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# **REVIEW**

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# A comprehensive overview of vaccines developed for pandemic viral pathogens over the past two decades including those in clinical trials for the current novel SARS-CoV-2

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The unprecedented coronavirus disease 2019 (COVID-19) is triggered by a novel strain of coronavirus namely, Severe Acute Respiratory Syndrome-Coronavirus-2 (SARS-CoV-2). Researchers are working around the clock to control this pandemic and consequent waves of viral reproduction, through repurposing existing drugs as well as designing new vaccines. Several countries have hastened vaccine design and clinical trials to quickly address this outbreak. Currently, more than 250 aspirants against SARS-CoV-2 are in progress, including mRNA-replicating or non-replicating viral vectored-, DNA-, autologous dendritic cell-based-, and inactivated virus-vaccines. Vaccines work by prompting effector mechanisms such as cells/molecules, which target quickly replicating pathogens and neutralize their toxic constituents. Vaccine-stimulated immune effectors include adjuvant, affinity, avidity, affinity maturation, antibodies, antigen-presenting cells, B lymphocytes, carrier protein, CD4+ T-helper cells. In this review, we describe updated information on the various vaccines available over the last two decades, along with recent progress in the ongoing battle developing 63 diverse vaccines against SARS-CoV-2. The inspiration of our effort is to convey the current investigation focus on registered clinical trials (as of January 08, 2021) that satisfy the safety and efficacy criteria of international wide vaccine development.

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research at Tamkang University, Taiwan. Then he was employed as a SERB-post-doctoral fellow at the CSIR-Central Leather Research Institute, Chennai, India. He has synthesized substituted alkynylated hybrid molecules and utilized them for NIR and EL applications; and liquid crystalline-, steroidal-based materials for sensor and biomedical applications; he also worked on the characterization of natural product transitmycin.



Senthilkumar obtained his M.Sc. chemistry at Bharathidasan University, where he was involved in mechanistic studies of anticancer drugs. He explored new discovery and development from marine and medicinal plants, and their synthesis which is been developed at IIT Madras, IISc Bangalore and IIT Kanpur. He developed variegated candidates at IIT Madras, such

as phosphate binder resin, sevcar (two patents); and tolvaptan, hyponatremia and cystagon; and drugs from Terminalia arjuna. He then engaged in the peptidomimetic domain for antitumor screening at IISc Bangalore. Then back at IIT Madras worked on transitmycin, which is in phase-II pre-clinical trials. He has also discovered new blood, breast, and liver cancer drugs, and diabetic treatments. Recently, he also discovered, arjunetin, that is a better candidate for COVID-19 treatment than FDA approved drugs.

## Introduction

One of the most successful therapeutic strategies to prevent or control various diseases is by "vaccination" protocol. 1,2 Millions of lives have been saved because of vaccinations, which cover a number of diseases including certain types of cancer, HIV and many viral infections.3 Vaccination was initially accomplished by Edward Jenner, who was the pioneer for smallpox in the late 18th century.4 In the 1980s, the development of vaccines to fight against pathogenic microorganisms was tentatively introduced. Vaccines are now employed to improve and increase the protection capability (immunity) of the body to fight severe infection and disease,5 and moreover, primarily intend to make immunity stronger and reduce resistivity of diseases by reducing the reproduction of target pathogens. The majority of vaccines actively exist in the immune system, since anti-bodies are constantly generated in the body to sustain a healthy immunity system.6,7

#### General components of vaccines

A vaccine consists of an antigen, stabilizer, adjuvant, antibiotic, preservative, and chemical reagents such as formaldehyde. The advantages and disadvantages of vaccinations are listed in Table 1.

**Antigen.** The component matching the structural array of disease-oriented organisms, wherein, they are identified by the immune system as 'foreign' and cause an active immune response.

**Stabilizer.** This module is employed to assist the vaccine by sustaining its efficiency during storage. Instability of the vaccine can lead to reduced antigenicity and decreased infectivity of live attenuated vaccine (LAV). Magnesium chloride (MgCl $_2$ ) for oral polio vaccine (OPV), magnesium sulfate (MgSO $_4$ ) in measles vaccines, and lactose and gelatin associated with sorbitol, are current representatives of stabilizing factors.

**Adjuvant.** They are responsible for enhancing the efficacy of the vaccine by motivating the generation of antibodies.



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mingham, UK) and has postdoctoral experience (University of Cambridge, UK, and Texas A & M, USA). He has authored 320 technical papers, 10 books, and filed 15 patents and is a director of two startup biotechnology companies.

Chemically, adjuvants are a highly heterogeneous group of compounds (including Al salts).

Antibiotics. Used in lower amounts during the development phase, to circumvent bacterial infection during tissue culture cells where the viruses are grown. MMR (measles, mumps, rubella) and IPV (inactivated polio vaccine) associated vaccines have a minimum amount ( $<25~\mu g$ ) of neomycin for each dose.

The MMR vaccine was developed by Maurice Hilleman in 1971. Mumps, like measles infections, are caused by an RNA based virus from the Paramyxoviridae family. Moreover, measles and mumps belong to the genus Rubulavirus, it is a human disease with no animal reservoirs. Generally, the MMR vaccine exhibits side effects of a painful arm from the shot, minor rashes, generally in teen/adult women who have no earlier immunity; then the rubella vaccine component can result in joint and tendon stiffness. This vaccine is also associated with the minor threat of seizures/jerking instigated by fever, but is not connected with any enduring effects. The threat of febrile seizures increases as infants get older, hence this vaccine is recommended at a young age. Some people may experience cheek/neck inflammation, impermanent low platelet counts that generally do not require treatment and are also not life threatening.9b

In 1955, Jonas Salk initiated an inactivated polio vaccine (IPV), after that, Albert Sabin further developed the live, OPV. Even though poliovirus has three serotypes, both vaccines are trivalent and offer good resistivity against poliomyelitis, a limited number of countries have continuously provided IPV. Sabin's OPV vaccine is underused in most countries due to minimising oral management, but it advances immunity in the intestine, which is capable of spreading to others, and is associated with lower cost. 9c

**Preservatives.** They are added in multi-dose vaccines, which can be used to control bacterial and fungal growth, including thiomersal (sodium(2-carboxylatophenyl)sulfanyl-ethyl mercury), formaldehyde, or phenolic derivatives. In fact, formaldehyde is responsible for inactivating the viruses (*e.g.* IPV) and removing toxicity in bacterial strains, including the toxin employed in diphtheria and tetanus vaccines.

#### Types of vaccines

Various kinds of vaccines are available and those which are administered to infants and adults can be classified (Fig. 1) as follows:

- Live-attenuated.
- Inactivated.
- Toxoid.
- Conjugate.
- Subunit.

Live attenuated vaccines. These vaccines are adapted from the existing bacteria or virus, they have been weakened, and thus will not result in serious disease in people with strong immunity. LAV vaccines are more similar to the actual infection. A few representative examples are the MMR and varicella (chickenpox) vaccines. Albeit, it is efficient, but not everyone **RSC Advances** Review

Table 1 Advantages and disadvantages of vaccines

| Advantages   | Disadvantages   |
|--|---|
| Protection of inhabitants against disease                | Uncertainty about complete protection                                       |
| Preventing epidemic and pandemic diseases                | May have some possible side effects   |
| Prevents diseases spreading to others                    | Requires NHS/individual outlay  |
| Avoids large cost for the treatment of infected patients | Injections are not pleasant/more immune booster injections are inconvenient |
|  |   |

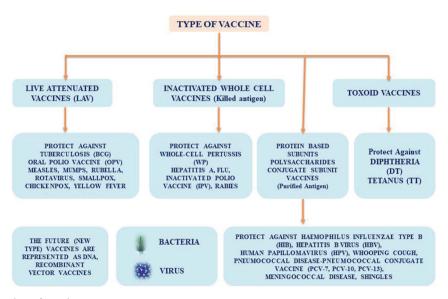


Fig. 1 Pictorial representation of vaccine types.

can be administered these vaccines, including children and patients undergoing chemotherapy as their immune system is too weak.

Inactivated vaccines. These vaccines are made from inactivated or dead organisms. The polio vaccine is an example of an inactivated vaccine, which generates an immune response through various routes different to the live attenuated vaccine. It requires a higher dosage to increase and/or sustain immunity.

**Toxoid vaccines.** This vaccine prevents the disease resulting from the toxins released by a virus into the body. Since the toxic substances administered are weak they are unable to cause the illness, while the immune system which encounters this toxoid vaccine, becomes able to repel the natural toxin. The DTaP vaccine for diphtheria and tetanus toxoids falls into this category.

Conjugate vaccines. They defend the system against different kinds of bacteria which have antigens that are surface coated with polysaccharide units. It is obvious, that this sugar coated unit masks the antigen. These vaccines append (or conjugate) themselves to the polysaccharide units on the antigens. Haemophilus influenzae type B (Hib) vaccine is an example of the conjugate type.

**Subunit vaccines.** These vaccines are made up of simple fractions of the virus/bacteria or subunits, rather than the complete germ. Since the subunit vaccines have an essential antigen only and are not the molecular array constituting the

germ, they have less side effects. The DTaP vaccine-pertussis (whooping cough) module is a paradigm of subunit vaccines.

#### Common characteristics of vaccines

Live attenuated vaccines are typically grown in animal cell lines under poor development conditions. The development of an inactivated vaccine involves use of thermal or chemical methods in the beginning, and its mode of action involved in conferring immunity is not fully known. However, the live attenuated or execution of entire organism-supported vaccines have shown a lot of success in the control and inhibition of severe transmittable diseases in human, including animal infectious cattle plague, classic swine fever, equine infectious anaemia, measles, mumps, polio, rubella, smallpox, and so on. In recent times, the use of LAVs, subunit and peptide based vaccines have become possible because of progressive technologies in molecular biology. LAVs are based on the mechanism of action associated with the immune response. Inactivated vaccines, based on antibodies have been mostly used to prevent and manage microbial infectious diseases. LAVs introduce stronger cell immune responses that are decisive to remove several intracellular viral pathogens. However, these pathogens sometimes bypass inactivated vaccines9d by mutating peripheral antigens. On the other hand, subunit and peptide based vaccines are less efficient in drawing a strong CD8<sup>+</sup> immune response.

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Progressive vaccination involves the use of non-viral distributed nucleic acid-supported vaccines, which imitate live microorganism infection- or immunization. This leads to T-helper cellular immune responses. In addition, this vaccine development set is harmless and consumes less. It does not require extreme infectious organisms, so is safe from infectivity through live transmittable agents and the discharge of harmful pathogenic organisms. These vaccines fill the gap between a virus outbreak and design of a desirable vaccine, wherein they are classified as DNA/RNA based pentose-carbon sugar motif. The remarkable growth of RNA-associated vaccines resulted in the growth of mRNA based vaccines. It is quite significant to note that mRNA vaccines provide many valuable benefits when compared to viral vectored- and DNA vaccines. 10

# Proceedings of vaccination against viral diseases

In 1970, to evade the prevalent spread of foot and mouth disease, scientists discovered a vaccine using a single protein from the virus. Despite their achievements in virology, in particular vaccine studies and their development, the lack of understanding of immunological mechanisms of action during induced defensive immunity, has prevented the use of existing vaccines during global pandemic outbreaks of related diseases with similar viral pathogens. The immune system protects against various pathogens, including distinctive units of Thelper cells that are useful to protect against various unusual pathogens. Besides, the follicular T-helper cells (TFH cells) generate interleukins (ILs) and support the partition of B cells (they are lymphocytes, and take part in the humoral immune response) and generation of Memory B cells. Furthermore, Memory T cells can be sub classified as CD4<sup>+</sup>/CD8<sup>+</sup> T cells (Cluster of Differentiation) and their functionalities are (a) central memory and (b) effectors memory, which provide various responses upon vaccination against different pathogens.11 Different vaccines attempted for identical pathogens depend on the perceptions of the scientist, 12 many healthcare professionals do not pay sufficient attention to vaccines, which may result in uncertainty in their efficacy, side effects and toxicity. Table 2 lists the various vaccines discovered that are available in the current market for protection against viral infections. They are listed on the basis of various factors, 13,14 including the apparent protection level, plausible mechanism of action, possibility of usage for other diseases. 15,16

#### Mode of action stimulated by vaccines

Normally antibodies prevent/minimize infections from extracellular pathogens:

- (a) use enzymatic active sites to fuse to toxins to break their diffusion;
  - (b) prophylactic action preventing viral replication;
  - (c) facilitate opsonophagocytosis of extracellular bacteria;
  - (d) inducing the complement cascade.

CD8<sup>+</sup> T cells do not inhibit infection but work to minimize, regulate, and remove intracellular pathogens through:

- (a) direct destruction of infected cells (discharge of perforin a pore forming cytolytic protein present in cytotoxic T lymphocytes (CTLs) and natural killer (NK) cells), granzyme (serine proteases delivered through cytoplasmic granules inside cytotoxic T cells and NK cells);
  - (b) destroy infected cells with antimicrobial cytokine release.
- CD4<sup>+</sup> T cells do not inhibit infection, however they contribute in the minimization, regulation, and refinement of extra- and intra-cellular pathogens with their control and cytokine-development capabilities. The main examples of CD4<sup>+</sup> T cells are:
- (a) Follicular T-helper (Tfh) cells yielding predominantly interleukin (IL)-21 and providing assistance to B-cells;
- (b) T-helper 1 (Th1) effector cells yield interferon (IFN)- $\gamma$ , (TNF)- $\alpha$ /TNF- $\beta$ , IL-2, and provide a major role in controlling intracellular pathogens *e.g.*, viruses and bacteria such as *M. tuberculosis*;
- (c) Th2 effector cells generate IL-4, IL-5, IL-13, and impact extracellular pathogens such as bacteria and helminths;
- (d) Th9 effector cells generate IL-9 and also defend against extracellular pathogens;
- (e) Th17 effector cells generate IL-17, IL-22, and IL-26 and participate in mucosal protection (for example against, *S. pneumoniae, B. pertussis, M. tuberculosis*).

#### Main effectors of vaccine responses

Vaccines prevent disease by inducing effector modes of action in cells/molecules to reduce the development of pathogens and deactivate their toxic effects. Vaccine-stimulating immune effectors are resourceful antibodies generated by B lymphocytes' ability to interact to a particular toxin/pathogen. A pictographic representation of vaccine immunological function is shown in Fig. 2.

**Isotype switching.** Control of immunoglobulin (Ig) expression and production from IgM yielding IgG, IgA, or IgE that ensues through B-cell differentiation by DNA recombination.

**Marginal zone.** The zone between the spleen's red pulp and white pulp is known as marginal zone. Its main function is to catch particulate antigens from the circulation and distribute them to lymphocytes.

**Pattern recognition receptors.** These germline-encoded receptors sense the existence of infection through the identification of pathogenic microbe molecular arrays, and stimulate innate immune responses.

Regulatory T cells-t. T cells secrete cytokines (IL-10, transforming growth factor [TGF]- $\beta$ /surface markers) and react to reduce immune system response through different modes of action, this sustains immune homeostasis and tolerance to self-antigens.

**Resident memory T cells.** Effector memory T cells exist in particular tissues (such as the lungs, gut, and skin) and are an instant and early line of protection against various viral and bacterial pathogens.

**Somatic hypermutation.** This is a process which intercalates unsystematic mutations in the B-cell receptor region (*i.e.*,

Table 2 The various antiviral vaccines available in the last two decades, and their characteristics

| S.   | Generic name                                     | Trade name  | Type   | Mode of action  | Treatment for other diseases   | Year of approval | Def |
|------|--|---|--|---|--|------------------|-----|
| 110. | Generic name                                     | Trade name  | Туре   | Mode of action  | uiseases   | арріочаі         | Kei |
| 1    | AJ vaccines                                      | Picovax IPV vaccine, Danish Medicines Agency  | Inactivated Polio<br>Vaccine (IPV)   | IPV provides serum immunity to each poliovirus (three types) and defence against any ensuing paralysis causative disease (poliomyelitis). The mucosal immunity level in the intestine is less than that afforded by OPV, this difference may slightly show in the pharyngeal mucosal lining. Prevents poliovirus in the nervous system; immunostimulant |  | 2019             | 17  |
| 2    | Grippol plus                                     | Quadrivalent AbbVie   | Polymer supported<br>inactivated influenza<br>vaccine 3-valent, egg<br>derived adjuvanted<br>influenza vaccine,<br>polyoxidonium as<br>a distinct adjuvant | Vaccine primes<br>advanced meticulous<br>anti-influenza immunity<br>formation;<br>immunostimulant   | To treat influenza virus<br>infection (A/H1N1 + A/<br>H3N2)  | 2018             | 18  |
| 3    | Ys-On-001  | Yivyka polyinosinic-<br>polycystidylic acid/<br>inactivated rabies virus<br>Yisheng Biopharma | Cancer vaccines anti-<br>neoplastics   | B-cell stimulant,<br>cytokines stimulant,<br>dendritic cell stimulant,<br>immune-modulator,<br>macrophage modulator,<br>natural cell stimulant,<br>regulatory T lymphocyte<br>inhibitor, Th1 cell<br>stimulant  | Cancer vaccines<br>pancreatic cancer   | 2017             | 19  |
| 4    | Adimmune's<br>quadrivalent flu vaccine<br>(4142) | Adimflu-S (QIS)BioB2B<br>Taiwan   | Monovalent vaccine,<br>egg-based inactivated<br>split virus, influenza<br>H1N1 vaccine   | Immunostimulants;<br>inactivated virus from<br>chick embryo culture<br>split virus  | Influenza A (H1N1)<br>vaccine during<br>pregnancy  | 2017             | 20  |
| 5    | Quadrivalent influenza<br>vaccine (J0&BB02)      | Vaxigrip Tetra QIVSanofi<br>Pasteur   | Split influenza virus<br>vaccine   | Induces the humeral antibodies against hemagglutinins   | Potent immunization<br>against the four<br>influenza virus strains (A<br>and B types have two<br>each) | 2017             | 21  |
| 6    | NBP-608  | SKY Zoster  | Attenuated zoster-,<br>varicella-type vaccine  | Inducing both humoral and cellular immune response, which creates an IgG humoral immune response; varicellazoster specifically activate CD4 <sup>+</sup> T-helper and CD8 <sup>+</sup> T-lymphocyte cells   | (VZV), a human<br>neurotropic alpha-   | 2017             | 22  |
| 7    | HBV-ISS (Dynavax)                                | Heplisav-B  | Hepatitis B virus used<br>for vaccine development  | Immune-stimulatory DNA sequence (ISS) ISS- 1018, adjuvant proceeds TLR-9 agonist, which is used for potential prevention and treatment of HIV infection   | Treatment of HBV-, HIV viruses   | 2017             | 23  |

Table 2 (Contd.)

| S.<br>no. | Generic name   | Trade name  | Туре   | Mode of action  | Treatment for other diseases   | Year of approval | Ref. |
|-----------|--|---|--|---|--|------------------|------|
| 8         | GSK-1437173A   | Shingrix GSK  | Non-live, recombinant<br>subunit vaccine;<br>varicella vaccine herpes<br>zoster (Hz/su)<br>2q4vaccine candidates | system AS01B develop  | To prevent herpes zoster (HZ) infection  | 2017             | 24   |
| 9         | DTPa-hepB-IPV-Hib  | EasySix, Hexaxim<br>Panacea Biotech                   | 6 in 1 combination<br>Infanrix hexa whole cell<br>pertussis antigen  | The body produces its own antibodies to   | Diphtheria, DTP,<br>Hepatitis B, Hib, IPV,<br>Pertussis, Polio, Tetanus<br>vaccine | 2017             | 25   |
| 10        | Ad5-EBOVJ07BX02  | ErveboBIB, CanSinoBIO                                 | Glycoprotein<br>recombinant adenovirus<br>type 5 Ebola virus<br>vaccine  | Recombinant   | To treat Ebola virus   | 2017             | 26   |
| 11        | Tetravalent inactivated influenza vaccine (TIV)            | Vaxiflu-4 Zydus Cadila                                | Influenza vaccine  | To increase immunity by<br>generating antibody<br>proteins, which defend<br>against infection caused<br>by the virus present in<br>the vaccine        | viruses-H1N1, H3N2,<br>Type B (Brisbane and  | 2017             | 27   |
| 12        | Flu Vac Qs 2019–20 (4 Yr<br>Up) CD,BL-125408               | Flucelvax Quadrivalent<br>Seqirus, Inc                | Cell based flu vaccines  |   | To prevent influenza A and B viruses   | 2016             | 28   |
| 13        | Quadrivalent influenza<br>vaccine (QIV) (GQM-10)           | Vaxigrip TetraSanofi                                  | Inactivated influenza vaccine (split version)  | To enhance the  | To protect against influenza (flu) viruses   | 2016             | 29   |
| 14        | Live attenuated influenza vaccine (LAIV) J07BB03           | FluMist Quadrivalent<br>Influenza Tetra-<br>Medimmune | Attenuated virus   | LAIVs induce T cell<br>antibody-reactions<br>against the surface<br>protein of HA and NA;<br>LAIVs provide hetero-<br>subtype protection in<br>humans | To prevent common flu; influenza A-(H1N1), (H3N2), and B-viruses                   | 2016             | 30   |
| 15        | Enterovirus type 71<br>vaccine (Vero cells)                | Inlive SINOVAC  | Inactivated EV 71 virus antigen vaccine  | Generate immune<br>reactions against EV71<br>virus  | To prevent hand-foot-<br>mouth disease (HFMD)<br>caused by EV-71                   | 2016             | 31   |
| 16        | Inactivated influenza vaccine                              | Cadiflu-S CBL<br>Biologicals Pvt. Ltd                 | Inactivated vaccine  | To develop immunity against the disease by forming antibodies   | To prevent influenza<br>and protect against its<br>effects                         | 2016             | 32   |
| 17        | Inactivated quadrivalent influenza vaccine (split version) | •   | Influenza quadrivalent<br>vaccine 2020   | Against influenza-A and influenza-B type viruses  |  | 2016             | 33   |
| 18        | Riv-4 (RIV4)   | Flublok-QSonofi Pasteur                               | Quadrivalent<br>recombinant influenza<br>vaccine   | Humoral immune<br>response measured by<br>hemagglutination<br>inhibition antibodies   | To prevent influenza A (H1N1) and B (H3N2)   | 2016             | 34   |
| 19        | Gam Evac Combi   | Combined Vector Based<br>Vaccine                      | Heterologous VSV and<br>adenovirus type-5<br>vectored Ebola virus  | Heterologous prime-<br>boost vaccine humoral<br>immune response, cell<br>facilitated immune<br>response to Ebola                                      | To prevent Ebola viral disease (EVD)   | 2015             | 35   |

| S.<br>no. | Generic name                               | Trade name                                       | Туре  | Mode of action   | Treatment for other diseases  | Year of approval | Ref. |
|-----------|--|--|---|--|---|------------------|------|
| 20        | In32 Activated Sabin<br>polio vaccine      | Ai Bi Wei IMB China                              | Inactivated Poliomyelitis<br>(IPV)  | Inactivated vaccines<br>offer immunity by<br>delivering an inactivated<br>antigen. This vaccine<br>cannot cause disease,<br>thus, it may be<br>administered to an<br>immuno-compromised<br>host                  | To prevent polio in new<br>born babies  | 2015             | 36   |
| 21        | Rotavirus Orv-116E                         | ROTAVAC 5D Vero cells<br>Derived Bharath Biotech |   | Rotarix (vaccine prevents rotavirus infectivity) reproduces in the small intestine and induces immunity; the particular mechanistic action of immunology by rotarix against rotavirus gastroenteritis is unknown | To prevent rotavirus gastroenteritis  | 2015             | 37   |
| 22        | NBP-607-QIV                                | SKY cell flu Quadrivalent<br>SK Bioscience       | Cell cultured<br>quadrivalent inactivated<br>subunit influenza<br>vaccine | •  | To treat influenza virus infections   | 2015             | 38   |
| 23        | GC-3110A, GC-3110B                         | GCflu Quad GC Pharma                             | Quadrivalent influenza virus vaccine                                      | Hemagglutination inhibition antibody response  | To treat influenza virus infections   | 2015             | 39   |
| 24        | Chimerivax-dengue<br>(CYD-TDV)             | Dengvaxia Sanofi<br>Pasteur                      | Live attenuated tetravalent chimeric vaccine                              | Antibody dependent improvement   | To prevent Dengue virus-1, 2, 3 and 4 fever in humans   | 2015             | 40   |
| 25        | H5N1 influenza (avian<br>flu) vaccine (Rx) | Audenz Vn-101 Sonali<br>Pasteur GSK              | Egg based H5N1<br>vaccines, inactivated<br>influenza virus vaccine        | Induces immunity (antibodies), which act against viral HA in the vaccine, after interrupting viral attachment to human respiratory cells, provides immunity to influenza A virus subtype H5N1                    | To treat influenza virus  | 2014             | 41   |
| 26        | 9-Valent HPV vaccine [9v<br>HPV]           | GARDASIL 9                                       | Recombinant virus   | Prevent HPV through<br>humoral immune<br>responses induced by<br>the vaccine   | Prevention of cervical,<br>vuvar, vaginal, anal, or<br>pharyngeal, head and<br>neck cancer; by<br>preventing papilloma<br>virus (HPV) infection | 2014             | 42   |
| 27        | Cell cultured-H5N1<br>vaccine KD-295       | GSK  | Emulsion cell culture<br>influenza HA vaccine<br>(Prototype)              | Antibody titer calculated through hemagglutination inhibition (HI): hightiter virus generation led to suspension of growth of MDCK (Madin–Darby Canine Kidney) and Vero cells in a serum-free distribution       | To prevent influenza A<br>virus H5N1  | 2014             | 43   |

Table 2 (Contd.)

| S.<br>no. | Generic name   | Trade name   | Туре  | Mode of action   | Treatment for other diseases  | Year of approval | Ref. |
|-----------|--|--|---|--|---|------------------|------|
| 28        | Live modified vaccine<br>virus Ankara Ankara-<br>Bavarian Nordic (MVA-<br>BN)-J07BX  | IMVANEX/IMVAMUNE<br>JYNNEOS  | Non-replicating small pox vaccine   | Non-replicating and capable of generating humoral and cellular immune response to orthopox-viruses                                       | To prevent smallpox   | 2013             | 44   |
| 29        | Japanese encephalitis<br>vaccine BBIL/JEV  | JENVAC Bharath Biotech   | Inactivated Vero cell-<br>derived viral vaccine   | JENVAC is sufficient to elicit an immune response  | To protect against<br>Japanese encephalitis<br>virus (JEV)              | 2013             | 45   |
| 30        | Fluzone quadrivalent BL<br>103914/J07B B   | FLUZONE Quadrivalent<br>Sanofi Pasteur Inc.                          | Inactivated quadrivalent<br>influenza virus vaccine<br>type A and type B (split<br>version)                               |  | Prevents influenza<br>diseases, type A and B                            | 2013             | 46   |
| 31        | FLU-Q-QIV flu laval<br>quadrivalent GSK-<br>2282511A   | FluLaval $^{\text{TM}}$ Quadrivalent Glaxo Smith Kline (GSK)         | Influenza virus vaccine   | Vaccines which improve<br>immunity against the<br>viral pathogen leading to<br>influenza: they induce<br>the generation of<br>antibodies | Prevents influenza<br>diseases, type A and B                            | 2013             | 47   |
|           | DTa-IPV-HePB-<br>HibHexavalent vaccine6-<br>in-1 vaccine J07CA09   | Hexyon, Infanrix, Vaxelis  |   | Booster vaccination  | To treat DTaP, hepatitis<br>B, polio, haemophilus<br>influenza diseases | 2013             | 48   |
| 33        | GSK-2282512a BL<br>125127  | Fluarix Quadrivalent<br>(FLU Q-QIV) Glaxo Smith<br>Kline Biologicals | Inactivated influenza<br>vaccine, quadrivalent,<br>seasonal   | Increases immunity for<br>treatment of disease<br>originating from<br>influenza-A subtype and<br>type B viruses                          | triggered by influenza A subtype and type B                             | 2012             | 49   |
| 34        | ChimeriVax™-JE   | IMOJEV Sanofi  | Live attenuated<br>Japanese encephalitis<br>vaccine (JEV),<br>monovalent  | IMOJEV is highly<br>immunogenic and able<br>to induce continuing<br>immunity through both<br>preclinical and clinical<br>trials          | To prevent yellow fever virus   | 2012             | 50   |
| 35        | Prepandemic influenza<br>vaccine (H5N1) J07BB01  | Vepacel  | Inactivated flu strain<br>known as A/Vietnam/<br>1203/2004 (H5N1) whole<br>virion, derived from<br>inactivated Vero cells | Overall the vaccine primes the immune system   | Protect against influenza<br>H5N1 (bird flu)                            | 2012             | 51   |
| 36        | Medi-3250STN: 125020   | FLUMIST Quadrivalent<br>Med-Immune                                   | Influenza vaccine, live<br>attenuated influenza<br>vaccine (LAIV)   | To provide immunity against influenza virus caused by subtypes A and B   | Protects against influenza  | 2012             | 52   |
| 37        | Hepatitis E hecolin<br>(HEV-239)   | Hecolin Xiamen Innovax<br>Biotech                                    | Non enveloped virus<br>with positive sense HEV<br>vaccine, a recombinant<br>vaccine                                       |  | Fights against hepatitis-E virus  | 2012             | 53   |
| 38        | Measles/rubella vaccine  | German measles   | Live attenuated<br>(weakened) viruses   | Immunostimulant,<br>produces antibodies<br>(associated proteins<br>fight and kill measles,<br>mumps, and rubella<br>(MMR) viruses        | Prevent MMR viruses   | 2011             | 54   |
| 39        | Human inactivated<br>influenza vaccine<br>(H1N1) 2009 vaccine.<br>Pandemic influenza<br>strain A/California/7/<br>2009/nyMC X-179A | HNVAC (Bharath<br>Biotech)   | Inactivated Influenza A<br>virus vaccine (H1N1).<br>Cell culture derived<br>vaccine                                       | Active immunization<br>agent, which acts<br>against the influenza A<br>(H1N1) 2009 virus   | Activity against<br>influenza A   | 2010             | 55   |

| Cha       | aracteristic vaccines   |  |   |   |  |                  |      |
|-----------|---|--|---|---|--|------------------|------|
| S.<br>no. | Generic name  | Trade name                                       | Туре  | Mode of action  | Treatment for other diseases   | Year of approval | Ref. |
| 40        | Influenza A virus<br>(H1N1), monovalent<br>vaccine  | 2009 Influenza A<br>(H1N1), Sonali Pasteur       | Monovalent vaccine, adjuvant  | Active immunization for preventing disease caused by influenza virus A (H5N1)   | To prevent influenza A viral disease   | 2010             | 56   |
| 41        | Quadrivalent flu vaccine  | FLUCELVAX  | Cellular influenza<br>vaccine   | Active immunization for preventing influenza subtypes A and B causative diseases  | To defend against four<br>different strains of<br>influenza for both<br>subtypes A and B | 2010             | 57   |
| 42        | Influenza vaccine (whole virion). Inactivated combination   | Vaxiflu-S Fluzone Zydus<br>Cadila Healthcare Ltd | Inactivated influenza vaccine (NZ)  | Active immunization for preventing Vaxiflu-X  | <b>7</b> I   | 2010             | 58   |
| 43        | H1N1 pandemic<br>influenza vaccine H5N1<br>strain of the flu virus A/<br>Vietnam/1203/2004Flu<br>strain/California/07/<br>2009 (H1N1) virus               | Celvapan Baxter<br>International                 | Whole virion in Vero<br>cell-based influenza<br>vaccine, inactivated                  | Motivates the immune<br>system to produce<br>antibodies when<br>exposed to the virus  | To protect against the influenza strains of H5N1 virus                                   | 2009             | 59   |
| 44        | Influenza A virus vaccine<br>H5N1   | Fluval P-H5N1<br>Omninvest                       | Fluart innovative vaccine   | Fluval affords active<br>immunization against<br>four virus strains; two<br>for influenza A subtype,<br>and two for B type  | To prevent influenza<br>a (H5N1)   | 2009             | 60   |
| 45        | Non adjuvant influenza<br>A (H1N1) 2009<br>monovalent vaccine.<br>Influenza<br>hemagglutinin (HA) A/<br>California/07/2009<br>(H1N1) V-like virus         | Panenza Sanofi-Pasteur                           | Non-adjuvanted pandemic vaccine   |   | To prevent influenza A (H5N1)  | 2009             | 61   |
| 46        | Monovalent, cell culture-<br>derived, inactivated<br>subunit influenza<br>vaccine. Produced from<br>A/California/07/2009<br>(H1N1) with adjuvant<br>MF-59 | Celtura  | MF-59 adjuvanted cell<br>cultured derivative A/<br>H1N1 pandemic<br>influenza vaccine | MF-59 induces a strong response in adults and substantially develops the response with growth of HA-specific Tfh (CD4 <sup>+</sup> , ICOS+, CXCR5+, IL-21+) cells         | Induces an immune<br>response for protection<br>against influenza virus                  | 2009             | 62   |
| 47        | Pandemic (H1N1) ASO3<br>adjuvanted influenza<br>vaccine   | Pandremix GSK                                    | Combination of H1N1<br>virus antigen and<br>adjuvant system of<br>H1N1                | Enhances the natural immunity of the body   | To prevent influenza A (H1N1), swine flu viral infections                                | 2009             | 63   |
| 48        | Influenza vaccine<br>(H1N1) flu strain from A/<br>California/7/2009<br>(H1N1) derived strain<br>NYMC-181 J07BB02  | Focetria   | Surface antigen<br>(hemagglutinin and<br>neuraminidase)<br>inactivated adjuvant       | Vaccine acts by priming<br>the immune system  | To protect against<br>influenza type A (H1N1)<br>2009 virus                              | 2009             | 64   |
| 49        | Cell culture-derived<br>adjuvanted influenza<br>virus vaccine (Grippol<br>TC  | Grippol NeoSolvay<br>pharmaceutical AbbVie       | Cell based adjuvanted influenza vaccine   | Activates the endosomal receptor, which leads non-specific activation of the surface TLRs, which induce the intracellular signals contributing to the antiviral mechanism | To prevent influenza virus infections  | 2009             | 65   |
| 50        | Inactivated H5N1<br>influenza (avian flu)<br>vaccine A/Vietnam/1194/<br>2003/(H5N1) RG  | Pan-flu (Sinovac<br>Biotech)                     | Single shot vaccine<br>against H1N1 influenza   | Body reacts by creating antibodies  | To prevent H5N1<br>pandemic influenza  | 2008             | 66   |

Table 2 (Contd.)

| S.<br>no. | Generic name   | Trade name                                | Туре  | Mode of action   | Treatment for other diseases                    | Year of approval | Ref. |
|-----------|--|---|---|--|---|------------------|------|
| 51        | Pandemic influenza<br>vaccine  | Panvax H1N1 vaccine                       | Split virion inactivated vaccine  | Panvax, is an inactive<br>viral part of H1N1, the<br>immune system<br>responds by developing<br>antibodies to the virus<br>particle  | Influenza virus (H1N1),<br>swine flu infections | 2008             | 67   |
| 52        | Pre-pandemic influenza<br>vaccine H5N1 (split<br>virion, adjuvanted,<br>inactivated) AS03-H5N1<br>vaccine A/Vietnam/1194/<br>2004; or A/Indonesia/5/<br>2005 GSK-1562902a  | •   | Split virion, inactivated, adjuvanted; hemagglutinin, antigen, adjuvanted                   | Potential immunization against influenza A   | Influenza (H5N1) virus,<br>swine flu infection  | 2008             | 68   |
| 53        | Influenza vaccine<br>(surface antigen,<br>inactivated, prepared in<br>cell cultures) J07BB02   | Optaflu Flucelvax TETRA<br>Novartis       | Influenza virus surface<br>antigens, hemagglutinin<br>and neuraminidase<br>subunit vaccines |  | Prevent influenza viral infection               | 2008             | 69   |
| 54        | Smallpox (vaccinia)<br>vaccine ACAM2000  | Acam-2000 Sanofi<br>Pasteur Biologics Co. | The vaccine is made<br>from a live virus,<br>vaccinia                                       | Possessing potent<br>immunization against<br>smallpox  | Prevention of smallpox                          | 2007             | 70   |
| 55        | Inactivated quadrivalent influenza vaccine (split virion)  | Afluria Seqirus Pty. Ltd.                 | Quadrivalent split<br>virion, influenza virus<br>hemagglutinin as the<br>active ingredient. | Immunity against 3 (type A (2) and type B (1)) or 4 (type A (2) and type B (2)) microbial strains. Suitable for the annual flu season  | To stop infection caused<br>by influenza virus  | 2007             | 71   |
| 56        | H5N1 avian flu vaccine,<br>avian influenza or bird<br>flu, vaccine is derived<br>from A/Vietnam/1203/<br>2004 influenza virus  | Fluzone Sanofi Pasteur                    | Inactivated influenza<br>virus vaccine  | Provides protection<br>against the H5N1<br>influenza virus<br>stimulating the immune<br>response   | To prevent infection caused by influenza virus  | 2007             | 72   |
| 57        | Adjuvanted H5N1 pre-<br>pandemic vaccine   | Daronix, GSK                              | Second generation pandemic vaccine  | Prepare the body's immune system to prevent a flu epidemic   | Prevent common flu disease                      | 2007             | 73   |
| 58        | Influenza vaccine<br>surface antigen,<br>inactivated, prepared in<br>cell culture: A/<br>California/7/2009<br>(H1N1) pdm09-like<br>strain, A/Switzerland/<br>9715293/2013 (H3N2)-<br>like strain, B/Phuket/<br>3073/2013-like strain | Optaflu                                   | Vaccine containing flu<br>surface antigen   | 1 1  | virus   | 2007             | 74   |
| 59        | Birch pollen allergy<br>vaccine  | Oralgen Birch Pollen<br>ALK-Abello        | Peptide based vaccine   | Suppression of allergic reactions after immunization with fusion protein, which is caused by releasing the immune messenger interleukin-10 (IL-10), an autologous cytokine, functioning to decrease the overacting immune response | To prevent allergy                              | 2007             | 75   |

Table 2 (Contd.)

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| S.<br>no. | Generic name   | Trade name   | Туре   | Mode of action   | Treatment for other diseases  | Year of<br>approval | Ref. |
|-----------|--|--|--|--|---|---------------------|------|
| 60        | Influenza vaccine, split<br>virion, inactivated<br>hemagglutinin of A/<br>California/7/2009<br>Hemagglutinin of A/<br>Victoria/210/2009<br>Hemagglutinin of B/<br>Brisbane/60/2008 | Anflu Sinovac  | Split virion, inactivated                        | Body generates<br>immuno-reactions<br>against influenza virus  | To prevent infection<br>caused by influenza<br>virus  | 2006                | 76   |
| 61        | Rec hepatitis B vaccine  | Supervax   | Hepatitis B vaccine, immunoglobulin (HBIG)       | Active immunization of<br>hepatitis B vaccine<br>induces the immune<br>system to create anti-<br>HBs without an active<br>infection risk | To prevent infection against hepatitis B virus  | 2006                | 77   |
| 62        | Antirabies vaccine   | RABIRIX Bharath<br>Biotech                           | Vero cell based rabies<br>vaccine (PVRV)         | Inactivated virus vaccine boosts immunization against rabies   | To prevent infection against rabies virus   | 2006                | 78   |
| 63        | Live herpes zoster vaccine   | Zostavax Merck                                       | Live attenuated virus vaccine                    | Enhancing VZV –<br>specific immunity<br>against zoster   | Treatment for herpes<br>zoster/postherpetic<br>neuralgia (PHN),<br>a neuropathic pain                         | 2006                | 79   |
| 64        | Rotavirus vaccine, live<br>rotavirus 116E strain<br>derived in Vero cells  | RotaTeq Bharath<br>Biotech                           | Live attenuated, oral, monovalent                | Active immunization for infants from the age of 6 weeks  | To protect against<br>rotavirus gastro-enteritis<br>disease in infants and<br>young children                  | 2006                | 80   |
| 65        | Human papillomavirus<br>quadrivalent (type 6, 11,<br>16, and 18) vaccine (HPV<br>vaccine) J07BM01  | Gardasil, Cervarix Merck                             | Protein subunit<br>quadrivalent,<br>recombinant  | Body's immune system<br>identifies the viral<br>proteins in Gardasil,<br>develops antibodies<br>against them                             | To protect against either<br>two or four or nine types<br>of HPV (cervical, vaginal<br>and vulvar in females) | 2006                | 81   |
| 66        | Rotavirus vaccine with<br>five strains of rotavirus,<br>from both human and<br>animal sources  | ROTARIX  | Live, attenuated, oral                           | Usually to develop<br>immunity against<br>rotavirus-based disease  | Rotarix vaccine assists to prevent this disease in children   | 2005                | 82   |
| 67        | Hepatitis B (r DNZ)<br>vaccine, adjuvanted,<br>absorbed J07BC01  | Fendrix GSK  | Adjuvanted, absorbed, recombinant DNA technology | Vaccine works by priming the immune system   | Prevents hepatitis B virus infection  | 2005                | 83   |
| 68        | MR vaccine freeze-dried, live, attenuated,   | Mearubik, Mitsubishi<br>Tanabe Pharma<br>Corporation |  | •  | To prevent rubella virus  | 2005                | 11   |
| 69        | Inactivated hepatitis A<br>and hepatitis B (rDNA) –<br>HAB adsorbed vaccine  | Bilive Sinovac                                       | Recombinant DNA technology                       | Suboptimal immune response to the vaccine  | Prevents hepatitis B viral infection  | 2005                | 84   |
| 70        | Virosomal influenza<br>vaccine, Invivac<br>influenza vaccine   | Invivac Solvay<br>Pharmaceuticals                    | Adjuvant   | The virosome<br>mechanism remains<br>complex; it is the<br>transporter as well as an<br>immune stimulant                                 | To treat influenza  | 2004                | 85   |
| 71        | Influenza virus (live)<br>(LAIV), inactivated<br>influenza vaccine (IIV)   | Flumist  | Wild-type  | Disease-causing viruses<br>has been attenuated and<br>inactivated, using the<br>influence of heat/<br>chemicals such as<br>formaldehyde  | Protects against<br>infection from influenza<br>viruses   | 2003                | 86   |
| 72        | Hepatitis A and B Vac<br>DB10989, DB11627  | Ambirix  | Vaccine based                                    | B-lymphocytes anti-HBs antibodies  | Immunization against<br>hepatitis A and B viral<br>infectivity  | 2003                | 87   |

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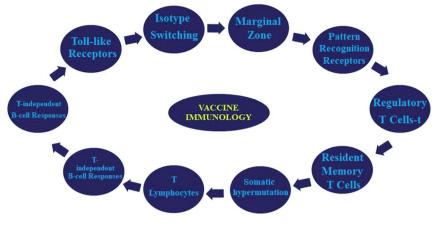


Fig. 2 General features of vaccine immunology.

immunoglobulin) at a particularly high rate throughout the proliferation of B-cells. This mechanistic process occurs as an effect of the cytidine deaminase enzyme and results in antibody diversification.

**T lymphocytes.** Cells that mature in the thymus, and become stimulated in the spleen or nodes if their T-cell receptors interact with an antigen marked with MHC molecules, and they obtain supplementary co-stimulation indications motivating them to kill the infected cell (mainly CD8<sup>+</sup> T cells)/secondary (mainly CD4<sup>+</sup> T cells) roles.

**T-Independent B-cell responses.** This B cell differentiation pathway is mostly triggered by polysaccharides, in the marginal zone and extra-follicular regions of the spleen or nodes. It provides a fast response (days), and generates transient (over months) low affinity antibodies without impacting immune memory.

**T-Dependent B-cell responses.** This B cell differentiation occurs due to protein antgens which recruit T and B cells into germinal centers of the spleen/nodes. It is slower (weeks), but provides enduring stimulation (years) with high affinity antibodies formation and immune memory.

**Toll-like receptors.** A cluster of 10 receptors (TLR1 to TLR10) existing at immune cell external regions, which identify pathogens and trigger characteristic immunity.

The main target of immunization through vaccination is to inhibit specific infections and their unavoidable difficulties. The best vaccine is the one which concomitantly accomplishes the following criteria such as:

- (a) Actively inhibit the infective disease or else minimize the adverse effects of the disease;
- (b) Offer a strong and continuing defence against a specific disease;
- (c) Improves immunity through a minimum quantity of administrations;
- (d) Deliver abundant antigens to afford wide-ranging safety against infection;
  - (e) Never results in side effects, or keeps them to a minimum;
- (f) Remains stable under storage conditions, preferably mild storage conditions, for its shelf-life;

- (g) Can be produced on a huge scale;
- (h) Should be economical and easily available.

Herein we consider the COVID-19 pandemic, the main resolution of vaccination against SARS-CoV-2 are:

- (a) Inhibition of characteristic clinical symptoms so hospitalization is avoided, and reduces severe infectivity;
- (b) Prevention of disease spreading before the corresponding antibodies are produced (sero conversion)
- (c) Producing a strong neutralizing immune response able to link with the viral protein spike (S) that must prevent it from attaching to human cells.

From this perspective, the various immunological response effects which neutralize antibodies and CD8<sup>+</sup> T cells are most significant.

The antibodies of anti-SARS-CoV-2 alert the host organism's immune response to the presence of the virus; such antibodies are immunoglobulins, which are appropriately split into IgA, IgM, IgG, and less frequently IgD. Prior serological antibody model responses to viral infections have usually proven the subsequent sequence of these antibodies resulting from virus infection: the antibodies of IgA are primary, which are followed by IgM, IgG-type continues at high levels for a longer time than the preceding ones (IgA and IgM). For certain viruses, sometimes the antigen (the virus itself) co-occurs with antibodies, particularly with the antibodies of primary IgA and IgM. Further, viruses have a "serological window," i.e., a period between the initial arrival of the antigen (in the blood) and the antibody response, thus phase intervals of infection occur. Eventually, IgG-type antibodies are specific to the novel coronavirus (SARS-CoV-2) and can be examined through chemiluminescence immunological routes, which is an automated laboratory process using enzyme-linked immunosorbent assays with higher arrangement. The examination mostly identifies the body's immune response to SARS CoV-2 infection.

#### Significant characteristics of (corona) virus

Before 2019, there were two pandemics caused in the past two decades by coronavirus; namely SARS during 2002–2003 and Middle East Respiratory Syndrome (MERS) in 2011. According

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to the International Committee on Taxonomy of Viruses (ICTV), the coronaviruses (CoVs) are sub-classified as Othocoronavirinae, which in turn consist of four categories - alpha-CoV, beta-CoV, gamma-CoV, and delta-CoV. The alpha- and beta-CoVs transmit disease in mammals such as bats, pigs, cats and mice. Gamma- and delta-CoVs usually infect birds. In addition, the seven different types of human CoVs - which include HCoV-229E, HCoV-NL63 - belong to alpha-CoVs. The HCoV-OC43, HCoV-HKU1, SARS-, MERS- CoVs, and the current pandemic SARS-CoV-2 belong to beta-CoVs, and the genus of such beta coronavirus occurrences are zoonotic infections. The December 2019 outbreak coronavirus, which leads to the respiratoryassociated syndrome, originated from Wuhan, China, and is called the novel corona virus disease 2019 or nCOVID-19, and its genome is fully sequenced.88 The genetic sequential arrangement of SARS-CoV-2 has an identical genomic array of SARS-/ MERS-CoV.89,90

Initially, the taxonomy of coronaviruses are split in to three groups, on account of genetic and serological interactions, the first set (Group-1) comprised several viruses including the porcine epidemic diarrhoea virus (PEDV), porcine transmissible gastroenteritis virus (TGEV), canine coronavirus (CCoV), feline infectious peritonitis virus (FIPV), the previously identified coronavirus of HCoV-229E, and HCoV-NL63. Whilst the second combination (Group-2) consists of murine hepatitis virus (MHV), bovine coronavirus (BCoV), human coronavirus OC43 (HCoV-OC43), rat sialodacryoadenitis virus (SDAV), porcine hemagglutinating encephalomyelitis virus (PHEV), canine respiratory coronavirus (CRCoV), and equine coronavirus (ECoV). In a similar pattern, the third group (Group-3) contains the avian infectious bronchitis virus (IBV) and Turkey coronavirus (TCoV). Now, the SARS coronaviruses (SARS-CoV) cannot be associated with any of these representative groups, however they possess some similarities along with the second group coronaviruses.91a

The battle between scientists and viral infections is a perpetual process and thus the identification of specific potential drugs with high efficiency and low toxicity is a continuous aim. Globally, this is the first time, that scientific researchers, diplomats, politicians, and capitalists have convened to work towards a common objective. The FDA approved drugs chloroquine and hydroxychloroquine to be utilized in critical illness cases, but clinical practices were still becoming overwhelmed by CoV cases. The suggestion to employ extensive use of these antiviral drugs was not sufficient. Certain polymerase nucleoside/nucleotide inhibitors are promising agents. Favipiravir a selective viral RdRp inhibitor, has been tested in clinical trials against COVID-19. Furthermore, the antiviral drugs lopinavir, ritonavir, remdesivir, nelfinavir, serine based protease inhibitors of nafamostat, camostat, efficient lipid reducing statin, rosovastatin, TNF alpha inhibitors, interleukin 1 receptor antagonists; Janus associated kinases (JAK) as well as monoclonal antibodies, tocilizumab, baricitinib and ruxolitinib etc., along with their combinations and other antiviral components, are also under investigation in clinical studies to combat COVID-19. In the future, researchers are requested to accumulate their results to provide more

knowledge to repurpose significant drugs appropriately, and provide cheap drugs with the minimum toxicity profile. <sup>91b,c</sup>

# The manufacture of SARS-CoV-2 vaccines through various approaches

Vaccines have to be approved based on sufficient evidence. Many scientific researchers have reviewed the current literature and worked towards the development of a specific treatment for COVID-19. Current studies have exposed several beneficial opportunities, a few of them are more established and are in preclinical trials in addition to some in clinical trials. Yaccines endeavor to represent the antibody to the antigen, and they should help the immune system (innate and adaptive immune responses, Fig. 3).

#### Virus vaccines

A number of vaccines are being developed for SARS-CoV-2, including inactivated and weakened, replicating and non-replicating viral vectors, and the tentative use of nucleic acid in the form of DNA and RNA. Now, we can distinguish each vaccine characteristic as follows:

Weakened virus. Viruses cause disease by reproducing in a rapid manner. A weakened virus reproduces inside the human or animal host very poorly thereby decreasing its virulence and disease causing ability. One pharmaceutical company in the USA (Codagenix, New York) is collaborating with an Indian research team (Serum Institute at Pune) for the manufacture of SARS-CoV-2 with changed genetic sequence – making viral proteins with lower potency.

**Inactivated virus.** Chemicals such as formaldehyde, or thermal heating, are applied to inactivate a virus which is then used as a vaccine.

**Viral-vector vaccines.** In this technique other virus types are modified to render them safe and then used as a vaccine. They combine the qualities of DNA vaccines with those of live attenuated vaccines. The viral vector vaccine consists of a live attenuated virus which is genetically modified to carry the DNA encoding protein antigens from a different organism. For example, the virus belonging to measles/adenovirus is genetically modified and thus made safe. Currently there are products for veterinary use but not for human use. There are two types of viral vectors, <sup>94</sup> one is still able to replicate inside the cells, while the other is non-replicating, since the key genes have been rendered inoperative.

Replicating viral vector (weakened measles). One of the approved Ebola vaccines is a typical paradigm for a viral-vector vaccine. Even if replication occurs inside cells, this vaccine is safe and will intensify the immune response.

Non-replicating viral vector (adenovirus variety). In general, viral vectors are genetically transformed to generate defective replications which are termed non-replicating vectors. Numerous viruses like adenovirus, adeno-linked, measles, and human parainfluenza viruses has been extensively utilized as viral vectors. Ultimately, the virus attains an attenuated state in which they can activate the anticipated human immune

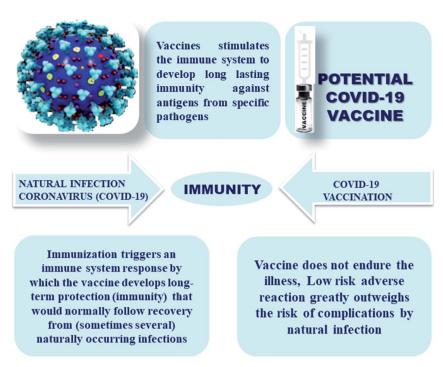


Fig. 3 Pictorial representation of characteristic immunity.

responses, however, they are unable to reproduce in human cells.

Nucleic-acid vaccines. A number of groups are aspiring to utilize gene sequences (based on DNA or RNA) for a coronavirus protein to instantaneously provide an immune response. The majority of these vaccines focus on the spike protein present in the virus.95

Protein-based vaccines. Several researchers have focused on injecting proteins present in coronavirus into the body. Also, protein fragments or shells of protein as copies of the coronavirus' surface could also be utilized.

Protein subunits. Nearly 28t groups are working on vaccines using subunits of viral proteins, predominantly on the spike protein at the receptor binding arena. Vaccines for protecting monkeys against SARS virus infection are available which have not been examined for humans. In fact, to employ such vaccines, adjuvant-immune-stimulation drug molecules should be administered together with the vaccine.96

Virus-like particles. Shells (outer regions) of an empty virus resemble the CoV structure, however, it is non-transmittable due to the absence of the genetic factors. Several research groups are engaged in vaccines based on virus-like particles that can activate a good immune response, however it is quite complex to synthesize such particles, and 70% of the teams directing their research in this way are from industry or private companies rather than academic laboratories. 97,98

Table 3 and 4 provide a comprehensive overview of the current development of vaccines for SARS-CoV-2, their mode of action, their utility and the name of the manufacturer.99 More than 10 vaccines have almost reached the approval stage, while many are in the preclinical stage. Fig. 4 portrays the number of vaccine aspirants which are under clinical (~63 drug aspirants) and preclinical (~172 drug aspirants) stages against nCOVID-19.99a Protein-based techniques seem to be the most popular among the various mechanisms (see https://www.who.int/publications/ m/item/draft-landscape-of-covid-19-candidate-vaccines).

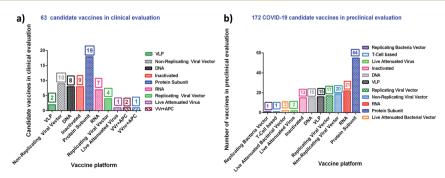


Fig. 4 nCOVID-19 vaccine aspirants at the clinical (63 drug aspirants) and preclinical (172 drug aspirants) stage.

 Table 3
 COVID-19 aspirant developers and their significant details

|     |  | Archard Real Reals Area Security Security  |            | -             | Clinical t  | rials"                               |                      |
|-----|--|--|------------|---------------|-------------|--------------------------------------|----------------------|
| no. | Vaccine aspirant developer   | Vaccine aspirant category  | Phase<br>I | Phase<br>I/II | Phase<br>II | Phase<br>II/III                      | Phase                |
| 1   | Sinovac Research and Development Co., Ltd  | SARS-CoV-2 vaccine (inactivated)   |            |               |             |                                      | 2                    |
|     | Sinopharm + China National Biotec Group<br>Co + Wuhan Institute of Biological Products                         | Inactivated SARS-CoV-2 vaccine (Vero cell)   |            |               |             |                                      | Ø                    |
|     | Sinopharm + China National Biotec Group<br>Co + Beijing Institute of Biological Products                       | Inactivated SARS-CoV-2 vaccine (Vero cell)   |            |               |             |                                      | •                    |
|     | AstraZeneca + University of Oxford   | ChAdOx1-S - (AZD1222) (Covishield)   |            |               |             |                                      | Ø                    |
| 5   | CanSino Biological Inc./Beijing Institute of<br>Biotechnology  | Recombinant novel coronavirus vaccine<br>(Adenovirus type 5 vector)  |            |               |             |                                      | •                    |
| 5   | Gamaleya Research Institute; Health<br>Ministry of the Russian Federation                                      | Gam-COVID-Vac Adeno-based (rAd26-<br>S+rAd5-S)   |            |               |             |                                      | <b>2</b>             |
| ,   | Janssen Pharmaceutical   | Ad26.COV2.S  |            |               |             |                                      | •                    |
| ı   | Novavax  | SARS-CoV-2 rS/Matrix M1-Adjuvant (full<br>length recombinant SARS CoV-2<br>glycoprotein nanoparticle vaccine adjuvanted<br>with Musrix M)    |            |               |             |                                      | ŭ                    |
| ,   | Moderna + National Institute of Allergy and<br>Infectious Diseases (NIAID)                                     | mRNA-1273  |            |               |             |                                      | •                    |
| 10  | Pfizer/BioNTech + Fosun Pharma   | BNT162 (3 LNP-mRNAs)   |            |               |             | <b>3</b>                             |                      |
| _   |  | Recombinant SARS-CoV-2 vaccine (CHO  |            |               |             | _ ≝                                  | _                    |
| 11  | Anhui Zhifei Longeom Biopharmaceutical +<br>Institute of Microbiology, Chinese Academy<br>of Sciences          | Cell)  |            |               |             |                                      | Ø                    |
| 12  | CureVac AG   | CVnCoV vaccine   |            |               |             |                                      | •                    |
| 3   | Institute of Medical Biology + Chinese<br>Academy of Medical Sciences  | SARS-CoV-2 vaccine (Vero cells)  |            |               |             |                                      | Ø                    |
| 14  | Research Institute for Biological Safety<br>Problems, Rep of Kazakhstan  | QazCovid-in® – COVID-19 inactivated vaccine  |            |               |             |                                      | Ø                    |
| 15  | Inovio Pharmacouticals + International<br>Vaccine Institute + Advaccine (Suzhou)<br>Biopharmacoutical Co., Ltd | INO-4800 + electroporation   |            |               |             |                                      |                      |
| 16  | AnGes + Takara Bio + Osaka University  | AG0301-COVID-19  |            |               |             | •                                    |                      |
| 17  | Cadila Healthcare Ltd.   | nCov vaccine   |            |               |             |                                      | ī.                   |
| 18  | Genexine Consortium  | GX-19  |            |               |             |                                      |                      |
| 9   | Bharat Biotech International Limited   | Whole-Virion Inactivated SARS-CoV-2<br>Vaccine (BBV152)  |            | - 2           |             |                                      |                      |
| 20  | Kentucky Bioprocessing Inc.  | Vaccine (BBV152)  KBP-COVID-19 (RBD-based)   |            | 8             |             |                                      |                      |
| 11  | Sanofi Pasteur + GSK   | SARS-CoV-2 vaccine formulation 1 with<br>adjuvant 1 (S protein (Baculovirus<br>production)   |            | 2             |             |                                      |                      |
| 2   | Arcturus Therapeutics  | ARCT-021   |            |               | •           |                                      |                      |
| 13  | Serum Institute of India + Accelagen Pty   | RBD SARS-CoV-2 HBsAg VLP vaccine   |            | <b>2</b>      | _           |                                      |                      |
| 24  | Shenzhen Kangtai Biological Products Co.,  | Inactivated SARS-CoV-2 vaccine (Vero cell)   |            | _             |             |                                      |                      |
| 25  | Ltd.  ReiThera + Leukocare + Univercells   | GRAd-COV2 (replication defective Simian  | <b>3</b>   |               |             |                                      |                      |
| 26  | Vaxart   | Adenovirus (GRAd) encoding S)  VXA-CoV2-1 Ad5 adjuvanted oral vaccine  |            |               |             |                                      |                      |
| 27  |  | platform<br>MVA-SARS-2-S   | <b>2</b>   |               |             |                                      |                      |
|     | University of Munich (Ludwig-Maximilians)  | MVA-SARS-2-S<br>SCB-2019 + AS03 or CpG 1018 adjuvant   | ◙          | _             |             |                                      |                      |
| 28  | Clover Biopharmaceuticals<br>Inc./GSK/Dynavax  | plus alum adjuvant (native like trimerie<br>subunit spike protein vaccine)   |            |               |             | <b>3</b>                             |                      |
| 29  | Vaxine Pty Ltd. + Medytox  | COVID-19 vaccine   | ☑          |               |             |                                      |                      |
|     | CSL Ltd. + Seqirus + University of<br>Queensland   | MF59 adjuvanted SARS-CoV-2 Sclamp vaccine  | •          | Do<br>aspir   | ant have    | ent of this<br>suspended<br>sosition | vaccine<br>I from th |
| 30  | Medigen Vaccine Biologics + Dynavax +<br>National Institute of Allergy and Infectious<br>Diseases (NIAID)      | MVC-COV1901 (S-2P protein + CpG 1018)  | •          |               |             |                                      |                      |
| 31  | Instituto Finlay de Vacunas  | FINLAY-FR anti-SARS-CoV-2 Vaccine<br>(RBD + adjuvant)  |            |               | •           |                                      |                      |
| 32  | Federal Budgetary Research Institution State<br>Research Center of Virology and<br>Bietechnology "Vector"      | EpiVacCorona (EpiVacCorona vaccine based<br>on peptide antigens for the prevention of<br>COVID-19)  RBD (Baculovirus production expressed in |            | Ø             |             |                                      |                      |
| 33  | West China Hospital + Sichuan University   | Sf9 cells) recombinant SARS-CoV-2 vaccine<br>(Sf9 cell)  |            |               | •           |                                      |                      |
| 34  | University Hospital Tuebingen  | IMP CoVac-1 (SARS-CoV-2 HLA-DR peptides)   | ⊠          |               |             |                                      |                      |
| 35  | COVAXX + United Biomedical Inc   | UB-612 (multitope peptide based S1-RBD-<br>protein based vaccine)  |            |               |             | •                                    |                      |
| 36  | Merck & Co. + Themis + Sharp & Dohme +<br>Institute Pasteur + University of Pittsburgh                         | V591-001 - Measles-vector based (TMV-<br>038)  |            | <b>2</b>      |             |                                      |                      |
| 37  | Jiangsu Provincial Center for Disease<br>Prevention and Control  | DelNS1-2019-nCoV-RBD-OPT1 (Intranasal<br>flu-based-RBD)  |            |               | •           |                                      |                      |
| 17  | Symvivo Corporation  | bucTRL-Spike   | ⊠          |               |             |                                      |                      |
| 18  | ImmunityBio, Inc.  | hAd5-S-Fusion + N-ETSD vaccine   | ■          |               |             |                                      |                      |
| 19  | City of Hope Medical Center + National<br>Cancer Institute   | COH04S1 (MVA-SARS-2-S)   | ■          |               |             |                                      |                      |
| 10  | Cancer Institute  Israel Institute for Biological Research   | rVSV-SARS-CoV-2-S vaccine  |            | ■             |             |                                      |                      |
| i1  | Aivita Biomedical, Inc., National Institute of<br>Health Research and Development, Ministry                    | Dendritic cell vaccine AV-COVID-19. A<br>vaccine consisting of autologous dendritic<br>cells loaded with antigens from SARS-CoV-             |            | •             |             |                                      |                      |
|     | of Health Republic of Indonesia  | 2, with or without GM-CSF  |            |               | _           |                                      |                      |
| 52  | Codagenix/Serum Institute of India  Center for Genetic Engineering and   | COVI-VAC   | ◙          | L             |             |                                      |                      |
| 53  | Bietechnology (CIGB)  Center for Genetic Engineering and   | CIGB-669 (RBD + AgnHB)   |            | •             |             |                                      |                      |
| 14  | Biotechnology (CIGB)   | CIGB-66 (RBD + aluminium hydroxide)  |            |               |             |                                      |                      |
| 15  | Valneva, National Institute for Health<br>Research, United Kingdom<br>Biological E Limited                     | VLA2001<br>BECOV2  |            | E             | H           |                                      |                      |
| 57  | Cellid Co., Ltd.   | AdCLD-CoV19  |            | •             |             |                                      |                      |
| 58  | GeneOne Life Science, Inc.   | GLS-5310   |            | _<br>_        |             |                                      |                      |
| 59  | Nanogen Pharmaceutical Biotechnology   | Recombinant Saru-CoV-2 spike protein.  |            |               |             |                                      |                      |
| 50  | Shienogi   | aluminum adjuvanted  Recombinant protein vaccine S-268019  |            | <u>=</u>      | H           |                                      |                      |
| ~   |  | (using Baculovirus expression vector system)  AdCOVID, Adenovirus-based platform   |            | 2             | _           |                                      |                      |
| _   |  | ,  | <b>B</b>   |               |             |                                      |                      |
| i1. | Altimmune, Inc."   | expresses the receptor-binding domain (RBD) of the SARS-CoV-2 spike protein  |            |               |             |                                      |                      |
| 51  | Altimmune, Inc."  University Medical Center Groningen + Akston Biosciences Inc."                               | expresses the receptor-binding domain<br>(RBD) of the SARS-CoV-2 spike protein<br>SARS-CoV-2-RBD-Fc fusion protein                           | •          | 5             |             |                                      |                      |

<sup>&</sup>lt;sup>a</sup> Vaccine candidate list and relevant information shown according to the World Health Organization (WHO) statistical data for COVID-19 during 2019–2020. Clinical research data (last updated on January 08, 2021) may be variable.

# Comparison between previous vaccines and COVID-19 vaccines

Comparing COVID-19 vaccine novelty may seem challenging, but it is normal for queries to arise on a vaccine's overall potential, safety, toxicity and its side effects *etc.*, 1000 and some of the efficacy, efficiency and safety of the COVID-19 frontrunner vaccines can be determined by comparison with other vaccines, like flu vaccines. For example, Pfizer/BioNTech released their COVID-19 vaccine initially in the UK and USA, and the rest of the world could see their effectiveness. However, prior to this demonstration the three prominent COVID-19 vaccines – Pfizer/BioNTech claims an efficacy of 95%, the Oxford/AstraZeneca provides 70%, and the Moderna is stated as 94.1% efficacy – were compared with other available vaccines such as the flu, polio, and measles, which helped provide their effectiveness and efficiency predictions.

It is important to discuss here that "effectiveness" and "efficacy" are different. Efficacy states how a vaccine performs in ideal lab circumstances, like those present in clinical trials. Whereas, effectiveness means how the vaccine works in normal, non-controlled conditions. For example, in a clinical trial, 90% efficacy refers to 90% lower disease rates in the group getting the vaccine compared with the sample group. But, the members in a group selected for a clinical trial have to be in good health and young and they usually have no underlying health conditions. Besides, medical researchers will not generally consider some demographics in these clinical research studies like children/pregnant women. Thus, once a vaccine is able to prevent disease in clinical trials, we may observe the effectiveness drop when directed to different demographics.

Vaccines do not necessarily need high effectiveness to protect several thousands of lives from disease. For instance, the vaccine for flu<sup>100c</sup> has 40–60% effectiveness according to CDC data. During 2018–19, this vaccine prevented millions of influenza cases and its associated illness, but determining the exact effectiveness rate is challenging. Dosages can also increase effectiveness for some vaccines. The two doses of a vaccine can give a protection boost, nevertheless this advantage is sometimes limited to only certain groups like children/organ transplanted people. The booster dosage may not provide an advantage in people aged 65 years.

Through comparing vaccines, like the ones for polio and measles, we see heavy dosages are needed to realize effectiveness.

Polio vaccines<sup>100d</sup> should be up to 100% effective. According to the CDC, "Two inactivated polio vaccine (IPV) dosages have 90% effectiveness; three dosages are 99–100% effective." The IPV vaccine prevents poliomyelitis (poliovirus), which can activate infection in the brain and spinal cord leading to paralysis.

The MMR vaccine<sup>100e</sup> defends against measles, mumps, and rubella, which tends to have up to 97% effectiveness at inhibiting measles once directed in two dosages. A single dose is around 93% effective, as reported by the CDC. They suggest to give the initial dose at "12–15 months of age, followed by the second dose at 4–6 years."

Table 4 Range of SARS-CoV-2 vaccine aspirants and their representative data<sup>16a,b,99c</sup>

| ΙD |         | Vaccine sector description         | Vaccine category and composition                                   |   | Producers  | Clinical phases   | Vaccine utility   | Ref         |
|----|---------|------------------------------------|--|---|--|---|---|-------------|
| ш  | acronym | description                        | and composition  | vaccination   | Producers  | Cliffical phases  | ioi ottiei viruses  | Re          |
| 1  | IV      | Inactivated virus                  | CoronaVac,<br>PiCoVaccine,<br>based on vaccine<br>cultured in Vero | Based on an inactivated pathogen, body generates a varied immune response against several viral antigens, producing neutralizing antibodies   | Sinovac R&D Co. Ltd  | Phase III NCT04456595, Phase 1/2 NCT04383574, NCT04352608   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 101         |
| 2  | IV      | Inactivated virus                  |  | Vaccine from non-<br>living viral particles,<br>bacteria, and other<br>pathogens which are<br>developed in<br>a culture medium.<br>No potential for<br>infection, but<br>induces an immune<br>system response | Sinopharm + WIBP   | Phase 3, Phase-I/II/III<br>ChiCTR2000031809   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 102         |
| 3  | IV      | Inactivated virus                  | similar to virus   | The foremost vaccine, does not exhibit any adverse side effects with favored immunogenicity and safety; also an inactivated new crown vaccine which completely neutralizes the antibodies in 28 days          |  | Phase 3 ChiCTR2000032459  | Vaccine being<br>developed for<br>treatment of<br>similar viruses | 92 <i>d</i> |
| 4  | VVnr    | Viral vector (non-<br>replicating) | Covishield<br>ChAdOx1-S-<br>(AZD1222)                              | Adenovirus vector<br>based on<br>chimpanzee<br>adenovirus   |  | Phase 3 ISRCTN89951424,<br>Phase2b/3 2020-001228-32,<br>Phase 1/2<br>PACTR2020069221651322020-<br>001072-15 | MERS, influenza,<br>TB,<br>Chikungunya,<br>Zika, MenB,<br>plague  | 103         |
| 5  |         | Viral vector (non-<br>replicating) | Ad5-nCoV<br>recombinant<br>vaccine for CoV                         | Recombinant<br>adenovirus type 5<br>vector based vaccine<br>aspirant, which is<br>genetically modified<br>with replication-<br>deficient groups,<br>mimics SARS-CoV-2<br>spike protein                        | Beijing<br>Biotechnology<br>Institute/CanSino<br>Biological Inc.             |   | EBOV (Ebola<br>virus)   | 104         |
| 6  |         | Viral vector (non-<br>replicating) | adeno-based (rad26-S+rAd5-S),                                      | Develop immunity<br>against the<br>coronavirus, and<br>strengthens the<br>immune system   | Gamaleya Research<br>Institute; Ministry of<br>Health, Russian<br>Federation | Phase-I/III NCT04436471,<br>NCT04437875   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 105         |
| 7  | VVnr    | Viral vector (non-<br>replicating) |  | Activates specific<br>acquired immunity<br>for Ebola virus  | Janssen<br>Pharmaceutical  | Phase 3 NCT04505722,<br>NCT04614948   | Vaccine being<br>developed for<br>treatment of<br>similar viruses | 106         |

Table 4 (Contd.)

| Vaccine<br>division<br>ID acronym | Vaccine sector<br>description | Vaccine category and composition  |   | Producers   | Clinical phases   | Vaccine utility for other viruses  | Ref |
|-----------------------------------|-------------------------------|---|---|---|---|--|-----|
| 8 PS                              | Protein subunit               | SARS CoV-2 GP<br>nanoparticle<br>based vaccine<br>with Matrix Mas   | Matrix-M, an<br>adjuvant which<br>improves the<br>immune response<br>and induces<br>advanced<br>neutralizing<br>antibodies  | Novavax   | Phase 3 NCT04611802,<br>NCT04583995                                 | Ebola, Lassa,<br>MERS, Nipah,<br>Rift Valley Fever<br>and<br>Chikungunya | 107 |
| 9 RNA                             | RNA based<br>vaccine          | adjuvant) Moderna mRNA- 1273 vaccine, LNP encapsulated cell bank; mRNA, VAX (non- replicating viral vector) | A vaccine with<br>mRNA encapsulated<br>in LNP, encoding for<br>perfusion stabilized<br>spike (S) protein.<br>Host generates an<br>immune response<br>against the spike<br>protein on SARS-<br>CoV-2         | Moderna + NIAID   | Phase 3 NCT04470427, Phase 2 NCT04405076, Phase 1 NCT04283461       | Multiple agents  | 108 |
| 10 RNA                            | mRNA based<br>vaccine         | BNT162 (3 LNP-mRNAs)  | BNT162 contains<br>a nucleoside<br>modified mRNA<br>(modRNA) encoding<br>the viral spike (S)<br>glycoprotein  | BioNTech + Fosun<br>Pharma; Jiangsu<br>Provincial CDC +<br>Pfizer | Phase 2/3 NCT04368728   | To be<br>implemented   | 109 |
| 11 PS                             | Protein subunit               | Recombinant<br>COVID-19<br>vaccine, CHO<br>(Chinese<br>hamster ovary)<br>cell system,<br>prevents CoVs      | Increases rate of new<br>coronavirus<br>neutralizing S<br>protein antibody<br>(IgG) and RBD<br>protein antibody<br>(IgG)  | Anhui Zhifei<br>Longcom<br>Biopharmaceutical +<br>IMCAS           | Phase-I NCT04453852,<br>NCT04445194, Phase 3<br>NCT04646590         | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2             | 110 |
| 12 RNA                            | mRNA based<br>vaccine         | CVnCoV vaccine  | Nucleotides without<br>chemical<br>modifications in the<br>mRNA   | CureVac AG  | NCT04449276 Phase 1,<br>NCT04515147 Phase 2,<br>NCT04674189 Phase 3 | CureVac's<br>vaccine<br>candidate<br>against SARS-<br>CoV-2              | 111 |
| 13 IV                             | Inactivated virus             | Vaccine for SARS-<br>CoV-2 based on<br>Vero cells   | ELISA derived<br>antibodies (IgGs)<br>and neutralizing<br>antibodies, target<br>the spike protein, N<br>protein virion and<br>the specific positive<br>CTL responses<br>against N, S and<br>virion antigens | IMB + CAMS  | Phase 3 NCT04659239, Phase 1/2 NCT04470609, NCT04412538             |  | 112 |
| 14 IV                             | Inactivated virus             | QazCovid-in®-<br>COVID-19<br>inactivated<br>vaccine   | There are specific antibodies that activate upon receiving small dilutions given in the vaccine, they constantly counterbalance the novel coronavirus with a virulence dose of 3000 TCD50                   | RIBSP, Republic of<br>Kazakhstan                                  | Phase 1/2 NCT04530357   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2             | 113 |

Table 4 (Contd.)

| Vaccine<br>division<br>ID acronym | Vaccine sector<br>description | Vaccine category and composition  |   | Producers                                  | Clinical phases   | Vaccine utility<br>for other viruses                                 | Ref. |
|-----------------------------------|-------------------------------|---|---|--|---|--|------|
| 15 DNA                            | DNA based<br>vaccine          | INO-4800 +<br>electroporation<br>vaccine with<br>DNA-plasmid<br>based<br>connecting<br>electroporation<br>device          | DNA plasmids<br>distribute by electro-<br>permeabilization<br>(electro-transfer),<br>a computationally<br>sequenced design to<br>produce a specific<br>immune response  | Inovio<br>Pharmaceuticals +<br>IVI         | Phase 1 NCT04336410,<br>NCT04447781, Phase 2<br>ChiCTR2000040146, Phase 2/<br>3 NCT04642638 | Multiple agents,<br>Cancer, HBV,<br>HIV, HPV, Lassa,<br>Nipah, Zika, | 114  |
| 16 DNA                            | DNA based<br>vaccine          | AG0301-<br>COVID19  | Plasmid DNA vaccine developed through an intradermal gene transfer targeting the S protein, which increases the efficiency of gene expression and antibody production   | AnGes + Takara Bio +<br>Osaka University   | Phase 2/3 NCT04655625,<br>Phase 1/2 NCT04463472,<br>NCT04527081                             | To be<br>implemented   | 115  |
| 17 DNA                            | DNA based vaccine             | nCov vaccine<br>viral vector,<br>membrane<br>protein based<br>vaccine   | Progression of DNA vaccine with the viral membrane protein liable for CoV cell entry; plasmid DNA incorporated into the host cell changes the viral protein, leading to a strong immune response intervened by cellular and humoral immunity. Also used to produce live attenuated recombinant measles viral vector vaccine | Ltd.                                       | Phase 1/2 CTRI/2020/07/<br>026352   | To be implemented  | 116  |
| 18 DNA                            | DNA based<br>vaccine          | GX-19   | Designed to make<br>antigens by adding<br>nucleic acids into<br>the body, the<br>antigens then<br>produce an immune<br>response   | Genexine<br>Consortium                     | Phase-1/2 NCT04445389   | To be<br>implemented   | 117  |
| 19 IV                             | Inactivated virus             | Whole-virion<br>inactivated<br>vaccine<br>(BBV152),<br>Covaxin for<br>SARS-CoV-2,<br>India's trials on<br>nCoV-19 vaccine | Isolated from<br>asymptomatic<br>COVID-19 patient at<br>NIV Pune, India   | Bharat Biotech<br>International<br>Limited | Phase 1/2 NCT04471519,<br>Phase 3 NCT04641481; CTRI/<br>2020/11/028976                      | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2         | 118  |
| 20 PS                             | Protein subunit               | KBP-COVID-19<br>(RBD-based),<br>plant based<br>technology   | A distinctive plant-<br>based vaccine<br>technology for the<br>generation of<br>antigens to various<br>diseases   | KBP., Inc.                                 | Phase 1/2 NCT04473690   | To be implemented  | 119  |

| Vaccine<br>division<br>ID acronym | Vaccine sector<br>description                                       | Vaccine category and composition   |   | Producers                                     | Clinical phases                  | Vaccine utility for other viruses                            | Ref   |
|-----------------------------------|---|--|---|---|----------------------------------|--|-------|
| 21 PS                             | Protein subunit   | Adjuvanted<br>vaccine of SARS-<br>CoV-2<br>formulation<br>spike protein<br>(baculovirus<br>production) | _   | Sanofi Pasteur +<br>Glaxo SmithKline<br>plc.  | Phase 1/2 NCT04537208            |  | 120   |
| 22 RNA                            | RNA based<br>vaccine  | ARCT-021   | Powerful single<br>dose, vaccine is<br>based upon self-<br>replicating mRNA,<br>self-transcribing and<br>replicating RNA<br>along with its liquid-<br>enabling and<br>resolving nucleo-<br>monomer facilitator<br>altered RNA | Arcturus<br>Therapeutics                      | Phase 1/2 NCT0448095             | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2 | 121   |
| 23 VLP                            | Virus like<br>particle  | RBD SARS-CoV-2<br>HBsAg VLP<br>vaccine   | *******   | SII + Accelagen Pty                           | Phase 1/2<br>ACTRN12620000817943 | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2 | 122   |
| 24 VVnr +<br>APC                  | Viral vector (non-<br>replicating) +<br>APC IV<br>inactivated virus | SARS-CoV-2<br>vaccine<br>(inactivated)   | minumzation   | Shenzhen Geno-<br>Immune Medical<br>Institute | Phase 1/2 NCT04276896            | Vaccine for SARS-<br>CoV-2                                   | - 123 |
| 25 VVnr                           | Viral vector (non-<br>replicating)                                  | replication using  | Able to generate<br>immune response<br>(antibodies and T<br>cells)  | ReiThera +<br>Leukocare +<br>Univercells      | Phase 2 ChiCTR2000039462         | Ebola and RSV<br>(respiratory<br>syncytial virus)            | 124   |
| 26 VVnr                           | Viral vector (non-replicating)                                      | VXA-CoV2-1 Ad5<br>adjuvanted oral  | Vaccine mainly provides mucosal immunity, this vital factor targets mucosal pathogens, including the current coronavirus. Stimulates antigenspecific CD4+ and CD8+ T cells at low and high dosage levels                      | Vaxart  | Phase 1 NCT04563702              | Influenza<br>infection H1N1                                  | 125   |

Table 4 (Contd.)

| Vaccine<br>division<br>ID acronym | Vaccine sector<br>description      | Vaccine category and composition  |  | Producers  | Clinical phases   | Vaccine utility for other viruses                            | Ref. |
|-----------------------------------|------------------------------------|---|--|--|---|--|------|
| 27 VVnr                           | Viral vector (non-<br>replicating) |   | Immune responses and neutralizing antibodies directing the S antigen used to defend against infectivity. The double recombinant sMVA-CoV-2 vectors give S-specific CD8 <sup>+</sup> and CD4 <sup>+</sup> T cells to both S and N antigens in Balb/c mice, respectively | (Ludwig-   | Phase 1 NCT04569383   | Infectious<br>diseases and<br>cancer                         | 126  |
| 28 PS                             | Protein subunit                    | *   | used to target the   | Clover<br>Biopharmaceuticals<br>Inc./GSK/Dynavax           | Phase 1 NCT04405908   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2 | 127  |
| 29 PS                             | Protein subunit                    | COVID19 vaccine   | Effective T cells and<br>neutralizing<br>antibody response   | Vaxine Pty Ltd. +<br>Medytox                               | Phase 1 NCT04453852   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2 | 128  |
|                                   | Protein subunit                    | MF59 adjuvanted<br>SARS-CoV-2 S<br>clamp vaccine  | Advanced antibodies<br>are able to neutralize<br>infectivity through<br>live virus in the cell<br>culture  | -  | Phase 1, development was<br>suspended and the candidate<br>vaccine was removed from the<br>summary analysis | •  | ı    |
| 30 PS                             | Protein subunit                    | MVC-COV1901<br>(S-2P protein +<br>CpG 1018)   | High titer of<br>neutralizing<br>antibodies are<br>prompted against<br>pseudo type novel<br>CoV in sera of<br>immunized mice   | Medigen Vaccine<br>Biologics Corp +<br>NIAID + Dynavax     | Phase 1 NCT04487210   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2 | 129  |
| 31 PS                             | Protein subunit                    | FINLAY-FR anti-<br>SARS-CoV-2<br>vaccine (RBD +<br>adjuvant)  | It has the capability<br>to generate<br>a substantial<br>immune reaction   | Instituto Finlay de<br>Vacunas                             | Phase 2 RPCEC00000347,<br>Phase 1/2 RPCEC00000332,<br>Phase 1 RPCEC00000338                                 | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2 | 130  |
| 32 PS                             | Protein subunit                    | EpiVacCorona,<br>based on peptide<br>antigens   | This antigens-based preparation stimulates the immune reaction   | Federal Budgetary<br>Research Institution<br>SRC VB VECTOR | Phase 1/2 NCT04527575   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2 | 131  |
| 33 PS                             | Protein subunit                    | RBD, baculovirus<br>production<br>expressed in Sf9<br>cells,<br>recombinant<br>SARS-CoV-2<br>vaccine (Sf9 cell) |  | WCH, Sichuan<br>University                                 | Phase 2 NCT04640402,<br>ChiCTR2000039994, Phase 1<br>NCT04530656  | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2 | 132  |
| 34 PS                             | Protein subunit                    | IMP CoVac-1<br>(SARS-CoV-2<br>HLA-DR<br>peptides)   |  | University Hospital<br>Tuebingen (UKT)                     | Phase 1   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2 | 133  |

Table 4 (Contd.)

|                 | Vaccine sector<br>description          | Vaccine category and composition  |  | Producers   | Clinical phases   | Vaccine utility for other viruses                                 | Rei |
|-----------------|--|---|--|---|---|---|-----|
| 35 PS           | Protein subunit                        | UB-612,<br>multitope<br>peptide based S1-<br>RBD-protein<br>based vaccine   | This vaccine has the<br>immunity potential<br>to reduce future<br>pandemic rates   |   | Phase 1 NCT04545749                                     | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 134 |
| 36 VVr          | Viral vector<br>(replicating)          | V591-001-<br>measles-vector<br>based (TMV-038)  | Attains the target immune response in humans   | Merck & Co.,<br>Themis + Sharp &<br>Dohme + Pasteur<br>Institute + Pittsburgh<br>University | Phase 1 CT04498247, Phase1/<br>2 NCT04497298NCT04569786 |   | 135 |
| 37 VVr          | Viral vector<br>(replicating)          | DelNS1-2019-<br>nCoV-RBD-<br>OPT1, intranasal<br>flu-based-RBD  | SARS-CoV-2 RBD protein based vaccine used to motivate cross-reactivity or cross-neutralizing antibodies; moreover it blocks previous CoV pseudovirus and the advanced novel CoV pseudovirus into hACE2 expressing 293 T cells ( $IC_{50} = 4.1$ and $11.63$ $\mu$ g ml <sup>-1</sup> ) | Jiangsu Provincial<br>CDC   | Phase 2 ChiCTR2000039715,<br>Phase 2 ChiCTR2000037782   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 136 |
| 38 RNA          | RNA based<br>vaccine                   | LNP-nCoVsaRNA   | The saRNA vaccine can activate a strong immune response; it is suggested that the IM vaccination antigen is expressed in muscle cells, then moves to antigen presenting cells (APC), representing a cross priming mode of potentiality to prominent CD8 <sup>+</sup> T cells           | London  | Phase 1 ISRCTN17072692                                  | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 137 |
| 39 RNA          | RNA based<br>vaccine                   | SARS-CoV-2<br>mRNA vaccine  | _  | Shulan (Hangzhou)<br>Hospital and CDC at<br>Guangxi Zhuang<br>Autonomous Region             | Phase 1   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 138 |
| 40 VLP          | Virus-like<br>particle                 | Coronavirus-like<br>particle COVID-<br>19 (CoVLP)   | Provides immunity<br>by generating<br>a harmless spike<br>protein member,<br>reduces severe<br>infection effects   | Medicago Inc.   | Phase 2/3   | Vaccine being<br>developed for<br>treatment of<br>similar viruses | 139 |
| 41 VVr +<br>APC | Viral vector<br>(replicating) +<br>APC | COVID-19/aAPC vaccine produced from a variation of lentivirus with immune modulatory genes and viral minigenes to the aAPCs | Artificial antigen-<br>pathogen-specific   |   | Phase 1 NCT04299724                                     | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 140 |

Table 4 (Contd.)

| Vaccine<br>division<br>ID acronym | Vaccine sector<br>description               | Vaccine category and composition   |  | Producers                     | Clinical phases       | Vaccine utility for other viruses                                 | Ref |
|-----------------------------------|---|--|--|-------------------------------|-----------------------|---|-----|
| 42 VVnr +<br>APC                  | Viral vector (non-<br>replicating) +<br>APC | LV-SMENP-DC vaccine. Dendritic cells are modified with lentivirus vectors transmitting COVID-19 minigene SMENP and immune modulatory genes. Cytotoxic T lymphocytes (CTLs) are activated by LV-DC offering COVID-19 specific antigens. | Vaccine directed<br>with antigen-specific<br>CTLs  | Same medical institute        | Phase 1/2 NCT04276896 | Vaccine being<br>developed for<br>treatment of<br>similar viruses | 141 |
| 43 PS                             | Protein subunit                             | AdimrSC-2f<br>(recombinant<br>RBD ±<br>aluminium)  | _  | Adimmune<br>Corporation       | Phase 1 NCT04522089   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 142 |
| 44 DNA                            | DNA based<br>vaccine                        | Covigenix VAX-<br>001  | Induces neutralizing<br>antibody levels and<br>stable T helper cell<br>immunity  | Entos<br>Pharmaceuticals Inc. | Phase 1 NCT04591184   | Vaccine being<br>developed for<br>treatment of<br>SARS-CoV-2      | 143 |
| 45 DNA                            | DNA based<br>vaccine                        | CORVax   | CORVax12 initiates a coordinated vaccine response, able to expose the innate adaptive humoral and cellular arms. These cellular immune responses have the potential to generate a strong antiviral response  | Services                      | Phase 1 NCT04627675   | Vaccine being<br>developed for<br>treatment of<br>similar viruses | 144 |
| 46 RNA                            | RNA based<br>vaccine                        | ChulaCov19<br>mRNA vaccine   | The mRNA based vaccine encodes a protein antigen, while RNA is considered to be unstable; the design and development of this novel vaccine is improving its constancy and protein translation efficacy, so it efficiently enhances immune response | Chulalongkorn<br>University   | Phase 1 NCT04566276   | Vaccine being<br>developed for<br>treatment of<br>similar viruses | 145 |

Table 4 (Contd.)

| Vaccine<br>division<br>ID acronym | Vaccine sector<br>description          | Vaccine category and composition   |  | Producers               | Clinical phases                                  | Vaccine utility for other viruses                                 | Ref. |
|-----------------------------------|--|--|--|-------------------------|--|---|------|
| 47 DNA                            | DNA based<br>vaccine                   | bacTRL-Spike   | Symvivo's bac'TRL gene therapy platform associates an advanced gene-expression plasmid with a probiotic bacterium to resolve restrictions and distribute the DNA vaccine directly to the gut. The bacterium triggers the immune response | Symvivo Corporation     | Phase 1 NCT04334980                              | Vaccine being<br>developed for<br>treatment of<br>similar viruses | 146  |
| 48 VVnr                           | Viral vector (non-<br>replicating)     | hAd5-S-Fusion +<br>N-ETSD vaccine  | A viral spike protein.<br>Virus enters the host<br>cells through the<br>ACE2 receptor with<br>a fusion linker, S is<br>antigenic, and<br>generates an<br>efficient immune<br>response  | •                       | Phase 1 NCT04591717,<br>NCT04710303, NCT04732468 | Vaccine being<br>developed for<br>treatment of<br>similar viruses | 147  |
|                                   | Viral vector (non-<br>replicating)     | COH04S1 (MVA-<br>SARS-2-S)   | Vaccine holds the SARS-CoV-2 spike and nucleocapsid proteins inserted into the MVA platform that can replicate DNA within cells. Thus it generates novel CoV protein expression to trigger host immunity against the virus               | Medical Center          | Phase 1 NCT04639466                              | Vaccine in<br>development   | 148  |
| 50 VVr                            | Viral vector<br>(replicating)          | rVSV-SARS-CoV-<br>2-S vaccine  | rVSV- $\Delta$ G-spike<br>stimulated a safe,<br>efficient and<br>adequate<br>neutralizing<br>antibody.<br>Vaccination leads to<br>lower morbidity,<br>protects lungs, and<br>provides fast viral<br>clearance                            | IIBR, Israel            | Phase 1/2 NCT04608305                            | Vaccine in<br>development   | 149  |
| 51 VVr +<br>APC                   | Viral vector<br>(replicating) +<br>APC | Dendritic cell<br>vaccine AV-<br>COVID-19:<br>contains<br>autologous<br>dendritic cells<br>load with<br>antigens from<br>SARS-CoV-2,<br>with/without<br>GM-CSF | Produced from isolated peripheral blood monocytes from patients. Monocytes are then distinguished into dendritic cells with GM-CSF and IL=4  | Aivita Biomedical, Inc. | Phase 1/2 NCT04690387<br>NCT04386252             | Vaccine in development  | 150  |

Table 4 (Contd.)

| Vaccine<br>division<br>ID acronym | Vaccine sector<br>description | Vaccine category and composition          |   | Producers                     | Clinical phases                   | Vaccine utility<br>for other viruses | Ref. |
|-----------------------------------|-------------------------------|---|---|-------------------------------|-----------------------------------|--------------------------------------|------|
| 52 LAV                            | Live attenuated<br>virus      | COVI-VAC                                  | Codagenix's synthetic attenuated virus engineering (SAVE) platform utilizes synthetic biology to re-code the virus genes into the vaccine. Since this design involves distributing a benign, live attenuated version of SARS-CoV-2 it may stimulate a strong and enduring immune response |                               | Phase 1 NCT04619628               | Vaccine in<br>development            | 151  |
| 53 PS                             | Protein subunit               | CIGB-669 (RBD +<br>AgnHB)                 |   | CIGB                          | Phase 1/2 RPCEC00000345           | Vaccine in<br>development            | 130  |
| 54 PS                             | Protein subunit               | CIGB-66 (RBD +<br>aluminium<br>hydroxide) | Stimulates the generation of antibodies that boost the targeted immune response to the virus  | CIGB                          | Phase 1/2 RPCEC00000346           | Vaccine in<br>development            | 130  |
| 55 IV                             | Inactivated virus             | VLA2001                                   | Vero-cell supported the refined inactivated candidate, based on Valneva's JE vaccine, uses the spike protein normal structural array with CpG 1018 and can stimulate an immune response with a high titer of neutralizing antibodies  | United Kingdom                | Phase 1/2 NCT04671017             | Vaccine in<br>development            | 130  |
| 56 PS                             | Protein subunit               | BECOV2                                    |   | Biological E Limited          | Phase 1/2 CTRI/2020/11/<br>029032 | Vaccine in development               | 130  |
| 57 VVr                            | Viral vector<br>(replicating) | AdCLD-CoV19                               | Immunotherapeutic vaccine based on cells  | Cellid Co., Ltd.              | Phase 1/2 NCT04666012             | Vaccine in development               | 152  |
| 58 DNA                            | DNA based vaccine             | GLS-5310                                  | Nucleic acid-based<br>vaccine platform  | GeneOne Life<br>Science, Inc. | Phase 1/2 NCT04673149             | Vaccine in development               | 152  |

Table 4 (Contd.)

|       | Vaccine sector<br>description | Vaccine category and composition                                      |   | Producers                                  | Clinical phases          | Vaccine utility for other viruses | Ref. |
|-------|-------------------------------|---|---|--|--------------------------|-----------------------------------|------|
| 59 PS | Protein subunit               | Recombinant<br>SARS-CoV-2<br>spike protein,<br>aluminum<br>adjuvanted | Protein subunit containing the recombinant SARS-CoV-2 S1 domain of the spike protein, also integrates either CoVaccine HTTM or alhydrogel. CoVaccine HTTM, a single dose stimulated high titers of antigenbinding IgG. This accelerates the affinity maturation and switching of class to higher values, enhancing cell-induced immunity and virus antibodies | Nanogen<br>Pharmaceutical<br>Biotechnology | Phase 1/2 NCT04683484    | Vaccine in<br>development         | 153  |
| 60 PS | Protein subunit               | protein vaccine<br>S-268019 (based<br>on Baculovirus                  | Vaccine is based on recombinant protein units together with GSK; development of mRNA vaccine by Sanofi, in collaboration with Translate Bio.  Preclinical data revealed that the two immunizations of the mRNA based vaccine stimulated high neutralizing antibody levels equal to those produced in infected humans  |  | Phase 1/2 jRCT2051200092 | Vaccine in<br>development         | 130  |

<sup>&</sup>lt;sup>a</sup> Globally, vaccine producers have the ability to achieve rapid development of highly efficient, safe – minimum toxicity – vaccines.

In order to determine the similarities and differences between previously available vaccines and COVID-19 vaccines, the Bacillus Calmette–Guérin (BCG) vaccine against tuberculosis is provided as one more example; it confers a wide range of immunity against other infections, and it also may minimize the intensity of COVID-19. Moreover one epidemiological analyses provided universal connections between the vaccinations of BCG and COVID-19 mortality: the suggestion of BCG vaccination results on COVID-19 fatality are dominated by socio-economical and demographical variations between countries. In the wake of reducing the manifold distracting factors, many substantial connections between the BCG vaccination and decreased COVID-19 fatalities were perceived. Obviously this investigation emphasizes the necessity for an intrinsic mechanism of studies supporting BCG vaccination

effects on COVID-19, and also for clinical evaluation to control the COVID-19 pandemic. 99h

COVID-19 is a serious respiratory related disease, so the scientific and medicinal community are working hard across the globe to develop a vaccine. Presently, around sixty vaccine candidates are on trial in many countries. Now, nearing twenty candidate vaccines are in phase 3 clinical trials. Gratifyingly, seven vaccines have been approved in many countries.

#### Conclusions

Globally, as of January 2021, there have been around 88 million COVID-19 cases, including nearly 1.9 million fatalities, WHO have registered (see WHO Covid-19 case report). These numbers are expected to increase further, so there is an

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emergency requirement to produce vaccines to protect people. Several candidates vaccines are being developed which are in preclinical and clinical trials. The mechanism of action of these candidates varies significantly, as better knowledge becomes available about this virus, researchers can adapt their design so if one candidate shows low efficacy, another one may be more

Similarly, different demographics may necessitate the need for designing vaccines with different mechanisms of action. The time required for the various stages of clinical trials needs to be shortened (without compromising on the ethics and safety) to achieve the desired goal in a short period of time. 154b Finally, manufacturing such large quantities of the vaccine or vaccines, quickly (without compromising on the quality, purity and efficacy) and distributing them to all parts of the world is another problem which the present planners have not faced before. This requires sufficient manufacturing capacity, availability of raw materials, logistics and several other factors. Also, as all resources are diverted towards SARS-CoV-2, epidemiologists and public health organizations should not lose track of their fight against other viruses.155

The current assemblage of vaccine developers <sup>156a</sup> may reward researchers with increased capability, as numerous basic, transformational and preclinical statistical data have become available during coronavirus exploration. These factors combine together as a substantial promising source for rapid vaccine development.156b

Since, December 2020, some new vaccines have emerged and also have been approved by certain national regulatory authorities for use against COVID-19. Amongst these, as per the universal expectations of the WHO EUL/PQ assessment, the Pfizer vaccine and some other candidates have been approved. More studies on vaccine aspirant efficacy and safety results, 157 including on the Moderna and AstraZeneca vaccine, therein have been widely reported, and AstraZeneca have published their results in well reputed journals. Thus we are eagerly expecting more potential COVID-19 vaccine candidates will be offered to governing authorities for approval in the coming years. Gratifyingly, the growth of many efficient COVID-19 vaccine aspirants under clinical trials is fascinating. When the vaccine candidates are proven to be benign and efficient, they need to be acknowledged by the governing authorities, produced to the necessary standard, and distributed. WHO is collaborating all over the world to assist with the roles in this process, which includes facilitating reasonable access to safe and effective COVID-19 vaccines for everyone.

## Author contributions

KD and SA equally contributed to the collection of data and developed the entire manuscript. MD corrected the manuscript. SG and ST assisted in drafting.

### Conflicts of interest

We have no conflicts of interest.

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#### References

- 1 J. M. Ryan, R. L. Jonathan and V. Olivia, Chem. Rev., 2020, **120**, 3210-3229.
- 2 Y. Tingting, Z. Zifu, G.-S. Adolfo, S. Michael and G. D. G. Bruno, Angew. Chem., Int. Ed., 2020, 59, 18885-18897
- 3 B. Greenwood, Philos. Trans. R. Soc., B, 2014, 369, 20130433.
- 4 K. A. Smith, Front. Immunol., 2011, 2, 1-6.
- 5 A. S. Clem, J. Global Infect. Dis., 2011, 3, 73-78.
- 6 L. B. Nicholson, Essays Biochem., 2016, 60, 275-301.
- 7 J. Lu, G. Lu, S. Tan, J. Xia, H. Xiong, X. Yu, Q. Qi, X. Yu, L. Li, H. Yu, N. Xia, T. Zhang, Y. Xu and J. Lin, Cell Res., 2020, 30, 936-939.
- 8 B. E. Eldred, A. J. Dean, T. M. McGuire and A. L Nash, Med. J. Aust., 2006, 184, 170-175.
- 9 (a) W. Shang, Y. Yang, Y. Rao and X. Rao, npj Vaccines, 2020, 5, 18; (b) C. Edwards, West. J. Med., 2001, 174, 197-198; (c) K. Chumakov, E. Ehrenfeld, E. Wimmer and V. I. Agol, Nat. Rev. Microbiol., 2007, 5, 952-958; (d) H. Wang, Y. Zhang, B. Huang, W. Deng, Y. Quan, W. Wang, W. Xu, Y. Zhao, N. Li, J. Zhang, H. Liang, L. Bao, Y. Xu, L. Ding, W. Zhou, H. Gao, J. Liu, P. Niu, L. Zhao, W. Zhen, H. Fu, S. Yu, Z. Zhang, G. Xu, C. Li, Z. Lou, M. Xu, C. Qin, G. Wu, G. F. Gao, W. Tan and X. Yang, Cell, 2020, 82, 713-721; (e) N. Chauhan, S. Soni, A. Gupta, M. Aslam and U. Jain, J. Med. Virol., 2021, 93, 1967-1982; (f) N. Lurie, M. Saville, R. Hatchett and J. Halton, N. Engl. J. Med., 2020, 382, 1969-1973.
- 10 C. Zhang, G. Maruggi, H. Shan and J. Li, Front. Immunol., 2019, 10, 594-606.
- 11 B. Pulendran and R. Ahmed, Nat. Immunol., 2011, 12, 509-517.
- 12 V. Vetter, G. Denizer, L. R. Friedland, J. Krishnan and M. Shapiro, Ann. Med., 2018, 50, 110-120.
- 13 A. H. Ellebedy and R. Ahmed, The Vaccine Book, 2nd edn, 2016, pp. 283-310. DOI: 10.1016/B978-0-12-802174-3.00015-1.
- 14 E. De Clercq and G. Li, Clin. Microbiol. Rev., 2016, 29, 695-747.
- 15 K. Bharati and S. Vrati, Proc. Natl. Acad. Sci., India, Sect. B, 2012, 82, 181-198.
- 16 (a) https://clinicaltrials.gov; (b) https://www.drugbank.ca; (c) D. E. Speiser and M. F. Bachmann, Vaccines, 2020, 8, 404; (d) S. Tavakol, M. S. Alavijeh and A. M. Seifalian, Curr. Pharm. Des., 2021, 27, 1553-1563; (e) S. P. Kaur and V. Gupta, Virus Res., 2020, 288, 198114.
- 17 J. P. Fox, Rev. Infect. Dis., 1984, 6, S352-S355.
- 18 M. P. Kostinov, A. P. Cherdantsev, A. I. Kuselman, N. K. Akhmatova, A. M. Kostinova, E. Viktorovna,

D. E. Olegonva and A. M. Kostinov, *Hum. Vaccines Immunother.*, 2018, 14, 2971.

19 ClinicalTrials.govIdentifier: NCT03131765, April 27, 2017.

**RSC Advances** 

- 20 (a) J. S. Tregoning, R. F. Russell and E. Kinnear, *Hum. Vaccines Immunother.*, 2018, **14**, 550–564; (b) https://www.prnewswire.com/news-releases/adimmunes-quadrivalent-flu-vaccine-approved-by-tfda-300455062.html.
- 21 Quadrivalent-inactivated-influenza-vaccine-adimmune-corporation, *Reactions Weekly*, 2021, **1837**, 611.
- 22 (*a*) ClinicalTrials.gov Identifier: NCT03120364, April 19, 2017; (*b*) S. J. Lee, H. J. Park, H. L. Ko, J. E. Lee, H. J. Lee, H. Kim and J. H. Nam, *Immun., Inflammation Dis.*, 2020, **8**, 216–227.
- 23 J. J. Y. Sung and H. L. Yuen, Curr. Opin. Mol. Ther., 2006, 8, 150–155.
- 24 ClinicalTrials.govIdentifier: NCT00920218 December 12, 2017.
- 25 K. Mahmood, S. Pelkowski and J. J. Donnelly, *Hum. Vaccines Immunother.*, 2013, **9**, 1894–1902.
- 26 A. K. Mc Elory, R. S. Akondy, C. W. Davis, A. H. Ellebedy, A. K. Mehta, C. S. Kraft, G. M. Lyon, B. S. Ribner, J. Varkey and J. Sidney, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, 112, 4719–4724.
- 27 (*a*) www.adisinsight.springer.com; (*b*) http:// zyduscadila.com/research.
- 28 (a) www.rxlist.com/flucelvax-drug.html; (b) http://europa.eu/en/documents/product-information/flucelvax-tetra-epar-product-information\_en.pdf.
- 29 (a) www.medbroadcast.com/drug/getdrug/vaxigrip; (b) http://products.sanofi.com.au/vaccines/ VAXIGRIP\_NZ\_CMI.pdf.
- 30 (a) B. Brandenburg, W. K. Klaren, C. Tang, M. V. Bujny, H. J. W. Korse, T. Kwaks, J. J. Otterstrom, J. Juraszek, A. M. Van Oijen and R. Vogels, PLoS One, 2013, 8, 80034;
  (b) C. Yamazaki, M. Sugiyama, T. Ohta, H. Hemmi, E. Hamada, I. Sasaki, Y. Fukuda, T. Yano, M. Nobuoka, T. Hirashima, A. Iizuka, K. Sato, T. Tanaka, K. Hoshino and T. Kaisho, J. Immunol., 2013, 190, 296–306.
- 31 H. Juin, H. Lim, J. A. Lee, H. J. Kim, J. W. Kim, J. Y. Hyeon, S. G. Yeo, J. W. Lee, J. S. Yoo, Y. K. Choi and S. W. Lee, *PLoS One*, 2017, 12, 0178259.
- 32 http://www.lybrate.com/amp/medicine/cadiflu-s-vaccine.
- 33 P. T. Boer, J. K. Kelso, N. Halder, T. P. L. Nguyen, J. Modyes, C. Cohen, I. G. Barr, M. J. Postma and G. J. Milne, *Vaccine*, 2018, 836, 997–1007.
- 34 Influenza virus RIV4 vaccine (Flublok Quadrivalent): VAERS reports, *Reactions Weekly*, 2021, 1847, 8.
- 35 I. V. Dolzhikova, O. V. Zubkova, A. I. Tukhvatulin,
  A. S. Dzharullaeva, Z. M. Yukhvatulina,
  D. V. Shcheblyakov, M. M. Shmarov, E. A. Tokarskaya,
  Y. V. Simakova, D. A. Egorova, D. N. Scherbinin,
  I. L. Tutykhina, A. A. Lysenko, A. V. Lostranoy,
  - P. G. Gancheva, T. A. Ozharovskaya, B. V. Belugin,
  - L. V. Kolobukhina, V. B. Pantyukhov,
  - S. I. Syromyatnikova, I. V. Shatokhina, T. V. Sizikova,
  - I. G. Rumyantseva, A. F. Andrus, N. V. Boyarskaya,
  - A. N. Voytyuk, V. F. Babira, S. V. Volchikhina,

- D. A. Kutaev, A. N. Bel'skih, K. V. Zhdanov, S. M. Zakharenko, S. V. Borisevich, D. Y. Logunov, B. S. Naroditsky and A. L. Gintsburg, *Hum. Vaccines Immunother.*, 2017, **13**, 613–620.
- 36 (a) https://www.who.int/teams/health/vaccines-quality/poliomyelitis; (b) A. R. Hinman, J. P. Koplan, W. A. Orenstein, E. W. Brink and B. M. N. Kowane, *Am. J. Public Health*, 1998, **78**, 291–295.
- 37 N. Bhandari, T. R. Chandola, A. Bavdekar, J. John, K. Antony, S. Taneja, N. Goyal, A. Kawade, G. Kang, S. Singh Rathore, S. Juvekar, J. Muliyil, A. Arya, H. Shaikh, V. Abraham, S. Vrati, M. Proschan, R. Kohberger, G. Thiry, R. G. Harry, B. Greenberg, G. Curlin, K. Mohan, G. V. J. A. Harshavardhan, S. Prasad, T. S. Rao, J. Boslego and M. K. Bhan, *Lancet*, 2014, 383, 2136–2143.
- 38 W. S. Choi, J. Y. Noh, J. Y. Song, H. J. Cheong, S. H. Wie, J. S. Lee, J. Lee, S. W. Kim, H. W. Jeong, S. I. Jung, Y. S. Kim, H. J. Woo, K. H. Kim, H. Kim and W. J. Kim, Hum. Vaccines Immunother., 2017, 13, 1653–1660.
- 39 J. Lee, K. Y. Lee, J. H. Kim, C. S. Kim, B. W. Eun, H. M. Kim, D. H. Kim, Y. J. Hong, Y. Y. Choi, D. S. Jo, S. H. Ma and J. H. Kang, J. Korean Med. Sci., 2018, 26, 100.
- 40 J. Arroyo, C. Miller, J. Catalan, G. A Myers, M. S. Ratterree, D. W Trent and T. P Monath, *J. Virol.*, 2004, 78, 12497.
- 41 (*a*) J. Schon, W. R. M. Gorka, M. Schwemmle, M. Beer and D. Hoffmann, *npj Vaccines*, 2020, 5, 40; (*b*) http://www.who.int/influenza/resources/avin\_influenza/en.
- 42 A. Luxembourg, D. Brown, C. Bouchard, A. R. Giuliano, O. E. Lversen, E. A. Joura, M. E. Penny, J. A. Restrepo, J. Romaguera, R. Maansson, E. Moeller, M. Ritter and J. Chen, *Hum. Vaccines Immunother.*, 2015, 11, 1313–1322.
- 43 M. Endo, M. Tanishima, K. Lbarai, K. Hayashida, T. Fukuda, T. Tanable, T. Naruse, Y. Kino and K. Ueda, *Influenza Other Respir. Viruses*, 2020, **14**, 551–563.
- 44 ClinicalTrials.govIdentifier: NCT02977715, September, 23, 2020.
- 45 L. Turtle and T. Solomon, *Nat. Rev. Neurol.*, 2018, **14**, 298–313.
- 46 http://www.who.int/immunizati\_standards/ vaccine\_quality/ fluzone\_sanofi\_pasteur\_product\_insert.pdf.
- 47 (*a*) http://www.fda.gov/media/115785/download; (*b*) http://www.immunizationinfo.com/flulaval-vaccine/amp.
- 48 Y. Y. Syed, Paediatr. Drugs, 2019, 21, 501.
- 49 ClinicalsTrials.govIdentifier: NCT02148211, May 28, 2014.
- 50 (a) J. Arroyo, C. Miller, J. Catalan, G. A Myers, M. S Ratterree,
  D. W. Trent and T. P. Monath, J. Virol., 2004, 78, 12497–12507; (b) M. G. Moloney, A. P. Goncalvez, J. Catalan,
  V. Lecouturier, Y. G. Chambaz, F. Diaz, F. M. Arocho,
  R. C. Gomila, M. C. Bernard, R. Oomen, S. Delagrave,
  N. Burdin, H. Kleanthous, N. Jackson, J. Heinrichs and
  K. V. Pugachev, Sci. Rep., 2018, 8, 13206.
- 51 G. M. Keating, G. L Plosker and K. A. L. Williamson, *Biodrugs*, 2012, **26**, 425–430.
- 52 (a) J. H. C. M. Kreijtz, R. A. M. Fouchier and G. F. Rimmelzwaan, *Virus Res.*, 2011, **162**, 19–30; (b)

S. S. Wong and R. J. Webby, *Clin. Microbiol. Rev.*, 2013, **26**, 476–492.

- 53 X. Wu, P. Chen, H. Lin, X. Hao and Z. Liang, *Hum. Vaccines Immunother.*, 2016, **12**, 2603–2610.
- 54 B. M. Laksono, R. D. Vries, R. J. Verburgh, E. G. Visser, A. deong, P. L. A. Fraaij, W. L. M. Ruijs, D. F. Nieuwenhuijse, H. J. Ham, M. P. G. Koopmans, M. C. van Zelm, A. D. M. E. Osterhaus and R. L. de Swart, *Nat. Commun.*, 2018, 9, 4944.
- 55 N. R. Hegde, D. Kumar, P. P. Rao, P. K. Kumari, Y. Kaushik, R. Ravikrishnan, S. D. Prasad and K. M. Ella, *Vaccine*, 2014, 32, 3636.
- 56 A. Choudhry, S. Singh, S. Khare, A. Rai, D. S. Rawat, R. K. Aggarwal and L. S. Chauhan, *Indian J. Med. Res.*, 2012, 135, 534–537.
- 57 Y. N. Lamb, Drugs, 2019, 79, 1337-1348.
- 58 C. M. Trombetta, E. Gianchecchi and E. Montomoli, *Hum. Vaccines Immunother.*, 2018, **14**, 657–670.
- 59 K. J. Kallen, R. Heidenreich, M. Schnee, B. Petsch, T. Schlake, A. P. Baumh, B. Scheel, S. D. Koch and M. F. Mleczek, *Hum. Vaccines Immunother.*, 2013, 9, 2263– 2276.
- 60 J. S. Tregoning, R. F. Russell and E. Kinnear, *Hum. Vaccines Immunother.*, 2018, **14**, 550–564.
- 61 N. Chotirosniramit, P. Sugandhavesa, L. Aurpibul, Thetket, Kosashunhanan, N. T. Supindham, P. Wongkulab, Q. Kaewpoowat, K. Chaiklang, O. Kaewthip, P. Sroysuwan, Wongthanee, H. Lerdsamran, P. Puthavathana and K. Suparatpinyo, Hum. Vaccines Immunother., 2012, 8, 1854-1859.
- 62 H. Reynales, P. Astudillo, S. de Vallière, C. Hatz, P. Schlagenhauf, B. Rath, P. Velentgas, A. Fariña, V. Sales Carmona and N. Groth, *Vaccine*, 2012, 30, 6436–6443.
- 63 P. Hallberg, H. Smedje, N. Eriksson, H. Kohnke, M. Daniilidou, I. Öhman, Q. Y. Yue, M. Cavalli, C. Wadelius, P. K. E. Magnusson, A. M. Landtblom and M. Wadelius, EBioMedicine, 2019, 40, 595–604.
- 64 N. A. T. van der Maas, S. Godefrooij, P. E. Vermeer-de Bondt, H. E. de Melker and J. Kemmeren, *Hum. Vaccines Immunother.*, 2016, **12**, 1027–1032.
- 65 V. Talayev, I. Zaichenko, M. Svetlova, A. Matveichev, O. Babaykina, E. Voronina and A. Mironov, *Vaccine*, 2020, 38, 6645–6655.
- 66 (a) https://www.sciencedirect.com/topics/medicine-and-dentistry/hong-kong-influenza; (b) W.-S. Ryu, *Molecular Virology of Human Pathogenic Viruses*, 2017.
- 67 J. M. Vernon and T. Nolan, Expert Rev. Vaccines, 2011, 10, 35–43.
- 68 N. J. Carter and G. L. Plosker, BioDrugs, 2008, 22, 279-292.
- 69 I. Manini, A. Domnich, D. Amicizia, S. Rossi, T. Pozzi, R. Gasparini, D. Panatto and E. Montomoli, *Expert Rev. Vaccines*, 2015, 14, 789–804.
- 70 A. Nalca and E. E. Zumbrun, *Drug Des., Dev. Ther.*, 2010, **4**, 71–79.
- 71 (a) https://www.immunizationinfo.com/afluria-vaccine; (b) V. A. Statler, F. R. Albano, J. Airey, D. C. Sawlwin,

- A. Graves, J. V. Matassa, E. H. J. Edelman and G. S. Marshall, *Vaccine*, 2019, 37, 343–351.
- 72 C. A. Robertson, C. A. D. Granados, M. D. Decker, A. Chit, M. Mercer and D. P. Greenberg, Expert Rev. Vaccines, 2016, 15, 1495–1505.
- 73 (a) https://www.ema.europa.eu/en/documents/product-information/covid-19-vaccine moderna-epar-product-information\_en.pdf; (b) N. Pardi, M. J. Hogan, F. W. Porter and D. Weissman, *Nat. Rev. Drug Discovery*, 2018, 17, 261.
- 74 I. Manini, A. Domnich, D. Amicizia, S. Rossi, T. Pozzi, R. Gasparini, D. Panatto and E. Montomoli, *Expert Rev. Vaccines*, 2015, 14, 789–804.
- 75 G. Pauli, T. H. Larsen, S. Rak, F. Horak, E. Pastorello, R. Valenta, A. Purohit, M. Arvidsson, A. Kavina, J. W. Schroeder, N. Mothes, S. Spitzauer, A. Montagut, S. Galvain, M. Melac, C. André, L. K. Poulsen and H. J. Malling, J. Allergy Clin. Immunol., 2008, 122, 951–960.
- 76 M. Margarita, G. Lorenzo and M. J. Fenton, *Chest*, 2013, **143**, 502–510.
- 77 D. T. O. Hagan, L. R. Friedland, E. Hanon and A. M. Didierlaurent, *Curr. Opin. Immunol.*, 2017, 47, 93–102.
- 78 J. P. Mc Gettigan, Expert Rev. Vaccines, 2010, 9, 1177-1186.
- 79 M. Sanford and G. M. Keating, *Drugs Aging*, 2010, 27, 159–176.
- 80 G. M. Keating, BioDrugs, 2016, 30, 243-254.
- 81 D. M. H. Leslie and R. De Mars, *Gynecol. Oncol.*, 2017, **146**, 196–204.
- 82 J. M. Hyser and M. K. Estes, Curr. Opin. Gastroenterol., 2009, 25, 36–43.
- 83 G. L. Roels, Med. Microbiol. Immunol., 2015, 204, 69-78.
- 84 Y. L. Zhao, Y. G. Chen, J. Li, G. X. Han, C. Tian, J. L. Liang, Z. G. Wang, Y. G. Zhu, Z. N. Tian, H. Y. Zhang, Z. J. Wan, Z. L. Liang, S. L. Bi, Z. L. X. Bing and X. Z. Zhi, *Zhonghua Liuxingbingxue Zazhi*, 2004, **25**, 470–473.
- 85 I. A. de Bruijn, J. Nauta, L. Gerez and A. M. Palache, *Vaccine*, 2006, 24, 6629–6631.
- 86 S. S. Wong and R. J. Webby, Clin. Microbiol. Rev., 2013, 26, 476–492.
- 87 B. Jarvis and D. P. Figgitt, Drugs, 2003, 63, 214-215.
- 88 F. Wu, S. Zhao, B. Yu, Y.-M. Chen, W. Wang, Z.-G. Song, Y. Hu, Z.-W. Tao, J.-H. Tian, Y.-Y. Pei, M.-L. Yuan, Y.-L. Zhang, F.-H. Dai, Y. Liu, Q.-M. Wang, J.-J. Zheng, L. Xu, E. C. Holmes and Y.-Z. Zhang, *Nature*, 2020, 579, 265–269.
- 89 (a) H. Wang, X. Li, T. Li, S. Zhang, L. Wang, X. Wu and J. Liu, Eur. J. Clin. Microbiol. Infect. Dis., 2020, 39, 1629–1635; (b)
  N. Wang, J. Shang, S. Jiang and L. Du, Front. Microbiol., 2020, 11, 298; (c) Y. D. Li, W. Y. Chi, J. H. Su, L. Ferrall, C. F. Hung and T. C. Wu, J. Biomed. Sci., 2020, 27, 104.
- 90 P. Zhou, X.-L. Yang, X.-G. Wang, B. Hu, L. Zhang, W. Zhang, H.-R. Si, Y. Zhu, B. Li, C.-L. Huang, H.-D. Chen, J. Chen, Y. Luo, H. Guo, R.-D. Jiang, M.-Q. Liu, Y. Chen, X.-R. Shen, X. Wang, X.-S. Zheng, K. Zhao, Q.-J. Chen, F. Deng, L-L Liu, B. Yan, F.-X. Zhan, Y.-Y. Wang, G.-F. Xiao and Z.-L. Shi, *Nature*, 2020, 579, 270–273.

91 (a) L. Vijgen, E. Keyaerts, E. Moës, I. Thoelen, E. Wollants, P. Lemey, A.-M. Vandamme and M. V. Ranset, J. Virol., 2005, 79, 1595–1604; (b) J. Sultana, S. Crisafulli, F. Gabbay, E. Lynn, S. Shakir and G. Trifirò, Front. Pharmacol., 2020, 11, 588654; (c) S. Dotolo, A. Marabotti, A. Facchiano and R. Tagliaferri, Brief. Bioinform., 2020; (d) Y. Zhou, F. Wang, J. Tang, R. Nussinov and F. Cheng, Lancet Glob. Health, 2020, 2, e667–e676.

- 92 (a) L. Cynthia, Q. Zhou, Y. Li, V. L. V. Garner, S. P. Watkins, L. J. Carter, J. Smoot, A. C. Gregg, A. D. Daniels, S. Jervey and D. Albaiu, ACS Cent. Sci., 2020, 6, 315–331; (b) D. Calina, C. Sarkar, A. L. Arsene, B. Salehi, A. Docea, M. Mondal, M. T. Islam, A. Zaliand and J. S. Rad, Immunol. Res., 2020, 68, 315–324; (c) Y. Dong, T. Dai, Y. Wei, L. Zhang, M. Zheng and F. Zhou, Signal Transduction Targeted Ther., 2020, 5, 237; (d) G. N. A. Rego, M. P. Nucci, A. H. Alves, F. A. Oliveira, L. C. Marti, L. P. Nucci, J. B. Mamani and L. F. Gamarra, Vaccines, 2020, 8, 474; (e) D. D. Li and Q. H. Li, Mil. Med. Res., 2021, 8, 1–15.
- 93 H. Li, Y. Zhou, M. Zhang, H. Wang, Q. Zhao and J. Liu, Antimicrob. Agents Chemother., 2020, 64, e00483.
- 94 (a) R.-G. Marjorie, Curr Opin Biotechnol., 2007, 18, 546-556;
  (b) G. N. A. Rego, M. P. Nucci, A. H. Alves, F. A. Oliveira, L. C. Marti, L. P. Nucci, J. B. Mamani and L. F. Gamarra, Vaccines, 2020, 8, 474.
- 95 W. H. Chen, U. Strych, P. J. Hotez and M. E. Bottazzi, *Curr. Trop. Med. Rep.*, 2020, 7, 61–64.
- 96 W. Ning, S. Jian, J. Shibo and D. Lanying, *Front. Microbiol.*, 2020, **11**, 298.
- 97 R. António, C. M. M. Maria, R. C. Leda, J. T. C. Manuel and M. A. Paula, *Expert Rev. Vaccines*, 2010, 9, 1149–1176.
- 98 L. Jie, U. Laura, S. Erica, R. T. Deborah and V. Raphael, *Viral Immunol.*, 2013, 126–132.
- 99 (a) https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines; (b)
  L. E. Escobara, A. M. Cruz and C. B. Mury, Proc Natl. Acad. Sci., 2020, 117, 27741–27742; (c) https://www.clinicaltrials.gov; (d) https://covid19.who.int/.
- 100 (a) S. Geoghegan, K. P. O'Callaghan and P. A. Offit, Front. Microbiol., 2020, 11, 372; (b) T. H. T. Quach, N. A. Mallis and J. F. Cordero, Matern. Child Health J., 2020, 24, 229–240; (c) L. K. Boerner, ACS Cent. Sci., 2020, 6, 89–92; (d) M. Famulare, C. Selinger, K. A. McCarthy, P. A. Eckhoff and G. C. Couture, PLoS Biol., 2018, 16, e2002468; (e) D. A. Geier, J. K. Kern and M. R. Geier, BMC Pediatr., 2019, 19, 325.
- 101 R. Palacios, E. G. Patiño, R. de Oliveira Piorelli, M. Tilli Reis Pessoa Conde, A. Paula Batista, G. Zeng, Q. Xin, E. G. Kallas, J. Flores, C. F. Ockenhouse and C. Gast, *Trials*, 2020, 21, 853.
- 102 S. Xia, K. Duan, Y. Zhang, D. Zhao, H. Zhang, Z. Xie, X. Li, C. Peng, Y. Zhang, W. Zhang, Y. Yang, W. Chen, X. Gao, W. You, X. Wang, Z. Wang, Z. Shi, Y. Wang, X. Yang, L. Zhang, L. Huang, Q. Wang, J. Lu, Y. Yang, J. Guo, W. Zhou, X. Wan, C. Wu, W. Wang, S. Huang, J. Du, Z. Meng, A. Pan, Z. Yuan, S. Shen, W. Guo and X. Yang, JAMA, J. Am. Med. Assoc., 2020, 324, 951–960.

- 103 M. Oysey, S. A. C. Clemens, S. A. Madhi, et al., Lancet, 2021, 397, 99–111.
- 104 S. Ahamad, S. Branch, S. Harrelson, M. K. Hussain, M. Saquib and S. Khan, *Eur. J. Med. Chem.*, 2021, 209, 112862
- D. Y. Logunov, I. V. Dolzhikova, O. V. Zubkova,
  A. I. Tukhvatulin, D. V. Shcheblyakov, A. S. Dzharullaeva,
  D. M. Grousova, A. S. Erokhova, A. V. Kovyrshina,
  A. G. Botikov, F. M. Izhaeva, O. Popova,
  T. A. Ozharovskaya, I. B. Esmagambetov, I. A. Favorskaya,
  D. I. Zrelkin, D. V. Voronina, D. N. Shcherbinin,
  A. S. Semikhin, Y. V. Simakova, E. A. Tokarskaya,
  N. L. Lubenets, D. A. Egorova, M. M. Shmarov,
  N. A. Nikitenko, L. F. Morozova, E. A. Smolyarchuk,
  E. V. Kryukov, V. F. Babira, S. V. Borisevich,
  B. S. Naroditsky and A. L. Gintsburg, Lancet, 2020, 396, 887.
- 106 J. Sadoff, M. Le Gars, G. Shukarev, D. Heerwegh, C. Truyers, A. M. de Groot, J. Stoop, S. Tete, W. Van Damme, I. Leroux-Roels, P.-J. Berghmans, M. Kimmel, P. Van Damme, J. de Hoon, W. Smith, K. E. Stephenson, S. C. De Rosa, K. W. Cohen, M. J. McElrath, E. Cormier, G. Scheper, D. H. Barouch, J. Hendriks, F. Struyf, M. Douoguih, J. Van Hoof and H. Schuitemaker, N. Engl. J. Med., 2021, 384, 1824–1835.
- 107 E. Tumban, Viruses, 2021, 13, 54.
- L. R. Baden, H. M. El Sahly, B. Essink, K. Kotloff, S. Frey, R. Novak, D. Diemert, S. A. Spector, N. Rouphael, C. B. Creech, J. McGettigan, S. Khetan, N. Segall, J. Solis, A. Brosz, C. Fierro, H. Schwartz, K. Neuzil, L. Corey, P. Gilbert, H. Janes, D. Follmann, M. Marovich, J. Mascola, L. Polakowski, J. Ledgerwood, B. S. Graham, H. Bennett, R. Pajon, C. Knightly, B. Leav, W. Deng, H. Zhou, S. Han, M. Ivarsson, J. Miller and T. Zaks, N. Engl. J. Med., 2021, 384, 403-416.
- 109 F. P. Polack, S. J. Thomas, N. Kitchin, J. Absalon, A. Gurtman, S. Lockhart, J. L. Perez, G. P. Marc, E. D. Moreira, C. Zerbini, R. Bailey, K. A. Swanson, S. Roychoudhury, K. Koury, P. Li, W. V. Kalina, D. Cooper, R. W. Frenck, L. L. Hammitt, Ö. Türeci, H. Nell, A. Schaefer, S. Ünal, D. B. Tresnan, S. Mather, P. R. Dormitzer, U. Şahin, K. U. Jansen and W. C. Gruber, N. Engl. J. Med., 2020, 383, 2603–2615.
- 110 R. Chakraborty and S. Parvez, *Biochem. Pharmacol.*, 2020, 180, 114184.
- 111 G. A. Poland, I. G. Ovsyannikova and R. B. Kennedy, *Lancet*, 2020, **396**, 1595–1606.
- 112 T. Li, T. Zhang, Y. Gu, S. Li and N. Xi, Fundam. Res., 2021, 1, 139–150.
- 113 J. Y. Chung, M. N. Thone and Y. J. Kwon, *Adv. Drug Delivery Rev.*, 2021, **170**, 1–25.
- 114 L. Calzetta, B. L. Ritondo, A. Coppola, M. G. Matera, N. D. Daniele and P. Rogliani, *Vaccines*, 2021, 9, 341.
- 115 C. Corti and G. Curigliano, Ann. Oncol., 2021, 32, 569-571.
- 116 A. K. Yadav, S. Ghosh and A. Kotwal, *J. Mar. Med. Soc.*, 2020, 22, 110.
- 117 E. U. Haq, J. Yu and J. Guo, *Exp. Hematol. Oncol.*, 2020, 9, 24.

Review **RSC Advances** 

- 118 P. Yadav, R. Ella, S. Kumar, D. Patil, S. Mohandas, A. Shete, G. Bhati, G. Sapkal, H. Kaushal, S. Patil, R. Jain, G. R. Deshpande, N. Gupta, K. Agarwal, M. Gokhale, B. Mathapati, S. Metkari, C. Mote, D. D. Patil, B. S. S. Prasad, A. Suryawanshi, M. Kadam, A. Kumar, S. Daigude, S. Gopale, T. Majumdar, D. Mali, P. Sarkale, S. Baradkar, P. Gawande, Y. Joshi, S. Fulari, H. Dighe, S. Sharma, R. Gunjikar, A. Kumar, K. Kalele, V. K. Srinivas, K. Mohan, R. Gangakhedkar, K. Ella, P. Abraham, S. Panda and B. Bhargava, Nat. Commun.,
- 119 M. F. Haidere, Z. A. Ratan, S. Nowroz, S. B. Zaman, Y.-J. Jung, H. Hosseinzadeh and J. Y. Cho, Biomol. Ther., 2021, 29, 1.

2021, 12, 1386-1394.

- 120 Y.-D. Li, W.-Y. Chi, J.-H. Su, L. Ferrall, C.-F. Hung and T.-C. Wu, J. Biomed. Sci., 2020, 27, 104.
- 121 C. Chakraborty, A. R. Sharma, M. Bhattacharya, G. Sharma, R. P. Saha and S.-S. Lee, *Immune Network*, 2021, 21, e5.
- 122 L. Scarabel, M. Guardascione, M. D. Bo and G. Toffoli, Int. J. Infect. Dis., 2021, 104, 441.
- 123 R. Simoneaux and S. L. Shafer, ASA Monitor., 2020, 84, 17-
- 124 T. M. Karpiński, M. Ożarowski, A. S. Mrozikiewicz, H. Wolski and D. Wlodkowic, Theranostics, 2021, 11, 1690.
- 125 Q. Huang and J. Yana, Fundam. Res., 2021, 1, 131.
- 126 S. M. Vrba, N. M. Kirk, M. E. Brisse, Y. Liang and H. Ly, Vaccines, 2020, 8, 680.
- 127 J. G. Liang, D. Su, T. Z. Song, Y. Zeng, W. Huang, J. Wu, R. Xu, P. Luo, X. Yang, X. Zhang, S. Luo, Y. Liang, X. Li, J. Huang, Q. Wang, X. Huang, Q. Xu, M. Luo, A. Huang, D. Luo, C. Zhao, F. Yang, J. B. Han, Y. T. Zheng and P. Liang, Nat. Commun., 2021, 12, 1346.
- 128 R. Chakraborty and S. Parvez, Biochem. Pharmacol., 2020, 180, 114184.
- 129 K. Kucukoglu, N. Faydalı and D. Bul, Med. Chem. Res., 2020, 10, 1.
- 130 S. Kashte, A. Gulbake, S. F. El-Amin and A. Gupta, Hum. Cell, 2021, 34, 711-733.
- 131 N. Chauhan, S. Soni, A. Gupta, M. Aslam and U. Jain, J Med Virol, 2021, 93, 1967.
- 132 L. Scarabel, M. Guardascione, M. D. Bo and G. Toffoli, Int. J. Infect. Dis., 2021, 104, 441.
- 133 I. Jatoi and J. Fan, Biomater. Transl. Med., 2021, 2, 30.
- 134 J. S. Tregoning, E. S. Brown, H. M. Cheeseman, K. E. Flight, S. L. Higham, N.-M. Lemm, B. F. Pierce, D. C. Stirling, Z. Wang and K. M. Pollock, Clin. Exp. Immunol., 2020, 202, 162.

- 135 Z. Strizova, J. Smetanova, J. Bartunkova and T. Milotaa, Int. Arch. Allergy Immunol., 2021, 182, 339.
- 136 M. Galdiero, M. Galdiero, V. Folliero, C. Zannella, A. De Filippis, A. Mali, L. Rinaldi and G. Franci, Eur. Rev. Med. Pharmacol. Sci., 2021, 25, 2752.
- 137 S. P. Kaur and V. Gupta, Virus Res., 2020, 288, 198114.
- 138 B. Stav, B. Tal, P. S. Cederna and R. J. Rohrich, Plast. Reconstr. Surg., 2020, 8, e3206.
- 139 T. M. Belete, Infect. Drug Resist., 2021, 14, 151.
- 140 M. P. Lythgoe and P. Middleton, Trends Pharmacol. Sci., 2020, 41, 363.
- 141 A. Muacevic and J. R. Adler, Cureus, 2020, 12, e8342.
- 142 J. Pollet, W.-H. Chen and U. Strycha, Adv. Drug Delivery Rev., 2021, 170, 71.
- 143 M. M. Silveira, G. M. S. G. Moreira and M. Mendonc, Life Sci., 2021, 267, 118919.
- 144 D. Pushparajah, S. Jimenez, S. Wonga, H. Alattas, N. Nafissi and R. A. Slavcev, Adv. Drug Delivery Rev., 2021, 170, 113.
- 145 J. Kim, Y. Eygeris, M. Gupta and G. Sahaya, Adv. Drug Delivery Rev., 2021, 170, 83.
- 146 M. Bhatta, S. Nandi, S. Dutta and M. K. Saha, Hum. Vaccines Immunother., 2021, 1.
- 147 Y. Li, R. Tenchov, J. Smoot, C. Liu, S. Watkins and Q. Zhou, ACS Cent. Sci., 2021, 7, 512-533.
- 148 K. Lundstrom, Viruses, 2021, 13, 317.
- 149 M. E. Dieterle, D. Haslwanter, R. H. Bortz III, A. S. Herbert, K. Chandran and R. K. Jangra, Cell Host Microbe, 2020, 28, 486.
- 150 M. K. Saadeldin, A. K. Abdel-Aziz and A. Abdellatif, Med. Hypotheses, 2021, 146, 110365.
- 151 P. McIntyre, Y. J. Joo, C. Chiu, K. Flanagan and K. Macartney, Aust. Prescr., 2021, 44, 19.
- 152 J.-H. Yoo, J. Korean Med. Sci., 2021, 36, e54.
- 153 M. Kool, T. Soullie, M. van Nimwegen, M. A. M. Willart, F. Muskens, S. Jung, H. C. Hoogsteden, H. Hammad and B. N. Lambrecht, J. Exp. Med., 2008, 205, 869.
- 154 (a) M. Peiris and G. M. Leung, Lancet, 2020, 396, 1467–1469; (b) D. B. Fogel, Contemp. Clin. Trials Commun., 2018, 156-164.
- 155 K. Goodarz, A. Mohammad, M. S. Hossein, T. Niloufar, R. Sajjad, I. Neda and S. H. N. Seyed, Arch. Acad. Emerg. Med., 2020, 8, e41.
- 156 (a) A. Awadasseid, Y. Wu, Y. Tanaka and W. Zhang, Int. J. Biol. Sci., 2021, 17, 8-19; (b) L. Dai and G. F. Gao, Nat. Rev. Immunol., 2021, 21, 73-82, DOI: 10.1038/s41577-020-00480-0.
- 157 M. C. Castells and E. J. Phillips, N. Engl. J. Med., 2021, 384, 643-649.