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Medicinal chemistry inspired by ginger: exploring the chemical space around 6-gingerol

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Ginger (*Zingiber officinale* Roscoe) has been used as a spice and as a traditional remedy since ancient times, especially in traditional Chinese medicine. It has been applied as a treatment for many diseases either alone or in combination with other remedies. Many studies were conducted on ginger and its constituents and a wide array of bioactivities were reported, e.g., antioxidant, anti-inflammatory, antiemetic, and anticancer activity. Most of these had been correlated to gingerols and shogaols, the most abundant secondary metabolites in ginger. This inspired several research groups to explore the biomedical value of the chemical space around these compounds, and many of their synthetic or semi-synthetic analogues have been prepared and studied for various bioactivities. Thanks to this, many valuable structure activity relationships have been revealed for such compounds. Herein, we provide a brief summary on the synthetic derivatization efforts that had so far been implemented on 6-gingerol, the main constituent of fresh ginger. This review covers 160 natural, semisynthetic, or synthetic 6-gingerol derivatives and their reported bioactivities.

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Introduction

Ginger, *Zingiber officinale* Roscoe (Zingiberaceae) is widely renowned and has been historically used for culinary and medicinal purposes.^{1,2} The roots of this plant have been used as a herbal remedy for the prevention and/or treatment of nausea and vomiting, diarrhea, abdominal discomfort, headache, rheumatism and respiratory illnesses (e.g. common cold),² and have also been studied for a wide range of bioactivities including anti-inflammatory, antioxidant,³ antimicrobial,⁴ antidiabetic,⁵ antihypertensive, cardioprotective,⁶ neuroprotective,⁷ anti-obesity,⁸ anti-migraine,⁹ and anticancer effects.^{2,10} Most recently, ginger gained a significant public attention when it was also suggested as a natural home remedy that may help against COVID-19,^{11–13} and even though at this time this is by no means an evidence-based use,¹⁴ the anti-inflammatory effect of ginger may hold some promise in the relief of respiratory symptoms connected to the SARS-CoV-2 infection.¹⁵

A wide array of bioactive compounds have been identified in ginger such as phenolic compounds and terpenes; these have recently been reviewed.² Among these constituents, the so-called gingerols are present in by far the most significant amount in ginger (23–25%), and this is accompanied by relatively lower levels

of other, related compounds such as shogaols and paradols.^{10,16,17} The main compound in the gingerol series, 6-gingerol (Fig. 1) is partially responsible for the strong pungent taste of ginger. This compound has been correlated with many bioactivities of ginger, and it is present in much higher amounts in fresh ginger roots compared to the dried roots because drying converts it into 6-shogaol through a water elimination.^{16,18}

Much research has been devoted to the biomedical value of 6-gingerol and its semisynthetic derivatives, and several clinical trials had been performed using ginger extract and its constituents, and some of these are still in progress. The high interest in ginger is shown well by the fact that searching the term “ginger” in Scopus gives over a thousand hits only for the year 2020. There are many recent reviews on ginger constituents and their potential therapeutic applications.^{2,10,16,18,19} However, to the best of our knowledge, currently no reviews are available on the semi- and total-synthetic efforts to explore the biomedical value of the chemical space around 6-gingerol, i.e. the medicinal chemistry inspired by this compound. Therefore, the aim of this paper is to provide such a coverage with a hope that it may draw a roadmap for further possible structural manipulations

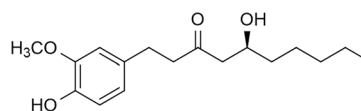


Fig. 1 The structure of 6-gingerol, i.e. 5-hydroxy-1-(4-hydroxy-3-methoxyphenyl)decan-3-one.

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of 6-gingerol towards new promising lead compounds based on this simple but versatile bioactive natural product.

Review scope and coverage

The information included in this review is based on papers collected from a search in four online databases: PubMed, Scopus, Embase and SciFinder, with no time limits, while limiting the language to English and the search domain to title, abstract and keywords. The search strategy included different combinations of the following keywords: (“Ginger” OR “gingerol”) AND (“semisynthetic derivative(s)” OR “synthetic derivative(s)” OR “semisynthetic analog(s)” OR “synthetic analog(s)” OR “semisynthetic analogue(s)” OR “synthetic analogue(s)” OR “analogs” OR “derivatives” OR “semisynthetic” OR “synthetic” OR “synth*”). The wild-card term “*” was used to increase the sensitivity of the search. The search in SciFinder database was conducted as a sub-structure search using 6-gingerol structure to access publications reporting related synthetic or semi-synthetic work. The hits from the four databases together with the Cochrane Library returned no review papers in the subject of the current review.

This paper aims to provide an as complete as possible coverage of reports dealing with semisynthetic derivatives prepared from 6-gingerol regardless of its origin (*i.e.* isolated from ginger roots or prepared by total synthesis). Some diarylheptanoids are included if they were not the focus of the referred publication, *e.g.* if they are mentioned as compounds synthesized together with other gingerol derivatives. The same applies for derivatives of 8- and 10-gingerol, shogaol, zingerone, and other ginger constituents.

When presenting chemical structures in the following sections, the therapeutic aim was taken as the primary organizing principle. In the descriptive text, semi- and total-synthetic gingerol analogues are presented after a brief overview on the current knowledge about ginger and ginger constituents, particularly 6-gingerol, in relation to the targeted bioactivities.

Anticancer analogues

A growing body of research suggests the possibility for the use of ginger in cancer prevention and treatment. Ginger extract was claimed to have promising activity against various types of cancer (*e.g.* oral squamous cell carcinoma,²⁰ chronic myeloid leukemia,²¹ lung cancer²² *etc.*). It was also studied for reducing chemotherapy-induced nausea and vomiting.^{23–25}

A number of biochemical pathways were implied in the possible anticancer activity of ginger and its constituents.^{16,26} 6-Gingerol was reported to induce cell cycle arrest and exert anti-invasive and apoptosis promoting effects through acting on multiple signaling pathways in different types of cancer cell lines.^{27–29} This seems to be at least partially connected to the antioxidant–prooxidant properties of 6-gingerol. This compound was reported to induce reactive oxygen species (ROS) generation leading to DNA damage in cancer cells.¹⁸ It is also of interest that gingerol was found to have antioxidant and

chemopreventive activity through modulating nuclear factor erythroid 2-related factor 2 (Nrf2);^{30,31} this transcription factor is considered as a master switch in cellular antioxidant defense and redox signaling, and has implications as a potential anti-tumor target.^{32–34} Unsurprisingly, the anticancer effect of gingerol appears to be the result of a multitarget action. According to a most recent review on this subject, transcription factors (NF- κ B, activator protein-1; AP-1), β -catenin, mitogen activated protein kinases (MAPK), growth factor receptors (EGFR, VEGFR) and pro-inflammatory mediators (COX-2, TNF α) were reported to contribute to the anticancer activity exerted by this compound.³⁵

A series of clinical trials were conducted to evaluate the possible efficacy of a 50% aqueous ethanol extract of ginger roots (normalized to 5% of total gingerols) in preventing colorectal cancer (CRC). Results from a pilot, randomized controlled trial in patients with high risk of developing colorectal cancer suggested that this extract may increase apoptosis and differentiation and reduce proliferation of normal-appearing colon mucosa cells.³⁶ Increased eicosanoid, and mainly prostaglandin E2 (PGE2) level is a marker of early stages of CRC development. Consumption of ginger root extract was found to decrease cyclooxygenase-1 (COX-1) expression and consequentially lower PGE2 levels in people with increased risk of developing colorectal cancer (CRC) but not in participants with normal risk,³⁷ and similar results were found in a phase II clinical study on the PGE2 levels in the colon mucosa of healthy people at normal risk for developing CRC.³⁸ The PGE2-decreasing activity was later not confirmed in volunteers with increased CRC risk, which certainly does not rule out chemopreventive action through other mechanisms.³⁹

The apparent antitumor potential of ginger inspired several research groups to take 6-gingerol as a lead compound aiming at various bioactivities with a special emphasis on cancer. Based on our literature survey, 160 compounds have been synthesized, some of which are naturally present in ginger; these compounds are discussed hereinafter.

Gingerol derivatives with antiproliferative and/or cytotoxic activity

De Lima Silva *et al.* recently reported the synthesis of 6-gingerol derivatives, its *O*-propargyl ether (1) and several compounds where this moiety was transformed to a 1,2,3-triazol linker to a second phenolic ring (2–8). The compounds were tested on colon adenocarcinoma (HCT-116) and breast metastatic adenocarcinoma (MCF-7) cell lines. The propargyl substitution (1) did not influence the effect of 6-gingerol on cell viability, however, most of the triazole derivatives exerted somewhat increased activity that was the strongest in case of the *meta*-bromine-substituted compound 8. This was still moderate, *i.e.* ca. 87% and 71% inhibition in MCF-7 and HCT-116 cell lines, respectively, at 50 μ M concentration, while these values were 37% and 36% for 6-gingerol. The cells were much more sensitive to the positive control doxorubicin (IC₅₀ = 0.5 and 1.9 μ M against HCT-116 and MCF-7 cells, respectively).⁴⁰



Another study introduced different changes in the skeleton of 6-gingerol (compounds **9–14**, **17** *i.e.* 6-shogaol, **19**, and **22–24**), with the aim of understanding the structure activity relationships (SAR) concerning cytotoxicity of these compounds against MCF-7 breast cancer cells. Among these compounds, only a 4-allyloxy derivative (**10**) showed higher inhibitory activity against MCF-7 cells ($IC_{50} = 21 \mu\text{M}$) as compared to 6-gingerol ($IC_{50} = 30.3 \mu\text{M}$). While these values seem to indicate moderate activity, the same experimental setup resulted in unusually high IC_{50} values for the positive controls doxorubicin and 5-fluorouracil ($IC_{50} = 120 \mu\text{M}$ and $158.5 \mu\text{M}$, respectively, after 72 h incubation), which may suggest the involvement of an unknown resistance mechanism in the cells line used. Through comparing different substitution patterns, the authors claimed that the aromatic ring and a free hydroxyl group on the aliphatic side chain are important for the activity against breast cancer. It was also noted that the length of the alkyl side chain is optimal for this activity. Dehydrated products of 6-gingerol and its analogue **11** (compounds **17** and **14**, respectively) showed lower inhibitory activity compared to 6-gingerol. Further, a surprising opposite activity was found for the dimerization product of 6-gingerol (**24**) that exerted a concentration-dependent increase in cell viability.⁴¹

Another semi-synthetic effort yielded compound **25** that was cytotoxic on triple negative breast cancer (TNBC) cell line MDA-MB-231 ($IC_{50} = 22.9 \mu\text{M}$ after 48 h incubation), whereas 6-gingerol was technically inactive ($IC_{50} = 404.5 \mu\text{M}$). The mechanism of action was postulated to be the induction of early autophagy, in addition to a significant increase in ROS levels leading to caspase-independent cellular death at later periods of the incubation. When comparing the cytotoxicity of **25** on MDA-MB-231 to that on a non-tumorigenic epithelial cell line MCF-10A, a mild selectivity was found ($IC_{50} = 26.13$ and $40.46 \mu\text{M}$, respectively, after 24 h incubation). Further, compound **25** also inhibited migration and invasion of TNBC cells, caused cell cycle arrest at the G1-phase, and promoted apoptosis.^{42,43}

An interesting hybrid molecule of 6-gingerol and acetylsalicylic acid (**46**) was synthesized by Zhu *et al.* with an aim to combine the chemo-preventive and gastroprotective effect of the former with the anti-inflammatory activity of the latter, and to simultaneously counteract the well-known gastric irritative action of Aspirin. *In vitro*, compound **46** showed superior activity as compared to the two compounds alone or in combination, and it exerted protective effect against acute gastric ulceration in mice, suggesting that the hybrid could potentially be used as a multitarget chemo-preventive agent against gastrointestinal malignancies.⁴⁴

Gingerol derivatives with LTA4H inhibitory activity

Leukotriene A4 hydrolase (LTA4H), a bifunctional metalloenzyme with aminopeptidase and epoxide hydrolase activities, plays an important role in chronic inflammation associated with carcinogenesis.^{45,46} Badria *et al.* reported the synthesis and LTA4H inhibitory activity of natural and semi-synthetic gingerol derivatives (**9–17**, and **19–24**). Compound **11**, a prenylated gingerol derivative, demonstrated the strongest

activity against both functions of the enzyme (aminopeptidase: $IC_{50} = 3.0 \mu\text{M}$, epoxide hydrolase: $IC_{50} = 7.3 \mu\text{M}$) followed by methylshogaol (**16**; aminopeptidase: $IC_{50} = 4.9 \mu\text{M}$, epoxide hydrolase: $IC_{50} = 11.3 \mu\text{M}$).⁴⁷ Concerning the cytotoxicity of these compounds against HCT-116 colorectal cancer cells, compound **16** was the most effective ($IC_{50} = 1.5 \mu\text{M}$), much more potent than the positive controls used in the study (betastin and 4-(4-benzylphenyl)-thiazol-2-amine (4BSA) with $IC_{50} = 42.5$ and $30.5 \mu\text{M}$, respectively). 6-Shogaol (**17**) itself was less potent ($IC_{50} = 12.9 \mu\text{M}$) than compound **16**, *i.e.* the *ortho*-dimethoxy group in the aromatic ring increased the cytotoxic activity by nearly an order of magnitude. The same increase in efficacy was, however, not observed when 6-gingerol was similarly methylated to compound **19** ($IC_{50} = 76.5 \mu\text{M}$), which highlights the importance of the enone group in the side chain for this bioactivity. The compounds showed no toxicity to normal cells, and several of them had high selectivity towards HCT-116 cells as compared to normal TIG-1 cells: the selectivity index was over 52 in case of the most potent compound **16**.⁴⁷

Gingerol derivatives with antioxidant and/or HDAC inhibitory activity

In the last two decades, histone deacetylase (HDAC) enzymes have been emerging as anticancer targets.⁴⁸ In 2019, A combination of tucidinostat, an HDAC enzyme inhibitor, and exemestane, an aromatase enzyme inhibitor was studied in a randomized, double-blind, placebo-controlled, phase III trial on postmenopausal patients with advanced hormone receptor-positive breast cancer, and encouraging results were reported.⁴⁹ A link between the HDAC inhibitory activity and ROS levels had been highlighted. A suggested mechanism for HDAC inhibitors in cancer is through increasing intracellular ROS levels and downregulating antioxidant pathways resulting in increased level of DNA damage,⁵⁰ and the ability to repair this damage is impaired in cancer cells.⁵¹ In connection to this, it is of interest that many plant-originated phenolic antioxidants have been reported as HDAC inhibitors.^{52–54}

The anticancer activity of natural and semi-synthetic 6-gingerol derivatives (**9**, **17**, **18** and **26–29**, (3*R*,5*S*)-**30** and **31–39**), was studied through assessment of their HDAC enzyme inhibition and antioxidant activity by Kunboonma *et al.* All these compounds showed HDAC inhibitory activity in the micromolar concentration range, and compound **29** was the most active among all ($IC_{50} = 42 \mu\text{M}$). Compound **18**, a demethylated 6-shogaol derivative was the most potent semi-synthetic compound ($IC_{50} = 45 \mu\text{M}$; compared to $61 \mu\text{M}$ for gingerol), a role of the catechol moiety was suggested for the increase in activity as compared with that of 6-shogaol itself. In case of compounds **31–39**, the oxime orientation did not influence the bioactivity. When testing the compounds by the DPPH (2,2-diphenyl-1-picryl-hydrazyl-hydrate) scavenging assay, most of the investigated derivatives (except compounds **28**, (3*R*,5*S*)-**30**, and **38**), showed higher antioxidant activity compared to 6-gingerol ($IC_{50} = 81 \mu\text{M}$) but lower than the applied positive control, gallic acid ($IC_{50} = 37 \mu\text{M}$), and **35** and **38** were the most active ($IC_{50} = 42 \mu\text{M}$). Among the tested compounds, **29** ($IC_{50} =$



58 μM) was reported to exert the highest antioxidant activity. Furthermore, based on *in silico* docking studies it was suggested that compounds **17**, **18**, and **29** may serve as promising anti-HDAC leads with different isoform selectivity. Nevertheless, the reported activities were still moderate and the compounds would require further structural optimization to exert pharmacologically relevant HDAC inhibitory effect.⁵⁵

Influence of the length and structure of the side chain on the antioxidant properties of 6-gingerol was studied on its analogues **17** and **40–45**. Four experimental models were used including DPPH scavenging, ferric reducing antioxidant power (FRAP), DNA strand breakage inhibition and human red blood cell haemolysis protection. Regarding DPPH scavenging activity, shogaols were found the most effective, followed by gingerols, while dehydrogingerols and dehydroshogaols were the least effective; therefore, the C4–5 double bond may have a role in boosting the activity. Increasing the side chain length had no remarkable effect on the DPPH scavenging activity, however in case of FRAP measurements it had a negative effect on the potency. Increasing the side chain length significantly decreased the DNA strand breakage ability, while enhanced the anti-haemolysis activity. Thus, it was concluded that the antioxidant activity largely depends on the side chain.⁵⁶

Altogether, concerning the antitumor potential of gingerol derivatives, the currently known compounds are not very cytotoxic, still, they seem to have antitumor potential due to their abilities to interfere with several pathways relevant to antitumor drug discovery. This concerns mainly the chemo-preventive potential of the semi-synthetic derivatives, similarly to the inspiring compound 6-gingerol itself. Nevertheless, the potent and selective cytotoxic activity of compound **16** against colon cancer cells suggests that related analogues may also be developed with a potential to fight the already developed disease.

Anti-inflammatory gingerol analogues

The anti-inflammatory effect of ginger is among its most deeply studied medicinal properties. The effects of ginger were studied against many inflammatory conditions in human subjects, including knee osteoarthritis^{57,58} joint pain,⁵⁹ and obesity as a risk for cardiovascular complications.⁶⁰ The anti-inflammatory effect of ginger supplementation was studied on subjects who are free of any inflammatory conditions and at varying degree of exercise levels (NCT03219463). After ginger consumption a drop in the levels of inflammatory biomarkers tumor necrosis factor α (TNF- α) and interleukin-6 (IL-6) was reported. Therefore, it had been concluded that ginger may serve as a potential adjuvant to prevent diabetes, heart diseases and other chronic disorders connected to inflammation. Ginger consumption was also reported to reduce inflammatory mediators C-reactive protein (CRP) and interleukin-1 β (IL-1 β) in patients with rheumatoid arthritis (RA),⁶¹ and 6-gingerol was reported to inhibit sepsis development through interfering with pro-inflammatory cytokines' secretion and attenuating pyroptosis in macrophages,⁶² *i.e.*, an inflammatory-type programmed cell death.⁶³

These findings on ginger inspired the synthesis of several 6-gingerol analogues aiming to develop new anti-inflammatory agents. A set of natural and semi-synthetic ginger constituents, *i.e.*, compounds **17**, **20**, **21**, **42** (Fig. 2), and compounds **47–58** (Fig. 3) were studied for their cyclooxygenase-2 (COX-2) inhibitory activity in intact A549 cells that are known to express this enzyme. Compound **56** was found to be the most active ($\text{IC}_{50} = 1.4 \mu\text{M}$) among these compounds, followed by compounds **17**, **58**, **47**, 8-paradol (**21**), 10-gingerol (**57**), and **49** that was still active with an IC_{50} value of $5.5 \mu\text{M}$. All other compounds showed moderate activity, and 6-gingerol itself did not reach 50% inhibition at up to $50 \mu\text{M}$. Concerning SAR, an aromatic group substituted with a free hydroxyl group at C3 or C4 was found important for a potent COX-2 inhibition: compound **53** with the hydroxyl group at C2 of the aromatic ring and compound **50** with methoxy-substituents at both C3 and C4 showed only moderate activity ($\text{IC}_{50} > 50 \mu\text{M}$). Also, importance of the length of the alkyl chain was highlighted, and a 14-C length was suggested as the optimum. A significant increase in the activity was observed with a hydroxyl group on the alkyl chain, while replacement of the carbonyl group with a hydroxyl group had no remarkable effect on the potency of compounds, as in, *e.g.*, compound **54** ($\text{IC}_{50} = 12.5 \mu\text{M}$) compared with compound **20**, *i.e.*, 8-gingerol ($\text{IC}_{50} = 10 \mu\text{M}$). Nevertheless, reduction of the carbonyl group to a methylene boosted the activity as in compound **56** ($\text{IC}_{50} = 1.4 \mu\text{M}$).⁶⁴

Anti-inflammatory activity of two racemic gingerol derivatives, a stable metabolite of 6-gingerol (**58**), and another derivative named Capsarol (**49**), joining some molecular properties of gingerol and capsaicin, were evaluated as possible anti-inflammatory agents by Aktan *et al.* The compounds were tested for their effect on inducible nitric oxide synthase (iNOS) and found to suppress NO production in murine macrophages through a partial inhibition of the enzyme and by simultaneously decreasing iNOS expression through its NF- κ B-mediated transcriptional regulation.⁶⁵

As seen from the above examples, the anti-inflammatory activity of 6-gingerol analogues seems to be encouraging to further studies that may have a chance for the development of a suitable anti-inflammatory drug candidate in the future.

Antimicrobial gingerol analogues

The potential antibacterial activity of ginger was also assessed. Ginger extract showed activity against dental caries-causing bacteria; *Streptococcus mutans* and *S. sanguinis* with minimum inhibitory concentration (MIC) values of 20 and $300 \mu\text{g ml}^{-1}$, respectively, and it was suggested that this activity may be attributed to gingerols.⁶⁶ In another, *in vitro* and *in vivo* study on the effect of ginger on *S. mutans* bacteria, ginger extract inhibited bacterial growth with $\text{MIC} = 256 \mu\text{g ml}^{-1}$; differences in MIC values might be attributed to different experimental conditions and/or extraction method. Glucan synthesis, bacterial adhesion, and biofilm formation were also reduced *in vitro*, and a reduction of caries development was observed in ginger-treated rats *versus* the control group.⁶⁷



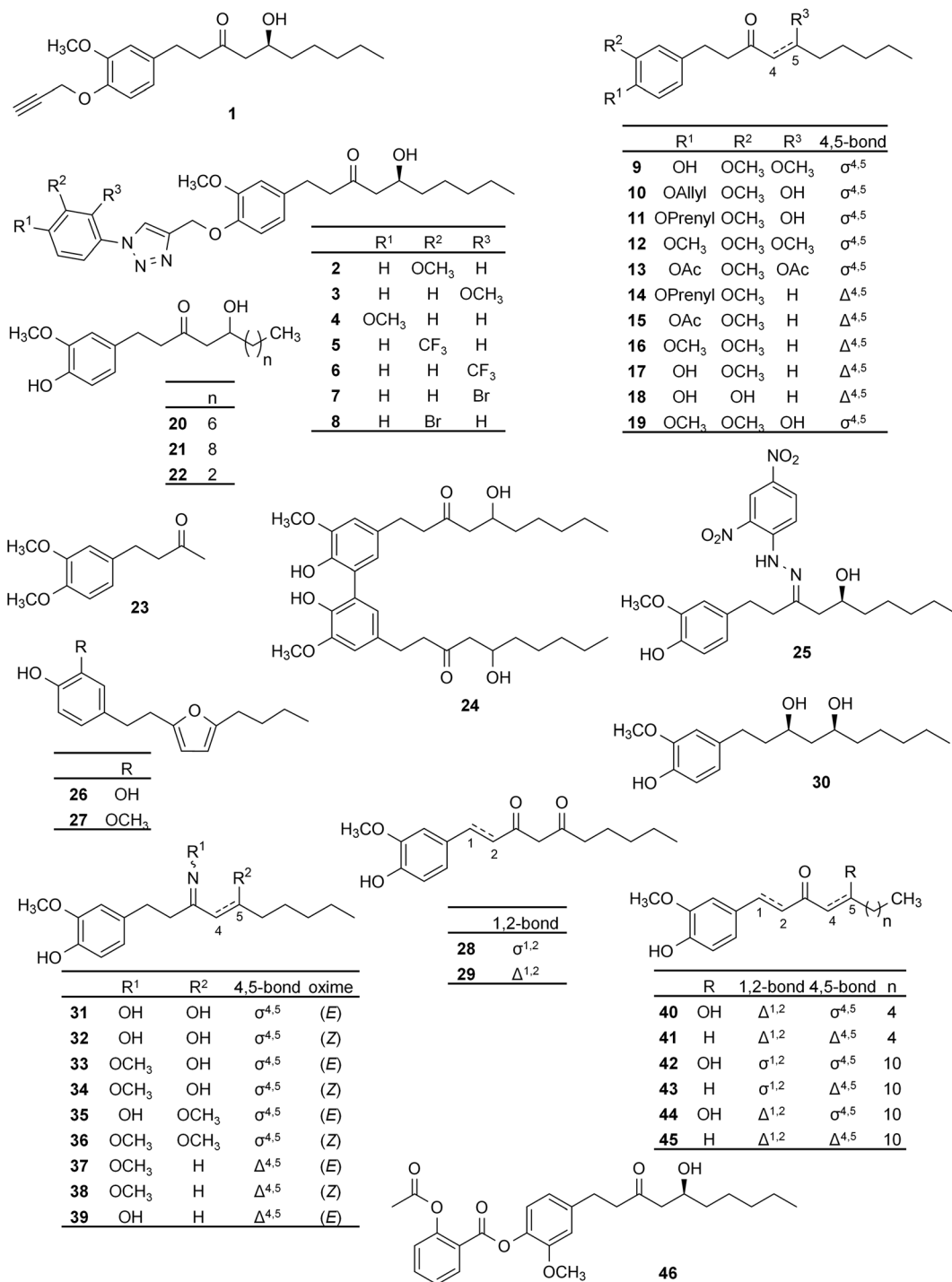


Fig. 2 Semi-synthetic gingerol derivatives prepared and directly or indirectly evaluated for their *in vitro* antitumor potential.

Components of ginger were investigated for anti-virulence and antibiofilm activities against a fluconazole resistant *Candida albicans* strain. It was reported that 6-gingerol, 8-gingerol and 6-shogaol effectively inhibited biofilm formation. Furthermore, 6-gingerol and 6-shogaol also reduced virulence of the fungus.⁶⁸

Gingerol analogues including 6-gingerol and 6-shogaol were also studied for their antibacterial activity against a range of

multi-drug resistant bacteria. Plasmid conjugal transfer property was also assessed. It was concluded that the investigated compounds are valuable antibacterial agents with an ability to reverse horizontal antibiotic resistance spread in bacteria.⁶⁹

In combination with tea polyphenols, 6-gingerol was also reported to maintain the quality of shrimp paste during storage, and the reduction of bacterial growth was a suggested mechanism for this effect.⁷⁰



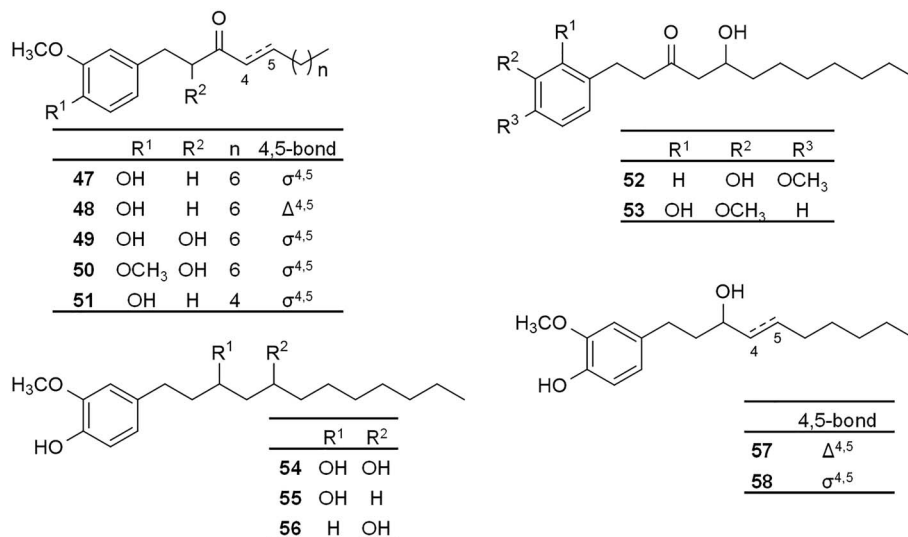


Fig. 3 Semi-synthetic gingerol derivatives prepared and evaluated for their *in vitro* anti-inflammatory activity.

6-Gingerol was found to reduce virulence and biofilm formation of *Pseudomonas aeruginosa* via the inhibition of quorum sensing (QS),⁷¹ i.e., a mechanism of bacterial cell to cell

communication that is of crucial importance in controlling their colony-wide functions and plays an important role in their virulence.⁷² In further research into the bioactivities of ginger

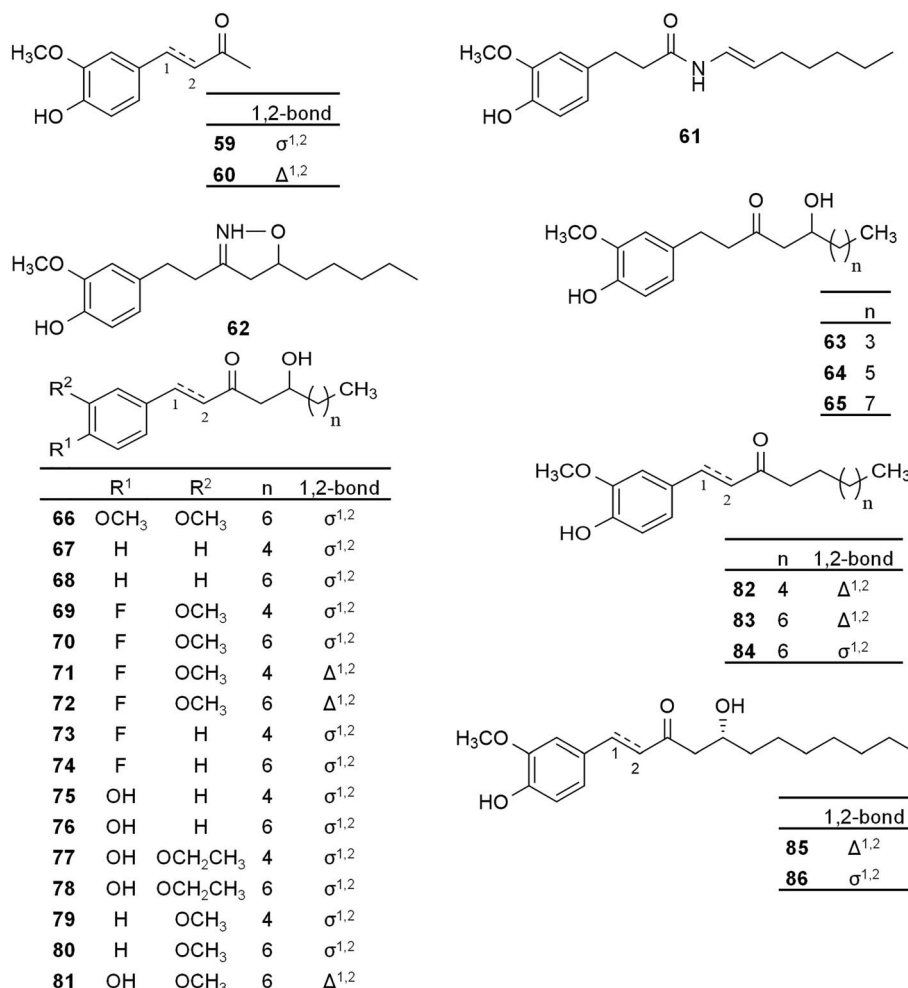


Fig. 4 Semi-synthetic gingerol derivatives prepared and evaluated for their *in vitro* antibacterial activity.



constituents, the antibacterial activity of 6-gingerol, 6-shogaol (compound **17**, Fig. 2), zingerone (**59**) and two new synthetic analogues, namely 6-azashogaol (**61**) and an isoxazole derivative of 6-gingerol (**62**) was assessed (Fig. 4). The antibacterial activity of these compounds was investigated by assessing their growth inhibitory activity on *Pseudomonas aeruginosa* and *Chromobacterium violaceum* bacterial strains, and by measuring the concentration of pyocyanin pigment produced by *P. aeruginosa*.⁷³ The expression of this pigment is controlled by QS.⁷² Compound **61** was reported to have the highest activity with the lowest MIC against both strains and 90% reduction of pyocyanin pigment produced by *P. aeruginosa*, while zingerone (**59**) was the least active. The authors highlighted that low molecular weight compounds with long side chain are needed for anti-QS activity which may explain the low activity of zingerone (only 4 carbons long side chain). It was also observed that the presence of an amide linker enhances the activity, as evidenced by the activity of compound **61**, while the isoxazoline linker had only minor influence on the activity.⁷³

Investigating the same pathway of QS and the possibility of discovering new antibacterial gingerol derivatives, Choi *et al.* reported the synthesis of 6-gingerol and further derivatives (**19–22**, **40**, Fig. 2; **47**, Fig. 3; **59**, **60**, and **63–86**, Fig. 4), some of them naturally present in ginger root. The compounds were tested for their binding to LasR, a transcriptional regulator protein playing a major role in the processes of QS in *P. aeruginosa* and biofilm formation, which in turn confers virulence to the bacteria and poses a health problem especially to immunocompromised patients. Bacterial inhibition was also monitored. It was concluded that (*R*)-8-gingerol (**86**) and its C1–C2 unsaturated analogue (**85**) were the most promising, and the importance of stereochemistry regarding the activity against *P. aeruginosa* was emphasized. At 10 μM concentration, compound **85** decreased bacterial biofilm thickness from 34.5 μm (negative control, DMSO) to 10.3 μm , which was 13.9 μm in case of compound **86** and 17 μm for the naturally occurring (*S*)-8-gingerol (compound **20**), even though bacterial growth inhibition was not observed for either compound even at 100 μM concentration. The authors emphasized the importance of rotational rigidity between the head section and the carbonyl group for LasR-binding affinity and for inhibition of biofilm formation as evidenced by the higher activity of compound **85** as compared to that of **86**. This was also supported by *in silico* docking. SAR evaluation showed that an increase of the alkyl chain length up to 12 carbons led to an increase of activity, which may explain why 8-gingerol analogues (compounds **47**, **66**, **68**, **70**, **72**, **74**, **76**, **78**, **80**, **81**, **82**, **85** and **86**) were found to be more active than 6-gingerol derivatives (compounds **19**, **40**, **67**, **69**, **71**, **73**, **75**, **77**, and **79**). Also, it was reported that a hydrogen bond acceptor is needed at position C4 on the aromatic ring for a higher potency. In addition, the presence of a hydroxyl group substituent on the alkyl chain has been correlated with higher antibacterial activity.⁴

A recent study, published in 2019, reported the application of 6-gingerol and two of its derivatives (**85**, **86**) to reduce biofouling in reverse osmosis water treatment systems by disrupting the QS processes of *P. aeruginosa*. The study aimed to

propose effective but harmless solution to membrane biofouling as the biocides in use may pose toxicity problems. In accordance with the above-mentioned results reported by Choi *et al.*, compound **85** was the most effective in inhibiting biofilm formation followed by compound **86** and then 6-gingerol (38%, 35%, and 22% reduction in biofilm formation, respectively), while only 4% inhibition was observed for sodium hypochlorite (NaOCl) used as a positive control in this study (only 10 μM NaOCl concentration was used in the study for the purpose of comparison, while its MIC is ranging between 33.6–40.3 mM). These results were confirmed by the reduction in QS-responsive gene expression. However, bacterial growth was not affected by the compounds investigated.⁷⁴

Anti-platelet gingerol analogues

The anti-platelet activity of ginger was also investigated, and ginger constituents like 6-gingerol and 6-shogaol were reported to exert potent antiplatelet activity.^{75,76}

Shih *et al.* reported the synthesis of 6-gingerol and a group of 45 derivatives that are either naturally present in ginger root or new synthetic analogues (compounds **17**, **20**, **21**, **29**, **40**, and **41**, Fig. 2; **47**, **48**, and **51**, Fig. 3; **63–65**, **81**, **82**, and **83**, Fig. 4; and **87–116**, Fig. 5), and testing these compounds for anti-platelet aggregation activity. It was demonstrated that at 10 $\mu\text{g ml}^{-1}$ concentration most of the compounds exert an over 90% inhibition of platelet aggregation induced by 100 μM of arachidonic acid. Compounds of the paradol series (**47**, **51** and **103–107**) were the most active, and 6-paradol (**51**) showed the highest activity ($\text{IC}_{50} = 0.070 \mu\text{g ml}^{-1}$ compared to 1 $\mu\text{g ml}^{-1}$ for 6-gingerol). A decrease in the activity was observed with the introduction of a double bond (as in shogaols, or dehydroparadols) or a hydroxyl group (as in gingerols) into the paradol side chain, however, increasing the alkyl side chain length increased the activity (*e.g.* the dehydroparadol compound **110** showed an IC_{50} value of 0.160 $\mu\text{g ml}^{-1}$). The epoxide derivatives (**111–116**) showed a lower potency compared to *n*-paradols ($\text{IC}_{50} = 0.96–2.38 \mu\text{g ml}^{-1}$). On the other hand, it was reported that the compounds showed negligible activity against platelet aggregation induced by platelet activating factor (PAF) or thrombin (Thr), suggesting that compound **51** is a selective inhibitor.⁷⁷

Koo *et al.* studied the effect of 6-gingerol and its synthetic analogues (**20**, Fig. 2; **54**, **55**, Fig. 3; and **117–119**, Fig. 5) on the arachidonic acid-induced platelet serotonin release and aggregation, and found lower platelet aggregation inhibitory activity for all compounds ($\text{IC}_{\text{max}} = 10–25 \mu\text{M}$) as compared to acetylsalicylic acid ($\text{IC}_{\text{max}} = 6 \mu\text{M}$). However, the compounds acted in a similar dose range as Aspirin when tested on arachidonic acid-induced platelet serotonin release. To examine the underlying mechanism, COX-inhibitory activity of these compounds was assessed. Compounds **55**, **20**, and **54** exerted similarly potent inhibitory activity ($\text{IC}_{50} = 1.2$, 1.5, and 3.3 μM , respectively) as the positive control indomethacin ($\text{IC}_{50} = 0.76 \mu\text{M}$), unlike 6-gingerol ($\text{IC}_{50} = 50 \mu\text{M}$). It is worth mentioning that the COX inhibitory activity of the compounds correlated with their hydrophobicity, with compound **55** being the most active and the most hydrophobic at the same time.⁷⁸



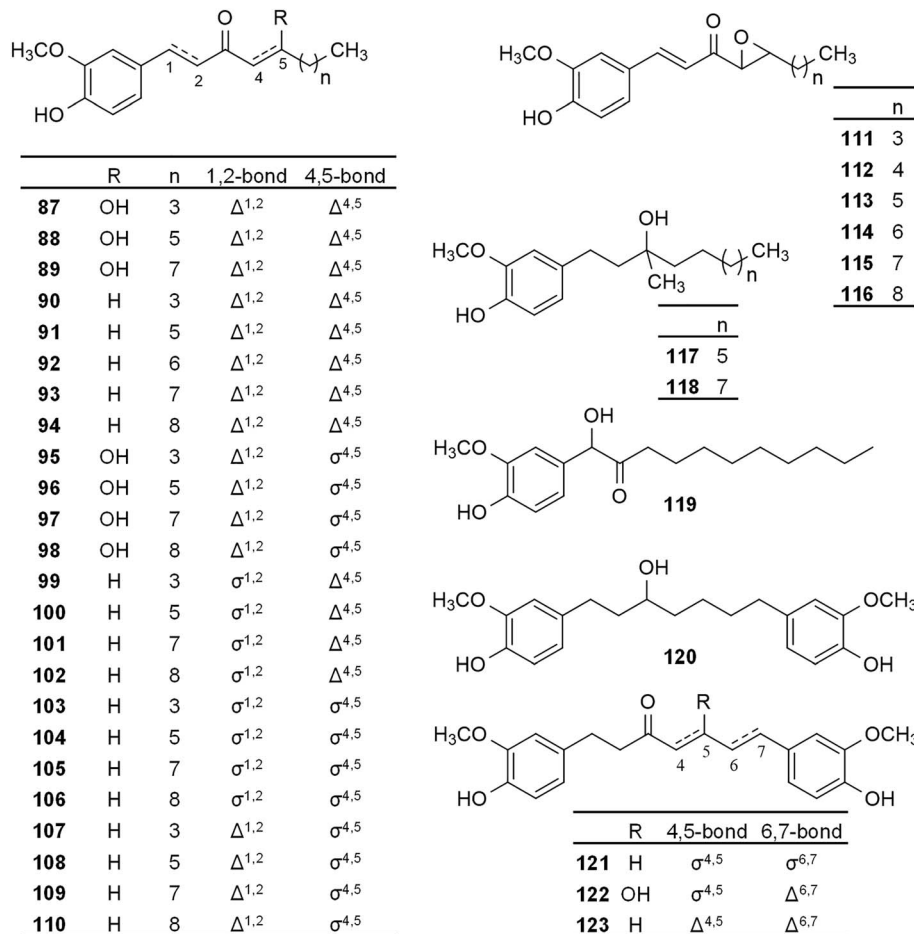


Fig. 5 Semi-synthetic gingerol derivatives prepared and evaluated for their *in vitro* anti-platelet activity.

The anti-platelet and COX-1 inhibitory activity were also studied for another set of gingerol derivatives (17, 20, 21, 30, 42, Fig. 2; and 47–49, 51, 54–58, Fig. 3; 65, Fig. 4; and 120–123, Fig. 5). 8-Paradol (47) was reported as the most effective anti-platelet agent among the investigated analogues (75% inhibition at 2 μM while 6-gingerol exerted 3.4% inhibition at the same concentration). In case of compounds 47, 49 and 58 the COX-1 inhibitory activity was also assessed through monitoring the amount of the pro-aggregatory product thromboxane A₂. Compound 47 also exerted the highest activity and it was more potent than aspirin under the conditions described in the study ($\text{IC}_{50} = 4 \mu\text{M}$ vs. 20 μM). Investigations into the SAR revealed that presence of the carbonyl function at C3 is important for activity, any other substituents on the alkyl chain interfere with the COX-1 inhibitory activity. This was evidenced through comparing the activity of compound 47 with that of 49 ($\text{IC}_{50} = 20 \mu\text{M}$). No correlation between the molecular hydrophobicity and anti-platelet aggregation activity was found in this case.⁷⁹

The above-mentioned *in vitro* results seem to be promising and may also point towards cardiovascular protective drug development. Nevertheless, further research would be needed to evaluate the *in vivo* efficacy and safety of gingerol analogues to assess their potential as anti-platelet lead or candidate drugs.

Miscellaneous bioactivities

Several other bioactivities were reported for ginger and its constituents, *e.g.* smooth muscle relaxant, painkiller, anti-emetic, and anti-obesity activity. Altogether, these reports suggest a versatile pharmacology to these natural products. Anti-emetic activity of 6-gingerol was also confirmed in a phase II randomized double-blind placebo-controlled study in cancer patients treated with highly emetogenic chemotherapeutic agents.²⁵ A recent study on rats revealed that the underlying mechanism of action is the modulation of serotonin levels.⁸⁰ Although the effect of ginger in nausea and vomiting prevention and treatment was thoroughly studied and showed promising results, to the best of our knowledge no related studies were dedicated for semisynthetic derivatives of 6-gingerol.

Gingerol analogues with activity on muscles' Ca²⁺ homeostasis

In 1988, 6-gingerol and compounds 20–22, 28, 30, 42 (Fig. 2), and 124–131 (Fig. 6) were investigated for their effect on spontaneous Ca²⁺ spikes and contraction of portal veins isolated from mice. Most of the tested compounds suppressed spontaneous contractions; however, the most promising activity was



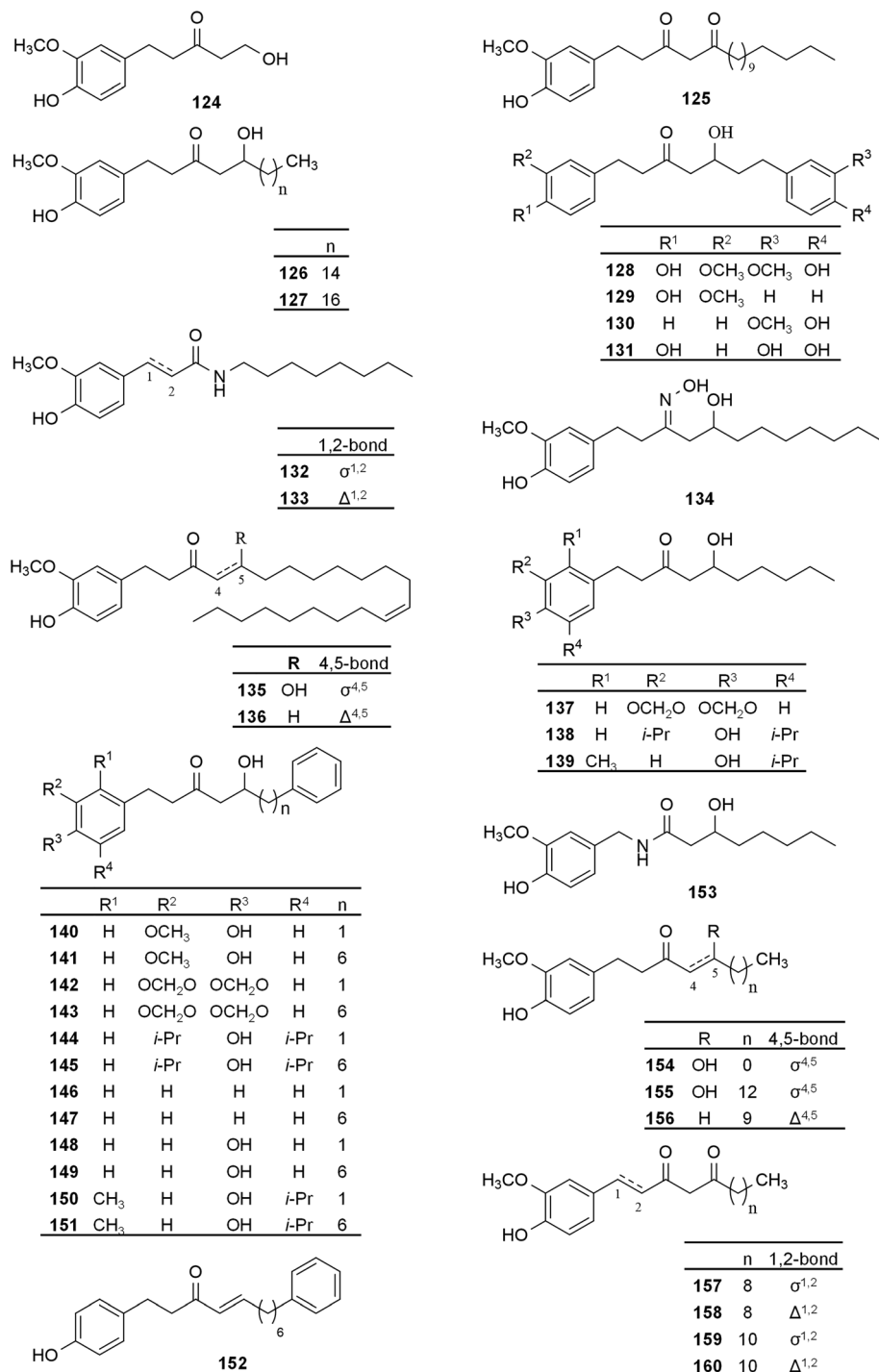


Fig. 6 Semi-synthetic gingerol derivatives with miscellaneous bioactivities. Compounds are presented that were prepared and evaluated for their effect on the intracellular Ca²⁺ homeostasis, TRPV1 and TRPA1 ion channels, or as potential antidiabetic or hepatoprotective agents.

found for 6-gingerol and derivative **129**. Interestingly, compound **20** (8-gingerol) also inhibited spontaneous contractions but without affecting Ca²⁺ spikes. It was suggested that the inhibition of spontaneous contraction is due to Ca²⁺ spike suppression in case of 6-gingerol, while 8-gingerol acts through a different mechanism of action. SAR evaluation revealed that the optimal alkyl chain length for higher potency is ten carbons

(*i.e.*, the chain length of 6-gingerol), and the reduction of carbonyl group on C3 of the alkyl side chain or oxidation of the 5-hydroxyl group decreases smooth muscle relaxant activity.⁸¹

Based on its stimulating activity on cardiac sarcoplasmic reticulum (CSR) Ca²⁺ ATPase, 6-gingerol was reported as a potent cardiotonic agent by Kobayashi *et al.*, and it was also found active on skeletal muscles.⁸² Based on a streptozotocin-



treated diabetic mouse model, this bioactivity also confers 6-gingerol a beneficial activity in diabetes-related diastolic dysfunction.⁸³ When studying this bioactivity for 6-, 8-, and 10-gingerol and their derivatives (20, 21; Fig. 2, and 132–134; Fig. 6), all the tested compounds were found to increase the SR-ATPase activity in a concentration-dependent manner. Therefore, it was postulated that they may play a role in Ca^{2+} – pumping from the cytoplasm to the SR lumen causing skeletal muscle relaxation, and both the hydrocarbon chain and the *o*-methoxyphenol parts were postulated as necessary for the activity.⁸⁴

Pain management: TRPV and TRPA modulation

Ginger has a proven painkiller activity operating *via* multiple mechanisms: COX and lipoxygenase (LOX) enzymes inhibition, inhibition of NF- κ B, or acting as vanilloid receptor agonist.⁸⁵ The transient receptor potential (TRP) cation channel subfamily V, member 1 (TRPV1), and subfamily A, member 1 (TRPA1) are known for their integral role in pain and neurogenic inflammation. Recent studies revealed that TRP channels are crucial in other physiological and pathological conditions, like cancer, cardiac health, and renal physiology. However, their precise function *in vivo* is still yet to be revealed.^{86–88}

In 2007, inspired by the activity of the oleyl moiety on TRPV1 receptor,⁸⁹ the synthesis of related gingerol and shogaol analogues (oleylgingerol; 135, and oleylshogaol; 136) was reported. When testing TRPV1 activating effect of 6-gingerol and its derivatives (17, 20, 21; Fig. 2, 48; Fig. 3, 101; Fig. 5, 135 and 136; Fig. 6), all compounds showed higher activity ($\text{EC}_{50} = 0.26\text{--}4.17\ \mu\text{M}$) than 6-gingerol ($\text{EC}_{50} = 4.55\ \mu\text{M}$) but lower than the positive control capsaicin ($\text{EC}_{50} = 0.082\ \mu\text{M}$). Oleylgingerol was the most active, while oleylshogaol the least active of them, suggesting that the 5-hydroxyl group has a significant role in activating the TRPV1 channel.⁹⁰

To explore the chemical space around 6-gingerol concerning its potential for vanilloid receptor modulation, Morera *et al.* reported the synthesis of racemic 6-gingerol analogues (17; Fig. 1, 67, 75; Fig. 4 and 137–152; Fig. 6) and their biological evaluation on TRPV1 and TRPA1 channels.⁹¹ With regard to the activity on TRPV1, compound 141 exerted the highest potency and selectivity among all ($\text{EC}_{50} = 0.11\ \mu\text{M}$, compared to $3.3\ \mu\text{M}$ for 6-gingerol), whereas compounds 138, 144, and 145, each containing two isopropyl groups in *ortho* position to the phenolic hydroxyl group, were found to be inactive. This indicates that steric hindrance around the phenolic hydroxyl function negatively affects the activity. Further studies into the SAR showed importance of the phenolic hydroxyl group and noted an increase in activity with the increase in molecular lipophilicity. The free hydroxyl group on the side chain was found to be of less importance as proved by the efficacy of 6-shogaol (17) and its analogue (152) on TRPV1 channels. On the contrary, activity on TRPA1 channels was the highest in case of compound 139, with favoured branched alkyl substituents at the *ortho* position to the phenolic hydroxyl group. It is noteworthy that compounds 17, 141 and 151 acted as selective

TRPV1 agonists, while compounds 143, 144, 146, and 147 were selective TRPA1 antagonists.⁹¹

Antidiabetic and antihepatotoxic activities

Results from a randomized clinical trial on the possible anti-diabetic potential of ginger consumption suggested a decrease in blood glucose and cholesterol levels.⁹²

Moreover, synergistic effect of 6-gingerol and quercetin was investigated in streptozotocin-induced type 2 diabetes and poloxamer P-407 induced hyperlipidaemia on rats. The combination treatment was found to exert remarkable antidiabetic and beneficial cardiac effects, and the synergism was suggested to occur through the modulation of serotonergic system.⁹³

In metabolic syndrome, the prophylactic effect of 6-gingerol and its synthetic analogue aza-6-gingerol (153) was investigated on high fat diet-fed type 2 diabetic mice. Both compounds caused a reduction of lipogenic genes, which was postulated to occur through downregulation of sterol regulatory element-binding protein 1c (SREBP-1c), a protein that is responsible for regulating the transcription of many lipogenic genes. Compound 153 was found more effective than 6-gingerol in enhancing metabolism and reducing the extent of lipogenesis. It was suggested that compound 153 might possess potential therapeutic value to reduce the risk of obesity-associated diseases, however, further studies are needed to uncover the underlying mechanism of action.⁹⁴

Liu *et al.* reported the repressive activity of 6-gingerol on nutritional steatohepatitis induced in mice. The protective effect was postulated to occur due to regulation of key genes related to oxidative stress, inflammation and fibrogenesis.⁹⁵ Antihepatotoxic effects of gingerols and shogaols on carbon tetrachloride- and galactosamine-induced cytotoxicity in primary cultured rat hepatocytes were also studied. Compounds 17, 20–22, 28–30, 42, 43 (Fig. 2), 48 (Fig. 3), 59 (Fig. 4), 100–102 (Fig. 5), 124, 126, and 154–160 (Fig. 6) were evaluated, and both gingerols and shogaols were found to exert anti-hepatotoxic actions, with gingerols being superior in this regard. Studies into the SAR revealed importance of the side chain length with the highest activity achieved with (7) and (8)-congeners. Reduction of the carbonyl group on the side chain, oxidation of the 5-hydroxyl group, or introduction of a double bond into the side chain were found to decrease the activity of gingerols.⁹⁶

Conclusions

6-Gingerol, the main constituent of *Zingiber officinale* (Zingiberaceae) has been extensively studied in the last century for its potential anti-inflammatory, antioxidant, antimicrobial and anticancer effects. Several clinical studies have been performed and some are currently in progress to confirm these observations. The beneficial activities of ginger inspired several research groups to explore the chemical space around 6-gingerol: chemical modifications have led to numerous derivatives interfering with multiple cellular pathways leading to improved anticancer, anti-inflammatory, antimicrobial anti-platelet, and/or various other bioactivities. Some of these derivatives seem to



show serious potential to drug discovery, concerning their sub-micromolar efficacy as anti-inflammatory and/or antiplatelet agents, or vanilloid receptor agonist. Nevertheless, *in vivo* studies are missing for many of these compounds so that their efficacy and safety profile could be better evaluated, and to the best of our knowledge none of the semi-synthetic derivatives of gingerol reached a clinical trial until now. Therefore, there is still much to learn on the true drug discovery potential that can be attributed to the chemical space surrounding this relatively simple, but versatile natural product, 6-gingerol.

Conflicts of interest

There are no conflicts of interest to declare.

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References

- 1 Y. a. Han, C. w. Song, W. s. Koh, G. h. Yon, Y. s. Kim, S. y. Ryu, H. j. Kwon and K. h. Lee, *Phytother. Res.*, 2013, **27**, 1200–1205.
- 2 Q. Q. Mao, X. Y. Xu, S. Y. Cao, R. Y. Gan, H. Corke, T. Beta and H. B. Li, *Foods*, 2019, **8**, 185.
- 3 S. H. Nile and P. Se Won, *Ind. Crops Prod.*, 2015, **70**, 238–244.
- 4 H. Choi, S. Y. Ham, E. Cha, Y. Shin, H. S. Kim, J. K. Bang, S. H. Son, H. D. Park and Y. Byun, *J. Med. Chem.*, 2017, **60**, 9821–9837.
- 5 C. K. Wei, Y. H. Tsai, M. Korinek, P. H. Hung, M. El-Shazly, Y. B. Cheng, Y. C. Wu, T. J. Hsieh and F. R. Chang, *Int. J. Mol. Sci.*, 2017, **18**, 168.
- 6 A. J. Akinyemi, G. R. Thome, V. M. Morsch, N. Stefanello, J. F. Goularte, A. Belló-Klein, G. Oboh and M. R. C. Schetinger, *J. Funct. Foods*, 2015, **17**, 792–801.
- 7 S. C. Ho, K. S. Chang and C. C. Lin, *Food Chem.*, 2013, **141**, 3183–3191.
- 8 S. Suk, G. T. Kwon, E. Lee, W. J. Jang, H. Yang, J. H. Kim, N. R. Thimmegowda, M. Y. Chung, J. Y. Kwon, S. Yang, J. K. Kim, J. H. Y. Park and K. W. Lee, *Mol. Nutr. Food Res.*, 2017, **61**, DOI: 10.1002/mnfr.201700139.
- 9 L. B. Martins, A. Rodrigues, D. F. Rodrigues, L. C. Dos Santos, A. L. Teixeira and A. V. M. Ferreira, *Cephalalgia*, 2019, **39**, 68–76.
- 10 M. Jalali, M. Mahmoodi, S. P. Moosavian, R. Jalali, G. Ferns, A. Mosallanezhad, M. H. Imanieh and Z. Mosallanezhad, *Phytother. Res.*, 2020, 1723–1733.
- 11 O. E. Orisakwe, C. N. Orish and E. O. Nwanaforo, *Scientific African*, 2020, e00620, DOI: 10.1016/j.sciaf.2020.e00620.
- 12 D. Sen and P. Debnath, *J. Biomol. Struct. Dyn.*, 2020, 1–22.
- 13 S. Kumar, P. Kashyap, S. Chowdhury, S. Kumar, A. Panwar and A. Kumar, *Phytomedicine*, 2020, 153317, DOI: 10.1016/j.phymed.2020.153317.
- 14 WHO, <https://www.who.int/southeastasia/outbreaks-and-emergencies/novel-coronavirus-2019/fact-or-fiction>, accessed 18.12.2020.
- 15 D. Silveira, J. M. Prieto-Garcia, F. Boylan, O. Estrada, Y. M. Fonseca-Bazzo, C. M. Jamal, P. O. Magalhães, E. O. Pereira, M. Tomczyk and M. Heinrich, *Front. Pharmacol.*, 2020, **11**, 581840.
- 16 M. F. Mahomoodally, M. Z. Aumeeruddy, K. R. R. Rengasamy, S. Roshan, S. Hammad, J. Pandohee, X. Hu and G. Zengin, *Semin. Cancer Biol.*, 2021, **69**, 140–149.
- 17 M.-J. Ko, H.-H. Nam and M.-S. Chung, *Food Chem.*, 2019, **270**, 149–155.
- 18 R. M. T. de Lima, A. C. Dos Reis, A. P. M. de Menezes, J. V. O. Santos, J. Filho, J. R. O. Ferreira, M. de Alencar, A. da Mata, I. N. Khan, A. Islam, S. J. Uddin, E. S. Ali, M. T. Islam, S. Tripathi, S. K. Mishra, M. S. Mubarak and A. A. C. Melo-Cavalcante, *Phytother. Res.*, 2018, **32**, 1885–1907.
- 19 S. Wang, C. Zhang, G. Yang and Y. Yang, *Nat. Prod. Commun.*, 2014, **9**, 1027–1030.
- 20 A. Dehghani Nazhvani, N. Sarafraz, F. Askari, F. Heidari and M. Razmkhah, *Asian Pac. J. Cancer Prev.*, 2020, **21**, 479–484.
- 21 P. Mega Tiber, S. Kocyyigit Sevinc, O. Kilinc and O. Orun, *Gene*, 2019, **692**, 217–222.
- 22 N. Kaewtunjai, R. Wongpoomchai, A. Imsumran, W. Pompimon, A. Athipornchai, A. Suksamrarn, T. R. Lee and W. Tuntiwechapikul, *ACS Omega*, 2018, **3**, 18572–18581.
- 23 M. Crichton, S. Marshall, W. Marx, A. L. McCarthy and E. Isenring, *J. Acad. Nutr. Diet.*, 2019, **119**, 2055–2068.
- 24 A. Uthaipaisanwong and S. Oranratanaphan, *Support. Care Cancer*, 2019, 3831–3838.
- 25 J. Konmun, K. Danwilai, N. Ngamphaiboon, B. Sripanidkulchai, A. Sookprasert and S. Subongkot, *Med. Oncol.*, 2017, **34**, 69.
- 26 A. Almatroudi, M. A. Alsahli, F. Alrumaihi, K. S. Allemailem and A. H. Rahmani, *Curr. Pharm. Biotechnol.*, 2019, **20**, 5–16.
- 27 C. Chatupheeraphat, C. Nantasenamat, K. Deesrisak, S. Roytrakul, U. Anurathapan and D. Tanyong, *EXCLI J.*, 2020, **19**, 582–595.
- 28 N. Sp, D. Y. Kang, J. M. Lee, S. W. Bae and K. J. Jang, *Int. J. Mol. Sci.*, 2021, **22**, 4660.
- 29 J. Czarnik-Kwasniak, K. Kwasniak, P. Kwasek, E. Swierzowska, A. Strojewska and J. Tabarkiewicz, *Nutrients*, 2019, **12**, 96.
- 30 Y. Sun, J. Ren and F. Wang, *J. Biochem. Mol. Toxicol.*, 2021, **35**, e22689.
- 31 C. J. Weng and G. C. Yen, *Cancer Treat. Rev.*, 2012, **38**, 76–87.
- 32 C. Tonelli, I. I. C. Chio and D. A. Tuveson, *Antioxid. Redox Signaling*, 2018, **29**, 1727–1745.
- 33 M. G. van der Wijst, R. Brown and M. G. Rots, *Biochim. Biophys. Acta*, 2014, **1846**, 494–509.
- 34 S. Vomund, A. Schäfer, M. J. Parnham, B. Brüne and A. von Knethen, *Int. J. Mol. Sci.*, 2017, **18**, 2772.
- 35 S. Nafees, M. Zafaryab, S. H. Mehdi, B. Zia, M. A. Rizvi and M. A. Khan, *Anti-Cancer Agents Med. Chem.*, 2021, **21**, 428–432.



- 36 J. Citronberg, R. Bostick, T. Ahearn, D. K. Turgeon, M. T. Ruffin, Z. Djuric, A. Sen, D. E. Brenner and S. M. Zick, *Cancer Prev. Res.*, 2013, **6**, 271–281.
- 37 Y. Jiang, D. K. Turgeon, B. D. Wright, E. Sidahmed, M. T. Ruffin, D. E. Brenner, A. Sen and S. M. Zick, *Eur. J. Cancer Prev.*, 2013, **22**, 455–460.
- 38 S. M. Zick, D. K. Turgeon, S. K. Vareed, M. T. Ruffin, A. J. Litzinger, B. D. Wright, S. Alrawi, D. P. Normolle, Z. Djuric and D. E. Brenner, *Cancer Prev. Res.*, 2011, **4**, 1929–1937.
- 39 S. M. Zick, D. K. Turgeon, J. Ren, M. T. Ruffin, B. D. Wright, A. Sen, Z. Djuric and D. E. Brenner, *Mol. Carcinog.*, 2015, **54**, 908–915.
- 40 W. C. de Lima Silva, R. Conti, L. C. de Almeida, P. A. B. Morais, K. B. Borges, V. L. Junior, L. V. Costa-Lotufu and W. de Souza Borges, *Curr. Top. Med. Chem.*, 2020, **20**, 161–169.
- 41 A. S. Ibrahim, M. A. Sobh, H. M. Eid, A. Salem, H. H. Elbelasi, M. H. El-Naggar, F. M. AbdelBar, H. Sheashaa, M. A. Sobh and F. A. Badria, *Tumor Biol.*, 2014, **35**, 9941–9948.
- 42 L. Luna-Dulcey, J. A. da Silva and M. R. Cominetti, *Anti-Cancer Drugs*, 2020, **31**, 35–43.
- 43 L. Luna-Dulcey, R. Tomasin, M. A. Naves, J. A. da Silva and M. R. Cominetti, *Oncotarget*, 2018, **9**, 30787–30804.
- 44 Y. Zhu, F. Wang, Y. Zhao, P. Wang and S. Sang, *Sci. Rep.*, 2017, **7**, 40119.
- 45 X. Chen, S. Wang, N. Wu and C. S. Yang, *Curr. Cancer Drug Targets*, 2004, **4**, 267–283.
- 46 N. Oi, H. Yamamoto, A. Langfald, R. Bai, M. H. Lee, A. M. Bode and Z. Dong, *Carcinogenesis*, 2017, **38**, 728–737.
- 47 M. H. El-Naggar, A. Mira, F. M. Abdel Bar, K. Shimizu, M. M. Amer and F. A. Badria, *Bioorg. Med. Chem.*, 2017, **25**, 1277–1285.
- 48 J. E. Bolden, M. J. Peart and R. W. Johnstone, *Nat. Rev. Drug Discovery*, 2006, **5**, 769–784.
- 49 Z. Jiang, W. Li, X. Hu, Q. Zhang, T. Sun, S. Cui, S. Wang, Q. Ouyang, Y. Yin, C. Geng, Z. Tong, Y. Cheng, Y. Pan, Y. Sun, H. Wang, T. Ouyang, K. Gu, J. Feng, X. Wang, S. Wang, T. Liu, J. Gao, M. Cristofanilli, Z. Ning and X. Lu, *Lancet Oncol.*, 2019, **20**, 806–815.
- 50 C. Robert and F. V. Rassool, *Adv. Cancer Res.*, 2012, **116**, 87–129.
- 51 J.-H. Lee, M. L. Choy, L. Ngo, S. S. Foster and P. A. Marks, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 14639–14644.
- 52 I. A. M. Groh, C. Chen, C. Lüske, A. T. Cartus and M. Esselen, *J. Nutr. Metab.*, 2013, **2013**, 821082.
- 53 A. K. Kiss, in *Handbook of Nutrition, Diet, and Epigenetics*, ed. V. Patel and V. Preedy, Springer International Publishing, Cham, 2017, DOI: DOI: 10.1007/978-3-319-31143-2_105-1, pp. 1–21.
- 54 L. W. Evans and B. S. Ferguson, *Nutrients*, 2018, **10**, 1120.
- 55 P. Kumboonma, T. Senawong, S. Saenglee, C. Yenjai and C. Phaosiri, *Med. Chem. Res.*, 2017, **26**, 650–661.
- 56 D.-L. Lu, X.-Z. Li, F. Dai, Y.-F. Kang, Y. Li, M.-M. Ma, X.-R. Ren, G.-W. Du, X.-L. Jin and B. Zhou, *Food Chem.*, 2014, **165**, 191–197.
- 57 M. Rondanelli, A. Riva, P. Allegrini, M. A. Faliva, M. Naso, G. Peroni, M. Nichetti, C. Gasparri, D. Spadaccini, G. Iannello, V. Infantino, T. Fazia, L. Bernardinelli and S. Perna, *J. Pain Res.*, 2020, **13**, 761–770.
- 58 R. D. Altman and K. C. Marcussen, *Arthritis Rheum.*, 2001, **44**, 2531–2538.
- 59 D. C. Nieman, R. A. Shanely, B. Luo, D. Dew, M. P. Meaney and W. Sha, *Nutr. J.*, 2013, **12**, 154.
- 60 S. Atashak, M. Peeri, M. A. Azarbayjani, S. R. Stannard and M. M. Haghghi, *J. Sports Sci. Med.*, 2011, **10**, 685–691.
- 61 N. Aryaeian, M. Mahmoudi, F. Shahram, S. Poursani, F. Jamshidi and H. Tavakoli, *Med J. Islam Repub. Iran.*, 2019, **33**, 154.
- 62 F. L. Zhang, B. W. Zhou, Z. Z. Yan, J. Zhao, B. C. Zhao, W. F. Liu, C. Li and K. X. Liu, *Cytokine*, 2020, **125**, 154854.
- 63 T. Bergsbaken, S. L. Fink and B. T. Cookson, *Nat. Rev. Microbiol.*, 2009, **7**, 99–109.
- 64 E. Tjendraputra, V. H. Tran, D. Liu-Brennan, B. D. Roufogalis and C. C. Duke, *Bioorg. Chem.*, 2001, **29**, 156–163.
- 65 F. Aktan, S. Henness, V. H. Tran, C. C. Duke, B. D. Roufogalis and A. J. Ammit, *Planta Med.*, 2006, **72**, 727–734.
- 66 A. Azizi, S. Aghayan, S. Zaker, M. Shakeri, N. Entezari and S. Lawaf, *Int. J. Dent.*, 2015, **2015**, 489842.
- 67 S. Hasan, M. Danishuddin and A. U. Khan, *BMC Microbiol.*, 2015, **15**, 1.
- 68 J. H. Lee, Y. G. Kim, P. Choi, J. Ham, J. G. Park and J. Lee, *Front. Cell. Infect. Microbiol.*, 2018, **8**, 299.
- 69 B. O. Oyedemi, E. M. Kotsia, P. D. Stapleton and S. Gibbons, *J. Ethnopharmacol.*, 2019, **245**, 111871.
- 70 L. Cai, S. Liu, L. Sun, Y. Wang, H. Ji and J. Li, *Front. microbiol.*, 2015, **6**, 981.
- 71 H. S. Kim, S. H. Lee, Y. Byun and H. D. Park, *Sci. Rep.*, 2015, **5**, 8656.
- 72 S. T. Rutherford and B. L. Bassler, *Cold Spring Harbor Perspect. Med.*, 2012, **2**, a012427.
- 73 N. V. Kumar, P. S. Murthy, J. R. Manjunatha and B. K. Bettadaiah, *Food Chem.*, 2014, **159**, 451–457.
- 74 S.-Y. Ham, H.-S. Kim, Y. Jang, P.-F. Sun, J.-H. Park, J. Lee, Y. Byun and H.-D. Park, *Fuel*, 2019, **250**, 79–87.
- 75 G. E. Hirsch, P. R. N. Viceli, A. S. de Almeida, S. Nascimento, F. G. Porto, J. Otero, A. Schmidt, B. da Silva, M. M. Parisi and J. Z. Klafke, *Curr. Pharm. Des.*, 2017, **23**, 1228–1246.
- 76 Y. R. Liao, Y. L. Leu, Y. Y. Chan, P. C. Kuo and T. S. Wu, *Molecules*, 2012, **17**, 8928–8937.
- 77 H. C. Shih, C. Y. Chern, P. C. Kuo, Y. C. Wu, Y. Y. Chan, Y. R. Liao, C. M. Teng and T. S. Wu, *Int. J. Mol. Sci.*, 2014, **15**, 3926–3951.
- 78 K. L. Koo, A. J. Ammit, V. H. Tran, C. C. Duke and B. D. Roufogalis, *Thromb. Res.*, 2001, **103**, 387–397.
- 79 E. Nurtjahja-Tjendraputra, A. J. Ammit, B. D. Roufogalis, V. H. Tran and C. C. Duke, *Thromb. Res.*, 2003, **111**, 259–265.
- 80 Q. Cheng and X. Feng, *Drug Des., Dev. Ther.*, 2020, **14**, 4085–4099.
- 81 I. Kimura, L. R. Pancho, T. Shiori and M. Kimura, *Jpn. J. Pharmacol.*, 1988, **48**, 257–262.
- 82 M. Kobayashi, N. Shoji and Y. Ohizumi, *Biochim. Biophys. Acta, Biomembr.*, 1987, **903**, 96–102.



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- 83 I. Namekata, S. Hamaguchi, Y. Wakasugi, M. Ohhara, Y. Hirota and H. Tanaka, *Eur. J. Pharmacol.*, 2013, **706**, 48–55.
- 84 Y. Ohizumi, S. Sasaki, K. Shibusawa, K. Ishikawa and F. Iketomo, *Biol. Pharm. Bull.*, 1996, **19**, 1377–1379.
- 85 M. Rondanelli, F. Fossari, V. Vecchio and C. Gasparri, *Phytother. Res.*, 2020, **34**, 2843–2856.
- 86 E. S. Fernandes, M. A. Fernandes and J. E. Keeble, *Br. J. Pharmacol.*, 2012, **166**, 510–521.
- 87 M. M. Moran, M. A. McAlexander, T. Bíró and A. Szallasi, *Nat. Rev. Drug Discovery*, 2011, **10**, 601–620.
- 88 A. Samanta, T. E. T. Hughes and V. Y. Moiseenkova-Bell, *Subcell. Biochem.*, 2018, **87**, 141–165.
- 89 A. Morita, Y. Iwasaki, K. Kobata, T. Iida, T. Higashi, K. Oda, A. Suzuki, M. Narukawa, S. Sasakuma, H. Yokogoshi, S. Yazawa, M. Tominaga and T. Watanabe, *Life Sci.*, 2006, **79**, 2303–2310.
- 90 A. Morita, Y. Iwasaki, K. Kobata, H. Yokogoshi and T. Watanabe, *Biosci., Biotechnol., Biochem.*, 2007, **71**, 2304–2307.
- 91 E. Morera, L. De Petrocellis, L. Morera, A. S. Moriello, M. Nalli, V. Di Marzo and G. Ortar, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 1674–1677.
- 92 G. C. N. Carvalho, J. C. G. Lira-Neto, M. F. M. d. Araújo, R. W. J. F. d. Freitas, M. L. Zanetti and M. M. C. Damasceno, *Rev. Lat.-Am. Enferm.*, 2020, **28**, e3369.
- 93 Y. Shao, Y. Yu, C. Li, J. Yu, R. Zong and C. Pei, *RSC Adv.*, 2016, **6**, 12235–12242.
- 94 M. Okamoto, H. Irii, Y. Tahara, H. Ishii, A. Hirao, H. Udagawa, M. Hiramoto, K. Yasuda, A. Takanishi, S. Shibata and I. Shimizu, *J. Med. Chem.*, 2011, **54**, 6295–6304.
- 95 T.-F. Tzeng, S.-S. Liou and I. M. Liu, *RSC Adv.*, 2014, **4**, 61427–61436.
- 96 H. Hikino, Y. Kiso, N. Kato, Y. Hamada, T. Shioiri, R. Aiyama, H. Itokawa, F. Kiuchi and U. Sankawa, *J. Ethnopharmacol.*, 1985, **14**, 31–39.

