

Cite this: *Chem. Sci.*, 2019, 10, 1677

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The biobehavior, biocompatibility and theranostic application of SPNS and Pd@Au nanoplates in rats and rabbits†

Jingchao Li,^a Hongzhi Liu,^c Jiang Ming,^a Duo Sun,^a Xiaolan Chen,^{ID} *^{ab}
Xiaolong Liu,^{ID} ^c and Nanfeng Zheng,^{ID} *^a

On account of the fascinating surface plasmon resonance (SPR) properties, the ability of passively targeting tumors and remarkable biocompatibility, two-dimensional (2D) Pd-based nanomaterials have demonstrated wide application prospects in cancer theranostics. However, the used animal models for exploring the bioapplications and biosafety of 2D Pd-based nanomaterials were usually limited to mice. To further widen their biomedical applications and promote future clinical transformation, it is necessary to make a breakthrough in animal models. In this work, Sprague Dawley (SD) rats and New Zealand rabbits were used as the experimental animals and orthotopic liver tumors or subcutaneous tumors were induced in these animals. Taking ≈ 5 nm small Pd nanosheets (SPNS) and 30 nm Pd@Au nanoplates (Pd@Au) as the representative 2D Pd-based nanomaterials, we investigated their biobehaviors and biosafety in rat liver & subcutaneous tumor models and rabbit liver tumors. The results indicated that SPNS and Pd@Au could still effectively accumulate on the tumor sites of these bigger animal models by the enhanced permeability and retention (EPR) effect, and the accumulation effects were closely related to their sizes. Metabolism studies confirmed that SPNS could be excreted out of rats through urine. Moreover, based on the sufficient uptake by cancer cells and passive accumulation of SPNS and Pd@Au in subcutaneous tumors in rats, we performed photothermal therapy (PTT) *in vitro* and *in vivo*. Significant tumor growth inhibition illustrated that even though the animal model was dozens of times bigger than the mouse model, the 2D Pd-based nanomaterials satisfied the requirements of being an outstanding photothermal reagent. Finally, the hematological and histological examination results suggested that SPNS and Pd@Au had favorable biocompatibility in rats and rabbits at a given dose. We hope this work will drive the development of 2D Pd-based nanomaterials towards practical clinical applications and provide a guide for other theranostic nanoplatfoms that will be applied in bigger animal tumor models in the future.

Received 28th September 2018
Accepted 24th November 2018

DOI: 10.1039/c8sc04318c

rsc.li/chemical-science

^aState Key Laboratory for Physical Chemistry of Solid Surfaces, Collaborative Innovation Center of Chemistry for Energy Materials, National & Local Joint Engineering Research Center of Preparation Technology of Nanomaterials, College of Chemistry and Chemical Engineering, Xiamen University, Xiamen 361005, China. E-mail: chenxl@xmu.edu.cn; nfhzheng@xmu.edu.cn

^bState Key Laboratory for Chemistry and Molecular Engineering of Medicinal Resources, Guangxi Normal University, Guilin 541004, China

^cThe United Innovation of Mengchao Hepatobiliary Technology Key Laboratory of Fujian Province, Mengchao Hepatobiliary Hospital of Fujian Medical University, Fuzhou, 350025, China

† Electronic supplementary information (ESI) available: The photos of the rabbit orthotopic liver tumor, and orthotopic liver tumor and subcutaneous tumor of rats, molar absorption coefficients of SPNS at different wavelengths, TEM images of Pd@Au-PEG and Pd@Au-PEG after laser irradiation, the biodistribution results of SPNS and Pd@Au in rats and rabbits calculated in % ID, accumulation amount changes of SPNS in rat kidney, excretion of Pd@Au by feces, loading efficiencies of Cy5.5 on SPNS and Pd@Au, color changes of rat subcutaneous tumors after intravenous injection of SPNS and Pd@Au, cellular uptake of SPNS and Pd@Au by W256 cells and VX2 cells, and PTT efficiency for *in vitro* killing VX2 cells. See DOI: 10.1039/c8sc04318c

Introduction

With the advantage of size which is close to those of some biological molecules like DNA, enzymes and proteins, nanomaterials have been widely used in the biomedical field.¹ Therein, the application of nanomaterials in cancer theranostics has become a hot research focus and attracted much attention.^{2–5} Many nanomaterial-based theranostic platforms have been reported to greatly improve the imaging and therapeutic efficiency for cancer.^{6–12} To evaluate the theranostic effects as well as biosafety of the designed nanoplatfoms, certain animal models need to be adopted. In general, mice are common and widely used laboratory animals because they are mammals like humans, and the price of mice is low and they can reproduce quickly.¹³ The strains of mice for experimental use contain BALB/C, C57BL/6, DBA/2 and so on.^{14–17} Different types of tumors including subcutaneous tumors, orthotopic liver tumors and metastatic tumors could be induced on these



The corresponding molar absorption coefficients of SPNS (40 ppm) at different wavelengths were also measured based on the Beer-Lambert Law and the ϵ value could reach around $1534 \text{ L mol}^{-1} \text{ cm}^{-1}$ at a wavelength of 808 nm (Fig. S2†). To extend the blood circulation half-lives of SPNS and Pd@Au in rats and rabbits and compare the *in vivo* results with those in mice models, SPNS and Pd@Au were further surface-modified with thiol-polyethylene glycol (mPEG-SH) to obtain SPNS-PEG and Pd@Au-PEG, respectively. The morphologies, sizes and absorption spectra of SPNS-PEG and Pd@Au-PEG were similar to those of the corresponding SPNS and Pd@Au (Fig. 1c and S3†). And after laser irradiation for 10 min, the morphology of Pd@Au-PEG was still maintained well which demonstrated their good photothermal stability (Fig. S4†). Therefore, the excellent absorptions of SPNS-PEG and Pd@Au-PEG in the NIR region together with their high photothermal conversion capability (Fig. 1d), stability and good biocompatibility endowed them with great potential for application in cancer theranostics.

In vivo pharmacokinetics, excretion and biodistribution studies in rats bearing an orthotopic liver tumor model

As demonstrated above, 2D Pd-based nanomaterials with high photothermal conversion efficiency and good biocompatibility have brilliant prospects for application in cancer PTT and PTT-based combined therapy. However so far, the *in vivo* behavior explorations and theranostic applications of 2D Pd-based nanomaterials are still limited to mice. To further promote the biomedical applications of Pd-based 2D nanomaterials, more animal models are required to assess their biosafety and theranostic effects. Here, we first investigated the blood circulation, biodistribution and urinary excretion of SPNS and Pd@Au in rats. In our previous work, it was found that the blood circulations of 2D Pd-based nanomaterials in mice were closely related to their sizes and surface modifications.^{28,32} A similar size-dependent circulation time was also observed in rats. As shown in Fig. 2a, the circulation half-lives of SPNS-PEG and Pd@Au-PEG in rats by measuring the Pd concentrations in blood by ICP-MS were 10 h and 2.5 h, respectively. It was easier for SPNS with a super small size ($\approx 5 \text{ nm}$) to escape the capture of the reticuloendothelial system (RES), so the blood circulation time of SPNS was longer than that of 30 nm Pd@Au. The long circulation half-life also gave nanoparticles more contact time to accumulate in the target tissues. Fig. 2c and d show the biodistribution results of SPNS and Pd@Au in rats with the orthotopic liver tumor model which was fabricated by surgical operation and injection of cancer cells, respectively. Owing to their ultra-small size as well as long circulation time, SPNS had a relatively broad distribution in various organs, including the heart, liver, spleen, lung, kidney and tumor. Especially the high concentration in kidneys indicated that SPNS might be excreted out of the body through the urinary system. Compared to SPNS, the larger-sized Pd@Au mainly accumulated in the spleen. It was worth noting that the accumulations of SPNS and Pd@Au in the orthotopic liver tumors increased with time. At 48 h post injection, the accumulation amounts were 2.03% ID per g and



Fig. 2 The investigation of biobehavior *in vivo* ($n = 4$ for each group): blood circulation (a) and urinary excretion (b) of SPNS-PEG and Pd@Au-PEG; biodistribution of SPNS-PEG (c) and Pd@Au-PEG (d) in rats with the orthotopic liver tumor model. Asterisks suggest statistically significant differences (* $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$).

2.41% ID per g for SPNS and Pd@Au, respectively (Fig. 2c and d). Moreover, the biodistribution results that used % ID as the unit are also presented in Fig. S5(a and b†). The size effect was also apparent in urinary excretion. It is generally recognized that nanomaterials that are less than 10 nm in size would be excreted out from the glomerulus through urine.^{33,34} In our study, 10.3% ID SPNS was excreted out from the body through urine within one day but some (0.56% ID) Pd@Au could be detected in urine during the same period and the amount remained invariable (Fig. 2b). For SPNS, the metabolic rate started to go down after 2 days and the total amount of SPNS excreted out through urine within 8 days was about 22% ID. The trend was also confirmed by the accumulation amount changes of SPNS in the kidney with time (Fig. S6†). Furthermore, the possible metabolic pathway of Pd@Au was also further investigated, and about 9% ID of Pd@Au could be excreted out from the body through feces within 8 days (Fig. S7†), indicating the larger-sized Pd@Au could not be effectively cleared out of the body through urine and feces metabolism routes. The feature of SPNS that can be easily eliminated out of the body *via* urine enables it to be an ideal nanoplatform for application in cancer theranostics.

In vivo biodistribution of SPNS and Pd@Au in rat subcutaneous tumor model

Due to the simplicity of fabrication, the subcutaneous tumor model was frequently used in mice to appraise the performance of the theranostic nanoplatform. For better comparison with the results in the mouse subcutaneous tumor model, we further



investigated the biodistribution of SPNS and Pd@Au in the rat subcutaneous tumor model. Fluorescence imaging is widely used for tumor diagnosis because of its high resolution and sensitivity. With facile surface functionalization, 2D Pd-based nanosheets (NSs) could serve as a suitable fluorescence probe carrier for *in vitro* and *vivo* fluorescence imaging. By way of covalent linkage, Cy5.5 modified NSs were successfully synthesized and intravenously injected into rats. The loading efficiencies of Cy5.5 on SPNS (18.8%, w/w) and Pd@Au (25.6%, w/w) were calculated by the difference of adding Cy5.5 and the Cy5.5 in the supernatant after purification according to the standard absorption curve of Cy5.5 at 678 nm (Fig. S8†). As shown in Fig. 3a, free dye could not accurately target subcutaneous tumor sites so the fluorescence signal tended to be dispersive in the scanned area. Obviously, the intensity of the signal at tumor sites was not higher than that in the normal area. However, SPNS-Cy5.5 or Pd@Au-Cy5.5 by tail vein injection rapidly accumulated in tumors and there was a strong fluorescence signal at 24 h post injection. As time went by, signals at tumor sites became stronger at 48 h than before. The fluorescence imaging results not only indicated that SPNS and Pd@Au had the potential to be the carriers of some molecule imaging contrast agents, but also qualitatively confirmed that the SPNS and Pd@Au possessed nice tumor passive targeting ability in rats. Moreover, after intravenous injection of nano-materials, the subcutaneous tumor sites obviously grew black

within 6 h for SPNS (Fig. S9†) and 24 h for Pd@Au (Fig. S10†), which more intuitively showed the specific accumulation of SPNS and Pd@Au in target tissue.

Meanwhile, the precise quantitation of biodistribution in the rat subcutaneous tumor model by ICP-MS was also performed. As seen from Fig. 3b and c, the accumulation amounts of SPNS or 30 nm Pd@Au in tumors kept continuously increasing at 2 h, 12 h, 24 h and 48 h post injection, which was similar to that in the orthotopic liver tumor model due to the similar internal structures of these two tumor models. And at 48 h the enrichment reached 2.48% ID per g for SPNS and 3.25% ID per g for Pd@Au, respectively (the % ID distribution data are also shown in Fig. S5(c and d)†). These results further suggested that both SPNS and 30 nm Pd@Au had fantastic passive targeting ability to recognize lesions in the subcutaneous tumor of rats.

In vitro and *in vivo* PTT of tumor cells and subcutaneous tumors of rats

The excellent absorptions in the NIR region of SPNS and Pd@Au together with their abilities of passively targeting tumors could



Fig. 3 Live fluorescence imaging and signals in major tissues (H: heart, Li: liver, S: spleen, Lu: lung, K: kidney and T: tumor) of Cy5.5, SPNS-Cy5.5 and Pd@Au-Cy5.5 (a), respectively. The red circles represent the location of the tumors. Biodistribution of SPNS-PEG (b) and Pd@Au-PEG (c) in the rat subcutaneous tumor model ($n = 4$ for each group). In all fluorescence images, the amounts of Cy5.5 in different groups were 200 μ g. The scale bar in a is 1 cm.

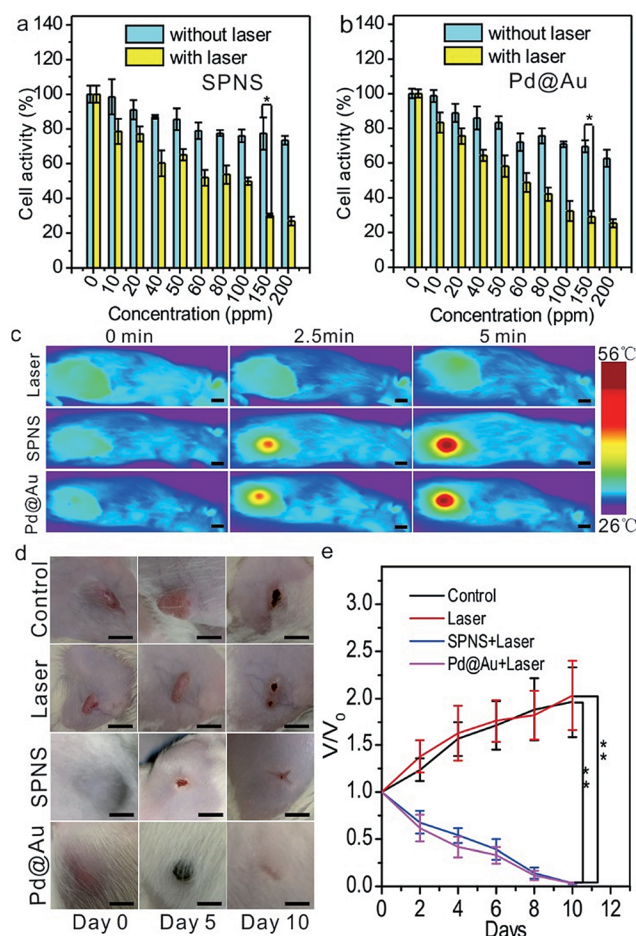


Fig. 4 *In vitro* PTT of SPNS (a) and Pd@Au (b) ($n = 8$ for each group). The temperature increases of tumor sites upon laser irradiation after intravenous injection with SPNS and Pd@Au (c). *In vivo* PTT of SPNS and Pd@Au (d) and the tumor growth curves in different treatment groups (e) ($n = 4$ for each group). Asterisks suggest statistically significant differences (* $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$).





Fig. 5 Biodistribution of SPNS (a) and Pd@Au (b) in rabbit orthotopic liver tumor models ($n = 4$). (c) PA imaging comparison of the spleen and tumor of Pd@Au-injected and PBS-injected rabbits. The scale bar for (c) and (d) is 1 cm.

be incorporated to perform the PTT in the rat subcutaneous tumor model. To visually observe the uptake of nanomaterials by W256 rat tumor cells, SPNS and Pd@Au which were surface modified with the dye of FITC were co-cultured with the tumor cells for 6 h in 24-well plates. Observed using a Fluorescence Inversion Microscope System, we found that both SPNS and

Pd@Au could be effectively taken in by W256 cells (Fig. S11†). At the same time, the cell toxicity and PTT effect of SPNS and Pd@Au on W256 cells were also measured by MTT assay. The cell activity remained at a high level when co-cultured with SPNS or Pd@Au in the absence of 808 nm laser irradiation, while the cell activity decreased to less than 30% with laser irradiation (0.5 W cm^{-2}) for 5 min. The favourable therapeutic efficiency on the cellular level predicted that the solid tumor *in vivo* might be cured by PTT depending on the photothermal conversion capability of SPNS and Pd@Au.

Before *in vivo* treatment, we first investigated if SPNS and Pd@Au accumulated in the subcutaneous tumor sites of rats after intravenous administration could still generate a good rising temperature effect, which was the precondition for PTT. To perform this, 5 mg kg^{-1} SPNS or Pd@Au were intravenously injected into the subcutaneous tumor-bearing rats. At 48 h post injection, the rats were exposed to 808 nm laser (0.5 W cm^{-2}) irradiation for 5 min, and the temperature changes of the tumor sites were recorded using a visual IR camera. As observed from Fig. 4c, in the groups treated with SPNS and Pd@Au for 5 min, the temperature of the tumor sites could reach about $55 \text{ }^\circ\text{C}$ (Fig. 4c), whereas for the group treated with only laser irradiation for 5 min, the tumor surface temperature could only reach $33.6 \text{ }^\circ\text{C}$. The results suggested that SPNS and Pd@Au accumulated in the subcutaneous tumors of rats could still effectively convert the absorbed light energy into heat for PTT treatment.



Fig. 6 The studies of biocompatibility: (a) blood routine and (b) blood serum biochemistry examination of SPNS and Pd@Au in rats.





Fig. 8 Histological examination of SPNS and Pd@Au in (a) rats and (b) rabbits, respectively. The scale bar is 50 μm .

have a great influence on the *in vivo* biobehaviors of 2D Pd-based nanomaterials in mice. In brief, these results suggested that the excellent EPR effect of 2D Pd-based nanomaterials was still retained in the rabbit orthotopic liver tumor model. Considering their capabilities for cellular internalization and *in vitro* killing VX2 cancer cells by PTT (Fig. S12 and S13[†]), 2D-based nanomaterials hold promise for realizing PTT or PTT-based combined therapy in the rabbit tumor model, which further convincingly proved their value for clinical translation.

Biosafety of SPNS and Pd@Au in rats and rabbits

As mentioned above, SPNS and Pd@Au have excellent accumulation capacities in both rat and rabbit tumor sites by the EPR effect; however, it was still unclear if they could result in potential toxicity in some biochemical analysis including total proteins (TP), albumin (ALB), globulin (GLO), albumin/globulin (A/G), alkaline phosphatase (ALP), alanine aminotransferase (ALT), aspartate aminotransferase (AST) and blood urea nitrogen (BUN). From the results presented in Fig. 6 and 7, in the rat or rabbit models, all these detection indicators were changed within the normal range, which indicated that SPNS and Pd@Au would not cause an abnormal blood composition and side-effects on liver/kidney functions. What's more, histological analysis was performed to discuss whether SPNS and Pd@Au could give rise to inflammation or lesions on five major organs, including the heart, liver, spleen, lung and kidney. As shown in Fig. 8, the liver and spleen of rats and rabbits which were injected with the nanomaterial displayed a slight inflammation effect, possibly because of the high amount

accumulation of nanomaterials in these two organs. Compared with the control group, other organs basically remained normal and there was no obvious damage. The above results manifested the superior biocompatibility of SPNS and Pd@Au for bigger animal models at the given dose, which was a valid supplement to the safety evaluation of 2D Pd-based nanomaterials.

Conclusions

Based on the prominent theranostic effects of 2D Pd-based nanomaterials on mouse tumor models before, in this work we successfully fabricated SD rat and New Zealand rabbit tumor models and systematically investigated the biobehaviors and biosafety of SPNS and 30 nm Pd@Au in these bigger animals. The results indicated that both SPNS and Pd@Au could efficiently accumulate in orthotopic liver tumors or subcutaneous tumors through the EPR effect. Utilizing the passive targeting ability of SPNS and Pd@Au in subcutaneous tumor together with their high photothermal conversion properties, highly efficient PTT was carried out to effectively eradicate tumors. Similar to the results obtained from mice, the urinary excretion and blood circulation of SPNS and Pd@Au were directly related to their sizes. Owing to the advantages of long blood circulation time and easy-elimination from the body, SPNS have been shown to have great clinical translation value. Moreover, SPNS and Pd@Au were proved to be safe in bigger animal models from the results of blood biochemical analysis and histopathology.



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