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Designing photolabile ruthenium polypyridyl crosslinkers for hydrogel formation and multiplexed, visible-light degradation†

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Photoresponsive materials afford spatiotemporal control over desirable physical, chemical and biological properties. For advanced applications, there is need for molecular phototriggers that are readily incorporated within larger structures, and spatially-sequentially addressable with different wavelengths of visible light, enabling multiplexing. Here we describe spectrally tunable ($\lambda_{\text{max}} = 420\text{--}530\text{ nm}$) ruthenium polypyridyl complexes functionalized with two photolabile nitrile ligands that present terminal alkynes for subsequent crosslinking reactions, including hydrogel formation. Two Ru crosslinkers were incorporated within a PEG–hydrogel matrix, and sequentially degraded by irradiation with 592 nm and 410 nm light.

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Introduction

Photoresponsive molecules and materials are transforming multiple areas of research, from drug delivery,^{1–6} to materials engineering,^{7–15} and biology.^{16–26} Many natural biological processes are not photoresponsive, making light a versatile trigger for controlling complex biological systems.²⁷ The incorporation of photoactive moieties within biomolecules,²⁴ small-molecule drugs,²⁸ and materials⁷ provides a method for modulating their activity. Likewise, photoactive moieties incorporated within soft materials, *e.g.*, polymers, hydrogels, and elastomers, enable spatiotemporally precise, light-guided modulation of structure–function properties. Photoresponsive hydrogels in particular have long been used as platforms for cell growth and delivery, for small and large molecule drug delivery,^{29,30} and for basic materials applications.³¹ To expand methods for tuning soft material properties, *e.g.*, shape and viscosity, we developed differentially photoresponsive ruthenium moieties suitable for hydrogel formation and subsequent multiplexed ligand dissociation.

A drawback to most current photoresponsive molecules is the high-energy light required for bond dissociation. Common photoresponsive organic chromophores, *e.g.*, *o*-nitrobenzyl,³² azobenzene,¹⁴ and coumarin,^{30,33} respond to near-UV and blue light, which barely penetrates most

biomaterials or live tissue. Attempts to red-shift the activation wavelength have focused on multiphoton excitation,^{10,34–37} coupling with upconverting nanoparticles^{36,38} or chemically modified chromophores.³⁹ Some limiting factors include the small activation volume of multiphoton processes, the potential toxicity of embedded nanoparticles, low quantum yields (leading to sample heating and photo-damage during repeated illumination), and synthetic complexity.

To address these challenges, we have worked to develop inorganic photoactive molecules that absorb orange-red light, which has greater penetration depth and is less prone to photodamage in clinical applications.³⁷ Our laboratory has expanded the use of photolabile ruthenium crosslinkers for applications in biology and soft materials. The first Ru-based crosslinker, (Ru(bipyridine)₂(3-ethynylpyridine)₂) (Ru-BEP), presented two alkynes for the circularization of antisense bis-azide-modified oligonucleotides for light-activated gene knockdown in zebrafish embryos.²¹ A related compound, Ru(bipyridine)₂(3-pyridinaldehyde)₂ (RuAldehyde), provided a light-responsive crosslinker for hydrogel formation, site-selective degradation and protein release.²⁹

These Ru polypyridyl complexes share the unique ability to exchange a monodentate pyridine ligand with solvent upon irradiation with visible light. Single-site photo-substitution has been observed for other [Ru(polypyridyl)₂X₂]²⁺ complexes, where X = pyridine-^{2,40–42} or sulphur-containing⁴³ ligands. Alternatively, two nitrile ligands^{44,45} can both undergo rapid photo-substitution (Fig. 1A). Excitation into the singlet metal-to-ligand charge transfer (¹MLCT) band initiates intersystem crossing to a low-lying triplet state (Fig. 1B). In most photo-responsive Ru-polypyridyl complexes this triplet state is primarily ³MLCT in character, with another triplet metal-

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Table 1 Absorptivities and quantum yields for 1–3

	ϵ ($M^{-1} \text{ cm}^{-1}$)	Φ_{pr}
1	6140 \pm 100	0.16 \pm 0.02@450 nm
2	1900 \pm 100	0.19 \pm 0.005@532 nm
3	7400 \pm 400	0.07 \pm 0.01@532 nm

70 nm was observed with the first substitution of a bipyridine for biquinoline ligand, from 1 to 2 (Fig. 2A), followed by a nearly 40 nm red-shift from 2 to 3. This shows good agreement with previously published spectra for Ru(phen)₂(MeCN)₂ ($\lambda_{\text{max}} = 420$ nm), Ru(phen)(biq)(MeCN)₂ ($\lambda_{\text{max}} = 497$ nm), and Ru(biq)₂(MeCN)₂ ($\lambda_{\text{max}} = 535$ nm).⁴⁶

The photolysis of ruthenium polypyridyl compounds can be observed directly using UV-Vis spectroscopy. As the compound undergoes ligand exchange of a coordinated ligand for a solvent molecule, a significant red shift is observed in the MLCT band. Under continuous irradiation, compounds 1–3 sequentially exchanged both nitrile ligands (Fig. 3A, S2[†]). The UV-Vis photolysis curve for 3 is shown in Fig. 3B, where peaks at 560 and 590 nm indicated a stepwise process, with a monoaquated intermediate. The clear isosbestic points at 550 and 570 nm also indicated the stepwise transition from 3 to monoaquated 3' to bisaquated 3'', although the first transition point at 550 nm included early formation of 3'' under continuous irradiation.

The loss of the second nitrile ligand in 3 was slower, occurring on the order of 40 min (Fig. 3B), compared to the first ligand exchange event, which was completed within 4 min of constant irradiation in the bulk sample. This trend was observed for 1 and 2 as well (Fig. S2[†]). The Ru MLCT band extends well beyond the λ_{max} , which can be used to induce ligand exchange at longer wavelengths of light; irradiation at 600–700 nm (red incandescent light bulb, 5 mW) was less efficient but led to complete photolysis of 3 in 4 h (Fig. S3[†]).

Photolysis data were fit to an equation derived from a pseudo-first order kinetics process, and the time constants were determined (Fig. S4[†]). The value of Φ_{pr} was found for the

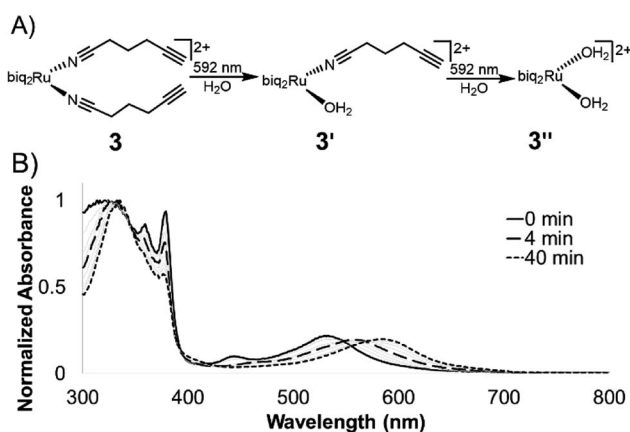


Fig. 3 Photolysis of 3 in water. (A) Compounds 1–3 undergo a stepwise ligand exchange of both nitrile ligands when irradiated in water. The second step takes much longer than the first. (B) Photolysis trace of 3 in water under irradiation from 592 nm LED (25 mW cm⁻²).

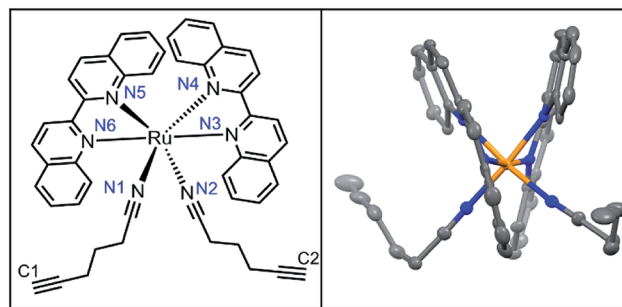


Fig. 4 Crystal structure of 3.

first exchange event from the rate constant coupled with the laser power (Fig. S4[†]). As expected, Φ_{pr} decreased roughly 2-fold as the MLCT band was shifted further to the red, from 0.16 (in 1) to 0.07 (in 3), Table 1.^{52,53} Similarly, the efficiency of photolysis for 3 ($\epsilon \times \Phi_{\text{pr}} = 520 \text{ M}^{-1} \text{ cm}^{-1}$) was lower than 1 at $980 \text{ M}^{-1} \text{ cm}^{-1}$. Although these values were lower than other published ruthenium caging groups, with efficiencies that range from 2000 (ref. 46) to $4000 \text{ M}^{-1} \text{ cm}^{-1}$,²⁹ they are significantly improved over other green-light-sensitive caging groups like BODIPY, with Φ_{pr} on the order of $100 \text{ M}^{-1} \text{ cm}^{-1}$.^{54,55}

Diffraction-quality crystals of 3 as PF₆⁻ salt were grown *via* vapour diffusion from 3–5 mg dissolved in acetonitrile/methanol/THF (0.1 mL each) with diethyl ether, stored at -20 °C for 2 weeks (Fig. 4). Bond lengths between Ru²⁺ and ligands were within expected ranges, with variations due to the steric strain in the system. The angle between the nitrile ligands is stretched significantly to $>95^\circ$ perhaps due to the strain caused by bulkier biq ligands coordinated to Ru²⁺. In the less crowded Ru(bpy)(biq)(5-hexynenitrile)₂ compound, the nitrile–Ru–nitrile angle is $\sim 90^\circ$ (Fig. S5[†]). Crystal structure of 3 shows alkynes positioned 4.3 Å and 4.9 Å from biquinolines, angled such that they are accessible for cycloaddition with azide and copper catalyst (Table 2).

The conformational flexibility of the nitrile-alkyl ligands required synthesis and testing of several Ru compounds to identify competent crosslinkers. Initially, Ru compounds employing a shorter 4-pentynenitrile ligand were synthesized and found to be incapable of Cu(I)-mediated PEG gelation (Fig. S6[†]). Incorporation of longer 5-hexynenitrile ligands led to functional Ru crosslinkers, but only after mild synthetic conditions for nitrile coordination were employed. Ru-(5-hexynenitrile) coordination performed at elevated temperatures and longer reaction times resulted in Ru compounds found to be incapable of Cu(I)-mediated PEG gelation. X-ray crystal

Table 2 Select bond lengths, from X-ray crystal structure of 3

Bond		3 (Å)
Ru–biq	Ru–N4	2.084(6)
	Ru–N3	2.093(6)
Ru–N≡C	Ru–N1	2.025(6)
	Ru–N2	2.024(6)
C≡C to biq	C1–biq	4.267
	C2–biq	4.926

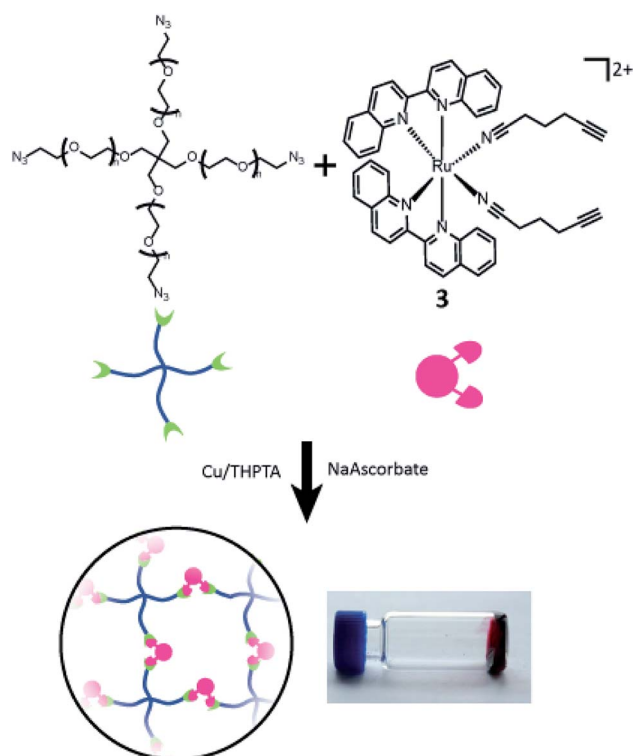


structure analysis of one such example shows alkyne positioned much closer to the biquinoline, only 3.7 Å (Fig. S5†). The *cis*-alkane conformation should disfavour Cu(I)-mediated alkyne-azide cycloaddition chemistry. Ru compounds 1–3 were synthesized using the mild conditions detailed in the Synthetic Procedure and confirmed to be excellent crosslinkers in gelation studies.

CuAACs have been widely used for materials design, with several studies showing the generation of hydrogel materials. Hyaluronic acid,^{56,57} polyethylene glycol (PEG),⁵⁸ dextran,⁵⁹ poly(vinyl) alcohol (PVA),⁶⁰ along with several other polymers have been modified with azides and terminal alkynes to facilitate hydrogel formation. The need for a Cu(I) catalyst has limited some bio-applications as it can be toxic to cells,⁶¹ but can also provide spatiotemporal control. In one example Bowman and co-workers used a photocatalyst to reduce Cu(II) for the formation of a hydrogel with precise control.⁶² Copper can be dialysed away from preformed hydrogels, which is acceptable for many drug delivery platforms.

Compounds 1–3 were tested for crosslinking reaction with azido-PEG (MW 10 000 Da) in the presence of CuSO₄, THPTA ligand, and sodium ascorbate reducing agent (Scheme 1), forming a strong hydrogel within 30 s (results shown for 3, Fig. 5). Hydrogels formed at a final weight percent of 7.5 wt% with stoichiometric ruthenium crosslinker, generating elasticity nearing 1 kPa (Fig. 5). As expected, when exposed to visible light (400–500 nm) the hydrogel rapidly lost its elastic properties, becoming a viscous liquid within 5 min (Fig. 5).

Next, a striped hydrogel was formed for multiplexing experiment *via* “layer-by-layer” reaction of azido-PEG with crosslinker



Scheme 1 Gelation of branched PEG *via* crosslinking reaction with 3.

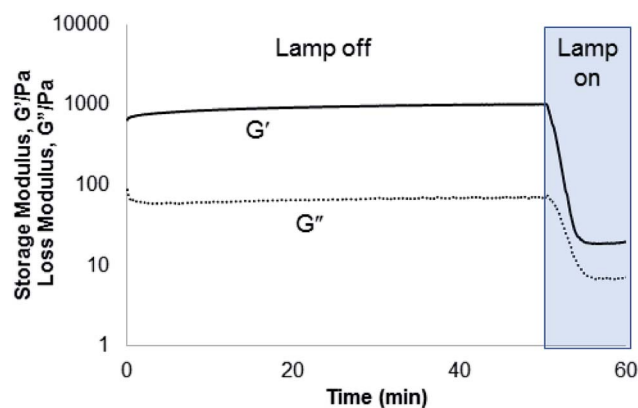


Fig. 5 Rheometry demonstrating gelation formed from the incorporation of 3 into a PEG hydrogel. The hydrogel was rapidly degraded under irradiation with 400–500 nm light (25 mW cm⁻²).

1, alternating with crosslinker 3 (Fig. 6). Orange light (592 nm) was used to degrade 3 selectively while leaving 1 intact, as demonstrated both in a solution experiment with equal parts 1 and 3 (Fig. 6A) and in the gel (Fig. 6B). A significant increase in absorbance at 590 nm confirmed the formation of the bis-quated product Ru(biq)₂(H₂O)₂, 3'' (Fig. 6A). Finally, irradiation at 410 nm led to a significant decrease in absorbance at 423 nm and small increase at 590 nm due to formation of Ru(bpy)₂(-H₂O)₂, 1'' (Fig. 6A, S1†), and rapidly degraded the remaining hydrogel sections crosslinked by 1 (Fig. 6B). The sequence of irradiation is important in this case, as compound 3 absorbs

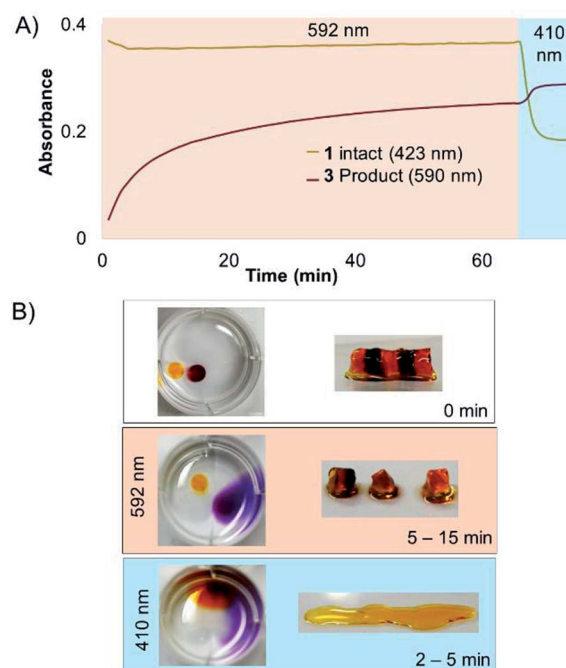


Fig. 6 Selective degradation of 1 and 3 in solution and hydrogel. (A) Irradiation at 592 nm photolyzed 3 in solution while leaving 1 intact, until irradiation at 410 nm. (B) Striped hydrogel incorporating alternating sections of 1 and 3 was selectively degraded at 592 nm, leaving the orange gel regions intact. The remaining sections crosslinked by 1 were degraded by 410 nm light.



both 410 and 592 nm light and will be degraded by both wavelengths.

We developed spectrally tuneable ruthenium polypyridyl crosslinkers with two pendant alkynes for hydrogel formation, and high photolysis efficiency for multiplexed, visible-light gel degradation. Replacing bipyridine with more pi-conjugated biquinoline ligands red-shifted the absorbance, generating a series of sequentially red-shifted compounds. Incorporation of two flexible 5-hexynenitrile ligands at the Ru²⁺ center enabled CuAAC crosslinking reactions, while also facilitating subsequent photodegradation of gels incorporating these Ru crosslinkers. This represents the first example to our knowledge of a two-color hydrogel system that can be selectively activated by two different visible wavelengths. Such Ru crosslinkers may be applied broadly in materials chemistry, or alternately employed for generating photoactive versions of circular oligos,²¹ peptides, or other bis-azide containing molecules.

Conflicts of interest

There are no conflicts to declare.

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References

- N. A. Smith, P. Zhang, S. E. Greenough, M. D. Horbury, G. J. Clarkson, D. McFeely, A. Habtemariam, L. Salassa, V. G. Stavros, C. G. Dowson, *et al.*, *Chem. Sci.*, 2017, **8**, 395–404.
- A. Li, C. Turro and J. J. Kodanko, *Chem. Commun.*, 2018, **54**, 1280–1290.
- F. Reefsing and W. Szymanski, *Curr. Med. Chem.*, 2018, **24**, 4905–4950.
- A. Döbber, A. F. Phoa, R. H. Abbassi, B. W. Stringer, B. W. Day, T. G. Johns, M. Abadleh, C. Peifer and L. Munoz, *ACS Med. Chem. Lett.*, 2017, **8**, 395–400.
- M. M. Dcona, D. Mitra, R. W. Goehe, D. A. Gewirtz, D. A. Lebman and M. C. T. Hartman, *Chem. Commun.*, 2012, **48**, 4755.
- S. Ki Choi, T. Thomas, M.-H. Li, A. Kotlyar, A. Desai and J. R. Baker Jr, *Chem. Commun.*, 2010, **46**, 2632.
- A. M. Kloxin, A. M. Kasko, C. N. Salinas and K. S. Anseth, *Science*, 2009, **324**, 59–63.
- A. Ovsianikov, A. Deiwick, S. Van Vlierberghe, P. Dubruel, L. Möller, G. Dräger and B. Chichkov, *Biomacromolecules*, 2011, **12**, 851–858.
- D.-S. Shin, J. You, A. Rahimian, T. Vu, C. Siltanen, A. Ehsanipour, G. Stybayeva, J. Sutcliffe and A. Revzin, *Angew. Chem., Int. Ed.*, 2014, **53**, 8221–8224.
- K. Peng, I. Tomatsu, B. van den Broek, C. Cui, A. V. Korobko, J. van Noort, A. H. Meijer, H. P. Spaink and A. Kros, *Soft Matter*, 2011, **7**, 4881.
- C. M. Kirschner, D. L. Alge, S. T. Gould and K. S. Anseth, *Adv. Healthcare Mater.*, 2014, **3**, 649–657.
- C. K. Arakawa, B. A. Badeau, Y. Zheng and C. A. DeForest, *Adv. Mater.*, 2017, **29**, 1703156.
- D. R. Griffin and A. M. Kasko, *J. Am. Chem. Soc.*, 2012, **134**, 13103–13107.
- A. M. Rosales, K. M. Mabry, E. M. Nehls and K. S. Anseth, *Biomacromolecules*, 2015, **16**, 798–806.
- T. E. Brown, I. A. Marozas and K. S. Anseth, *Adv. Mater.*, 2017, **29**, 1605001.
- L. Wu, Y. Wang, J. Wu, C. Lv, J. Wang and X. Tang, *Nucleic Acids Res.*, 2013, **41**, 677–686.
- L. Kröck and A. Heckel, *Angew. Chem., Int. Ed.*, 2005, **44**, 471–473.
- D. D. Young, M. O. Lively and A. Deiters, *J. Am. Chem. Soc.*, 2010, **132**, 6183–6193.
- S. Yamazoe, Q. Liu, L. E. McQuade, A. Deiters and J. K. Chen, *Angew. Chem., Int. Ed.*, 2014, **53**, 10114–10118.
- S. Yamazoe, I. A. Shestopalov, E. Provost, S. D. Leach and J. K. Chen, *Angew. Chem., Int. Ed.*, 2012, **51**, 6908–6911.
- J. C. Griepenburg, T. L. Rapp, P. J. Carroll, J. Eberwine and I. J. Dmochowski, *Chem. Sci.*, 2015, **6**, 2342–2346.
- J. C. Griepenburg, B. K. Ruble and I. J. Dmochowski, *Bioorg. Med. Chem.*, 2013, **21**, 6198–6204.
- B. N. Goguen, B. D. Hoffman, J. R. Sellers, M. A. Schwartz and B. Imperiali, *Angew. Chem., Int. Ed.*, 2011, **123**, 5785–5788.
- C. W. Riggsbee and A. Deiters, *Trends Biotechnol.*, 2010, **28**, 468–475.
- N. Ankenbruck, T. Courtney, Y. Naro and A. Deiters, *Angew. Chem., Int. Ed.*, 2018, **57**, 2768–2798.
- J. Hemphill, E. K. Borchardt, K. Brown, A. Asokan and A. Deiters, *J. Am. Chem. Soc.*, 2015, **137**, 5642–5645.
- S. Mura, J. Nicolas and P. Couvreur, *Nat. Mater.*, 2013, **12**, 991–1003.
- M. Huisman, J. K. White, V. G. Lewalski, I. Podgorski, C. Turro and J. J. Kodanko, *Chem. Commun.*, 2016, **52**, 12590–12593.
- T. L. Rapp, C. B. Highley, B. C. Manor, J. A. Burdick and I. J. Dmochowski, *Chem.–Eur. J.*, 2018, **24**, 2328–2333.
- M. A. Azagarsamy and K. S. Anseth, *Angew. Chem., Int. Ed.*, 2013, **52**, 13803–13807.
- D. Y. Wong, D. R. Griffin, J. Reed and A. M. Kasko, *Macromolecules*, 2010, **43**, 2824–2831.
- J. W. Walker, J. A. Mccray and G. P. Hess, *Biochemistry*, 1986, **25**, 1799–1805.
- M. A. Azagarsamy, D. D. McKinnon, D. L. Alge and K. S. Anseth, *ACS Macro Lett.*, 2014, **3**, 515–519.
- I. Aujard, C. Benbrahim, M. Gouget, O. Ruel, J.-B. Baudin, P. Neveu and L. Jullien, *Chem.–Eur. J.*, 2006, **12**, 6865–6879.



- 35 R. G. Wylie and M. S. Shoichet, *J. Mater. Chem.*, 2008, **18**, 2716.
- 36 B. Yan, J.-C. Boyer, N. R. Branda and Y. Zhao, *J. Am. Chem. Soc.*, 2011, **133**, 19714–19717.
- 37 X. Zeng, X. Zhou and S. Wu, *Macromol. Rapid Commun.*, 2018, 1800034.
- 38 B. Yan, J.-C. Boyer, D. Habault, N. R. Branda and Y. Zhao, *J. Am. Chem. Soc.*, 2012, **134**, 16558–16561.
- 39 D. Wang, M. Wagner, H.-J. Butt and S. Wu, *Soft Matter*, 2015, **11**, 7656–7662.
- 40 Z. Leonardo, C. Cecilia, A. Pablo, L. Baraldo and R. Etchenique, *J. Am. Chem. Soc.*, 2003, **125**, 882–883.
- 41 M. Huisman, J. K. White, V. G. Lewalski, I. Podgorski, C. Turro and J. J. Kodanko, *Chem. Commun.*, 2016, **52**, 12590–12593.
- 42 L. N. Lameijer, D. Ernst, S. L. Hopkins, M. S. Meijer, S. H. C. Askes, S. E. LeDévédec and S. Bonnet, *Angew. Chem., Int. Ed.*, 2017, **56**, 11549–11553.
- 43 R. N. Garner, L. E. Joyce and C. Turro, *Inorg. Chem.*, 2011, **50**, 4384–4391.
- 44 R. Sharma, J. D. Knoll, P. D. Martin, I. Podgorski, C. Turro and J. J. Kodanko, *Inorg. Chem.*, 2014, **53**, 3272–3274.
- 45 T. Respondek, R. Sharma, M. K. Herroon, R. N. Garner, J. D. Knoll, E. Cueny, C. Turro, I. Podgorski and J. J. Kodanko, *ChemMedChem*, 2014, **9**, 1306–1315.
- 46 B. A. Albani, C. B. Durr and C. Turro, *J. Phys. Chem. A*, 2013, **117**, 13885–13892.
- 47 J. D. Knoll, B. A. Albani, C. B. Durr and C. Turro, *J. Phys. Chem. A*, 2014, **118**, 10603–10610.
- 48 X. Zeng, X. Zhou and S. Wu, *Macromol. Rapid Commun.*, 2018, 1800034.
- 49 C. Xie, W. Sun, H. Lu, A. Kretschmann, J. Liu, M. Wagner, H.-J. Butt, X. Deng and S. Wu, *Nat. Commun.*, 2018, **9**, 3842.
- 50 V. V. Rostovtsev, L. G. Green, V. V. Fokin and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2002, **41**, 2596–2599.
- 51 C. Tornøe, C. Christensen and M. Meldal, *J. Org. Chem.*, 2002, **67**, 3057–3064.
- 52 L. N. Lameijer, D. Ernst, S. L. Hopkins, M. S. Meijer, S. H. C. Askes, S. E. LeDévédec and S. Bonnet, *Angew. Chem., Int. Ed.*, 2017, **56**, 11549–11553.
- 53 L. M. Loftus, A. Li, K. L. Fillman, P. D. Martin, J. J. Kodanko and C. Turro, *J. Am. Chem. Soc.*, 2017, **139**, 18295–18306.
- 54 P. P. Goswami, A. Syed, C. L. Beck, T. R. Albright, K. M. Mahoney, R. Unash, E. A. Smith and A. H. Winter, *J. Am. Chem. Soc.*, 2015, **137**, 3783–3786.
- 55 N. Umeda, H. Takahashi, M. Kamiya, T. Ueno, T. Komatsu, T. Terai, K. Hanaoka, T. Nagano and Y. Urano, *ACS Chem. Biol.*, 2014, **9**, 2242–2246.
- 56 V. Crescenzi, L. Cornelio, C. Di Meo, S. Nardecchia and R. Lamanna, *Biomacromolecules*, 2007, **8**, 1844–1850.
- 57 X. Hu, D. Li, F. Zhou and C. Gao, *Acta Biomater.*, 2011, **7**, 1618–1626.
- 58 S. Q. Liu, P. L. Rachel Ee, C. Y. Ke, J. L. Hedrick and Y. Y. Yang, *Biomaterials*, 2009, **30**, 1453–1461.
- 59 D. A. Heller, Y. Levi, J. M. Pelet, J. C. Doloff, J. Wallas, G. W. Pratt, S. Jiang, G. Sahay, A. Schroeder, J. E. Schroeder, *et al.*, *Adv. Mater.*, 2013, **25**, 1449–1454.
- 60 D. A. Ossipov and J. Hilborn, *Macromolecules*, 2006, **39**, 1709–1718.
- 61 Y. Jiang, J. Chen, C. Deng, E. J. Suuronen and Z. Zhong, *Biomaterials*, 2014, **35**, 4969–4985.
- 62 B. J. Adzima, Y. Tao, C. J. Kloxin, C. A. DeForest, K. S. Anseth and C. N. Bowman, *Nat. Chem.*, 2011, **3**, 256–259.

