

PAPER

View Article Online
View Journal | View Issue

Cite this: *Biomater. Sci.*, 2025, **13**, 1323

A thiol–ene click-based strategy to customize injectable polymer–nanoparticle hydrogel properties for therapeutic delivery†

Sophia J. Bailey, ^a Noah Eckman, ^b Elisa S. Brunel,^a Carolyn K. Jons, ^a Samya Sen ^a and Eric A. Appel ^{*a,c,d,e,f}

Polymer–nanoparticle (PNP) hydrogels are a promising injectable biomaterial platform that has been used for a wide range of biomedical applications including adhesion prevention, adoptive cell delivery, and controlled drug release. By tuning the chemical, mechanical, and erosion properties of injected hydrogel depots, additional control over cell compatibility and pharmaceutical release kinetics may be realized. Here, we employ thiol–ene click chemistry to prepare a library of modified hydroxypropylmethylcellulose (HPMC) derivatives for subsequent use in PNP hydrogel applications. When combined with poly(ethylene glycol)-*b*-poly(lactic acid) nanoparticles, we demonstrate that systematically altering the hydrophobic, steric, or π stacking character of HPMC modifications can readily tailor the mechanical properties of PNP hydrogels. Additionally, we highlight the compatibility of the synthetic platform for the incorporation of cysteine-bearing peptides to access PNP hydrogels with improved bioactivity. Finally, through leveraging the tunable physical properties afforded by this method, we show hydrogel retention time *in vivo* can be dramatically altered without sacrificing mesh size or cargo diffusion rates. This work offers a route to optimize PNP hydrogels for a variety of translational applications and holds promise in the highly tunable delivery of pharmaceuticals and adoptive cells.

Received 1st October 2024,
Accepted 29th December 2024

DOI: 10.1039/d4bm01315h

rsc.li/biomaterials-science

1. Introduction

Hydrogels are water-swollen soft materials with diverse biomedical applications ranging from regenerative medicine¹ to drug delivery.² Injectable hydrogel materials are particularly advantageous for minimally invasive delivery,³ where they have been employed for cell delivery^{4,5} and as reservoirs for the sustained delivery of pharmaceuticals.⁶ Several strategies to access injectable hydrogels exist, including *in situ* curing,⁷ microparticle hydrogel assemblies,⁸ dynamic covalent chemistries,^{9,10} and supramolecular interactions.¹¹ Of these, polymer–nanoparticle (PNP) hydrogels are a promising class of supramolecu-

lar hydrogel that are formed by dynamic bridging interactions between polymers and nanoparticles. Our group has previously demonstrated that dodecyl-modified hydroxypropylmethylcellulose (HPMC-C12) can form supramolecular hydrophobic interactions with poly(ethylene glycol)-*b*-poly(lactic acid) nanoparticles (PEG-*b*-PLA NPs), providing injectable hydrogels well-suited for the sustained delivery of pharmaceuticals and cell-based therapies (Fig. 1a and b).^{12–14} The physical properties of PNP hydrogels are a key consideration for designing materials for specific applications, as the stiffness, mesh size, and *in vivo* retention time of hydrogels are directly related to cell viability,^{4,13} cargo diffusion^{2,15} and pharmacokinetics,² respectively.

To date, modulation of the physical properties and *in vivo* retention time of PNP hydrogels has largely been limited to altering the concentration of polymer and nanoparticles in solution. Thus, the stiffest, longest-lasting PNP hydrogels are accessed at high network densities, with the typical formulation being 2 wt% HPMC-C12 with 10 wt% PEG-PLA NPs, while softer, faster-dissolving hydrogels are prepared by lowering the concentrations of each component. Although much successful work has been demonstrated by this simple concentration-based method, this strategy fails to independently tailor hydrogel mechanics without changing network mesh

^aDepartment of Materials Science & Engineering, Stanford University, Stanford, CA, 94305, USA. E-mail: eappel@stanford.edu

^bDepartment of Chemical Engineering, Stanford University, Stanford, CA, 94305, USA

^cDepartment of Bioengineering, Stanford University, Stanford, CA, 94305, USA

^dChEM-H Institute, Stanford University, Stanford, CA, USA

^eWoods Institute for the Environment, Stanford University, Stanford, CA, USA

^fDepartment of Pediatrics – Endocrinology, Stanford University School of Medicine, Stanford, CA, 94305, USA

†Electronic supplementary information (ESI) available: HPMC ¹H NMRs, cell encapsulation studies, supplemental rheological data, and photographs of *in vivo* excisions. See DOI: <https://doi.org/10.1039/d4bm01315h>

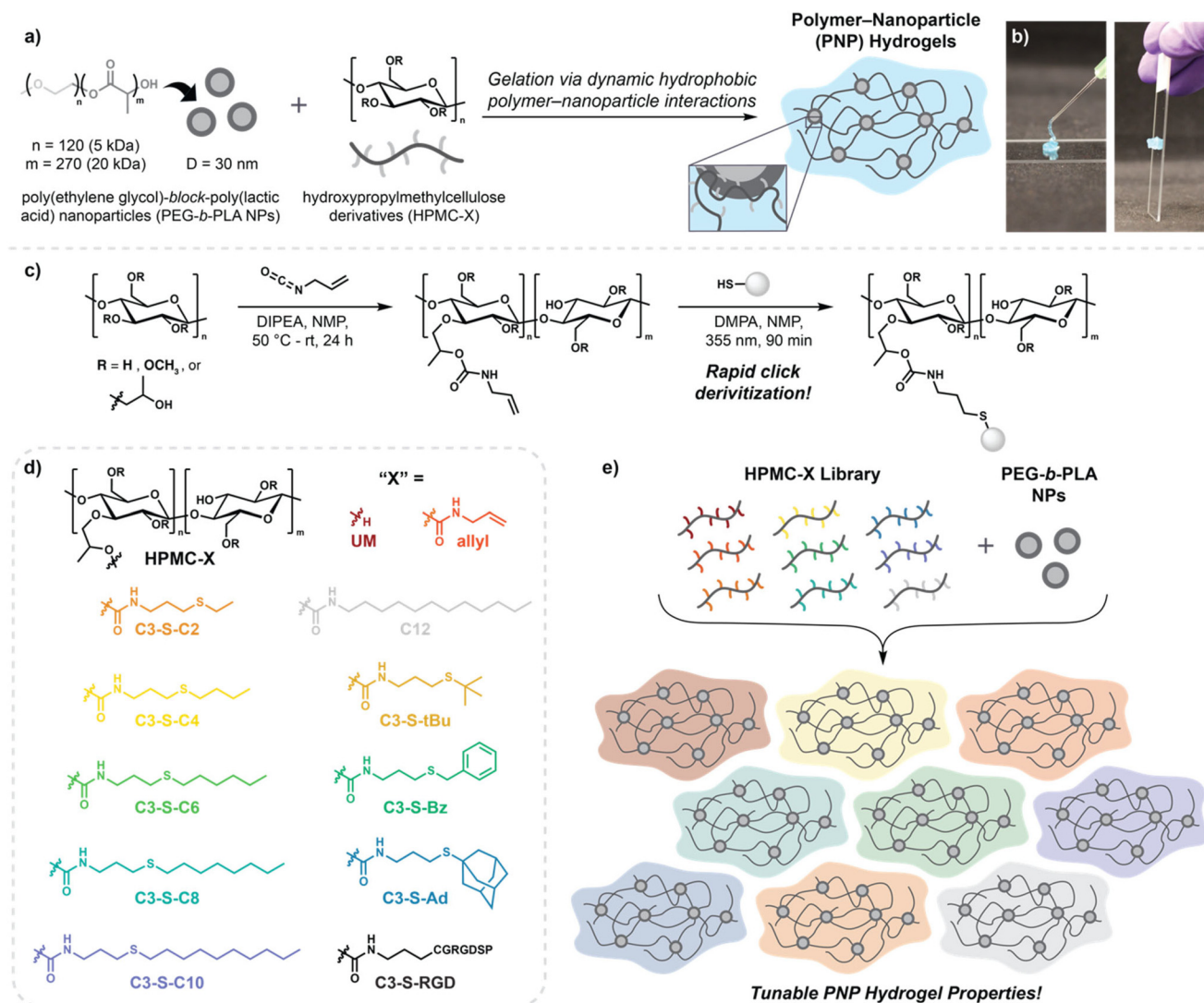



Fig. 1 Preparation of PNP hydrogel library. (a) General schematic of PNP hydrogel preparation. (b) Photographs demonstrating injectability and recovery of PNP hydrogels. Blue color corresponds to a dye used for visualization only. (c) Synthetic scheme to prepare HPMC-X derivatives. (d) Library of HPMC-X derivatives prepared. (e) Schematic depiction of PNP hydrogel library preparation.

size, and thus cargo diffusivity. Previous strategies to instead tune the strength of the supramolecular polymer-nanoparticle interaction have involved changing the identity of the nanoparticles, yielding unique hydrogel properties.¹⁶ However, this strategy is limited by the arduous synthetic requirements of block copolymer libraries and need for unique nanoprecipitation conditions. Thus, alternative chemical methods to tune PNP physical properties would enable better customization for broad biomedical applications.

We proposed that derivatization of HPMC might provide a more accessible route to tune polymer-nanoparticle interactions. In previous explorations of HPMC pendant group identity, it has been revealed that dodecyl modification results in stiffer, more robust hydrogels than hexyl or adamantyl analogs.^{12,23} However, a thorough investigation of pendant group hydrophobicity and steric effects has been plagued by

synthetic challenges. Previous approaches to functionalize HPMC have relied upon reactions with isocyanates¹² or electrostatic interactions,¹⁷ which are both limited by the commercial availability of potential pendant groups. Furthermore, isocyanates are highly reactive and suffer from poor shelf-stability, greatly limiting batch-to-batch reproducibility. Because of this, systematic studies comparing the effects of pendant group identity on polymer-nanoparticle interactions have been hindered by variable HPMC functionalization efficiencies. To thoroughly investigate pendant group effects on polymer-nanoparticle interactions and access therapeutic materials with new properties, an approach that provides consistent HPMC functionalization is needed. We envisioned click chemistry^{18,19} as a solution to this challenge, where a click handle could be installed on HPMC then easily derivatized *via* quantitative click reactions. With this strategy in mind, we



selected thiol–ene click chemistry^{20,21} as an ideal candidate as it would allow for diverse derivatization with commercially available thiols and provide access to functionalization with cysteine-bearing peptides. Herein, we use this thiol–ene click strategy to investigate how systematically altering the hydrophobic, steric, or pi stacking character of HPMC modifications can be used to tailor the properties of subsequent PNP hydrogels. Notably, we demonstrate that PNP hydrogels can be prepared with consistent network density and diffusion properties while providing varied *in vivo* retention times, offering a new approach to access superior control over therapeutic cargo release.

2. Results and discussion

2.1 Derivatization of HPMC

To access a uniformly functionalized series of HPMC derivatives, we opted to first prepare a large-batch parent material by treatment of HPMC with allyl isocyanate, followed by clicking to various thiol pendant groups (Fig. 1c). Although isocyanates have previously lead to inconsistent batch-to-batch functionalization, we envisioned the preparation of one large batch of parent material that would be used for all subsequent thiol–ene click reactions would allow us to circumvent these issues while taking advantage of previously established HPMC functionalization strategies using isocyanates.¹² For our library, we chose pendant groups which we hypothesized would readily impact the strength of the polymer–nanoparticle interaction, including thiols with varied alkyl chain length (C2–C10); steric bulk (*t*Bu and Ad) and competitive pi stacking interactions (Bz). To allow for comparison to previously reported HPMC-C12 PNP hydrogels, isocyanate feed ratio was maintained at 0.12 equivalents per cellulose repeat¹² and incorporation of 5% relative to hydroxypropyl units was confirmed by ¹H NMR (ESI Fig. S1†). This single batch of parent HPMC-allyl was then exposed to various thiol–ene conditions to install our suite of pendant groups (Fig. 1d). Successful HPMC functionalization was assessed by ¹H NMR for each derivative, where consumption of allylic protons was confirmed and peaks corresponding to the various pendant functionalities could be readily observed (ESI Fig. S2†).

2.2 Peptide–HPMC conjugates for bioactive PNP hydrogels

Adjacently, this strategy offers a route to functionalize PNP hydrogels through the conjugation of cysteine-bearing peptides to HPMC. Previous routes to functionalize PNP hydrogels with peptides have relied on clicking peptides to PEG-*b*-PLA;²² however, these peptide-bearing block copolymers suffer from poor solubility, and can pose challenges in the subsequent nanoprecipitation. Thus, we envisioned the thiol–ene click strategy to provide an additional tool to introduce bioactivity to PNP hydrogels. To demonstrate the applicability of this alternative strategy for PNP functionalization, we conjugated a cell adhesive peptide, CRGDSP, to a 15% functionalized HPMC-allyl material, providing an RGD-dense biopolymer that

could be introduced into PNP hydrogels. We found that doping this HPMC-RGD into traditional HPMC-C12 PNP hydrogels for cell encapsulation led to improved cell morphology over traditional nonfunctionalized PNP hydrogels (ESI Fig. S5†). Although outside of the scope of the current study, this functionalization strategy offers exciting potential to improve cell biocompatibility and elicit specific cellular behaviors through the incorporation of diverse peptides.

2.3 Rheological properties of PNP library

With a series of HPMC derivatives in hand, corresponding PNP hydrogels were prepared to investigate the effects of pendant group identity on rheological properties. Previous reports have demonstrated that increasing the hydrophobicity HPMC modifications greatly improves the strength of the supramolecular interactions with polymer nanoparticles, leading to stiffer materials.^{12,23} Based on these observations, we hypothesized that our library with well-matched functionalization would allow us to better detect nuances in the polymer–nanoparticle interactions *via* oscillatory shear rheometry.

Hydrogels were prepared by combining 1 wt% of each HPMC-X derivative with 5 wt% PEG-*b*-PLA NPs (Fig. 1e) in phosphate buffered saline (PBS) (Fig. 1e). For simplicity, PNP hydrogels are named after the “X” functionalization of the HPMC. Unmodified HPMC and dodecyl-modified HPMC following previously reported protocols¹² were used for control PNP hydrogels denoted “UM” and “C12”, respectively. Upon mixing, all PNP derivatives form solid-like materials, with storage moduli (*G'*) greater than loss moduli (*G''*) across all frequencies tested (Fig. 2a, b and ESI Fig. S6b–S16b†). In contrast, without the addition of nanoparticles, frequency sweeps revealed liquid-like character for all 1 wt% solutions of HPMC-X derivatives (ESI Fig. S6a–S16a†). These frequency sweeps do reveal the presence of polymer–polymer interactions due to hydrophobic modifications as increased moduli and decreased tan δ values in comparison to unmodified HPMC. Additionally, polymer solutions possessed increasing opacity with increasing hydrophobic modification, indicative of some interaction between pendant groups (ESI Fig. S4†). Despite these modest contributions, PNP properties are predominated by the polymer–nanoparticle interactions as evidence by the significant increases in moduli and elastic-like behavior following addition of PEG-*b*-PLA NPs (Fig. S6b–S16b†). We also prepared hydrogels out of unmodified HPMC exposed to the radical reaction conditions, and observed no changes in the rheological properties, highlighting the mild nature of the click conditions (ESI Fig. S17†). Satisfyingly, we found that increasing pendant chain length led to a gradual increase in PNP hydrogel modulus likely due to increased interactions between the hydrophobic pendant groups and the PEG-*b*-PLA NPs. We also found that nuances between sterically bulky adamantyl groups were easily detected in comparison to linear C3-S-C10; although, no impact was noted when comparing the smaller C3-S-*t*Bu and C3-S-C4 groups. A decrease in stiffness was also observed for C3-S-Bz groups over linear C3-S-C6, indi-



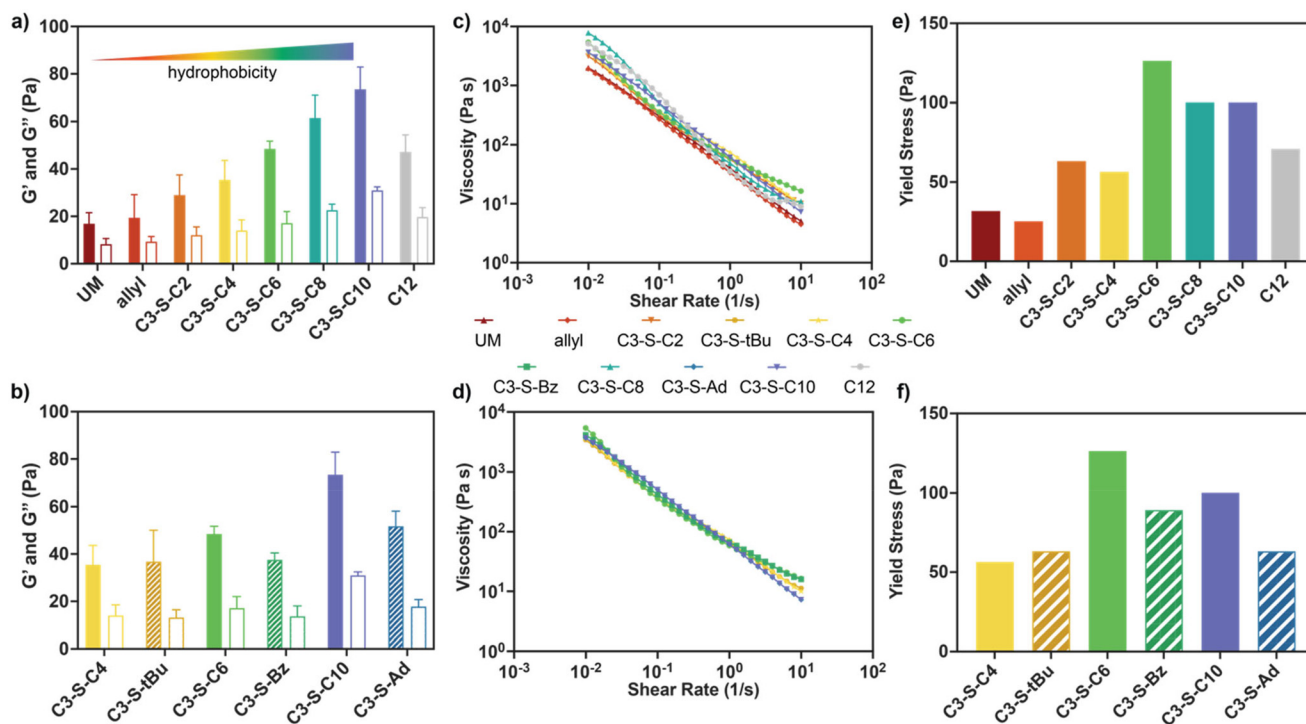


Fig. 2 Rheological properties of PNP library. Average storage (G' , filled bars) and loss (G'' , empty bars) moduli of (a) linear and (b) steric/pi stacking PNP derivatives at a representative angular frequency in the linear viscoelastic regime ($\omega = 1 \text{ rad s}^{-1}$, 2% strain). Values reported are means $\pm$ SE of $n = 3$. Steady shear rheology of (c) linear and (d) steric/pi stacking PNP derivatives. Yield stresses of (e) linear and (f) steric/pi stacking PNP derivatives determined from stress ramps where viscosity dropped below half of the initial plateau viscosity.

cating competitive pi stacking between HPMC chains might reduce the overall number of polymer–nanoparticle interactions contributing to the material properties. In comparison to polymer–nanoparticle interactions, however, we found that pi stacking interactions alone did not appear to enhance the storage modulus (ESI Fig. S18†). As previously reported, we also demonstrated that increasing the concentration of polymer and nanoparticle components provides a route to further modulate stiffness.¹⁵ Thus, hydrogels prepared with 2 wt% HPMC-X and 10 wt% PEG-*b*-PLA NPs provide a stiffer library while maintaining the trends observed with lower concentration hydrogels (ESI Fig. S20†). Interestingly, in amplitude sweeps of all PNP hydrogel derivatives, we observed characteristic G'' peaks at the crossover with G' , indicative of a jamming transition between polymer–nanoparticle units (ESI Fig. S6d–S16d†).²⁴ Observation of this transition had been reported for PNP hydrogels prepared with HPMC-C12, but previous synthetic routes and rheological methods had precluded the observation of this phenomenon when using other HPMC derivatives.²³

The ability for PNP hydrogels to be readily injected then form robust depots *in vivo* has enabled versatile biomedical applications to be realized; thus, we sought to confirm that these essential properties were preserved across the new PNP library. First, we conducted steady shear rheometry, where we were pleased to observe the viscosities for all materials decreased multiple orders of magnitude with increasing shear

rates (Fig. 2c and d). We then ran stress ramp measurements to determine the yield stress for each derivative, a predictor of depot persistence *in vivo*.^{14,25} Similar to the trends observed in moduli, we found that increased pendant hydrophobicity led to higher zero shear viscosities and yield stresses (Fig. 2e and ESI Fig. S21†), indicative of stronger interactions between nanoparticles with pendant groups of increasing chain lengths. Likewise, steric and pi stacking interference led to decreased zero shear viscosities and yield stresses compared to linear analogs (Fig. 2f and ESI Fig. S21†).

Flow sweeps and stress ramp measurements provided confirmation of the hydrogels' ability to flow under high shear; however, we wished to further investigate the dynamics of the polymer–nanoparticle interactions which would be applicable to depots post-injection. Even after the cessation of high shear, dynamic supramolecular interactions allow PNP hydrogels to exhibit viscoelastic behaviors, which play important roles on depot persistence and cargo delivery when exposed to the *in vivo* environment.²⁵ Given the observed differences in stiffness and shear-thinning properties across our PNP library, we predicted our stronger interactions would slow the rate of network dissociation and lead to observed differences in viscoelastic behaviors like stress relaxation.^{26,27} Stress relaxation experiments revealed that all PNP hydrogels readily relax stress (Fig. 3a and b), with characteristic relaxation times on the order of 10^2 – 10^3 seconds that increased with pendant hydrophobicity (Fig. 3c and d). Interestingly, C3-S-Bz pendants led



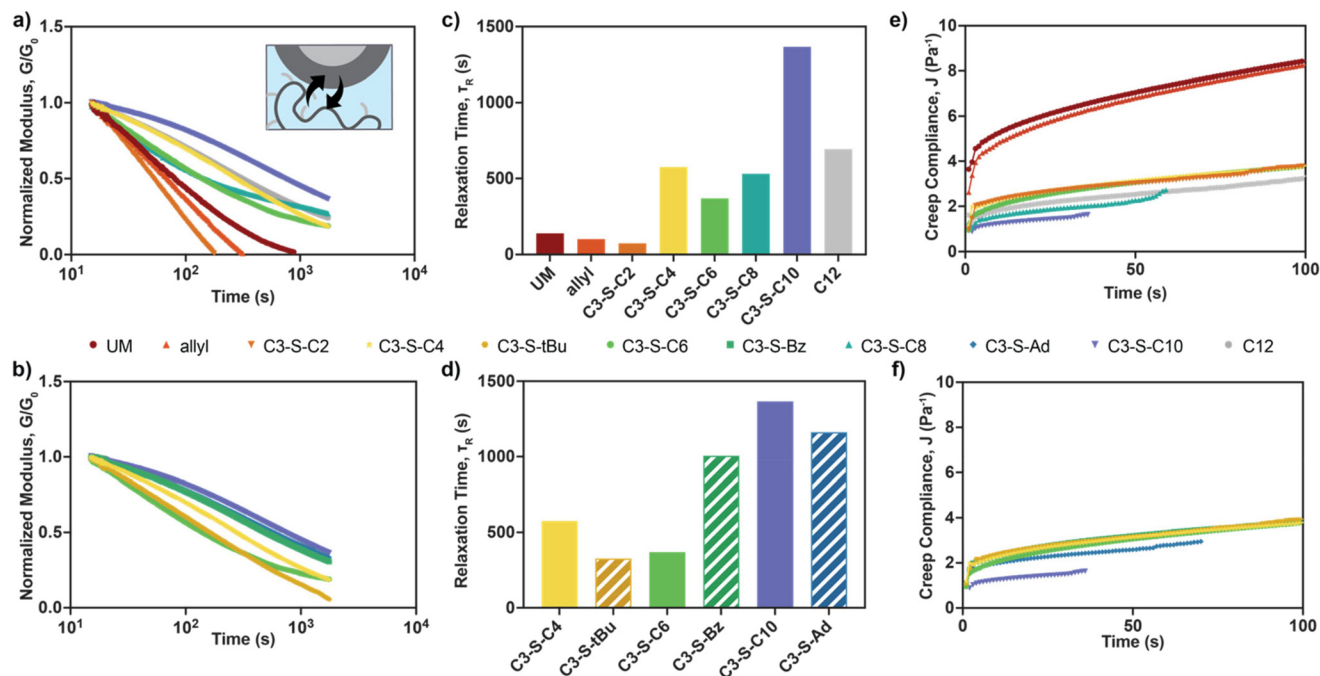


Fig. 3 Dynamic properties of PNP library. Stress relaxation measurements of (a) linear and (b) steric/pi stacking PNP derivatives (5% strain). Relaxation times, τ_R , of (c) linear and (d) steric/pi stacking PNP derivatives determined from fitting of relaxation curves in (a) and (b). Creep compliance of (e) linear and (f) steric/pi stacking PNP derivatives under a constant applied stress of 20 Pa.

to an unexpected increase in relaxation time, which could be attributed to breaking of pi stacking interactions. Fitting of frequency sweep data to continuous viscoelastic relaxation spectra was also conducted to determine G'/G'' crossovers for each PNP hydrogel derivative, revealing similar, but less dramatic trends compared to stress relaxation measurements (ESI Fig. S23 and 24†). We next conducted creep tests to investigate material deformation under a subcutaneously relevant stress of 20 Pa, an ideal predictor of depot flattening and retention for *in vivo* mouse models.²⁵ We observed rapid deformation of PNP hydrogels prepared with unmodified HPMC and allyl pendants, while increasing pendant chain lengths again provided a satisfying trend, with the most hydrophobic C3-S-C10 material exhibiting the lowest compliance (Fig. 3e). For sterically hindered and pi stacking pendants, notable differences in creep behavior were only observed between C3-S-Ad and its linear analog C3-S-C10 (Fig. 3f). To test stress values relevant to other organisms, we also investigated the creep response of each PNP hydrogel derivative under applied stresses up to 100 or 200 Pa and found similar trends (ESI Fig. S25 and 26†).

In addition to probing network dissociation dynamics, we were also interested in the thixotropic behaviors of hydrogels: an important consideration for rapid depot formation after injection and prevention of burst release.²⁸ To investigate these behaviors, we conducted stress overshoot experiments to obtain characteristic material recovery times. Stress overshoot experiments were conducted as previously described,¹⁶ where stress overshoots during shear were measured following

increasing wait times (Fig. 4a–c). Fitting of stress overshoots to an exponential plateau provides characteristic timescales for this healing behavior, τ_H .^{16,29} These experiments revealed that long, linear pendant groups C3-S-C8, C3-S-C10 and C12 required nearly twice the time to heal as all other derivatives (Fig. 4d). Small differences were also noted when comparing linear derivatives to their sterically hindered and pi stacking counterparts (Fig. 4e). These results indicate that although these pendant groups ultimately form stronger polymer–nanoparticle interactions, the rate of association is also significantly slowed. This observation highlights that there could be an important tradeoff between material retention time and initial burst release due to slowed network recovery post-injection.

2.4 Cargo diffusion and release

Previous work has established that timescales of biotherapeutic cargo delivery can be readily tuned through altering PNP hydrogel network density;¹⁵ however, this strategy simultaneously changes both cargo diffusivity and hydrogel lifetime, limiting the overall control over cargo release. In contrast, we hypothesized that through chemical modification of the polymer–nanoparticle interaction we would effectively maintain mesh size and cargo diffusivity, providing control over exposure exclusively through material retention. To test this, we investigated the diffusivity of a model cargo, bovine albumin serum (BSA) across the PNP library (Fig. 5a) using fluorescence recovery after photobleaching (FRAP) experiments. PNP hydrogels loaded with 0.05 wt% fluorescein-labeled BSA as a model were prepared, and fluorescence recov-

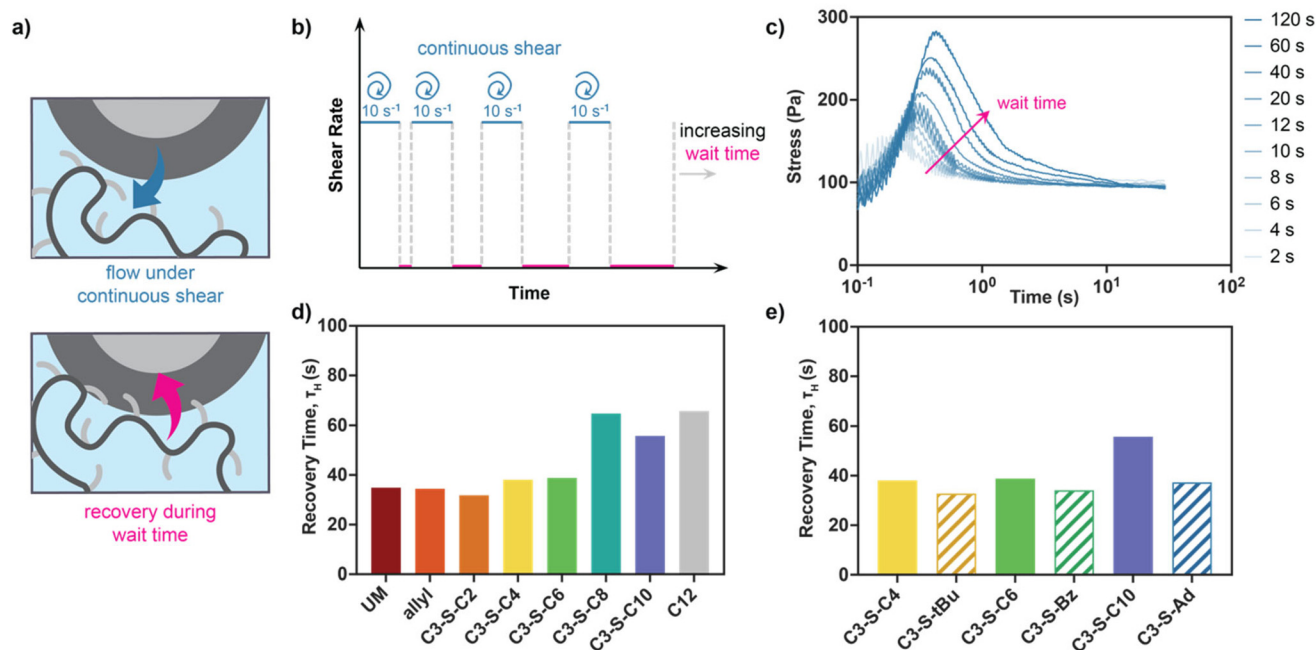


Fig. 4 Recovery properties of PNP library. (a) Simplified depiction of polymer–nanoparticle interaction under flow and wait time conditions. (b) Diagram describing experimental protocol to determine recovery times. (c) Representative stress overshoot measurement of a C3-S-Ad PNP hydrogel with increasing wait times. Recovery times, τ_r , of (d) linear and (e) steric/pi stacking PNP derivatives determined by fitting stress overshoots.

ery was monitored after bleaching with high intensity light. Diffusivity values for BSA within each PNP hydrogel derivative were then determined from these recovery curves. We found that across all derivatives, diffusion is more than an order of magnitude slower than calculated for PBS – indicative of successful entrapment within the hydrogel mesh (Fig. 5b). Additionally, minimal differences in diffusivity were observed between the most mechanically distinct PNP hydrogels UM and C3-S-C10, indicating cargo release could be tuned selectively through material erosion properties rather than varied cargo diffusivity. To further investigate release, PNP hydrogels containing BSA conjugated to AlexaFluor 647 (0.025 wt%) were incubated in phosphate buffered saline at 37 °C for two weeks while media was monitored *via* fluorescence. As illustrated in Fig. 5c, all derivatives possess similar diffusional release profiles with less than 30% release over two weeks. Together, these release experiments suggest that using this chemical modification strategy will allow for exquisite control over biotherapeutic cargo exposure purely through polymer–nanoparticle interaction strength, rather than network density.

In addition to delivery of large biotherapeutics which can be trapped within the hydrogel mesh and slowly delivered, PNP hydrogels also offer a route for the sustained release of hydrophobic small molecule cargo which can be incorporated within the hydrophobic cores of PEG-*b*-PLA NPs.¹² Of note, our group has recently demonstrated the promise of PNP hydrogels for the extended delivery of hydrophobically modified glucagon-like peptide 1 receptor agonists, semaglutide and liraglutide, for the prolonged treatment of type 2 diabetes in rat models following a single hydrogel injection.³⁰ These previous

studies have suggested that hydrophobic cargo conservation within PNP hydrogels can be explained by adsorption to the PEG-*b*-PLA NP cores; however, the impact of HPMC hydrophobicity has not been considered. To probe these effects, we also conducted a release study comparing the release of a fluorescently conjugated lipid, DSPE-Fluor 488, across our PNP hydrogel library (Fig. 5d). In contrast to the similar release profiles observed for BSA cargo, which were dictated by mesh size, we found that release rates of this lipid containing model cargo were more dependent upon the inclusion of hydrophobic pendant groups on the HPMC (Fig. 5e). These results reveal that the hydrophobic HPMC-C12 used in previous studies likely played a crucial role in the sustained delivery of hydrophobic small molecule cargo, which was previously attributed solely to interactions with PEG-*b*-PLA NP cores. In addition to these insights about the mechanism of molecular delivery from PNP hydrogels, modulation of HPMC hydrophobicity provides a new strategy to tailor the pharmacokinetics of hydrophobic small molecule release from these hydrogels that had not previously been accessible.

2.5 *In vivo* material retention

Having demonstrated that PNP hydrogel mechanical properties are directly related to the polymer–nanoparticle affinity through HPMC derivatization we proposed that we could also tune material retention *in vivo*. Four derivatives were chosen to test that spanned the range of physical properties obtained from the library: UM, C3-S-C4, C3-S-C10, and the previously reported C12 (Fig. 6a–c). Female C57BL/6 mice were injected subcutaneously with PNP hydrogels and depots were measured



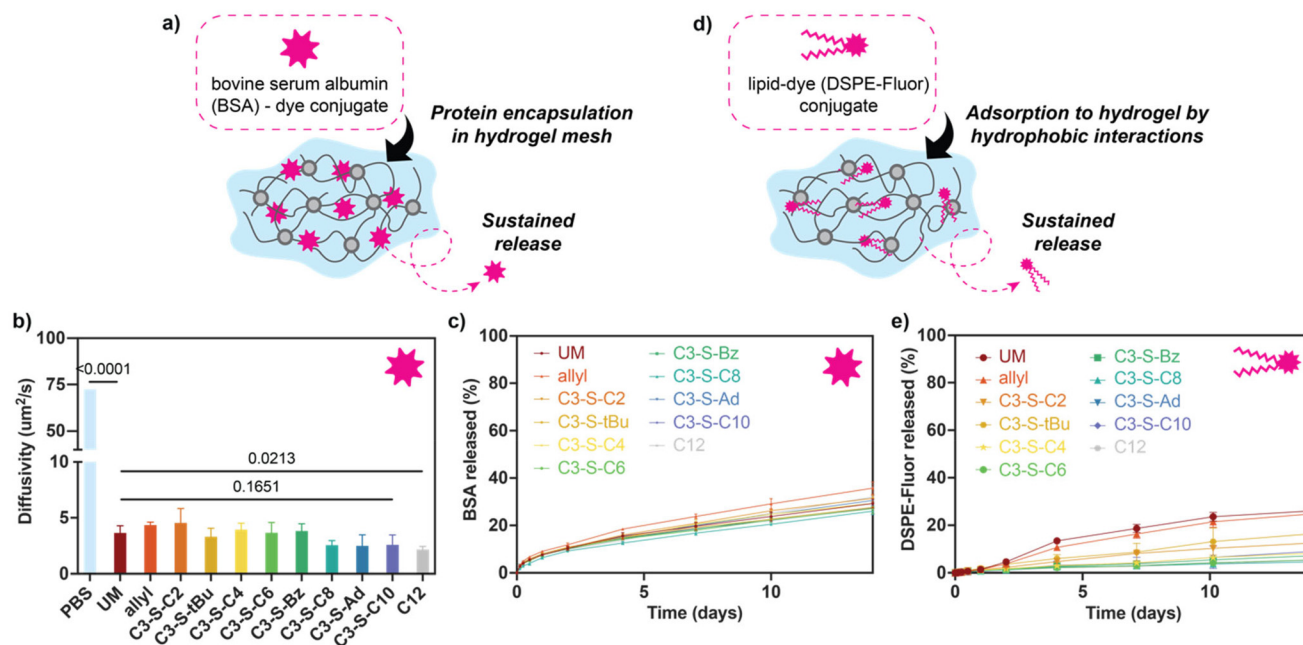


Fig. 5 (a) Schematic depiction of sustained BSA release. (b) Bar chart plotting diffusivities of BSA-FITC determined by FRAP ($n = 3$) for each PNP hydrogel derivative. PBS value calculated by Stokes–Einstein equation. Statistical analysis between groups was run using GraphPad Prism using an unpaired t-test with two-tails. (c) *In vitro* release of BSA-AlexaFluor 647 from each PNP hydrogel derivative into PBS over two weeks at 37 °C. Values reported are means $\pm$ SE of $n = 3$. (d) Schematic depiction of sustained DSPE-Fluor release. (e) *In vitro* release of DSPE-Fluor 488 from each PNP hydrogel derivative into PBS over two weeks at 37 °C. Values reported are means $\pm$ SE of $n = 3$.

with calipers at designated time points for 70 days, at which point mice were euthanized and remaining depots were excised to assess fibrosis and mass retention (Fig. 6d). As highlighted in Fig. 6e–g, creep and yielding behaviors remain a good predictor of *in vivo* retention.²⁵ Notably, our UM hydrogels which exhibit the greatest creep compliance under subcutaneously relevant stresses led to the most rapid depot dissolution, with all five undetectable by day 46. The next most compliant material, C3-S-C4, had improved material retention, with only one depot still observable by the end of the 10-week study. In contrast, the most robust C3-S-C10 and C12 materials maintain observable depots through the endpoint of the study. At day 70, excisions of all remaining depots were conducted and revealed no fibrotic capsule formation (ESI Fig. S29†). C3-S-C10 and C12 explants were easily removed with 9.2% and 11.4% mass retention, respectively (Fig. 6h). In contrast, C3-S-C4 had lower mass retention of 4.1%, with only 3 detectable gels excised. Likewise, only one explant was obtained from the UM group, with an overall mass retention of 0.2%. This *in vivo* work highlights the tunability of PNP hydrogel retention times afforded by this platform, including access to depots that dissolve much faster than the previously reported HPMC-C12-based formulation,¹² yet maintain low diffusivity as demonstrated in section 2.3. Thus, this newfound control has the potential to provide well-matched dissolution rates with desired drug release profiles. Notably, faster dissolving hydrogels could also circumvent the need for multiple injection

sites necessitated by hydrogel persistence beyond drug dosing regimens, a common challenge with depot technologies.

3. Conclusions

Methods to gain control over mechanical properties and delivery timescales from PNP hydrogels can have critical impacts on therapeutic efficacy. For example, physical PNP hydrogel properties play a key role in the viability of adoptive cells¹³ and the magnitude of immune responses.^{14,15,31} We anticipate that this thiol-ene derivatization strategy will allow for greater control over mechanical properties and delivery profiles by specifically modifying the strength of polymer–nanoparticle interactions without altering hydrogel mesh size or component concentrations. Notably, by conducting a thorough investigation of rheological properties of our PNP hydrogel library, we have observed an interesting tradeoff between material retention time and recovery post-shear. Given that initial burst release from hydrogels after injection has remained an unmet problem,²⁸ increasing recovery rates of materials post-injection might be a tunable handle to maintain drug exposure within the desired therapeutic window. Thus, future investigation into intermediate formulations that provide sustained release while offering more rapid recovery post-injection could reduce adverse effects associated with burst release.



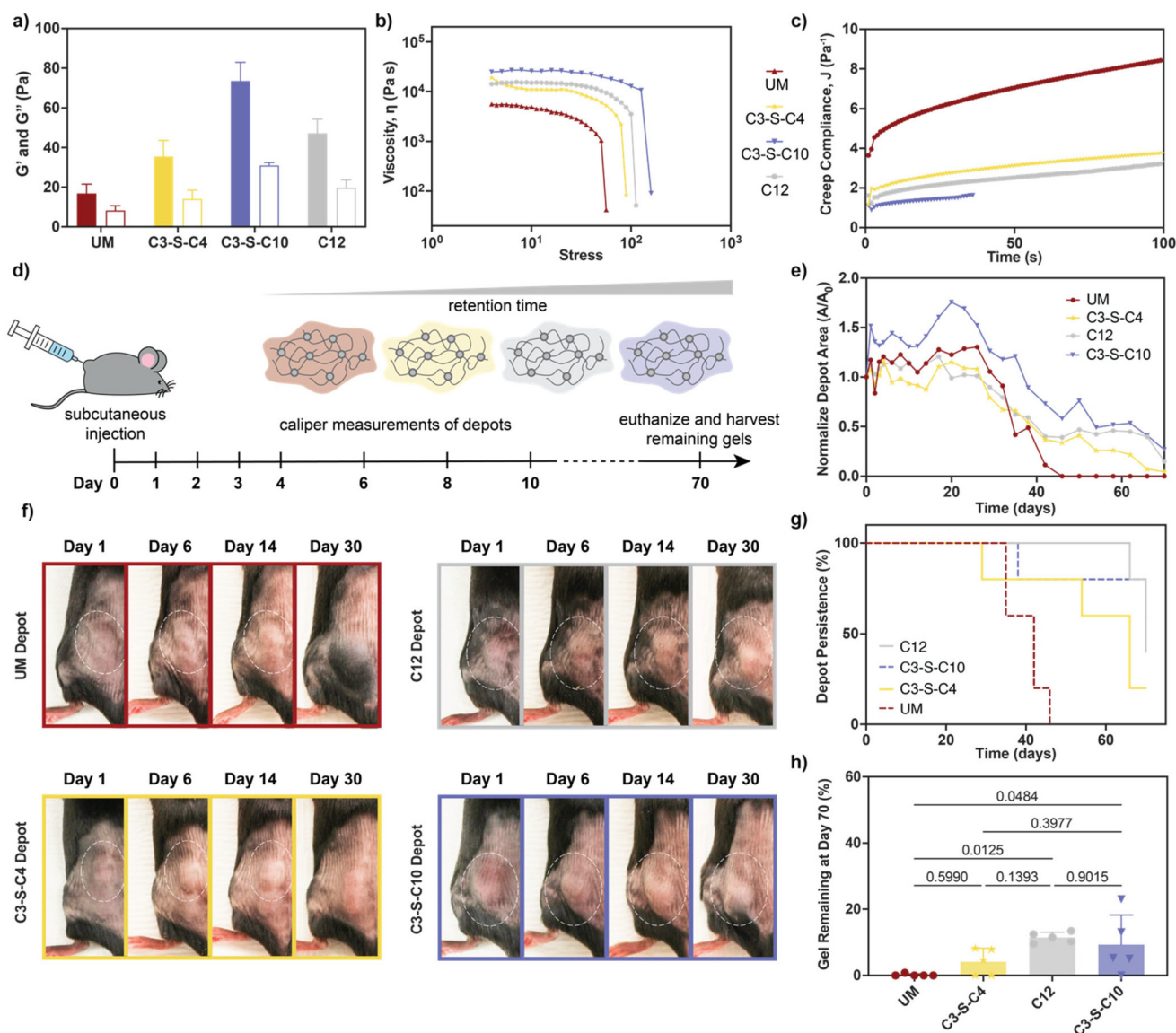


Fig. 6 *In vivo* hydrogel retention. (a) Bar chart comparing average storage (G' , filled bars) and loss (G'' , empty bars) moduli of PNP derivatives investigated *in vivo* ($\omega = 1 \text{ rad s}^{-1}$, 2% strain). Values reported are means $\pm$ SE of $n = 3$. (b) Comparison of stress sweeps of PNP derivatives investigated *in vivo*. (c) Comparison of creep compliance of PNP derivatives investigated *in vivo* when exposed to subcutaneously relevant 20 Pa stress. (d) Timeline of *in vivo* experiment. (e) Normalized observable depot areas over 70 days. (f) Representative images of hydrogel depots of each PNP hydrogel derivative over 30 days. (g) Fraction of mice with observable depots over 70 days. (h) Gel mass remaining at day 70 measured from *in vivo* explants. Statistical analysis was run using GraphPad Prism using one-way ANOVA test with a Tukey's multiple comparisons test across multiple groups. Values reported in plots d and h are means $\pm$ SE of $n = 5$.

Additionally, as previewed by our demonstration of RGD functionalization, this synthetic strategy is easily translatable to any cysteine-bearing peptide. Not only does this provide a route to easily functionalize with cell adhesive moieties that can improve cell biocompatibility, but it also opens exciting opportunities in the covalent attachment of peptides and small molecules with broad functions to the PNP scaffold. Small and hydrophilic cargo have traditionally been incompatible with delivery *via* PNP hydrogels as they readily diffuse out of the comparatively large mesh. Thus, strategies to incorporate small molecule agents have required burdensome conju-

gation to PEG-*b*-PLA nanoparticles.^{22,32,33} Thus, this HPMC modification strategy offers a new modular route to the delivery of thiol containing biologically active small molecules and peptides *via* PNP hydrogels.

In summary, we have presented a facile strategy for tuning PNP hydrogels based on facile thiol-ene click HPMC derivatization. This method takes advantage of the wealth of commercially available thiol containing compounds and cysteine-bearing peptides, greatly expanding the scope of previously accessible polymer-nanoparticle interactions that can be investigated. We used rheometry to probe the impact of HPMC

pendant group identity on bulk material properties, and demonstrated that stiffness, flow and recovery behaviors could be readily tuned by changing HPMC pendant group identity. Finally, we showcase that these tunable mechanical properties could be leveraged to provide varied hydrogel retention times *in vivo*, highlighting the potential for this strategy for tailored therapeutic delivery.

4. Experimental

4.1 Materials

Hydroxypropylmethylcellulose 2910 (HPMC) was purchased from Sigma Aldrich (Product Number H3785). CGRGDSP peptide was custom ordered from Peptide 2.0. Unless otherwise noted, chemicals were purchased from Sigma Aldrich, TCI Chemicals or Fisher Scientific and used as received without further purification.

4.2 Polymer characterization

All ^1H NMR spectra were recorded on a Bruker Neo 500 MHz spectrometer and are reported relative to deuterated dimethyl sulfoxide ($\text{DMSO-}d_6$, $\delta = 2.50$ ppm) or chloroform (CDCl_3 , $\delta = 7.26$ ppm). Average molecular weight and dispersity values were determined by size exclusion chromatography (SEC) eluting with *N,N*-dimethylformamide (0.01 M LiBr) and calibrated relative to poly(ethylene glycol) standards. Separation was done through two Jordi Labs Resolve Mixed Bed Low Divinylbenzene (DVB) columns in series and data was collected by a Dionex Ultimate 3000 Variable Wavelength detector and RefractoMax521 RI detector.

4.3 Synthesis of HPMC derivatives

4.3.1 HPMC-allyl Preparation. To an oven-dried round bottom flask containing stirred *N*-Methyl-2-pyrrolidone (NMP) (280 mL, anhydrous) was slowly added HPMC (7 g, ~ 30 mmol cellulose repeat units). The solution was allowed to stir at room temperature under inert atmosphere until HPMC had fully dissolved. The solution was then heated to 50°C before adding *N,N*-Diisopropylamine (0.89 mL, 5.09 mmol, 0.17 eq. per repeat) dropwise. In a separate vial, allyl isocyanate (0.82 mL, 3.6 mmol, 0.12 eq. per repeat) was pre-mixed with NMP (10 mL NMP) before it was added dropwise to the stirred HPMC solution. Once added, the reaction was allowed to cool to room temperature and stir under nitrogen atmosphere for and additional 24 hours. The material was isolated by precipitation into a stirred mixture of acetone/methanol (70 : 30, 2 L). The isolated solid was then washed thoroughly with acetone before it was dried under reduced pressure. For thiol-ene click reactions, this HPMC-allyl was taken forward without additional purification. HPMC-allyl used for subsequent rheological, *in vitro* and *in vivo* experiments was further purified by dialysis against Milli-Q water (MWCO = 3.5 kDa) for 2 days followed by lyophilization. Allyl incorporation was determined to be 4% by ^1H NMR.

4.3.2 General procedure for thiol-ene click. To a 20 mL scintillation vial equipped with stir bar, HPMC-allyl (300 mg, ~ 1.2 mmol cellulose repeat units, ~ 0.14 mmol ene) and the desired thiol (7.5 eq. per ene) were dissolved in NMP (12 mL). A stock solution of 2,2-dimethoxy-2-phenylacetophenone in NMP (185 mg mL^{-1}) was then added to the vial (0.1 mL, 0.07 mmol, 0.5 eq. per ene) before irradiating in an LZC-4V photoreactor at 355 nm (4 mW cm^{-2}) for 90 minutes. After irradiation the material was isolated by precipitation into acetone (200 mL). The isolated solid was then washed thoroughly with acetone before it was dissolved in Milli-Q water (30 mL) and transferred into hydrated 3.5 kDa MWCO dialysis tubing. The polymer was purified by dialysis against Milli-Q water (1 L) for 2 days followed by lyophilization. Quantitative conversion of allyl groups was confirmed by ^1H NMR.

4.4 Synthesis of poly(ethylene glycol)-*b*-poly(lactic acid) (PEG-*b*-PLA)

PEG-*b*-PLA was prepared by ring opening polymerization of D,L -lactide from poly(ethylene glycol) monomethyl ether (PEG-OMe) (5 kDa, Sigma Aldrich) similar to previous reports.^{12,16,25} To ensure exclusion of water, the following purifications were conducted prior to polymerization: 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) was purified by distillation under reduced pressure and stored over molecular sieves until use; D,L -lactide was recrystallized 3 times from anhydrous ethyl acetate; PEG-OMe was dried at 100°C under vacuum for 3 hours directly before use; and dichloromethane (DCM) was dispensed from a solvent purification system immediately before use. In an oven-dried round bottom flask under nitrogen atmosphere, D,L -lactide (8 g, 55.5 mmol, 139 eq.) was dissolved in DCM (36 mL). Once dissolved, PEG-OMe (2 g, 0.4 mmol, 1 eq.) dissolved in DCM (4 mL) was added. To initiate the polymerization a stock solution of DBU (150 μL mL^{-1} DCM) was quickly added (400 μL stock, 0.4 mmol, 1 eq.) and the polymerization was conducted for 8 minutes. The polymerization was then terminated by addition of 0.5 mL stock solution of acetic acid in acetone (3–4 drops per mL). Conversion was determined to be 90% ($M_{n,\text{theor}} = 23\,000$ Da) by ^1H NMR. The polymer was purified by precipitation into a 1 : 1 mixture of hexanes/diethyl ether (400 mL). Molecular weight was determined by ^1H NMR: $M_{n,\text{NMR}} = 24\,600$ Da and GPC (DMF) $M_{n,\text{GPC}} = 21\,000$ Da, $D = 1.09$.

4.5 Nanoparticle preparation and characterization

PEG-*b*-PLA NPs were prepared within 24 hours of use by nanoprecipitation as previously reported.^{12,16,25} A solution of PEG-*b*-PLA (50 mg mL^{-1}) was prepared in a mixture of acetonitrile and dimethyl sulfoxide (75 : 25). For each batch of nanoparticles, 1 mL of this PEG-*b*-PLA solution was added dropwise into 10 mL Milli-Q water stirred at 600 rpm. Nanoparticles were then concentrated by centrifugal filtration (MWCO 10 kDa) and resuspended in PBS to afford a 20 wt% stock solution. Hydrodynamic diameter and dispersity of nanoparticle batches were assessed by dynamic light scattering (DynaPro II



plate reader, Wyatt Technology): $D_H = 28\text{--}32\text{ nm}$, $PDI < 15\%$ for all batches.

4.6 General PNP hydrogel preparation

PNP hydrogels were prepared similar to previous reports.^{12,16,25} Stock solutions of each HPMC-X were prepared by dissolving at 6 wt% in PBS over several days on a nutator with additional agitation introduced as needed to ensure sufficient dissolution. Once dissolved, stock solutions were stored at 4 °C until use. For rheological experiments, hydrogels (600 μL) were prepared by mixing 6 wt% HPMC-X stock solutions (100 mg), 20 wt% PEG-*b*-PLA NPs stock solution (150 μL) and PBS (350 μL) in Eppendorf tubes to afford final concentrations of 1 wt% HPMC-X and 5 wt% PEG-*b*-PLA. Gels were periodically placed on a benchtop centrifuge to remove bubbles during mixing. For *in vitro* and *in vivo* experiments, PEG-*b*-PLA NPs stock solution, PBS, and dyes were loaded into one syringe, HPMC-X was loaded into a second syringe, and the components were mixed using an elbow connector. After mixing, the elbow mixer was replaced with a needle for injection into capillary tubes or mice.

4.7 Rheology

Rheological experiments on PNP hydrogels were conducted at 25 °C using a 20 mm diameter serrated parallel plate at a 500 μm gap on a stress-controlled TA Instruments DHR-2 rheometer equipped with solvent trap. For 1 wt% HPMC-X solutions, testing was conducted using a 40 mm, 2.018° cone and plate geometry. Frequency sweeps for all PNP hydrogels were performed at a strain of 1% within the linear viscoelastic regime. Flow sweeps were performed from high to low shear rates with steady state sensing. Stress controlled flow measurements were performed from low to high stress with steady state sensing and 10 or 20 points per decade. For stress relaxation experiments, a 5% step strain was applied, and the corresponding stress and modulus was measured for 1000 s. Relaxation times were determined by fitting to Kohlrausch's stretched-exponential relaxation model³⁴ in GraphPad Prism (ESI Fig. S22†). Creep experiments measured strain rate at fixed stress. An initial stress of 0.5 Pa was applied to the material for 20 s. The desired stress (5–200 Pa) was then applied, and corresponding strain percent was measured for 300 s. Strain rate at a given stress was obtained by fitting a slope for the linear region of the strain vs. time curve in GraphPad Prism. Stress overshoot experiments were conducted similar to previously described.¹⁶ For each step, a flow step was applied (10 s^{-1}) followed by a specified wait time (2, 4, 6, 10, 12, 20, 40, 60 or 120 s) before moving onto the next step (Fig. 4b). Characteristic relaxation times were determined by fitting the stress overshoot as a function of wait time to an exponential plateau in GraphPad Prism (ESI Fig. S27 and 28†).

4.8 Fluorescence recovery after photobleaching (FRAP) analysis

FRAP analysis was conducted as previously described.¹⁵ PNP hydrogels containing 0.5 mg mL^{-1} BSA-FITC (Sigma) were pre-

pared, placed on glass slides and imaged using a confocal ZEISS LSM780 microscope ($n = 3$ per hydrogel type). Samples were imaged to obtain an initial level of fluorescence (excitation = 488 nm) before a high intensity laser (488 nm) was used to bleach a region of interest (ROI) with 12.5 μm diameter for 10 s. Fluorescence data was then recorded for 200 seconds to obtain an exponential fluorescence recovery curve. Diffusivity values were calculated according to,

$$D = \frac{\gamma_D \omega^2}{4\tau_{1/2}}$$

where ω is the radius of the bleached ROI and the constant γ_D is calculated as $\tau_{1/2}/\tau_D$. Both $\tau_{1/2}$, the half-time of recovery, and τ_D , the characteristic recovery time, are given by the ZEN software.

The diffusivity of BSA in PBS was calculated according to the Stokes–Einstein equation,

$$D = \frac{k_B T}{6\pi\eta R_H}$$

where k_B is Boltzmann's constant, T is temperature in Kelvin, η is solvent viscosity (0.8872 mPa s for PBS at 25 °C),³⁵ and R_H is the hydrodynamic radius (3.4 nm for BSA).³⁶

4.9 *In vitro* capillary release

Glass capillary tubes (inner diameter = 3 mm) were plugged at one end with epoxy before 100 μL of each PNP hydrogel containing 25 μg BSA-Alexa 647 (Invitrogen) or 0.5 nmol DSPE-Fluor 488 (BroadPharm) was injected ($n = 3$). Hydrogels were carefully topped with 200 μL of PBS and capillaries were sealed with parafilm and incubated at 37 °C. At each time point, the PBS was removed and replaced with fresh PBS. Release into PBS was determined by fluorescent measurement (650/668 nm for BSA-Alexa 647, 490/525 nm for DSPE-Fluor 488) relative to calibration curve of known concentrations.

4.10 *In vivo* methodology

Seven-week-old female C57BL/6 mice were purchased from Charles River and housed in the animal facility at Stanford University. Mice were cared for following the Institutional Animal Care and Use guidelines. All animal studies were performed in accordance with the National Institutes of Health guidelines and the approval of Stanford Administrative Panel on Laboratory Animal Care (protocol APLAC-32109). Mice were shaved on both flanks prior to injections. Each mouse received two subcutaneous injections (100 μL) of PNP hydrogel on either flank while under brief isoflurane anesthesia. Depots were measured with calipers in the x and y plane at each time-point and depot areas were estimated as an ellipse. After 70 days, mice were euthanized and remaining PNP gels were excised and weighed to determined remaining mass percentage relative to initial injection.



Author contributions

S.J.B., N.E., and E.A.A. conceived the ideas. S.J.B., N.E., and E.A.A. designed the experiments. S.J.B., N.E., E.B., and C.K.J. conducted the experiments. S.J.B., N.E., and S.S. analyzed the data. S.J.B. wrote the paper. S.J.B., N.E., and E.A.A. revised the paper. All authors have given approval to the final version of the manuscript.

Data availability

All data reported in the paper are included as part of the ESI.†

Conflicts of interest

E.A.A. is listed as an inventor on several patents related to the PNP hydrogel materials discussed in this manuscript. E.A.A. is an equity holder and advisor to Appel Sauce Studios LLC, which holds an exclusive license to several patents related to the PNP hydrogel materials discussed in this manuscript. All other authors have no conflicts to declare.

Acknowledgements

The research reported here was financially supported by the Bill & Melinda Gates Foundation (INV-027411). The instrument used for ¹H NMR data acquisition was funded significantly by an NIH High End Instrumentation grant (1 S10 OD028697-01). S. J. B. would like to thank Stanford's Office for Postdoctoral Affairs for support through a PRISM Baker Fellowship. N.E. was supported by an NSF Graduate Research Fellowship (DGE-2146755). C. J. was supported by an NSF Graduate Research Fellowship. The authors would also like to thank Prof. Sarah Heilshorn for use of a Leica DMi8 microscope for cellular imaging.

References

- 1 B. V. Slaughter, S. S. Khurshid, O. Z. Fisher, A. Khademhosseini and N. A. Peppas, *Adv. Mater.*, 2009, **21**(32–33), 3307–3329.
- 2 J. Li and D. J. Mooney, *Nat. Rev. Mater.*, 2016, **1**(12), 1–17.
- 3 P. Bertsch, M. Diba, D. J. Mooney and S. C. G. Leeuwenburgh, *Chem. Rev.*, 2023, **123**(2), 834–873.
- 4 A. A. Foster, L. M. Marquardt and S. C. Heilshorn, *Curr. Opin. Chem. Eng.*, 2017, **15**, 15–23.
- 5 S. J. Bidarra, C. C. Barrias and P. L. Granja, *Acta Biomater.*, 2014, **10**(4), 1646–1662.
- 6 F. Rizzo and N. S. Kehr, *Adv. Healthcare Mater.*, 2021, **10**(1), 2001341.
- 7 R. Dimatteo, N. J. Darling and T. Segura, *Adv. Drug Delivery Rev.*, 2018, **127**, 167–184.
- 8 L. Riley, L. Schirmer and T. Segura, *Curr. Opin. Biotechnol.*, 2019, **60**, 1–8.
- 9 S. Tang, B. M. Richardson and K. S. Anseth, *Prog. Mater. Sci.*, 2021, **120**, 100738.
- 10 H. Wang, D. Zhu, A. Paul, L. Cai, A. Enejder, F. Yang and S. C. Heilshorn, *Adv. Funct. Mater.*, 2017, **27**(28), 1605609.
- 11 M. J. Webber, E. A. Appel, E. W. Meijer and R. Langer, *Nat. Mater.*, 2016, **15**(1), 13–26.
- 12 E. A. Appel, M. W. Tibbitt, M. J. Webber, B. A. Mattix, O. Veisheh and R. Langer, *Nat. Commun.*, 2015, **6**(1), 6295.
- 13 A. K. Grosskopf, L. Labanieh, D. D. Klysz, G. A. Roth, P. Xu, O. Adebawale, E. C. Gale, C. K. Jons, J. H. Klich, J. Yan, C. L. Maikawa, S. Correa, B. S. Ou, A. I. d'Aquino, J. R. Cochran, O. Chaudhuri, C. L. Mackall and E. A. Appel, *Sci. Adv.*, 2022, **8**(14), eabn8264.
- 14 G. A. Roth, E. C. Gale, M. Alcántara-Hernández, W. Luo, E. Axpe, R. Verma, Q. Yin, A. C. Yu, H. Lopez Hernandez, C. L. Maikawa, A. A. A. Smith, M. M. Davis, B. Pulendran, J. Idoyaga and E. A. Appel, *ACS Cent. Sci.*, 2020, **6**(10), 1800–1812.
- 15 O. M. Saouaf, G. A. Roth, B. S. Ou, A. A. A. Smith, A. C. Yu, E. C. Gale, A. K. Grosskopf, V. C. T. M. Picece and E. A. Appel, *J. Biomed. Mater. Res.*, 2021, **109**(11), 2173–2186.
- 16 A. K. Grosskopf, J. L. Mann, J. Baillet, H. Lopez Hernandez, A. A. A. Autzen, A. C. Yu and E. A. Appel, *Macromolecules*, 2022, **55**(17), 7498–7511.
- 17 E. A. Appel, M. W. Tibbitt, J. M. Greer, O. S. Fenton, K. Kreuels, D. G. Anderson and R. Langer, *ACS Macro Lett.*, 2015, **4**(8), 848–852.
- 18 C. Barner-Kowollik and A. J. Inglis, *Macromol. Chem. Phys.*, 2009, **210**(12), 987–992.
- 19 H. C. Kolb, M. G. Finn and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2001, **40**(11), 2004–2021.
- 20 C. E. Hoyle and C. N. Bowman, *Angew. Chem., Int. Ed.*, 2010, **49**(9), 1540–1573.
- 21 L. M. Campos, K. L. Killops, R. Sakai, J. M. J. Paulusse, D. Damiron, E. Drockenmuller, B. W. Messmore and C. J. Hawker, *Macromolecules*, 2008, **41**(19), 7063–7070.
- 22 A. K. Grosskopf, G. A. Roth, A. A. A. Smith, E. C. Gale, H. L. Hernandez and E. A. Appel, *Bioeng. Transl. Med.*, 2020, **5**(1), e10147.
- 23 A. C. Yu, A. A. A. Smith and E. A. Appel, *Mol. Syst. Des. Eng.*, 2020, **5**(1), 401–407.
- 24 A. Agrawal, H.-Y. Yu, A. Sagar, S. Choudhury and L. A. Archer, *Macromolecules*, 2016, **49**(22), 8738–8747.
- 25 C. K. Jons, A. K. Grosskopf, J. Baillet, J. Yan, J. H. Klich, O. M. Saouaf and E. A. Appel, *Adv. Funct. Mater.*, 2022, **32**(40), 2203402.
- 26 B. Marco-Dufort, R. Iten and M. W. Tibbitt, *J. Am. Chem. Soc.*, 2020, **142**(36), 15371–15385.
- 27 P. C. Cai, B. Su, L. Zou, M. J. Webber, S. C. Heilshorn and A. J. Spakowitz, *ACS Cent. Sci.*, 2022, **8**(9), 1318–1327.
- 28 S. Correa, A. K. Grosskopf, H. Lopez Hernandez, D. Chan, A. C. Yu, L. M. Stapleton and E. A. Appel, *Chem. Rev.*, 2021, **121**(18), 11385–11457.
- 29 C. J. Dimitriou and G. H. McKinley, *Soft Matter*, 2014, **10**(35), 6619–6644.
- 30 A. I. d'Aquino, C. L. Maikawa, L. T. Nguyen, K. Lu, I. A. Hall, C. K. Jons, C. M. Kasse, J. Yan, A. N. Prossnitz,



- E. Chang, S. W. Baker, L. Hovgaard, D. B. Steensgaard, H. B. Andersen, L. Simonsen and E. A. Appel, *Cell Rep. Med.*, 2023, **4**(11), 101292–101292.
- 31 B. S. Ou, O. M. Saouaf, J. Yan, T. U. J. Bruun, J. Baillet, X. Zhou, N. P. King and E. A. Appel, *Adv. Healthcare Mater.*, 2023, **12**(28), 2301495.
- 32 G. A. Roth, O. M. Saouaf, A. A. A. Smith, E. C. Gale, M. A. Hernández, J. Idoyaga and E. A. Appel, *ACS Biomater. Sci. Eng.*, 2021, **7**(5), 1889–1899.
- 33 B. S. Ou, J. Baillet, V. C. T. M. Picece, E. C. Gale, A. E. Powell, O. M. Saouaf, J. Yan, A. Nejatfard, H. Lopez Hernandez and E. A. Appel, *ACS Nano*, 2024, **18**(4), 3214–3233.
- 34 Q. Li, D. G. Barrett, P. B. Messersmith and N. Holten-Andersen, *ACS Nano*, 2016, **10**(1), 1317–1324.
- 35 C. Li, J. Ma, Q. Fan, Y. Tao and G. Li, *Chem. Commun.*, 2016, **52**(50), 7850–7853.
- 36 A. Valstar, M. Almgren, W. Brown and M. Vasilescu, *Langmuir*, 2000, **16**(3), 922–927.

