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# Antibacterial activity of the micro and nanostructures of the optical material tris(8-hydroxyquinoline)aluminum and its application as an antimicrobial coating†

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Although tris(8-hydroxyquinoline)aluminum (Alq<sub>3</sub>), a fluorescent optical organometallic material, is known for its applications in optoelectronics, it has only few and limited applications in the biological field. In this study, the antibacterial activity of Alq<sub>3</sub> micro and nanostructures was investigated. We prepared Alq<sub>3</sub> nanostructures. We prepared nanosized Alq<sub>3</sub> as rice-like structures that assembled into flower shapes with an  $\alpha$ -crystal phase. Then, Alq<sub>3</sub> micro and nanostructure antibacterial activities were estimated against seven human pathogenic bacterial strains. Besides, we compared their antibacterial activities with those of standard antibiotics. The minimal inhibitory concentration (MIC), minimum bactericidal concentration (MBC), and IC50 were evaluated. Alq<sub>3</sub> micro and nanostructure antibacterial activity showed considerable values compared to standard antibiotics. Besides, the obtained data revealed that the antibacterial activity of Alq<sub>3</sub> in nanostructures with a new morphology is more than that in microstructures. The antibacterial activity of Alq<sub>3</sub> nanostructures was attributed to their more surface interactions with the bacterial cell wall. The molecules of 8-hydroxyquinoline in the Alq<sub>3</sub> structure could play crucial roles in its antibacterial activity. To apply the achieved results, Alq<sub>3</sub> was incorporated with polystyrene (PS) in a ratio of 2% to fabricate a PS/Alq<sub>3</sub> composite and used to coat glass beakers, which showed inhibition in the bacterial growth reduced to 65% compared with non-coated beakers. The finding of this study showed that Alq<sub>3</sub> could be used as a promising antimicrobial coating.

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## 1. Introduction

Tris(8-hydroxyquinoline)aluminum (Alq<sub>3</sub>) is a well-known optical material that has been used in the manufacture and design of optoelectronic devices, especially in organic light-emitting diode (OLED) designs.<sup>1,2</sup> It is an organometallic semiconductor, which is also considered one of the most widely used small molecules. It is a chelate complex with polymorph

crystal structures.<sup>3–5</sup> Where its crystal could be a meridional (*mer*) or facial (*fac*) isomer, it was proved that Alq<sub>3</sub> could exist in five different crystal phases, namely,  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\epsilon$ , and  $\delta$ .<sup>3–5</sup> It has an efficient luminescence property with thermal stability at  $\geq 350$  °C. Although Alq<sub>3</sub> has been utilized mainly in optoelectronic manufacturing, new applications have been performed; recently, it has been used in the direction of radiation detection as an X-ray<sup>6</sup> and ultraviolet (UV) dosimeter.<sup>7</sup> In the field of medical and bio-applications, it was used to recognize DNA.<sup>8</sup> Besides, in a previous study, we have found that Alq<sub>3</sub> can kill human colon cancer cell lines (HCT116); its fluorescence was also utilized to follow gelatin degradation.<sup>9</sup> Recently, in more detail, Alq<sub>3</sub> toxicity was investigated<sup>10</sup> with HCT116, human cell line breast cancer (MCF7), and human foreskin fibroblasts cell line (HFF-1); besides, the Alq<sub>3</sub> fluorescence was investigated to be used as a dye for *in vitro* bioimaging for the first time.<sup>10</sup>

The presence of microorganisms that cause harms to human health is a great concern of societies, peoples, countries, and humanity in general. The diversity of infections and the emergence of new diseases further complicate the matter and exacerbates the issue. What is more, despite the existence and innovation of many antibiotics, these microorganisms are

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rapidly developing their resistance to antibiotics. Accordingly, bacterial contamination has become one of the serious threats to human health. It is there in all human activities and daily life; for instance, it can occur in food.<sup>11</sup> Moreover, it occurs on surfaces in hospitals,<sup>12</sup> where it may spread to the hands of caregivers as a result of direct contact with patients and people with multi-infection.<sup>13</sup> Although many materials have been synthesized for antibacterial applications,<sup>14–25</sup> finding new antimicrobial materials is essential in the light of emerging antibiotic resistance.

In our previous studies,<sup>9,10</sup> we have found the effect of Alq<sub>3</sub>, especially its nanoparticles (NPs),<sup>9</sup> in killing cancer cells, which could indicate the possibility of using it as an antibacterial material. Therefore, we conducted this study to investigate the antibacterial activity of Alq<sub>3</sub> optical materials. To the best of our knowledge, despite many studies devoted to studying Alq<sub>3</sub> and its properties and uses, no investigation has been dedicated to studying its antibacterial activity. In this study, we investigated the antibacterial activity of its micro and nanostructures. The antibacterial activity of Alq<sub>3</sub> was studied on seven different types of Gram-positive and gram-negative bacteria to obtain a comprehensive and detailed study. We evaluated the antibacterial activity by an agar well diffusion (AWD) method and a resazurin assay. Furthermore, we added Alq<sub>3</sub> to polystyrene (PS) to form a PS/Alq<sub>3</sub> composite; then, we studied the bacterial growth in a glassware coated with a PS/Alq<sub>3</sub> composite. Many details of the study and its results are included in the following sections.

## 2. Methodology

### 2.1. Materials

Alq<sub>3</sub> was obtained from (Tokyo Chemical Industry Company Limited, Japan). Resazurin, phosphate-buffered saline (PBS), and gentamicin (GM) solution (50 mg ml<sup>-1</sup>) were purchased from (Sigma-Aldrich, Germany). Mueller-Hinton agar (MHA) (Condalab, Spain) and Mueller-Hinton broth (MHB) (Scharlau, Spain) were used as the bacteriological media. Commercially standard antibiotics (St. antibiotics) discs (Mast Group Ltd, UK) were used as control antibiotics. Petri dishes, glassware, and sterilized swap cotton were obtained from the local market (Jeddah, Saudi Arabia). Microtiter plates (96-Well with lid) were obtained from SPL Life Sciences Co. (SPL, South Korea). PS polymer was obtained from SABIC (SABIC Co., Saudi Arabia). The materials were used directly without further purification.

### 2.2. Sampling and Alq<sub>3</sub> nanoparticle (NP) preparation

Alq<sub>3</sub> original powders were named (Alq<sub>3</sub> as rec) sample. We used the original powders of Alq<sub>3</sub> to prepare Alq<sub>3</sub> NPs. We prepared the Alq<sub>3</sub> NPs based on the published work with modifications if necessary. Briefly, we dissolved the original powders of Alq<sub>3</sub> in chloroform (15 mM); chloroform is a preferred solvent over other solvents due to its excellent properties as a solvent for Alq<sub>3</sub> with a reasonable boiling temperature (61.15 °C). Then, the Alq<sub>3</sub> chloroform solution was placed in an ultrasonic bath for completely dissolving Alq<sub>3</sub>. After that, the solution was added

dropwise into deionized water at a temperature of 60 °C under stirring. For chloroform to get volatilized, the stirring was continued at 60 °C for 15 minutes after the dropwise addition. Then, the Alq<sub>3</sub>-water solution was centrifuged, and the precipitate was dried at 70 °C. The obtained powders were named Alq<sub>3</sub> NP sample. Finally, Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples were used to perform characterizations and antibacterial activity experiments.

### 2.3. Characterization techniques

We investigated the morphology and particle size of both Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples using a field-emission scanning electron microscope (FESEM), model JSM-7600F (JEOL, Japan). We recorded their X-ray powder diffraction (XRPD) patterns to study their crystallinity and crystal phases using an X-ray diffractometer, model Ultima IV (Rigaku, Japan). The XRPD patterns of the Alq<sub>3</sub> samples were scanned at room temperature and an angular range ( $2\theta$ ) of 5–28° in the step of 0.05° and a continuous mode ( $\lambda = 1.54 \text{ \AA}$ ,  $I = 40 \text{ mA}$ , and  $V = 40 \text{ kV}$ ). The chemical functional groups of Alq<sub>3</sub> were verified and characterized using a Fourier transform infrared (FTIR) spectrometer, model Nicolet iS10 FTIR (Thermo Scientific, USA). We recorded their FTIR spectra using the attenuated total reflection (ATR) sampling technique in which the ATR occurred in a germanium crystal. We recorded both photoluminescence (PL) excitation and emission spectra of the Alq<sub>3</sub> samples using a fluorescence spectrofluorophotometer, model RF-5301 PC (Shimadzu, Japan).

### 2.4. Antibacterial activity tests

**2.4.1. Bacterial strains and growth.** We tested the antibacterial activity of the Alq<sub>3</sub> samples on seven different human pathogenic bacterial strains representing gram-positive and gram-negative bacteria. These seven bacteria are *Escherichia coli* ATCC 11775 (EC), *Enterococcus faecalis* ATCC 29212 (EF), *Klebsiella pneumoniae* ATCC 13883 (KP), *methicillin-resistant Staphylococcus aureus* ATCC 33591 (MRSA), *Pseudomonas aeruginosa* ATCC 9027 (PA), *Staphylococcus aureus* ATCC 12600 (SA), and *Salmonella typhimurium* ATCC 14028 (ST). We obtained them from Microbiology Unit at King Fahd Medical Research Center (KFMRC) (Jeddah, Saudi Arabia). We grew the human pathogenic bacterial strains on blood agar and preserved them on MHA plates in a refrigerator at 4 °C until use.<sup>26</sup>

**2.4.2. AWD assay.** We tested the antibacterial activity of both Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples against EC, EF, KP, MRSA, PA, SA, and ST bacteria by AWD methods according to the published work<sup>27</sup> with needed modifications. The samples were suspended in PBS at four concentrations, namely, 120, 240, 360, and 480  $\mu\text{g ml}^{-1}$ . The MHA solution was prepared according to the manufacturer's instructions, sterilized and then poured into Petri dishes to obtain gel MHA plates. We measured the absorbance at 600 nm using a UV-visible spectrophotometer, model Genesys 10s UV-vis (Thermo Scientific, USA), to determine the bacterial turbidity that was adjusted to  $0.1 \pm 0.02$ . Then, 50  $\mu\text{l}$  of bacteria was aseptically introduced and spread using sterilized cotton swabs on the surface of gel MHA plates.



After that, wells were punched on the gel MHA plates (four wells/plate) using a sterile cork borer of 6.0 mm diameter. Then, 170  $\mu\text{l}$  of  $\text{Alq}_3$  sample suspension was added into the wells (one concentration/well – four concentrations/plate). The gel MHA plates were incubated in an incubator with an air atmosphere at 37  $^\circ\text{C}$ . After 24 h, the plates were checked, where the inhibition zones' diameter, resulting from the tested  $\text{Alq}_3$  concentration, was measured using a ruler. These procedures were performed similarly with all seven human pathogenic bacterial strains.

**2.4.3. St. antibiotics.** We utilized six commercially purchased St. antibiotics discs, namely, *ampicillin* 10  $\mu\text{g}$  (AP), *augmentin* 30  $\mu\text{g}$  (AU), *gentamicin* 10  $\mu\text{g}$  (GM), *cefoxitin* 30  $\mu\text{g}$  (FOX), *cephalothin* 30  $\mu\text{g}$  (KF), and *cotrimoxazole* 25  $\mu\text{g}$  (TS). We placed them on the inoculated MHA plates (one plate/bacterium strain). The plates were incubated at 37  $^\circ\text{C}$  for 24 h. Then, we measured the inhibition zones that rounded each antibiotic disc using a ruler. We used these St. antibiotics as the control antibiotics to compare their results with those obtained from  $\text{Alq}_3$  samples.

**2.4.4. Resazurin assay.** We used the resazurin assay to estimate the minimal inhibitory concentration (MIC) and minimum bactericidal concentration (MBC); the resazurin assay was performed according to the published work<sup>28</sup> with necessary modification (Fig. 1 shows the illustration of the resazurin assay). Briefly, under aseptic conditions, sterile 96-well microtiter plates were used. Both  $\text{Alq}_3$  as rec and  $\text{Alq}_3$  NP samples were prepared in MHB media with a concentration of

500  $\mu\text{M}$ ; then, 100  $\mu\text{l}$  of this concentration was added to column 1, and at the same time, 50  $\mu\text{l}$  of MHB media (media only) was added using a multichannel pipette to columns 2–10, the same volume being added to column 12 as the positive control (PC). In contrast, we added 100  $\mu\text{l}$  MHB only for column 11 as a negative control (NC). Then, serial dilutions were performed in columns 1–10 to obtain concentrations of 120, 60, 30, 15, 7.5, 3.75, 1.88, 0.94, 0.47, and 0.23  $\mu\text{g ml}^{-1}$ . The highest concentration of 120  $\mu\text{g ml}^{-1}$  was in column 1 and the lowest concentration of 0.23  $\mu\text{g ml}^{-1}$  was in column 10. After that, 50  $\mu\text{l}$  of the bacterial suspension was added to all the wells except those in column 11; simultaneously, the turbidity was adjusted to be around  $5 \times 10^5$  colony-forming units (CFU) per milliliter in columns 1–10 and 12. Then, the 96-well microtiter plates were incubated in an air atmosphere at 37  $^\circ\text{C}$  for 24 h. After 24 h incubation, for the estimation of the half-maximal inhibitory concentration (IC<sub>50</sub>), the absorbance values of 96-well microtiter plates were read at 620 nm<sup>29</sup> using a plate reader spectrometer, model SYNRGY2 reader, operated using the Gen5 Software (BioTek, USA). The recorded absorbance values for the 96-well microtiter plates were used to estimate the growth inhibition of bacteria ratio based on the following equation:<sup>29</sup>

$$\text{Inhibition \%} = \left(1 - \frac{A_x - A_{\text{NC}}}{A_{\text{PC}} - A_{\text{NC}}}\right) \times 100\% \quad (1)$$

where  $A_x$  is the average absorbance of the wells at a concentration  $X$ ,  $A_{\text{NC}}$  is the average absorbance of the NC wells, and  $A_{\text{PC}}$  is

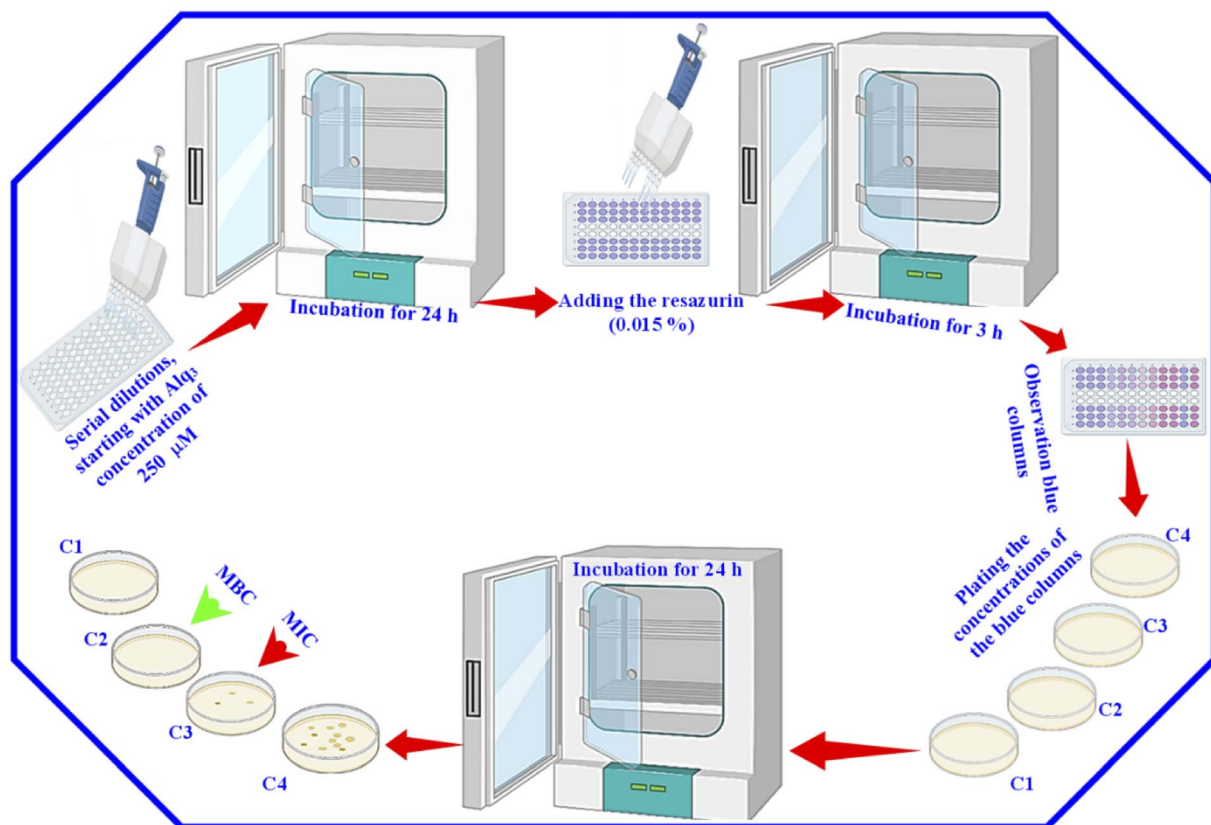


Fig. 1 Illustration of the resazurin assay using 96-well microtiter plates.



the average absorbance of the PC wells (see 2.4.4 subsection). The inhibition percentages were plotted *versus* common logarithms of the concentration; then, a nonlinear curve fit was carried out for every sample and bacterial strain based on a sigmoidal curve (dose–response curve) with the Levenberg–Marquardt as the iteration algorithm; this procedure was performed using OriginPro 2019b version 9.65. Then, the IC50 was extracted from the fitted curve.

After that, for the estimation of MIC and MBC, 30  $\mu\text{l}$  of resazurin (0.015%) was added to all wells, and the 96-well microtiter plates were again incubated for 3 h. Then, we observed the color change. We considered the columns with blue color (unchanged color) as above the MIC value; we plated the content of these columns to determine the MIC and MBC. Besides, we used GM as the *St.* antibiotic; the serial dilutions were performed with the GM to obtain the same concentrations of the  $\text{Alq}_3$  samples, and the same procedures were applied.

## 2.5. PS/ $\text{Alq}_3$ composite preparation

After we studied the effectiveness of  $\text{Alq}_3$  as an antimicrobial material, we incorporated it into a PS polymer to form an antibacterial composite; Fig. 2 shows a scheme that summarizes the fabrication of PS/ $\text{Alq}_3$  composites. The PS granules and the  $\text{Alq}_3$  powder were dissolved in chloroform separately under stirring at

40  $^\circ\text{C}$  to get solutions A and B; when they got completely dissolved in chloroform, we added the  $\text{Alq}_3$  chloroform solution (A) to the PS chloroform solution (B) to form solution (C). The stirring for solution C continued for 1 h till it became homogenous. Then, 10 ml glass beakers were coated with a PS polymer and a PS/ $\text{Alq}_3$  composite by immersing them in solutions B and C. Then, they were pulled out slowly from the solutions; they were dried in an air atmosphere at room temperature. The thickness of the coated layer was investigated by SEM.

After that, we grew bacterial suspensions in coated beakers (3 ml/beaker), and also in the non-coated beakers as NC; non-coated beakers that contained 5  $\mu\text{g ml}^{-1}$  of *St.* antibiotic GM served as PC group. The turbidity was adjusted to be the same for all beakers. Then, the beakers were covered with parafilm; they were incubated in an air atmosphere at 37  $^\circ\text{C}$  inside the incubator. The turbidity was recorded by measuring the absorbance at 600 nm after incubation. The bacterial growth was calculated from the absorbance as follows:

$$\text{Bacteria growth \%} = \frac{A_s - A_b}{A_c - A_b} \times 100\% \quad (2)$$

where  $A_s$  and  $A_c$  are respectively the absorbance of the sample and negative control beakers after incubation;  $A_b$  is the absorbance before incubation. The absorbance before incubation for

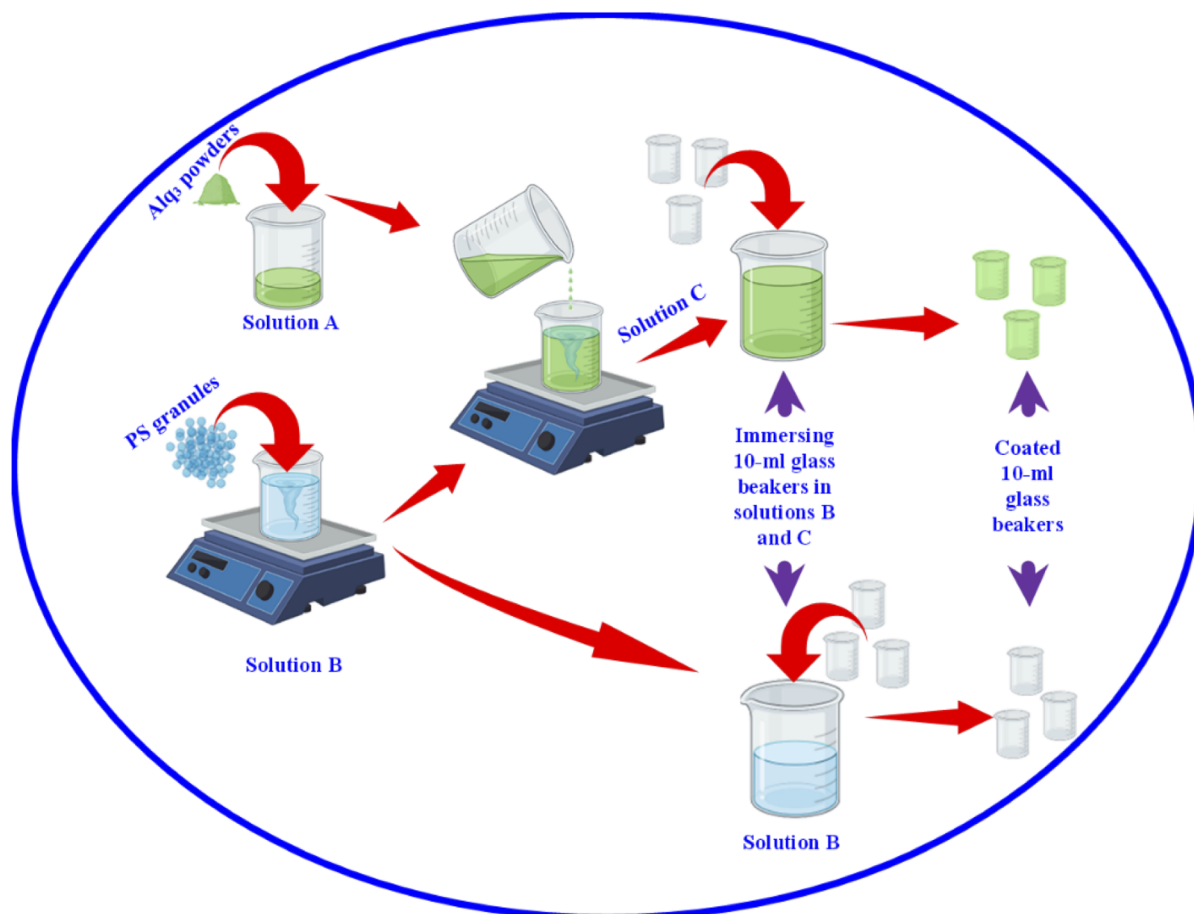


Fig. 2 Schematic diagram showing the fabrication of PS/ $\text{Alq}_3$  as an antibacterial composite.



all samples and control was adjusted to be the same value for all tested samples.

## 2.6. Statistical and data analysis

The experiments were repeated five times; then, the data were expressed as average values, standard deviation (SD). IC50s were extracted from dose–response curves by fitting the data measured during antibacterial activity tests. In this work, all statistical analyses and extraction of data were performed using OriginPro 2019b version 9.65.

# 3. Results and discussion

## 3.1. Alq<sub>3</sub> sample characterizations

We prepared the Alq<sub>3</sub> nanostructures and investigated their morphology and sizes by SEM. Fig. 3a and 3b display the captured SEM images of both Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples, respectively. Besides, we photographed the samples at a higher magnification, shown in Fig. 3c and d, to get a clear view of the particle size of the Alq<sub>3</sub> samples. As we can observe in Fig. 3a and c, the Alq<sub>3</sub> as rec sample appear to have rod shapes that are aggregative in bundles; the length of the rods is in the range between ~500 nm and ~2 μm with a diameter of ~200 nm, whereas the prepared Alq<sub>3</sub> NPs show small particles with a length up to ~130 nm and a diameter less than ~70 nm, which are nearly rice-like shapes, that are aggregative with flower-like shapes (Fig. 3b and d); hence, it can be said that the prepared Alq<sub>3</sub> NPs are rice-like nanostructures aggregated into flower shapes. This result is in agreement with the published works<sup>9,30</sup> reporting the preparation of Alq<sub>3</sub> NPs. The change in

morphology between the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples can be understood as the ratio of solvent (chloroform) to water playing a crucial role in controlling the particle growth, shape, and size, where the morphology transformation process is different because of the various chemical and physical properties, such as surface tension, viscosity, and density between the solvent and the water during Alq<sub>3</sub> re-precipitation.<sup>9,30</sup>

The XRPD patterns of the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples' powders are shown in Fig. 4a, which displays different diffraction peaks for both samples, indicating the samples' crystallization. The diffracted peaks (Miller indices) were indexed based on ICDD PDF card no. 00-026-1550 and published works.<sup>30–32</sup> All diffracted peaks in the XRPD patterns of both Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples' powders matched with the α-Alq<sub>3</sub> crystal phase. The difference that can be observed between the two samples is that the intensities of the diffracted peaks of the Alq<sub>3</sub> as rec sample's powder are higher than those of the diffracted peaks of the Alq<sub>3</sub> NP powder sample. This observation points out the differences in the crystallite size; it could also indicate that the crystallinity degree of the Alq<sub>3</sub> as rec sample is greater than that of the Alq<sub>3</sub> NP sample. This result was observed in a published study,<sup>32</sup> in which the Alq<sub>3</sub> NPs prepared without adding surfactants showed a lower degree of crystallization. During the preparation of Alq<sub>3</sub> nanoparticles, we avoided adding any other material as a surfactant to prevent the influence on the results of the antibacterial activity experiments.

The infrared spectra of both Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples' powders were recorded; Fig. 4b displays the FTIR absorbance spectra recorded in the spectral range between 1650 and 600 cm<sup>-1</sup>. Both samples have the same spectral features,

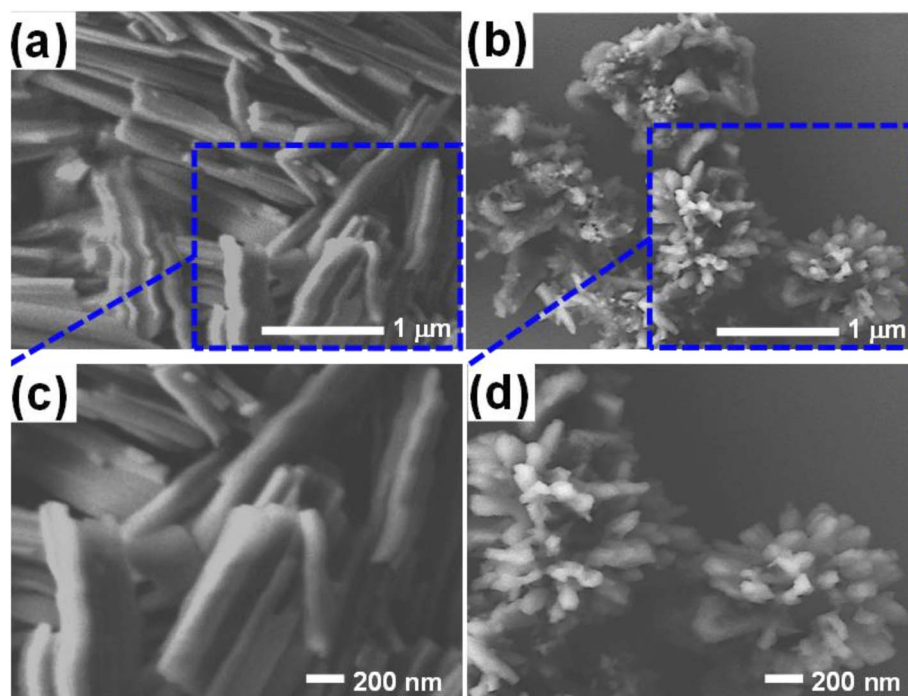


Fig. 3 SEM images of the Alq<sub>3</sub> samples: (a) and (b) images of Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples, respectively; (c) and (d) their images at a higher magnification.



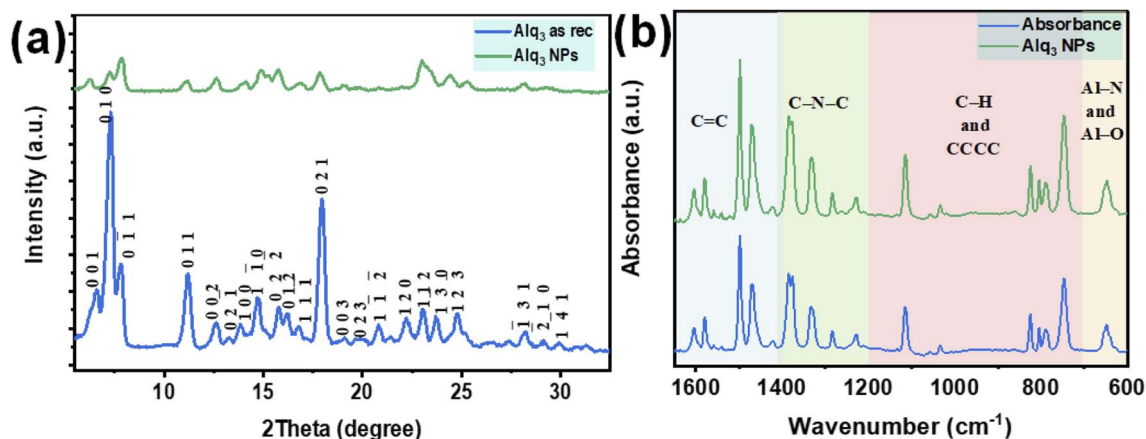


Fig. 4 (a) XRPD patterns and (b) FTIR spectra of the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples' powders.

indicating that the chemical bonds did not change during the preparation of the nanostructures. In general, and based on the published works,<sup>9,33,34</sup> the FTIR spectral peaks can be divided into four spectral regions resulting from the vibration of chemical bonds between the atoms that make up the Alq<sub>3</sub> molecule. The peaks in the spectral range starting from 1650 to 1400 cm<sup>-1</sup> could be ascribed to the stretching vibration of the aromatic C=C bonds, while the peaks in the spectral range of 1400–1200 cm<sup>-1</sup> were assigned to the vibration of the aromatic amine resonances C–N–C bonds. The peaks between 1200 and 700 cm<sup>-1</sup> were attributed to the in-plane bending/out-of-plane wagging vibrations of the aromatic C–H bonds, besides the CCCC torsional motions of the quinoline skeleton. The FTIR absorbance peaks in the spectral range 700–600 cm<sup>-1</sup> belonged to the stretching vibration modes of the metal–ligand bonds (Al–N and Al–O).

Alq<sub>3</sub> is an optical material well known for its excellent electroluminescence and fluorescence; therefore, we recorded the PL of the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples. The PL spectra of both samples are shown in Fig. 5; the PL excitation spectra recorded in the spectral range of 250–470 nm ( $\lambda_{em} = 500$  nm) show two peaks: one peak at 380, which is in the UV region, and the other at 419 nm, which is in the visible region (Fig. 5a). These two PL

peaks result from the ligands' electronic  $\pi$ – $\pi^*$  transitions.<sup>35,36</sup> The PL emission spectra of the samples, in the spectral range of 400–750 nm ( $\lambda_{ex} = 380$  nm), are shown in Fig. 5b; the spectra show a broadband peak centered at 495 nm. This peak results from the electronic  $\pi$ – $\pi^*$  transitions in the quinolinolate ligands,<sup>35,36</sup> in which the electrons predominantly at the phenoxide ring transit to the pyridyl ring.<sup>37</sup> The Alq<sub>3</sub> NP sample shows a significantly lower intensity than that of the Alq<sub>3</sub> as rec sample. This observation could be due to the higher crystallinity<sup>32</sup> in the microrods of the Alq<sub>3</sub> as rec (see XRPD results), where the PL originates from Alq<sub>3</sub> molecules' effective  $\pi$ – $\pi$  interactions, which increase with the increase in crystallinity. These results agree with those reported in the published work.<sup>32</sup>

### 3.2. Alq<sub>3</sub> antibacterial activity

We prepared Alq<sub>3</sub> colloids at concentrations of 120, 240, 360, and 480  $\mu\text{g ml}^{-1}$ . Fig. S1† shows these concentrations for the Alq<sub>3</sub> NP sample, where an optical image in visible light (Fig. S1a†) and a fluorescence image under UV irradiation at an excitation wavelength of 385 nm (Fig. S1b†). The fluorescence image shows the expected observation, where the fluorescence

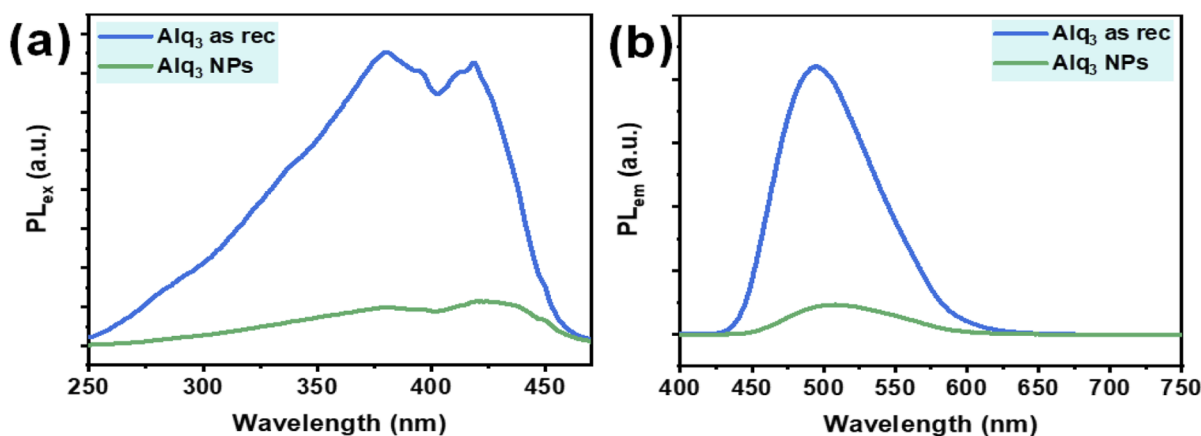


Fig. 5 PL spectra of the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples: (a) excitation spectra ( $\lambda_{em} = 500$  nm) and (b) emission spectra ( $\lambda_{ex} = 380$  nm).



shining increased with the concentration of Alq<sub>3</sub> NPs as the fluorescence material.

We have studied these concentrations on seven different types of human pathogenic bacterial strains (HPBS), gram-positive and gram-negative, and compared them with six types of St. antibiotics. Fig. S2† shows the antibacterial activity performed by the AWD against the bacteria of EC, EF, KP, MRSA, PA, SA, and ST, besides the St. antibiotics. Fig. S2† includes seven rows representing the seven types of HPBS. The first column represents the Alq<sub>3</sub> as rec sample antibacterial activity, the second column represents the Alq<sub>3</sub> NP sample antibacterial activity, and the third column represents the St. antibiotic antibacterial activity. The bacterial growth inhibition zones for the Alq<sub>3</sub> NP sample (Fig. S2,† second column) are larger than those for the Alq<sub>3</sub> as rec sample (Fig. S2,† first column). Interestingly, the bacterial growth inhibition zones for the Alq<sub>3</sub> NP sample at a concentration of 480 μg ml<sup>-1</sup> (Fig. S2,† second column) are larger than those for the St. antibiotics (Fig. S2,† third column) in KP, MRSA, and SA.

To quantitatively evaluate the results of the AWD method, we measured the diameter of the bacterial growth inhibition zones for each concentration of the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples against each bacterial strain as well as the St. antibiotics. Table 1 summarizes the inhibition zone diameters for Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples at 120, 240, 360, and 480 μg ml<sup>-1</sup>, besides the St. antibiotics AP 10 μg per disc, AU 30 μg per disc, GM 10 μg per disc, FOX 30 μg per disc, KF 30 μg per disc, and TS 25 μg per disc against the EC, EF, KP, MRSA, PA, SA, and ST. It can be seen that the Alq<sub>3</sub> NP sample showed better bacterial activity than the Alq<sub>3</sub> as rec sample against most of the seven bacterial strains. Besides, the sample Alq<sub>3</sub> NPs showed bacterial activity similar to St. antibiotics. Moreover, they showed even better than the activity of the St. antibiotics tested against bacteria, especially at 360 and 480 μg ml<sup>-1</sup>. For instance, the Alq<sub>3</sub> NPs antibacterial activity at 480 μg ml<sup>-1</sup> was the best (inhibition zone diameter ~27 mm) against KP, followed by GM 10 μg per disc, TS 25 μg per disc (inhibition zone diameter ~24 mm), and then the Alq<sub>3</sub> NPs at a concentration of 360 μg ml<sup>-1</sup> (inhibition zone diameter ~23 mm). The Alq<sub>3</sub> NP sample at 240, 360, and 480 μg ml<sup>-1</sup> concentrations recorded the best antibacterial activity against the bacterial strain MRSA (inhibition zone diameter ~22, 27, and 31 mm, respectively) compared with the Alq<sub>3</sub> as rec sample and all the St. antibiotics. Against the bacterial strain PA, although the six St. antibiotics showed the best antibacterial activity, the Alq<sub>3</sub> NP sample showed good antibacterial activity (inhibition zone diameter ~18 to 30 mm). The recorded antibacterial activity of the Alq<sub>3</sub> NP sample at a concentration of 480 μg ml<sup>-1</sup> against the SA and ST (inhibition zone diameter > 27 mm) was better than those recorded with the six St. antibiotics (inhibition zone diameter ~25 mm). Generally, the antibacterial activity of the Alq<sub>3</sub> NP sample showed better results than those of the Alq<sub>3</sub> as rec sample; besides, its antibacterial activity was excellent compared with those of the St. antibiotics. A histogram in Fig. S3† represents the data listed in Table 1.

We found that the best antibacterial activity recorded *via* the AWD method is for the Alq<sub>3</sub> NP sample, which showed larger inhibition zones than those of the Alq<sub>3</sub> as rec sample. The AWD

**Table 1** Growth inhibition zones (mm) against the bacterial strains EC, EF, KP, MRSA, PA, SA, and ST, which were incubated for 24 h; the growth inhibition zones were recorded by the AWD assay for both the Alq<sub>3</sub> as rec and the Alq<sub>3</sub> NP samples (170 μl per well) at concentrations of 120, 240, 360, and 480 μg ml<sup>-1</sup>, besides the St. antibiotics AP 10 μg per disc, AU 30 μg per disc, GM 10 μg per disc, FOX 30 μg per disc, KF 30 μg per disc, and TS 25 μg per disc

Tested bacteria	Alq <sub>3</sub>												St. antibiotics														
	Alq <sub>3</sub> as rec			Alq <sub>3</sub> NPs						AP			AU			GM			FOX			KF			TS		
	120 μg ml <sup>-1</sup>	240 μg ml <sup>-1</sup>	480 μg ml <sup>-1</sup>	120 μg ml <sup>-1</sup>	240 μg ml <sup>-1</sup>	360 μg ml <sup>-1</sup>	480 μg ml <sup>-1</sup>	120 μg ml <sup>-1</sup>	240 μg ml <sup>-1</sup>	360 μg ml <sup>-1</sup>	480 μg ml <sup>-1</sup>	10 μg per disc	30 μg per disc	10 μg per disc	30 μg per disc	10 μg per disc	30 μg per disc	30 μg per disc	30 μg per disc	25 μg per disc	30 μg per disc	30 μg per disc	25 μg per disc	30 μg per disc	25 μg per disc		
EC	0	0	0	0	11 ± 3.1	15 ± 1.9	22 ± 1.5	14 ± 2.1	16 ± 3.33	19 ± 1.9	23 ± 2.6	12 ± 1.7	23 ± 2.6	19 ± 1.9	23 ± 2.6	12 ± 1.7	23 ± 2.6	12 ± 1.7	23 ± 2.6	23 ± 1.1	12 ± 1.7	23 ± 1.1	23 ± 1.1	23 ± 1.1	23 ± 1.1		
EF	0	0	11 ± 1.8	11 ± 3.2	12 ± 4.0	14 ± 2.1	15 ± 1.3	22 ± 2.1	23 ± 1.7	17 ± 1.1	20 ± 2.9	21 ± 3.3	21 ± 3.3	17 ± 1.1	20 ± 2.9	21 ± 3.3	21 ± 3.3	21 ± 3.3	21 ± 3.3	25 ± 1.1	21 ± 3.3	25 ± 1.1	25 ± 1.1	25 ± 1.1	25 ± 1.1		
KP	0	10 ± 2.0	12 ± 2.9	15 ± 3.0	20 ± 2.5	23 ± 2.4	27 ± 2.1	0	20 ± 2.1	24 ± 0.1	21 ± 1.9	21 ± 2.1	21 ± 2.1	24 ± 0.1	21 ± 1.9	21 ± 2.1	21 ± 2.1	21 ± 2.1	21 ± 2.1	24 ± 1.9	21 ± 2.1	24 ± 1.9	24 ± 1.9	24 ± 1.9	24 ± 1.9		
MRSA	0	0	16 ± 0.9	19 ± 1.1	22 ± 0.9	27 ± 0.3	31 ± 0.2	9 ± 3.9	15 ± 0.9	21 ± 1.6	0	7 ± 4.9	7 ± 4.9	21 ± 1.6	0	7 ± 4.9	7 ± 4.9	7 ± 4.9	7 ± 4.9	20 ± 2.9	7 ± 4.9	20 ± 2.9	20 ± 2.9	20 ± 2.9	20 ± 2.9		
PA	12 ± 3.9	13 ± 1.1	16 ± 2.1	18 ± 1.1	16 ± 1.1	22 ± 0.7	30 ± 0.8	>35	>35	25 ± 1.4	>35	>35	>35	25 ± 1.4	>35	>35	>35	>35	>35	>35	>35	>35	>35	>35	>35		
SA	11 ± 4.1	12 ± 1.7	14 ± 3.9	18 ± 2.1	23 ± 1.3	26 ± 0.8	30 ± 1.4	0	21 ± 1.4	25 ± 0.7	24 ± 0.4	24 ± 0.9	24 ± 0.9	25 ± 0.7	24 ± 0.4	24 ± 0.9	24 ± 0.4	24 ± 0.9	26 ± 0.9	26 ± 0.9	24 ± 0.9	26 ± 0.9	26 ± 0.9	26 ± 0.9	26 ± 0.9		
ST	0	0	10 ± 3.5	15 ± 2.5	19 ± 2.1	23 ± 0.9	28 ± 1.5	25 ± 0.9	26 ± 1.4	22 ± 1.6	25 ± 0.9	26 ± 2.1	26 ± 2.1	22 ± 1.6	25 ± 0.9	26 ± 2.1	25 ± 0.9	26 ± 2.1	26 ± 2.1	28 ± 0.3	26 ± 2.1	28 ± 0.3	28 ± 0.3	28 ± 0.3	28 ± 0.3		



method results indicated that the best results of the antibacterial activity were against MRSA and PA. Fig. S4† presents the optical image and the fluorescence image by photographing the agar plates with wells filled with Alq<sub>3</sub> NP sample concentrations and cultured with bacteria of MRSA (Figs. S4a and S4b†) and PA (Figs. S4c and S4d†) under the visible light and UV irradiation ( $\lambda = 385$  nm). As seen in the optical and fluorescence images, the inhibition zones resulting from the Alq<sub>3</sub> NP sample increased with the increase in the concentration of the sample. The fluorescence images clearly show the increase in the fluorescence glow with the increase in the concentration of the sample.

MIC and MBC are among the most important parameters to know to study the antibacterial activity of any material. That is why we used the resazurin assay to identify them. We performed a resazurin assay by serial dilutions for both Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples, besides GM as St. antibiotic. The serial dilutions started at 120  $\mu\text{g ml}^{-1}$  concentration in 96-well microtiter plates; the resulting serial dilution concentrations are 120, 60, 30, 15, 7.50, 3.75, 1.88, 0.94, 0.47, and 0.23  $\mu\text{g ml}^{-1}$ , diluted in 10 columns of 96-well microtiter plates in addition to the PC and NC in columns 11 and 12. After incubation for 24 hours, the absorbance reading of the 96-well microtiter plates was measured to calculate the IC<sub>50</sub>, which will be discussed later. Fig. S5† shows the 96-well microtiter plates where the wells were stained pink and blue. The wells that appear pink in color contain active bacteria and indicate that the samples' concentrations were not enough to kill bacteria. In contrast, the wells with blue color in which there was no bacterial activity indicate that the wells included samples' concentrations that reduced the activity of bacteria and killed them. Therefore, the more the number of blue columns, the more the efficiency of the antibacterial activity and the more the material effectiveness at lower concentrations. As shown in Fig. S5,† both the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples showed antibacterial activity, noting that the Alq<sub>3</sub> NP sample showed more antibacterial activity than that of the Alq<sub>3</sub> as rec sample. This figure also pointed out that the best antibacterial activity for both samples was against MRSA and PA. This result is consistent with those obtained by the AWD method (Fig. S2†). Besides, as shown in Fig. S5,† the concentrations in the columns colored in blue were considered the concentrations in which MIC and MBC could be. The obtained MIC values are summarized in Table 2. We did not find the MIC for both the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples against the bacterial strains EC, EF, and ST, while

testing all the diluted concentrations, including the concentration of 120  $\mu\text{g ml}^{-1}$ , which means that the values of MIC are higher than 120  $\mu\text{g ml}^{-1}$  (Table 2). At the same time, the MIC values for GM against EC, EF, and ST were 15, 15, and 3.75, respectively. The values of MIC in units of  $\mu\text{g ml}^{-1}$  for Alq<sub>3</sub> as rec, Alq<sub>3</sub> NPs, and GM against KP, MRSA, PA, and SA were (60, 30, 3.75), (30, 15, 7.5), (30, 15, 3.75), and (60, 30, 3.75), respectively. It can be noticed that the MIC for both the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples against MRSA and PA recorded the lowest concentrations. These results indicated that antibacterial activities against MRSA and PA for Alq<sub>3</sub> nanostructures (MIC  $\sim 15$   $\mu\text{g ml}^{-1}$ ) were higher than those of the microstructures (MIC  $\sim 30$   $\mu\text{g ml}^{-1}$ ). At the same time, they showed the lowest antibacterial activities against EC, EF, and ST (MIC  $> 120$   $\mu\text{g ml}^{-1}$ ). Besides, by comparing with St. antibiotic GM, the Alq<sub>3</sub> nanostructures recorded good values of the antibacterial activity against MRSA and PA (Table 2). The MBC values of both the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples, besides St. antibiotic against tested, were determined, and are listed in Table 2. We found that the MBC values showed the same tendency as the MIC values.

The IC<sub>50</sub>s of both samples against human pathogenic bacterial strains EC and EF have not been extracted because the fitted curves were not performed, the tested concentrations being lower than that required to produce enough inhibition to achieve the entire fitted curve (Fig. 6a and b). In contrast, the IC<sub>50</sub>s of the St. antibiotic GM were obtained against all tested human pathogenic bacterial strains (Table 2). The IC<sub>50</sub> of the microstructure of the Alq<sub>3</sub> as rec sample against human pathogenic bacterial strains KP, MRSA, PA, SA, and ST were 34.88, 9.25, 9.41, 19.74, and 52.15  $\mu\text{g ml}^{-1}$ , respectively. In contrast, for the nanostructures of the Alq<sub>3</sub> NP sample, IC<sub>50</sub>s were 21.69, 3.85, 3.96, 7.21, and 22.91  $\mu\text{g ml}^{-1}$ . At the same time, the IC<sub>50</sub>s of the microstructure of the Alq<sub>3</sub> as rec sample against human pathogenic bacterial strains EC, EF, KP, MRSA, PA, SA, and ST were 5.07, 6.24, 1.65, 1.91, 1.43, 1.57, and 2.48  $\mu\text{g ml}^{-1}$ , respectively (Table 2), where we could obtain the dose–response curves for the St. antibiotic against all tested human pathogenic bacterial strains (Fig. 6).

Although the IC<sub>50</sub>s were not extracted against bacterial strains of EC and EF, the number of blue columns can be used as an indicator to compare the samples of Alq<sub>3</sub> as rec and Alq<sub>3</sub> NPs. Figs. S5a and S5b† show that the number of blue columns when Alq<sub>3</sub> nanostructures were used as the antimicrobial material is higher than that of blue columns when using Alq<sub>3</sub>

Table 2 MIC, MBC, and IC<sub>50</sub> of the Alq<sub>3</sub> as rec sample, Alq<sub>3</sub> NP sample, and St. antibiotic GM against the tested EC, EF, KP, MRSA, PA, SA, and ST

Tested bacteria	MIC ( $\mu\text{g ml}^{-1}$ )			MBC ( $\mu\text{g ml}^{-1}$ )			IC <sub>50</sub> ( $\mu\text{g ml}^{-1}$ )		
	Alq <sub>3</sub> as rec	Alq <sub>3</sub> NPs	GM	Alq <sub>3</sub> as rec	Alq <sub>3</sub> NPs	GM	Alq <sub>3</sub> as rec	Alq <sub>3</sub> NPs	GM
EC	>120	>120	15	>120	>120	30	—	—	5.07
EF	>120	>120	15	>120	>120	30	—	—	6.24
KP	60	30	3.75	120	60	7.5	34.88	21.69	1.65
MRSA	30	15	7.5	60	30	15	9.25	3.85	1.91
PA	30	15	3.75	60	30	7.5	9.41	3.96	1.43
SA	60	30	3.75	120	60	7.5	19.74	7.21	1.57
ST	>120	>120	7.5	>120	>120	15	52.15	22.91	2.48





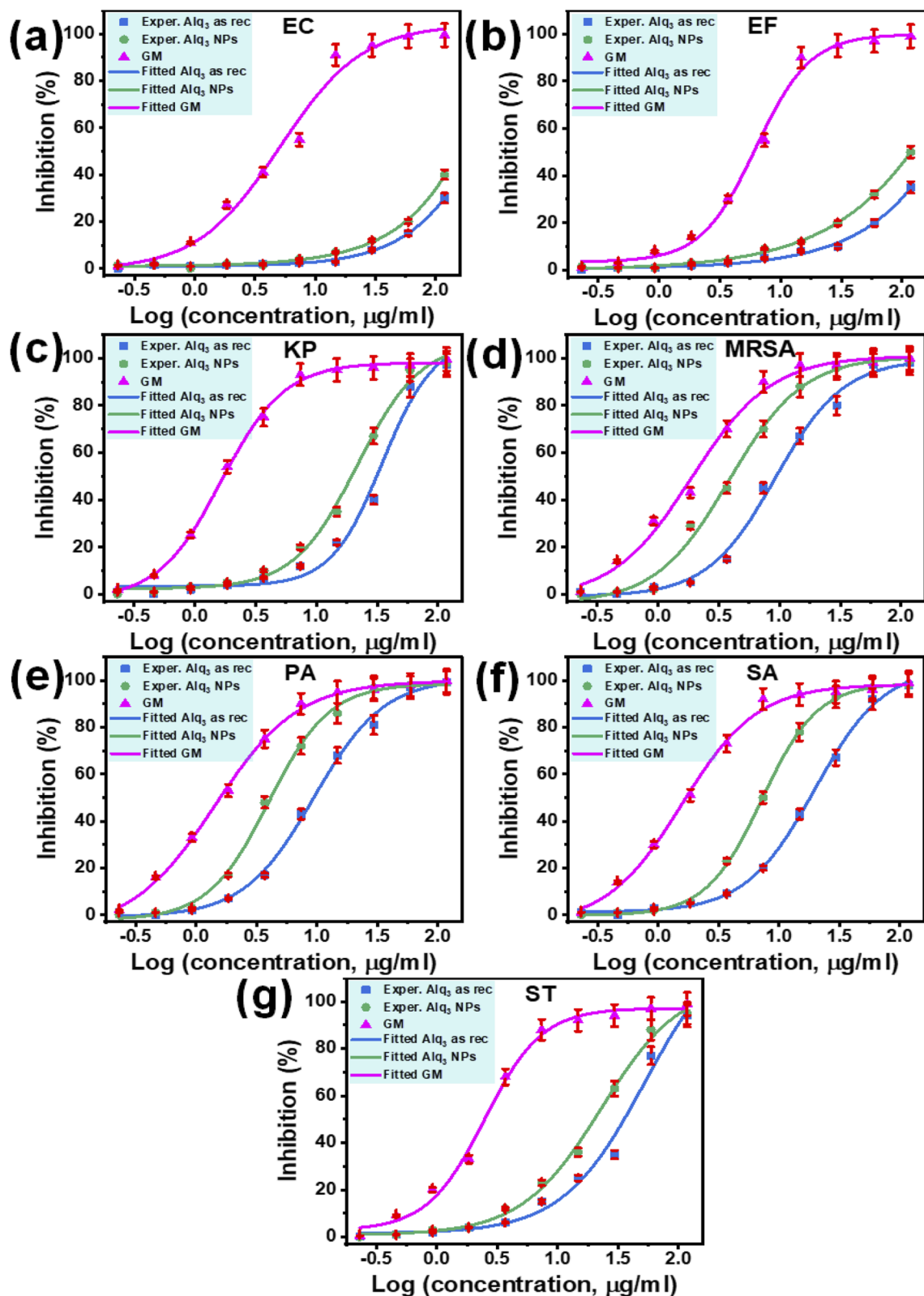


Fig. 6 Bacterial growth inhibition experimentally from the Alq<sub>3</sub> as rec sample, Alq<sub>3</sub> NP sample, and St. antibiotic GM versus common logarithms of the concentrations of 120, 60, 30, 15, 7.5, 3.75, 1.88, 0.94, 0.47, and 0.23  $\mu\text{g ml}^{-1}$  against (a) EC, (b) EF, (c) KP, (d) MRSA, (e) PA, (f) SA, and (g) ST. The lines are the fitted dose–response curves.



microstructures. These obtained results indicated that the antibacterial activities of Alq<sub>3</sub> nanostructures against all tested human pathogenic bacterial strains are more than those of the microstructures.

### 3.3. Antimicrobial activity mechanism

We found that the Alq<sub>3</sub> NP sample showed a higher antibacterial activity than that of the Alq<sub>3</sub> as rec sample through the AWD method and resazurin assay results. To explain this result, we put a possible scenario for the interaction of the two tested samples with bacterial strains, as shown in Fig. 7. Here, Fig. 7a and b represent the samples Alq<sub>3</sub> as rec and Alq<sub>3</sub> NPs, respectively; the Alq<sub>3</sub> as rec sample has microstructures, while the Alq<sub>3</sub> NP sample has nanostructures. At the same time, Fig. 7c and d represent the coccus and bacillus bacteria, respectively. Fig. 7c depicts coccus strains such as EC, EF, MRSA, and SA, while Fig. 7d represents bacillus bacterial strains such as KP, PA, and ST. When the bacterial strains are incubated with the sample Alq<sub>3</sub> as rec, which has a microstructure, the contact area between the sample and the cell walls of the bacteria is less because of the particle sizes of the sample Alq<sub>3</sub> as rec in microstructures, which cannot completely cover the cell walls of the bacteria that leads to an area with no contact (Fig. 7e), subsequently reducing the interaction between the sample's particles and the bacteria.

We think that the interaction between particles and bacteria could depend on direct contact, whereas in the case of sample Alq<sub>3</sub> NPs, whose particles have nanostructures, they allow for complete connection between them and the bacteria walls (Fig. 7f), leading to an increase in the interaction between the particles of the sample Alq<sub>3</sub> NPs and the bacteria, and thus an increase in the efficiency of the antibacterial activity of the Alq<sub>3</sub> NP sample. Hence, the surface area of the nanostructures increases as one of the features of the nanostructures, leading to an increase in the efficiency of the nanostructures during interactions. Although the increase in the contact between Alq<sub>3</sub> and the bacteria (surface interactions) could interpret why Alq<sub>3</sub>

nanostructures have more antibacterial activity than the microstructures, this not answer why Alq<sub>3</sub> stops the bacteria from growing; in other words, why Alq<sub>3</sub> kills the bacteria. The chemical structure of the Alq<sub>3</sub> could be the key to answering this question; where Alq<sub>3</sub> is an organometallic material, every Alq<sub>3</sub> molecule contains three molecules of 8-hydroxyquinoline. Herein, 8-hydroxyquinoline derivatives were reported to treat a microbial infection.<sup>38</sup> Moreover, it was reported that 8-hydroxyquinoline rapidly inhibits RNA synthesis.<sup>39</sup> Many studies<sup>40–42</sup> also demonstrated that its derivatives inhibit the methionine aminopeptidases, which remove the amino-terminal methionine residue from newly synthesized proteins. Accordingly, inhibiting the bacterial RNA synthesis and methionine aminopeptidase by 8-hydroxyquinoline could inhibit the growth of the bacteria and kill them.

To test the hypothesis illustrated in Fig. 7, we carried out an experiment in which three groups of MRSA were grown in the MHB media for 3 h; one group was grown in MHB only; the other two groups were grown in MHB with Alq<sub>3</sub> as rec and Alq<sub>3</sub> NPs at 20 μg ml<sup>-1</sup>. Then, 100 μl was placed on the silicon wafer from each group to obtain SEM images. The captured SEM images are shown in Fig. 8, including the SEM images of the powders Alq<sub>3</sub> as rec (Fig. 8a) and Alq<sub>3</sub> NPs (Fig. 8b). While at two magnifications, Fig. 8c and d show the SEM images of MRSA without the addition of Alq<sub>3</sub> samples into their MHB. Fig. 8e and f display the SEM images at two magnifications for MRSA grown in MHB with the Alq<sub>3</sub> as rec sample at 20 μg ml; these two images show the contact between MRSA cells and Alq<sub>3</sub> in microstructures, where they cannot contact the cell surface completely; this result is in agreement with the hypothesis illustrated in Fig. 7e. While Fig. 8g and h display the SEM images at two magnifications for MRSA grown in MHB with Alq<sub>3</sub> NPs at 20 μg ml<sup>-1</sup>, it can be seen that the nanostructures of Alq<sub>3</sub> surrounded the MRSA cells and more point contact between the Alq<sub>3</sub> and the cells; this result could confirm the hypothesis in Fig. 7f. It can also be seen that the shapes of the MRSA cells somewhat changed, which could be attributed to the killing of MRSA by Alq<sub>3</sub> NPs.

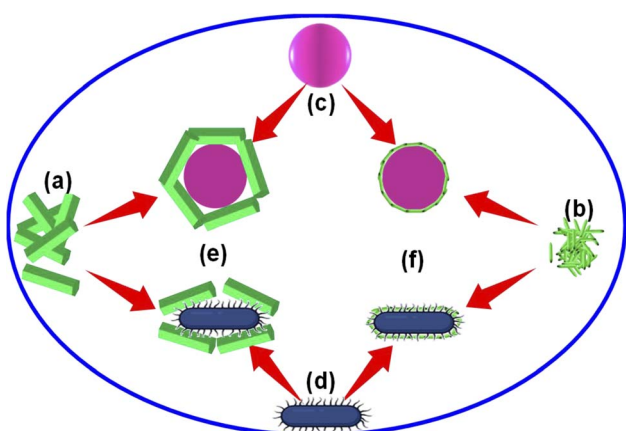


Fig. 7 Schematic diagram presenting the antimicrobial activity mechanism of the Alq<sub>3</sub> samples: (a) Alq<sub>3</sub> as rec, (b) Alq<sub>3</sub> NPs, (c) coccus bacteria, (d) bacillus bacteria, (e) cross-section of bacteria surrounded by the microparticles of the sample Alq<sub>3</sub> as rec and (f) cross-section of bacteria surrounded by the nanoparticles of the sample Alq<sub>3</sub> NPs.

### 3.4. PS/Alq<sub>3</sub> as an antibacterial coating

The results of the Alq<sub>3</sub> as rec and Alq<sub>3</sub> NP samples tested against EC, EF, KP, MRSA, PA, SA, and ST showed antibacterial activity, particularly in MRSA and PA strains, which indicates the possible use of Alq<sub>3</sub> in new biological applications. In applying these results, we prepared the PS/Alq<sub>3</sub> composite, where Alq<sub>3</sub> constituted 2% of the content of the entire composite. We tested the PS/Alq<sub>3</sub> composite as an antibacterial composite against the strain of bacteria MRSA because this strain of bacteria was the one in which the Alq<sub>3</sub> samples showed the best antibacterial activity. The composite was used to coat the 10 ml glass beakers. The MRSA strain was grown in four groups of 10 ml glass beakers: the negative control group (NC), a group coated with a PS polymer only, a group coated with a PS/Alq<sub>3</sub> composite, and a group with St. antibiotic GM (5 μg ml<sup>-1</sup>) as the positive control (see 2.5. subsection). Fig. 9a presents the image of the 10 ml glass beakers coated with a PS polymer and a PS/Alq<sub>3</sub> composite under visible light irradiation. At the same time,



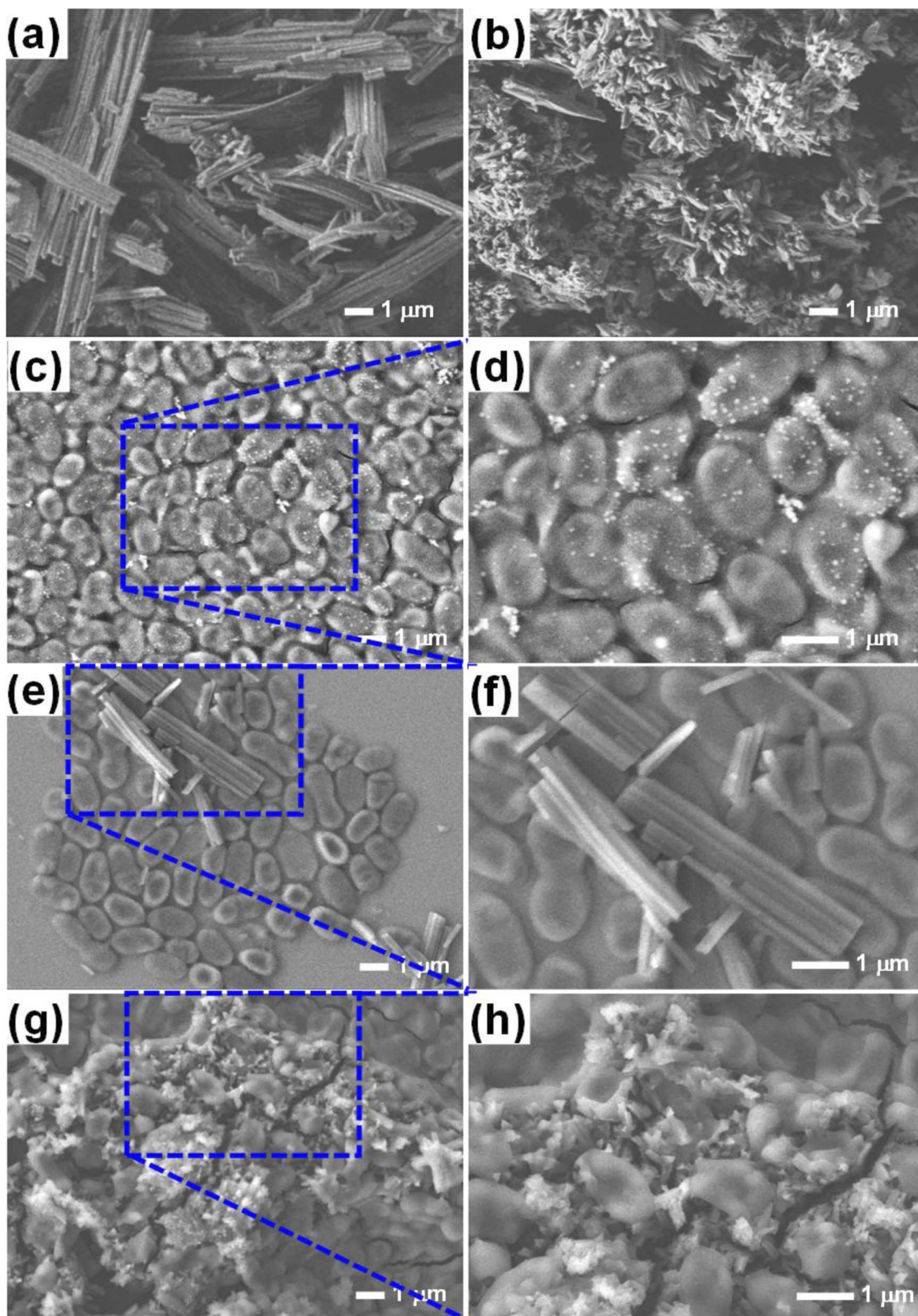


Fig. 8 SEM images of (a) Alq<sub>3</sub> as rec, (b) Alq<sub>3</sub> NP, (c and d) MRSA growth in MHB without treatment with Alq<sub>3</sub>, (e and f) MRSA growth in MHB including 20 µg ml<sup>-1</sup> of Alq<sub>3</sub> as rec, and (g and h) MRSA growth in MHB including 20 µg ml<sup>-1</sup> of Alq<sub>3</sub> NPs.

Fig. 9b shows the fluorescence image under UV irradiation at an excitation wavelength of 385 nm. It can be observed that 10 ml glass beakers coated with a PS/Alq<sub>3</sub> composite appear to glow

green, which belongs to Alq<sub>3</sub>. Fig. 9c presents the SEM image of the coated 10 ml glass beaker cross-sections, where the coating thickness is 3 µm. Fig. 9d shows the histogram of the bacterial



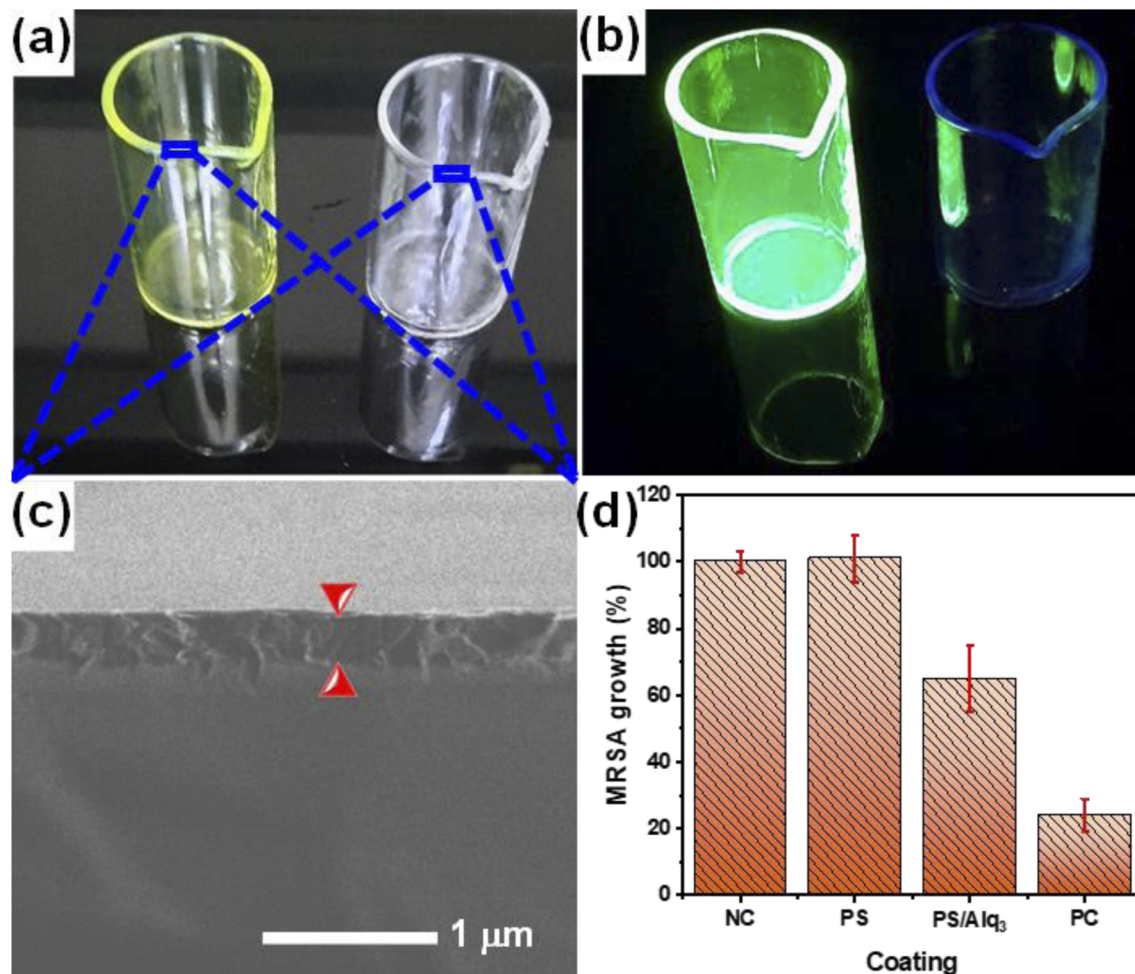


Fig. 9 PS/Alq<sub>3</sub> as an antibacterial composite: (a) and (b) 10 ml glass beaker coated with a PS polymer and a PS/Alq<sub>3</sub> composite under visible light and UV irradiation ( $\lambda = 385$  nm), respectively; (c) SEM image of the cross-sections of the coated beakers; and (d) histogram demonstrating the MRSA growth ratio after incubation for 24 h in the NC, PS, PS/Alq<sub>3</sub> (2% Alq<sub>3</sub>), and PC ( $5 \mu\text{g ml}^{-1}$ ) group beakers.

growth ratio for MRSA incubated for 24 h in the four groups of beakers, where it shows that the bacterial growth ratio in the beakers coated with the PS polymer did not change compared with the control group. In contrast, the percentage of bacteria growth in the beakers coated with a PS/Alq<sub>3</sub> composite decreased to 65% compared with the NC group. The percentage of bacteria growth in the PC ( $5 \mu\text{g ml}^{-1}$  of GM) group decreased to 24% compared with that in the NC group. The results indicated that the presence of Alq<sub>3</sub> in the PS/Alq<sub>3</sub> composite inhibited bacterial activity and could be used as a coating antimicrobial composite.

## 4. Conclusion

The antibacterial activity of Alq<sub>3</sub> has been extensively studied in its micro and nanostructures, where it was tested against seven different types of HPBS and compared with six kinds of standard antibiotics. The results indicated that the Alq<sub>3</sub> as the rec sample in microrod shapes and nanostructures in rice-like shapes that assembled into flower shapes have inhibition properties against bacterial growth. However, the nanostructures with a new

morphology revealed more activity against the bacterial growth than the microstructures, even revealing an activity similar to the standard antibacterial activity. To apply these results, Alq<sub>3</sub> was added to PS to form an antimicrobial composite coating, which showed a reduction in bacterial growth to 65% in the glass beakers coated with this fabricated composite. From the results of this study, we can conclude that Alq<sub>3</sub> can be used as a promising antimicrobial material; it can also be added to polymers to form composites as an antimicrobial coating.

## Conflicts of interest

There are no conflicts of interest to declare.

## Abbreviations

Alq <sub>3</sub>	Tris(8-hydroxyquinoline)aluminum
AP	<i>Ampicillin</i>
AU	<i>Augmentin</i>
AWD	Agar well diffusion



## Paper

CFU	Colony-forming units
EC	<i>Escherichia coli</i> ATCC 11775
EF	<i>Enterococcus faecalis</i> ATCC 29212
fac	Facial isomer
FOX	<i>Cefoxitin</i>
FTIR	Fourier transform infrared
GM	Gentamicin
HCT116	Human colon cancer cell lines
HFF-1	Human foreskin fibroblasts cell line
HPBS	Human pathogenic bacterial strains
IC50	Half-maximal inhibitory concentration
KF	<i>Cephalothin</i>
KP	<i>Klebsiella pneumoniae</i> ATCC 13883
MBC	Minimum bactericidal concentration
MCF7	Human breast cancer cell line
mer	Meridional isomer
MHA	Mueller-Hinton agar
MHB	Mueller-Hinton broth
MIC	Minimal inhibitory concentration
MRSA	Methicillin-resistant <i>Staphylococcus aureus</i> ATCC 33591
NC	Negative control
OLED	Organic light-emitting diode
PA	<i>Pseudomonas aeruginosa</i> ATCC 9027
PBS	Phosphate-buffered saline
PC	Positive control
PL	Photoluminescence
PS	Polystyrene
SA	<i>Staphylococcus aureus</i> ATCC 12600
SD	Standard deviation
SEM	Scanning electron microscopy
ST	<i>Salmonella Typhimurium</i> ATCC 14028
St.	Standard antibiotics
antibiotics	
TS	Cotrimoxazole
UV	Ultraviolet
XRPD	X-ray powder diffraction

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