

Nanoscale

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ARTICLE

One- and Two-Photon Brightness of Proteins Interacting with Gold. A Closer Look at Gold-Insulin Conjugates

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Received 00th January 20xx,
Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

Red luminophores displaying large Stokes shift and high-quantum yields are obtained when gold salts are reacted with proteins under strongly alkaline conditions. Although bovine serum albumin (BSA) has mainly been used as a protein template, other attempts to prepare red luminophores have been proposed using other proteins. Here, we report on the structural characterization and nonlinear optical properties of insulin-gold conjugates. Such conjugates display strong luminescence at ≈ 670 nm with quantum yields that reach 5.4%. They also display long luminescence lifetimes allowing efficient reactive oxygen species generation, with a quantum yield of $^1\text{O}_2$ generation reaching $\sim 10\%$. In addition, they exhibit remarkable nonlinear optical properties and in particular a strong two-photon excited fluorescence (TPEF) cross section in the range of 800–1100 nm. By combining experimental studies and time-dependent density functional theory simulations (TD-DFT), we show the formation of Insulin-Au(III) conjugates. The interaction of Au(III) ions with the aromatic rings of tyrosine induces charge transfer-like excitations in the visible range. Experimental investigations, together with molecular dynamics simulations of insulin and calculations of electronic properties in a model system, are performed to explore the origin of optical features and the structure-optical property relationship, leading the way to new concepts for nonlinear optics using protein-Au(III) conjugates.

Introduction

When gold salts are reacted with proteins at a basic pH, the reaction products exhibit large Stokes shift, red luminescence, and high-quantum yields.^{1–8} In particular, many applications using mainly bovine serum albumin (BSA) as protein templates and gold have been reported as diagnostic and therapeutic platforms for biomedical applications.^{9, 10}

Exemplified with BSA, two possible scenarios of this luminescence have been suggested: one scenario considers that BSA acts as a reacting and protecting “cage” in which gold ions are reduced by tyrosine residues in alkaline conditions and nucleate into a gold nanocluster, composed of dozen of gold atoms.¹¹ The other scenario asserts that the red luminophore formation involves Au(III)

coordination in a specific domain of serum albumin.^{12, 13} These observations and protein structural analyses with various proteins (bovine serum albumin, ovalbumin, trypsin, and insulin) show that red luminophore formation involves Au(III) coordination with a common motif within these proteins.¹⁴

Insulin is a small protein, or more precisely a polypeptide hormone comprising only 51 amino acids. Its function primarily lies in the regulation of insulin-responsive tissues and it is related to many diseases. Insulin has no free cysteine and only 4 tyrosine residues, so it could display unique binding motifs with gold and thus be a good candidate for producing red luminophores. Attempts to characterize prepared insulin-gold samples by mass spectrometry have proved unsuccessful, emphasizing the difficulty of exploring the structure-optical property relationship taking place during the interaction of proteins with gold.^{5, 15}

In this paper, we synthesize water-soluble insulin-gold conjugates. The goal of the current preparation was to develop highly luminescent gold-insulin conjugates. Although insulin functionalized nanoclusters were reported earlier, their preparation was time consuming whereas the current synthesis process is fast and completed within 4 hours. Moreover, apart from the new preparations, the mass spectrometric characterization¹⁶ and quantitative analysis of two-photon fluorescence properties¹⁷ are new findings which have also been correlated with theoretical calculations. In particular, our quantitative analysis of two-photon properties will be very attractive for two-photon bioimaging applications.

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Electronic Supplementary Information (ESI) available: [Experimental. synthesis and purification. XPS spectra. ESI-MS spectra. DLS and CD spectra and analysis. PAGE Gels. Optical spectra and ROS measurements. TD-DFT results]. See DOI: 10.1039/x0xx00000x



Results and discussion

Synthesis and characterization of Insulin-gold conjugate

Here, we synthesized highly luminescent insulin-gold conjugates using a one pot “green” synthesis method with a modification of protocol reported previously.⁵ The reaction was carried out at physiological temperature (37°C) at elevated pH (~ 11.5), as described in detail in Methods (see supporting information Figs. S1-S2). The pH of the insulin solution is in the range of pH~3.5-4.0. Insulin has less solubility in water, thus when we added insulin in Au³⁺ solution, the latter transformed into yellowish white aggregates. Quantitative XPS measurements were compatible with one gold and one insulin stoichiometry in gold-insulin conjugates (see Table S1 and Fig. S3). We also tried to use XPS to investigate the valence state of the gold atoms in gold-insulin conjugates. A doublet peak structure should be observed with fitted binding energies of ~87.4 and ~91.4 eV for Au(III) conjugates (respectively ^{4f}_{7/2} and ^{4f}_{5/2} doublet peaks). However, the ^{4f}_{5/2} peak of Au(0) was also located at 87.6 eV.¹⁸ The fact that the content of gold (0.3%) is very low and the overlap of 4f peaks between different oxidation states of gold failed to provide clear evidence of Au(III) by XPS, while mass spectrometry measurements led to an unambiguous assignment (see below). As already mentioned in the Introduction, early attempts to measure the mass of the as prepared insulin–Au NCs were either unsuccessful⁵ or only one gold atom attached to insulin was detected in mass spectra.¹⁵ We continued the mass characterization of Insulin-gold using high-resolution electrospray mass spectrometry (HR-ESI-MS). The full ESI-MS spectrum of the insulin-gold conjugates in SI is shown in Fig. S4, where charge states from 2+ to 4+ can be observed. For insulin, charge states n+ are given by extra protonations [Insu+nH]ⁿ⁺. The isotopic pattern of insulin (4+ charge state, [Insu+4H]⁴⁺) is reproduced using the following formula: C₂₅₇H₃₈₃N₆₅O₇₇S₆ (see insets in Fig. 1). When Au is complexed with insulin (4+ charge state), thanks to the isotopic distribution analysis, we observed that 3 hydrogen ions were removed from insulin. Then the isotopic pattern of Insulin is reproduced using the following formula: C₂₅₇H₃₈₀N₆₅O₇₇S₆Au (see insets in Fig. 1). The number of H atoms is reduced from 383 to 380 assigned to the 3 hydrogen ions removed from the insulin. At pH=11.5 used for the synthesis, phenolic groups of tyrosine are deprotonated. One can thus assume that deprotonations on insulin occur on tyrosine residues.

The interaction between insulin and gold has also been investigated using a circular dichroism (CD) spectropolarimeter. Figure S5 shows the CD spectra of pure insulin and insulin-Au conjugates. The pure insulin in neutral pH showed two negative absorption bands at ~211 and ~224 nm (in fair agreement with the reported CD spectrum of human insulin).¹⁹ When Au was added into insulin, only the first negative band at ~209 nm remained, and a broad negative band emerged at ~350 nm. The popular method for estimating protein secondary structure fractions from CD spectra (implemented in software packages CDSSTR)²⁰ was tested on insulin and insulin-Au conjugates. The results presented in Figure S6 indicate that the native structure of insulin is not much affected by the presence of gold (Table S2). Only the local alpha-helix environment is affected (where 3 of the 4 tyrosine residues are in helix motifs, see Fig. 3a). It is noteworthy that crystallization

tests were carried out, but no crystals were formed in the presence of gold.

DOI: 10.1039/D4NR01697A

Although the presence of one gold atom bound to insulin was demonstrated by HR-ESI-MS, we cannot rule out the formation of oligomers (as expected since insulin is known to easily form multimers). Indeed, in a previous study, we used a simple and reliable fluorescence-based technique to measure the hydrodynamic size of monolayer protected gold nanoclusters.^{8, 21} This technique uses the relation between the time resolved fluorescence anisotropy decay and the hydrodynamic volume. The comparison of sizes obtained between bare proteins and gold containing proteins allowed us to address the volume changes induced by the formation of oligomers for insulin during gold incorporation.⁸ Interestingly, Dixon and Egusa further evidenced the formation of oligomers (approximately between dimers and tetramers) for insulin during gold incorporation, by performing native gel electrophoresis of protein–Au(III) conjugates.¹⁴ We performed native gel electrophoresis on our as-synthesized insulin-gold conjugates, also evidencing the tendency to oligomerization during gold incorporation (see Fig. S7). Oligomer formation was further observed by mass spectrometry (both MALDI²² and ESI-MS, see Fig. S8). Interestingly, the VT-ESI source was used to fix the sample temperature at 17°C, thereby slowing down the aggregation.²³ In parallel, we performed DLS analysis on insulin and insulin-gold conjugates, see Fig. S9. As can be seen, dynamic light scattering shows that the mean sizes of a) insulin and b) insulin-Au conjugates are 3.8 nm and 7 nm, respectively. Thus, the presence of gold in the sample affects insulin oligomerization. To sum-up, the as-synthesized Insulin-gold conjugates are mainly composed by Insulin-Au(III) complex resulting in oligomers (ranging from monomers to trimers).

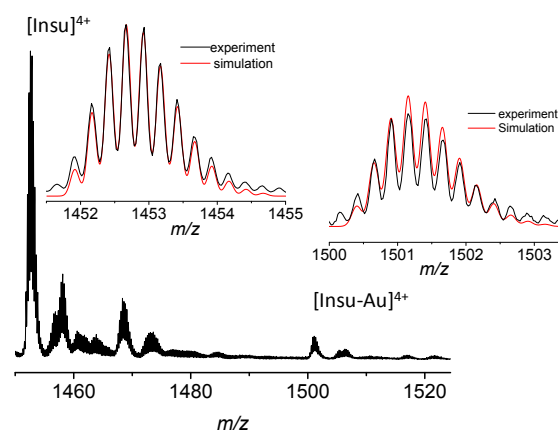


Figure 1. Zoom of the VT-nanoESI-MS spectrum of insulin-gold conjugate, corresponding to the 4+ charge state (Insu for Insulin). Insets. Isotopic pattern (black: experimental, and red: simulation) of Insulin (C₂₅₇H₃₈₃N₆₅O₇₇S₆) on left and of Insulin with Au(III) (C₂₅₇H₃₈₀N₆₅O₇₇S₆Au) on right. The other minor peaks are mostly alkali adducts.

Optical properties of Insulin-gold conjugate

Linear optical properties. We recorded the absorption and photoluminescence spectra of insulin-gold conjugates, as shown in



Figure S10. The as-synthesized insulin-Au conjugates show an absorption hump around 510 nm. The excitation spectrum exhibits a similar band corresponding to an absorption band at 510 nm which indicates that the PL band is excited by the HOMO–LUMO transition. We observed an intense emission peak at 667 nm. The emission spectra of Au-insulin only changed in intensity when the conjugates were irradiated at different wavelengths. The Stokes shift can reach 350 nm (see Fig. S11). We recorded the luminescence quantum yield (QYs) using the integrating sphere method and found it to be 5.40 %, with observed lifetimes fitted at 237 ns and 1723 ns (Figure S12).

Since insulin-gold conjugates exhibit a long photoluminescence lifetime, suggesting that they can be capable of singlet oxygen generation,^{24–28} as demonstrated for other protein template gold systems.^{26, 27} As a typical mechanism of singlet oxygen generation, after photoexcitation to an excited singlet state, the photosensitizer undergoes intersystem crossing to its triplet state, followed by energy transfer to ground-state (triplet) dioxygen. Generally, excited triplet states are relatively long-lived compared with excited singlet states because of the spin-forbidden nature of the $T_1 \rightarrow S_0$ transition. Here, the generation of singlet oxygen was triggered by the excitation of the conjugates with a cw laser emitting at 473 nm with different exposure times. With 1,3-Diphenylisobenzofuran (DPBF, an absorption probe for detection of 1O_2), 1O_2 generation can be monitored by a change in the absorption of DPBF at 412 nm. Absorption spectra and the change in the absorption of DPBF monitored over time are presented in Figure S12. The rate of 1O_2 generation was obtained by the initial DPBF concentration change over time ($\Delta[\text{DPBF}]/\Delta t$, at a shorter time where the slope has a linear behavior) divided by the concentration of the conjugates. To demonstrate the efficiency of insulin-Au for 1O_2 production, we compared 1O_2 production by insulin-Au to that of the conventional dye photosensitizer, new methylene blue (NMB). Under 473 nm irradiation, a high 1O_2 production efficiency was observed for Insulin-Au (1O_2 generation rate of 1.5 ± 0.3 compared with NMB of 2.3). The quantum yield of singlet oxygen generation allows normalization by the absorbance at 473 nm of the cw laser. Using a comparative method (as described in Methods and Figure S13), we were able to evaluate the 1O_2 generation quantum yield of insulin-Au conjugates at $13\% \pm 5\%$. However, we cannot exclude that the generation of oxidizing species (H_2O_2 , radicals) upon irradiation may also occur. To better address this point, we used a DCF-dye,²⁵ for probing the oxidizing agents produced following photoexcitation. We confirm that insulin-Au conjugates are prone to produce such oxidizing agents in addition to 1O_2 species. Figure S13c) displays the fluorescence spectra of DCF dye without and with 10 min laser irradiation. The strong increase in fluorescence intensity is a signature of generation of type I species.

Non-linear optical properties. We measured the two-photon excited luminescence (TPEL) spectra in a wide range of 800–1100 nm excitation wavelengths. Based on TPEL, we calculated two-photon absorption cross-sections (TPA) – σ_2 . The features observed in TPA cross sections as a function of wavelength are fairly comparable to the features observed in one-photon excitation spectra when plotted at doubled wavelength, for which resonance effects in a two-photon process may occur.^{29–31} Indeed, significant

resonance enhancement occurs when the excitation energy of a one-photon absorption state is close to half that of a TPA state (“double resonance” condition, see scheme S1). TPA cross-sections can be calculated by dividing the TPEL cross-sections with the one-photon luminescence QY (0.054) and the values range from 1.5 to 6 GM for excitation wavelengths in the range 800–1100 nm.

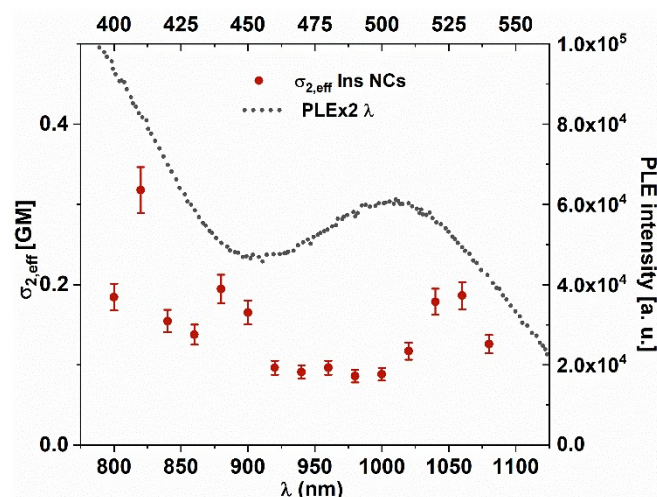


Figure 2. Two-photon brightness (TPEL) spectra of insulin-gold conjugate (red dots, left and bottom axes) correlated with the photoluminescence excitation spectrum of insulin-gold (black dotted line, right and upper axes).

Structure-optical property relationship. Insights from calculations

This detailed structural characterization, completed with circular dichroism measurements, leads us to suggest that deprotonations on insulin occur on tyrosine residues (that are naturally deprotonated at pH=11.5 used for synthesis). The interaction of Au(III) ions with the aromatic rings of tyrosine may induce charge transfer-like excitations in the visible range, as we demonstrated for tryptophan bound to gold cations.³² Clearly, at high pH, other chemical groups can be deprotonated (e.g. carboxylic group, thiols). We demonstrated in our previous works on peptides interacting with gold that complexes of gold with carboxylic residues or thiol residues lead to absorption bands in the near-UV range and not in the visible range.^{33, 34} Only deprotonated tyrosine residues interacting with gold can induce absorption in the visible range. This argumentation can thus indirectly support deprotonations of tyrosine in the insulin-gold conjugates.

To benchmark the calculations, we first explored the optical properties of neutral tyrosine and doubly deprotonated tyrosine. TD-DFT absorption spectra and the lowest energy structures of tyrosine are presented in Fig. S14 and compared with the experimental spectra of tyrosine recorded at neutral and highly basic pHs. In comparison to neutral tyrosine, the overall spectrum of the deprotonated tyrosine is redshifted. The qualitative agreement between experimental and theoretical spectra is present. TD-DFT computations revealed that the nature of the well-known $\pi\pi^*$ transition was preserved and that the bathochromism of the absorption spectrum could be explained by the energy shift of



both the π and the π^* orbitals under the influence of the nearby negatively charged group. A similar behavior had already been reported for tryptophan anions.³⁵ Then, as model systems, we explored the effect of different oxidation states of gold atoms (Au(0), Au(I) and Au(III)) when added to doubly deprotonated tyrosine. The complexation of deprotonated tyrosine with gold led to the emergence of excitations in the visible range with a strong charge-transfer character (see Fig. S15).

Insulin contains 4 tyrosines which can participate in the coordination of Au(III), their position in the structure is shown in Figure 3a). Tyrosine Y26 is located at the end of chain B and it is not part of the secondary structure elements of insulin. This allows for more flexibility and permits approaching other tyrosines (Y16, Y19) in the structure and coordinating the Au(III) atom, as demonstrated by molecular dynamics simulations (see Fig. 3a). Additionally, the absorption spectrum of 4 tyrosines with Au(III) (see Fig. S17) presents very similar features to those of 3 tyrosines with Au(III). This underlines the role of 3 tyrosines which coordinate Au(III). Indeed, the addition of Au(III) to doubly deprotonated tyrosine leads to the dissociation of tyrosine. However, Au(III) is strongly stabilized by the complexation with three tyrosines (see Fig. 3a). The corresponding TD-DFT absorption and analysis of the main excitations responsible for the characteristic absorption features confirm the presence of excitations in the visible range with a strong charge-transfer character, as displayed in Fig. 3b). Low-lying states in the visible range are key to obtaining large two-photon absorption cross sections, which would make them ideal labels for multiphoton excited luminescence (Fig. S16).

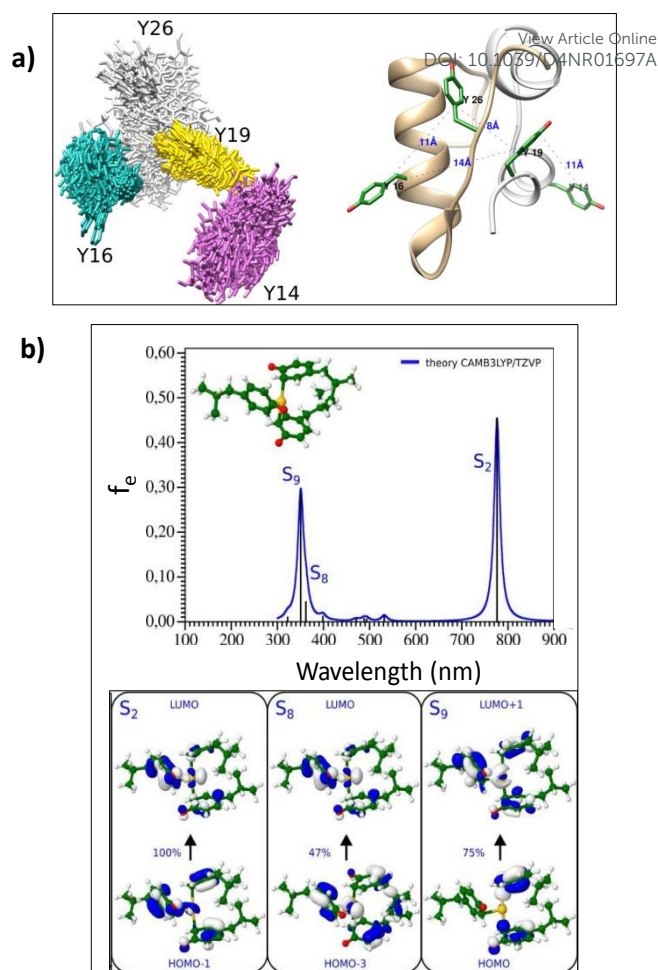


Figure 3. a) (left) Overlap of the snapshots from the molecular dynamics of insulin to check for possible tyrosine positions. The Y26 located near the end of the B chain was found to be the most flexible (silver stick representation). (right) The position of the tyrosine residues in insulin's crystal structure (pdb: 3i40, human insulin). Chains A and B are colored gray and beige, respectively. Tyrosine residues are shown in stick representation, and colored green. Blue colored numbers indicate the distance between tyrosine residues. b) (top panel) TD-DFT absorption spectra and lowest energy structure of 3 Tyrosines with Au(III) (both carboxylic and amino groups exchanged with methyl groups while phenolic groups are deprotonated). (bottom panel) Leading excitations responsible for the characteristic features of absorption.

Conclusions

We synthesized highly luminescent red-emitting insulin-gold conjugates using a one pot “green” synthesis method. The current synthesis process is fast and completed within 4 hours. The as-synthesized Insulin-gold conjugates were characterized by mass spectrometry and mainly composed of an Insulin-Au(III) conjugates resulting in oligomers. The main effect of oligomerization is a rigidification effect leading to the enhancement of luminescent properties, both in the linear and nonlinear optical regimes. They present long luminescence lifetimes enabling efficient reactive oxygen species generation that may create oxidative stress, pointing to potential antimicrobial applications.^{25, 36} Regarding other



bio-applications, it was demonstrated by Liu et al.⁵ that the blood glucose-lowering activities of humulin R and insulin–Au nanoclusters showed no significant difference, which implies that the as prepared insulin–Au NCs retained bioactivity in reducing blood glucose. We postulate that similar blood glucose-lowering activities would be reported with the present Insulin–Au(III) conjugates, since the native structure of insulin is not greatly affected by the presence of a single Au(III) in the conjugate. Detailed structural characterization allows us to suggest that deprotonations on insulin occur on tyrosine residues. The interaction of Au(III) ions with the aromatic rings of tyrosine induces charge transfer-like excitations³⁷ in the visible range. Deprotonated phenolic groups in tyrosine residues serve as anchors for stabilizing Au(III) ions. Model calculations can establish new concepts for nonlinear optics using protein–Au(III) conjugates. In particular, engineering mutations in proteins with tyrosines will allow more selective control of the number and the position of Au(III) ions in proteins, thereby opening new routes for the rational control of their photophysical properties. Finally, we point out the unique character of insulin having no free cysteine and only 4 tyrosine residues, thus it serves as an ideal template for coordinating Au(III). Other proteins have high numbers of tyrosine (and other aromatic acid groups) and free cysteines where the reduction of gold (Au(III)→Au(I)→Au(0)) enters into competition and where nanoclusters can be produced and anchored.

Conflicts of interest

There are no conflicts to declare.

Author contributions

R.A. conceived the initial idea and coordinated the work. D.B., H.Y. synthesized and prepared the nanoclusters. C.B., C.C.Z. and F.C. recorded and analyzed mass spectra. H.Y. conducted ROS experiments with nanoclusters. A.H. A.P. and D.B. conducted circular dichroism and nonlinear optical experiments, supervised by J.O.B. Z.S.M., and V.B.K. performed and analyzed the theoretical results. V.G.C. carried out crystallization tests and DLS measurements. R.A., D.B., and H.Y. wrote the paper. All authors provided critical feedback and helped to shape the final manuscript.

Acknowledgements

H.Y. is grateful for PhD fellowships donated by the China Scholarship Council (CSC). D.B. is grateful for post-doc fellowship donated by Agence Nationale de la Recherche (project MANBAMM, ANR-21-CE29-0020). C.B. is grateful for PhD fellowship donated by Agence Nationale de la Recherche (project nanoGOLD, ANR-22-CE29-0022). R.A. and H.Y. acknowledge Shanghai Science and Technology Innovation Program (22520712500) for support. R.A. and D.B. acknowledge Agence Nationale de la Recherche (project MANBAMM, ANR-21-CE29-0020) for support. R.A. and C.C.Z. acknowledge Agence Nationale de la Recherche (project MOONSTONE, ANR-22-CE42-0031) for support. R.A. C.B. and F.C. acknowledge Agence Nationale de la

Recherche (project nanoGOLD, ANR-22-CE29-0022) for support. R.A., D.B., H.Y., A.H., A.P., and J.O.B. acknowledge Agence Nationale de la Recherche (project POLONIUM (Project: 49252PM, “Gold nanoclusters in chiral nanoassemblies - nonlinear optical properties”) for support. VBK acknowledges STIM-REI project, Contract KK.01.1.1.01.0003. The authors would like to thank Carol Grossiord from Science et Surface (<http://www.science-et-surface.fr>) for XPS spectra. We acknowledge the contribution of SFR Biosciences (Universite Claude Bernard Lyon 1, CNRS UAR3444, Inserm US8, ENS de Lyon) Protein Science Facility.

Notes and references

1. N. Goswami, K. Zheng and J. Xie, *Nanoscale*, 2014, **6**, 13328-13347.
2. R. Antoine, D. Maysinger, L. Sancey and V. Bonačić-Koutecký, *Commun. Chem.*, 2022, **5**, 47.
3. N. Alkudaisi, B. A. Russell, B. Jachimska, D. J. S. Birch and Y. Chen, *J. Mat. Chem. B*, 2019, **7**, 1167-1175.
4. F. Wen, Y. Dong, L. Feng, S. Wang, S. Zhang and X. Zhang, *Anal. Chem.*, 2011, **83**, 1193-1196.
5. C.-L. Liu, H.-T. Wu, Y.-H. Hsiao, C.-W. Lai, C.-W. Shih, Y.-K. Peng, K.-C. Tang, H.-W. Chang, Y.-C. Chien, J.-K. Hsiao, J.-T. Cheng and P.-T. Chou, *Angew. Chem. Int. Ed.*, 2011, **50**, 7056-7060.
6. X. Xia, Y. Long and J. Wang, *Anal. Chim. Acta*, 2013, **772**, 81-86.
7. H. Shi, M. Y. Ou, J. P. Cao and G. F. Chen, *RSC Adv.*, 2015, **5**, 86740-86745.
8. A. Soleilhac, F. Bertorelle and R. Antoine, *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, 2018, **193**, 283-288.
9. I. Zare, D. M. Chevrier, A. Cifuentes-Rius, N. Moradi, Y. Xianyu, S. Ghosh, L. Trapiella-Alfonso, Y. Tian, A. Shourangiz-Haghighi, S. Mukherjee, K. Fan and M. R. Hamblin, *Mater. Today*, 2023, **66**, 159-193.
10. V. Bonačić-Koutecký, X. Le Guével and R. Antoine, *ChemBioChem*, 2023, **24**, e202200524.
11. J. Xie, Y. Zheng and J. Y. Ying, *J. Am. Chem. Soc.*, 2009, **131**, 888-889.
12. J. M. Dixon, J. Tomida and S. Egusa, *J. Phys. Chem. Lett.*, 2020, **11**, 3345-3349.
13. J. M. Dixon and S. Egusa, *J. Am. Chem. Soc.*, 2018, **140**, 2265-2271.
14. J. M. Dixon and S. Egusa, *J. Phys. Chem. Lett.*, 2021, **12**, 2865-2870.
15. M. Shamsipur, E. Babaee, M.-B. Gholivand, F. Molaabasi, F. Mousavi, A. Barati, B. Hajipour Verdom, B. Shojaedin-Givi and H. Naderi-Manesh, *ACS Appl. Nano Mater.*, 2023, **6**, 5939-5951.
16. C. Comby-Zerbino, X. Dagany, F. Chiro, P. Dugourd and R. Antoine, *Mater. Adv.*, 2021, **2**, 4896-4913.
17. F. Bertorelle, C. Moulin, A. Soleilhac, C. Comby-Zerbino, P. Dugourd, I. Russier-Antoine, P.-F. Brevet and R. Antoine, *ChemPhysChem*, 2018, **19**, 165-168.
18. S. Müllegger, W. Schöffberger, M. Rashidi, T. Lengauer, F. Klappenberger, K. Diller, K. Kara, J. V. Barth, E. Rauls, W. G. Schmidt and R. Koch, *ACS Nano*, 2011, **5**, 6480-6486.
19. S. G. Melberg and W. C. Johnson Jr., *Proteins: Struct., Funct., Bioinf.* 1990, **8**, 280-286.



20. L. Whitmore and B. A. Wallace, *Nucleic Acids Res.*, 2004, **32**, W668-W673.
21. A. Soleilhac, F. Bertorelle, C. Comby-Zerbino, F. Chiro, N. Calin, P. Dugourd and R. Antoine, *J. Phys. Chem. C*, 2017, **121**, 27733-27740.
22. H. Yuan, D. Lima, C. Comby-Zerbino, C. Bouanchaud, F. Chiro, D. Bain, S. Zhang and R. Antoine, *Int. J. Mass Spectrom.*, 2024, **503**, 117276.
23. W. Mohammad, K. D. Wegner, C. Comby-Zerbino, V. Trouillet, M. P. Ogayar, J.-I. Coll, R. Marin, D. J. Garcia, U. Resch-Genger, R. Antoine and X. Le Guével, *J. Mater. Chem. C*, 2023, **11**, 14714-14724.
24. H. Fakhouri, M. P. Bakulić, I. Zhang, H. Yuan, D. Bain, F. Rondepierre, P.-F. Brevet, Ž. S. Maršić, R. Antoine, V. Bonačić-Koutecký and D. Maysinger, *Commun. Chem.*, 2023, **6**, 97.
25. H. Yuan, M. Theresa, D. Bain, H. Fakhouri, K. Sreekanth, A. Ravi, S. Thomas, N. Kalarikkal, R. Antoine and E. K. Radhakrishnan, *Inorg. Chem. Commun.*, 2024, **159**, 111799.
26. V. Poderys, G. Jarockyte, S. Bagdonas, V. Karabanovas and R. Rotomskis, *J. Photochem. Photobiol. B*, 2020, **204**, 111802.
27. M. Yamamoto, I. Osaka, K. Yamashita, H. Hasegawa, R. Arakawa and H. Kawasaki, *J. Lumin.*, 2016, **180**, 315-320.
28. H. Kawasaki, S. Kumar, G. Li, C. Zeng, D. R. Kauffman, J. Yoshimoto, Y. Iwasaki and R. Jin, *Chem. Mater.*, 2014, **26**, 2777-2788.
29. Z. Hu and L. Jensen, *Chem.Sci.*, 2017, **8**, 4595-4601.
30. I. Russier-Antoine, F. Bertorelle, N. Calin, Ž. Sanader, M. Krstić, C. Comby-Zerbino, P. Dugourd, P.-F. Brevet, V. Bonačić-Koutecký and R. Antoine, *Nanoscale*, 2017, **9**, 1221-1228.
31. Ž. Sanader, M. Krstić, I. Russier-Antoine, F. Bertorelle, P. Dugourd, P.-F. Brevet, R. Antoine and V. Bonačić-Koutecký, *Phys.Chem.Chem.Phys.*, 2016, **18**, 12404-12408.
32. R. Antoine, F. Bertorelle, M. Broyer, I. Compagnon, P. Dugourd, A. Kulesza, R. Mitrić and V. Bonačić-Koutecký, *Angew. Chem. Int. Ed.*, 2009, **48**, 7829-7832.
33. B. Bellina, I. Compagnon, F. Bertorelle, M. Broyer, R. Antoine, P. Dugourd, L. Gell, A. Kulesza, R. Mitric and V. Bonacic-Koutecky, *J. Phys.Chem.C*, 2011, **115**, 24549-24554.
34. I. Russier-Antoine, F. Bertorelle, A. Kulesza, A. Soleilhac, A. Bensalah-Ledoux, S. Guy, P. Dugourd, P.-F. Brevet and R. Antoine, *Prog. Nat. Sci.: Mater. Int.*, 2016, **26**, 455-460.
35. I. Compagnon, A.-R. Allouche, F. Bertorelle, R. Antoine and P. Dugourd, *Phys.Chem.Chem.Phys.*, 2010, **12**, 3399-3403.
36. S. Chakraborty, P. Sagarika, S. Rai, C. Sahi and S. Mukherjee, *ACS Appl. Mat. Interfaces*, 2021, **13**, 36938-36947.
37. R. Antoine, T. Tabarin, M. Broyer, P. Dugourd, R. Mitrić and V. Bonačić-Koutecký, *ChemPhysChem*, 2006, **7**, 524-528.

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DOI: 10.1039/D4NR01697A



Data availability

View Article Online
DOI: 10.1039/D4NR01697A

The data supporting the findings of this study are available within the article and its ESI files.† Additional data that support the findings of this study are available from the corresponding author upon reasonable request. This includes computational codes and raw data files generated during the study.

