140] During the ion exchange process for the removal of PFAS, the positively (or negatively) charged ion exchange site is attracted to the negatively (or positively) charged functional group(s) of the PFAS. Simultaneously, the hydrophobic backbone on the resin attracts the hydrophobic C-F tail of the PFAS.[139] Anion exchange resins (e.g., Dowex) are the most widely used ion exchange resins[141-146] because of the abundance of anionic PFAS. Ionic exchange resins capable of removing cationic and zwitterionic PFAS[36, 147] are also increasingly reported.[148, 149] Ion exchange is a cost-effective and efficient way to remove PFAS from water; however, this technology has been shown to be much less effective for short or ultra-short chain PFAS as well as complex matrices. It is most useful in the treatment of drinking water in a final 'polishing' step.

Adsorption is a reversible process whereby adsorbate molecules are transferred from a fluid bulk phase and stick to a solid surface phase.[43, 150] Adsorption of PFAS onto solid surfaces has been explored in literature as one of the effective methods of PFAS removal from contaminated media because of their easy design, cost-effectiveness and simple operation.[151] Popular adsorbent materials include granular activated carbon, biochar, aminated rice husk, clay, molecular imprinted polymer, and zeolite. Granular activated carbon, clay, and biochar materials were used in situ to immobilise short and long-chain PFAS in contaminated artificial soil samples.[133] One study reported that the addition of granular activated carbon to contaminated soils reduced leachability through chemical stabilisation between 55.8-99.9%. In addition, the leachability was further reduced to 87.1-99.9% by binding with cement.

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Foam fractionation is an adsorptive separation technique that functions without the use of solid adsorbents.[152] The use of the foam fractionation method in PFAS remediation is based on the surface-active properties of PFAS which facilitates the separation of PFAS from contaminated liquid samples by extraction using rising foam. The foam is collected and collapsed (known as foamate) to give a low-volume residual liquid waste stream.[153] In a typical setup for foam fractionation, gas bubbles are introduced into a liquid containing surface-active substances, where the surface-active substances attach to the gas-liquid interface of the bubbles to form foam. The foam is stabilised by decreasing the surface tension of the gas-liquid interface, thereby enabling the formed bubbles to create an emerging foam column above the liquid level. The foam column is collected and then collapsed mechanically or by reduced pressure.[154] Foam fractionation can be operated by batch or continuous modes.[154] Buckley et al.[155] explored foam fractionation to remove long-chain PFAS from contaminated water. The study found 10-90% removal efficiency for the PFHxS, PFOA, and PFOS studied. However, the method was unable to remove short-chain PFAS.

4.1.2. Membrane Filtration

Removal of PFAS from contaminated aqueous streams using membrane filtration has been widely studied.[156-158] Filtration can be electrical, pressure, or temperature gradient-driven, and involve the selective removal of solutes from the solvent using a semipermeable or porous membrane.[159] Nanofiltration and reverse osmosis are two high-pressure membrane filtration processes that have been widely studied for removal of PFAS from wastewater at bench and pilot scales.[159] In specific examples, a 2D material MXene-polyanide thin-film nanocomposite hollow fibre made of Mxene nanosheets was used in a bench study for nanofiltration of PFOS from water.[160] Mxene are transition metal carbides, carbonitrides or nitrides represented as $M_{n+1}X_nT_x$, (M = Ti, V, Cr, etc.; X = C, N, T_x = O, F, Cl, H, n = 1-3).[161, 162] The results found up to 96% removal of PFOS from the contaminated water compared to 72% removal for thin-film composite material without Mxene nanosheets.[160] In a reverse osmosis study,[163] three commercial aromatic polyamide membranes were used to remove perfluorohexanoic acid (PFHxA) from contaminated wastewater. It was found that two of the three studied membranes achieved removal efficiency of 99%, while one of the membranes attained suitable water permeability.

4.1.3. Phytoremediation

Another technology, useful in mobilising contaminants in soil, air, and water is phytoremediation.[164] In phytoremediation (as the name implies phyto- = of a plant; relating to plants), living plants uptake PFAS *in situ* from contaminated sites.[164] Studies have shown that phytoremediation is a good candidate for removing PFAS from contaminated media, although it is a slow process.[165-168]

In a study using three native Australian plant species, the study suggested that the plants have potential to reduce PFAS concentration in stormwater, with mean overall percentage removal efficiency of about 53%.[169] In general, the report found that less PFOA than PFOS accumulated in the roots, while more PFOA than PFOS accumulated in the shoots.

Table 2. A summary of the performance of common and emerging separation and destruction methods for PFAS (classified into **Acceptable** technologies, methods showing **Some Promise** and methods that are considered **Unacceptable** or for which the feasibility is **Unknown**)

Methods	Media	Efficiency for Long-Chain PFAS	Efficiency for (Ultra) Short-Chain PFAS	Maturity	Practicality
Separation					
Ion Exchange Resins[136-138,139, 140]	Liquids (water)				
Adsorption by AC[133,151]	Liquids (water)				
Adsorption by Polymers[123,124]	Liquids (water)				
Filtration[156-158,159]	Liquids (water)				
Foam Fractionation[155]	Liquids, leachate (water)				
Phytoremediation[165-168]	Liquids and Solids				
Destruction					
Incineration[122]	Solids (soil, resin)				
Electrochemical Oxidation[170-172]	Liquids (water)				
Photochemical Oxidation[173]	Liquids (water)				
Sonolysis[174]	Liquid (water)				
Supercritical Water Oxidation[175]	Liquids (water)				
Advanced Reductive Processes[176]	Liquids (water)				
Hydrothermal Alkaline Treatment[177-179]	Liquids (water)				
Mechanochemical Degradation[180-167]	Solids (soil, sludge)				
Bioremediation[181, 182,183, 184,185,186]	Liquids, soil, sediment				
Non-Thermal Plasma[187, 188]	Liquids (water)				
Smouldering[189-191]	Solids (soil, sludge)				

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4.2. Destruction technologies

While incineration is industrially viable and widely used to degrade PFAS on solids, it results in the formation of toxic HF gas and other gaseous fluorocarbons.[41, 43, 180] In addition, incineration requires high temperatures (> 1000°C), enough to break the hydrophobic C-F bonds in PFAS, and is an energy intensive process.[35, 43] To this end, the US-EPA challenged researchers to discover new technologies that can remove >99% PFAS in unused AFFF with less harmful by-products than incineration.[192] Several PFAS destruction technologies have since emerged (Table 2).

4.2.1. Oxidative processes

The most promising oxidative processes include, electrochemical, photochemical, and sonochemical oxidation. Reactive free radical oxidising agents such as hydroxyl and chlorine radicals ($HO^{\bullet}$ and $Cl^{\bullet}$) are used to break down the strong C-F bonds of PFAS, degrading them into harmless by-products.[24]

In an electrochemical oxidation process, a potential difference or an electric current is applied between an anode and a cathode to generate reactive oxidising radicals at the electrode surface, which are used to degrade pollutants.[171] Electrochemical oxidation processes can occur through direct and indirect oxidation mechanisms. In the indirect oxidation mechanism, reactive oxidising radicals are electrochemically generated *in situ* at the electrode, which are used to degrade PFAS. However, in the direct oxidation mechanism, PFAS are degraded at the anode through a direct electron transfer reaction between the PFAS and the anode surface.[170-172]

During photochemical oxidation reactions, PFAS are degraded as a result of their interactions with excited state oxidising species (e.g., $HO^{\bullet}$, $CO_3^{\bullet}$, $H^{\bullet}$) in the presence of light.[193, 194] Efficient photodegradation of PFAS can occur at UV light wavelengths <220 nm (in the vacuum UV range). However, at longer wavelengths (>220 nm), PFAS does not absorb UV light,[194] thereby direct photodegradation of PFAS becomes ineffective.[195] Liu et al. reported 93-100% total defluorination of perfluorinated and fluorotelomer carboxylates and sulfonates through integrated redox photochemical processes[173] as well as near complete defluorination and mineralisation of most PFAS in AFFF.[196] The oxidation process was through hydroxyl radical treatment, while the reduction process was through UV-sulfite treatment.

In sonochemistry, ultrasound radiation in the range of 20-1000 kHz is applied to molecules to achieve chemical reactions through a physical phenomenon called acoustic cavitation. This process creates localised spots for chemical reactions, which can reach pressures and temperatures of 500 atm and 5000 K, respectively.[197] Reactive radical species such as $HO^{\bullet}$ and $Cl^{\bullet}$ can be generated in the hot spots (e.g., from the decomposition of H_2O to yield $HO^{\bullet}$ and $H^{\bullet}$)[194, 197] and are able to degrade pollutants, including PFAS. Sonochemical reactions can occur as quickly as a fraction of a second.[198] A study of low frequency (20 kHz) sonochemical degradation of PFOS and PFOA in water showed >90% degradation efficiency at 20°C under acidic conditions.[174]

Supercritical water oxidation is regarded as an advancement in wet air oxidation,[199] and is postulated to mineralise organic waste samples.[200] This method, based on chemical oxidation in supercritical H_2O (>374°C and 22.11 Mpa) uses H_2O_2, air, or O_2 as the oxidising agent.[201] In a typical supercritical water oxidation process, H_2O and CO_2 are formed from the oxidation of organic compounds, while heteroatoms such as F, S, P and Cl react to form HF, H_2SO_4, H_3PO_4 and HCl, respectively.[199] The supercritical water oxidation technology was applied to successfully achieve 99.99% destruction of PFAS in aqueous matrices, and its efficiency was not hampered by co-contaminants such as volatile organic compounds and petroleum hydrocarbons.[175]

4.2.2. Reductive processes

In contrast to advanced oxidation processes (AOPs) which use highly reactive species such as $HO^{\bullet}$ and $SO_4^{\bullet-}$, advanced reductive processes (ARP) typically employ hydrated electrons (e_{aq}^-),[202] hydrogen atoms ($H^{\bullet}$) and other species (e.g., $SO_3^{\bullet-}$ and $SO_2^{\bullet-}$ depending on activation method and solute) to cleave the resistant C-F bonds for PFAS degradation.[203] The degradation of PFAS is dependent on the type of reductive species and the chemical structure of PFAS.[204] The concentration of highly reactive and reductive e_{aq}^- (E = −2.9 V) must be maximized in an ARP treatment process to break the resistant C-F bonds. Depending on the activation methods and chemical agents to form reactive radicals, ARP systems can be of different types. Parameters such as F index, overall defluorination ratio and molecular defluorination ratio are used to quantify the extent of defluorination.

The degradation of PFOA using photoionization in a UV/sulfite system (254 nm) was found to be dependent on the pH of the initial solution in generating e_{aq}^- species. In acidic media, e_{aq}^- are quenched by reacting with H^+ thereby suppressing the decomposition of PFOA in water while under alkaline conditions

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the $H^{\bullet}$ and HO^- react to regenerate the e_{aq}^- which enhances the defluorination of PFOA (Scheme 2).[205] Additionally, the distribution of sulfite species (e.g., SO_3^{2-} and HSO_3^-) varies with pH. Hydrogen sulfite, HSO_3^-, dominates at pH values lower than its pKa (ca. 7) and at pH 9.2 the concentration of sulfite, SO_3^{2-}, is at its maximum (99%) efficiently able to reduce PFOA. Sulfite is more photoactive, hence able to generate more e_{aq}^-, as compared to hydrogen sulfite due to its more significant spectral overlap at wavelengths typical of UV irradiation sources.[206] Overall, basic conditions are favored in UV/sulfite system.

$$H^+ + e_{aq}^- \longrightarrow H^{\bullet}$$

$$H^{\bullet} + HO^- \longrightarrow H_2O + e_{aq}^-$$

Scheme 2. pH dependence on the regeneration of e_{aq}^- in UV/sulfite system

Typical degradation mechanisms include shortening of the chain and H/F exchange. Also, degradation efficiency is affected in the presence of anions which quench e_{aq}^-. Production of e_{aq}^- can be improved by coupling the ARP process with chemical agents. For example, in high energy vacuum ultraviolet light systems (VUV) at 185 nm, coupling with iron(III) in acid aqueous solution increased the defluorination rate for PFOA by the formation of a complex between ferric ions and PFOA.[207] It is important to note that e_{aq}^- can also be generated by other sources to decompose PFAS, for example iodide,[208] ferrocyanide,[209] dithionite and indole derivatives[210, 211] and the effect of pH depends on the specific system and reductive species. Recently, electrochemical reductive treatment[212] has also appeared as a feasible alternative due to ease of operation and milder reaction conditions.

Though ARP has emerged as a promising option as oxidation processes; more understanding of their degradation pathways and mechanisms, types of materials/catalysts, effectiveness of degradation, pH adjustment and implication of solution chemistry is needed. For realistic applications, the influence of effluent total dissolved solids (TDS) needs to be considered because the formation of inert salt residuals, during ARP would increase TDS levels which makes it impractical for drinking water treatment or municipal wastewater reuse (but okay for other PFAS contaminated water applications).[176] The concentration of dissolved oxygen in water also needs to be controlled, which is challenging for real-world scenarios.[203]

4.2.3. Other processes

Another PFAS destruction method developed recently that has the potential to remediate PFAS is low-temperature decarboxylation. The method, which reported up to nearly 100% defluorination of PFAS within 24 h, used a DMSO/NaOH solvent mixture at 120°C.[213] Although the method reported the low-temperature mineralisation of PFAS using readily available reagents and produced less toxic by-products, and could be adapted for small-scale or lab-scale destruction, the applicability of using large volume of DMSO for a pilot plant is a major drawback.

Hydrothermal alkaline treatment thermochemically degrades pollutants by leveraging the alkaline properties of H_2O (in the presence of NaOH) along with high pressures (2-22 Mpa) and temperatures (170-350°C).[214, 215] Originally used in the degradation of halogenated waste, HALT has been recently applied to degrade PFAS, with more than 90% reported efficiency.[178, 216] Proposed mechanisms for hydrothermal alkaline treatment include the nucleophilic substitution of the charged head (e.g., SO_3^-),[179] or of F^- from the C-F tail of PFAS with HO^- from the alkaline solution to form a less thermodynamically stable C-OH bond.[178] These unstable intermediates undergo hydrolysis, decarboxylation or cleavage to release F^- as HF or fluoride salts until the PFAS is completely degraded.[177-179]

Mechanochemical degradation involving high-energy ball milling has been shown to produce sufficient energy to achieve the degradation of PFAS.[152] Although partial degradation of halogenated compounds occurred in previous studies using this technique,[217] the introduction of tribomaterials or co-milling agents (e.g., CaO, NaOH, SiO_2, KOH, La_2O_3) has improved the degradation efficiency significantly. The co-milling agents generate triboplasma that emit high energy particles at the sites where intramolecular bonds are broken,[152, 217] thereby facilitating the degradation of PFAS into safe inorganic salts (e.g., KF, K_2CO_3 and K_2SO_4, R_3SiF).[180] Mechanochemical degradation is able to mineralise halogenated pollutants, such as PFAS, in short reaction times.[218] For example, MCD of 6:2 fluorotelomer sulfonate (6:2 FTS) with KOH reported nearly 100% mineralisation in less than 1 h.[219] Furthermore, it is possible for real-world samples of contaminated soil, to directly undergo destruction using this method.[220]

Bioremediation is another potential way to degrade PFAS, particularly with partially fluorinated substances.[221] However, as with other reductive dehalogenation routes, bacterial defluorination depends heavily on the bacterial strain and, generally does not completely degrade PFAS.[181, 182] The transformation products are typically shorter-chain derivatives. The process is quite slow compared to other routes (e.g. requires days).[183, 184] Recent developments in this area have shown significant improvements, for example, using a biomimetic multifunctional lignocellulosic nano-framework to concentrate the PFAS prior to fungal bioremediation yielding shorter chain derivatives;[185] and the use of a dual biocatalysed microbial electrosynthesis system resulting in 91% biodegradation of PFOA.[186]

Plasma is a state of matter with charged gaseous molecules able to degrade PFAS. The ionization can be induced by adding energy which leads to the formation of highly reactive species. Depending on the energy of the electrons compared to the temperature of the background gas, plasma can be categorized as either thermal (same temperature) or non-thermal (NTP, cold plasma, lower gas temperature).[122] NTP is preferable due to its lower energy. The effectiveness of the degradation of PFAS by NTP can be correlated with the length of the perfluorinated carbon backbone. Kavanagh *et al.* investigated

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the NTP degradation of various PFAS in aqueous solution and observed a more effective degradation of long chain PFAS compared to shorter PFBA and PFBS.[187] This is due to lower accumulation of hydrophilic SC-PFAS at the liquid-plasma interface thereby limiting their degradation. To increase the surface activity of SC-PFAS, Thagard *et al.* used a cationic surfactant, hexadecyltrimethylammonium bromide (CTAB) to destroy the recalcitrant PFBS in contaminated water.[188] The PFBS-CTAB complex increased the PFBS mass transport to the interface via argon bubbles thereby increasing its degradation. Overall, plasma treatment is an effective technology with short treatment times and has minimal impact from the presence of organic or inorganic contaminants. Further research is required with regards to PFAS mineralisation pathways, observed formation of SC-PFAS, and acidification of treated water.

Smouldering is a thermal degradation technique that is flameless and occurs on the surface of condensed fuel. It is self-sustained once ignited and is more energy and cost efficient compared to incineration, which requires continuous energy input.[189] Conventionally, contaminants are the fuel for smouldering combustion of hydrocarbon contaminated soils, but as PFAS cannot support smouldering on its own, some surrogate fuels (like carbon particles) are needed in small concentrations. Gerhard *et al.* mixed GAC (fresh or PFAS-loaded) with PFAS contaminated soil to support sufficient smouldering temperatures that could destroy PFAS.[190] This method could be promising to capture emitted or transformed PFAS on the sorbent. The same researchers also used the smouldering technique to treat PFAS in sewage sludge by adding CaO (5-10 mg/kg of dried sludge), which served the dual purpose of mineralising the fluorine, as well as minimising hazardous PFAS and HF emissions.[191] Though smouldering combustion is a promising and energy efficient thermal decomposition technique; the amount of surrogate fuel needed, completeness of PFAS combustion, removal of decomposed PFAS and the careful management of the amount of HF emissions need to be further evaluated. In addition, the solid spent sorbent is destroyed in the process of smouldering combustion, preventing it from reuse.

4.3 Opportunities for contributions from chemists

The complete destruction of PFAS is incredibly challenging, due to a wide variety of samples containing different concentrations, types of PFAS, and contaminated media to manage. Some techniques are practical, yet not well developed, while others look promising for long chain PFAS but have not shown efficacy for short chain PFAS. It is too early to tell with some of the emerging techniques whether or not they will compete with the current state of the art (i.e., adsorption plus incineration or landfilling). There is not yet one technique that checks all of the boxes. Ways that chemists can support the existing destruction and removal of PFAS in products and the environment are:

1. Develop robust protocols to investigate and measure adsorption capacity of both control samples (e.g., deionized water) and realistic samples (e.g., drinking water, waste water) using realistic concentrations of PFAS
2. Develop selective, efficient, and reusable separation strategies, particularly for hydrophilic short- and ultra-short chain PFAS from liquid media that can be integrated into existing processes.
3. Find or develop new solutions for the complete destruction of multiple C-F bonds across all media (e.g., soil, sediment, leachate, water, air) in the environment as well as spent products.
4. Improved analysis of thermal- or chemical-induced PFAS destruction by-products alongside an increased understanding of the mechanisms involved.
5. Develop on-site solutions to manage existing and future waste, including contaminated equipment, soil, waters, and PFAS-containing products.

5.0 The Need for Alternatives to PFAS

PFAS have been detected in environmental media with confirmed pernicious effects on human health.[5, 66] The risks associated with PFAS, alongside government regulations, have stimulated the development of alternative substances with comparable chemical and physical properties. Initially, the historic long-chain PFAS (e.g., PFOA, PFOS) were replaced by shorter chain PFAS (e.g., GenX, PFHxS). This strategy is problematic as the shorter-chain alternatives are highly mobile, persistent, difficult to remediate[222] and there is no evidence that they are less toxic.[223] Substituting PFAS with alternatives without a proper assessment is deleterious for health, ecology and the economy and results in the alternatives being regrettable.[224-228] A different strategy is a benign-by-design approach, where alternatives that are safer, sustainable and which can perform the necessary functions to create lasting solutions are sought.[22]

5.1 PFAS-free certification

To promote the use of sustainable alternatives, GreenScreen has certified PFAS-free products based on hazard endpoints to help consumers in minimising their PFAS footprint by making healthier and informed choices. Another approach in the global phaseout of PFAS is grouping them into non-essential, substitutable, and essential types based on societal needs and the availability of alternatives.[26, 27] In contrast to non-essential (e.g., ski wax) and substitutable categories (e.g., aqueous fire-fighting foams), certain applications of PFAS are specialised and essential (e.g., occupational protective clothing) and will require resources for future innovation and product development as no established alternatives currently exist.[27]

To this end, some manufacturers of consumer products are aiming to avoid intentionally adding PFAS to their products, where possible. Progress has been made in developing non-fluorinated alternatives for PFAS in various categories. These alternatives typically target specific properties which are

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desired for an application, such as water-repellency, surfactant structure, or thermal stability (Figure 5). For example, melamine and its derivatives are nonfluorinated chemicals used as textile additives to achieve water-repellency. In addition, commercial PFAS-free AFFFs have been developed, such as ENVIRO 2-3% FFF and NFD 20-391.[227] Biobased materials are also attractive choices as alternatives, such as starch and zein, which have been employed in food packaging as degradable water- and oil-repellent paper packaging products.[229]

5.2 Safer alternatives across sectors

5.2.1 Firefighting

Arguably, aqueous fire-fighting foams are one of the largest contributors to environmental pollution due to PFAS. In part, this is because effective fire suppression to minimise damage to life and property is a high priority. However, fire suppression needs to be balanced with its environmental impact and the health of the firefighters. Numerous Class B Fluorine-free PFAS replacement foams are being used for aviation, in the military, as well as in oil and gas companies. These alternatives are chemically grouped into hydrocarbons, detergents, siloxanes, and proteins.[230] However, transitioning between foam types requires review, modification, redesign, and rinsing of storage, discharge, and application systems to avoid incompatibility and cross-contamination with the new formulations.[227, 228, 231] Moreover it is essential to optimise the performance of fluorine-free foams (F3) and assess their potential human health, safety, and environmental risks, when replacing PFAS-containing foams with F3. Lastly, transitioning from PFAS to safer F3 is proposed to decrease the impact on the environment but currently requires higher application rates and densities to achieve similar results as PFAS-containing AFFFs.[232]

5.2.2 Construction

Safer alternatives to PFAS exist in the construction sector. For example, roofing solutions include silicone-modified polyester, acrylic or PVC-coated membranes. Siloxanes, silicone polymers, paraffins and polyurethanes have been employed to replace PFAS in flooring and or in glass coatings.[233] However, for coatings, paints, and varnishes, PFAS function as anti-blocking, anti-soiling, oil-repellent, and UV-cooling additives, and no suitable alternatives have been identified that contain all of these properties. Although polyolefin- and polyurethane-based alternatives are suitable and cost-effective, they do not meet the high-performance requirements for most coating applications. Additionally, glass and polyester materials do not perform well in comparison to fluoropolymers for solar panels. Overall, further research and development are required to improve PFAS alternatives in coatings, paints, and varnishes. In the metal plating and finishing industry, mechanical controls and viable non-fluorinated surfactants are commercially available to suppress chromium-bearing mist emissions, but metal plating facilities continue to be a source of PFAS pollution.

5.2.3 Textiles

In the textile sector, non-fluorinated alternatives based on silicones, hydrocarbons, and other chemicals meet the requirements for water-repellency for most outdoor gear and fabrics. For example, silicone based on a polydimethylsiloxane (PDMS) backbone provides a soft feel and has moderate durability to laundering. However, these alternatives have potential health and ecotoxicity hazards due to residual levels of cyclic volatile methyl siloxane (D4 and D5).[30] Hydrocarbon-based durable water repellents e.g., crystallised linear n-alkyl chains, paraffin waxes, acrylic copolymers have promising biodegradation over time, but they suffer from poor durability to dry cleaning. Other dendrimer (hyperbranched polymeric structures consisting of ester or polyurethane segments) and inorganic nanoparticle chemistries (manufactured using SiO_2 or Al_2O_3 materials) are degradable in principle but have not been well studied. More importantly, the non-fluorinated DWR does not provide sufficient repellence to liquids of varying polarities required for occupational protective clothing, which is an essential use.[30, 234]

5.2.4 Other sectors

PFAS are used in personal care products including cosmetics and can be found in lipsticks, foundations, and waterproof mascaras.[235-237] They are incorporated specifically to improve the product texture and ease of application, making them long lasting and water resistant. Recently, some major cosmetic manufacturers have committed to phasing out PFAS in their products; however, there is limited information available on their alternatives (if any).

For cookware, several non-stick alternatives are available, including ceramic, cast iron, and stainless-steel options that does not contain PFAS-based coatings which have been linked to harmful health effects.

There are some alternatives available for PFAS use in the electronics sector;[238] however, PFAS still have several specialised uses in the production of semiconductors, fuel cells, lithium-ion batteries, high-speed telecommunication, ICT and acoustic equipment, high voltage cable insulation and wiring. Further research and development are needed to innovate viable alternatives for these essential uses.[239]

Due to their effective hydrophobic properties, PFAS are used in drilling and fracking liquids to enhance the flow of oil and gas from underground geological formations.[240] A number of alternatives are available, including non-fluorinated silicone/siloxane-based anti-foaming agents.

5.3 Opportunities for contributions from chemists

Currently, there is a growing awareness of the environmental and health risks associated with PFAS, which has led to the ongoing regulations to limit their non-essential use. Concurrently, collaborative research efforts are being made to develop methods for destruction and remediation as well as

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identifying safer PFAS alternatives. A combination of i) changes in consumer behaviours, ii) action by manufacturers, iii) legislation, and iv) litigation will see an increase in uptake of alternatives to PFAS. However, substituting PFAS needs to be done responsibly by switching to safer alternatives after a thorough evaluation of their hazard profiles, sustainability, performance, and practicality. Specifically, chemists are challenged to work with collaborators to:

1. Perform life-cycle analyses on PFAS, products containing intentionally added PFAS, and alternatives to PFAS.
2. Develop new molecules and materials to be used in as alternatives to PFAS that can be transitioned in an efficient and cost-effective manner.
3. Perform assessment of the key material properties (*e.g.*, fire-, water-, oil-, grease-, chemical- and thermal-resistance) for new and existing alternatives to ensure they meet performance standards set by PFAS-containing products.
4. Determine the environmental fate and effect of new and existing alternatives to support robust risk assessments prior to placement on the market.

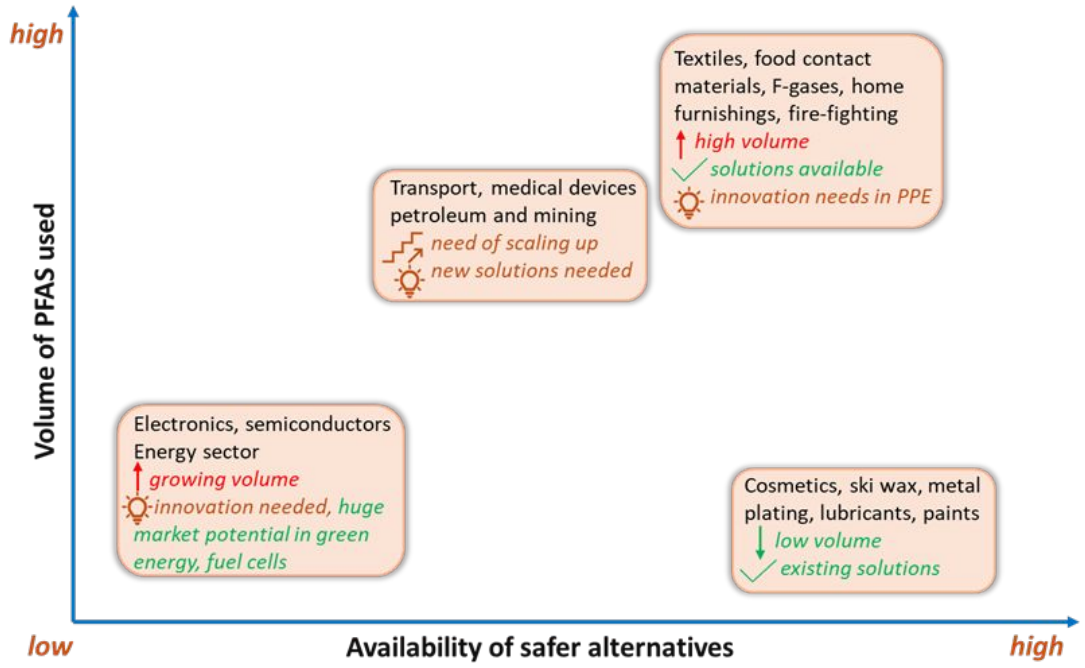

Figure 5. The volume of PFAS used across sectors and the availability of suitable alternatives. Original source credit: ChemSec, 2024.[241] *Note: there are uncertainties in volume use and available alternatives in the fluoropolymers, pharmaceuticals, and medical sectors. PPE = personal protective equipment.*

6.0 Summary and Future Outlook

The exceptional properties of PFAS, combined with the challenges of finding viable alternatives, mean they will continue to be used in numerous essential applications. The unique properties of PFAS also prevent their degradation, which, in light of their potential for bioaccumulation and toxicity, has resulted in a dramatic shift in policy around PFAS production and usage. Importantly, the US EPA has recently finalised a rule (effective since November 2023) requiring manufacturers to report and keep records of PFAS under the Toxic Substances Control Act.[242] This mandates includes disclosing usage, production volumes, disposal, exposure, and hazards for PFAS manufacturing as well as the manufacture of PFAS-containing articles. Thus, there is an urgent need to develop novel methods for the effective management of PFAS, using best practice strategies,[120] and to design safe and suitable alternatives. This multifaceted problem requires contributions from multiple areas of chemistry and provides an opportunity for chemists to work alongside engineers, epidemiologists, social scientists, and policymakers to develop sustainable solutions to this global issue.

Author Contributions

E.J.I. – Conceptualization, Investigation, Writing- original draft. S.D. – Investigation, Visualization, Writing – original draft. J.C. - Conceptualization, Validation, Writing- review & editing, Supervision, Funding acquisition. M.K. - Conceptualization, Validation, Writing- review & editing, Supervision, Funding acquisition. L.P.P. - Conceptualization, Validation, Writing- review & editing, Supervision, Funding acquisition. E. M. L. – Conceptualization, Visualization, Validation, Writing- review & editing, Supervision, Funding acquisition.

Conflicts of interest

There are no conflicts to declare.

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References

1. Z. Wang, A. M. Buser, I. T. Cousins, S. Demattio, W. Drost, O. Johansson, K. Ohno, G. Patlewicz, A. M. Richard, G. W. Walker, G. S. White and E. Leinala, *Environ. Sci. Technol.*, 2021, **55**, 15575-15578.
2. B. Langenbach and M. Wilson, *Int. J. Env. Res. Public Health*, 2021, **18**, 11142.
3. R. C. Buck, J. Franklin, U. Berger, J. M. Conder, I. T. Cousins, P. de Voogt, A. A. Jensen, K. Kannan, S. A. Mabury and S. P. van Leeuwen, *Integr. Environ. Assess. Manage.*, 2011, **7**, 513-541.
4. G. T. Ankley, P. Cureton, R. A. Hoke, M. Houde, A. Kumar, J. Kurias, R. Lanno, C. McCarthy, J. Newsted, C. J. Salice, B. E. Sample, M. S. Sepúlveda, J. Steevens and S. Valsecchi, *Environ. Toxicol. Chem.*, 2021, **40**, 564-605.
5. E. Panieri, K. Baralic, D. Djukic-Cosic, A. Buha Djordjevic and L. Saso, *Toxics*, 2022, **10**, 44.
6. A. F. Ojo, C. Peng and J. C. Ng, *J. Hazard. Mater.*, 2021, **407**, 124863.
7. R. A. Dickman and D. S. Aga, *J. Hazard. Mater.*, 2022, **436**, 129120.
8. R. E. Banks and J. C. Tatlow, *J. Fluorine Chem.*, 1986, **33**, 71-108.
9. H.-J. Lehmler, *Chemosphere*, 2005, **58**, 1471-1496.
10. N. V. Ignat'ev, A. Kucherina and P. Sartori, *Acta Chem. Scand.*, 1999, **53**, 1110-1116.
11. Y. W. Alsmeyer, W. V. Childs, R. M. Flynn, G. G. I. Moore and J. C. Smeltzer, in *Organofluorine Chemistry: Principles and Commercial Applications*, eds. R. E. Banks, B. E. Smart and J. C. Tatlow, Springer US, Boston, MA, 1994, DOI: 10.1007/978-1-4899-1202-2_5, pp. 121-143.
12. E. Karbassiyazdi, M. Kasula, S. Modak, J. Pala, M. Kalantari, A. Altaee, M. R. Esfahani and A. Razmjou, *Chemosphere*, 2023, **311**, 136933.
13. A. G. Paul, K. C. Jones and A. J. Sweetman, *Environ. Sci. Technol.*, 2009, **43**, 386-392.
14. K. Prevedouros, I. T. Cousins, R. C. Buck and S. H. Korzeniowski, *Environ. Sci. Technol.*, 2006, **40**, 32-44.
15. G. Kostov and B. Ameduri, in *Modern Synthesis Processes and Reactivity of Fluorinated Compounds*, eds. H. Groult, F. R. Leroux and A. Tressaud, Elsevier, 2017, DOI: https://doi.org/10.1016/B978-0-12-803740-9.00018-4, pp. 533-559.
16. I. J. Neuwald, D. Hübner, H. L. Wiegand, V. Valkov, U. Borchers, K. Nödler, M. Scheurer, S. E. Hale, H. P. H. Arp and D. Zahn, *Environ. Sci. Technol.*, 2022, **56**, 6380-6390.
17. M. K. Björnsdotter, L. W. Y. Yeung, A. Kärrman and I. E. Jogsten, *Environ. Sci. Technol.*, 2019, **53**, 11093-11101.
18. M. Ateia, M. Arifuzzaman, S. Pellizzeri, M. F. Attia, N. Tharayil, J. N. Anker and T. Karanfil, *Water Res.*, 2019, **163**, 114874.
19. F. Li, J. Duan, S. Tian, H. Ji, Y. Zhu, Z. Wei and D. Zhao, *Chem. Eng. J.*, 2020, **380**, 122506.
20. A. O. De Silva, C. Spencer, B. F. Scott, S. Backus and D. C. G. Muir, *Environ. Sci. Technol.*, 2011, **45**, 8060-8066.
21. D. Szabo, D. Moodie, M. P. Green, R. A. Mulder and B. O. Clarke, *Environ. Sci. Technol.*, 2022, **56**, 8231-8244.
22. Z. Wang, J. C. DeWitt, C. P. Higgins and I. T. Cousins, *Environ. Sci. Technol.*, 2017, **51**, 2508-2518.
23. L. P. Wackett, *mSphere*, 2021, **6**, e00721-00721.
24. M. Trojanowicz, A. Bojanowska-Czajka, I. Bartosiewicz and K. Kulisa, *Chem. Eng. J.*, 2018, **336**, 170-199.
25. R. W. Helmer, D. M. Reeves and D. P. Cassidy, *Water Res.*, 2022, **210**, 117983.
26. I. T. Cousins, J. C. De Witt, J. Glüge, G. Goldenman, D. Herzke, R. Lohmann, M. Miller, C. A. Ng, S. Patton, M. Scheringer, X. Trier and Z. Wang, *Environ. Sci. Processes Impacts*, 2021, **23**, 1079-1087.
27. I. T. Cousins, G. Goldenman, D. Herzke, R. Lohmann, M. Miller, C. A. Ng, S. Patton, M. Scheringer, X. Trier, L. Vierke, Z. Wang and J. C. DeWitt, *Environ. Sci. Processes Impacts*, 2019, **21**, 1803-1815.
28. L. G. T. Gaines, *Am. J. Ind. Med.* 2023, **66**, 353-378.
29. M. Sajid and M. Ilyas, *Environ. Sci. Pollut. Res.*, 2017, **24**, 23436-23440.
30. H. Holmquist, S. Schellenberger, I. van der Veen, G. M. Peters, P. E. G. Leonards and I. T. Cousins, *Environ. Int.*, 2016, **91**, 251-264.
31. Y. Fujii, K. H. Harada and A. Koizumi, *Chemosphere*, 2013, **93**, 538-544.
32. L. Schultes, R. Vestergren, K. Volkova, E. Westberg, T. Jacobson and J. P. Benskin, *Environ. Sci. Processes Impacts*, 2018, **20**, 1680-1690.
33. J. Glüge, M. Scheringer, I. T. Cousins, J. C. DeWitt, G. Goldenman, D. Herzke, R. Lohmann, C. A. Ng, X. Trier and Z. Wang, *Environ. Sci. Processes Impacts*, 2020, **22**, 2345-2373.
34. M. Gilchrist, 2022. Minnesoa Pollution Control Agency. PFAS in the metal plating and finishing industry. https://www.pca.state.mn.us/sites/default/files/gp3-05.pdf
35. J. Wang, Z. Lin, X. He, M. Song, P. Westerhoff, K. Doudrick and D. Hanigan, *Environ. Sci. Technol.*, 2022, **56**, 5355-5370.
36. W. J. Backe, T. C. Day and J. A. Field, *Environ. Sci. Technol.*, 2013, **47**, 5226-5234.
37. H. T. Nguyen, M. S. McLachlan, B. Tscharke, P. Thai, J. Braeunig, S. Kaserzon, J. W. O'Brien and J. F. Mueller, *Chemosphere*, 2022, **293**, 133657.
38. P. Hopewell and K. Darvodelsky, 2017. Assessment of Emergent Contaminants in Biosolids. https://www.biosolids.com.au/wp-content/uploads/Emerging-Contaminants-in-Biosolids-Research-report.pdf
39. N. Bolan, B. Sarkar, M. Vithanage, G. Singh, D. C. W. Tsang, R. Mukhopadhyay, K. Ramadass, A. Vinu, Y. Sun, S. Ramanayaka, S. A. Hoang, Y. Yan, Y. Li, J. Rinklebe, H. Li and M. B. Kirkham, *Environ. Int.*, 2021, **155**, 106600.
40. J. R. Lang, B. M. Allred, J. A. Field, J. W. Levis and M. A. Barlaz, *Environ. Sci. Technol.*, 2017, **51**, 2197-2205.

View Article Online DOI: 10.1039/D4SU00410D

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DOI: 10.1039/D4SU00272D

41. E. W. Tow, M. S. Ersan, S. Kum, T. Lee, T. F. Speth, C. Owen, C. Bellona, M. N. Nadagouda, A. M. Mikelonis, P. Westerhoff, C. Mysore, V. S. Frenkel, V. deSilva, W. S. Walker, A. K. Safulko and D. A. Ladner, *AWWA Water Sci.*, 2021, **3**, e1233.
42. E. Hepburn, C. Madden, D. Szabo, T. L. Coggan, B. Clarke and M. Currell, *Environ. Pollut.*, 2019, **248**, 101-113.
43. M. A. Uriakhil, T. Sidnell, A. De Castro Fernández, J. Lee, I. Ross and M. Bussemaker, *Chemosphere*, 2021, **282**, 131025.
44. L. J. Winchell, J. J. Ross, M. J. M. Wells, X. Fonoll, J. W. Norton Jr and K. Y. Bell, *Water Environ. Res.*, 2021, **93**, 826-843.
45. Per- and Polyfluoroalkyl Substances (PFAS): Incineration to Manage PFAS Waste Streams, 2019. https://www.epa.gov/sites/default/files/2019-09/documents/technical_brief_pfas_incineration_ioaa_approved_final_july_2019.pdf
46. K. V. Martin, T. J. Hilbert, M. Reilly, W. J. Christian, A. Hoover, K. G. Pennell, Q. Ding and E. N. Haynes, *Environ. Sci. Pollut. Res.*, 2023, **30**, 80643-80654.
47. S. Liu, S. Zhao, Z. Liang, F. Wang, F. Sun and D. Chen, *Sci. Total Environ.*, 2021, **795**, 148468.
48. S. Tartu, J. Aars, M. Andersen, A. Polder, S. Bourgeon, B. Merkel, A. D. Lowther, J. Bytingsvik, J. M. Welker, A. E. Derocher, B. M. Jenssen and H. Routti, *Environ. Sci. Technol.*, 2018, **52**, 3211-3221.
49. K. R. Miner, H. Clifford, T. Taruscio, M. Potocki, G. Solomon, M. Ritari, I. E. Napper, A. P. Gajurel and P. A. Mayewski, *Sci. Total Environ.*, 2021, **759**, 144421.
50. W. F. Hartz, M. K. Björnsdotter, L. W. Y. Yeung, A. Hodson, E. R. Thomas, J. D. Humby, C. Day, I. E. Jogsten, A. Kärrman and R. Kallenborn, *Sci. Total Environ.*, 2023, **871**, 161830.
51. D. Q. Andrews, T. Stoiber, A. M. Temkin and O. V. Naidenko, *Sci. Total Environ.*, 2023, **901**, 165939.
52. M. S. Shimizu, R. Mott, A. Potter, J. Zhou, K. Baumann, J. D. Surratt, B. Turpin, G. B. Avery, J. Harfmann, R. J. Kieber, R. N. Mead, S. A. Skrabal and J. D. Willey, *Environ. Sci. Technol. Lett.*, 2021, **8**, 366-372.
53. N. Yu, H. Guo, J. Yang, L. Jin, X. Wang, W. Shi, X. Zhang, H. Yu and S. Wei, *Environ. Sci. Technol.*, 2018, **52**, 8205-8214.
54. M. Chen, C. Wang, X. Wang, J. Fu, P. Gong, J. Yan, Z. Yu, F. Yan and J. Nawab, *J. Geophys. Res. Atmos.*, 2019, **124**, 7442-7456.
55. G. Casas, A. Martínez-Varela, J. L. Roscales, M. Vila-Costa, J. Dachs and B. Jiménez, *Environ. Pollut.*, 2020, **267**, 115512.
56. B. Sha, J. H. Johansson, P. Tunved, P. Bohlin-Nizzetto, I. T. Cousins and M. E. Salter, *Environ. Sci. Technol.*, 2022, **56**, 228-238.
57. B. Sha, J. H. Johansson, M. E. Salter, S. M. Blichner and I. T. Cousins, *Sci. Adv.*, 2024, **10**, eadl1026.
58. R. McGregor, *Rem. J.*, 2023, **33**, 265-278.
59. I. T. Cousins, J. C. DeWitt, J. Glüge, G. Goldenman, D. Herzke, R. Lohmann, C. A. Ng, M. Scheringer and Z. Wang, *Environ. Sci. Processes Impacts*, 2020, **22**, 2307-2312.
60. A. J. Lewis, X. Yun, D. E. Spooner, M. J. Kurz, E. R. McKenzie and C. M. Sales, *Sci. Total Environ.*, 2022, **822**, 153561.
61. L. Lesmeister, F. T. Lange, J. Breuer, A. Biegel-Engler, E. Giese and M. Scheurer, *Sci. Total Environ.*, 2021, **766**, 142640.
62. X. Yun, A. J. Lewis, G. Stevens-King, C. M. Sales, D. E. Spooner, M. J. Kurz, R. Suri and E. R. McKenzie, *Sci. Total Environ.*, 2023, **866**, 161208.
63. F. Pérez, M. Nadal, A. Navarro-Ortega, F. Fàbrega, J. L. Domingo, D. Barceló and M. Farré, *Environ. Int.*, 2013, **59**, 354-362.
64. W. Cheng, J. A. Doering, C. LaLone and C. Ng, *Toxicol. Sci.*, 2021, **180**, 212-223.
65. K. S. Betts, *Environ. Health Perspect.*, 2007, **115**, A250-A256.
66. S. E. Fenton, A. Ducatman, A. Boobis, J. C. DeWitt, C. Lau, C. Ng, J. S. Smith and S. M. Roberts, *Environ. Toxicol. Chem.*, 2021, **40**, 606-630.
67. *Per- and Polyfluoroalkyl Substances (PFAS) Report*, National Science and Technology Council, 2023. https://www.whitehouse.gov/wp-content/uploads/2023/03/OSTP-March-2023-PFAS-Report.pdf
68. S. Gaballah, A. Swank, J. R. Sobus, X. M. Howey, J. Schmid, T. Catron, J. McCord, E. Hines, M. Strynar and T. Tal, *Environ. Health Perspect.*, 2020, **128**, 047005.
69. K. Roth, Z. Imran, W. Liu and M. C. Petriello, *Front. Toxicol.*, 2020, **2**.
70. US-EPA, 2016, **81**. Lifetime Health Advisories and Health Effects Support Documents for Perfluorooctanoic Acid and Perfluorooctane Sulfonate. https://www.gpo.gov/fdsys/pkg/FR-2016-05-25/pdf/2016-12361.pdf
71. US-EPA, 2022. https://www.epa.gov/newsreleases/epa-announces-new-drinking-water-health-advisories-pfas-chemicals-1-billion-bipartisan
72. C. Hogue, *Chem. Eng. News,* 2022. https://cen.acs.org/policy/legislation/US-legislation-limit-PFAS-industrial/100/web/2022/07
73. Environmental Officials from Sweden, the Netherlands, Germany, and Denmark. Elements for an EU-strategy for PFAS, 2019. https://www.documentcloud.org/documents/6586418-EUStrategy-for-PFASs-FINAL-VERSION-December-19.html
74. ECHA publishes PFAS restriction proposal, 2023.https://echa.europa.eu/-/echa-publishes-pfas-restriction-proposal
75. https://echa.europa.eu/hot- topics/perfluoroalkyl-chemicals-pfas
76. https://www.pfasfree.org.uk/regulations #1527174044222-65be56c6-8e65
77. USA Department of Defense, 2023. https://media.defense.gov/2023/Jan/12/2003144157/-1/-1/1/MILITARY-SPECIFICATION-FOR-FIRE-EXTINGUISHING-AGENT-FLUORINE-FREE-FOAM-F3-LIQUID-CONCENTRATE-FOR-LAND-BASED-FRESH-WATER-APPLICATIONS.PDF
78. *Cosmetics Products Group Standard 2020*, Report HSR002552, New Zealand Environmental Protection Authority, 2024. https://www.epa.govt.nz/assets/RecordsAPI/Cosmetic-Products-Group-Standard-2020-Amended-January-2024.pdf
79. C. Schiavone and C. Portesi, *Applied Sciences*, 2023, **13**, 6696.
80. M. Wahlström, E. Pohjalainen, E. Yli-Rantala, D. Behringer, D. Herzke, S. Mudge, M. Beekman, A. Blaeij, J. Devilee, S.

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View Article Online DOI: 10.1039/D4SU00552D

Gabbert, M. Kuppevelt, M. Zare Jeddi, P. Gabrielsen and X. Trier, *Fluorinated polymers in a low carbon, circular and toxic-free economy*, 2021.
81. C. A. McDonough, J. L. Guelfo and C. P. Higgins, *Curr. Opin. Environ. Sci. Health*, 2019, **7**, 13-18.
82. R. A. Brase, E. J. Mullin and D. C. Spink, *Int. J. Molecular Sci.*, 2021, **22**, 995.
83. L. J. Winchell, M. J. M. Wells, J. J. Ross, X. Fonoll, J. W. Norton, S. Kuplicki, M. Khan and K. Y. Bell, *Sci. Total Environ.*, 2021, **774**, 145257.
84. Best Practices for Non-Targeted Analysis BP4NTA, https://nontargetedanalysis.org/reference-content/methods/study-design/).
85. R. F. Menger, E. Funk, C. S. Henry and T. Borch, *Chem. Eng. J.*, 2021, **417**, 129133.
86. R. Gonzalez de Vega, A. Cameron, D. Clases, T. M. Dodgen, P. A. Doble and D. P. Bishop, *J. Chromatogr. A*, 2021, **1653**, 462423.
87. PFAS Master list of PFAS Substances. https://comptox.epa.gov/dashboard/chemical-lists/pfasmaster.
88. E. F. Houtz and D. L. Sedlak, *Environ. Sci. Technol.*, 2012, **46**, 9342-9349.
89. R. Aro, U. Eriksson, A. Kärrman, I. Reber and L. W. Y. Yeung, *iScience*, 2021, **24**, 102968.
90. C. A. Moody, W. C. Kwan, J. W. Martin, D. C. G. Muir and S. A. Mabury, *Anal. Chem.*, 2001, **73**, 2200-2206.
91. D. Camdzic, R. A. Dickman and D. S. Aga, *J. Hazard. Mater. Lett.*, 2021, **2**, 100023.
92. E. E. Ritter, M. E. Dickinson, J. P. Harron, D. M. Lunderberg, P. A. DeYoung, A. E. Robel, J. A. Field and G. F. Peaslee, *Nucl. Instrum. Methods Phys. Res. B: Beam Interact. Mater. At.*, 2017, **407**, 47-54.
93. S. Heuckeroth, T. N. Nxumalo, A. Raab and J. Feldmann, *Anal. Chem.*, 2021, **93**, 6335-6341.
94. B. M.B, N. Rhakho, S. R. Jena, S. Yadav, A. Altaee, M. Saxena and A. K. Samal, *Sustain. Chem. Environ.*, 2023, **3**, 100031.
95. M. J. Davern, G. V. West, C. M. A. Eichler, B. J. Turpin, Y. Zhang and J. D. Surratt, *Analyst*, 2024, **149**, 3405-3415.
96. B. B. Bowers, J. A. Thornton and R. C. Sullivan, *Environ. Sci. Processes Impacts*, 2023, **25**, 277-287.
97. J. M. Mattila, J. D. Krug, W. R. Roberson, R. P. Burnette, S. McDonald, L. Virtaranta, J. H. Offenberg and W. P. Linak, *Environ. Sci. Technol.*, 2024, **58**, 3942-3952.
98. M. Hemida, A. Ghiasvand, V. Gupta, L. J. Coates, A. A. Gooley, H.-J. Wirth, P. R. Haddad and B. Paull, *Anal. Chem.*, 2021, **93**, 12032-12040.
99. S. Garg, P. Kumar, G. W. Greene, V. Mishra, D. Avisar, R. S. Sharma and L. F. Dumée, *J. Environ. Manage.*, 2022, **308**, 114655.
100. R. Khan, D. Andreescu, M. H. Hassan, J. Ye and S. Andreescu, *Angew. Chem. Int. Ed.*, 2022, **61**, e202209164.
101. H. Niu, S. Wang, Z. Zhou, Y. Ma, X. Ma and Y. Cai, *Anal. Chem.*, 2014, **86**, 4170-4177.
102. M. Takayose, K. Akamatsu, H. Nawafune, T. Murashima and J. Matsui, *Anal. Lett.*, 2012, **45**, 2856-2864.
103. C. Fang, X. Zhang, Z. Dong, L. Wang, M. Megharaj and R. Naidu, *Chemosphere*, 2018, **191**, 381-388.
104. R. F. Menger, J. J. Beck, T. Borch and C. S. Henry, *ACS Environ. Sci. Technol.*, 2022, **2**, 565-572.
105. H. Feng, N. Wang, T. Tran.T, L. Yuan, J. Li and Q. Cai, *Sens. Actuators B: Chem.*, 2014, **195**, 266-273.
106. Z. Zheng, H. Yu, W.-C. Geng, X.-Y. Hu, Y.-Y. Wang, Z. Li, Y. Wang and D.-S. Guo, *Nature Commun.*, 2019, **10**, 5762.
107. Q. Liu, A. Huang, N. Wang, G. Zheng and L. Zhu, *J. Lumin.*, 2015, **161**, 374-381.
108. M. Takayose, K. Nishimoto and J. Matsui, *Analyst*, 2012, **137**, 2762-2765.
109. H.-Q. Yin, K. Tan, S. Jensen, S. J. Teat, S. Ullah, X. Hei, E. Velasco, K. Oyekan, N. Meyer, X.-Y. Wang, T. Thonhauser, X.-B. Yin and J. Li, *Chem. Sci.*, 2021, **12**, 14189-14197.
110. V. Pichon and F. Chapuis-Hugon, *Anal. Chim. Acta*, 2008, **622**, 48-61.
111. R. B. Clark and J. E. Dick, *Chem. Commun.*, 2021, **57**, 8121-8130.
112. M. H. Hassan, R. Khan and S. Andreescu, *Electrochem. Sci. Adv.*, 2022, **2**, e2100184.
113. N. Karimian, A. M. Stortini, L. M. Moretto, C. Costantino, S. Bogialli and P. Ugo, *ACS Sensors*, 2018, **3**, 1291-1298.
114. R. Kazemi, E. I. Potts and J. E. Dick, *Anal. Chem.*, 2020, **92**, 10597-10605.
115. A. Concellón and T. M. Swager, *Angew. Chem. Int. Ed.*, **n/a**, e202309928.
116. S. Chen, A. Li, L. Zhang and J. Gong, *Anal. Chim. Acta*, 2015, **896**, 68-77.
117. Y. H. Cheng, D. Barpaga, J. A. Soltis, V. Shutthanandan, R. Kargupta, K. S. Han, B. P. McGrail, R. K. Motkuri, S. Basuray and S. Chatterjee, *ACS Appl. Mater. Interfaces*, 2020, **12**, 10503-10514.
118. X. Li, X. Wang, T. Fang, L. Zhang and J. Gong, *Talanta*, 2018, **181**, 147-153.
119. V. Trinh, C. S. Malloy, T. J. Durkin, A. Gadh and S. Savagatrup, *ACS Sensors*, 2022, **7**, 1514-1523.
120. C. M. Taylor, M. C. Breadmore and N. L. Kilah, *Chem.-Methods*, e202300017.
121. M. Zhang, X. Zhao, D. Zhao, T.-Y. Soong and S. Tian, *Waste Manage.*, 2023, **155**, 162-178.
122. J. N. Meegoda, B. Bezerra de Souza, M. M. Casarini and J. A. Kewalramani, *Int. J. Env. Res. Public Health*, 2022, **19**, 16397.
123. D. Lu, S. Sha, J. Luo, Z. Huang and X. Zhang Jackie, *J. Hazard. Mater.*, 2020, **386**, 121963.
124. T. Yamada, P. H. Taylor, R. C. Buck, M. A. Kaiser and R. J. Giraud, *Chemosphere*, 2005, **61**, 974-984.
125. W. Tsang, D. R. Burgess and V. Babushok, *Combust. Sci. Technol.*, 1998, **139**, 385-402.
126. P. H. Taylor, T. Yamada, R. C. Striebich, J. L. Graham and R. J. Giraud, *Chemosphere*, 2014, **110**, 17-22.
127. N. Bolan, B. Sarkar, Y. Yan, Q. Li, H. Wijesekara, K. Kannan, D. C. W. Tsang, M. Schauerte, J. Bosch, H. Noll, Y. S. Ok, K. Scheckel, J. Kumpiene, K. Gobindlal, M. Kah, J. Sperry, M. B. Kirkham, H. Wang, Y. F. Tsang, D. Hou and J. Rinklebe, *J. Hazard. Mater.*, 2021, **401**, 123892.
128. S. Takagi, F. Adachi, K. Miyano, Y. Koizumi, H. Tanaka, I. Watanabe, S. Tanabe and K. Kannan, *Water Res.*, 2011, **45**, 3925-3932.
129. R. Mukhopadhyay, B. Sarkar, K. N. Palansooriya, J. Y. Dar, N. S. Bolan, S. J. Parikh, C. Sonne and Y. S. Ok, *Adv. Colloid Interface Sci.*, 2021, **297**, 102537.
130. L. Zhao, J. Bian, Y. Zhang, L. Zhu and Z. Liu, *Chemosphere*, 2014, **114**, 51-58.
131. Q. Yu, R. Zhang, S. Deng, J. Huang and G. Yu, *Water Res.*, 2009, **43**, 1150-1158.

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RSC Sustainability Accepted Manuscript

132. D. Zhang, Q. Luo, B. Gao, S.-Y. D. Chiang, D. Woodward and Q. Huang, *Chemosphere*, 2016, **144**, 2336-2342.
133. E. Barth, J. McKernan, D. Bless and K. Dasu, *J. Environ. Manage.*, 2021, **296**, 113069.
134. F. Cao, L. Wang, Y. Tian, F. Wu, C. Deng, Q. Guo, H. Sun and S. Lu, *J. Chromatogr. A*, 2017, **1516**, 42-53.
135. Y. Wu, Y. Li, A. Tian, K. Mao and J. Liu, *Int. J. Photoenergy*, 2016, **2016**, 7368795.
136. Y. Dahman, in *Nanotechnology and Functional Materials for Engineers*, ed. Y. Dahman, Elsevier, 2017, DOI: https://doi.org/10.1016/B978-0-323-51256-5.00006-X, pp. 121-144.
137. S. J. Zaidi and H. Saleem, in *Reverse Osmosis Systems*, eds. S. J. Zaidi and H. Saleem, Elsevier, 2022, DOI: https://doi.org/10.1016/B978-0-12-823965-0.00006-7, pp. 141-177.
138. A. L. Kohl and R. B. Nielsen, in *Gas Purification (Fifth Edition)*, eds. A. L. Kohl and R. B. Nielsen, Gulf Professional Publishing, Houston, 1997, DOI: https://doi.org/10.1016/B978-088415220-0/50003-3, pp. 187-277.
139. S. Woodard, J. Berry and B. Newman, *Rem. J.*, 2017, **27**, 19-27.
140. F. Dixit, R. Dutta, B. Barbeau, P. Berube and M. Mohseni, *Chemosphere*, 2021, **272**, 129777.
141. F. Dixit, B. Barbeau, S. G. Mostafavi and M. Mohseni, *Environ. Sci. Water Res. Technol.*, 2019, **5**, 1782-1795.
142. M. F. Rahman, W. B. Anderson, S. Peldszus and P. M. Huck, *ACS Environ. Sci. Technol.*, 2022, **2**, 1195-1205.
143. Y.-L. Liu and M. Sun, *Water Res.*, 2021, **207**, 117781.
144. Y. Gao, S. Deng, Z. Du, K. Liu and G. Yu, *J. Hazard. Mater.*, 2017, **323**, 550-557.
145. A. Maimaiti, S. Deng, P. Meng, W. Wang, B. Wang, J. Huang, Y. Wang and G. Yu, *Chem. Eng. J.*, 2018, **348**, 494-502.
146. F. Dixit, B. Barbeau, S. G. Mostafavi and M. Mohseni, *J. Hazard. Mater.*, 2020, **384**, 121261.
147. A. Nickerson, A. E. Rodowa, D. T. Adamson, J. A. Field, P. R. Kulkarni, J. J. Kornuc and C. P. Higgins, *Environ. Sci. Technol.*, 2021, **55**, 313-323.
148. F. Dixit, G. Munoz, M. Mirzaei, B. Barbeau, J. Liu, S. V. Duy, S. Sauvé, B. Kandasubramanian and M. Mohseni, *Environ. Sci. Technol.*, 2022, **56**, 6212-6222.
149. C. Ching, M. J. Klemes, B. Trang, W. R. Dichtel and D. E. Helbling, *Environ. Sci. Technol.*, 2020, **54**, 12693-12702.
150. Y. Artioli, in *Encyclopedia of Ecology*, eds. S. E. Jørgensen and B. D. Fath, Academic Press, Oxford, 2008, DOI: https://doi.org/10.1016/B978-008045405-4.00252-4, pp. 60-65.
151. J. O'Connor, N. S. Bolan, M. Kumar, A. S. Nitai, M. B. Ahmed, S. S. Bolan, M. Vithanage, J. Rinklebe, R. Mukhopadhyay, P. Srivastava, B. Sarkar, A. Bhatnagar, H. Wang, K. H. M. Siddique and M. B. Kirkham, *Process Saf. Environ. Prot.*, 2022, **164**, 91-108.
152. S. C. E. Leung, P. Shukla, D. Chen, E. Eftekhari, H. An, F. Zare, N. Ghasemi, D. Zhang, N.-T. Nguyen and Q. Li, *Sci. Total Environ.*, 2022, **827**, 153669.
153. T. Buckley, X. Xu, V. Rudolph, M. Firouzi and P. Shukla, *Sep. Sci. Technol.*, 2022, **57**, 929-958.
154. B. Burghoff, *J. Biotechnol.*, 2012, **161**, 126-137.
155. T. Buckley, K. Karanam, X. Xu, P. Shukla, M. Firouzi and V. Rudolph, *Sep. Purif. Technol.*, 2022, **286**, 120508.
156. V. Franke, P. McCleaf, K. Lindegren and L. Ahrens, *Environ. Sci. Water Res. Technol.*, 2019, **5**, 1836-1843.
157. C. C. Murray, H. Vatankhah, C. A. McDonough, A. Nickerson, T. T. Hedtke, T. Y. Cath, C. P. Higgins and C. L. Bellona, *J. Hazard. Mater.*, 2019, **366**, 160-168.
158. S. Lath, E. R. Knight, D. A. Navarro, R. S. Kookana and M. J. McLaughlin, *Chemosphere*, 2019, **222**, 671-678.
159. S. Garg, J. Wang, P. Kumar, V. Mishra, H. Arafat, R. S. Sharma and L. F. Dumée, *J. Environ. Chem. Eng.*, 2021, **9**, 105784.
160. T. Le, E. Jamshidi, M. Beidaghi and M. R. Esfahani, *ACS Appl. Mater. Interfaces*, 2022, **14**, 25397-25408.
161. Y. Gogotsi and B. Anasori, *ACS Nano*, 2019, **13**, 8491-8494.
162. G. S. Park, D. H. Ho, B. Lyu, S. Jeon, D. Y. Ryu, D. W. Kim, N. Lee, S. Kim, Y. J. Song, S. B. Jo and J. H. Cho, *Sci. Adv.*, 2022, **8**, eabl5299.
163. Á. Soriano, D. Gorri and A. Urtiaga, *Ind. Eng. Chem. Res.*, 2019, **58**, 3329-3338.
164. T. G. Reichenauer and J. J. Germida, *ChemSusChem*, 2008, **1**, 708-717.
165. D. Q. Zhang, M. Wang, Q. He, X. Niu and Y. Liang, *Environ. Pollut.*, 2020, **257**, 113575.
166. A. Gredelj, C. Nicoletto, S. Valsecchi, C. Ferrario, S. Polesello, R. Lava, F. Zanon, A. Barausse, L. Palmeri, L. Guidolin and M. Bonato, *Sci. Total Environ.*, 2020, **708**, 134766.
167. E. Yamazaki, S. Taniyasu, K. Noborio, H. Eun, P. Thaker, N. J. I. Kumar, X. Wang and N. Yamashita, *Chemosphere*, 2019, **231**, 502-509.
168. B. Wen, L. Li, H. Zhang, Y. Ma, X.-Q. Shan and S. Zhang, *Environ. Pollut.*, 2014, **184**, 547-554.
169. J. Awad, G. Brunetti, A. Juhasz, M. Williams, D. Navarro, B. Drigo, J. Bougoure, J. Vanderzalm and S. Beecham, *J. Hazard. Mater.*, 2022, **429**, 128326.
170. A. Chavoshani, M. Hashemi, M. Mehdi Amin and S. C. Ameta, in *Micropollutants and Challenges*, eds. A. Chavoshani, M. Hashemi, M. Mehdi Amin and S. C. Ameta, Elsevier, 2020, DOI: https://doi.org/10.1016/B978-0-12-818612-1.00002-7, pp. 35-90.
171. L. Martín-Pozo, M. d. C. Gómez-Regalado, M. T. García-Córcoles and A. Zafra-Gómez, in *Emerging Contaminants in the Environment*, eds. H. Sarma, D. C. Dominguez and W.-Y. Lee, Elsevier, 2022, DOI: https://doi.org/10.1016/B978-0-323-85160-2.00015-9, pp. 381-406.
172. B. N. Nzeribe, M. Crimi, S. Mededovic Thagard and T. M. Holsen, *Crit. Rev. Environ. Sci. Technol.*, 2019, **49**, 866-915.
173. Z. Liu, M. J. Bentel, Y. Yu, C. Ren, J. Gao, V. F. Pulikkal, M. Sun, Y. Men and J. Liu, *Environ. Sci. Technol.*, 2021, **55**, 7052-7062.
174. D. Panda, V. Sethu and S. Manickam, *Chem. Eng. Process. Process Intensification*, 2019, **142**, 107542.
175. C. G. Scheitlin, K. Dasu, S. Rosansky, L. E. Dejarme, D. Siriwardena, J. Thorn, L. Mullins, I. Haggerty, K. Shqau and J. Stowe, *ACS Environ. Sci. Technol.*, 2023, **3**, 2053-2062.
176. E. Houtz, D. Kempisty and Y. Lester, *Curr. Opin. Chem. Eng.*, 2024, **44**, 101022.
177. J. Li, B. R. Pinkard, S. Wang and I. V. Novosselov, *Chemosphere*, 2022, **307**, 135888.
178. S. Hao, Y.-J. Choi, B. Wu, C. P. Higgins, R. Deeb and T. J. Strathmann, *Environ. Sci. Technol.*, 2021, **55**, 3283-3295.
179. B. Wu, S. Hao, Y. Choi, C. P. Higgins, R. Deeb and T. J. Strathmann, *Environ. Sci. Technol. Lett.*, 2019, **6**, 630-636.

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10.1039/D4SU00152D
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RSC Sustainability Accepted Manuscript

View Article Online DOI: 10.1039/D4SU00390E

180. M. Ateia, L. P. Skala, A. Yang and W. R. Dichtel, *J. Hazard. Mater. Adv.*, 2021, **3**, 100014.
181. B. G. Kwon, H.-J. Lim, S.-H. Na, B.-I. Choi, D.-S. Shin and S.-Y. Chung, *Chemosphere*, 2014, **109**, 221-225.
182. S. P. Chetverikov, D. A. Sharipov, T. Y. Korshunova and O. N. Loginov, *Appl. Biochem. Microbiol.*, 2017, **53**, 533-538.
183. S. Huang and P. R. Jaffé, *Environ. Sci. Technol.*, 2019, **53**, 11410-11419.
184. S. P. Chetverikov and O. N. Loginov, *Microbiology*, 2019, **88**, 115-117.
185. J. Li, X. Li, Y. Da, J. Yu, B. Long, P. Zhang, C. Bakker, B. A. McCarl, J. S. Yuan and S. Y. Dai, *Nature Commun.*, 2022, **13**, 4368.
186. K. Tahir, A. S. Ali, J. Kim, J. Park, S. Lee, B. Kim, Y. Lim, G. Kim and D. S. Lee, *Chemosphere*, 2023, **328**, 138584.
187. D. Alam, S. Lee, J. Hong, D. F. Fletcher, D. McClure, D. Cook, P. J. Cullen and J. M. Kavanagh, *J. Environ. Chem. Eng.*, 2023, **11**, 111588.
188. O. Isowamwen, R. Li, T. Holsen and S. M. Thagard, *J. Hazard. Mater.*, 2023, **456**, 131691.
189. B. Améduri and H. Hori, *Chem. Soc. Rev.*, 2023, **52**, 4208-4247.
190. A. L. Duchesne, J. K. Brown, D. J. Patch, D. Major, K. P. Weber and J. I. Gerhard, *Environ. Sci. Technol.*, 2020, **54**, 12631-12640.
191. T. Fournie, T. L. Rashwan, C. Switzer and J. I. Gerhard, *Waste Manage.*, 2023, **164**, 219-227.
192. B. N. Nzeribe, M. Crimi, S. Mededovic Thagard and T. M. Holsen, *Critical Reviews in Environ. Sci. Technol.*, 2019, **49**, 866-915.
193. O. C. Olatunde, A. T. Kuvarega and D. C. Onwudiwe, *Heliyon*, 2020, **6**, e05614.
194. M. B. Ahmed, M. M. Alam, J. L. Zhou, B. Xu, M. A. H. Johir, A. K. Karmakar, M. S. Rahman, J. Hossen, A. T. M. K. Hasan and M. A. Moni, *Process Saf. Environ. Protection*, 2020, **136**, 1-14.
195. M. Umar, *Water*, 2021, **13**, 3185.
196. Y. Guan, Z. Liu, N. Yang, S. Yang, L. E. Quispe-Cardenas, J. Liu and Y. Yang, *Nature Water*, 2024, **2**, 443-452.
197. N. Tran, P. Drogui and S. K. Brar, *Environ. Chem. Lett.*, 2015, **13**, 251-268.
198. C. G. Joseph, G. Li Puma, A. Bono and D. Krishnaiah, *Ultrason. Sonochem.*, 2009, **16**, 583-589.
199. P. Kritzer and E. Dinjus, *Chem. Eng. J.*, 2001, **83**, 207-214.
200. Z. Chen, K. Tong, F. Xu, M. Xue, H. Chen, Q. Chen, D. Wang and Y. Xu, *J. Environ. Chem. Eng.*, 2021, **9**, 105296.
201. C. Berg, B. Crone, B. Gullett, M. Higuchi, M. J. Krause, P. M. Lemieux, T. Martin, E. P. Shields, E. Struble, E. Thoma and A. Whitehill, *J. Air Waste Manage. Assoc.*, 2022, **72**, 540-555.
202. M. J. Bentel, Y. Yu, L. Xu, Z. Li, B. M. Wong, Y. Men and J. Liu, *Environ. Sci. Technol.*, 2019, **53**, 3718-3728.
203. J. Cui, P. Gao and Y. Deng, *Environ. Sci. Technol.*, 2020, **54**, 3752-3766.
204. M. J. Bentel, Y. Yu, L. Xu, H. Kwon, Z. Li, B. M. Wong, Y. Men and J. Liu, *Environ. Sci. Technol.*, 2020, **54**, 2489-2499.
205. Z. Ren, U. Bergmann and T. Leiviskä, *Water Res.*, 2021, **205**, 117676.
206. M. Fischer and P. Warneck, *J. Phys. Chem.*, 1996, **100**, 15111-15117.
207. I. M. F. Cardoso, L. Pinto da Silva and J. C. G. Esteves da Silva, *Nanomaterials*, 2023, **13**, 1668.
208. N. O'Connor, D. Patch, D. Noble, J. Scott, J. Koch, K. G. Mumford and K. Weber, *Sci. Total Environ.*, 2023, **888**, 164137.
209. W. A. Maza, J. A. Ridenour, B. L. Chaloux, A. Epshteyn and J. C. Owrutsky, *Envrion. Sci. Adv.*, 2023, **2**, 1641-1650.
210. X. Jin, Z. Wang, R. Hong, Z. Chen, B. Wu, S. Ding, W. Zhu, Y. Lin and C. Gu, *Water Res.*, 2022, **225**, 119147.
211. H. Tian, J. Gao, H. Li, S. A. Boyd and C. Gu, *Sci. Rep.*, 2016, **6**, 32949.
212. J. F. King and B. P. Chaplin, *Curr. Opin. Chem. Eng.*, 2024, **44**, 101014.
213. B. Trang, Y. Li, X.-S. Xue, M. Ateia, K. N. Houk and W. R. Dichtel, *Science*, 2022, **377**, 839-845.
214. S. Hao, Y. J. Choi, R. A. Deeb, T. J. Strathmann and C. P. Higgins, *Environ. Sci. Technol.*, 2022, **56**, 6647-6657.
215. B. R. Pinkard, C. Austin, A. L. Purohit, J. Li and I. V. Novosselov, *Chemosphere*, 2023, **314**, 137681.
216. B. R. Pinkard, *J. Environ. Eng.*, 2022, **148**, 05021007.
217. L. P. Turner, B. H. Kueper, K. M. Jaansalu, D. J. Patch, N. Battye, O. El-Sharnouby, K. G. Mumford and K. P. Weber, *Sci. Total Environ.*, 2021, **765**, 142722.
218. G. Cagnetta, H. Liu, K. Zhang, J. Huang, B. Wang, S. Deng, Y. Wang and G. Yu, *Sci. Rep.*, 2016, **6**, 28394.
219. M. Lu, G. Cagnetta, K. Zhang, J. Huang and G. Yu, *Sci. Rep.*, 2017, **7**, 17180.
220. K. Gobindlal, E. Shields, A. Whitehill, C. C. Weber and J. Sperry, *Envrion. Sci. Adv.*, 2023, **2**, 982-989.
221. O. Tiedt, M. Mergelsberg, K. Boll, M. Müller, L. Adrian, N. Jehmlich, M. v. Bergen and M. Boll, *mBio*, 2016, **7**, 10.1128/mbio.00990-00916.
222. M. Ateia, A. Maroli, N. Tharayil and T. Karanfil, *Chemosphere*, 2019, **220**, 866-882.
223. C. Hogue, *Chem. Eng. News*, 2021, 99, **40** https://cen.acs.org/environment/persistent-pollutants/US-EPA-deems-two-GenX-PFAS-chemicals-more-toxic-than-PFOA/99/i40
224. C. Ng, I. T. Cousins, J. C. DeWitt, J. Glüge, G. Goldenman, D. Herzke, R. Lohmann, M. Miller, S. Patton, M. Scheringer, X. Trier and Z. Wang, *Environ. Sci. Technol.*, 2021, **55**, 12755-12765.
225. S. V. Kabadi, J. Fisher, J. Aungst and P. Rice, *Food Chem. Toxicol.*, 2018, **112**, 375-382.
226. D. K. Jones, K. A. Quinlin, M. A. Wigren, Y. J. Choi, M. S. Sepúlveda, L. S. Lee, D. L. Haskins, G. R. Lotufo, A. Kennedy, L. May, A. Harmon, T. Biber, N. Melby, M. K. Chanov, M. L. Hudson, P. B. Key, K. W. Chung, D. W. Moore, J. G. Suski, E. F. Wirth and J. T. Hoverman, *Environ. Sci. Technol.*, 2022, **56**, 6078-6090.
227. F. Hossain, N. M. Dennis, S. Subbiah, A. Karnjanapiboonwong, J. L. Guelfo, J. Suski and T. A. Anderson, *Environ. Toxicol. Chem.*, 2022, **41**, 2003-2007.
228. X. Wu, H. Nguyen, D. Kim and H. Peng, *Sci. Total Environ.*, 2022, **850**, 158100.
229. G. Glenn, R. Shogren, X. Jin, W. Orts, W. Hart-Cooper and L. Olson, *Compr. Rev. Food Sci. Food Saf.*, 2021, **20**, 2596-2625.
230. Firefighting Foams, https://pfas-1.itrcweb.org/3-firefighting-foams/.
231. F. F. F. Coalition, https://www.fffc.org/.
232. J. Horst, J. Quinnan, J. McDonough, J. Lang, P. Storch, J. Burdick and C. Theriault, *Groundwater Monit. Rem.*, 2021, **41**, 19-26.

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RSC Sustainability Accepted Manuscript

233. S. R. K. Fernández, C.; Bruton, T., 2021. https://greensciencepolicy.org/docs/pfas-building-materials-2021.pdf
234. S. Schellenberger, P. J. Hill, O. Levenstam, P. Gillgard, I. T. Cousins, M. Taylor and R. S. Blackburn, *J. Cleaner Prod.*, 2019, **217**, 134-143.
235. H. D. Whitehead, M. Venier, Y. Wu, E. Eastman, S. Urbanik, M. L. Diamond, A. Shalin, H. Schwartz-Narbonne, T. A. Bruton, A. Blum, Z. Wang, M. Green, M. Tighe, J. T. Wilkinson, S. McGuinness and G. F. Peaslee, *Environ. Sci. Technol. Lett.*, 2021, **8**, 538-544.
236. K. J. Harris, G. Munoz, V. Woo, S. Sauvé and A. A. Rand, *Environ. Sci. Technol.*, 2022, **56**, 14594-14604.
237. K. W. Pütz, S. Namazkar, M. Plassmann and J. P. Benskin, *Environ. Sci. Processes Impacts*, 2022, **24**, 1697-1707.
238. D. K. Lay, Ian; Tiscione, Scott; Kreissig, Julius, *Check Your Tech: A guide to PFAS in electronics*, 2023.
239. J. Glüge, R. London, I. T. Cousins, J. DeWitt, G. Goldenman, D. Herzke, R. Lohmann, M. Miller, C. A. Ng, S. Patton, X. Trier, Z. Wang and M. Scheringer, *Environ. Sci. Technol.*, 2022, **56**, 6232-6242.
View Article Online
DOI: 10.1039/D4SU00152D
240. Fracking with "Forever Chemicals", https://psr.org/resources/fracking-with-forever-chemicals/.
241. https://www.slideshare.net/slideshows/oecd-global-forum-on-the-environment-dedicated-to-per-and-polyfluoroalkyl-substances-available-pfas-alternatives-and-innovation-needs-anna-lennquist/266308319, (accessed Feb 16, 2024).
242. US-EPA, 2023. TSCA Section 8(a) (&) Reporting and Recordkeeping Requirements for Perfluoroalkyl and Polyfluoroalkyl Substances. https://www.epa.gov/assessing-and-managing-chemicals-under-tsca/tsca-section-8a7-reporting-and-recordkeeping.

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No primary research results, software or code have been included and no new data were generated or analysed as part of this review.

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