

### CFD-accelerated bioreactor optimization: Reducing the hydrodynamic parameter space

Journal:	Environmental Science: Water Research & Technology
Manuscript ID	EW-PER-09-2021-000666.R1
Article Type:	Perspective



Optimization of promising water/wastewater treatment technologies requires significant resources in terms of time, labor and cost due to complex interactions between flow, microorganisms and reactions. The use of computational fluid dynamic simulations can shrink the possible parameter space, hence decreasing scale-up optimization costs.

#### Page 2 of 14

### Journal Name

### ARTICLE TYPE

Cite this: DOI: 00.0000/xxxxxxxxx

Received Date Accepted Date

DOI:00.0000/xxxxxxxxx

# CFD-accelerated bioreactor optimization: Reducing the hydrodynamic parameter space

Yinuo Yao,\*ab Oliver B. Fringer,b and Craig S. Criddlea

Optimization of bioreactor design can be experimentally challenging because of the complex interactions between hydrodynamic and biological processes. A promising prototyping strategy is the use of computational fluid dynamic (CFD) simulations to identify preferred hydrodynamic parameter spaces. In this work, we describe CFD simulations of flow in anaerobic fluidized-bed reactors (FBRs), with a focus on bed expansion and particle size. The results reveal regimes of putative high mass transfer where the diffusion layer thickness is impacted by a combination of flow velocity and particle collisions. These regimes are observed when bed expansion is narrowed from 10-70% (typically recommended) to 40-60%. Similarly, prospects for short circuiting are minimized by constraining the Archimedes number Ar of fluidized particles to Ar > 1000 (as opposed to the common wisdom that "smaller is better"). When membranes are added to an FBR design, fluidized particles can effectively scour and clean membranes by constraining Ar to values Ar > 7000 (a minimum is required). We conclude that CFD can provide valuable insights into reactor design and operation, reducing the hydrodynamic parameter space that must otherwise be explored by laboratory and pilot-scale validation thus decreasing time and cost for system optimization.

### 2 1 Introduction

3 Sustainability is a grand challenge for the 4 21st century<sup>1</sup>. Current human civilization 5 is largely supported by linear economies in 6 which resources are extracted, used, and dis-7 carded at end-of-life. This has created enor-8 mous challenges, increasing the need for circular 9 economies based upon recycling and reuse<sup>2–5</sup>. Microbial processes play an integral role in 10 the removal of organic carbon and nutrients<sup>6–8</sup>. 11 The prevailing technology first developed at the 12 turn of the 20th century is activated sludge, a 13 process that has since been modified to enable 14 nutrient removal. Many emerging technologies 15 cannot cross the "Valley of Death" because they 16 treat tiny flows (in many cases, just milliliters 17 per day) while adoption in practical applications 18 may require treatment of tens of millions of liters 19 per day. As biological and hydrodynamic com-20 plexity increases, the "Valley of Death" becomes 21

<sup>&</sup>lt;sup>a</sup> Codiga Resource Recovery Center at Stanford, Department of Civil and Environmental Engineering, Stanford University, Stanford, CA, 94305, USA. Email: yaoyinuo@stanford.edu

<sup>&</sup>lt;sup>b</sup> The Bob and Norma Street Environmental Fluid Mechanics Laboratory, Department of Civil and Environmental Engineering, Stanford University, Stanford, CA, 94305, USA.

deeper. An example would be bioelectrochemi-22 cal processes, such as microbial fuel cells<sup>9</sup> and 23 microbial batteries<sup>10,11</sup>, technologies that have 24 been demonstrated at bench- and, in some cases, 25 pilot-scale but not full-scale. Academia is a likely 26 source for such potentially disruptive innova-27 tion but lacks access to the facilities and fund-28 ing needed for long-term testing at a meaning-29 ful scale. To date, microbially-based technolo-30 gies have largely relied upon experiments for op-31 timization, but such testing is slow and costly. 32 Not surprisingly, practitioners and utilities tend 33 to innovate incrementally using existing systems. 34 There is thus a great risk of locking-out innova-35 tion. A pathway for lower cost and more rapid 36 scale-up of promising technology is needed. 37

In general, bioreactors can be classified as ei-38 ther dispersed growth systems, where substrate 39 gradients are minimal, or attached-growth/floc-40 based systems, where appreciable substrate gra-41 42 dients drive diffusion of substrate into floc or attached biofilms as products diffuse out. The clas-43 sic dispersed-growth example is activated sludge 44 (AS), a technology that efficiently removes or-45 ganic carbon in wastewater by converting solu-46 ble organics into CO<sub>2</sub>. Active microbial biomass 47 is concentrated by settling or membrane separa-48 tion. These microorganisms are typically present 49 as discrete cells or as floc within a size range 50 of 2-20  $\mu$ m<sup>12,13</sup>. This size range confers two 51 benefits: (1) the absence of a significant diffu-52 sion layer eliminates mass transfer limitations, 53 and (2) small particles follow flow trajectories 54 with negligible disruption of the overall hydro-55 dynamics. The combined effect of these pro-56 cesses is to weaken the dependence of biologi-57 cal activity on diffusion within floc or particles 58 under well-mixed conditions, with minimal en-59 ergy consumption and minimal short-circuiting. 60 To achieve excellent mixing, vigorous bubbling 61 is used for aeration and/or mechanical mixing. 62 63 To date, most research has focused on optimization of process-related parameters such as 64 HRT, solid retention time (SRT), and on micro- 65 bial kinetic parameters with minimal hydrody- 66 namic impacts. By excluding hydrodynamics in 67 such models, reactor design and operation are 68 greatly simplified. Examples include, but are not 69 limited to, prediction of biological activity using 70 ordinary differential equations rather than par-71 tial differential equations. In contrast to dis-72 persed growth reactions, attached growth and 73 biofilm reactors are much more complex and 74 more affected by process hydrodynamics. Exam- 75 ples would include trickling filters, granular reac- 76 tors, fluidized-bed reactors, membrane-aerated 77 reactors, microbial fuel cells, and microbial bat-78 teries. For these examples, well-mixed conditions 79 do not insure a reduction in the hydrodynamic 80 parameter space. The parameter space in com- 81 plex systems (such as microbial flocs, biofilm- 82 coated particles and biofilm-coated porous mate- 83 rials, and electrically conductive sponge) is much 84 larger than in dispersed-growth reactors, and a 85 thicker diffusion layer can increase mass transfer 86 limitations. In addition, floc and BC-Ps do not 87 necessarily follow the flow and flow-particle in- 88 teractions can eventually alter the flow trajecto-89 ries, creating more complex hydrodynamics. As 90 a result, biological activity and overall treatment 91 efficiency depend upon local hydrodynamics. To 92 optimize reactor design and operation, a quan- 93 titative understanding of hydrodynamic-related 94 parameters such as particle Reynolds number, 95 porosity and Archimedes number is critical. By 96 including hydrodynamic-related parameters, the 97 total number of parameters (both hydrodynamic-98 and process-related) increases, resulting in dras- 99 tic increases in resources in terms of cost and 100 time and the number of experiments required for 101 optimization. Simultaneously, the likelihood of 102 obtaining optimal performance diminishes due to 103 the high dimensional parameter space. 104



**Fig. 1** Workflow of CFD-accelerated scale-up. The red boxes indicate conventional CFD applications for reactor optimization. The blue box highlights iterative and integrative simulation and experiments. Mesoscale refers to simulations where the size of the computational domain is on the order of meters and the shape resembles an industrial reactor. Microscale refers to simulations at scales much smaller than reactors, on the order of millimeters or even microns.

#### 105 2 Computational fluid dynamics

Computational fluid dynamics (CFD) uses nu-106 107 merical methods to study problems that involve fluid flows. Over the past few decades, advances 108 in computational power and methods have ex-109 panded the range of problems that can be ad-110 dressed using CFD. A review by Karpinska and 111 Bridgeman<sup>14</sup> has evaluated different strategies 112 and models for optimization of wastewater treat-113 ment. In wastewater treatment (Figure 1), CFD 114 can (1) prospectively preview macroscopic reac-115 tor hydrodynamics and (2) retrospectively im-116 prove current design and operation. Studies are 117 carried out sequentially by first comparing simu-118 lations with experimental results (i.e., historical 119 results from an existing system in retrospective 120 applications or from a similar system in prospec-121 tive applications) then conducting simulations by 122 varying a parameter of interest. These studies are 123 mostly conducted at the mesoscale where the size 124 of the computational domain is on the order of 125 meters and the shape resembles an industrial re-126 127 actor. The main disadvantage of this approach is loss of microscale information where microscale 128 129 refers to simulations investigating scales that are much smaller than reactors and in the order of 130 micron or millimeters (i.e., interactions between 131 small particles on an industrial fluidized-bed re-132 actor). As such, most research adopting this ap-133 proach focuses on macroscopic properties such 134 as flow short-circuiting, reactor mixing and oxy-135 gen transfer efficiency using commercial soft-136 ware<sup>15-17</sup>. For dispersed growth reactors, with 137 a reduced hydrodynamic parameter space, CFD 138 studies have focused on aeration and mixing. An-139 other useful CFD application is disinfection. In 140 this case, short-circuiting is minimized to enable 141 efficient pathogen removal, a goal that must be 142 balanced against the need for minimization of 143 disinfection byproducts<sup>18-20</sup>. In these applica-144 tions, the simulations are reactor-specific, so the 145 knowledge gained from one system does not nec-146 essarily translate to another. 147

In more complex reactors (attached-growth or 148 floc-based), the parameter space increases sig-149 nificantly due to local interactions between hy-150 drodynamics and biological activity. Aerobic flu-151 idized bed reactors require aeration and mixing 152 simulations, but also guidelines on bed expan-153 sion, particle size, and other carrier properties of 154

interest. Focusing on aeration and mixing alone 155 is unlikely to result in optimized design and per-156 formance. In addition, because most interac-157 tions occur at the microscale, ignoring microscale 158 properties (e.g., floc diffusion layer or floc-floc in-159 teractions) with a singular focus on macroscopic 160 properties (bed expansion or mixing) is also un-161 likely to lead to correct conclusions. Understand-162 ing of systems that have shared physics can expe-163 dite translation across systems. Resources spent 164 on understanding of one system would benefit 165 other similar systems. An example is the effect 166 of Archimedes number (a combination of particle 167 and fluid properties) in upward flow reactors. A 168 quantitative understanding of this number would 169 170 be beneficial to both non-fluidized granular reactors and fluidized-bed reactors. The focus of 171 this approach is not to identify the exact values 172 for optimized parameters but rather to reduce 173 the parameter space within which optimized pa-174 rameters fall. By narrowing this space, reactor-175 specific experimental studies can be more tar-176 geted, enabling more efficient optimization and 177 scale-up with fewer resources. 178

179 In this paper, we propose a new framework for bioreactor optimization: a computational strat-180 egy in which CFD is used to understand funda-181 mental interactions involving fluid flow, particles, 182 microorganisms, membranes, and other porous 183 materials. We envision that this approach will 184 enable deeper insight into the underlying physics 185 and accelerated optimization. We use the Staged 186 Anaerobic Fluidized-bed Membrane bioreactors 187 (SAF-MBR) as a case study to demonstrate the 188 feasibility and potential of this framework. 189

### 190 3 A case study: Staged Anaerobic Fluidized-191 bed Membrane Bioreactors

The SAF-MBR is a recently developed biocarrierbased anaerobic treatment technology<sup>21,22</sup>. Aeration is eliminated because the active microorganisms are obligate anaerobes that do not toler-

ate oxygen. Energy is recovered as methane, en-196 abling net energy-positive secondary treatment 197 of domestic wastewater<sup>23</sup>. Because they are 198 slow-growing, the anaerobes also generate fewer 199 biosolids for disposal. These properties make the 200 SAF-MBR more attractive than conventional aer- 201 obic processes, such as AS<sup>24</sup>. 202

The SAF-MBR consists of two reactors in series 203 with a conventional anaerobic fluidized-bed reac-204 tor (AFBR) followed by an anaerobic membrane 205 bioreactor (AnMBR). AFBRs have been widely 206 used to treat industrial wastewater where the 207 Chemical Oxygen Demand (COD) and Biochem-208 ical Oxygen Demand (BOD) are much greater 209 than domestic wastewater. In SAF-MBR 1.0, the 210 AFBR discharges to a particle-sparged membrane 211 bioreactor (P-MBR), in which fluidized granu-212 lar activated carbon (GAC) functions as both a 213 biocarrier of slow-growing microorganisms (in-214 side the GAC pores) and as a scouring agent for 215 cleansing of membranes and prevention of bio-216 fouling  $^{21,22,24,25}$ . This strategy successfully con-217 trolled membrane biofouling in a pilot-scale SAF-218 MBR<sup>21,26</sup>, but also led to particle abrasion and 219 damage of the polymeric membranes<sup>27</sup>. As noted 220 by Shin et al.<sup>27</sup>, the GAC used in the P-MBR con-221 tained two size fractions - one at 1.18 - 1.4mm 222 (29%) and a second at 1.70 -4.00mm (47%).223 Significant membrane damage occurred in the 224 lower region of the membranes, and this dam-225 age was attributed to the larger GAC fraction. 226 In subsequent pilot-scale tests of SAF-MBR 2.0<sup>23</sup>, 227 membrane sparging was accomplished with bio-228 gas bubbles rather than solid particles. 229

The hydrodynamics of fluidized-bed reactors 230 have been investigated experimentally<sup>28,29</sup> and 231 with simulations<sup>30–34</sup>, but membrane bioreactor 232 studies of microbial activity have largely focused 233 on experimental testing<sup>35,36</sup>. These studies do not 234 track particle dynamics at high volume fraction 235 (low porosity), but instead focus on macroscopic 236 behavior such as fluidization stability and expan-237

sion<sup>37,38</sup>. The range of bed expansion in flu-238 idized beds fluctuates between 20% and 70% 239 with a qualitative understanding that low expan-240 sion leads to flow short-circuiting and high ex-241 pansion leads to biofilm loss. The optimal bed 242 expansion or porosity is thus an open question. 243 At present, most studies focus on bed expansion 244 without considering the impacts of particle prop-245 erties such as Archimedes Ar (or Galilei Ga) num-246 ber on the optimal bed expansion where Ar is de-247 248 fined as

$$Ar = Ga^{2} = \frac{(s-1)gd_{p}^{3}}{v^{2}},$$
(1)

where  $s = \rho_p / \rho_f$  is particle-fluid density ratio,  $d_p$ 249 is the clean particle diameter, g is the gravita-250 tional acceleration and v is the kinematic viscos-251 ity of water. Qualitatively, small particles are pre-252 ferred to enable more efficient mass transfer and 253 enhanced biological activity. Aslam et al.<sup>26</sup> stud-254 ied the effects of three different particles (PET 255 256 beads, silica and GAC) on membrane scouring efficiency and concluded that PET beads are best. 257

#### 258 3.1 Upflow velocity and porosity

In fluidized-bed reactors, upflow velocity con-259 260 trols bed expansion and hence porosity. Understanding particle dynamics as a function of poros-261 ity gives important insights into the biological ac-262tivity and design and operation of reactors. Re-263 cently, Yao et al.<sup>32</sup> investigated particle dynam-264 ics by varying upflow velocity in simulations of 265 a monodispersed/single-size fluidized bed with 266 particle properties similar to those of pilot-scale 267 and lab-scale reactors<sup>21</sup>. Within FBRs, poros-268 ity controls both the horizontal mixing and col-269 lisions between particles. Since no horizontal 270 flow is generated at the inlet, horizontal mixing is 271 mainly due to momentum transfer from the verti-272 cal to horizontal directions due to particle fluctu-273 ations. At low porosity, most fluctuations are in-274 275 duced by weak collisions. At intermediate porosity, collisions and hydrodynamic effects become 276 equally important, leading to an increase in par-277 ticle velocity fluctuations and stronger collisions. 278 At high porosity, hydrodynamic effects dominate, 279 and collisions are diminished. 280

#### 3.1.1 Hypothetical impacts on biofilm de-281 tachment 282

Accurate quantification of biofilm detachment 283 rate provides valuable information in modeling 284 biofilm reactor dynamics, such as the height of 285 expanded beds and insight into reaction- and 286 mass-transfer limitations<sup>39</sup>. The overall biofilm 287 detachment rate  $b_t$  is modeled as a combination 288 of first-order cell decay and mechanical detach-289 ment: 290

$$b_t = b + b_{det},\tag{2}$$

where *b* is the first-order cell decay constant and 291  $b_{det}$  is the mechanical detachment rate. Typically, 292  $b << b_{det}$  for most the engineered applications 293 such that  $b_t \approx b_{det}$ . There are two types of detach- 294 ment (continuous and discrete) and three mech- 295 anisms (shear stress, abrasion, and sloughing). 296 The shear stress is due to flow, while abrasion is 297 due to collisions between particles. Since slough- 298 ing is typically described as a discrete probabilis- 299 tic event that might lead to breakup of the entire 300 biofilm<sup>40</sup>, most models do not consider it. Chang 301 et al.<sup>41</sup> modeled  $b_{det}$  as 302

$$b_{det} = -3.14 + 0.0335C_p + 19.3Re_{p,b} - 3.46\sigma$$
, (3)

where  $C_p$  is the particle concentration in the flu- 303 idized bed,  $Re_{p,b} = u_0 d_b / v$  is the biofilm-covered 304 particle Reynolds number,  $u_0$  is the upflow veloc- 305 ity in the fluidized bed,  $d_b$  is the diameter of the 306 BC-P and  $\sigma$  is the shear stress. The author as- 307 sumed that  $C_p$ ,  $Re_{p,b}$  and  $\sigma$  account for abrasion, 308 turbulence and shear stress, respectively. The 309 main challenge with this model is related to de- 310 coupling flow ( $Re_{p,b}$ ) and abrasion effects ( $C_p$ ) 311



Fig. 2 Hypothetical impacts of upflow velocity and porosity on fluidized-bed reactor modeling, design and operation.

312 where both  $C_p$  and  $Re_{p,b}$  are functions of poros-313 ity.

Nicolella et al.<sup>42</sup> constructed an empirical model for a fluidized-bed reactor based on dimensional analysis and showed that the normalized detachment rate  $\hat{b} = d_p \tilde{b}_{det} / \rho_f v$  is given by

$$\hat{b} = 1.95 \times 10^{-10} Re_{p,c}^{1.49} d_*^{2.67},$$
 (4)

where  $\tilde{b}_{det}$  is the amount of biofilm detached per 318 unit area and time,  $Re_{p,c} = d_p u_0 / v$  is the clean 319 particle Reynolds number and  $d_* = d_b/d_p$  is the 320 diameter ratio of a biofilm-covered to a clean par-321 322 ticle. An interpretation of this model is that  $Re_{p,c}$ represents the effects of clean particle-related 323 flow whereas  $d_*$  includes the effects of biofilm 324 thickness on the detachment rate. The effect of 325 flow includes both turbulence and abrasion as 326 compared to equation 3<sup>41</sup>. Interestingly, detach-327 328 ment rates that were orders of magnitude higher were observed for  $d_* = 3$  and  $Re_{p,c} = 2.3 - 2.7$ 329

which cannot be explained by this model, al-330 though one plausible explanation is that detach-331 ment occurs near the inlet where turbulence is 332 strongest. Overall, this model better parameter-333 izes the mechanical biofilm detachment rate in 334 the sense that there is much less cross-correlation 335 between parameters. Instead of considering both 336 flow and abrasion, Gjaltema et al.<sup>43</sup> assume that 337 abrasion is the only dominant detachment mech-338 anism in an airlift reactor and used a model to 339 estimate the energy of abrasion. 340

All of the above models are unable to decouple 341 flow and abrasion or are limited to abrasion. By 342 comparing high- and low-strength FBRs, Shin et 343 al.<sup>44</sup> successfully modeled low-strength FBRs by 344 assuming small  $b_{det}$ . By examining particle dy-345 namics in a fluidized bed, Yao et al.<sup>32</sup> discovered 346 that collisions dominate over hydrodynamic ef-347 fects at low porosity. At intermediate porosity, 348 both collisions and hydrodynamic effects are im-349 portant while hydrodynamic effects dominate at 350

higher porosity. The combination of effects of
wastewater strength and hydrodynamics implies
that the biofilm detachment rate in a fluidizedbed reactor can be modeled with a stepwise function

$$b_{det} = \begin{cases} b_{col} f(C_{COD}), & \text{for } \varepsilon < \varepsilon_{c1}, \\ (b_{col} + b_{hydro}) f(C_{COD}), & \text{for } \varepsilon_{c1} \le \varepsilon < \varepsilon_{c2}, \\ (b_{hydro}) f(C_{COD}), & \text{for } \varepsilon \ge \varepsilon_{c2}, \end{cases}$$

where  $f(C_{COD})$  is a function that relates  $b_{det}$  and 356 chemical oxygen demand (COD) concentration 357 of wastewater,  $\varepsilon$  is the porosity,  $\varepsilon_{c1}$  and  $\varepsilon_{c2}$  are 358 critical porosities representing the boundaries of 359 the different regimes, and  $b_{col}$  and  $b_{hvdro}$  are 360 the detachment rates associated with collisional 361 and hydrodynamic effects. Biofilm detachment 362 363 is likely maximized to the coexistence of two different mechanisms at intermediate Reynolds 364 numbers. Although further experimental valida-365 tion is required, observing how particle dynam-366 ics change in fluidized bed simulations provides 367 insight into biofilm detachment and how detach-368 ment rates might best be modeled. Furthermore, 369 because  $\varepsilon_{c1}$  and  $\varepsilon_{c2}$  vary with particle properties 370 such as diameter and density, a universal scaling 371 law can be developed that confirms and general-372 izes this approach for different particle diameters 373 and densities. 374

### 375 3.1.2 Hypothetical impacts on mass transfer 376 and biological activities

377 Fluidized-bed reactors are known for their excellent mass transfer rate. When applied for 378 wastewater treatment, the AFBR can either be 379 mass transfer limited or reaction rate limited. 380 The latter usually occurs in shallow and fully-381 penetrated biofilms where substrates are me-382 tabolized at a much slower rate than diffusion 383 enables. Buffiere et al.<sup>35</sup> discovered that the 384 methanogenic step requires deep biofilms while 385 386 acidogenesis only requires shallow biofilms for treatment of high-strength wastewater. Conflict-387 ing results have been reported where increases in 388 flow rate can either increase<sup>45</sup> or decrease<sup>46</sup> the 389 mass transfer rate. Nicolella et al.<sup>47,48</sup> discov-390 ered that mass transfer of biofilm-covered parti-391 cles in airlift reactors is roughly 15% lower than 392 that of clean particles. 393

 $^{(5)}$  Due to the serial nature of process kinetics, 394 with mass transfer preceding biochemical kinet-395 ics, overall reactions can be mass transfer-limited 396 when the reaction step is fast or they can be 397 reaction-limited when the mass transfer step is 398 slow<sup>35</sup>. In AFBRs, the particle Reynolds num-399 ber based on superficial velocity leads to colli-400 sions and hydrodynamic effects that control mass 401 transfer. Higher flow rates reduce the thickness 402 of the diffusion layer thereby enhancing mass 403 transfer. Similarly, more frequent collisions dis-404 rupt the diffusion layer reducing its thickness 405 in fluidized-bed electrochemical cells<sup>49</sup>. The ef-406 fect of collisions alone can be accurately de-407 scribed by the collision pressure which is known 408 to have a maximum and zeros for both single-409 particle ( $\varepsilon \approx 1$ ) and close-packed reactors ( $\varepsilon \approx 410$  $(0.4)^{43,50}$ . After close examination of particle dy-411 namics in fluidized bed simulations, Yao et al.<sup>32</sup> 412 suggested that mass transfer is most likely maxi-413 mized within the intermediate porosity regime at 414 which point collisions and hydrodynamic factors 415 are equally important, leading to optimal biolog-416 ical performance. Although not yet experimen-417 tally validated, pilot- and lab-scale reactors oper-418 ated at this intermediate porosity (bed expansion 419 of 40% to 60%) have achieved optimal treatment 420 performance<sup>21,23,51</sup>. 421

# **3.1.3 Hypothetical impacts on membrane** 422 fouling control 423

The primary role of the P-MBR is to retain par-424 ticulate biodegradable organic matter in the re-425 actor because more time is required for hydroly-426 sis. The main challenge is to prevent membrane 427

biofouling, which can be accomplished by either 428 particle- or gas-sparging. Particle-sparged opera-429 tion enables low energy demand<sup>52</sup> but can lead 430 to severe membrane damage in the lower region 431 of the reactor<sup>53</sup>. Moreover, due to non-uniform 432 particle sizes, the fluidized bed in the P-MBR 433 forms segregated layers of particles with larger 434 particles (2-4 mm) located at the bottom of the 435 bed. Yao et al.<sup>32</sup> found that the maximum colli-436 sion frequency is attained at intermediate poros-437 ity for 2 mm particles. Low porosity is charac-438 terized by more frequent weak collisions while 439 high porosity is dominated by flow rather than 440 collisions. Comparing the collision frequency as a 441 function of porosity by Yao et al.<sup>32</sup> and the mem-442 brane integrity study by Shin et al.<sup>27</sup> with sim-443 ilar particle sizes, the porosity of the lower re-444 gion in the pilot-scale P-MBR corresponds to the 445 region of maximum effective collisions from the 446 simulations. This result implies that membrane 447 scouring efficiency can be controlled by varying 448 porosity, therefore the bed expansion. Maximum 449 membrane scouring is attained at the porosity 450 with maximum collisions. To avoid membrane 451 damage, varying the porosity to deviate from the 452 maxima say, by reducing or increasing it, is likely 453 to eliminate membrane damage. As discussed 454 below, instead of switching to alternative mem-455 brane fouling control methods, studying the ef-456 fects of the Archimedes number enables a retro-457 spective modification to both new and existing 458 reactors. 459

### 460 3.2 Particle properties and the Archimedes461 number

In addition to operating parameters such as upflow velocity and porosity, choosing optimal or
appropriate design parameters (i.e. Archimedes
number) is critical. As discussed in the previous section, particles with low Archimedes number are preferred for better mass transfer in the
AFBR. In reality, particles with the same prop-

erties are usually used for the P-MBR. Aslam et 469 al.<sup>26</sup> attempted to relate particle properties such 470 as materials, diameter and density to membrane 471 scouring efficiency and concluded that larger par-472 ticles are better at membrane fouling control. 473 Recently, Yao et al.<sup>33</sup> elucidated the role of the 474 Archimedes number on particle dynamics in a 475 fluidized bed. The Archimedes number com-476 bines different particle properties into a single 477 dimensionless number. Based on the simula-478 tions, the normalized particle velocity fluctuation 479 decreases as the Archimedes number increases, 480 indicating that the particles experience weaker 481 effects of wake interactions in which the par-482 ticle is weakly affected by neighbouring parti-483 cles. By using Voronoï tessellation, particle clus-484 tering is identified and the results suggest that 485 Archimedes number has a strong inverse rela-486 tionship on particle clustering lifespan such that 487 an increase in Archimedes number strongly de-488 creases the lifespan. Therefore, applications with 489 low Archimedes number are characterized by 490 long-lived clusters while applications with high 491 Archimedes number are characterized by short-492 The mechanism governing the 493 lived clusters. lifespan of particle clusters is the collision fre-494 quency. Increasing the Archimedes number in-495 creases the collision frequency, creating condi-496 tions more favorable for cluster breakup, leading 497 to short-lived clusters. 498

#### 3.2.1 Hypothetical impacts on flow short-499 circuting 500

A common practice in the operation of fluidized-501 bed reactors in wastewater treatment is to use 502 small particles that enhance both mass trans-503 fer and surface contact. In analogy to bound-504 ary layer thickness, the diffusion layer thickness 505 scales as 506

$$L \sim \sqrt{\frac{Dd_p}{\tilde{u}}},\tag{6}$$

527



Fig. 3 Hypothetical impacts of particle properties and Archimedes number on fluidized-bed reactor modeling, design and operation.

where D is the diffusion coefficient of the tar-507 geted compound and  $\tilde{u}$  is the fluid velocity over 508 the particle. From equation 6, L decreases as 509  $d_p$  decreases and  $\tilde{u}$  increases. Therefore, smaller 510 particles are less likely to be mass-transfer lim-511 ited due to the reduced diffusion layer thick-512 ness. In practice, particle size is chosen based on 513 the minimum particle size or Archimedes number 514 that can be easily retained in the system. How-515 ever, contrary to popular opinion, Yao et al.<sup>33</sup> 516 found that particles with Ar < 1000 tend to form 517 518 prolonged clusters while particles with  $Ar \ge 1000$ 519 are more likely to form short-lived clusters. Long-520 lived clusters tend to behave like a single large particle, resulting in more fluctuations in particle 521 dynamics. This result suggests that flow short-522 circuiting is more likely to occur when fluid flows 523 over a large particle cluster rather than each 524 individual particle, resulting in reduced surface 525 contact and exchange between biofilm and bulk 526

fluid.

# **3.2.2 Hypothetical impacts on scouring fre-** 528 **quency and membrane lifetime** 529

Hypothetical impacts

Besides porosity, particle properties controlling 530 the Archimedes number can affect membrane 531 scouring efficiency. As demonstrated in many 532 papers<sup>26,50,54</sup>, larger particles (high Archimedes 533 number) tend to result in more frequent impact 534 collisions that ultimately damage the membrane 535 over time while small particles (low Archimedes 536 number) do not induce effective collisions and 537 hence minimal membrane scouring<sup>33</sup>. Therefore, 538 choosing particles with appropriate Archimedes 539 number is critical. To ensure effective mem-540 brane biofouling control, particles with Ar > 1000541are likely to have effective collisions<sup>33</sup>. There-542 fore, the minimum Archimedes number for ef-543 fective membrane scouring is  $Ar \approx 1000$ . For 544 better scouring efficiency, particles with higher 545



Fig. 4 Power requirement as a function of porosity for (a)  $Ar = 2.3 \times 10^4$ , (b)  $Ar = 1.2 \times 10^5$  and (c)  $Ar = 2.3 \times 10^5$ .

546 Archimedes number will be more effective when547 there is a risk of membrane damage.

548 To alleviate membrane damage due to particle scouring, both the frequency and energy im-549 pacts of collisions must be reduced. This can be 550 551 achieved by operating the P-MBR at a porosity that favors weaker collisions. Collision frequency 552 and strength can both be reduced by changing 553 554 porosity (both by increasing and decreasing it). Because the expanded fluidized particles must be 555 able to access to the entire membrane module in 556 order to provide effective scouring, and because 557 the fluidized-bed height is predetermined, the to-558 tal mass of particles must change if the upflow ve-559 locity changes. The disadvantage of adding more 560 particles (reducing porosity) is that this leads to 561 higher headloss and increased pumping costs. To 562 increase porosity, a higher flow rate, hence a 563 higher power requirement, is essential. Since the 564 total headloss is proportional to both the hydro-565 static pressure loss and pipe friction loss, a more 566 detailed analysis of power requirements is re-567 quired to determine the optimal flow rate. As an 568 example, figure 4 shows the power requirement 569 as a function of porosity for different Archimedes 570 571 numbers Ar (model details can be found in the 572 Supplementary Information). As shown, when 573 the recirculation pipe diameter  $D_{pipe} > 0.2$  m,  $D_{pipe}$  is no longer an important parameter. For 574 low Ar, the power requirement is dominated by 575 the static head loss, and the wastewater must be 576 pumped from the bottom to the top of the reac- 577 tor, leading to a monotonically increasing func- 578 tion of Ar. For high Ar, the power needed to flu- 579 idize the particles exceeds static headloss, lead- 580 ing to a parabolic function of Ar. Therefore, to 581 reduce high energy collisions, the flow rate must 582 be reduced for small Ar and can be increased or 583 decreased for high Ar depending on the power 584 requirements. 585

#### 4 Conclusion and outlook 586

Simulations of particle dynamics in fluidized-bed 587 reactors using CFD suggest that the parameter 588 space for optimal bed expansion should decrease 589 from 10%-70% to 40%-60% because optimal 590 mass transfer is more likely to occur when both 591 collisional and hydrodynamic forces are compa-592 rably important. To design an efficient fluidized-593 bed reactor, particles with Ar > 1000 should be 594 chosen to avoid flow short-circuiting due to parti-595 cle clustering. Similarly, particles with Ar > 1000 596 or preferably Ar > 7000 are needed to induce 597 appreciable membrane scouring. The impact of 598 membrane scouring can be adjusted by varying 599 the porosity or flow rate.

Overall, high-fidelity CFD simulations enable a 601

650

602 close examination of fundamental hydrodynam-Although optimal de-603 ics within bioreactors. sign and operating conditions cannot be precisely 604 identified, the range of parameter space requir-605 ing experimental testing can be significantly re-606 duced, and the likelihood that optimal conditions 607 will be identified is greater. CFD simulations pro-608 vide an added tool for study of problems that 609 are difficult to investigate experimentally. Exper-610 iments can both validate and build upon CFD re-611 sults to optimize reactor performance. 612

613 Although CFD-accelerated strategies have tremendous potential for acceleration and opti-614 mization of wastewater treatment systems, more 615 work is clearly needed. More sophisticated com-616 617 putational methods are needed that incorporate 618 biological reactions. However, the main challenge in integrating biological reactions is the dif-619 ference in timescales. For biological reactions, 620 the timescales are typically much longer than the 621 time to reach hydrodynamic steady-state. As a re-622 sult, the total computational cost increases signif-623 icantly, and new methods are needed to address 624 this challenge. 625

#### 626 Funding Sources

This work was funded by the California Energy 627 Commission (CEC) under CEC project number 628 EPC-16-017, the U.S. NSF Engineering Center for 629 Reinventing of the Nation's Urban Water Infras-630 tructure (ReNUWIt) under Award No. 1028968, 631 and Office of Naval Research Grant N00014-16-632 633 1-2256. This paper was prepared as a result of work sponsored in part by the California Energy 634 Commission. It does not necessarily represent 635 636 the views of the Energy Commission, its employees, or the State of California. Neither the Com-637 mission, the State of California, nor the Commis-638 sion's employees, contractors, or subcontractors 639 makes any warranty, express or implied, or as-640 sumes any legal liability for the information in 641 642 this paper; nor does any party represent that the

use of this information will not infringe upon pri-643 vately owned rights. This paper has not been ap-644 proved or disapproved by the Commission, nor645 has the Commission passed upon the accuracy of 646 the information in this paper. 647

#### Conflicts of interest 648

There are no conflicts to declare. 649

- Notes and references
  - 1 T. Dooley, *Thirsting for a Future: Water and* 651 *Children in a Changing Climate*, United Na-652 tions Children's Fund, The (UNICEF), 2017. 653
  - 2 M. Smol, C. Adam and M. Preisner, J. Mater. 654 Cycles Waste Manage., 2020, **22**, 682–697. 655
  - 3 P. H. Nielsen, *Microb. Biotechnol.*, 2017, 10,656
     1102–1105.
     657
  - 4 E. Neczaj and A. Grosser, *Proc. AMIA Annu*. 658 *Fall Symp.*, 2018, **2**, 614. 659
  - 5 Y. D. Scherson, S.-G. Woo and C. S. Criddle, 660 Environ. Sci. Technol., 2014, **48**, 5612–5619. 661
  - 6 Y. Yao, Z. Wang and C. S. Criddle, *Environ*. 662 *Sci. Technol.*, 2021, **55**, 2016–2026. 663
  - 7 Z. Wang, S.-G. Woo, Y. Yao, H.-H. Cheng, Y.-J. 664
    Wu and C. S. Criddle, *Water Res.*, 2020, **173**, 665
    115575.
  - 8 Z. Wang, Y. Yao, N. Steiner, H.-H. Cheng, Y.-667
    J. Wu, S.-G. Woo and C. S. Criddle, *Envi*-668 *ron. Sci.: Water Res. Technol.*, 2020, 6, 3451–669
    3459.
- 9 B. H. Kim, I. S. Chang and G. M. Gadd, *Appl.* 671 *Microbiol. Biotechnol.*, 2007, 76, 485–494. 672
- 10 X. Xie, C. Criddle and Y. Cui, *Energy Environ*. 673 *Sci.*, 2015, **8**, 3418–3441. 674
- 11 X. Xie, M. Ye, C. Liu, P.-C. Hsu, C. S. Crid-675 dle and Y. Cui, *Energy Environ. Sci.*, 2015, 8,676 546–551.
- 12 D. Li and J. Ganczarczyk, Research Journal of 678 the Water Pollution Control Federation, 1991, 679
  63, 806–814.
- 13 B.-M. Wilén and P. Balmér, *Water Res.*, 1999, 681
  33, 391–400.

- 683 14 A. M. Karpinska and J. Bridgeman, *Water*684 *Res.*, 2016, **88**, 861–879.
- 685 15 M. W. D. Brannock, Y. Wang and G. Leslie, J.
  686 Memb. Sci., 2010, 350, 101–108.
- 687 16 Y. Le Moullec, C. Gentric, O. Potier and J. P.
  688 Leclerc, *Chem. Eng. Sci.*, 2010, 65, 343–350.
- 689 17 M. Gresch, M. Armbruster, D. Braun and
  690 W. Gujer, *Water Res.*, 2011, 45, 810–818.
- 691 18 J. Ducoste, K. Carlson and W. Bellamy,
  692 Journal of Water Supply: Research and
  693 Technology—AQUA, 2001, 50, 245–261.
- 694 19 D. J. Greene, B. Farouk and C. N. Haas, *J. Am.*695 *Water Works Assoc.*, 2004, **96**, 138–150.
- 696 20 J. M. Wilson and S. K. Venayagamoorthy, *Environ. Sci. Technol.*, 2010, 44, 9377–9382.
- 698 21 C. Shin, P. L. McCarty, J. Kim and J. Bae,
  699 *Bioresour. Technol.*, 2014, **159**, 95–103.
- Z J. Kim, K. Kim, H. Ye, E. Lee, C. Shin, P. L. McCarty and J. Bae, *Environ. Sci. Technol.*, 2011,
  45, 576–581.
- 703 23 C. Shin, S. H. Tilmans, F. Chen, P. L. Mc704 Carty and C. S. Criddle, *Water Res.*, 2021,
  705 204, 117598.
- 706 24 P. L. McCarty, J. Kim, C. Shin, P.-H. Lee and
  707 J. Bae, *Anaerobic Biotechnology*, IMPERIAL
  708 COLLEGE PRESS, 2015, pp. 211–242.
- 709 25 J. Bae, C. Shin, E. Lee, J. Kim and P. L. Mc710 Carty, *Bioresour. Technol.*, 2014, 165, 75–80.
- 711 26 M. Aslam, P. L. McCarty, J. Bae and J. Kim,
  712 Sep. Purif. Technol., 2014, 132, 10–15.
- 713 27 C. Shin, K. Kim, P. L. McCarty, J. Kim and
  714 J. Bae, Sep. Purif. Technol., 2016, 162, 101–
  715 105.
- 716 28 R. de Felice, *Chem. Eng. Sci.*, 1993, 48, 881–
  717 888.
- 718 29 C. Nicolella, S. Chiarle, R. Di Felice and
  719 M. Rovatti, *Water Sci. Technol.*, 1997, 36,
  720 229–235.
- 721 30 A. Esteghamatian, A. Hammouti, M. Lance
  722 and A. Wachs, *Phys. Fluids*, 2017, 29,
  723 033302–1–033302–14.

- 31 D. P. Willen and A. Prosperetti, *Phys. Rev. Flu-* 724 *ids*, 2019, 4, 014304.
  725
- 32 Y. Yao, C. S. Criddle and O. B. Fringer, J. Fluid 726 Mech., 2021, 927, A28.
   727
- 33 Y. Yao, C. S. Criddle and O. B. Fringer, J. Fluid 728
   Mech., 2021, 920, A40. 729
- 34 Y. Yao, C. S. Criddle and O. B. Fringer, *Phys.* 730
   *Rev. Fluids*, 2021, 6, 084306.
   731
- 35 P. Buffière, J. P. Steyer, C. Fonade and R. Mo-732 letta, *Biotechnol. Bioeng.*, 1995, **48**, 725–736.733
- 36 R. Lee, P. L. McCarty, J. Bae and J. Kim, J. 734
   *Chem. Technol. Biotechnol.*, 2015, 90, 391–735
   397. 736
- 37 S. Sundaresan, Annu. Rev. Fluid Mech., 2003, 737
   35, 63–88.
   738
- 38 J. F. Richardson and M. A. da S. Jerónimo, 739
   *Chem. Eng. Sci.*, 1979, 34, 1419–1422.
   740
- 39 B. E. Rittmann and P. L. McCarty, Environ-741 mental Biotechnology: Principles and Appli-742 cations, McGraw-Hill Education, Columbus,743 OH, 2018. 744
- 40 A. Gjaltema, L. Tijhuis, M. C. van Loosdrecht 745 and J. J. Heijnen, *Biotechnol. Bioeng.*, 1995, 746
  46, 258–269. 747
- 41 H. T. Chang, B. E. Rittmann, D. Amar, 748
  R. Heim, O. Ehlinger and Y. Lesty, *Biotechnol*. 749 *Bioeng.*, 1991, 38, 499–506. 750
- 42 C. Nicolella, R. Di Felice and M. Rovatti, 751 *Biotechnol. Bioeng.*, 1996, **51**, 713–719. 752
- 43 A. Gjaltema, J. L. Vinke, M. C. van Loosdrecht 753 and J. J. Heijnen, *Biotechnol. Bioeng.*, 1997, 754
  53, 88–99. 755
- 44 C. Shin, S. H. Tilmans, F. Chen and C. S. Crid-756
   dle, *Chem. Eng. J.*, 2021, 426, 131912.
- 45 A. Venu Vinod and G. Venkat Reddy, J. Haz-758 ard. Mater., 2006, **136**, 727–734. 759
- 46 L. P. Lakshmi and Y. P. Setty, *Chem. Eng. J.*, 760 2008, **135**, 135–140.
  761
- 47 C. Nicolella, M. C. van Loosdrecht, R. G. 762 van der Lans and J. J. Heijnen, *Biotechnol*. 763 *Bioeng.*, 1998, **60**, 627–635.

- 765 48 C. Nicolella, M. C. M. van Loosdrecht and J. J.
  766 Heijnen, *Chem. Eng. Sci.*, 1998, **53**, 2743–
- 767 2753.
- 768 49 N. A. Shvab, J. Appl. Electrochem., 2000, 30,
  769 1285–1292.
- 770 50 R. Zenit, M. L. Hunt and C. E. Brennen, J.
  771 *Fluid Mech.*, 1997, **353**, 261–283.
- 51 J. Jaafari, A. Mesdaghinia, R. Nabizadeh,
  M. Hoseini, H. Kamani and A. H. Mahvi, J *Environ Health Sci Eng*, 2014, 12, 139.
- 775 52 C. Shin, E. Lee, P. L. McCarty and J. Bae, *Bioresour. Technol.*, 2011, **102**, 9860–9865.
- 777 53 C. Shin and J. Bae, *Bioresour. Technol.*, 2018,
  778 247, 1038–1046.
- 779 54 A. Gjaltema, M. C. van Loosdrecht and J. J.
- 780 Heijnen, Biotechnol. Bioeng., 1997, **55**, 206–
- 781 215.