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Preparation and nanoaggregate formation ability in water of amphiphilic ladder-like polymers with parallelly linked hydrophilic polyether and hydrophobic polysiloxane chains†

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Aiming to enhance the high-temperature stability of nanoaggregates of amphiphilic polymers in water, an amphiphilic ladder-like (double-chain) polymer (PGE–PAMS) was successfully prepared *via* the thiol–ene reaction between 3-mercaptopropyltrimethoxymethylsilane and poly(allyl glycidyl ether), followed by intramolecular polycondensation (template polymerization) of the dimethoxysilyl groups in the side chains of the resulting polymer. Solubility tests, FT-IR spectroscopy, ¹H NMR spectroscopy, ²⁹Si NMR spectroscopy and transmission electron microscopy confirmed that PGE–PAMS possesses a ladder-like structure, where the hydrophilic poly(glycidyl ether) chain and the hydrophobic polyalkylmethylsiloxane chain are linked in parallel by sulfide bonds. PGE–PAMS formed nanoaggregates with a diameter of ca. 60–70 nm in water, which were stable even at high temperatures (80 °C–90 °C). Additionally, in water containing a small amount of *N,N*-dimethylformamide, the PGE–PAMS nanoaggregates could solubilize tetraphenylporphyrin, a hydrophobic dye, and stably retain it even at a high temperature of 90 °C.

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Introduction

Amphiphilic polymers contain both hydrophilic and hydrophobic segments within a single polymer chain, resulting in not only emulsifying, dispersing and solubilizing properties similar to those of low-molecular-weight surfactants but also diverse functionalities through self-assembly, such as the reversible formation of well-defined aggregates depending on the surrounding environment. By virtue of these characteristics, amphiphilic polymers are widely used as functional materials in various fields, including drug delivery systems,^{1,2} surface modification and functionalization³ and nanoscale patterning.^{4,5}

Amphiphilic molecules can be classified into ionic and nonionic types. Nonionic amphiphilic molecules are more commonly used because their self-assembly and interfacial activity functions are maintained even in salt-containing water, and they can also be combined with ionic amphiphilic molecules. Polyethylene glycol (PEG) is frequently used as the

hydrophilic segment of nonionic amphiphilic polymers, particularly in biotechnological applications like drug delivery systems, owing to its biocompatibility.^{6–15} As the hydrophobic segments of nonionic PEG-containing amphiphilic polymers, materials such as polypropylene glycol,^{6,7} poly(ϵ -caprolactone),^{8–10} polylactic acid,^{11,12} poly(lactic acid-*co*-glycolic acid)^{13,14} and polydimethylsiloxane (PDMS)¹⁵ are used.

In water, nonionic amphiphilic polymers form nanoaggregates such as micelles and vesicles, many of which exhibit temperature responsiveness because the hydrophilic PEG chain, which is surrounded by water molecules, undergo reversible hydration at low temperatures and dehydration at high temperatures.^{16–18} As a result, nanoaggregates of nonionic amphiphilic polymers can alter their shape in response to external stimuli such as temperature, which renders them promising candidates for smart materials applications.

However, only a limited number of nonionic amphiphilic polymers can stably maintain the nanoaggregates at high temperatures; instead, large aggregates that cause turbidity in water dispersions are formed. This is because, at high temperatures, water becomes a poor solvent not only for the hydrophobic segments but also for PEG.¹⁹ Considering that the hydration/dehydration of hydrophilic polymer chains is related to their conformation,¹⁶ suppressing the conformational change of the PEG chain could help maintain its hydrophilicity even at high temperatures.

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In fact, in a previous study, Honda *et al.* reported that a macrocyclic amphiphilic polymer composed of a hydrophilic PEG chain and a hydrophobic poly(butyl acrylate) chain exhibited a cloud point (T_c) of *ca.* 70 °C in water, which was *ca.* 40 °C higher than that of a linear amphiphilic polymer having the same components.²⁰ This suggests that the micelles formed by macrocyclic amphiphilic polymers remain stable even at relatively high temperatures (70 °C), which may be attributed to the inhibition of conformational changes due to the unique cyclic shape of the polymers. However, the relationship between the shape of amphiphilic polymers and the stability of their nanoaggregates at high temperatures has not yet been fully clarified. Unveiling this relationship is expected to contribute to the development of amphiphilic polymers with novel structures.

In this study, we were interested in investigating amphiphilic polymers with a ladder-like structure, which are expected to exhibit different properties from single-chain polymers due to the suppression of conformational changes.^{21,22} For example, Gu *et al.* reported that a low-defect conjugated ladder-like polymer forms stable aggregates in solution and remains aggregated at high temperatures up to 75 °C in chlorobenzene compared with a conjugated non-ladder-like polymer control.²³

Ladder-like polymers can be prepared using various methods. For instance, the hydrolytic polycondensation of limited organotrialkoxysilanes, *i.e.*, the sol-gel reaction, yields ladder-like polysilsesquioxanes,^{24–35} which exhibit more rigid properties than linear polysiloxanes. Meanwhile, template polymerization is a widely recognized method for preparing ladder-like organic polymers. In this approach, polymerizable groups are introduced into the side chains of a base polymer, which then undergoes intramolecular polymerization. Kämmerer *et al.* reported the preparation of a ladder-like oligomer *via* intramolecular free-radical polymerization of a *p*-cresol oligomer containing acrylic groups.^{36,37} Similarly, Jantas *et al.* prepared a ladder-like polymer by the intramolecular free-radical polymerization of multivinyl monomers based on polyvinyl alcohol and poly(meth)acrylate.^{38–42} Saito *et al.* prepared a ladder-like polymer through the intramolecular atom transfer radical polymerization of poly(2-methacryloyloxyethyl methacrylate).^{43,44} Recently, we reported a ladder-like polysiloxane with two siloxane main chains, which was prepared by introducing diethoxysilyl groups as polymerizable groups into the side chains of ammonium-containing polysiloxane,⁴⁵ followed by intramolecular polycondensation.⁴⁶

Herein, to investigate the effect of the inhibition of conformational changes due to the unique shape of the double chains on the stability of the self-assembly, we adopted the template polymerization method to prepare a ladder-like non-ionic amphiphilic polymer comprising a poly(glycidyl ether) (PGE) chain as the hydrophilic segment and a polyalkylmethylsiloxane (PAMS) chain as the hydrophobic segment, which were linked in parallel. To the best of our knowledge, a ladder-like polymer consisting of two polymer chains with such distinct properties has not been reported to date. The resulting PGE-PAMS polymer formed nanoaggregates with a diameter of

ca. 60–70 nm in water, and these nanoaggregates remained stable even at high temperatures (80 °C–90 °C). Furthermore, in water containing a small amount of *N,N*-dimethylformamide (DMF), the PGE-PAMS nanoaggregates could solubilize tetraphenylporphyrin (TPP), a hydrophobic dye. Even when the solution was heated to 90 °C, TPP did not precipitate, suggesting that the PGE-PAMS nanoaggregates can stably encapsulate TPP at high temperatures.

Results and discussion

Preparation of poly(allyl glycidyl ether) (PAGE)

PAGE was prepared as the precursor polymer *via* the anionic ring-opening polymerization of allyl glycidyl ether, following a previously reported procedure.^{47,48} PAGE was soluble in dimethylsulfoxide (DMSO), DMF, methanol (CH₃OH), acetone, chloroform, ethyl acetate, tetrahydrofuran (THF) and diethyl ether but insoluble in hexane and water. The number-average molecular weight (M_n) and molecular weight distribution (M_w/M_n) of PAGE were estimated to be 2.5×10^3 and 1.31, respectively, using gel permeation chromatography (GPC). Furthermore, its average degree of polymerization (DP) was calculated to be 46 according to end-group quantification using ¹H NMR spectroscopy. This calculation considered the formula weights of both the repeating and initiating units, yielding an M_n of 5.3×10^3 (Fig. S1†).

Preparation of PGE-PAMS

PGE-PAMS was prepared by means of the thiol-ene reaction between 3-mercaptopropyltrimethoxymethylsilane (MPDMMS) and PAGE, followed by intramolecular polycondensation (template polymerization) of the dimethoxysilyl groups in the side chains of the resulting polymer.

First, the thiol-ene reaction of MPDMMS with PAGE was performed using phenylbis(2,4,6-trimethylbenzoyl)phosphine oxide (BAPO) as a photoradical initiator in dehydrated THF, resulting in poly(glycidyl ether) containing dimethoxysilyl groups as side chains (PGE-DS, Scheme 1a). To confirm the progress of the thiol-ene reaction, we attempted to isolate the



Scheme 1 Preparation of (a) poly(glycidyl ether) containing dimethoxysilyl groups (PGE-DS) and (b) a ladder-like polymer with a poly(glycidyl ether) (PGE) chain as the hydrophilic segment and a polyalkylmethylsiloxane (PAMS) chain as the hydrophobic segment (PGE-PAMS).

solid product *via* reprecipitation. However, the reprecipitated product was insoluble in any solvent. The cause of insolubility is believed to be the partial hydrolysis of the diethoxysilyl groups, which leads to intermolecular dehydration condensation involving the formed silanol groups, resulting in the formation of intermolecular crosslinking. Therefore, after completing the reaction, a large amount of DMF was added to the reaction solution to prepare a dilute solution (1.0×10^{-3} mol unit per L) without isolating the product. To avoid the intermolecular reaction between the dimethoxysilyl groups in PGE-DS side chains, which should react only intramolecularly, the solution concentration was reduced during the subsequent polycondensation.

The intramolecular polycondensation of the dimethoxysilyl groups attached to the polymer chain of PGE-DS was performed by adding purified water, followed by stirring at 120 °C for 120 h (Scheme 1b). To isolate the product, the DMF solution was concentrated and reprecipitated from diethyl ether. The diethyl ether-insoluble fraction was then isolated by decantation and washed with diethyl ether. Finally, the product was dried under reduced pressure to yield PGE-PAMS. PGE-PAMS was soluble in water and in organic solvents such as DMSO, CH₃OH, ethanol (C₂H₅OH), and isopropyl alcohol (iso-C₃H₇OH) (Fig. 1a–e), demonstrating its amphiphilic nature. Meanwhile, it was insoluble in organic solvents such as DMF, acetone, THF, chloroform, toluene and hexane.

Characterization of PGE-PAMS

PGE-PAMS was characterized *via* ¹H NMR, ²⁹Si NMR and transmission electron microscopy (TEM) analyses, which confirmed its ladder-like structure with PGE and PAMS chains linked in parallel *via* sulfide bonds.

In the ¹H NMR spectrum of PGE-PAMS in CD₃OD, the signals attributed to the allyl groups (5.40–5.10 ppm) almost disappeared, indicating the progress of the thiol-ene reaction between MPDMMS and PAGE. Additionally, the average DP of the PAMS chain was estimated to be *ca.* 39.1 by calculating the integral ratio of signal **k** to signal **b**, which correspond to the phenolate-initiated end group and the methylene proton adjacent to the Si atom, respectively, in the ¹H NMR spectrum (Fig. 2). Furthermore, the average DP of the PGE chain was *ca.* 36.4, of which 33.5 were units linked to PAMS chains and 2.9 were units with unreacted allyl groups, indicating that 8.0% of



Fig. 2 ¹H NMR spectrum of PGE-PAMS in CD₃OD. The chemical shifts are referenced to CD₃OD (δ 3.31). *: Unknown signal.

the PGE chains contained units with unreacted allyl groups. This was determined from the integral ratio of signal **k** to signals **d** and **i**, which are attributed to the phenolate-initiated end group, methylene protons and double bonds, respectively, in the ¹H NMR spectrum (Fig. 2).

Thus, the ¹H NMR end-group quantification revealed a difference between the DP of the PGE chain (36.4) and that of the PAMS chain (39.1). This suggests that the PAMS chains contain units with mercapto groups in addition to units linked to the PGE chains. The integral ratio of signal **b** to signal **d** was 2.00 : 1.72, and that of signal **c** to signal **d** was 2.00 : 1.61 (Fig. 2), further supporting the higher DP of the PAMS chain compared with that of the PGE chain. According to the integral ratio of signal **b** to signal **d**, *ca.* 14.3% of the PAMS chain contains units with mercapto groups. Assuming an average DP of 39.1 for the PAMS chain, 33.5 units are linked to the PGE chains and 5.6 units contain mercapto groups. Furthermore, from the average DPs of the PGE and PAMS chains, the M_n of PGE-PAMS was estimated to be *ca.* 1.0×10^4 .

The ²⁹Si NMR spectrum of PGE-PAMS in CD₃OD at 40 °C exhibited two broad signals in the D² regions at –17 to –21 ppm and –21 to –24 ppm, with an integral ratio of 68 : 32, whereas no signals attributable to a D¹ or D⁰ structure were detected (Fig. 3a). The predominance of the D² structure indicates that sufficient polycondensation of the dimethoxysilyl groups in PGE-DS occurred, resulting in the formation of the Si–O–Si bond main chain.

As mentioned earlier, the signals attributed to the D² structure of the PAMS chain were split into two. These signals are presumed to correspond to units linked to the PGE chain *via* sulfide bonds (–17 to –21 ppm) and units with mercapto groups (–21 to –24 ppm) (Fig. 3a). According to the ¹H NMR spectrum (Fig. 2), *ca.* 14.3% of the repeating units in the PAMS chain contain mercapto group components, supporting this interpretation. Although the estimated proportions of mercapto-containing units in the PAMS chains, as determined by ²⁹Si and ¹H NMR measurements, differ (*ca.* 32% and 14.3%,



Fig. 1 Photographs of PGE-PAMS solutions in (a) water, (b) DMSO, (c) CH₃OH, (d) C₂H₅OH and (e) iso-C₃H₇OH (concentration: 0.1 w/v%).



Fig. 3 ^{29}Si NMR spectra of PGE-PAMS obtained *via* intramolecular polycondensation of PGE-DS for (a) 120 h, (b) 96 h and (c) 48 h in CD_3OD at 40°C . A small amount of $\text{Cr}(\text{acac})_3$ was added as a relaxation agent. The chemical shifts are referenced to tetramethylsilane (TMS, δ 0.0).

respectively), this discrepancy is considered to fall within the range of experimental error.

Since the repeating unit length of the PAMS chain, which consists of two main atoms (Si-O), is shorter than that of the PGE chain, which has three main atoms (C-C-O), introduction of mercapto-containing units in the PAMS chain most likely helps balance the chain lengths between PGE and PAMS, thereby alleviating strain. From an alternative perspective, the strain might also be alleviated by the long crosslinking units even without the introduction of MPDMMS. However, in such a case, it is speculated that the molecular chains would curve rather than straighten due to the mismatch in the repeating unit lengths of the PAMS and PGE chains. As evidenced by TEM observations (Fig. 4), described in detail below, the molecular chains are straight, suggesting that the introduction of MPDMMS plausibly explains this outcome.

Performing the intramolecular polycondensation of PGE-DS for shorter times (96 and 48 h) resulted in a decrease in the proportion of D^2 signals and an increase in the proportion of D^1 (D^1 : $\text{C}_2\text{Si}(\text{OSi})(\text{OR})$, $\text{R} = \text{H}$ or CH_3) and $(\text{D}^2)_{\text{c}3}$ signals ($(\text{D}^2)_{\text{c}3}$ is a cyclic trimer component⁴⁹) in the ^{29}Si NMR spectra of the resulting products (Fig. 3b and c). These results indicate that a reaction time of 120 h is essential for obtaining PGE-PAMS *via* the intramolecular polycondensation of PGE-DS. Therefore, subsequent experiments were performed using PGE-PAMS prepared with an intramolecular polycondensation time of 120 h.

To examine the structure of PGE-PAMS, TEM measurements were performed. A droplet of an aqueous solution of PGE-PAMS was placed onto a TEM grid and dried without electron staining. The TEM image of the resulting sample revealed straight polymer chains arranged in parallel stripe patterns (Fig. 4), indicating that PGE-PAMS possesses a one-dimensional structure. The width of a single black-and-white



Fig. 4 TEM image of a sample obtained by drying an aqueous solution of PGE-PAMS.

stripe set was estimated to be *ca.* 3.33 nm. The black-and-white regions can be attributed to the inorganic PAMS components (Si-rich regions) and the organic PGE components (C-rich regions), respectively. On the basis of the molecular dimensions of a model compound representing the repeating unit of PGE-PAMS (Fig. 5a), the molecular width of PGE-PAMS was estimated to be *ca.* 1.76 nm. Accordingly, the stripe widths of *ca.* 3.33 nm observed in the TEM image suggest that the polymer chains are arranged in a regular pattern in which identical segments are aligned face-to-face (Fig. 5b).



Fig. 5 (a) Molecular dimensions of a model compound representing the repeating unit of PGE-PAMS, energy-minimized using MM2 in the CS Chem3D program package. (b) Plausible stacking structure of PGE-PAMS.



Fig. 8 Transmittance curves ($\lambda = 500$ nm) of (a) PEG-*block*-PDMS-*block*-PEG, (b) PDMS-*graft*-PEG and (c) PGE-PAMS in water during heating. The polymer concentration was 0.1 w/v%. Inset: photographs of (d) PEG-*block*-PDMS-*block*-PEG, (e) PDMS-*graft*-PEG and (f) PGE-PAMS at 90 °C in water (0.1 w/v%).

cloudy at 90 °C (Fig. 8d and e). These findings suggest the formation of large aggregates above these temperatures, indicating that block and graft copolymers containing PDMS and PEG components have the T_c of 61 °C and 81 °C. In contrast, the transmittance of the PGE-PAMS aqueous solution did not decrease upon heating to 90 °C (Fig. 8c). Visual observation revealed that the PGE-PAMS solution remained transparent at 90 °C, without the formation of turbidity or a precipitate (Fig. 8f), indicating that the nanoaggregates remained stable even at high temperatures.

Furthermore, the volume-average particle size of PGE-PAMS was found to be 84.0 ± 29.8 nm by performing DLS measurements in water (0.1 w/v%) at high temperatures up to the limit of our instrument (80 °C), (Fig. 7b). This suggests that the rigid ladder-like structure of PGE-PAMS hinders conformational changes in the hydrophilic PGE chain, thereby preventing dehydration, which explains the lack of formation of large aggregates even at high temperatures of 80 °C–90 °C.

Solubilization of TPP by PGE-PAMS nanoaggregates

Amphiphilic molecules can encapsulate and solubilize hydrophobic molecules within the hydrophobic space of the nanoaggregates formed in water. Here, the encapsulation behaviour of the PGE-PAMS nanoaggregates in water containing a small amount of DMF was evaluated using the hydrophobic dye TPP in free base form as the guest molecule.

Before investigating the solubilization of TPP by PGE-PAMS in a water/DMF (9/1, v/v) mixed solvent, the solubility of TPP in the mixed solvent was determined in the absence of PGE-



Fig. 9 Photographs of water/DMF (9/1, v/v) solutions of (a) TPP at 25 °C and (b) a mixture of PGE-PAMS and TPP at 25 °C. (c) Volume-average particle size distribution of a mixture of PGE-PAMS and TPP measured by DLS in a water/DMF (9/1, v/v) mixed solvent at 25 °C (concentration: 1.0×10^{-2} w/v% for PGE-PAMS and 2.5×10^{-3} w/v% for TPP). Photographs of water/DMF (9/1, v/v) solutions of a mixture of PGE-PAMS and TPP (d) at 90 °C and (e) after being left at room temperature for 24 h.

PAMS. When 2 mL of a DMF solution containing TPP (0.5 mg) was slowly mixed with 18 mL of purified water under stirring, TPP gradually precipitated (Fig. 9a). In contrast, when a mixture of PGE-PAMS (2.0 mg) and TPP (0.5 mg) in DMF was slowly mixed with purified water (18 mL) under stirring, TPP remained soluble in the mixed solvent without precipitation (Fig. 9b). Additionally, DLS measurement of this mixture confirmed the formation of nanoaggregates with a particle size of *ca.* 65 nm (Fig. 9c). From these findings, it can be inferred that TPP was encapsulated within the hydrophobic space of the PGE-PAMS nanoaggregates.

The UV-Vis spectrum of a mixed solution of PGE-PAMS (2.0×10^{-3} w/v%) and TPP (5.0×10^{-4} w/v%) revealed that TPP existed in a monomeric state ($\lambda_{\max} = 420$ nm) when solubilized by PGE-PAMS (Fig. S6†). This was confirmed by comparing this UV-Vis spectrum with that of a dilute solution of TPP alone in chloroform (a good solvent for TPP, 5.0×10^{-4} w/v%) and with a previous report⁵⁰ (Fig. S7†).

In addition, to investigate whether the solubilized state of TPP could be maintained at high temperatures, the mixed solution of PGE-PAMS and TPP was heated to 90 °C, and it was found that no precipitate was formed (Fig. 9d). This suggests that TPP was stably encapsulated within the PGE-PAMS nanoaggregates even at 90 °C, since the collapse or aggregation of the nanoaggregates would cause the precipitation of either TPP or PGE-PAMS from the mixed solution. Additionally, after being left at room temperature for 24 h, no precipitate was observed, confirming that TPP remained solubilized (Fig. 9e).

In contrast, although PEG-*block*-PDMS-*block*-PEG and PDMS-*graft*-PEG could solubilize TPP in a water/DMF (9/1, v/v) mixed solvent at room temperature, precipitation occurred upon heating to 90 °C and standing for 1 h (Fig. S8†). These

results indicate that the nanoaggregates formed by the block and graft copolymers can solubilize TPP but are unable to stably maintain the TPP encapsulation in the water/DMF (9/1, v/v) mixed solvent at high temperatures, despite containing similar components.

It has been reported that the intermolecular interactions of a rigid conjugated ladder-like polymer are considerably more stable at higher temperatures than those of a non-ladder-like polymer with similar structures owing to the low entropy nature of the rigid ladder-like polymer.²³ Therefore, the stability of the PGE-PAMS nanoaggregates at high temperature can be attributed to the low entropy of its ladder-like structure as well as to the suppression of the conformational changes of the hydrophilic segment, which prevent dehydration.

Conclusions

In this study, PGE-PAMS, an amphiphilic ladder-like polymer, was successfully prepared *via* the thiol-ene reaction between MPDMMS and PAGE, followed by intramolecular polycondensation of the dimethoxysilyl groups in the side chains of the resulting polymer, PGE-DS. The results of solubility tests, GPC, ¹H NMR, ²⁹Si NMR and TEM suggest that PGE-PAMS possesses a ladder-like structure composed of PGE chains as hydrophilic segments and PAMS chains as hydrophobic segments, which are linked in parallel by sulfide bonds. This is the first report on the preparation of a ladder-like polymer with two chains exhibiting highly contrasting properties. In water, PGE-PAMS formed nanoaggregates with a diameter of *ca.* 60–70 nm, which remained stable even at high temperatures (80 °C–90 °C). This stability is likely due to the ladder-like structure of PGE-PAMS preventing conformational changes in the PGE chain, thereby inhibiting dehydration of the hydrophilic segments. Additionally, in water containing a small amount of DMF, the PGE-PAMS nanoaggregates could solubilize a hydrophobic dye (TPP), which remained encapsulated within the nanoaggregates even at a high temperature of 90 °C.

Author contributions

Conceptualization, Y. K.; data curation, S. M., S. N. and A. M.; formal analysis, S. M., S. N. and A. M.; investigation, S. M.; methodology, S. M.; supervision, Y. K.; writing – original draft, S. M. and S. N.; writing – review & editing, Y. K.; TEM measurement, K. S. All authors have read and agreed to the published version of the manuscript.

Data availability

The data supporting this article have been included as part of the ESI.†

Conflicts of interest

There are no conflicts to declare.

References

- 1 K. Kataoka, A. Harada and Y. Nagasaki, *Adv. Drug Delivery Rev.*, 2001, **47**, 113–131.
- 2 H. Cabral, K. Miyata, K. Osada and K. Kataoka, *Chem. Rev.*, 2018, **118**, 6844–6892.
- 3 T. Yoshida, M. Doi, S. Kanaoka and S. Aoshima, *J. Polym. Sci., Part A: Polym. Chem.*, 2005, **43**, 5704–5709.
- 4 T. Thurn-Albrecht, J. Schotter, G. A. Kästle, N. Emley, T. Shibauchi, L. Krusin-Elbaum, K. Guarini, C. T. Black, M. T. Tuominen and T. P. Russell, *Science*, 2000, **290**, 2126–2129.
- 5 A. C. Edrington, A. M. Urbas, P. DeRege, C. X. Chen, T. M. Swager, N. Hadjichristidis, M. Xenidou, L. J. Fetters, J. D. Joannopoulos, Y. Fink and E. L. Thomas, *Adv. Mater.*, 2001, **13**, 421–425.
- 6 T. H. Vanghn, H. R. Suter, L. G. Lundsted and M. G. Kramer, *J. Am. Oil Chem. Soc.*, 1951, **28**, 294–299.
- 7 C. Booth and D. Attwood, *Macromol. Rapid Commun.*, 2000, **21**, 501–527.
- 8 C. Y. Gong, S. Shi, P. W. Dong, B. Kan, M. L. Gou, X. H. Wang, X. Y. Li, F. Luo, X. Zhao, Y. Q. Wei and Z. Y. Qian, *Int. J. Pharm.*, 2009, **365**, 89–99.
- 9 M. Boffito, P. Sirianni, A. M. Di Rienzo and V. Chiono, *J. Biomed. Mater. Res., Part A*, 2015, **103A**, 1276–1290.
- 10 H. Deng, A. Dong, J. Song and X. Chen, *J. Controlled Release*, 2019, **297**, 60–70.
- 11 G. Bonacucina, M. Cespi, G. Mencarelli, G. Giorgioni and G. F. Palmieri, *Polymers*, 2011, **3**, 779–811.
- 12 J. Shi, L. Yu and J. Ding, *Acta Biomater.*, 2021, **128**, 42–59.
- 13 B. Jeong, Y. H. Bae and S. W. Kim, *Colloids Surf., B*, 1999, **16**, 185–193.
- 14 P. Wang, W. Chu, X. Zhuo, Y. Zhang, J. Gou, T. Ren, H. He, T. Yin and X. Tang, *J. Mater. Chem. B*, 2017, **5**, 1551–1565.
- 15 H. Kunieda, M. H. Uddin, M. Horii, H. Furukawa and A. Harashima, *J. Phys. Chem. B*, 2001, **105**, 5419–5426.
- 16 M. J. Hey, S. M. Ilett and G. Davidson, *J. Chem. Soc., Faraday Trans.*, 1995, **91**, 3897–3900.
- 17 K. Tasaki, *J. Am. Chem. Soc.*, 1996, **118**, 8459–8469.
- 18 C. de las Heras Alarcón, S. Pennadam and C. Alexander, *Chem. Soc. Rev.*, 2005, **34**, 276–285.
- 19 P. Alexandridis and T. A. Hatton, *Colloids Surf., A*, 1995, **96**, 1–46.
- 20 S. Honda, T. Yamamoto and Y. Tezuka, *J. Am. Chem. Soc.*, 2010, **132**, 10251–10253.
- 21 C. Arnold Jr., *J. Polym. Sci., Macromol. Rev.*, 1979, **14**, 265–378.
- 22 M. M. Tessler, *J. Polym. Sci., Part A-1*, 1966, **4**, 2521–2532.
- 23 G. Ma, M. Leng, S. Li, Z. Cao, Y. Cao, D. P. Tabor, L. Fang and X. Gu, *J. Mater. Chem. C*, 2022, **10**, 13896–13904.

- 24 J. F. Brown Jr., L. H. Vogt Jr., A. Katchman, J. W. Eustance, K. M. Kiser and K. W. Krantz, *J. Am. Chem. Soc.*, 1960, **82**, 6194–6195.
- 25 J. F. Brown Jr., L. H. Vogt Jr. and P. I. Prescott, *J. Am. Chem. Soc.*, 1964, **86**, 1120–1125.
- 26 Y. Abe and T. Gunji, *Prog. Polym. Sci.*, 2004, **29**, 149–182.
- 27 Y. Kaneko, N. Iyi, K. Kurashima, T. Matsumoto, T. Fujita and K. Kitamura, *Chem. Mater.*, 2004, **16**, 3417–3423.
- 28 Y. Kaneko, N. Iyi, T. Matsumoto and K. Kitamura, *Polymer*, 2005, **46**, 1828–1833.
- 29 X. Zhang, P. Xie, Z. Shen, J. Jiang, C. Zhu, H. Li, T. Zhang, C. C. Han, L. Wan, S. Yan and R. Zhang, *Angew. Chem., Int. Ed.*, 2006, **45**, 3112–3116.
- 30 S. S. Choi, H. S. Lee, S. S. Hwang, D. H. Choi and K. Y. Baek, *J. Mater. Chem.*, 2010, **20**, 9852–9854.
- 31 Z. Ren, D. Sun, H. Li, Q. Fu, D. Ma, J. Zhang and S. Yan, *Chem. – Eur. J.*, 2012, **18**, 4115–4123.
- 32 H. Toyodome, Y. Kaneko, K. Shikinaka and N. Iyi, *Polymer*, 2012, **53**, 6021–6026.
- 33 Y. Kaneko, H. Toyodome, T. Mizumo, K. Shikinaka and N. Iyi, *Chem. – Eur. J.*, 2014, **20**, 9394–9399.
- 34 A. Harada, K. Shikinaka, J. Ohshita and Y. Kaneko, *Polymer*, 2017, **121**, 228–233.
- 35 Y. Kaneko, *Polymer*, 2018, **144**, 205–224.
- 36 H. Kämmerer and S. Ozaki, *Makromol. Chem.*, 1966, **91**, 1–9.
- 37 H. Kämmerer and A. Jung, *Makromol. Chem.*, 1966, **101**, 284–295.
- 38 R. Jantas and S. Polowinski, *J. Polym. Sci., Part A: Polym. Chem.*, 1986, **24**, 1819–1827.
- 39 R. Jantas, *J. Polym. Sci., Part A: Polym. Chem.*, 1990, **28**, 1973–1982.
- 40 R. Jantas, G. Janowska, H. Szocik and S. Polowinski, *J. Therm. Anal. Calorim.*, 2000, **60**, 371–376.
- 41 R. Jantas, J. Szumilewicz, G. Strobin and S. Polowinski, *J. Polym. Sci., Part A: Polym. Chem.*, 1994, **32**, 295–300.
- 42 R. Jantas, S. Polowinski and G. Strobin, *Polym. Int.*, 1995, **37**, 315–318.
- 43 R. Saito, Y. Iijima and K. Yokoi, *Macromolecules*, 2006, **39**, 6838–6844.
- 44 R. Saito and Y. Iijima, *Polym. Adv. Technol.*, 2009, **20**, 280–284.
- 45 S. Kinoshita, S. Watase, K. Matsukawa and Y. Kaneko, *J. Am. Chem. Soc.*, 2015, **137**, 5061–5065.
- 46 M. Nobayashi, K. Shikinaka and Y. Kaneko, *Polym. J.*, 2022, **54**, 11–20.
- 47 S. Aoki, A. Koide, S. Imabayashi and M. Watanabe, *Chem. Lett.*, 2002, **31**, 1128–1129.
- 48 B. F. Lee, M. J. Kade, J. A. Chute, N. Gupta, L. M. Campos, G. H. Fredrickson, E. J. Kramer, N. A. Lynd and C. J. Hawker, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 4498–4504.
- 49 Y. Sugahara, S. Okada, K. Kuroda and C. Kato, *J. Non-Cryst. Solids*, 1992, **139**, 25–34.
- 50 M. Zhou, S. Ouyang, Z. Liu, G. Lu, S. Gao and Z. Li, *Vib. Spectrosc.*, 2009, **49**, 7–13.