

Cite this: *Anal. Methods*, 2021, 13, 34

Detection of RNA viruses from influenza and HIV to Ebola and SARS-CoV-2: a review

Rostislav Bukasov,^a Dina Dossym^a and Olena Filchakova^{*b}

RNA-based viruses likely make up the highest pandemic threat among all known pathogens in about the last 100 years, since the Spanish Flu of 1918 with 50 M deaths up to COVID-19. Nowadays, an efficient and affordable testing strategy for such viruses have become the paramount target for the fields of virology and bioanalytical chemistry. The detection of the viruses (influenza, hepatitis, HIV, Zika, SARS, Ebola, SARS-CoV-2, etc.) and human antibodies to these viruses is described and tabulated in terms of the reported methods of detection, time to results, accuracy and specificity, if they are reported. The review is focused, but not limited to publications in the last decade. Finally, the limits of detection for each representative publication are tabulated by detection methods and discussed. These methods include PCR, lateral flow immunoassays, LAMP-based methods, ELISA, electrochemical methods (e.g., amperometry, voltammetry), fluorescence spectroscopy, AFM, SPR and SERS spectroscopy, silver staining and CRISPR-Cas based methods, bio-barcode detection, and resonance light scattering. The review is likely to be interesting for various scientists, and particularly helpful with information for establishing interdisciplinary research.

Received 12th October 2020
Accepted 10th November 2020

DOI: 10.1039/d0ay01886d

rsc.li/methods

1. Introduction

Viruses are small, nanometer-scale carriers of genetic material. Unable to replicate by themselves, they live inside host cells and use their hosts for replication and assembly. Hence, they are intracellular parasites. In order to exist, viruses must be able to have multiple hosts. Viruses of bacteria, fungi, plants, and animals exist. Furthermore, a single virus can have hosts belonging to different species (e.g., rhinolophid bats and humans). Such viruses can jump from one host species to another. Viruses are not “interested” in killing their host instantaneously because the host’s death would lead to the termination of viral spread. Therefore, the majority of viruses do not cause lethality. Some viruses, however, are infectious agents capable of causing illnesses with lethal outcomes. According to the CDC, the flu caused 12 000–61 000 deaths in the USA each year since 2010.

Depending on the type of nucleic acid they carry, viruses are classified into DNA-containing and RNA-containing viruses. DNA-containing viruses can be single-stranded or double-stranded; they are typically more benign than RNA-viruses and mutagenize to a lesser degree. RNA-viruses can be positive-strand, negative-strand or ambisense. Positive-strand RNA viruses contain genomic RNA, which is identical to viral

mRNA, and can be translated by a host cell. Negative-strand viruses contain RNA in their genome, which is complimentary to mRNA and is used as a template for transcription by RNA-dependent RNA polymerase. Ambisense viruses contain genomic RNA, part of which behaves as positive-strand, while another part possesses negative-strand characteristics. For their replication, RNA-containing viruses rely on polymerases without stringent proofreading abilities. Contrarily, DNA-containing viruses rely on DNA-polymerase, which has proofreading properties. Therefore, the genome of RNA-containing viruses is subject to mutations at a much higher rate. This makes RNA-containing viruses rapidly evolving viruses that outnumber DNA-containing viruses. In addition, RNA-containing viruses are more pathogenic as compared to DNA-containing viruses. Examples of RNA-containing viruses that cause diseases include SARS-coronaviruses, influenza, hepatitis A, C, D, E, dengue, Ebola, HIV and other viruses.

Viruses are a diverse group of infectious agents, with more than 100 families characterized to date. Within the current review, we will focus on the laboratory diagnostics of RNA-containing viruses, which are causative agents of diseases with a high mortality rate or with a tendency for pandemic spread.

A multitude of different diagnostics tools exists. They include the detection of viruses, nucleic acids, and antibodies against viruses. The criteria that diagnostics tests should satisfy include a low limit of detection, high sensitivity, high specificity, high accuracy, and rapid speed of diagnosis. The limit of detection refers to the lowest concentration of analyte detected

^aChemistry Department, SSH, Nazarbayev University, Nur-Sultan, 010000, Kazakhstan

^bBiology Department, SSH, Nazarbayev University, Nur-Sultan, 010000, Kazakhstan.
E-mail: olena.filchakova@nu.edu.kz



by a particular test. Depending on the test, the limit of detection can span a few RNA species in a reaction (for assays that detect nucleic acids) or a few ng mL⁻¹ of analyte (for assays that detect antibodies). The sensitivity refers to the ability of a particular test to detect a virus when the virus is present in a sample, and is expressed in % (100% - % of false negative results). Specificity refers to the ability of a particular test to show a negative result when the virus is absent from a sample, and is expressed in % (100% - % of false positive results). Accuracy denotes the percentage of times at which the performed test results are correct. Thus, a high accuracy indicates low percentages of false-positive and false-negative results. The time it takes for a particular test to produce results depends on the test. Antibodies-detecting tests take less time to produce results than nucleic-acid-based tests (such as quantitative real-time PCR). The present review is focused on comparing different viral diagnostic tests through a quantitative perspective.

Cheng *et al.*'s review in *Analytical and Bioanalytical Chemistry* gave a detailed picture of viral diagnostics in 2009,¹ where the detection was described using a method-to-method approach. Since the publication of that review, new lethal viruses have appeared (Ebola and SARS-coronaviruses), and many tests were developed and validated for their detection and diagnostics. The present review is focused on the quantitative analytical parameters in viral diagnostics as well, but it covers the subject from virus to virus. We selectively covered viruses that are lethal to humans, which contain RNA, and which have at least some pandemic potential. Most attention is directed towards the

COVID-19 pandemic virus. The papers described in this review typically reported the limit of detection or some other analytical parameters describing the test accuracy (selectivity, sensitivity) (Fig. 1). Moreover, the orthogonal approach of viral detection classified by detection method is briefly shown as a table at the end of the review.

2. Detection of influenza viruses

Influenza is a respiratory disease of viral origin. Two types of viruses, influenza A and influenza B, are causative agents² of the disease. Both viruses are negative-strand single-strand RNA viruses. The viral genome is segmented and contains 8 segments. They code for RNA-dependent RNA polymerase (needed to convert negative-strand into positive-strand RNA), haemagglutinin (HA, glycoprotein, required for viral entry), neuraminidase (NA, glycoprotein, needed for viral release), nucleoprotein (NP), matrix protein, membrane protein, nuclear export protein (NEP), and nonstructural proteins (NS). HA and NA proteins have high antigenic variability, contributing to the great diversity of viral subtypes. Influenza is characterized by seasonal epidemics, with an ability to transform into pandemics. The occurrence of pandemics is due to the zoonotic origin of the influenza A virus. It is of particular significance that the influenza A virus can spread among both animals and birds. The pandemics of 1918 and of 2009 (Spanish flu and swine flu, respectively) were caused by the H1N1 viral strain. The pandemic of 1918 resulted in more than 40 million deaths

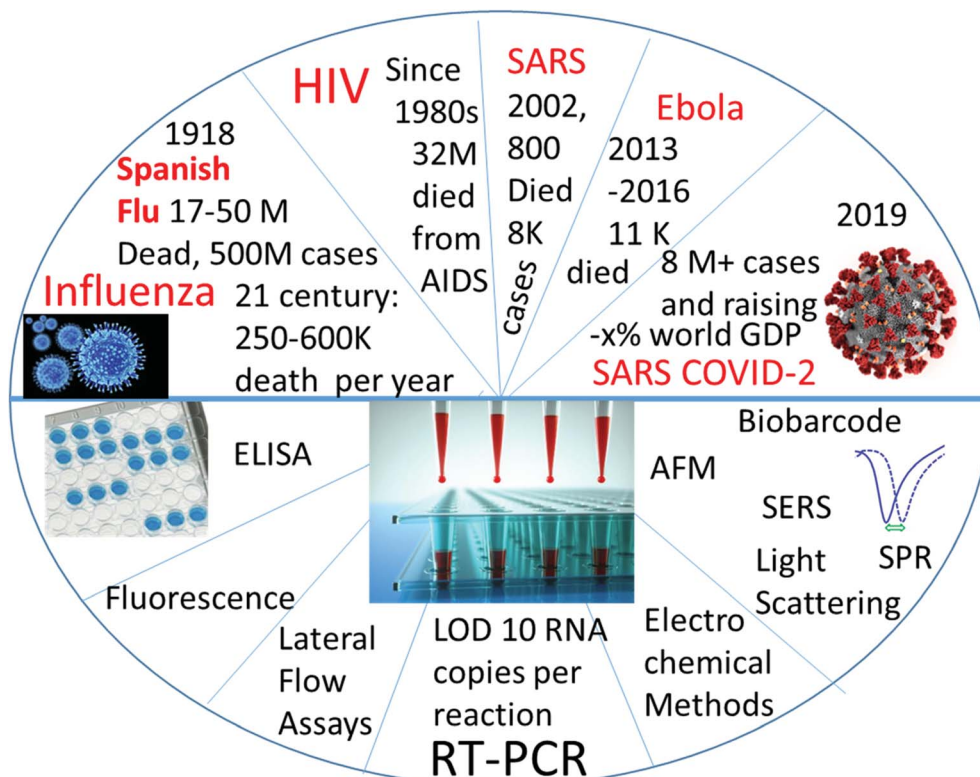


Fig. 1 Graphic Content of the review: detection of RNA-containing pandemic-prone viruses by virus (top half) and by method of detection (bottom half).



worldwide. The 2009 swine flu pandemic was due to the H1N1 influenza A virus, and caused an estimated 201 200 respiratory deaths, together with 83 300 cardiovascular deaths; only 18 500 deaths were laboratory-confirmed.³ The WHO estimates that 290 000 to 650 000 deaths occur annually worldwide due to influenza-associated respiratory diseases.⁴ The CDC estimates that influenza has resulted in between 9 million and 45 million illnesses, between 140 000 and 810 000 hospitalizations, and between 12 000 and 61 000 deaths annually, since 2010 in the USA.⁵ The representative test for the detection of the H5N1 influenza virus is shown in Fig. 2, while different methods are presented in Table 1.

3. Detection of RNA-containing viruses of health concern

HIV/AIDS is prevalent mostly in East and Southern Africa, with a prevalence of 7.4% in the region in 2014, which is over 3 times higher than the prevalence in Western and Central Africa. Eastern Europe and Central Asia are the only regions with a rising HIV prevalence, at 0.8% in 2014. The global prevalence was 0.8% in 2014. The morbidity rate is currently at 1.1 million per year (2015).¹⁵ A variety of methods exist for HIV diagnostics. Two such methods are presented in Fig. 2 and 3. Both methods detect the p24 antigen by colloidal gold immunochromatography, and by ELISA with the use of AFM, respectively. In 2014, 10 000 people were infected with Ebola, with 4922 fatalities. The mortality rate for Ebola can reach 90%.¹⁶

In 2016, more than 85 countries and territories had the Zika virus infection transmitted by mosquitos. Brazil had the

greatest impact, with more than 200 000 cases of the Zika virus disease.¹⁷ The Zika virus had a mortality rate of 8.3% in Brazil.¹⁸

Cases of viral hemorrhagic fever were seen in Zimbabwe, Uganda, the Democratic Republic of Congo, Kenya, and Angola, with a fatality rate of up to 15%.¹⁹

The number of dengue cases reported to the WHO has increased over 15 fold over the last two decades, from 505 430 cases in 2000 to over 2 400 138 in 2010 and 3 312 040 in 2015. Deaths increased from 960 in 2000 to more than 4032 in 2015.²⁰

There are an estimated 1.4 million cases per year of hepatitis A, with 0.5% of mortalities due to viral hepatitis.²¹ Globally, an estimated 71 million people have chronic hepatitis C virus infection. The WHO estimated that approximately 399 000 people died from hepatitis C in 2016, mostly from cirrhosis and hepatocellular carcinoma (primary liver cancer).²² Hepatitis D virus (HDV) globally affects nearly 5% of people who are chronically infected with the hepatitis B virus (HBV). The superinfection of HDV on chronic hepatitis B accelerates the progression to a more severe disease in all ages and in 70–90% of persons.²³ Every year, there are an estimated 20 million HEV infections worldwide. The WHO estimates that hepatitis E caused approximately 44 000 deaths in 2015 (accounting for 3.3% of the mortality due to viral hepatitis).²⁴ The summary of different methods to detect the outlined viruses of health concern is presented in Table 2.

4. Detection of coronaviruses

Coronaviruses are single-stranded, plus-strand, enveloped RNA-containing viruses, with a relatively large genome averaging at 30 kb pairs. Human coronaviruses (HCoV) 229E, NL63, OC43,

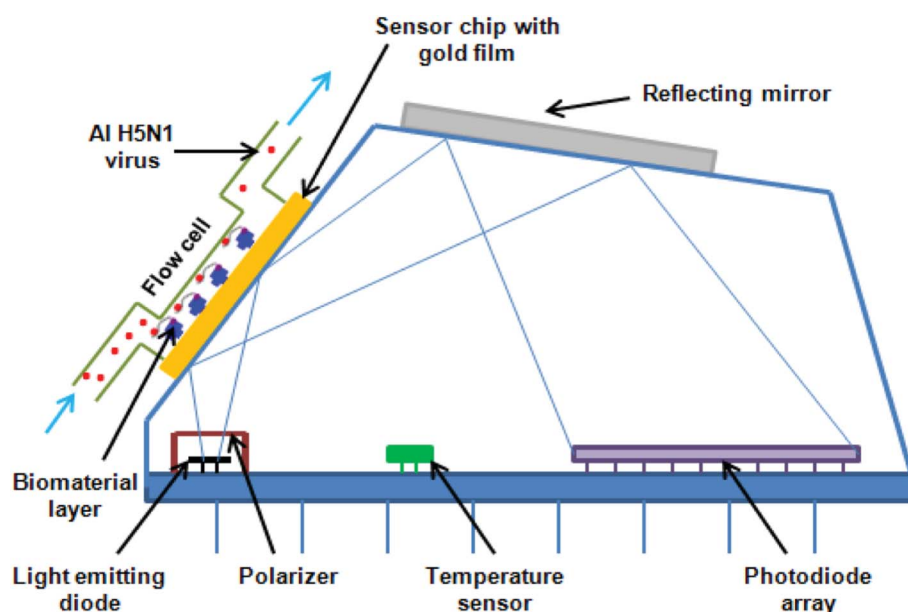


Fig. 2 SPR sensor for the detection of H5N1 avian influenza virus. The sensor is based on the changes of the refractive index of the plasmon as a result of binding the virion to the aptamer. The aptamer is bound to the gold surface of the sensor via biotin–streptavidin linkage. The portable sensor is capable of detecting 0.128 HAU, which is equal to $0.17 \times 10^{3.2}$ ELD₅₀ per mL. The image is reprinted with permission from H. Bai, R. Wang, B. Hargis, H. Lu and Y. Li, A SPR Aptasensor for Detection of Avian Influenza Virus H5N1, *Sensors*, 2012, 12(9), 12506–12518.



and HKU1 are known endemic human coronaviruses that cause mild respiratory infection with such symptoms as rhinorrhea and mild cough. Two other coronaviruses – Severe-Acute Respiratory Syndrome (SARS) and Middle East Respiratory Syndrome (MERS) coronaviruses – are more virulent, and lead to a severe respiratory disease with symptoms similar to influenza. The SARS coronavirus caused a global epidemic in 2002–2003,^{85–87} killing 774 people out of the 8096 infected, and thus having a 9.56% mortality rate. MERS-CoV caused an epidemic in the Middle East that started in 2012.⁸⁸ By January 2020, MERS-CoV had killed 866 people out of the 2519 that were sick, and so it has a 34.4% death rate.⁸⁹

Diagnostic tests that use Polymerase Chain Reaction (PCR) allow for the detection of viral nucleic acid. With DNA-containing viruses, the PCR technique is more straightforward, as it depends on the amplification step of isolated viral DNA. The detection of RNA-containing viruses requires an additional step of converting RNA into DNA by reverse transcription. PCR utilizes synthetic single-stranded DNA primers and probes, and depends on the hybridization. Because primers and/or probes might bind nonspecifically to a region different from the intended one, PCR tests can result in false-positive outcomes. On the other hand, false-negative results might result from improperly collected material. The focus of the present review is to compare the key parameters of different test assays, such as specificity and selectivity, to get a quantitative outlook on the test systems.

There are different modalities of PCR, which are useful in viral detection. Among them are quantitative real-time PCR (qPCR), which allows for rapid detection with the identification

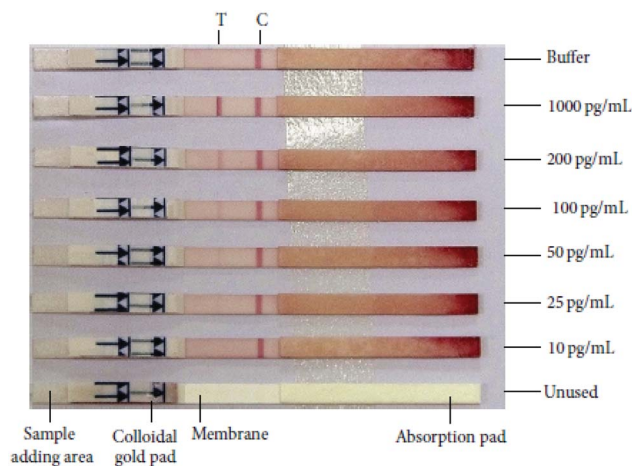


Fig. 3 Colloidal gold immunochromatographic assay to detect the recombinant p24 protein of HIV-1. The reported limit of detection for this method is 25 pg mL⁻¹. It is completed in 20 minutes with an accuracy of 98.03%. The image is reprinted with permission from Ma, Ni *et al.*, Development of Monoclonal Antibodies against HIV-1 p24 Protein and Its Application in Colloidal Gold Immunochromatographic Assay for HIV-1 Detection, *BioMed Research International*, 2016, 2016, 1–6.

of relative amounts of the nucleic acid. The quantitative real time RT-PCR (RT-qPCR) method is an efficient method of diagnostics, which allows for the rapid detection of viral nucleic acid. Many RT-qPCR methods are developed for the detection of coronaviruses. The range of methods for the detection of SARS and MERS coronaviruses is summarized in Table 3.

Table 1 Detection of influenza viruses

#	Method	Target/analyte	Volume	Accuracy	Time for test	Ref.
1	Commercial antigen detection tests and RT-qPCR	Swine-origin influenza virus (S-OIV) and seasonal influenza A (H1N1) isolate: antigens and M genes	N/A	N/A	N/A	6
2	– Rapid antigen test – Direct immunofluorescence (DFA) – R-Mix culture – Respiratory Virus Panel (RVP)	H1N1 influenza A antigen	200 µL of respiratory sample	Rapid antigen: 93.6% DFA: 94.5% R-Mix culture: 100% RVP: 100%	N/A	7
3	SPR aptasensor	Avian influenza virus H5N1 virions	N/A	N/A	1.5 hours	8
4	ELISA	Anti-influenza A virus nucleoprotein antibodies	15 µL of porcine sera	99.3% Sensitivity – 96.6%	N/A	9
5	A double-antigen sandwich ELISA	Antibodies to influenza A viruses	50 µL of serum sample	97.3% Sensitivity – 98%	N/A	10
6	RT-LAMP on an integrated centrifugal disc	Influenza A (subtypes H1, H3, H5, H7, and H9) and influenza B RNA	25 µL	N/A	45 min	11
7	A nanocomposite of AuNPs and polyols with a dual response	Influenza A virions	N/A	N/A	N/A	12
8	Aptamer-based field-effect transistor	H5N1 avian influenza virus hemagglutinin (HA) protein	3 µL	N/A	5 hours	13
9	Magnetic particle spectroscopy	Influenza A virus subtype H1N1 nucleoprotein	100 µL	N/A	10 s	14



Table 2 Detection of different RNA-containing viruses with health concern

#	Method summary	Analyte	Sample volume	Accuracy	Time	Ref.
1	Colloidal gold immunochromatographic assay (GICA)	p24 (viral protein of HIV-1)	75 μ L r-p24 and 75 μ L detector mAb	Specificity – 98.03% (1.96% – false-positive)	20 min	25
2	Chemiluminescent magnetic microparticle-based immunoassay (ARCHITECT HIV Ag/Ab Combo)	HIV-1 p24 antigen	N/A	N/A	N/A	26
3	Fluorescent microparticle enzyme immunoassay (AxSYM HIV Ag/Ab Combo)	HIV p24 antigen	N/A	N/A	N/A	26
4	Enzyme-linked fluorescent assay (VIDAS HIV DUO Quick)	HIV p24 antigen	N/A	N/A	N/A	26
5	Enzyme-linked fluorescent assay (VIDAS HIV DUO Ultra)	HIV p24 antigen	N/A	N/A	120 min	26
6	Quantitative enzyme-linked fluorescent immunoassay (VIDAS HIV p24 II)	HIV p24 antigen	N/A	N/A	N/A	26
7	Nanometer-scale antibody array-based analysis with AFM detection	HIV p24 antigen	1 μ L	N/A	6 hours; 2–3 hours before measurement	27
8	Ultrasensitive capacitive immunosensor functionalized by anti-HIV-1 p24 mAb	HIV p24 antigen	250 μ L	N/A	20 min	28
9	Boosted ELISA based on immune complex dissociation and amplified signal	HIV p24 antigen	100 μ L	N/A	>120 min	29
10	Nanoparticle-based biobarcode amplification assay	HIV p24 antigen	100 μ L	100%	>120 min	30
11	Colorimetric lateral diffusion immunochromatography	Antibody against HIV	Finger prick blood	N/A	3–30 min	31
12	Electrochemical ELISA	Antibody against HIV-1 and HIV-2 peptides	20 μ L	N/A	N/A	32
13	Electrochemical sensor, where antibodies bind to polypeptide epitopes	anti-HIV antibodies	200 μ L	N/A	8 min	33
14	Reverse transcription loop-mediated isothermal amplification (RT-LAMP) products are visualized using a lateral flow immunoassay (LFIA): microfluidic rapid and autonomous analysis device (microRAAD)	HIV-1 RNA	12 μ L of human whole blood	N/A	90 min	34
15	Luminescence assay	Ebola virus oligonucleotide (RNA)	N/A	N/A	2 days	35
16	Reverse transcription loop-mediated isothermal amplification (RT-LAMP)	Ebola RNA	1 μ L of RNA	N/A	1 hour	36
17	ELISA	Ebola virus nucleoprotein (NP)	100 μ L rNP	N/A	N/A	37
18	Fluorescence signal for quantitative detection and colorimetric signal for visual detection	Ebola virus glycoprotein	50 μ L	N/A	20 min	38



Table 2 (Contd.)

#	Method summary	Analyte	Sample volume	Accuracy	Time	Ref.
19	Chemiluminescent ELISA	Antibodies against ebola virus strains Zaire and Sudan	75 μ L	Low cross-reactivity	N/A	39
20	Electroluminescent nanospheres and immunomagnetic separation	Ebola virions	200 μ L	N/A	2 hours	40
21	Fluorescence assay on a micromagnetic platform	Ebola virions	200 μ L EBOV sample	Intra-assay CV = 4.9%	N/A	41
22	SERS immunoassay	Zika virions	10 μ L	N/A	N/A	42
23	Aptamer-based ELISA	Zika NS1 protein	100 μ L	N/A	N/A	43
24	Field effect biosensing	Zika viral antigen ZIKV NS1	75 μ L	1 : 10 dilution: CV = 19.89% 1 : 100 dilution: CV = 9.17%	N/A	44
25	Motion-based immunological detection	Zika virions	10 μ L of ZIKV	Correlation: 89.11% with the light microscopy; 100% with CDC Zika MAC-ELISA; 80% with Aptima Zika virus assay	>40 min for assay	45
26	Determination of isothermally Amplified Zika Virus RNA using a Universal DNA-Hairpin Probe	Zika Virus RNA	50 μ L	N/A	1 hour	46
27	Advanced strand exchange amplification (ASEA)	Zika Virus RNA	N/A	N/A	25 min	47
28	ELISA	anti-ZIKA IgM antibody	N/A	87.5% positive agreement between CDC and InBios MAC-ELISAs	N/A	48
29	Single particle interferometric reflectance imaging sensor (SP-IRIS) cartridge	HFV virions (model – Ebola virions)	100 μ L	N/A	20 min	49
30	Real-time reverse transcription-PCR	HFV RNA	2 μ L of RNA	No amplification of HIV-1, hepatitis B and C, herpes simplex type 1, cytomegalovirus, and Modoc viruses	>1 hour	50
31	qRT-PCR	HFV RNA	5 μ L of viral RNA	CV < 5%, no cross-reactivity	>1 hour	51
32	RT-LAMP	Crimean-Congo hemorrhagic fever (CCHF) RNA	2 μ L of the target RNA	100% agreement between RT-LAMP and the nested PCR	60 min	52
33	Fiber-optic biosensor with chemiluminescence	Crimean-Congo hemorrhagic fever (CCHF) IgG antibodies	200 μ L	N/A	90 min	53
34	Loop-mediated isothermal amplification, LAMP	Dengue virus RNA	2 μ L of prepared RNA	The area under the ROC curve (AUC) = 0.95	<1 hour	54
35	Biosensor with isothermal nucleic acid sequence-based amplification (NASBA)	Dengue virus RNA, serotypes 1, 2, 3, and 4	2 μ L of amplicon (amplified dengue virus RNA)	Serotype 3 displayed low cross reactivity with biosensors designed for the detection of serotypes 1 and 4	15 min	55
36	Tandem toehold-mediated displacement reactions (τ TMDR) with fluorescence	Dengue virus RNA	100 μ L	N/A	35 min	56
37	Magnetic paper-based ELISA	Dengue immunoglobulin M (IgM) antibodies	5 μ L	N/A	N/A	57
38	Long-range surface plasmon polariton (LRSP) gold (Au) waveguides	Dengue IgM antibody	~10 μ L plasma	N/A	N/A	58



Table 2 (Contd.)

#	Method summary	Analyte	Sample volume	Accuracy	Time	Ref.
39	Stacking flow immunoassay	Dengue-specific immunoglobulin antibody	100 μ L	N/A	N/A	59
40	Separative extended gate field-effect transistor (SEGFET) as an immunosensor	Dengue virus nonstructural protein 1 (NS1)	N/A	N/A	<1 hour	60
41	Lateral flow immunoassay (LFIA)	Dengue NS1 protein	10 mL	N/A	<1 hour	61
42	Magnetic separation and fluorescence detection	Dengue-2 virus virions	20 mL	High specificity in the presence of yellow fever virus	30–60 min	62
43	Electrochemical membrane-based nanobiosensor	Dengue-2 virus virions	5 μ L	RSD = 5.9%	N/A	63
44	RT-PCR	Hepatitis A virus (HAV) RNA	5 μ L of RNA extracts	N/A	>1 hour	64
45	qRT-PCR	HAV RNA	5 μ L of RNA extract	Regression coefficient of 0.9999	>1 hour	65
46	Indirect competitive electrochemical immunosensor	HAV antigen	100 μ L	RSD < 3%	N/A	66
47	Electrochemical immunosensor	HAV antigen	1.0 mL min ⁻¹	RSD = 3.1–5.7%	5 min	67
48	Solid-phase radioimmunoassay, HAVAB®-M	anti-HAV antibodies	100 μ L	RSD = 22%	N/A	68
49	Immunochemical assay (ICA)	anti-HAV IgM antibodies	5 μ L	Specificity = 100%	<20 min	69
50	Resonance light scattering (RLS) sensor	HAV virions	N/A	RSD = 1.3%	N/A	70
51	Multifunctional molecularly imprinted fluorescence sensor	HAV virions	200 μ L	RSD < 2.7%	20 min	71
52	A reduced graphene oxide-assisted hybridization chain reaction + fluorescence	Hepatitis C virus (HCV) RNA	N/A	RSD = 3–6.4%	>8 hours	72
53	Capture of RNA with probes and paramagnetic particle separation	HCV RNA	300 μ L of serum	93% sensitivity and 100% specificity	>1 hour	73
54	Electrochemical immunosensor	HCV antigen	1.0 mL min ⁻¹	RSD = 2.3–5.3%	5 min	67
55	Chemiluminescent magnetic particle-based immunoassay	HCV core Antigen	A few hundred μ L	99% specificity; 97.4% sensitivity	200 assays per hour	74
56	Sandwich electrochemical immunosensor	HCV core antigen	≈ 10 μ L	RSD = 3.1%	30 min	75
57	Nano-gold immunological amplification and silver staining (NIASS)	anti-HCV antibodies	10 μ L	N/A	<40 min	76
58	Antibody-induced DNA strand displacement and rolling circle amplification (RCA)	anti-HCV antibodies	1 μ L	“High specificity”	30 min	77
59	Immunogold electron microscopy	HCV virions	3 μ L	N/A	>3 hours	78
60	Dual-targeting real-time RT-PCR	Hepatitis D Virus (HDV) RNA	140 μ L viral sample	N/A	>1 hour	79
61	Real-time PCR	HDV RNA	200 μ L specimen	N/A	>20 min	80
62	Electrochemical immunosensor	HDV antigen	1.0 mL min ⁻¹	RSD = 3.4–6.8%	5 min	67



Table 2 (Contd.)

#	Method summary	Analyte	Sample volume	Accuracy	Time	Ref.
63	IgM capture enzyme immunoassay (EIA)	HDV antibodies IgM anti-HD	100 μ L	No cross-reactivity with other antibodies	2 days	81
64	Real-time RT-PCR	Hepatitis E virus (HEV) subtype 3b RNA	5 μ L of extracted RNA	N/A	>1 hour	82
65	Array-based nano-amplification and silver stain enhancement	HEV RNA	100 μ L	N/A	20–30 min	83
66	Electrochemical immunosensor	HEV antigen	1.0 mL min^{-1}	RSD = 3.4–6.9%	5 min	67
67	Enzyme immunoassay (EIA)	Antibody to the hepatitis E virus (anti-HEV)	250 μ L	N/A	N/A	84

5. Detection of SARS-CoV-2 (COVID-19 pandemic virus)

On December 31, 2019, the WHO Chinese Country Office was notified about pneumonia cases of an unknown nature, epidemiologically linked to the seafood market in Wuhan, Hubei province.⁹⁷ At the same time, the Chinese CDC conducted an investigation in the field.⁹⁸ The viral nature of the disease was identified, 3 viral genomes from the bronchoalveolar lavage of three disease-affected individuals were sequenced, and the sequences were submitted to GISAID (accession ID: EPI_ISL_402119; EPI_ISL_402120; EPI_ISL_402121). According to

the sequencing data, the novel virus belongs to the *Coronaviridae* family, *Orthocoronaviridae* subfamily, *Betacoronavirus* genus, *Sarbecovirus* subgenus. The viral genome contains a 5' untranslated region (5'-UTR), replicase gene (orf1ab), Spike gene (S gene), Envelope gene (E gene), M gene, Nucleocapsid gene (N gene), and open reading frames 3, 7, 8, 10b, 13, and 14. The virus was named novel coronavirus 2019-nCoV, and the disease it causes was named novel coronavirus-infected pneumonia (NCIP) by the Chinese CDC.⁹⁹ Later, the WHO renamed the virus into SARS-CoV-2, and the disease into COVID-19.¹⁰⁰ The virus is highly contagious with easy person-to-person transmission, has a variable incubation period (from 4 to 24

Table 3 Detection of SARS and MERS coronaviruses

#	Method summary	Analyte	Sample volume	Accuracy	Time	Ref.
1	RT-PCR and indirect immunofluorescence serologic testing	SARS-CoV RNA	2 mL nasopharyngeal aspirates and 2 g feces	RT-PCR: 60% for positive cases; 99.4% for negative cases Serologic testing: 92% for positive cases; 92% for negative cases	RT-PCR: N/A Serologic testing: >1 hour	90
2	Real time RT-PCR (2 assays: for upE and (ORF)1b)	SARS-CoV RNA: upstream of the E gene (upE) or within open reading frame (ORF)1b	5 μ L of extracted RNA	100% specificity for both for upE and (ORF)1b	36+ hours	91
3	Chemiluminescence immunosorbent assay with nanoarray RNA aptamer	SARS-CoV nucleocapsid protein (SARS-CoV N protein)	N/A	C-terminal domain or dimer form N protein is specifically recognized by the aptamer	N/A	92
4	Biosensor assay based on an optical QDs-based RNA aptamer	SARS-CoV nucleocapsid protein (SARS-CoV N protein)	N/A	QDs-conjugated RNA aptamer is selective against the SARS-CoV N protein	1 hour	93
5	Asymmetric five-primer reverse transcription loop-mediated isothermal amplification (RT-LAMP) assay	MERS-CoV RNA (3 genetic loci: ORF1a, ORF1b and E)	4 μ L RNA	N/A	30–50 minutes	94
6	ELISA	MERS-CoV nucleocapsid protein (NP)	50 μ L of sample	100% specificity	N/A	95
7	ELISA and plaque-reduction neutralization test (PRNT)	Antibodies against MERS-CoV	N/A	No serotypic discrimination between the MERS-CoV strains	N/A	96



Table 4 Detection of SARS-CoV-2

#	Method summary	Analyte	Sample volume	Accuracy	Time	Ref.
1	Real time RT-PCR Commercial kit from Altona diagnostics, Hamburg, Germany	E-gene RNA and S-gene RNA	30 µL	N/A	>1 hour	111 and 112
2	ePlex-based (DNA hybridization and electrochemical detection) SARS CoV-2	cDNA coding for nucleocapsid (N)	200 µL of nasopharyngeal swab	Detection of positive – 94.4%; 95% CI – 74.2– 99% Detection of negative – 100%; 95% CI – 92.4– 100%	>1 hour	111 and 113
3	Real-time RT-PCR	RNA coding for RdRp gene	N/A	N/A	>1 hour	114
4	Real-time RT-PCR	SARS-CoV-2 RNA coding for nucleocapsid gene (N1 probe)	20 µL	N/A	>1 hour	111 and 114
5	Real-time RT-PCR	SARS-CoV-2 RNA coding for nucleocapsid gene (N2 probe)	20 µL	N/A	>1 hour	114
6	Real-time RT-PCR (E-gene assay, and RdRp gene assay)	SARS-CoV-2 RNA coding for envelope (E) gene and RNA- dependent RNA polymerase (RdRp) gene	25 µL	No reactivity on human coronaviruses	>1 hour	105
7	Real-time RT-PCR	Co-V2 viral RNA; probe against nucleocapsid gene (N and N2 assays)	N/A	N/A	>1 hour	115
8	Real-time RT-PCR	SARS-CoV-2 RNA	5 µL RNA	No cross-reactivity with other human- pathogenic coronaviruses and respiratory pathogens	>1 hour	116
9	RT-PCR: Cepheid Xpert Xpress and Roche cobas assays	SARS-CoV-2 RNA	N/A	Both systems have agreement of 99%	45 min (Cepheid); 90 min (Roche)	117
10	RT-LAMP (reverse transcription loop- mediated isothermal amplification assay)	Viral RNA coding orf1ab gene and S gene	25 µL	Sensitivity – 100% (95% CI 92.3–100%) Specificity – 100% (95% CI 93.7–100%)	26.28 ± 4.48 min	107
11	Colorimetric LAMP	SARS-CoV-2 RNA	3 µL RNA	100% agreement with RT-PCR	30 min	118
12	RT-LAMP	Viral RNA coding for conserved region within nucleocapsid gene	N/A	Sensitivity – 100% Specificity – 98.7%	30 min (colorimetric visualization)	108
13	RT-LAMP	Viral RNA within RdRp gene	25 µL	100% consistency with RT-qPCR on positive samples	50 min real-time monitoring	109
14	DETECTR (SARS-CoV-2 DNA endonuclease- targeted CRISPR trans reporter) – CRISPR-Cas12- based assay	Viral RNA coding for nucleoprotein and envelope genes	N/A	95% for positive samples, 100% for negative samples	30–40 min (sample-to- result)	110
15	ELISA	SARS-CoV-2 neutralizing, spike- and nucleocapsid- specific antibodies	N/A	87–100% specificity	2 days	119
16	COVID-19 IgG/IgM rapid test Cassette	SARS-CoV-2-specific IgM and IgG	5 µL serum	Specificity 100% for IgM and 99.2% for IgG Sensitivity 69% for IgM and 93.1% for IgG	15 min	120
17	Colloidal gold Antibodies test	SARS-CoV-2-specific IgM and IgG	10 µL of serum	Specificity: IgM, 50.0%; IgG, 87.5%	N/A	121
18	Single molecule array (Simoa) immunoassay	SARS-CoV-2 nucleocapsid protein (N-protein)	20 µL of whole blood	100% specificity and 97.4% sensitivity	N/A	122
19	Pulse-controlled amplification (PCA)	SARS-CoV-2 RNA (E gene)	45 µL	100% agreement with RT-qPCR	20 min	123
20	LAMP	SARS-CoV-2 RNA	N/A	N/A	<30 min	124



Table 4 (Contd.)

#	Method summary	Analyte	Sample volume	Accuracy	Time	Ref.
21	Reverse-transcription recombinase-aided amplification (RT-RAA)	SARS-CoV-2 RNA	140 μ L nasopharyngeal swabs and sputum	100% agreement with RT-PCR	5–15 min	125

Table 5 Detection of RNA-containing lethal viruses: classification by method of detection

PCR-based nucleic acid detection					
Analyte	Ref.	Analytical parameters	Ref.	Analytical parameters	
Viral hemorrhagic fever (HF) viruses: Ebola, Marburg, Lassa, Crimean-Congo HF, Rift Valley fever, dengue, yellow fever viruses	50, 2002	LOD = 1545 to 2835 viral genome equivalents per mL of serum (8.6 to 16 RNA copies per assay)	51, 2014	LOD = between 45 and 150 cRNA/rxn	
Hepatitis A	64, 2009	LOD = 1 PFU/1.5 L	65, 2010	LOD = 10 PFU/1.5 L of bottled water, 100 PFU/1.5 L of tap water	
Hepatitis C	73, 2000	LOD = 33 cRNA per mL; ($\sim 1.74 \times 10^{-4}$ pg mL $^{-1}$)			
Hepatitis D	79, 2018	LOD = 575 IU mL $^{-1}$	80, 2013	LOD = 7500 HDV cRNA per mL; 190 cRNA/rxn (~ 0.28 pg mL $^{-1}$)	
Hepatitis E	82, 2013	LOD = 25 UI mL $^{-1}$			
Seasonal influenza A	6, 2009	LOD = log $_{10}$ 6.5–7.1 of M gene copies			
Swine-origin influenza A	6, 2009	LOD = log $_{10}$ 6.5–7.3 of M gene copies			
SARS-CoV	90, 2004	LOD = 10 cRNA/rxn; Ac = 60% for positive cases, 99.4% for negative cases	91, 2012	LOD = 3.4 cRNA/rxn for upstream of the E gene (upE) ($\sim 1.11 \times 10^{-2}$ pg mL $^{-1}$) and 64 cRNA/rxn for within open reading frame (ORF)1b, Sp = 100%	
SARS-CoV-2	116, 2020	LOD = 11.2 cRNA/rxn ($\sim 3.67 \times 10^{-2}$ pg mL $^{-1}$)	111, 2020	LOD = 24 cRNA/rxn ($\sim 1.97 \times 10^{-2}$ pg mL $^{-1}$)	
	105, 2020	LOD = 3.8 cRNA/rxn ($\sim 2.49 \times 10^{-3}$ pg mL $^{-1}$) – RdRp-gene assay; 5.2 cRNA/rxn ($\sim 3.41 \times 10^{-3}$ pg mL $^{-1}$) – E-gene assay	115, 2020	LOD = 25 and 250 cRNA/rxn	
Lateral flow immunoassays					
HIV-1 p24 antigen	25, 2016	LOD = 25 pg mL $^{-1}$			
Ab against HIV	31, 2006	LOD = sub pmol L $^{-1}$ range			
HIV-1 RNA	34, 2019	LOD = 3×10^5 HIV-1 viral particles, or 2.3×10^7 virus copies per mL of whole blood			
Dengue NS1 protein	61, 2020	LOD = 5 ng mL $^{-1}$			
ELISA					
HIV p24 antigen	29, 2003	LOD = 0.5 pg mL $^{-1}$			
Ab to HIV-1, HIV-2	32, 2013	LOD = 1 ng mL $^{-1}$ (6.7 pM)			
Ebola virus nucleoprotein (NP)	37, 2001	LOD = 30 ng of purified recombinant NP (rNP) ($\sim 3.00 \times 10^5$ pg mL $^{-1}$)			
Zika NS1 protein	43, 2017	LOD = 0.1 ng mL $^{-1}$			
IgM-dengue antibodies	57, 2017	LOD = 0.04 μ g mL $^{-1}$			
MERS-CoV nucleocapsid protein (NP)	95, 2015	LOD = 10 TCID $_{50}$ /0.1 mL			



Table 5 (Contd.)

PCR-based nucleic acid detection				
Analyte	Ref.	Analytical parameters	Ref.	Analytical parameters
AFM				
HIV p24 antigen	27, 2004	LOD = 25 fg mL ⁻¹		
Electrochemical detection				
HIV p24 antigen	28, 2010	LOD = 7.9 × 10 ⁻⁸ pg mL ⁻¹		
Anti-HIV antibodies	33, 2012	LOD = 1–10 nM (~1.50 × 10 ⁵ pg mL ⁻¹)		
Ebola virions	40, 2017	LOD = 5.2 pg mL ⁻¹		
Zika viral antigen ZIKV NS1	44, 2018	LOD = 450 pM		
Zika Virus RNA	46, 2019	LOD = 1.11 fg μL ⁻¹ (~0.3 fM)		
Dengue virus nonstructural protein 1 (NS1)	60, 2014	LOD = 0.25 μg mL ⁻¹		
Dengue-2 virions	63, 2012	LOD = 1 PFU mL ⁻¹		
H5N1 avian influenza virus hemagglutinin (HA) protein	13, 2020	LOD = 5.9 pM		
Hepatitis A antigen	66, 2017	LOD = 26 × 10 ⁻⁵ IU/mL	67, 2010	LOD = 0.5 ng mL ⁻¹
Hepatitis C antigen	67, 2010	LOD = 0.8 ng mL ⁻¹	75, 2017	LOD = 3 fg mL ⁻¹
Hepatitis D antigen	67, 2010	LOD = 0.5 ng mL ⁻¹		
Hepatitis E antigen	67, 2010	LOD = 1 ng mL ⁻¹		
Chemiluminescence detection				
HIV p24 antigen	26, 2011	LOD = 18–25 pg mL ⁻¹ or 1.24 IU mL ⁻¹		
Hepatitis C Virus (HCV) core antigen	74, 2006	LOD = viral concentration equivalent to the lowest titer of 2415 cRNA/mL		
Fluorescence detection				
HIV p24 antigen	26, 2011	LOD = 22–77.4 pg mL ⁻¹ or 1.94–2.25 IU mL ⁻¹	26, 2011	LOD = 13 pg mL ⁻¹ or 0.43 IU mL ⁻¹
	26, 2011	LOD = 11.5–25 pg mL ⁻¹ or 0.66 IU mL ⁻¹	26, 2011	LOD = 11.2 pg mL ⁻¹ or 0.73–1.15 IU mL ⁻¹
Ebola RNA	35, 2016	LOD = femtomolar level		
Ebola glycoprotein	38, 2017	LOD = 0.18 ng mL ⁻¹		
Ebola virions	41, 2018	LOD = 2.6 pg mL ⁻¹		
Dengue virus RNA	56, 2018	LOD = 6 cRNA per sample (~3.62 × 10 ⁻⁴ pg mL ⁻¹)		
Dengue-2 virions	62, 2008	LOD = 10 PFU mL ⁻¹		
HAV virions	71, 2019	LOD = 3.4 pmol L ⁻¹		
HCV RNA	72, 2019	LOD = 10 fM		
anti-HCV Ab	77, 2019	LOD = 0.998 pM		
SARS-CoV nucleocapsid protein (SARS-CoV N protein)	92, 2009	LOD = 2 pg mL ⁻¹		
LAMP-based nucleic acid detection				
Ebola	36, 2017	LOD = 100 cRNA (~1.04 pg mL ⁻¹)		
Crimean-Congo hemorrhagic fever	52, 2013	LOD = 0.1 fg of viral RNA (equivalent to 50 viral particles; ~0.05 pg mL ⁻¹)		
Dengue	54, 2020	LOD = 10 ² PFU per 200 μL of whole blood		
Influenza A	11, 2020	LOD of subtypes H1: 50 copies H3: 20–50 copies H5: 50 copies H7: 20–50 copies H9: 50–100 copies		



Table 5 (Contd.)

PCR-based nucleic acid detection				
Analyte	Ref.	Analytical parameters	Ref.	Analytical parameters
Influenza B	11, 2020	LOD = 50 copies		
MERS-CoV	94, 2015	LOD = 0.02 to 0.2 PFU (5 to 50 PFU mL ⁻¹)		
SARS-CoV-2	118, 2020	LOD = 120 cRNA/rxn (or 4.8 copies per μ L); $\sim 7.87 \times 10^{-2}$ pg mL ⁻¹	107, 2020	LOD = 20 cRNA/rxn - ORF1ab gene ($\sim 1.31 \times 10^{-2}$ pg mL ⁻¹) and 200 cRNA/rxn - S gene (~ 0.131 pg mL ⁻¹)
	109, 2020	LOD = 3 cRNA/rxn ($\sim 1.97 \times 10^{-3}$ pg mL ⁻¹)		
CRISPR-Cas based				
SARS-CoV-2	110 and 128, 2020	LOD = 10 cRNA/rxn (~ 0.164 pg mL ⁻¹)		
Biobarcode detection				
HIV p24 antigen	30, 2007	LOD = 0.1 pg mL; Ac = 100%		
Motion-based detection				
Zika virions	45, 2018	LOD = 1 particle/ μ L		
NASBA-based nucleic acid detection				
Zika	46, 2019	LOD = 1.11 fg μ L ⁻¹ (~ 0.3 fM)		
Dengue	55, 2002	LOD = 10 PFU mL ⁻¹		
Advanced strand exchange amplification-based nucleic acid detection				
Zika	47, 2018	LOD = 1.0×10^{-15} M; (~ 33 pg mL ⁻¹)		
Interferometric reflectance imaging				
HFV virions	49, 2017	LOD = 10 viruses per spot		
Surface plasmon polariton waveguides				
Dengue-specific immunoglobulin M (IgM) antibody	58, 2014	LOD = ~ 22 pg mm ⁻²		
SERS (Surface Enhanced Raman Spectroscopy)				
Zika virions	42, 2018	LOD = 10 ng mL ⁻¹		
FCV virions	129, 2005	LOD = 10 ⁶ viruses per mL		
Radioimmunoassay				
Anti-HAV Ab	68, 1993	LOD = 10 mIU mL ⁻¹		
Resonance light scattering				
HAV virions	70, 2017	LOD = 8.6 pmol L ⁻¹		
Silver staining				
HCV antibodies	76, 2005	LOD = 3 ng mL ⁻¹		
HEV RNA	83, 2006	LOD = 100 fM; (~ 237.6 pg mL ⁻¹)		
Electron microscopy				
HCV virions	78, 2006	LOD = 10 ⁷ virions per mL		
SPR				
AIV H5N1 virions	8, 2012	LOD = 0.128 HAU		
Confocal laser scanning microscopy				
SARS-CoV nucleocapsid protein	93, 2011	LOD = 0.1 pg mL ⁻¹		



Table 5 (Contd.)

PCR-based nucleic acid detection				
Analyte	Ref.	Analytical parameters	Ref.	Analytical parameters
Magnetic particle spectroscopy				
H1N1 nucleoprotein molecule	14, 2020	LOD = 4.4 pmoles		
Nanocomposite-based optical and mechanical detection				
Influenza A virions	12, 2020	LOD = 5×10^7 PFU mL ⁻¹		

days),¹⁰¹ and leads to the development of respiratory diseases with variable symptoms, from a mild cough to pneumonia.¹⁰² Sometimes, the infected person does not show any symptoms whatsoever.¹⁰³ Soon after the disease outbreak, close monitoring of the epidemiological situation around the globe started with a real-time count of new cases in the world.¹⁰⁴ Strict quarantine measures were implemented in the countries most affected by the disease, and on March 11th, the WHO declared a state of pandemic. Accurately monitoring the virus and its spread is not possible without reliable diagnostic tools. So, soon after the outbreak, the development of tests detecting SARS-CoV-2 itself or the immune response in the affected person was initiated.

A sample for the test includes material from the upper and lower respiratory tracts, and can include aspirates, oropharyngeal and nasopharyngeal swabs, bronchoalveolar lavage, and sputum, as well as nasal and nasopharyngeal aspirate. Test results depend on the quality of the collected material, as well as on the type of material.

Multiple assays were developed to test the presence of viral nucleic acid. The tests for SARS-CoV-2 detection employ a probe to detect the sequence within the RNA-dependent RNA polymerase viral gene (RdRp), as well as the nucleocapsid gene (NP), envelope protein gene (E), and spike protein gene (S). Some tests rely on the usage of fluorescently labeled TaqMan probes with a fluorescent reporter and a quencher attached to the 5'- and 3'-ends of the probe, respectively. Examples of such probes include probes with 6-carboxyfluorescein (FAM) or HEX dye at their 5'-end, and with a blackberry quencher (BBQ) or blackhole quencher (BHQ) at the probe's 3'-end.¹⁰⁵ The 5'-exonuclease ability of DNA-polymerase removes the fluorescently-labeled 5'-end of the hybridized probe, which leads to probe degradation and unquenching of the fluorescent reporter. There are 14 probes provided by the WHO, and one described by Zhu *et al.*⁹⁸ Most of them use TaqMan probes, with one exception from Japan that relies on nested PCR instead. Information about the limit of detection and the accuracy of the real-time reverse-transcription tests is gathered in Table 4.

Alongside real-time RT-PCR, isothermal hybridization is a method of choice for viral RNA detection in a shorter period of time, as compared to real-time RT-PCR. Originally invented by Notomi,¹⁰⁶ the so-called LAMP (loop-mediated isothermal amplification) coupled to reverse transcription is also used for SARS-CoV-2 detection,¹⁰⁷⁻¹⁰⁹ and allows for the rapid detection of viral

RNA within a time frame of less than 1 hour. The method relies on the strand-displacing ability of the Bst polymerase, and needs 4 to 6 primers, which increases the target selectivity of this method, as compared to regular PCR with only 2 primers.¹⁰⁶ The method can be run on a regular thermostat at 65 °C. The sensitivity of the RT-LAMP method is slightly lower (~10-fold) than that of real-time RT-PCR^{108,109} for SARS-CoV-2 detection.

The most novel method of viral detection, and possibly the most intriguing one, uses CRISPR-Cas 12 endonuclease and isothermal amplification.¹¹⁰ This method, named DETECTR, is very rapid, allowing for the detection of the viral RNA within minutes. It is a very promising point-of-care test that does not require expensive equipment and can be used in developing countries.

6. Summary of detection methods

Nucleic acid detection methods including PCR and LAMP

Comparing the different methods of viral detection, PCR-based methods can detect a few copies of RNA per reaction, which places them on the high-sensitivity spectrum, corresponding to a fg mL⁻¹ concentration range. PCR detection sensitivity is comparable between different viruses, with a range of detection from a few copies per reaction to a few hundreds of copies per reaction. PCR-based methods require trained personnel and expensive equipment, such as the PCR thermocycler. They are also time-consuming and can take several hours. In order to facilitate PCR-based techniques without compromising selectivity and specificity, alternatives to RT-qPCR exist. These include the LAMP technique. LAMP has a limit of detection comparable to that of RT-PCR, spanning from 3 copies of RNA per reaction to 100 copies of RNA per reaction, as evidenced from 5 sources presented in Table 5.

LAMP-based diagnostic tests allow for the rapid detection of the analyte, without a need for expensive equipment. Results can be obtained rapidly (within an hour), which makes these tests a suitable platform to be used for the development of point-of-care diagnostics tests. Alongside LAMP, the NASBA-based nucleic acid detection,^{46,55} as well as the advanced strand exchange amplification-based nucleic acid detection, are used and demonstrate a low limit of detection. A standard PCR test can also be incorporated into the assay, with other methods of separation such as laser-irradiated DNA extraction, paramagnetic particle separation, and others. Today, stationary PCR-based methods requiring expensive and complicated



equipment are accompanied by bench scale PCR detection with portable devices and automated procedure, where only one step of sample loading is necessary, such as with the “Cepheid Xpert Xpress” and “Roche cobas” assays.¹¹⁷

Immunoassay-based methods

The average limit of detection for ELISA-based methods is within the pg mL^{-1} range, but has been demonstrated to be as low as a fraction of pg mL^{-1} .²⁹ ELISA-based methods are routinely used and have demonstrated their applicability for an extended period. Despite this, they require trained personnel and are time-consuming. Therefore, alternative methods are being developed to make diagnostics tests relatively cheap and affordable without compromising their sensitivity and selectivity. One of these methods is lateral flow immunoassay. This method has shown a limit of detection at the pg mL^{-1} level, which is comparable to fine ELISA-based methods, but lateral flow immunoassays are faster. For example, a standard ELISA can take anywhere from 2 hours²⁹ to 2 days,¹¹⁹ as studies show. However, lateral flow immunoassays take from 20 minutes⁴⁷ to 90 minutes,³⁴ on average. This makes lateral flow immunoassays a good alternative to ELISA, especially because they can be performed at a small scale. The selectivity of immunoassay-based methods allows for the distinguishing of viruses with 100% specificity,⁹⁵ but not for making distinctions between different strains of the same virus.⁹⁶ Like PCR-based methods, immunoassay-based methods are now commercially available in bench scale, portable, fully automated devices, suitable for point-of-care testing.¹²⁰ However, these are currently qualitative, and their sensitivity is unknown. Immunoassay-based methods can detect a viral antigen or antibodies to the virus in the sample. The latter is more widespread for diagnostic testing, but can only be conclusive after the onset of symptoms (on average, a week after infection).

Electrochemical detection methods

Electrochemical detection methods provide sensitive detection of a wide range of analytes: virions,⁴⁰ viral antigen,⁴⁴ antibodies to the virus,³³ and viral nucleic acids.⁴⁶ The main advantage of these detection methods is that they are relatively inexpensive and not limited by a diffraction limit (like optical methods). The lowest limit of detection was reported by Teeparuksapun *et al.*²⁸ to be at a subattogram per milliliter concentration. Other studies show a pg mL^{-1} to fg mL^{-1} limit of detection range, which is comparable to PCR and immunoassays. The specificity of these methods is also high.³³ Notably, these detection methods also provide reproducible results with $\text{RSD} < 5\%$, such as the voltammetry method of Tang *et al.*,⁶⁷ which allows the multiplex detection of five analytes (viral antigens). Multiplex analyte detection has been reported in many research papers, which gives electrochemical methods an advantage over PCR and immunoassays. Electrochemical methods can be tuned to perform wash-free analysis³³ as well. Electrochemical detection methods usually use one of three approaches: voltammetry, amperometry, and detection of the change in capacitance. All three can be used in fast analysis,^{28,33,67} as the reported time of

experiments ranged between 5 and 20 minutes. This is much faster than the majority of PCR tests and immunoassays. However, these electrochemical methods require complicated equipment (for example, the use of gold nanoparticles with TEM and AFM imaging), highly trained operators, solution media, and are not suitable for point-of-care testing. Currently, the potential use of these methods in diagnostic testing is improbable because they require the fabrication of electrochemical sensors, often on a nanoscale level.

Fluorescence

Fluorescence spectroscopy is a well-established technique of sensitive detection. Its limit of detection is in the pg mL^{-1} range, on average, as can be seen in Table 5. Fluorescence signal detection can be incorporated in other procedures, such as immunoassays for the detection of antibodies or antigens.²⁶ Immunoassays with fluorescence signals have already been commercialized, and are even available in portable devices for home and point-of-care testing.²⁶ Fluorescence is reported as a reproducible method with $\text{RSD} < 5\%$.⁴¹ Other fluorescence-based methods use nanospheres, micromagnetic platforms, or upconversion materials for creating luminescence, which is further absorbed by nanoparticles conjugated to the target analyte. This method allows for more sensitive detection at the femtomolar level.³⁵ However, it is complicated and requires the fabrication of assay components, as well as highly trained personnel. Fluorescence signal acquisition is fast (around 20 minutes, on average), but the whole assay would take much longer to complete if the assay components need to be fabricated for fluorescence-based detection. Fluorescence-based immunoassays are faster (the whole assay takes around 2 hours), but their limit of detection is higher.²⁶ Fluorescence is

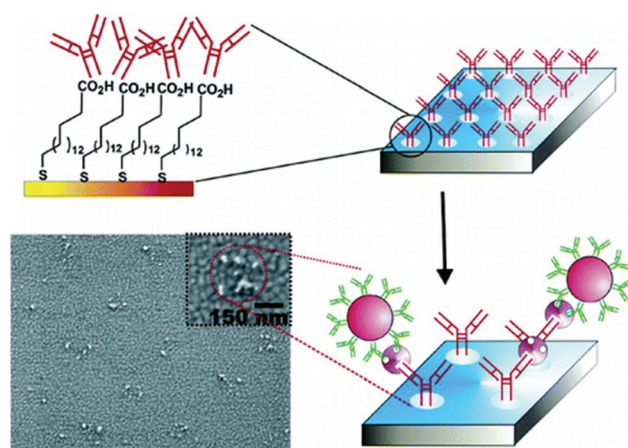


Fig. 4 AFM-based three-component ELISA using dip-pen nanolithography for HIV p24 antigen detection. The current method allows for detection of 25 fg mL^{-1} , which corresponds to 50 RNA copies per mL. The method shows at least 200-fold higher sensitivity compared to the 5 pg mL^{-1} limit of detection of conventional ELISA. The image is reprinted with permission from K. B. Lee, *et al.* The Use of Nanoarrays for Highly Sensitive and Selective Detection of Human Immunodeficiency Virus Type 1 in Plasma, *Nano Letters*, 2004, 4(10), 1869–1872. Copyright 2004 American Chemical Society.



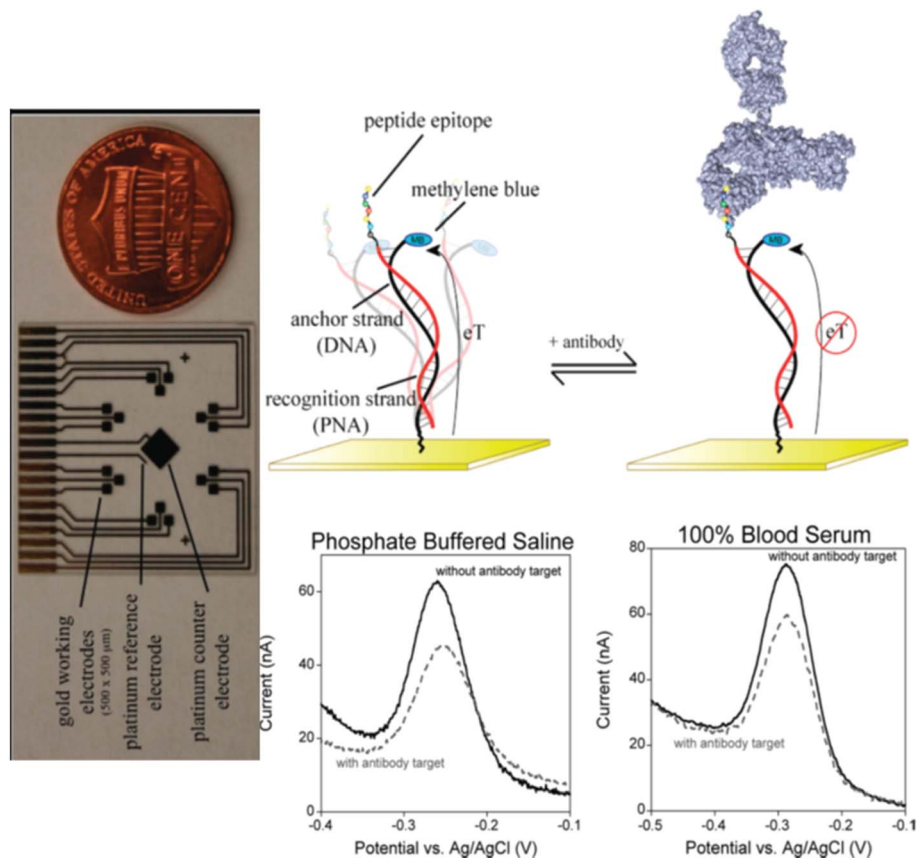


Fig. 5 E-DNA antibody sensor. The sensor (top) comprises an electrode-bound, redox-reporter-modified DNA strand, termed the "anchor strand," that forms a duplex with a complementary "recognition strand" (here composed of PNA) to which the relevant recognition element is covalently attached. In the absence of antibody binding (top middle), the flexibility of the surface attachment chemistry supports relatively efficient electron transfer between the redox reporter and the electrode surface. Binding to the relevant target antibody (top right) decreases electron transfer, presumably by reducing the efficiency with which the reporter collides with the electrode. Binding can thus be measured as a decrease in the peak current as observed *via* square wave voltammetry (bottom). As shown, sensors in this class are highly selective and perform equally well in buffered saline (bottom middle), undiluted blood serum (bottom right), or 1 : 4 diluted whole blood. The electrochemical E-DNA antibody sensor readily supports multiplexed detection. Here, (left, perpendicular orientation) a microfabricated chip containing eighteen $500 \times 500 \mu\text{m}$ sensors, arranged in six three-pixel clusters, was employed. Each cluster is directed against a different antibody. Thus, the device supports the simultaneous, triplicate measurement of six different targets. Copyright 2012 American Chemical Society. Reprinted with permission from ref. 33, R. J. White, H. M. Kallewaard, W. Hsieh, A. S. Patterson, J. B. Kasehagen, K. J. Cash, *et al.* Wash-free, electrochemical platform for the quantitative, multiplexed detection of specific antibodies. *Anal. Chem.*, 2012, **84**(2), 1098–1103.

subject to challenges such as photobleaching, autofluorescence, and dissociation of organic dyes used in live cells.

Other methods

There are other promising methods of detection: for example, silver staining, surface plasmon resonance (SPR), surface-enhanced Raman spectroscopy (SERS), confocal laser scanning microscopy, resonance light scattering, and radioimmunoassay. As can be seen in Table 5, silver staining and confocal laser scanning microscopy have the lowest limits of detection, in the pg to ng mL^{-1} and pg mL^{-1} ranges, respectively. They are also suitable for the detection of different analytes, such as viral nucleic acids, antigens, and antibodies to the virus.

Radioimmunoassay is another highly sensitive and inexpensive procedure, as can be seen in Table 5. The selectivity of this method is also very high, and the experimental procedure is

similar to that of a standard ELISA. However, special precautions must be taken during the experiment because radio-labeled reagents are used. Resonance light scattering is another valuable technique, whose convenience and sensitivity make it a potential diagnostic tool in health care.

SPR is mainly used to track the binding dynamics of biologically important molecules. It can also be used for the quantitative detection of analytes, as was demonstrated in ref. 8. This method has an advantage over other surface spectroscopy techniques because it does not need a vacuum, and is able to produce a linear dependence of resonant energy on the analyte concentration. SPR spectroscopy has potential for multiplexing, particularly when it is integrated with multi-channel microfluidic devices (Fig. 4).¹²⁶

SERS is a technique that allows for producing an enhancement in the order of millions and billions over standard Raman spectroscopy, which makes it useful for sensitive detection (in



ng mL⁻¹ range) of various analytes based on the characteristic Stokes shifts. This method requires a skilled operator and the use of a Raman spectrometer, which is now available as a portable, bench-size or even handheld device.¹²⁷ In comparison with fluorescence, Raman has great potential for multiplex detection of various analytes. Overall, this makes SERS a potential diagnostic test method. All abovementioned methods are listed in Table 5.

7. Post-COVID-19 trends and future perspectives

Comparing different methods, nucleic acid detection following amplification (real-time RT-PCR and LAMP) has higher sensitivity than ELISA, with a fg mL⁻¹ vs. pg mL⁻¹ average detection limit, respectively. The average limit of detection for real-time RT-PCR is 5 fg mL⁻¹ ($n = 7$), and 49 fg mL⁻¹ ($n = 6$) for the LAMP method. Having a detection limit higher than that for the ELISA method, the PCR-based tests are not free of drawbacks. Their disadvantages include the need for expensive equipment, trained personnel, and time. It takes several hours from the time of sample collection for a real-time RT-PCR test to produce results. The LAMP technique overcomes this limitation and enables a shorter (less than 1 hour) time until results, without the need for expensive equipment.

The limit of detection of electrochemical methods is comparable to the one for amplification-based methods of nucleic acid detection. The highest limit of detection is demonstrated by electrochemical methods with a capacitive immunosensor²⁸ for the detection of the HIV p24 antigen, which has a limit of detection of 7.9×10^{-5} fg mL⁻¹, and is accomplished in 20 minutes. The electrochemical methods are diverse in their principle, with some relying on capacitance measurements,²⁸ while others employ voltammetry⁴⁶ or amperometry.⁶⁰ They are also less restrictive in terms of the detected analyte, and are designed for the detection of viral particles, whether RNA⁴⁶ or protein²⁸, as well as antibodies (Fig. 5). The electrochemical methods are rapid and diverse. Some of them could be developed into low-cost point-of-care tests.

Amplification-based methods for the detection of nucleic acids, as well as ELISA-based methods for the detection of immune response in the form of antibodies, are two methods that are routinely used and demonstrate high sensitivity and specificity. However, they require time and proper equipment. The future of viral diagnostics lies in the point-of-care methods that can produce results within minutes, and do not require special equipment. The recent development of rapid diagnostics of SARS-CoV-2 in saliva¹³⁰ demonstrates the possibility of such a method. Other possible methods that can be used in point-of-care testing include portable antibodies/antigen test kits, fluorescence immunoassay devices, as well as portable RT-PCR and RT-LAMP devices.

Comparing the time it takes for different tests to produce results, lateral flow immunoassays offer a faster time to results, as compared to other methods (usually several minutes).

Electrochemical detection is another rapid diagnostics method, which takes less than 1 hour to produce results. Real-time RT-PCR takes the most time out of all test methods.

Enormous interest in virus detection on the wake of the COVID-19 pandemic is likely to reshape this area, directing efforts towards creating detection methods that have a fast time to results, high simplicity, high throughput, and are relatively low-cost. However, the specificity and selectivity of those tests should remain at least on par with well-established methods, such as PCR and ELISA.

Express testing for COVID-19 is necessary to contain the infection, and is particularly important for testing in airports, places of public gathering, transportation hubs and malls. During the pandemic, fast point-of-care tests on the scale of millions per day are required to minimize the mass quarantining of people, and to avert the harsh effects of lockdowns and travel bans on the economy and the wellbeing of people. Tests that take several days, such as the PCR test of the nasopharyngeal swabs, can result in the spread of infections since tested people will not know about their infection status until 5–7 days have passed. Today, there is a focus on developing express COVID-19 detection methods, especially using portable devices. There are several potential methods that provide test results within minutes or hours. One of them is the express RT-PCR test kit. These kits were developed by Cepheid (“Cepheid Xpert Xpress”), Roche (“Roche cobas”)¹¹⁷ and Abbott (“ID NOW”).¹³¹ These are already used in hospitals and ERs. According to published research, their accuracy matches that of the regular RT-PCR.¹¹⁷ This fact is a benefit of portable PCR devices, because with a sensitivity and specificity equal to that of stationary PCR, and with a significantly shorter time to results, they can be used for point-of-care testing. Some of these tests, such as “ID NOW”, are approved by the FDA. Another express COVID-19 testing method is RT-LAMP. RT-LAMP uses DNA polymerase with the capability of separating double-stranded DNA, which eliminates the need for cycling temperature and simplifies the procedure. Portable and rapid testing devices using this method have also been developed, such as “Talis One” by Talis Biomedical.¹³¹ This device provides results in 30 minutes, a time comparable to that of portable RT-PCR devices. Published research on the detection of RNA from SARS-CoV-2 by RT-LAMP shows 100% agreement between the RT-LAMP and RT-PCR methods.¹¹⁸ Both RT-LAMP and RT-PCR can be used for the detection of viral RNA in the saliva from a person. Currently, the main method of detection is the same as with nasopharyngeal swabs – RT-PCR. Saliva was reported to be a suitable sample for COVID-19 testing by the University of Illinois. Researchers at the Yale School of Public Health and Hokkaido University even commented that PCR testing of saliva provides more accurate results than testing of nasopharyngeal swabs. Collecting saliva does not need medical staff and is more comfortable for a patient. This allows for more frequent testing without putting pressure on medical staff. Several procedures have been developed to pretreat saliva before PCR because saliva is more of a chemically complex matrix than nasopharyngeal swabs. Some of these procedures were proven to be successful, such as the protocols from the University of Illinois



and Fluidigm's "Advanta Dx SARS-CoV-2 RT-PCR".¹³⁰ If the current saliva testing methods obtain governmental permission, they can replace the testing of nasopharyngeal swabs. The detection of viral antigens in blood or nasopharyngeal swabs is also a promising method. Benefits include it being relatively cheap, fast, and portable. One such example is the "Sofia SARS Antigen Fluorescent Immunoassay" by Quidel, which was approved for use in healthcare organizations. Detection of antibodies to the virus is also available in portable devices as a rapid test (e.g., "COVID-19 IgG/IgM Rapid Test Cassette" by Zhejiang Orient Gene Biotech Co., Ltd.). The main benefits are the ease of use and speed (results are available in 10 minutes, and the only requirement is the addition of a drop of blood and the provided buffer on a test slide).¹²⁰ Antibody detection, however, is efficient only after several days have passed since the onset of symptoms. Both antibody and antigen tests are easier to use and cheaper, but they are less sensitive than RT-PCR. They can be used after the onset of symptoms to avoid a false negative during early testing. There are other rapid tests, such as pulse-controlled amplification (20 minutes),¹²³ reverse-transcription recombinase-aided amplification (5–15 minutes),¹²⁵ and other methods. However, these approaches are all new research studies requiring proficient scientists, and are confined to the laboratory, so they are not the main focus of today's trends.

Abbreviations

AFM	Atomic-force microscopy
Ab	Antibodies
CDC	Center for Disease Control
CMV	Cytomegalovirus
CoV	Coronavirus
CRISPR	Clustered regularly interspaced short palindromic repeats
cRNA	RNA copies
DNA	Deoxyribonucleic acid
ELD ₅₀	50% egg lethal dose
ELISA	Enzyme-linked immunosorbent assay
HA	Haemagglutinin
HAU	Hemagglutination unit
HAV	Hepatitis A virus
HCV	Hepatitis C virus
HDV	Hepatitis D virus
HEV	Hepatitis E virus
HFV	Hemorrhagic fever virus
HIV	Human immunodeficiency virus
LAMP	Loop-mediated isothermal amplification
LOD	Limit of detection
MERS	Middle east respiratory syndrome
NA	Neuraminidase
NP	Nucleoprotein
NS	Nonstructural
PCR	Polymerase chain reaction
PFU	Plaque-forming unit
RNA	Ribonucleic acid
RSD	Relative standard deviation

RT-PCR	Reverse transcription polymerase chain reaction
Rxn	Reaction
SARS	Severe acute respiratory syndrome
SERS	Surface-enhanced Raman spectroscopy
SPR	Surface-plasmon resonance
TCID ₅₀	50% tissue culture infective dose
WHO	World Health Organization

Funding statement

This research is supported by the Nazarbayev University (NU) Faculty Development Competitive Research Grant no. 090118FD5352 (Kazakhstan), and the NU Faculty Development Competitive Research Grant no. 110119FD4517 (Kazakhstan).

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors would like to acknowledge Yunona Bukasova for proofreading the paper.

References

- X. Cheng, G. Chen and W. R. Rodriguez, Micro- and nanotechnology for viral detection, *Anal. Bioanal. Chem.*, 2009, **393**(2), 487–501.
- F. Krammer, G. J. D. Smith, R. A. M. Fouchier, M. Peiris, K. Kedzierska, P. C. Doherty, *et al.*, Influenza, *Nat. Rev. Dis. Primers*, 2018, **4**(1), 3.
- F. S. Dawood, A. D. Iuliano, C. Reed, M. I. Meltzer, D. K. Shay, P. Y. Cheng, *et al.*, Estimated global mortality associated with the first 12 months of 2009 pandemic influenza A H1N1 virus circulation: a modelling study, *Lancet Infect. Dis.*, 2012, **12**(9), 687–695.
- WHO, *Burden of disease*, WHO, 2020, https://www.who.int/influenza/surveillance_monitoring/bod/en/.
- CDC, *Disease Burden of Influenza CDC*, 2020, <https://www.cdc.gov/flu/about/burden/index.html>.
- K. H. Chan, S. T. Lai, L. L. Poon, Y. Guan, K. Y. Yuen and J. S. Peiris, Analytical sensitivity of rapid influenza antigen detection tests for swine-origin influenza virus (H1N1), *J. Clin. Virol.*, 2009, **45**(3), 205–207.
- C. C. Ginocchio, F. Zhang, R. Manji, S. Arora, M. Bornfreund, L. Falk, *et al.*, Evaluation of multiple test methods for the detection of the novel 2009 influenza A (H1N1) during the New York City outbreak, *J. Clin. Virol.*, 2009, **45**(3), 191–195.
- H. Bai, R. Wang, B. Hargis, H. Lu and Y. Li, A SPR aptasensor for detection of avian influenza virus H5N1, *Sensors*, 2012, **12**(9), 12506–12518.
- J. R. Ciacci-Zanella, A. L. Vincent, J. R. Prickett, S. M. Zimmerman and J. J. Zimmerman, Detection of anti-



- influenza A nucleoprotein antibodies in pigs using a commercial influenza epitope-blocking enzyme-linked immunosorbent assay developed for avian species, *J. Vet. Diagn. Invest.*, 2010, **22**(1), 3–9.
- 10 K. Watcharatanyatip, S. Boonmoh, K. Chaichoun, T. Songserm, M. Woratanti and T. Dharakul, Multispecies detection of antibodies to influenza A viruses by a double-antigen sandwich ELISA, *J. Virol. Methods*, 2010, **163**(2), 238–243.
 - 11 Y. Yao, X. Chen, X. Zhang, Q. Liu, J. Zhu, W. Zhao, *et al.*, Rapid Detection of Influenza Virus Subtypes Based on an Integrated Centrifugal Disc, *ACS Sens.*, 2020, **5**(5), 1354–1362.
 - 12 R. Randriantsilefisoa, C. Nie, B. Parshad, Y. Pan, S. Bhatia and R. Haag, Double trouble for viruses: a hydrogel nanocomposite catches the influenza virus while shrinking and changing color, *Chem. Commun.*, 2020, **56**(24), 3547–3550.
 - 13 J. Kwon, Y. Lee, T. Lee and J.-H. Ahn, Aptamer-Based Field-Effect Transistor for Detection of Avian Influenza Virus in Chicken Serum, *Anal. Chem.*, 2020, **92**(7), 5524–5531.
 - 14 K. Wu, J. Liu, R. Saha, D. Su, V. D. Krishna, M. C. J. Cheeran, *et al.*, Magnetic Particle Spectroscopy for Detection of Influenza A Virus Subtype H1N1, *ACS Appl. Mater. Interfaces*, 2020, **12**(12), 13686–13697.
 - 15 K. Danforth, R. Granich, D. Wiedeman, S. Baxi and N. Padian, in *Global Mortality and Morbidity of HIV/AIDS*, ed. K. K. Holmes, S. Bertozzi, B. R. Bloom and P. Jha, Washington, DC, 3rd edn, Major Infectious Diseases, 2017.
 - 16 BBC, *Ebola outbreak: Cases pass 10,000, WHO reports CDC*, 2014, <https://www.bbc.com/news/world-africa-29769782>.
 - 17 Medicine BCo, *Zika Virus*, 2020, <https://www.bcm.edu/departments/molecular-virology-and-microbiology/emerging-infections-and-biodefense/specific-agents/zika>.
 - 18 R. Noor and T. Ahmed, Zika virus: Epidemiological study and its association with public health risk, *J. Infect. Public Health*, 2018, **11**(5), 611–616.
 - 19 C. N. Fhogartaigh and E. Aarons, Viral haemorrhagic fever, *Clin. Med.*, 2015, **15**(1), 61–66.
 - 20 WHO, *Dengue and severe dengue 2020*, www.who.int/news-room/fact-sheets/detail/dengue-and-severe-dengue, accessed May 26, 2020.
 - 21 WHO, *Hepatitis A 2020*, <https://www.who.int/news-room/fact-sheets/detail/hepatitis-a>, accessed May 26, 2020.
 - 22 WHO, *Hepatitis C 2020*, <https://www.who.int/news-room/fact-sheets/detail/hepatitis-c>, accessed May 26, 2020.
 - 23 WHO, *Hepatitis D 2020*, www.who.int/news-room/fact-sheets/detail/hepatitis-d, accessed May 26, 2020.
 - 24 WHO, *Hepatitis E 2020*, www.who.int/news-room/fact-sheets/detail/hepatitis-e, accessed May 26, 2020.
 - 25 Y. Ma, C. Ni, E. E. Dzakah, H. Wang, K. Kang, S. Tang, *et al.*, Development of Monoclonal Antibodies against HIV-1 p24 Protein and Its Application in Colloidal Gold Immunochromatographic Assay for HIV-1 Detection, *BioMed Res. Int.*, 2016, **2016**, 6743904.
 - 26 M. Miedouge, M. Greze, A. Bailly and J. Izopet, Analytical sensitivity of four HIV combined antigen/antibody assays using the p24 WHO standard, *J. Clin. Virol.*, 2011, **50**(1), 57–60.
 - 27 K.-B. Lee, E.-Y. Kim, C. A. Mirkin and S. M. Wolinsky, The Use of Nanoarrays for Highly Sensitive and Selective Detection of Human Immunodeficiency Virus Type 1 in Plasma, *Nano Lett.*, 2004, **4**(10), 1869–1872.
 - 28 K. Teeparuksapun, M. Hedstrom, E. Y. Wong, S. Tang, I. K. Hewlett and B. Mattiasson, Ultrasensitive detection of HIV-1 p24 antigen using nanofunctionalized surfaces in a capacitive immunosensor, *Anal. Chem.*, 2010, **82**(20), 8406–8411.
 - 29 R. Sutthent, N. Gaudart, K. Chokpaibulkit, N. Tanliang, C. Kanoksinsombath and P. Chaisilwatana, p24 Antigen detection assay modified with a booster step for diagnosis and monitoring of human immunodeficiency virus type 1 infection, *J. Clin. Microbiol.*, 2003, **41**(3), 1016–1022.
 - 30 S. Tang, J. Zhao, J. J. Storhoff, P. J. Norris, R. F. Little, R. Yarchoan, *et al.*, Nanoparticle-Based biobarcode amplification assay (BCA) for sensitive and early detection of human immunodeficiency type 1 capsid (p24) antigen, *J. Acquired Immune Defic. Syndr.*, 2007, **46**(2), 231–237.
 - 31 J. L. Greenwald, G. R. Burstein, J. Pincus and B. Branson, A rapid review of rapid HIV antibody tests, *Curr. Infect. Dis. Rep.*, 2006, **8**(2), 125–131.
 - 32 A. Bhimji, A. A. Zaragoza, L. S. Live and S. O. Kelley, Electrochemical enzyme-linked immunosorbent assay featuring proximal reagent generation: detection of human immunodeficiency virus antibodies in clinical samples, *Anal. Chem.*, 2013, **85**(14), 6813–6819.
 - 33 R. J. White, H. M. Kallewaard, W. Hsieh, A. S. Patterson, J. B. Kasehagen, K. J. Cash, *et al.*, Wash-free, electrochemical platform for the quantitative, multiplexed detection of specific antibodies, *Anal. Chem.*, 2012, **84**(2), 1098–1103.
 - 34 E. A. Phillips, T. J. Moehling, K. F. K. Ejendal, O. S. Hoilett, K. M. Byers, L. A. Basing, *et al.*, Microfluidic rapid and autonomous analytical device (microRAAD) to detect HIV from whole blood samples, *Lab Chip*, 2019, **19**(20), 3375–3386.
 - 35 M. K. Tsang, W. Ye, G. Wang, J. Li, M. Yang and J. Hao, Ultrasensitive Detection of Ebola Virus Oligonucleotide Based on Upconversion Nanoprobe/Nanoporous Membrane System, *ACS Nano*, 2016, **10**(1), 598–605.
 - 36 P. Lam, R. A. Keri and N. F. Steinmetz, A Bioengineered Positive Control for Rapid Detection of the Ebola Virus by Reverse Transcription Loop-Mediated Isothermal Amplification (RT-LAMP), *ACS Biomater. Sci. Eng.*, 2017, **3**(3), 452–459.
 - 37 M. Niikura, T. Ikegami, M. Saijo, I. Kurane, M. E. Miranda and S. Morikawa, Detection of Ebola viral antigen by enzyme-linked immunosorbent assay using a novel monoclonal antibody to nucleoprotein, *J. Clin. Microbiol.*, 2001, **39**(9), 3267–3271.
 - 38 J. Hu, Y. Z. Jiang, L. L. Wu, Z. Wu, Y. Bi, G. Wong, *et al.*, Dual-Signal Readout Nanospheres for Rapid Point-of-Care Detection of Ebola Virus Glycoprotein, *Anal. Chem.*, 2017, **89**(24), 13105–13111.
 - 39 A. Petrosova, T. Konry, S. Cosnier, I. Trakht, J. Lutwama, E. Rwaguma, *et al.*, Development of a highly sensitive, field operable biosensor for serological studies of Ebola virus in central Africa, *Sens. Actuators, B*, 2007, **122**(2), 578–586.



- 40 Z. Wu, J. Hu, T. Zeng, Z. L. Zhang, J. Chen, G. Wong, *et al.*, Ultrasensitive Ebola Virus Detection Based on Electroluminescent Nanospheres and Immunomagnetic Separation, *Anal. Chem.*, 2017, **89**(3), 2039–2048.
- 41 S. L. Hong, Y. N. Zhang, Y. H. Liu, M. Tang, D. W. Pang, G. Wong, *et al.*, Cellular-Beacon-Mediated Counting for the Ultrasensitive Detection of Ebola Virus on an Integrated Micromagnetic Platform, *Anal. Chem.*, 2018, **90**(12), 7310–7317.
- 42 S. A. Camacho, R. G. Sobral-Filho, P. H. B. Aoki, C. J. L. Constantino and A. G. Brolo, Zika Immunoassay Based on Surface-Enhanced Raman Scattering Nanoprobes, *ACS Sens.*, 2018, **3**(3), 587–594.
- 43 K. H. Lee and H. Zeng, Aptamer-Based ELISA Assay for Highly Specific and Sensitive Detection of Zika NS1 Protein, *Anal. Chem.*, 2017, **89**(23), 12743–12748.
- 44 S. Afsahi, M. B. Lerner, J. M. Goldstein, J. Lee, X. Tang, D. A. Bagarozzi Jr, *et al.*, Novel graphene-based biosensor for early detection of Zika virus infection, *Biosens. Bioelectron.*, 2018, **100**, 85–88.
- 45 M. S. Draz, N. K. Lakshminaraasimulu, S. Krishnakumar, D. Battalapalli, A. Vasani, M. K. Kanakasabapathy, *et al.*, Motion-Based Immunological Detection of Zika Virus Using Pt-Nanomotors and a Cellphone, *ACS Nano*, 2018, **12**(6), 5709–5718.
- 46 C. A. Lynch III, M. V. Foguel, A. J. Reed, A. M. Balcarcel, P. Calvo-Marzal, Y. V. Gerasimova, *et al.*, Selective Determination of Isothermally Amplified Zika Virus RNA Using a Universal DNA-Hairpin Probe in Less than 1 Hour, *Anal. Chem.*, 2019, **91**(21), 13458–13464.
- 47 C. Ma, H. Jing, P. Zhang, L. Han, M. Zhang, F. Wang, *et al.*, An ultrafast one-step assay for the visual detection of RNA virus, *Chem. Commun.*, 2018, **54**(25), 3118–3121.
- 48 D. Granger, H. Hilgart, L. Misner, J. Christensen, S. Bistodeau, J. Palm, *et al.*, Serologic Testing for Zika Virus: Comparison of Three Zika Virus IgM-Screening Enzyme-Linked Immunosorbent Assays and Initial Laboratory Experiences, *J. Clin. Microbiol.*, 2017, **55**(7), 2127–2136.
- 49 S. M. Scherr, D. S. Freedman, K. N. Agans, A. Rosca, E. Carter, M. Kuroda, *et al.*, Disposable cartridge platform for rapid detection of viral hemorrhagic fever viruses, *Lab Chip*, 2017, **17**(5), 917–925.
- 50 C. Drosten, S. Gottig, S. Schilling, M. Asper, M. Panning, H. Schmitz, *et al.*, Rapid detection and quantification of RNA of Ebola and Marburg viruses, Lassa virus, Crimean-Congo hemorrhagic fever virus, Rift Valley fever virus, dengue virus, and yellow fever virus by real-time reverse transcription-PCR, *J. Clin. Microbiol.*, 2002, **40**(7), 2323–2330.
- 51 Z. Pang, A. Li, J. Li, J. Qu, C. He, S. Zhang, *et al.*, Comprehensive multiplex one-step real-time TaqMan qRT-PCR assays for detection and quantification of hemorrhagic fever viruses, *PLoS One*, 2014, **9**(4), e95635.
- 52 H. A. Osman, K. H. Eltom, N. O. Musa, N. M. Bilal, M. I. Elbashir and I. E. Aradaib, Development and evaluation of loop-mediated isothermal amplification assay for detection of Crimean Congo hemorrhagic fever virus in Sudan, *J. Virol. Methods*, 2013, **190**(1–2), 4–10.
- 53 F. Algaar, E. Eltzov, M. M. Vdovenko, I. Y. Sakharov, L. Fajs, M. Weidmann, *et al.*, Fiber-optic immunosensor for detection of Crimean-Congo hemorrhagic fever IgG antibodies in patients, *Anal. Chem.*, 2015, **87**(16), 8394–8398.
- 54 H. J. Yoo, C. Baek, M. H. Lee and J. Min, Integrated microsystems for the *in situ* genetic detection of dengue virus in whole blood using direct sample preparation and isothermal amplification, *Analyst*, 2020, **145**(6), 2405–2411.
- 55 A. J. Baeumner, N. A. Schlesinger, N. S. Slutzki, J. Romano, E. M. Lee and R. A. Montagna, Biosensor for dengue virus detection: sensitive, rapid, and serotype specific, *Anal. Chem.*, 2002, **74**(6), 1442–1448.
- 56 M. Gao, D. Daniel, H. Zou, S. Jiang, S. Lin, C. Huang, *et al.*, Rapid detection of a dengue virus RNA sequence with single molecule sensitivity using tandem toehold-mediated displacement reactions, *Chem. Commun.*, 2018, **54**(8), 968–971.
- 57 G. A. Ortega, S. Pérez-Rodríguez and E. Reguera, Magnetic paper-based ELISA for IgM-dengue detection, *RSC Adv.*, 2017, **7**(9), 4921–4932.
- 58 W. R. Wong, O. Krupin, S. D. Sekaran, F. R. Mahamd Adikan and P. Berini, Serological diagnosis of dengue infection in blood plasma using long-range surface plasmon waveguides, *Anal. Chem.*, 2014, **86**(3), 1735–1743.
- 59 Y. Zhang, J. Bai and J. Y. Ying, A stacking flow immunoassay for the detection of dengue-specific immunoglobulins in salivary fluid, *Lab Chip*, 2015, **15**(6), 1465–1471.
- 60 N. C. S. Vieira, A. Figueiredo, J. F. dos Santos, S. M. Aoki, F. E. G. Guimarães and V. Zucolotto, Label-free electrical recognition of a dengue virus protein using the SEG-FET simplified measurement system, *Anal. Methods*, 2014, **6**(22), 8882–8885.
- 61 T. Axelrod, E. Eltzov and R. S. Marks, Capture-Layer Lateral Flow Immunoassay: A New Platform Validated in the Detection and Quantification of Dengue NS1, *ACS Omega*, 2020, **5**(18), 10433–10440.
- 62 W. S. Chang, H. Shang, R. M. Perera, S. M. Lok, D. Sedlak, R. J. Kuhn, *et al.*, Rapid detection of dengue virus in serum using magnetic separation and fluorescence detection, *Analyst*, 2008, **133**(2), 233–240.
- 63 M. S. Cheng, J. S. Ho, C. H. Tan, J. P. Wong, L. C. Ng and C. S. Toh, Development of an electrochemical membrane-based nanobiosensor for ultrasensitive detection of dengue virus, *Anal. Chim. Acta*, 2012, **725**, 74–80.
- 64 S. Perelle, L. Cavellini, C. Burger, S. Blaise-Boisseau, C. Hennechart-Collette, G. Merle, *et al.*, Use of a robotic RNA purification protocol based on the NucliSens easyMAG for real-time RT-PCR detection of hepatitis A virus in bottled water, *J. Virol. Methods*, 2009, **157**(1), 80–83.
- 65 S. Blaise-Boisseau, C. Hennechart-Collette, L. Guillier and S. Perelle, Duplex real-time qRT-PCR for the detection of hepatitis A virus in water and raspberries using the MS2 bacteriophage as a process control, *J. Virol. Methods*, 2010, **166**(1–2), 48–53.
- 66 J. Mandli, A. Attar, M. M. Ennaji and A. Amine, Indirect competitive electrochemical immunosensor for hepatitis



- A virus antigen detection, *J. Electroanal. Chem.*, 2017, **799**, 213–221.
- 67 D. Tang, J. Tang, B. Su, J. Ren and G. Chen, Simultaneous determination of five-type hepatitis virus antigens in 5 min using an integrated automatic electrochemical immunosensor array, *Biosens. Bioelectron.*, 2010, **25**(7), 1658–1662.
- 68 W. J. Miller, W. Clark, W. Hurni, B. Kuter, T. Schofield and D. Nalin, Sensitive assays for hepatitis A antibodies, *J. Med. Virol.*, 1993, **41**(3), 201–204.
- 69 H. J. Lee, H. S. Jeong, B. K. Cho, M. J. Ji, J. H. Kim, A. N. Lee, *et al.*, Evaluation of an immunochromatographic assay for the detection of anti-hepatitis A virus IgM, *Virol. J.*, 2010, **7**, 164.
- 70 B. Yang, H. Gong, C. Chen, X. Chen and C. Cai, A virus resonance light scattering sensor based on mussel-inspired molecularly imprinted polymers for high sensitive and high selective detection of Hepatitis A Virus, *Biosens. Bioelectron.*, 2017, **87**, 679–685.
- 71 L. Luo, F. Zhang, C. Chen and C. Cai, Visual Simultaneous Detection of Hepatitis A and B Viruses Based on a Multifunctional Molecularly Imprinted Fluorescence Sensor, *Anal. Chem.*, 2019, **91**(24), 15748–15756.
- 72 J. Fan, L. Yuan, Q. Liu, C. Tong, W. Wang, F. Xiao, *et al.*, An ultrasensitive and simple assay for the Hepatitis C virus using a reduced graphene oxide-assisted hybridization chain reaction, *Analyst*, 2019, **144**(13), 3972–3979.
- 73 H. Miyachi, A. Masukawa, T. Ohshima, T. Hirose, C. Impraim and Y. Ando, Automated specific capture of hepatitis C virus RNA with probes and paramagnetic particle separation, *J. Clin. Microbiol.*, 2000, **38**(1), 18–21.
- 74 T. P. Leary, R. A. Gutierrez, A. S. Muerhoff, L. G. Birkenmeyer, S. M. Desai and G. J. Dawson, A chemiluminescent, magnetic particle-based immunoassay for the detection of hepatitis C virus core antigen in human serum or plasma, *J. Med. Virol.*, 2006, **78**(11), 1436–1440.
- 75 A. Valipour and M. Roushani, Using silver nanoparticle and thiol graphene quantum dots nanocomposite as a substratum to load antibody for detection of hepatitis C virus core antigen: Electrochemical oxidation of riboflavin was used as redox probe, *Biosens. Bioelectron.*, 2017, **89**(Pt 2), 946–951.
- 76 L. Duan, Y. Wang, S. S. Li, Z. Wan and J. Zhai, Rapid and simultaneous detection of human hepatitis B virus and hepatitis C virus antibodies based on a protein chip assay using nano-gold immunological amplification and silver staining method, *BMC Infect. Dis.*, 2005, **5**, 53.
- 77 T. Gao, W. Chai, L. Shi, H. Shi, A. Sheng, J. Yang, *et al.*, A new colorimetric assay method for the detection of anti-hepatitis C virus antibody with high sensitivity, *Analyst*, 2019, **144**(21), 6365–6370.
- 78 M. Kaito, S. Ishida, H. Tanaka, S. Horiike, N. Fujita, Y. Adachi, *et al.*, Morphology of hepatitis C and hepatitis B virus particles as detected by immunogold electron microscopy, *Med. Mol. Morphol.*, 2006, **39**(2), 63–71.
- 79 Y. Wang, J. S. Glenn, M. A. Winters, L. P. Shen, I. Choong, Y. L. Shi, *et al.*, A new dual-targeting real-time RT-PCR assay for hepatitis D virus RNA detection, *Diagn. Microbiol. Infect. Dis.*, 2018, **92**(2), 112–117.
- 80 M. Kodani, A. Martin, T. Mixson-Hayden, J. Drobeniuc, R. R. Gish and S. Kamili, One-step real-time PCR assay for detection and quantitation of hepatitis D virus RNA, *J. Virol. Methods*, 2013, **193**(2), 531–535.
- 81 A. G. Shattock, M. Morris, K. Kinane and C. Fagan, The serology of delta hepatitis and the detection of IgM anti-HD by EIA using serum derived delta antigen, *J. Virol. Methods*, 1989, **23**(3), 233–240.
- 82 C. Mokhtari, E. Marchadier, S. Haim-Boukobza, A. Jebblaoui, S. Tesse, J. Savary, *et al.*, Comparison of real-time RT-PCR assays for hepatitis E virus RNA detection, *J. Clin. Virol.*, 2013, **58**(1), 36–40.
- 83 H. H. Liu, X. Cao, Y. Yang, M. G. Liu and Y. F. Wang, Array-based nano-amplification technique was applied in detection of hepatitis E virus, *J. Biochem. Mol. Biol.*, 2006, **39**(3), 247–252.
- 84 E. E. Mast, M. J. Alter, P. V. Holland and R. H. Purcell, Evaluation of assays for antibody to hepatitis E virus by a serum panel. Hepatitis E Virus Antibody Serum Panel Evaluation Group, *Hepatology*, 1998, **27**(3), 857–861.
- 85 N. S. Zhong, B. J. Zheng, Y. M. Li, L. L. M. Poon, Z. H. Xie, K. H. Chan, *et al.*, Epidemiology and cause of severe acute respiratory syndrome (SARS) in Guangdong, People's Republic of China, in February, 2003, *Lancet*, 2003, **362**(9393), 1353–1358.
- 86 T. G. Ksiazek, D. Erdman, C. S. Goldsmith, S. R. Zaki, T. Peret, S. Emery, *et al.*, A novel coronavirus associated with severe acute respiratory syndrome, *N. Engl. J. Med.*, 2003, **348**(20), 1953–1966.
- 87 C. Drosten, S. Gunther, W. Preiser, S. van der Werf, H. R. Brodt, S. Becker, *et al.*, Identification of a novel coronavirus in patients with severe acute respiratory syndrome, *N. Engl. J. Med.*, 2003, **348**(20), 1967–1976.
- 88 A. M. Zaki, S. van Boheemen, T. M. Bestebroer, A. D. Osterhaus and R. A. Fouchier, Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia, *N. Engl. J. Med.*, 2012, **367**(19), 1814–1820.
- 89 WHO, *MERS situation update*, January 2020, <http://www.emro.who.int/health-topics/mers-cov/mers-outbreaks.html>.
- 90 K. H. Chan, L. L. Poon, V. C. Cheng, Y. Guan, I. F. Hung, J. Kong, *et al.*, Detection of SARS coronavirus in patients with suspected SARS, *Emerging Infect. Dis.*, 2004, **10**(2), 294–299.
- 91 V. M. Corman, I. Eckerle, T. Bleicker, A. Zaki, O. Landt, M. Eschbach-Bludau, *et al.*, Detection of a novel human coronavirus by real-time reverse-transcription polymerase chain reaction, *Eurosurveillance*, 2012, **17**(39), 20285.
- 92 D. G. Ahn, I. J. Jeon, J. D. Kim, M. S. Song, S. R. Han, S. W. Lee, *et al.*, RNA aptamer-based sensitive detection of SARS coronavirus nucleocapsid protein, *Analyst*, 2009, **134**(9), 1896–1901.
- 93 C. Roh and S. K. Jo, Quantitative and sensitive detection of SARS coronavirus nucleocapsid protein using quantum



- dots-conjugated RNA aptamer on chip, *J. Chem. Technol. Biotechnol.*, 2011, **86**(12), 1475–1479.
- 94 S. Bhadra, Y. S. Jiang, M. R. Kumar, R. F. Johnson, L. E. Hensley and A. D. Ellington, Real-time sequence-validated loop-mediated isothermal amplification assays for detection of Middle East respiratory syndrome coronavirus (MERS-CoV), *PLoS One*, 2015, **10**(4), e0123126.
- 95 Y. Chen, K. H. Chan, Y. Kang, H. Chen, H. K. Luk, R. W. Poon, *et al.*, A sensitive and specific antigen detection assay for Middle East respiratory syndrome coronavirus, *Emerging Microbes Infect.*, 2015, **4**(4), e26.
- 96 A. Liljander, B. Meyer, J. Jores, M. A. Muller, E. Lattwein, I. Njeru, *et al.*, MERS-CoV Antibodies in Humans, Africa, 2013–2014, *Emerging Infect. Dis.*, 2016, **22**(6), 1086–1089.
- 97 WHO, *Novel Coronavirus – China*, WHO, 2020, <https://www.who.int/csr/don/12-january-2020-novel-coronavirus-china/en/>.
- 98 N. Zhu, D. Zhang, W. Wang, X. Li, B. Yang, J. Song, *et al.*, A Novel Coronavirus from Patients with Pneumonia in China, 2019, *N. Engl. J. Med.*, 2020, **382**(8), 727–733.
- 99 T. Wenjie, Z. Xiang, M. Xuejun, W. Wenling, N. Peihua, X. Wenbo, *et al.*, *Notes from the Field: A Novel Coronavirus Genome Identified in a Cluster of Pneumonia Cases – Wuhan, China 2019–2020* *China CDC Weekly*, 2020.
- 100 WHO, *Naming the coronavirus disease (COVID-19) and the virus that causes it*, WHO, 2020, [https://www.who.int/emergencies/diseases/novel-coronavirus-2019/technical-guidance/naming-the-coronavirus-disease-\(covid-2019\)-and-the-virus-that-causes-it](https://www.who.int/emergencies/diseases/novel-coronavirus-2019/technical-guidance/naming-the-coronavirus-disease-(covid-2019)-and-the-virus-that-causes-it).
- 101 W. J. Guan, Z. Y. Ni, Y. Hu, W. H. Liang, C. Q. Ou, J. X. He, *et al.*, Clinical Characteristics of Coronavirus Disease 2019 in China, *N. Engl. J. Med.*, 2020, **382**(18), 1708–1720.
- 102 W. Wang, Y. Xu, R. Gao, R. Lu, K. Han, G. Wu, *et al.*, Detection of SARS-CoV-2 in Different Types of Clinical Specimens, *JAMA, J. Am. Med. Assoc.*, 2020, **323**(18), 1843–1844.
- 103 C. Rothe, M. Schunk, P. Sothmann, G. Bretzel, G. Froeschl, C. Wallrauch, *et al.*, Transmission of 2019-nCoV Infection from an Asymptomatic Contact in Germany, *N. Engl. J. Med.*, 2020, **382**(10), 970–971.
- 104 Worldometer, *COVID-19 Coronavirus Pandemic*, 2020, <https://www.worldometers.info/coronavirus/>.
- 105 V. M. Corman, O. Landt, M. Kaiser, R. Molenkamp, A. Meijer, D. K. Chu, *et al.*, Detection of 2019 novel coronavirus (2019-nCoV) by real-time RT-PCR, *Eurosurveillance*, 2020, **25**(3), 2000045.
- 106 T. Notomi, H. Okayama, H. Masubuchi, T. Yonekawa, K. Watanabe, N. Amino, *et al.*, Loop-mediated isothermal amplification of DNA, *Nucleic Acids Res.*, 2000, **28**(12), E63.
- 107 C. Yan, J. Cui, L. Huang, B. Du, L. Chen, G. Xue, *et al.*, Rapid and visual detection of 2019 novel coronavirus (SARS-CoV-2) by a reverse transcription loop-mediated isothermal amplification assay, *Clin. Microbiol. Infect.*, 2020, **26**(6), 773–779.
- 108 Y. H. Baek, J. Um, K. J. C. Antigua, J. H. Park, Y. Kim, S. Oh, *et al.*, Development of a reverse transcription-loop-mediated isothermal amplification as a rapid early-detection method for novel SARS-CoV-2, *Emerging Microbes Infect.*, 2020, **9**(1), 998–1007.
- 109 R. Lu, X. Wu, Z. Wan, Y. Li, L. Zuo, J. Qin, *et al.*, Development of a Novel Reverse Transcription Loop-Mediated Isothermal Amplification Method for Rapid Detection of SARS-CoV-2, *Virologica Sinica*, 2020, **35**, 344–347.
- 110 J. P. Broughton, X. Deng, G. Yu, C. L. Fasching, V. Servellita, J. Singh, *et al.*, CRISPR-Cas12-based detection of SARS-CoV-2, *Nat. Biotechnol.*, 2020, **38**, 870–874.
- 111 K. Uhteg, J. Jarrett, M. Richards, C. Howard, E. Morehead, M. Geahr, *et al.*, Comparing the analytical performance of three SARS-CoV-2 molecular diagnostic assays, *J. Clin. Virol.*, 2020, **127**, 104384.
- 112 A. Diagnostics, *Instructions for use RealStar SARS-CoV-2 RT-PCR Kit 1.0*, 2020.
- 113 ePLEX, *ePLEX SARS-CoV2 assay manual*, 2020.
- 114 WHO, *WHO in-house molecular assays*, 2020, https://www.who.int/docs/defaultsource/coronaviruse/whoinhouseassays.pdf?sfvrsn=de3a76aa_2.
- 115 K. Okamoto, K. Shirato, N. Nao, S. Saito, T. Kageyama, H. Hasegawa, *et al.*, An assessment of real-time RT-PCR kits for SARS-CoV-2 detection, *Jpn. J. Infect. Dis.*, 2020, **73**(5), 366–368.
- 116 J. F. Chan, C. C. Yip, K. K. To, T. H. Tang, S. C. Wong, K. H. Leung, *et al.*, Improved Molecular Diagnosis of COVID-19 by the Novel, Highly Sensitive and Specific COVID-19-RdRp/Hel Real-Time Reverse Transcription-PCR Assay Validated *In Vitro* and with Clinical Specimens, *J. Clin. Microbiol.*, 2020, **58**(5), e00310-20.
- 117 A. Moran, K. G. Beavis, S. M. Matushek, C. Ciaglia, N. Francois, V. Tesic, *et al.*, The Detection of SARS-CoV-2 using the Cepheid Xpert Xpress SARS-CoV-2 and Roche cobas SARS-CoV-2 Assays, *J. Clin. Microbiol.*, 2020, **58**(8), e00772-20.
- 118 Y. Zhang, N. Odiwuor, J. Xiong, L. Sun, R. O. Nyaruaba, H. Wei, *et al.*, Rapid Molecular Detection of SARS-CoV-2 (COVID-19) Virus RNA Using Colorimetric LAMP, *medRxiv*, 2020, 2020.02.26.20028373.
- 119 N. M. A. Okba, M. A. Muller, W. Li, C. Wang, C. H. GeurtsvanKessel, V. M. Corman, *et al.*, SARS-CoV-2 specific antibody responses in COVID-19 patients, *medRxiv*, 2020, 2020.03.18.20038059.
- 120 T. Hoffman, K. Nissen, J. Krambrich, B. Ronnberg, D. Akaberi, M. Esmaeilzadeh, *et al.*, Evaluation of a COVID-19 IgM and IgG rapid test; an efficient tool for assessment of past exposure to SARS-CoV-2, *Infect. Ecol. Epidemiology*, 2020, **10**(1), 1754538.
- 121 Y. Gao, Y. Yuan, T. T. Li, W. X. Wang, Y. X. Li, A. Li, *et al.*, Evaluation the auxiliary diagnosis value of antibodies assays for detection of novel coronavirus (SARS-Cov-2) causing an outbreak of pneumonia (COVID-19), *medRxiv*, 2020, 2020.03.26.20042044.
- 122 D. Shan, J. M. Johnson, S. C. Fernandes, M. Mendes, H. Suib, M. Holdridge, *et al.*, SARS-Coronavirus-2 nucleocapsid protein measured in blood using a Simoa ultra-sensitive immunoassay differentiates COVID-19



- infection with high clinical sensitivity, *medRxiv*, 2020, 2020.08.14.20175356.
- 123 K. Zwirgmaier, M. Weyh, C. Krueger, R. Ehmann, K. Mueller, R. Woelfel, *et al.*, Rapid detection of SARS-CoV-2 by pulse-controlled amplification (PCA), *medRxiv*, 2020, 2020.07.29.20154104.
- 124 W. S. Yoo, H. S. Han, J. G. Kim, K. Kang, H.-S. Jeon, J.-Y. Moon, *et al.*, Development of a tablet PC-based portable device for colorimetric determination of assays including COVID-19 and other pathogenic microorganisms, *RSC Adv.*, 2020, **10**(54), 32946–32952.
- 125 G. Xue, S. Li, W. Zhang, B. Du, J. Cui, C. Yan, *et al.*, Reverse-Transcription Recombinase-Aided Amplification Assay for Rapid Detection of the 2019 Novel Coronavirus (SARS-CoV-2), *Anal. Chem.*, 2020, **92**(14), 9699–9705.
- 126 J. Liu, M. A. Eddings, A. R. Miles, R. Bukasov, B. K. Gale and J. S. Shumaker-Parry, *In Situ* Microarray Fabrication and Analysis Using a Microfluidic Flow Cell Array Integrated with Surface Plasmon Resonance Microscopy, *Anal. Chem.*, 2009, **81**(11), 4296–4301.
- 127 R. Pilot, R. Signorini, C. Durante, L. Orian, M. Bhamidipati and L. Fabris, A Review on Surface-Enhanced Raman Scattering, *Biosensors*, 2019, **9**(2), 57.
- 128 H. C. Metsky, C. A. Freije, T.-S. F. Kosoko-Thoroddsen, P. C. Sabeti and C. Myhrvold, CRISPR-based surveillance for COVID-19 using genomically-comprehensive machine learning design, *bioRxiv*, 2020, 2020.02.26.967026.
- 129 J. D. Driskell, K. M. Kwarta, R. J. Lipert, M. D. Porter, J. D. Neill and J. F. Ridpath, Low-Level Detection of Viral Pathogens by a Surface-Enhanced Raman Scattering Based Immunoassay, *Anal. Chem.*, 2005, **77**(19), 6147–6154.
- 130 M. Peplow, *Saliva tests show promise for widespread COVID-19 surveillance at universities and workplaces C. E. N.*, 2020, <https://cen.acs.org/analytical-chemistry/diagnostics/Saliva-tests-show-promise-widespread/98/web/2020/08>.
- 131 M. Peplow, *Rapid COVID-19 testing breaks free from the lab C. E. N.*, 2020, <https://cen.acs.org/analytical-chemistry/diagnostics/Rapid-COVID-19-testing-breaks/98/web/2020/08>.

