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Polymeric biomaterials for the delivery of platinum-based anticancer drugs

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Since cisplatin, *cis*-diamminedichloroplatinum(II), received FDA approval for use in cancer treatment in 1978, platinum-based drugs have been one of the most widely used drugs for the treatment of tumors in testicles, ovaries, head and neck. However, there are concerns associated with the use of platinum-based anticancer drugs, owing to severe side effects and drug resistance. In order to overcome these limitations, various drug-delivery systems have been developed based on diverse organic and inorganic materials. In particular, the versatility of polymeric materials facilitates the tuning of drug-delivery systems to meet their primary goals. This review focuses on the progress made over the last five years in the application of polymeric nanoparticles for the delivery of platinum-based anticancer drugs. The present article not only describes the fundamental principles underlying the implementation of polymeric nanomaterials in platinum-based drug delivery, but also summarizes concepts and strategies employed in the development of drug-delivery systems.

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1. Introduction

Chemotherapy, *i.e.*, the treatment of diseases by administration of chemical compounds, is currently one of the most effective ways to treat cancer. Among a variety of anticancer agents, cisplatin, a leading platinum (Pt)-based anticancer drug, has been used for more than three decades, either as a

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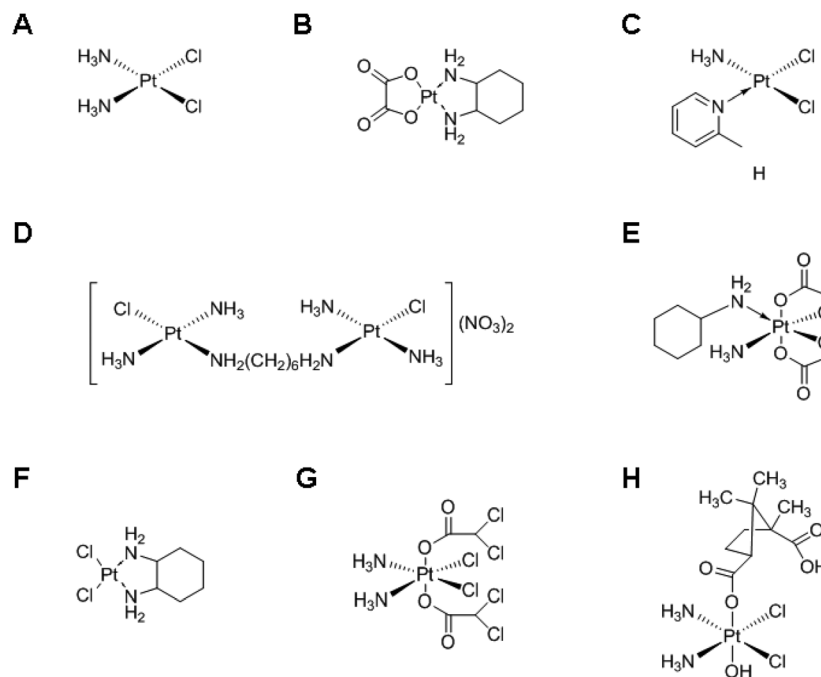
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single therapeutic agent or in combination with other agents.^{1,2} Following the success of cisplatin, various Pt-based drugs (Fig. 1) have been developed, although only a small subset has received US Food and Drug Administration (FDA) approval. Nowadays, they play a crucial role in the treatment of various cancers, such as cancers in testicles, ovaries, head, and neck.^{3–5} However, the use of Pt-based anticancer drugs is associated with drug resistance and inactivation by vascular and cellular components, and is associated with severe side effects incurred by the systemic delivery, such as nephrotoxi-

city, ototoxicity, neurotoxicity, and emetogenesis.^{6–9} Therefore, various delivery systems have been developed to prevent the shortcomings of Pt-based chemotherapy and to increase its efficacy.^{10–12}

Approaches like carrier-based drug delivery generally exploit the difference between normal and tumor tissues for increasing the efficacy and selectivity of a drug towards the target tissue. More specifically, the enhanced permeability and retention (EPR) effect is based on the increased permeability of nano-sized macromolecules coupled with the poor lymphatic



clearance and slow venous return in the tumor tissues.^{13–17} Long-circulating nanocarriers have greater chances to be exposed to a tumor site, compared to low-molecular weight drugs which are rapidly cleared from the circulation. Thus, the EPR effect results in passive targeting of drug-loaded nanoparticles to the tumor tissues. In some cases, this can be further enhanced by active targeting, the use of nanoparticles functionalized with ligands for selectively binding to tumor-specific moieties exposed at the target cells.^{16–19} Such moieties generally include transporters, antigens, or receptors whose quantity or functionality is higher in tumors compared to normal tissues. Drug-delivery systems based on nanocarriers include micelles,^{20,21} liposomes,²² dendrimers,²³ silica nanoparticles,²⁴ and organic/inorganic hybrid nanoparticles.²⁵ The most fascinating features of polymer-based delivery systems arise from the versatility of the polymer sources and their combinatorial synthesis. This versatility enables an easy tuning of the properties of the nanoparticles to meet the purpose of using delivery systems.²¹

For the successful drug delivery, drug encapsulation efficiency is one of the important indexes, especially in the case of poorly bioavailable drugs. To maximize the efficacy of anticancer drugs, the developed drug delivery systems must contain large amounts of drugs as much as possible. The polymeric nanoparticles facilitate efficient encapsulation of drugs into the nanocarriers by exploiting the compatibility between drugs and matrixes of polymeric nanoparticles or by employing the chemical conjugation between the polymeric backbone and drugs. In addition, the nanoparticles must release the drugs at a controlled rate. The controlled release of drugs has been achieved through the utilization of stimuli-responsive chemical bonds between drugs and the polymeric backbone or among polymeric blocks comprising the nanoparticles. Therefore, the ultimate goal of this review is to discuss the strategies for loading Pt-based drugs into the polymeric nanoparticles

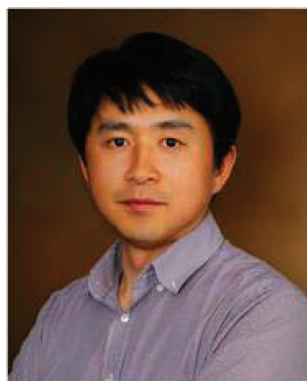
efficiently and for modulating their release in an appropriate manner, which affect significantly the anticancer efficacy.

In this review, we discuss the progress in the field of polymeric delivery systems of Pt-based anticancer drugs over the past five years. The article is organized into the following sections: first, we briefly introduce the characteristics of each structurally different Pt-based anticancer drug, involving both Pt(II)- and Pt(IV)-based drugs; then, we systematically discuss the encapsulation needed for delivering the Pt-based drugs; finally, we highlight the conjugation methods used for delivering Pt(II)- and Pt(IV)-based drugs.

2. Structure and cellular mechanism of Pt(II) and Pt(IV) compounds

Although numerous Pt-based drugs have been developed, most of them can be categorized into two groups, based on Pt(II) and Pt(IV), according to the molecular structure. As a background reference for the subsequent sections, here we provide a comprehensive summary of the structure and cellular mechanism of Pt(II) and Pt(IV)-based drugs.

The most representative Pt(II)-based drug is cisplatin, which is the *cis* isomer of the square planar configuration (Fig. 1A).^{26,27} According to the HSAB (Hard-Soft Acid-Base) theory, Pt can be defined as a soft metal with high affinity for soft non-metal sulfur.^{26,27} Therefore, during blood circulation, unintended deactivation of cisplatin may be caused by biomolecules containing thiol groups, such as cysteine and human serum albumin.^{26–29} Some studies have demonstrated that about 65–98% of Pt becomes bound to blood proteins one day after cisplatin administration.^{28,29} Therefore, only a small number of Pt(II)-based drugs can maintain their activity during blood circulation and enter cancer cells mainly by passive diffusion and/or by the copper transporter CTR1 (Fig. 2).^{3,26–30} The relatively low concentration of chloride in the cells facilitates the replacement of chloro ligands in Pt(II)-based drugs by water, resulting in the mono-aquated Pt complex. The aquated form is reactive to guanine and adenine in DNA. The resulting intra- and inter-strand crosslinks distort the DNA duplex, causing the apoptosis of cancer cells.^{3,26,27,31,32} However, in some cases, the efficacy of cisplatin may be limited by inherent or acquired cisplatin resistance. The major mechanisms of resistance can be classified into four categories: inactivation by increased thiol-containing molecules, an increased DNA-repair capacity, reduced uptake/enhanced efflux, and failure of apoptotic pathways.^{3,26,30,31} Oxaliplatin (Fig. 1B) is a successful alternative drug to overcome the limitations of cisplatin in the treatment of cisplatin-resistant tumors. It has been reported that the bulky diaminocyclohexane (DACH) ligand of oxaliplatin contributes to overcoming the drug resistance arising from the mechanism of DNA mismatch repair recognition.^{3,30,33–35} In addition, *cis*-ammine(2-methylpyridine)dichloroplatinum(II) (ZD0473) is another alternative drug candidate for overcoming the cisplatin resistance induced by increased thiol-containing cellular components (Fig. 1C). It has been proposed that the



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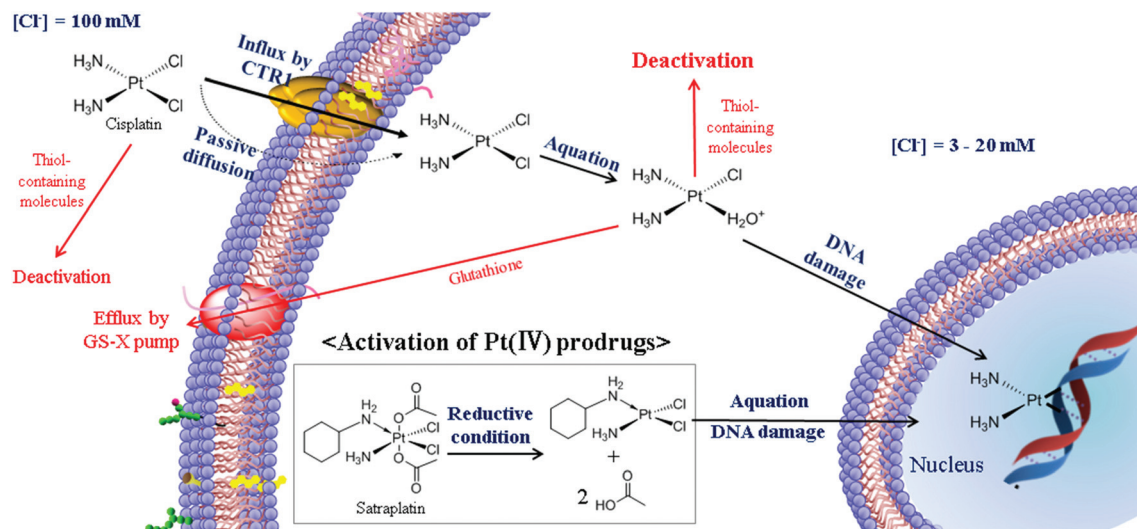


Fig. 2 Schematic diagram of the cytotoxic pathway of Pt-based drugs. Cisplatin is a representative. After entering the cell by copper transferase receptor (CTR1) or passive diffusion, cisplatin was activated by replacing a chloride ion with water molecules in low concentration of chloride ions (aquation). The aquated cisplatin binds to DNA. Distortion of DNA activates the programmed cell death. Pt(IV) prodrugs were activated by reduction of two axial positions, resulting in the release of active Pt(II) complexes. After this, the mechanism is similar to cisplatin.

steric hindrance of ZD0473 prevents the approach of glutathione (GSH). This strategy is not only able to maintain the cytotoxic ability of Pt-based drugs, but is also able to reduce deactivation by GSH.^{3,30,35} Another strategy to avoid cisplatin resistance involves BBR3005, a multinuclear Pt(II)-based drug (Fig. 1D). By inducing the irreversible B $\rightarrow$ Z transition in DNA structures with long range intra- and inter-strand crosslinks, multinuclear Pt(II)-based drugs prevent the detection of damaged DNA by DNA damage-recognition proteins.^{35,36}

Although a Pt(IV) complex was firstly identified together with cisplatin in Rosenberg's original work, its potential as an anticancer drug has been recently recognized.^{26,32,37–39} Pt(IV)-based drugs have a d^6 octahedral geometry, generally believed to be less reactive towards nucleophiles than Pt(II) complexes. The axial ligands of Pt(IV) complexes have an important role in making them relatively inert compared to the Pt(II) complexes. The enhanced stability of Pt(IV) complexes is expected to improve the stability of Pt-based drugs and increase their half-life in the blood stream. In addition, the axial ligands ideally can exhibit functional moieties that can be modified by targeting ligands to achieve active targeting, by other components in order to overcome drug resistance, and by polymers for developing drug-delivery systems.^{32,37–39} The Pt(IV)-based drugs undergo reduction to Pt(II) inside the cells, which eliminates the two axial ligands of the octahedral Pt(IV)-based drug (Fig. 2). The reactive Pt(II) analogs produced by the reduction process exert the anticancer activity.^{26,32,37–39} Because of the reactivity change before and after reduction, Pt(IV)-based drugs are generally considered to be prodrugs.³⁸ However, only a few Pt(IV) complexes such as satraplatin (Fig. 1E) have been involved in significant clinical trials, despite the rational design of Pt(IV) prodrugs.^{37–39} In order to proceed to further clinical practice, it is highly necessary to establish new syn-

thetic chemistry, optimize the kinetics of reduction, and develop well-designed drug delivery systems.^{38,39}

3. Physical encapsulation of Pt(II)

3.1. Traditional techniques

Traditional strategies for the delivery of Pt-based drugs are based on physical encapsulation—an approach that has been widely used in general drug delivery systems. These strategies have the merits of easy and simple procedures for loading drugs into the nanoparticles; however, they have been lamented for their low loading stability which causes the unintended loss of drugs during blood circulation.⁴⁰ Several techniques have been developed for the potential use of encapsulation methods.^{41–44} Desolvation is one of the most frequently used strategies for fabricating nanoparticles. Protein nanoparticles developed by the desolvation method represent ideal candidates for the delivery of anticancer drugs because of their biodegradable, less immunogenic, and non-toxic characteristics.^{41,42} Very recently, a folic acid-conjugated gelatin nanoparticle (Cis-GN-FA) for cisplatin delivery has been developed through a double desolvation method.⁴⁵ In the case of folate-negative A549 lung cancer cells, it was difficult to distinguish Cis-GN-FA from Cis-GNs. In contrast, the Cis-GN-FA nanoparticles show a significant decrease of the half maximal inhibitory concentration (IC_{50}) (8.3 mM) in folate receptor-active HeLa cells, compared to Cis-GNs (15.1 mM) and cisplatin (40.2 mM), implying that further *in vivo* investigations of Cis-GN-FA are needed.

The emulsion technique is another common encapsulation method, which is based on the compatibility between a drug and a matrix.^{41–43} In the case of poorly soluble and lipophilic

drugs, the oil-in-water (o/w) single emulsion method has been utilized for physical encapsulation of the drugs. In addition, double emulsion techniques like water-in-oil-in-water (w/o/w), solid-in-oil-in-water (s/o/w), water-in-oil-in-oil (w/o/o), and solid-in-oil-in-oil (s/o/o) have been employed for compounds with high solubility in water, including proteins, peptides, and drugs like cisplatin.^{41–43} These common techniques have been frequently applied in cisplatin-delivery systems. A study illustrating the development of cisplatin-encapsulated poly(lactic-co-glycolic acid) (PLGA) nanoparticles (NP) by the double emulsion technique has also been reported recently.⁴⁶ The developed PLGA NPs showed a sustained cisplatin release, with a cumulative release of 50% over 3 weeks, highlighting the potential of this system for avoiding multiple administrations of cisplatin in cancer treatment. In another study, cisplatin was emulsified into methoxy poly(ethylene glycol)-*block*-poly(lactic-co-glycolic acid)-*block*-poly(L-lysine) (mPEG-*b*-PLGA-*b*-PLL) nanoparticles (CDDP-NPs) using the w/o/w double emulsion method, followed by conjugation of an epidermal growth factor (EGF) ligand to the CDDP-NPs *via* EDC/NHS chemistry.⁴⁷ The developed CDDP-NP-EGF not only showed sustained drug release profiles *in vitro*, but also showed lower nephrotoxicity and improved anticancer effects on ovarian carcinoma *in vivo*. This simple and universal emulsion technique has been used to develop novel and complex delivery systems as well. In 2014,

Chen *et al.* reported an interesting nanocarrier system which releases both Pt(II)-based drugs and O₂ in response to the local H₂O₂ concentration typical of the tumor environment (Fig. 3).⁴⁸ In this system, Pt(II)-based drugs ([PtLCI]Cl) and catalase were incorporated into the aqueous core of PLGA nanoparticles *via* the double emulsion method. The catalase generated O₂ from intracellular H₂O₂, resulting in the rupture of the nanoparticle shell by the increased internal pressure. In addition, the Pt(II)-based drug and O₂ exhibited anticancer effects and the ability to overcome hypoxia-induced multidrug resistance (MDR),⁴⁹ respectively. Although further investigations are necessary, this novel system could enhance the anticancer efficiency of cisplatin at the *in vitro* level.

Nanoprecipitation, based on the interfacial deposition, is a convenient and commonly used method for the preparation of polymeric nanoparticles.^{43,44} Recently, a biodegradable multifunctional nanoparticle (MNP) system for co-delivery of cisplatin and genes has been developed. Cisplatin-loaded poly(lactic acid) (PLA) nanoparticles were prepared *via* nanoprecipitation, and the surface was then coated with chitosan for electrostatic complexation with genes.⁵⁰ The MNP is thus composed of a PLA inner core loaded with cisplatin and a cationic chitosan (CS) outer layer with P62 siRNA (siP62) and/or β5-expressing plasmid DNA (pβ5) (Fig. 4). The co-delivered genes could knock down P62 and restore β5 expression, resulting in a

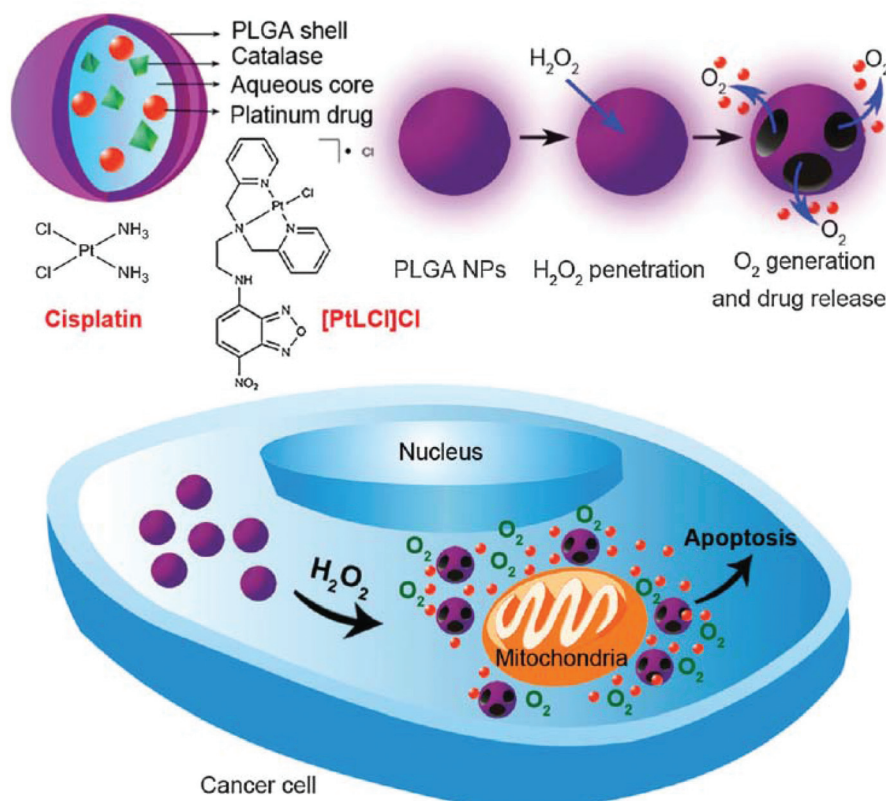


Fig. 3 Schematic illustration of H₂O₂-responsive PLGA nanoparticles containing Pt(II) drugs and O₂-generating catalase and the mechanism of drug release by H₂O₂ (reproduced from ref. 48 with permission from the Royal Society of Chemistry).



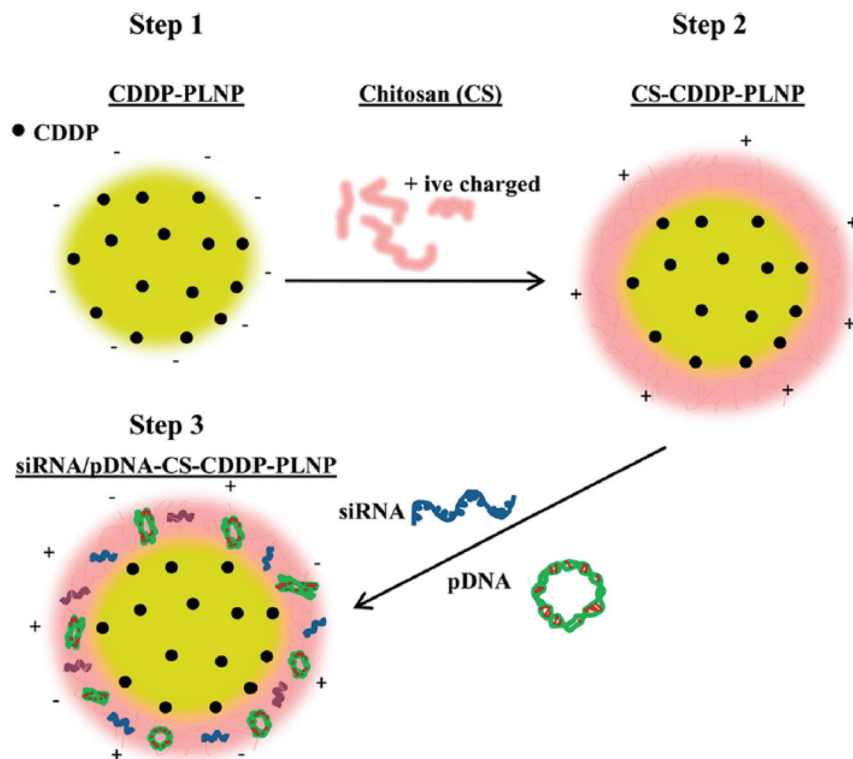


Fig. 4 Schematic diagram of the preparation of siRNA/pDNA-CS-CDDP-PLNP. CDDP-PLNP was prepared by the nanoprecipitation method (Step 1), which was subsequently coated with CS layers (Step 2). The siRNA/pDNA-CS-CDDP-PLNP complex was formed *via* electrostatic interactions between siRNA/pDNA and CS-CDDP-PLNP (Step 3) (reproduced from ref. 50 with permission from the American Chemical Society).

remarkable reduction in the IC_{50} value of cisplatin in cisplatin-resistant ovarian cancer cells. This simple co-delivery system has thus shown a potential ability to overcome the problems arising in the treatment of cisplatin-resistant tumors. Huang *et al.* have recently reported another co-delivery system capable of changing the tumor microenvironment for enhanced anticancer effects.⁵¹ To improve the encapsulation efficiency, cisplatin was coated with dioleoyl phosphatidic acid (DOPA). The DOPA-coated cisplatin cores facilitated the co-encapsulation of cisplatin and rapamycin, an mTOR inhibitor with antiangiogenic properties, into the PLGA *via* nanoprecipitation. The developed PLGA nanoparticles not only suppressed the tumor growth, but also altered the microenvironment of the tumor, inhibiting angiogenesis, reducing tumor-associated fibroblasts and collagen expression, and improving EPR effects of the nanoparticles in tumors. These findings have raised the expectations in the development of novel theranostic systems based on multifunctional nanoparticles such as quantum dots, superparamagnetic iron oxide nanoparticles, gold nanoparticles, and so on.

3.2. Micelles

Polymeric micelles are nanoscopic core-shell structures formed by amphiphilic block copolymers. They are generated when the hydrophobic portions of the polymers are driven to the interior of the structure, while the hydrophilic portions are

exposed to the aqueous environment. These micelles have been widely used for general drug-delivery systems, as the hydrophobic interior has the capacity to hold drugs that are poorly soluble in aqueous solution.^{12,20,52,53} However, as far as cisplatin delivery is concerned, the hydrophilic nature of cisplatin makes it difficult to obtain cisplatin-encapsulated micelles. The successful encapsulation of a hydrophobic Pt(II) compound, *cis*-(cha)₂Pt(NO₃)₂, into the amphiphilic cyclotriphosphazene [NP(MPEG750) (GlyPheLeu)₂Et]₃ (CP750) to form the micelle was recently reported.⁵⁴ Compared to free *cis*-(cha)₂Pt(NO₃)₂, the micelle not only exhibited prolonged blood circulation and large systemic exposure (AUC), but also showed excellent tumor accumulation and low acute toxicity. In particular, the micelle showed significant anticancer effects on the SNU638 cell line, a drug-resistant stomach carcinoma cell.

4. Physical encapsulation of Pt(IV)

The fundamental principles for physical encapsulation of a Pt(IV) prodrug in a nanoparticle are in line with those of Pt(II)-based drugs. The primary difference is that a Pt(IV) prodrug has two more ligands at its axial positions, and these additional ligands can be useful to encapsulate the Pt(IV)-based drug into the nanoparticle. For instance, a Pt(IV)-based



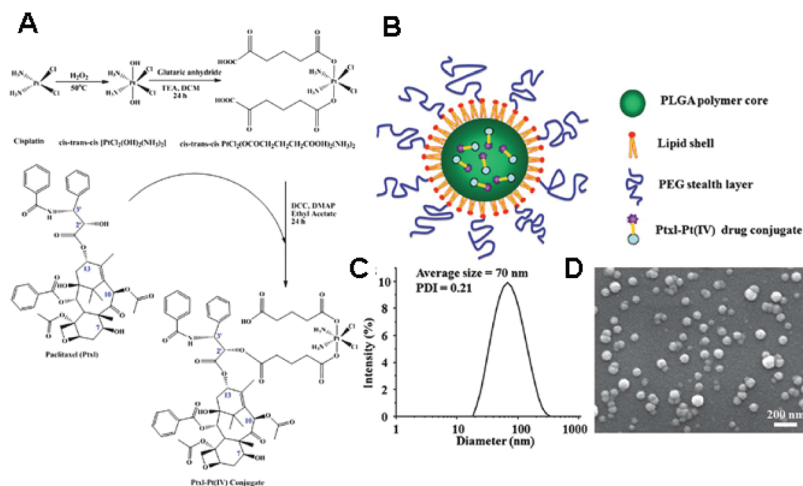


Fig. 5 (A) Synthesis of a drug conjugate composed of Ptxl and Pt(IV) prodrugs. (B) Schematic illustration of a Ptxl–Pt(IV) conjugate loaded polymeric nanoparticles. (C) Dynamic light scattering (DLS) analysis and (D) scanning electron microscopy (SEM) images of the nanoparticles containing a Ptxl–Pt(IV) conjugate (reproduced from ref. 59 with permission from the Royal Society of Chemistry).

drug can be easily encapsulated into the hydrophobic parts of a nanoparticle if its axial ligands are modified by hydrophobic groups.⁵⁵ Recently, Farokhzad and Lippard's group reported the development of targeted-delivery systems with reduced side effects by using simple nanoprecipitation and hydrophobic Pt(IV) prodrugs.^{56,57} Hydrophobic aliphatic chains at the axial position of Pt(IV) drugs helped in the efficient encapsulation of the drug into the hydrophobic core of methoxy poly(ethylene glycol)-*block*-poly(lactic-co-glycolic acid) (PEG-*b*-PLGA) nanoparticles *via* nanoprecipitation.^{55–57} In addition, the inertness of Pt(IV) is expected to mitigate side effects, like nephrotoxicity, compared to Pt(II)-based drugs, because the protection of the axial position of the drug can reduce the reactivity of Pt towards biomolecules. In particular, the conjugation of the cyclic arginine-glycine-aspartate (cRGD) ligand or the prostate-specific membrane antigen (PSMA) aptamer onto the nanocarriers facilitated the active targeting of drugs to the corresponding receptor on a tumor. In fact, cRGD-tethered and hydrophobic Pt(IV) drug-encapsulated mPEG-*b*-PLGA nanoparticles exhibited favorable anticancer effects on $\alpha_v\beta_3$ integrin-overexpressed cancer cell lines *in vitro*.⁵⁶ Furthermore, PSMA-tethered and hydrophobic Pt(IV) drug-encapsulated mPEG-*b*-PLGA nanoparticles showed significant anticancer effects on PSMA-overexpressed LNCaP cells in xenograft mouse models, as well as a remarkable pharmacokinetic index without nephrotoxicity and changes in body weight.⁵⁷ These basic principles have been extended to the development of several co-delivery systems. In 2013, Farokhzad and Walker's group modified the procedures for preparing mPEG-*b*-PLGA nanoparticles containing hydrophobic Pt(IV) prodrugs, in order to simultaneously load siRNA.⁵⁸ By adding a cationic lipid-like molecule (G0-C14) in the double emulsion process, they were able to obtain a nanoparticle whose structure involves an aqueous core, a cationic and hydrophobic layer formed by G0-C14 and PLGA, and a hydrophilic outer shell

composed of PEG. The G0-C14 has a pivotal role in the fabrication of the nanoparticle, because it provides flexible hydrophobic tails for self-assembly into the PLGA matrix, as well as efficient binding with the negatively-charged siRNA. These formulations enable the fabrication of nanoparticles with a dense structure, resulting in a sustained release of siRNA and Pt(IV) drug payloads. In addition, the nanoparticles exhibit a significant inhibition of tumor growth *in vivo* owing to the synergy of antitumor effects by Pt(IV) prodrugs and sensitization by siREV1/siREV3L. Moreover, hydrophobicity can be introduced by conjugating hydrophobic anticancer drugs at the axial position of a Pt(IV) prodrug. Zhang's group reported the synthesis of lipid-polymer hybrid nanoparticles for the delivery of a drug conjugate composed of a hydrophilic Pt(IV) prodrug and the hydrophobic paclitaxel (Ptxl) (Fig. 5).⁵⁹ The drug conjugate can be loaded into the lipid-polymer hybrid nanoparticles *via* nanoprecipitation. The loading of a drug conjugate system within a polymeric nanocarrier is expected not only to provide a route for improving loading of Pt-based drugs, but also to prevent limitations in anticancer effects arising from the variation of pharmacokinetics and biodistribution of different drugs in cocktail chemotherapy strategies.

5. Conjugation of Pt(II)

5.1. Pt(II)-conjugated micelles *via* labile leaving ligands

Pt(II)-based drugs typically consist of two permanently bound ligands, usually amines, and two labile leaving ligands like chlorides or carboxylate functional groups. The conjugation of Pt(II)-based drugs to a polymeric carrier has been accomplished either *via* leaving ligands or *via* permanent ligands. Conjugation *via* leaving ligands is widely used, as inspired by carboplatin which carries carboxylate as a leaving ligand. It is relatively straightforward to obtain polymer-Pt(II) conjugates

structurally similar to carboplatin by conjugating a Pt(II)-based drug to a polymer having pendant carboxyl functionalities.^{10,37} In addition, monofunctional conjugation, bifunctional conjugation, and/or crosslinking of polymer chains can be obtained upon coordinating a Pt(II)-based drug to a polymer containing carboxylate groups.^{60,61}

There have been several notable reports on the development and evaluation of micelles incorporating Pt(II)-based drugs *via* coordination between carboxylate functional groups of a block copolymer and leaving ligands of the drugs.^{62–77} For example, Sengupta *et al.* developed self-assembled micelles by complexation *via* mono- and di-carboxylate linkages between poly(ethylene glycol)-poly(isobutylene-*alt*-maleic anhydride) (PEG-PIMA) and diaquated cisplatin. The nano-sized particles exhibited superior antitumor efficiency compared with free cisplatin, owing to the EPR effect.⁶⁵ Moreover, tumor-targeted micelles for the delivery of Pt(II)-based drugs against specific receptor-positive cell lines have been frequently reported.^{66–68} For instance, Zhang and Yu's group developed micelles fabricated from folate-conjugated poly(ethylene glycol)-*graft*- α,β -poly [*N*-amino acidyl]-aspartamide] (FA-PEG-*g*-PAAsp) and Pt(II)-based drugs, and demonstrated their favorable antitumor effects.⁶⁸ In addition, as a representative example of co-delivery, we discuss the polymeric co-delivery system inspired by the GEMOX chemotherapy regimen, *i.e.*, co-administration of gemcitabine and oxaliplatin.⁶⁹ Gemcitabine is a nucleoside derivative of deoxycytidine and is involved in the repair and synthesis of DNA. It has been demonstrated that gemcitabine is effective on non-small cell lung cancer, pancreatic cancer, bladder cancer, colon, and breast cancer.^{69,70} However, the low stability of gemcitabine *in vivo* requires the administration of high doses, which causes adverse side effects. To overcome these problems, Jing's group developed micelles containing oxaliplatin-polymer (P(Pt)) and gemcitabine-polymer (P(Gem)) conjugates.⁶⁹ Due to the identical polymer backbones, poly(ethylene glycol)-*block*-poly(L-lactide-*co*-2-methyl-2-carboxyl-propylene carbonate) (mPEG-*b*-P(LA-*co*-MCC)), P(Pt) and P(Gem) could co-assemble to form hybrid micelles with controlled gemcitabine/oxaliplatin ratios. The co-assembled micelle exhibited enhanced drug accumulation in the tumor site, resulting in the efficient inhibition of tumor growth compared to single-drug administration and co-administration.

Traditionally, poly(aspartic acid), poly(glutamic acid), and poly(methacrylic acid) were employed for generating polymer-Pt(II) complexes. However, these common polymers coordinate to the Pt(II)-based drugs in a non-specific geometry, resulting in uncontrolled crosslinking and mixed configurations such as monodentate or bidentate coordination.^{71,72} Therefore, Stenzel's group aimed to develop a structurally defined Pt(II)-based drug carrier by investigating the effects of polymer architectures or by using neighboring carboxylate functional groups.^{71,72} Their analysis confirmed that polymers that form 7-membered rings with Pt enhance the solubility of polymer-Pt(II) conjugates, while those that form 11-membered rings led to low solubility due to inter-polymer crosslinking.⁷¹ In addition, it was revealed that statistical copolymers showed

complete drug release, in contrast to block copolymers that released around 40% of a Pt-based drug.⁷¹ However, it was found that the cytotoxicity was not closely related to the drug release, but was related to the cellular uptake.⁷¹ Stenzel's group also tried to develop polymeric micelles with defined geometry by using bidentate carboxylate ligands.⁷² They demonstrated that the formation of 6-membered rings with Pt provided stability to the complex, and that drug release was increasingly more effective as the hydrophobic block with cisplatin became shorter.⁷²

A major disadvantage of micelles is their instability during blood circulation, due to the unintended contact with blood components and the dilution to below critical micellar concentration (CMC). Covalent crosslinking has attracted significant interest as one of the options for improving the stability of micelles.^{73,74} This methodology has also been applied to the delivery of Pt(II)-based drugs using micelles.^{71,75–77} For example, Stenzel's group used 2,2'-(ethylenedioxy)bis(ethylamine) as a crosslinker of micelles, not only to stabilize the micelles, but also to improve the cytotoxic effects of cisplatin by inducing efficient cellular uptake.⁷¹ However, permanently stable crosslinkers have a potential risk to limit the drug release and interrupt the renal clearance of the polymeric carriers.^{76,77} Therefore, Huynh *et al.* utilized an acid-degradable acetal crosslinker to prepare cisplatin-conjugated micelles composed of poly(oligo(ethylene glycol) methylether methacrylate)-*block*-poly(*N*-hydroxysuccinic methacrylate)-*block*-poly(1,1-di-*tert*-butyl 3-(2-(methacryloyloxy)ethyl) butane-1,1,3-tricarboxylate) (POEGMEMA-*b*-PNHSMA-*b*-PMAETC).⁷⁶ POEGMEMA occupies the hydrophilic shell, and the PMAETC block is responsible for the complex formation with cisplatin. The hydroxyl succinimide groups in the PNHSMA block react with the diamine of the acetal crosslinker to produce the cross-linked micelle. Qiao's group employed the same strategy to develop pH-sensitive micelles for cisplatin delivery (Fig. 6).⁷⁷ The linear-brush diblock copolymer contains anhydride groups and grafted PEG, which self-assemble into the micelle. Subsequently, the micelle was crosslinked by the reaction between anhydride groups of diblock copolymers and diamine of the acetal crosslinkers. These two types of pH-sensitive micelles showed a faster drug release under acidic conditions and a higher cellular uptake compared to free drugs and uncross-linked micelles, thus resulting in enhanced anticancer effects.

5.2. Metallosomes

Besides the regular spherical micelles, amphiphilic block copolymers can self-assemble into diverse ordered structures, such as worm-like micelles, multilayer structures, and polymersomes. It has been reported that the structure of amphiphilic block copolymers in aqueous solutions is determined by the ratio of hydrophilic block to total polymer mass. The polymersome can be defined as a polymer-based liposome formed from synthetic polymers, rather than lipids or natural polymers, *via* supramolecular assembly.^{78–84} Polymersomes are typically hollow spheres consisting of an aqueous core and a polymeric bilayer shell. The shells are composed of three



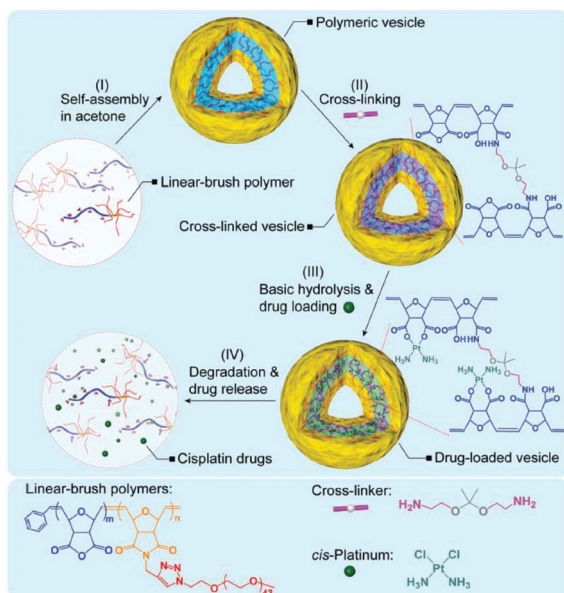


Fig. 6 Schematic diagram of the preparation of a Pt(II)-loaded polymeric vesicle with an acid-degradable crosslinker (reproduced from ref. 77 with permission from the Royal Society of Chemistry).

parts: an inner hydrophilic region, a middle hydrophobic part, and an outer hydrophilic part. The polymersomes have attracted considerable attention in the field of drug-delivery systems owing to their superior properties. First, they can encapsulate hydrophilic agents in their aqueous core and integrate hydrophobic drugs within the hydrophobic regions of the shells. In addition, the dense structure of the hollow shell not only provides structural stability to the polymersomes, but also reduces the permeability of the inner components. This reduced permeability can enhance the drug-loading efficiency and reduce the leakage of the drugs during blood circulation. Furthermore, the diverse synthetic routes for block copolymers provide an easy way to tune the properties of the polymersomes. In 2012, Kataoka's group reported a polymersome with a remarkable structure for the delivery of Pt(II)-based drugs (Fig. 7).⁸⁵ In contrast with general polymersomes formed by

hydrophobic or electrostatic interactions, they developed metallosomes by exploiting the coordination of Pt(II)-based drugs. The complex between (1,2-diaminocyclohexane)platinum(II) (DACHPt) (Fig. 1F) and the carboxylic functional groups of Y-shaped block copolymer (PEGasus-*b*-PLGA-Chole) composed of ω -cholesteroyl-poly(L-glutamic acid) and two-armed poly(ethylene glycol) triggered the metallosome formation. The metallosome showed sustained release of active Pt(II)-based drugs and effective suppression of tumor growth. In addition, by showing the feasibility of loading water-soluble fluorescent molecules into the metallosomes, this strategy shows potential applications to the co-delivery of other therapeutic agents.

5.3. Micelles crosslinked by Pt(II)-based drugs

The ability of Pt(II)-based drugs to coordinate to two carboxylic groups provides a route to crosslink nanoparticles composed of polymers with carboxylic functional groups.^{62–64,66,67,86–89} That is, conjugation *via* leaving ligands of Pt(II)-based drugs can perform an important role in stabilizing the nanoparticles, as well as in conjugating the drug to the carrier.^{60–64,66,67,86–89} Song *et al.* developed a polymeric micelle using the biocompatible triblock copolymer, methoxy poly(ethylene glycol)-*block*-poly(L-glutamic acid)-*block*-poly(L-phenylalanine) (mPEG-*b*-P(Glu)-*b*-P(Phe)), for the co-delivery of Ptxl and cisplatin.⁸⁷ The triblock copolymer was rationally designed: the hydrophobic P(Phe) block is responsible for the Ptxl encapsulation, the P(Glu) segment performs the conjugation of cisplatin and the crosslinking of the micelles, and mPEG in the outer corona can prolong the blood circulation time. It was demonstrated that the crosslinks created *via* coordination of cisplatin contributed to alleviate the initial burst release of Ptxl from the micelles. In addition, the micelles showed an enhanced anti-tumor efficacy with reduced side effects *in vivo*. Kim *et al.* reported an interesting strategy to form micelles containing cisplatin and photodynamic agents.⁸⁸ They employed cisplatin as a crosslinker among components containing carboxylic functional groups, such as phthalocyanine (DPC) and poly(ethylene glycol)-*block*-poly(L-aspartic acid) (PEG-*b*-PLAN). The developed polymer-metal complex micelles (PMCM) formed

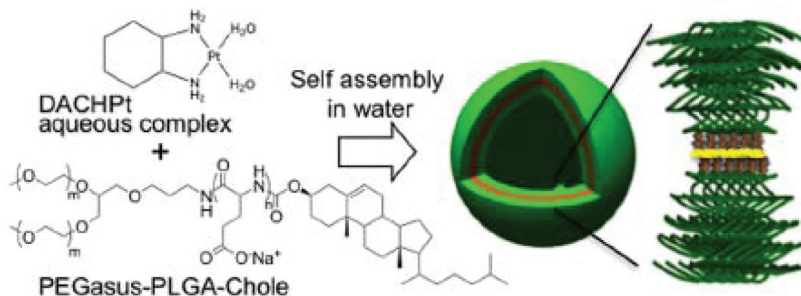
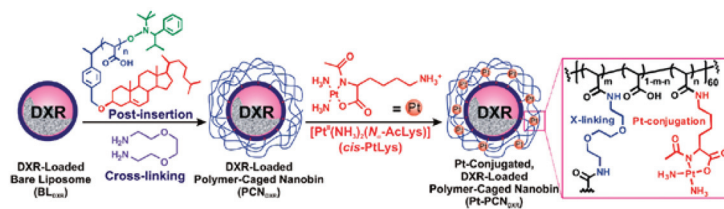


Fig. 7 Schematic diagram of the formation of metallosomes *via* self-assembly of the polymer-metal complex. A Pt in the DACHPt was located between two polymer chains by coordinating with carboxylates in the poly(L-glutamic acid) (PLGA) segment (reproduced from ref. 85 with permission from the American Chemical Society).



A



B

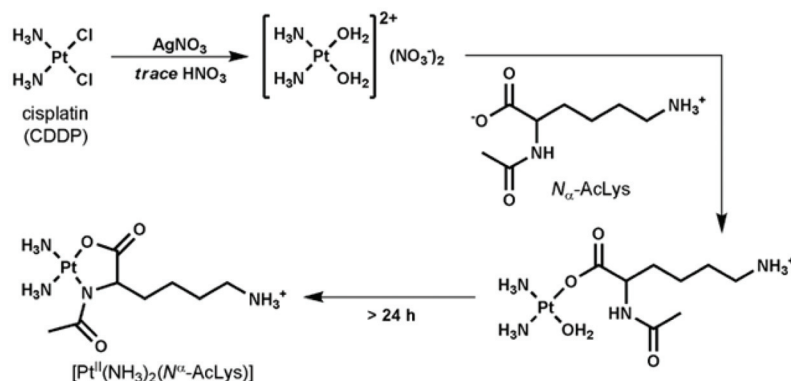


Fig. 8 (A) Synthesis of Pt(II)-conjugated and doxorubicin (DXR)-loaded Polymer-Caged Nanobins (PCNs). The DXR-loaded liposomes were coated with polymers which could act as a reservoir for Pt(II) drugs. Polymer shells provide structural stability to liposomes and controlled release of DXR and Pt(II) drugs. (B) Synthetic scheme of $[\text{Pt}^{\text{II}}(\text{NH}_3)_2(\text{N}_\alpha\text{-AcLys})]$ (reproduced from ref. 92 with permission from the American Chemical Society).

unimodal nano-sized particles that exhibited high stability in buffer solution. Although the generation of singlet oxygen was confirmed, the synergistic effects of cisplatin and singlet oxygen still need to be investigated. Koseva's group exploited the cross-linking ability of cisplatin to develop a reversible PEGylated strategy.⁸⁹ A residual leaving ligand of PEG-cisplatin was used as a glue between PEG and the carboxylic functional groups in star-type block copolymers. This strategy, capable of simultaneously loading Pt(II)-based drugs, is expected to control the pharmacological behavior of the drugs and the nanocarriers.

5.4. Pt(II)-conjugated nanoparticles *via* permanently bound ligands

Chemical conjugation *via* permanently bound ligands has been occasionally employed to obtain polymer-Pt(II) conjugates because of the difficulty in modification of permanent ligands.^{10,37} In particular, the biodegradability of Pt(II)-conjugated nanoparticles is important in the case of conjugation *via* permanently bound ligands. According to various studies, it has been hypothesized that the stable macromolecular structures of polymer-Pt(II) conjugates formed *via* permanent ligands suppress the chemotherapeutic efficiency of the Pt(II)-based drugs.¹² Therefore, many research studies have employed acid-degradable linkers for developing polymer-Pt(II) conjugates *via* permanently bound ligands.

In the process of developing and exploiting polymer-caged nanobins (PCNs) formed by crosslinking polymers which are

incorporated into bare liposomes, Lee *et al.* evaluated the ability of PCNs to load and deliver two types of cytotoxic agents, doxorubicin and Pt(II)-based drugs (Fig. 8A).^{90–92} The doxorubicin could be loaded into the liposomes by hydrophobic interactions. By conjugating the pH-sensitive C6-amine moiety of a cisplatin derivative ($\text{Pt}^{\text{II}}(\text{NH}_3)_2(\text{N}_\alpha\text{-AcLys})$) (Fig. 8B) to the carboxylic acid in the polymeric shell composed of cholesterol-terminated poly(acrylic acid), the developed PCN also contains a cisplatin derivative in the shell.⁹² This co-delivery system exhibited higher synergistic cytotoxicity than the combination of either the free drugs or the separately nanopackaged drugs. These results clearly demonstrate that the PCN platform can offer new means for building synergy into combination chemotherapy. Binauld *et al.* reported a strategy to introduce acid-degradable linkages between the polymer backbone and the permanently bound amine ligands of Pt(II)-based drugs (Fig. 9).⁹³ Their methodology exploited the high stability of the bonds between Pt and permanently bound ligands to enhance the pH responsiveness. The micelle formed by self-assembly of pH-sensitive polymer-Pt(II) conjugates showed an acid-accelerated drug release behavior.

6. Conjugation of Pt(IV)

6.1. Traditional strategies with Pt(IV)-polymer conjugates

The axial ligands of Pt(IV) complexes provide the drugs an opportunity to be encapsulated efficiently into the nanoparti-



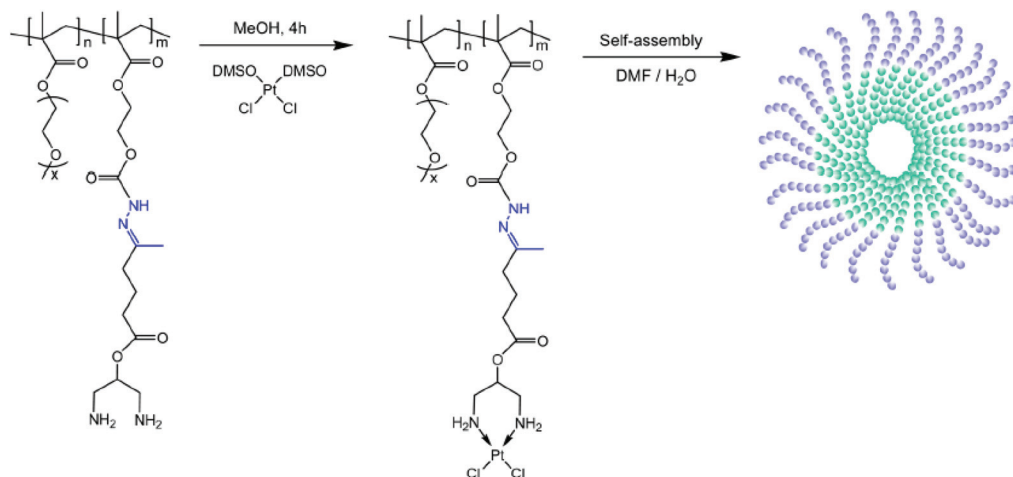


Fig. 9 Preparation of the hydrazone copolymer–Pt(II) complex and illustration of micelles through self-assembly of polymer–drug conjugates. The hydrazone bond was cleavable under acidic conditions, resulting in the release of Pt(II) (reproduced from ref. 93 with permission from the American Chemical Society).

cles *via* traditional and simple methods.^{32,37–39,55} In addition, combination chemotherapy has emerged as a possible solution for drug resistance, due to its ability to interrupt one or more cellular mechanisms related to apoptosis and survival.^{94,95} In particular, a prodrug comprised of Pt(IV)-based drugs and polymers can contribute to effectively encapsulate other anticancer agents, as well as Pt(IV)-based drugs, into the nanoparticles developed by nanoprecipitation. Mi *et al.* developed a strategy for targeted co-delivery of a Pt(IV) prodrug and docetaxel.⁹⁴ D- α -Tocopheryl-co-poly(ethylene glycol) 1000 succinate (TPGS), which has a water-soluble amphiphilic structure, was employed to incorporate a Pt(IV) prodrug. TPGS-COOH, the TPGS-Pt(IV) prodrug, and PLA-TPGS were nanoprecipitated with docetaxel to form the nanoparticles (HTCP-NP) for combination chemotherapy. Herceptin, which targets the HUR-2 receptor overexpressed in breast cancer, could react with carboxylic groups exposed on the HTCP-NP surface. This approach can be applied to the targeted co-delivery of various hydrophilic and hydrophobic drugs. Kolishetti *et al.* illustrated a platform technology for combination chemotherapy involving a Pt(IV)-polymer conjugate and the microfluidic nanoprecipitation method.⁹⁵ The Pt(IV)-based drugs were tethered to the pendant hydroxyl group of a hydrophobic poly(lactide) derivative, followed by the formation of nanoparticles with docetaxel and the PEG-*b*-PLGA block copolymer through the microfluidic nanoprecipitation method. This platform technology provides excellent drug encapsulation efficiency and reproducibility. In addition, the introduction of the A10 aptamer supplemented the nanoparticles with the ability to target PSMA-overexpressed prostate cancer cells. PSMA-targeted nanoparticles encapsulating a Pt(IV) prodrug and docetaxel showed enhanced cytotoxicity compared to nanoparticles containing the individual single drugs. This approach showed significant potential for providing conventional methods

capable of developing uniform nanoparticles for a targeted co-delivery system incorporating Pt-based drugs.

6.2. Pt(IV)-conjugated micelles

In addition to providing hydrophobicity to Pt-based drugs, the axial ligands of Pt(IV) complexes can enable the conjugation of the drug directly with the nanoparticles,^{32,37–39} which would allow the successful and efficient incorporation of Pt-based drugs into the micelles. Stenzel's group demonstrated the ability of a polymeric micelle containing oxoplatin and folate ligands to deliver the Pt(IV)-based drug into folate receptor-overexpressed ovarian cancer cells (Fig. 10).⁹⁶ In this system, oxoplatin was successfully loaded into the micelles *via* an ester linkage between the carboxyl group of hydrophobic segments of the amphiphilic block copolymer and the hydroxyl group of oxoplatin. The micelle was designed to release cisplatin by hydrolysis of the ester linkage and subsequent reduction of oxoplatin in the cytosol. Folate ligands were introduced to the ends of hydrophilic segments through the interaction between phenylboronic acid and dopamine. This simple chemistry enables easy control of the amount of folate ligands on the micelle, which affects the IC₅₀ value of oxoplatin. In addition, the micellar core was crosslinked to provide structural stability to the micelle, allowing the drugs to enter the cell efficiently.

Although it has been reported that Pt(IV)-based drugs are activated by the cellular environments, the drug efficacy can be limited due to the dependency of the drug release on indirect processes such as acid hydrolysis and/or reduction.^{97–100} Following a high demand for systems capable of releasing Pt(II) complexes directly, Sadler and co-workers reported a photosensitive Pt(IV)-diazido complex which is converted to Pt(II) species under mild UV irradiation.^{97,98} The UV irradiation breaks the Pt–N₃ bond, resulting in the release of the Pt(II) complex.^{97,98} Based on the photosensitive characteristics of



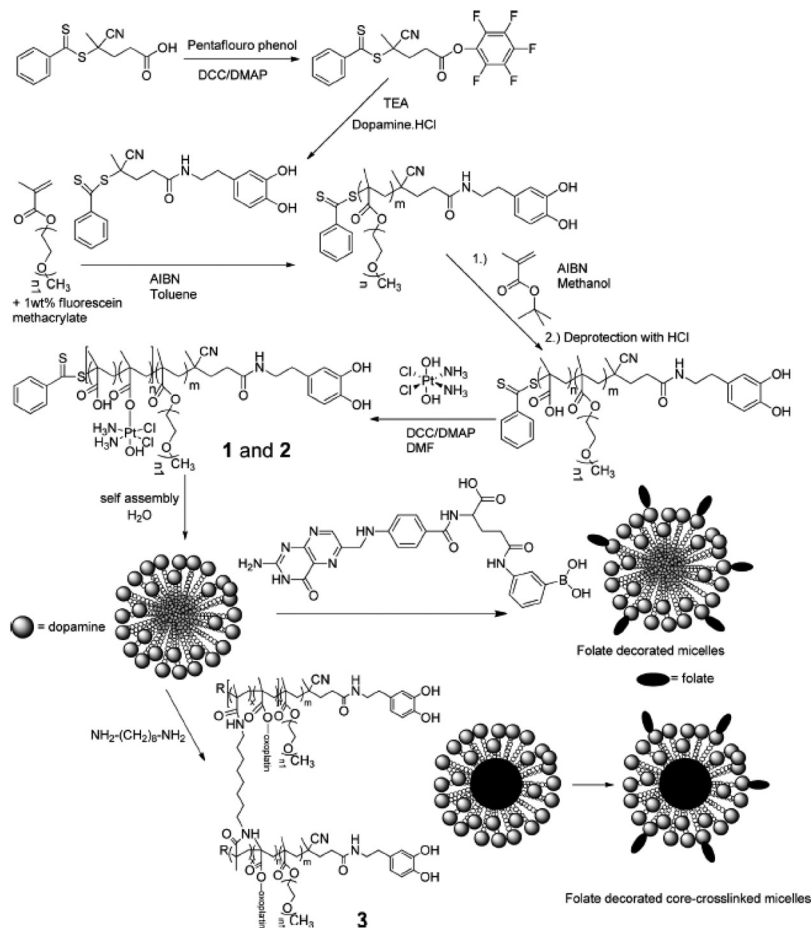


Fig. 10 Synthesis of Pt(IV) prodrug attached poly(oligo(ethylene glycol) methyl ether methacrylate)-*b*-poly(methacrylic acid) block copolymers (p(OMGMEMA)-*b*-p(MMA)) with a pendant dopamine group. After self-assembling, the folate ligands were introduced on the surface of micelles via dopamine–phenylboronic ester interaction and the cores of micelles were crosslinked for structural stability (reproduced from ref. 96 with permission from the American Chemical Society).

the Pt(IV)–diazido complex, polymeric micelles which showed light-responsive release of active Pt(II)-based drugs have been developed by conjugating the Pt(IV)–diazido complex to polymers *via* an ester or amide linkage.^{99,100} The micelles showed rapid release of Pt(II) species under UV irradiation, whereas they were stable under dark conditions. These micelles also exhibited enhanced cytotoxicity under UV irradiation, as well as inhibition of tumor growth *in vivo*, with reduced side effects.

Li *et al.* reported multi-stage Pt(IV) prodrug-delivery systems mimicking the common polymeric gene-delivery systems.¹⁰¹ The mono-carboxylated Pt(IV) complexes were conjugated with cationic dendrimers, and the as-synthesized cationic prodrugs formed polyplex micelles with the anionic diblock copolymer. According to the design, the cationic Pt(IV)–dendrimer conjugates are dissociated from the polyplex micelles due to the charge conversion of anionic segments of the diblock copolymer at tumoral acidic pH (~6.0). Subsequently, cisplatin can be released from Pt(IV)–dendrimer conjugates by the reductive environment in the cytosol. The polyplex micelles showed an ability for deep penetration in tumor tissue and enhanced anticancer effects compared to cisplatin.

6.3. Pt(IV)-conjugated micelles for co-delivery

Approaches involving drug-conjugated micelles have attracted considerable attention for the development of co-delivery systems. Unlike the physical encapsulation strategies, conjugated micelle systems provide Pt(IV)-based drugs and other combinatorial drugs with structural stability and ability of controlled release. Xiao *et al.* investigated micelles for co-delivery of daunomycin (DRB) and oxaliplatin.¹⁰² The DRB and the prodrug of oxaliplatin were attached to a biodegradable amphiphilic polymer, and the resulting two types of drug–polymer conjugates were co-assembled into the micelle. The different linkages between each drug and the polymer have a different effect on the drug release. The polymer–Pt(IV) conjugate exhibited rapid release of the Pt(II) complex obtained by reduction, while the polymer–DRB conjugate showed sustained release due to acid hydrolysis. In addition, the micelles showed reduced systemic toxicity and synergistic effects both *in vivo* and *in vitro*.

The two axial ligands of Pt(IV)-based drugs can be used for incorporating additional functionalities. For example, if one



ligand contains agents for combination therapy and the other is conjugated with the nanocarriers, a co-delivery system with synergistic effects, enhanced stability, and passive targeting ability can be developed. Inspired by mitaplatin (Fig. 1G), which contains two dichloroacetate (DCA) ligands in the axial positions of cisplatin, Xiao *et al.* developed micelles containing a Pt(IV) prodrug with a DCA ligand and a carboxylic group in its axial positions.¹⁰³ DCA is a mitochondria-targeting small molecule which inhibits glycolysis, enhancing the mitochondrial apoptosis in cancer and suppressing tumor growth. The carboxylic group at the axial positions of a DCA-Pt(IV) conjugate was linked to an amphiphilic biodegradable block copolymer (methoxy poly(ethylene glycol)-*block*-poly(caprolactone)-*block*-poly(L-lysine), mPEG-*b*-PCL-*b*-PLL) for the formation of micelles. The enhanced anticancer effects of this polymeric delivery system might be attributed to the individual pharmacological mechanism of each anticancer drug. Qi *et al.* described a similar Pt(IV) prodrug delivery system.¹⁰⁴ Camphor anhydride is a derivative of camphor that can down-regulate the intracellular levels of Bcl-2 expression, enabling to overcome the resistance to cisplatin. Conjugation of camphor anhydride with one of the axial positions of Pt(IV) prodrugs generated camplatin (Fig. 1H) and the other axial position was further linked to biocompatible mPEG-*b*-PCL-*b*-PLL. The as-prepared polymer-camplatin conjugate could self-assemble into micelles. The micelles exhibited enhanced cytotoxicity on both cisplatin-sensitive and cisplatin-resistant cell lines compared to free cisplatin, because of the synergistic effects of the cytotoxic Pt-based drug and the camphoric acid chemosensitizer.

6.4. Micelles crosslinked by Pt(IV) prodrugs

In addition to providing functional moieties that can be modified with other therapeutic agents, the two axial ligands can act as crosslinkers among polymers, leading to the formation and stabilization of the micelle structures. By taking advantage of this characteristic, Aryal *et al.* reported a pH-responsive delivery system, with the Pt(IV) prodrug employed as both an anticancer agent and a crosslinker between diblock copolymers.¹⁰⁵ In this system, the Bi(PEG-PLA)-Pt(IV) conjugate was obtained by crosslinking two PEG-*b*-PLA block copolymers by one Pt(IV) prodrug *via* pH-responsive hydrazone bonds (Fig. 11). The amphiphilic Bi(PEG-PLA)-Pt(IV) could self-assemble into micelles. This pH-responsive micelle showed a rapid release of cisplatin in acidic pH, which contributed to the increased cytotoxicity compared to free cisplatin. The strategy involving the use of a Pt(IV) prodrug as a crosslinker was extended to develop shell- or core-crosslinked micelles for stabilizing the micelle structure and preventing the premature release of the drug. Song *et al.* reported shell-crosslinked mPEG-*b*-PCL-*b*-PLL micelles based on a dicarboxyl Pt(IV) prodrug.¹⁰⁶ Two axial succinic moieties of the Pt(IV) prodrug could react *via* EDC/NHS chemistry with free amine groups in the PLL segment occupying the hydrophilic shell of the micelles. Compared with free cisplatin and prodrugs, shell-crosslinked micelles not only showed a sustained release of

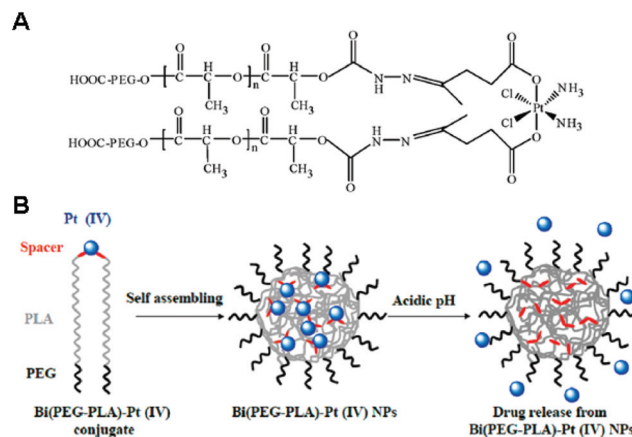


Fig. 11 (A) Structure of the Bi(PEG-PLA)-Pt(IV) polymer-prodrug conjugate. (B) Schematic illustration of the preparation and operation of the pH-responsive Pt(IV) delivery system. Bi(PEG-PLA)-Pt(IV) NPs were formed through self-assembly of polymer-drug conjugates. Under acidic conditions, the hydrazone bonds were hydrolyzed, resulting in the rapid release of cisplatin (reproduced from ref. 105 with permission from the American Chemical Society).

cisplatin with enhanced cytotoxicity *in vitro*, but also exhibited selective accumulation into tumor sites *in vivo*.

7. Conclusions

Since the successful clinical transfer of cisplatin and oxaliplatin, various Pt-based drugs have been developed and they have shown significant antitumor effects *in vitro*. Nevertheless, most free Pt-based drugs have failed to enter clinical use due to the severe side effects and lower activity than cisplatin. Polymeric nanoparticles have emerged as one of the alternative strategies for overcoming the disadvantages of bare Pt-based drugs. For example, the NC-6004 micelle comprised of cisplatin-conjugated PEG-*b*-poly(glutamic acid) (PEG-*b*-P(Glu)) was developed by Kataoka's group and has shown reduced nephrotoxicity and improved antitumor efficacy in pancreatic cancer. This is currently in a Phase III clinical trial.^{10–12,107} Inspired by the successful development of NC-6004, NC-4016 was also developed by conjugating DACHPt with the PEG-*b*-P(Glu). NC-4016 has also shown potential therapeutic efficacy in various tumor models and is currently in a Phase I clinical trial.^{10–12,108,109} ProlindacTM (AP5346) is one of the polymer-Pt(II) conjugates, which contains DACHPt conjugated with *N*-(2-hydroxypropyl)methacrylamide (HPMA) *via* a pH-sensitive coordinating group. This is currently in Phase II clinical studies for advanced ovarian cancer.^{10,12,110} In addition to the polymeric delivery systems, various liposomal nanoparticles containing Pt-based drugs have been reported for the clinical trials.^{10,111–113} The increasing number of Pt-based drug delivery systems under clinical evaluation have motivated many researchers to develop a more advanced strategy for effective anticancer therapy.



In this review, we have summarized and discussed the progress attained in the development of polymeric nanoparticles for effective delivery of Pt-based drugs, focusing on drug configurations and incorporation methods. Whereas micelles have been the most employed systems in the past five years, standard techniques such as desolvation, emulsion, and nanoprecipitation have also been frequently used for the delivery of Pt-based drugs. Crosslinking strategies and integration of targeting ligands have also been employed to improve the loading efficiency and the tumor accumulation of drugs. Co-delivery of other therapeutic agents or chemosensitizers has provided a method for overcoming the resistance to cisplatin. In particular, unique polymeric nanoparticles like metallosomes and PCN have been developed as potential routes to improve anticancer effects and mitigate side effects. Most strategies described in this review offer rational alternatives to existing conventional methods. These efforts are expected to pave the way for the practical advancement and clinical use of Pt-based anticancer therapy.

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Notes and references

- 1 A. S. Abu-Surrah and M. Kettunen, *Curr. Med. Chem.*, 2006, **13**, 1337–1357.
- 2 X. Wang and Z. Guo, *Dalton Trans.*, 2008, 1521–1532.
- 3 L. Kelland, *Nat. Rev. Cancer*, 2007, **7**, 573–584.
- 4 M. J. Hannon, *Pure Appl. Chem.*, 2007, **79**, 2243–2261.
- 5 S. P. Fricker, *Dalton Trans.*, 2007, 4903–4917.
- 6 C. A. Rabik and M. E. Dolan, *Cancer Treat. Rev.*, 2007, **33**, 9–23.
- 7 A. A. Argyriou, P. Polychronopoulos, G. Iconomou, E. Chroni and H. P. Kalofonos, *Cancer Treat. Rev.*, 2008, **34**, 368–377.
- 8 S. R. McWhinney, R. M. Goldberg and H. L. McLeod, *Mol. Cancer Ther.*, 2009, **8**, 10–16.
- 9 X. Yao, K. Panichpisal, N. Kurtzman and K. Nugent, *Am. J. Med. Sci.*, 2007, **334**, 115–124.
- 10 H. S. Oberoi, N. V. Nukolova, A. V. Kabanov and T. K. Bronich, *Adv. Drug Delivery Rev.*, 2013, **65**, 1667–1685.
- 11 H. Cabral and K. Kataoka, *J. Controlled Release*, 2014, **190**, 465–476.
- 12 M. Callari, J. R. Aldrich-Wright, P. L. de Souza and M. H. Stenzel, *Prog. Polym. Sci.*, 2014, **39**, 1614–1643.
- 13 K. Maruyama, *Adv. Drug Delivery Rev.*, 2011, **63**, 161–169.
- 14 A. K. Iyer, G. Khaled, J. Fang and H. Maeda, *Drug Discovery Today*, 2006, **11**, 812–818.
- 15 V. Torchilin, *Adv. Drug Delivery Rev.*, 2011, **63**, 131–135.
- 16 T. Lammers, W. E. Hennink and G. Storm, *Br. J. Cancer*, 2008, **99**, 392–397.
- 17 F. Danhier, O. Feron and V. Preat, *J. Controlled Release*, 2010, **148**, 135–146.
- 18 J. D. Byrne, T. Betancourt and L. Brannon-Peppas, *Adv. Drug Delivery Rev.*, 2008, **60**, 1615–1626.
- 19 F. Marcucci and F. Lefoulon, *Drug Discovery Today*, 2004, **9**, 219–228.
- 20 K. Kataoka, A. Harada and Y. Nagasaki, *Adv. Drug Delivery Rev.*, 2001, **47**, 113–131.
- 21 K. S. Soppimath, T. M. Aminabhavi, A. R. Kulkarni and W. E. Rudzinski, *J. Controlled Release*, 2001, **70**, 1–20.
- 22 T. M. Allen and P. R. Cullis, *Adv. Drug Delivery Rev.*, 2013, **65**, 36–48.
- 23 E. R. Gillies and J. M. J. Frechet, *Drug Discovery Today*, 2005, **10**, 35–43.
- 24 B. Ahn, J. Park, K. Singha, H. Park and W. J. Kim, *J. Mater. Chem. B*, 2013, **1**, 2829–2836.
- 25 T. L. Doane and C. Burda, *Chem. Soc. Rev.*, 2012, **41**, 2885–2911.
- 26 R. A. Alderden, M. D. Hall and T. W. Hambley, *J. Chem. Educ.*, 2006, **83**, 728–734.
- 27 D. Gibson, *Dalton Trans.*, 2009, 10681–10689.
- 28 A. I. Ivanov, J. Christodoulou, J. A. Parkinson, K. J. Barnham, A. Tucker, J. Woodrow and P. J. Sadler, *J. Biol. Chem.*, 1998, **273**, 14721–14730.
- 29 R. C. DeConti, B. R. Toftness, R. C. Lange and W. A. Creasey, *Cancer Res.*, 1973, **33**, 1310–1315.
- 30 D. Wang and S. J. Lippard, *Nat. Rev. Drug Discovery*, 2005, **4**, 307–320.
- 31 M. Ohmichi, J. Hayakawa, K. Tasaka, H. Kurachi and Y. Murata, *Trends Pharmacol. Sci.*, 2005, **26**, 113–116.
- 32 M. D. Hall and T. W. Hambley, *Coord. Chem. Rev.*, 2002, **232**, 49–67.
- 33 D. Fink, S. Nebel, S. Aebi, H. Zheng, B. Cenni, A. Nehme, R. D. Christen and S. B. Howell, *Cancer Res.*, 1996, **56**, 4881–4886.
- 34 E. Raymond, S. Faivre, S. Chaney, J. Woynarowski and E. Cvitkovic, *Mol. Cancer Ther.*, 2002, **1**, 227–235.
- 35 Y.-P. Ho, S. C. F. Au-Yeung and K. K. W. To, *Med. Res. Rev.*, 2003, **23**, 633–655.
- 36 P. Perego, C. Caserini, L. Gatti, N. Carenini, S. Romanelli, R. Supino, D. Colangelo, I. Viano, R. Leone, S. Spinelli, G. Pezzoni, C. Manzotti, N. Farrell and F. Zunino, *Mol. Pharmacol.*, 1999, **55**, 528–534.
- 37 E. Wong and C. M. Giandomenico, *Chem. Rev.*, 1999, **99**, 2451–2466.
- 38 M. D. Hall, H. R. Mellor, R. Callaghan and T. W. Hambley, *J. Med. Chem.*, 2007, **50**, 3403–3411.
- 39 E. Gabano, M. Ravera and D. Osella, *Dalton Trans.*, 2014, **43**, 9813–9820.
- 40 S. R. Croy and G. S. Kwon, *Curr. Pharm. Des.*, 2006, **12**, 4669–4684.



- 41 C. P. Reis, R. J. Neufeld, A. J. Ribeiro and F. Veiga, *Nano-medicine*, 2006, **2**, 8–21.
- 42 S. Sundar, J. Kundu and S. C. Kundu, *Sci. Technol. Adv. Mater.*, 2010, **11**, 014104.
- 43 S. Vrignaud, J.-P. Benoit and P. Saulnier, *Biomaterials*, 2011, **32**, 8593–8604.
- 44 S. Hornig, T. Heinze, C. R. Becer and U. S. Schubert, *J. Mater. Chem.*, 2009, **19**, 3838–3840.
- 45 N. Dixit, K. Vaibhav, R. S. Pandey, U. K. Jain, O. P. Katore, A. Katyal and J. Madan, *Biomed. Pharmacother.*, 2015, **69**, 1–10.
- 46 A. C. Jayasuriya and A. J. Darr, *J. Biomed. Sci. Eng.*, 2013, **6**, 586–592.
- 47 Y. Wang, P. Liu, L. Qui, Y. Sun, M. Zhu, L. Gu, W. Di and Y. Duan, *Biomaterials*, 2013, **34**, 4068–4077.
- 48 H. Chen, W. He and Z. Guo, *Chem. Commun.*, 2014, **50**, 9714–9717.
- 49 A. L. Harris, *Nat. Rev. Cancer*, 2002, **2**, 38–47.
- 50 A. Babu, Q. Wang, R. Muralidharan, M. Shanker, A. Munshi and R. Ramech, *Mol. Pharmaceutics*, 2014, **11**, 2720–2733.
- 51 S. Guo, C. M. Lin, Z. Xu, L. Miao, Y. Wang and L. Huang, *ACS Nano*, 2014, **8**, 4996–5009.
- 52 J. Gong, M. Chen, Y. Zheng, S. Wang and Y. Wang, *J. Controlled Release*, 2012, **159**, 312–323.
- 53 K. Miyata, R. J. Christie and K. Kataoka, *React. Funct. Polym.*, 2011, **71**, 227–234.
- 54 V. B. Jadhav, Y. J. Jun, J. H. Song, M. K. Park, J. H. Oh, S. W. Chae, I.-S. Kim, S.-J. Choi, H. J. Lee and Y. S. Sohn, *J. Controlled Release*, 2010, **147**, 144–150.
- 55 S. Dhar, F. X. Gu, R. Langer, O. C. Farokhzad and S. J. Lippard, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 17356–17361.
- 56 N. Graf, D. R. Bielenberg, N. Kolishetti, C. Muus, J. Banyard, O. C. Farokhzad and S. J. Lippard, *ACS Nano*, 2012, **6**, 4530–4539.
- 57 S. Dhar, N. Kolishetti, S. J. Lippard and O. C. Farokhzad, *Proc. Natl. Acad. Sci. U. S. A.*, 2011, **108**, 1850–1855.
- 58 X. Xu, K. Xie, X.-Q. Zhang, E. M. Pridgen, G. Y. Park, D. S. Cui, J. Shi, J. Wu, P. W. Kantoff, S. J. Lippard, R. Langer, G. C. Walker and O. C. Farokhzad, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 18638–18643.
- 59 S. Aryal, C.-M. Jack Hu, V. Fu and L. Zhang, *J. Mater. Chem.*, 2012, **22**, 994–999.
- 60 N. Nishiyama, S. Okazaki, H. Cabral, M. Miyamoto, Y. Kato, Y. Sugiyama, K. Nishio, Y. Matsumura and K. Kataoka, *Cancer Res.*, 2003, **63**, 8977–8983.
- 61 L. L. Komane, E. H. Mukaya, E. W. Neuse and C. E. J. van Rensburg, *J. Inorg. Organomet. Polym.*, 2008, **18**, 111–123.
- 62 Y. Xiong, W. Jiang, Y. Shen, H. Li, C. Sun, A. Ouahab and J. Tu, *Biomaterials*, 2012, **33**, 7182–7193.
- 63 Z. Ahmad, Z. Tang, A. Shah, S. Lv, D. Zhang, Y. Zhang and X. Che, *Macromol. Biosci.*, 2014, **14**, 1337–1345.
- 64 F. Zhang, M. Elsbahy, S. Zhang, L. Y. Lin, J. Zou and K. L. Wooley, *Nanoscale*, 2013, **5**, 3220–3225.
- 65 A. Paraskar, S. Soni, S. Basu, C. J. Amarasiriwardena, N. Lupoli, S. Srivats, R. S. Roy and S. Sengupta, *Nanotechnology*, 2011, **22**, 265101.
- 66 N. V. Nukolova, H. S. Oberoi, Y. Zhao, V. P. Chekhonin, A. V. Kabanov and T. K. Bronich, *Mol. Pharmaceutics*, 2013, **10**, 3913–3921.
- 67 M. Li, Z. Tang, Y. Zhang, S. Lv, H. Yu, D. Zhang, H. Hong and X. Chen, *J. Mater. Chem. B*, 2014, **2**, 3490–3499.
- 68 Y. Xue, X. Tang, J. Huang, X. Zhang, J. Yu, Y. Zhang and S. Gui, *Colloids Surf., B*, 2011, **85**, 280–288.
- 69 H. Song, H. Xiao, M. Zheng, R. Qi, L. Yan and X. Jing, *J. Mater. Chem. B*, 2014, **2**, 6560–6570.
- 70 C. M. Galmarini, J. R. Mackey and C. Dumontet, *Lancet Oncol.*, 2002, **3**, 415–424.
- 71 V. T. Huynh, G. Chen, P. de Souza and M. H. Stenzel, *Bio-macromolecules*, 2011, **12**, 1738–1751.
- 72 V. T. Huynh, P. de Souza and M. H. Stenzel, *Macromolecules*, 2011, **44**, 7888–7900.
- 73 A. Rosler, G. W. M. Vandermeulen and H. A. Klok, *Adv. Drug Delivery Rev.*, 2012, **64**, 270–279.
- 74 R. K. O'Reilly, C. J. Hawker and K. L. Wooley, *Chem. Soc. Rev.*, 2006, **35**, 1068–1083.
- 75 J. Peng, T. Qi, J. Liao, B. Chu, Q. Yang, W. Li, Y. Qu, F. Luo and Z. Qian, *Biomaterials*, 2013, **34**, 8726–8740.
- 76 V. T. Huynh, S. Binauld, P. L. de Souza and M. H. Stenzel, *Chem. Mater.*, 2012, **24**, 3197–3211.
- 77 Q. Fu, J. Xu, K. Ladewig, T. M. A. Henderson and G. G. Qiao, *Polym. Chem.*, 2015, **6**, 35–43.
- 78 B. M. Discher, Y.-Y. Won, D. S. Ege, J. C.-M. Lee, F. S. Bates, D. E. Discher and D. A. Hammer, *Science*, 1999, **284**, 1143–1146.
- 79 D. E. Discher and F. Ahmed, *Annu. Rev. Biomed. Eng.*, 2006, **8**, 323–341.
- 80 D. E. Discher, V. Ortiz, G. Srinivas, M. L. Klein, Y. Kim, D. Christian, S. Cai, P. Photos and F. Ahmed, *Prog. Polym. Sci.*, 2007, **32**, 838–857.
- 81 D. H. Levine, P. P. Ghoroghchian, J. Freudenberger, G. Zhang, M. J. Therien, M. I. Greene, D. A. Hammer and R. Murali, *Methods*, 2008, **46**, 25–32.
- 82 O. Onaca, R. Enea, D. W. Hughes and W. Meier, *Macromol. Biosci.*, 2009, **9**, 129–139.
- 83 H. K. Cho, I. W. Cheong, J. M. Lee and J. H. Kim, *Korean J. Chem. Eng.*, 2010, **27**, 731–740.
- 84 J. S. Lee and J. Feijen, *J. Controlled Release*, 2012, **161**, 473–483.
- 85 K. Osada, H. Cabral, Y. Mochida, S. Lee, K. Nagata, T. Matsuura, M. Yamamoto, Y. Anraku, A. Kishimura, N. Nishiyama and K. Kataoka, *J. Am. Chem. Soc.*, 2012, **134**, 13172–13175.
- 86 R. Wang, X. Hu, H. Xiao, Z. Xie, Y. Huang and X. Jing, *J. Mater. Chem. B*, 2013, **1**, 744–748.
- 87 W. Song, Z. Tang, M. Li, S. Lv, H. Sun, M. Deng, H. Liu and X. Chen, *Acta Biomater.*, 2014, **10**, 1392–1402.



- 88 J. Kim, H.-J. Yoon, S. Kim, K. Wang, T. Ishii, Y.-R. Kim and W.-D. Jang, *J. Mater. Chem.*, 2009, **19**, 4627–4631.
- 89 E. Stoyanova, V. Mitova, P. Shestakova, A. Kowalczyk, G. Momekov, D. Momekova, A. Marcinkowski and N. Koseva, *J. Inorg. Biochem.*, 2013, **120**, 54–62.
- 90 S.-M. Lee, H. Chen, C. M. Dettmer, T. V. O'Halloran and S. T. Nguyen, *J. Am. Chem. Soc.*, 2007, **129**, 15096–15097.
- 91 S.-M. Lee, H. Chen, T. V. O'Halloran and S. T. Nguyen, *J. Am. Chem. Soc.*, 2009, **131**, 9311–9320.
- 92 S.-M. Lee, T. V. O'Halloran and S. T. Nguyen, *J. Am. Chem. Soc.*, 2010, **132**, 17130–17138.
- 93 S. Binauld, W. Scarano and M. H. Stenzel, *Macromolecules*, 2012, **45**, 6989–6999.
- 94 Y. Mi, J. Zhao and S.-S. Feng, *J. Controlled Release*, 2013, **169**, 185–192.
- 95 N. Kolishetti, S. Dhar, P. M. Valencia, L. Q. Lin, R. Karnik, S. J. Lippard, R. Langer and O. C. Farokhzad, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 17939–17944.
- 96 W. Scarano, H. T. T. Duong, H. Lu, P. L. De Souza and M. H. Stenzel, *Biomacromolecules*, 2013, **14**, 962–975.
- 97 P. J. Bednarski, F. S. Mackay and P. J. Sadler, *Anticancer Agents Med. Chem.*, 2007, **7**, 75–93.
- 98 F. S. Mackay, J. A. Woods, P. Heringová, J. Kašpárková, A. M. Pizarro, S. A. Moggach, S. Parsons, V. Brabec and P. J. Sadler, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 20743–20748.
- 99 H. Xiao, G. T. Noble, J. F. Stefanick, R. Qi, T. Kiziltepe, X. Jing and B. Bilgicir, *J. Controlled Release*, 2014, **173**, 11–17.
- 100 R. Du, H. Xiao, G. Guo, B. Jiang, X. Yan, W. Li, X. Yang, Y. Zhang, Y. Li and X. Jing, *Colloids Surf., B*, 2014, **123**, 734–741.
- 101 J. Li, Y. Han, Q. Chen, H. Shi, S. ur Rehman, M. Siddiq, Z. Ge and S. Liu, *J. Mater. Chem. B*, 2014, **2**, 1813–1824.
- 102 H. Xiao, W. Li, R. Qi, L. Yan, R. Wang, S. Liu, Y. Zheng, Z. Xie, Y. Huang and X. Jing, *J. Controlled Release*, 2012, **163**, 304–314.
- 103 H. Xiao, L. Yan, Y. Zhang, R. Qi, W. Li, R. Wang, S. Liu, Y. Huang, Y. Li and X. Jing, *Chem. Commun.*, 2012, **48**, 10730–10732.
- 104 R. Qi, H. Xiao, S. Wu, Y. Li, Y. Zhang and X. Jing, *J. Mater. Chem. B*, 2015, **3**, 176–179.
- 105 S. Aryal, C.-M. J. Hu and L. Zhang, *ACS Nano*, 2010, **4**, 251–258.
- 106 H. Song, R. Wang, H. Xiao, H. Cai, W. Zhang, Z. Xie, Y. Huang, X. Jing and T. Liu, *Eur. J. Pharm. Biopharm.*, 2013, **83**, 63–75.
- 107 P. Plummer, R. H. Wilson, H. Calvert, A. V. Boddy, M. Griffin, J. Sludden, M. J. Tilby, M. Eatock, D. G. Pearson, C. J. Ottley, Y. Matsumura, K. Kataoka and T. Nishiya, *Br. J. Cancer*, 2011, **104**, 593–598.
- 108 T. Ueno, K. Endo, K. Hori, N. Ozaki, A. Tsuji, S. Kondo, N. Wakisaka, S. Muro, K. Kataoka, Y. Kato and T. Yoshizaki, *Int. J. Nanomedicine*, 2014, **9**, 3005–3012.
- 109 H. Wu, H. Cabral, K. Toh, P. Mi, Y.-C. Chen, Y. Matsumoto, N. Yamada, X. Liu, H. Kinoh, Y. Miura, M. R. Kano, H. Nishihara, N. Nishiyama and K. Kataoka, *J. Controlled Release*, 2014, **189**, 1–10.
- 110 D. P. Nowotnik and E. Cvitkovic, *Adv. Drug Delivery Rev.*, 2009, **61**, 1214–1219.
- 111 M. J. A. de Jonge, M. Slingerland, W. J. Loos, E. A. C. Wiemer, H. Burger, R. H. J. Mathijssen, J. R. Kroep, M. A. G. den Hollander, D. van der Biessen, M.-H. Lam, J. Verweij and H. Gelderblom, *Eur. J. Cancer*, 2010, **46**, 3016–3021.
- 112 G. P. Stathopoulos, D. Antoniou, J. Dimitroulis, J. Stathopoulos, K. Marosis and P. Michalopoulou, *Cancer Chemother. Pharmacol.*, 2011, **68**, 945–950.
- 113 N. Seetharamu, E. Kim, H. Hochster, F. Martin and F. Muggia, *Anticancer Res.*, 2010, **30**, 541–545.

