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## Fatty acid based transient nanostructures for temporal regulation of artificial peroxidase activity†

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Natural systems access transient high energy self-assembled structures for temporal regulation of different biological functions through dissipative processes. Compartmentalization within self-assembled structures is used by living systems to organize vital biochemical reactions that define cellular metabolism. Herein, we demonstrate a simple fatty acid based system where a redox active base (dimethylaminomethyl ferrocene, **Fc-NMe<sub>2</sub>**) acts as a counteraction to access unique hexagonal compartments resulting in the formation of a self-supporting gel. An oxidizing environment helps in the dissipation of energy by converting **Fc-NMe<sub>2</sub>** to oxidized waste and the gel autonomously undergoes transition to a sol. Hence, the system requires the addition of the fuel **Fc-NMe<sub>2</sub>** to access the temporal gel state. Notably, these transient compartments were able to temporally upregulate and downregulate hemin-catalyzed oxidation reactions mimicking peroxidase, a ubiquitous enzyme in extant biology. An order of magnitude variation in  $k_{\text{cat}}$  values was observed with time and the chemical reaction persists as long as the gel state was present.

### Introduction

Natural systems have exceptional capabilities to access transient non-equilibrium nanostructures by energy consuming processes.<sup>1–4</sup> Through remarkable spatiotemporal control over the size and conformation in the nanoscale domain, biological systems access these high-energy transient morphologies which are utilized to perform work. For instance, vital functions such as cell motility are achieved by transient length changes of microtubules and actin filaments in the presence of nucleoside triphosphates as a chemical fuel.<sup>5,6</sup> Elegant strategies to access out of equilibrium artificial synthetic systems have utilized energy from light or chemical energy stored in high-energy, kinetically stable molecules which are termed as chemical fuels.<sup>7–27</sup> As such, there is mounting interest to develop synthetic transient assemblies that are capable of displaying temporal control of functions foreshadowing the pathways followed by living systems. To this end, we were interested to develop fuel driven temporal generation of compartments or confinements which are capable of controlling the chemical reactivity of another reaction. Nature uses the strategy of compartmentalization to organize essential chemical reactions that define cellular metabolism.<sup>28</sup> Many of these reactions if performed in a single environment without compartments would produce futile chemical cycles, undesired by-products, or suffer from unfavourable thermodynamics from the bulk

environment.<sup>29</sup> Compartments are also used to store and regulate enzymes in extant biology.<sup>30</sup> Interestingly, reports have argued the importance of transient compartmentalization in early Earth to protect RNA replicators from parasitic mutants.<sup>31</sup> We asked whether a transient compartment of natural fatty acids can be achieved under dissipative conditions and if the transient changes of the microenvironment would lead to temporal control of the entrapped guest, as seen in organelles in living systems.

### Results and discussion

To achieve the transient formation of compartments, the challenge is to find a subtle balance of activation–deactivation rates, where the rate of the reaction which will create the proposed compartment should be faster than the deactivation reaction leading to its degradation.<sup>7–9</sup> For the fast activation step, we envisaged to use a redox responsive metastable base which can react with the fatty acid to form a salt that undergoes thermodynamically driven assembly (Fig. 1). For the slow deactivation step, experimental conditions can be set up which promote dissipation by gradual oxidation of the counteraction, leading to transient accumulation of the proposed compartment as shown in the reaction scheme (Fig. 1a and b and the representative SEM image of the fibre like morphology is shown in Fig. 1c). Thus the redox responsive base will work as the fuel to drive the transient assembly. To test our hypothesis, we mixed stearic acid (**C18A**) and dimethylaminomethyl ferrocene (**Fc-NMe<sub>2</sub>**) at varying molar ratio in DMSO/water (10%, v/v). Although **C18A** and **Fc-NMe<sub>2</sub>** are separately not soluble in 10% v/v DMSO/water, their mixtures dissolved in molar ratios of

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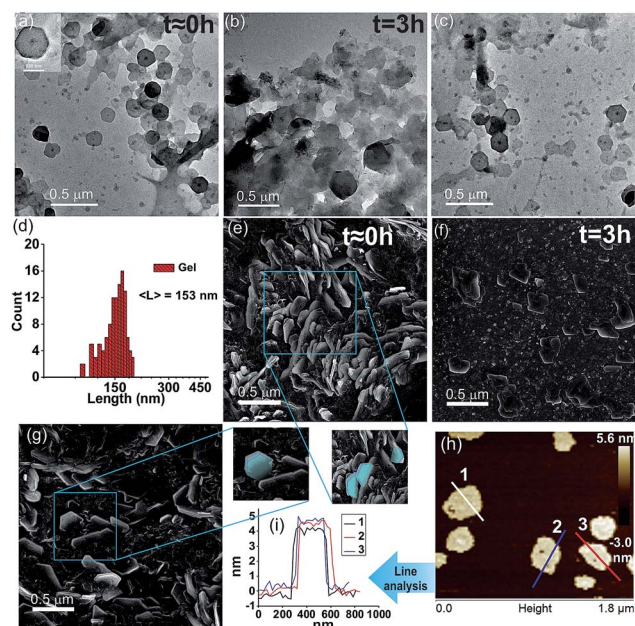


**Fig. 1** (a) Reaction scheme used in the present work and (b) schematic representation of a typical reaction cycle. Fuel driven transient assembly is shown along with the model of packing of interdigitated fatty acids, predicted from atomic force microscopy (AFM). The average of height profiles obtained from 15 randomly chosen hexagon like morphologies is calculated as  $4.5 \pm 0.2$  nm. The average size of the hexagon is about 153 nm (distance between the apexes) as obtained from transmission electron microscopy (TEM). (c) SEM image of the transient assembly showing fibre like morphology at 30 min, (d) chemical structures of the molecules involved and (e–j) representative images of temporal sol–gel–sol transition cycles prepared with **C18A** (245 mM) with **Fc-NMe<sub>2</sub>** (245 mM) and **Fe(ClO<sub>4</sub>)<sub>3</sub>** (10 mM) in 10% (v/v) DMSO/water.

1 : 0.75, 1 : 1 and 1 : 1.25 (**C18A** : **Fc-NMe<sub>2</sub>**), suggesting colloidal stability of the organic salt. The mixtures with molar ratios of  $\approx$  1 : 1 gradually turned more viscous suggesting assembly (see the ESI† for details). Notably, when we increased the concentration of the acid and counterion (*ca.* 210 mM, 1 : 1), rapid gelation was observed within 25 s with viscoelastic behavior (Fig. S1–S3, ESI†). Since ferrocene is known to acutely respond to changes in the redox environment, to design the deactivation step, the acid–base reaction was done under conditions promoting its oxidation (Fig. 1). Orthogonal routes of activation (salt formation) and deactivation (oxidation of the counterion) would give us the opportunity to manipulate the kinetics of the competing reactions, leading to accumulation of the transient assemblies under dissipative conditions (Fig. 1).<sup>7–9,22</sup> Clearly,

since both the acid–base neutralization and subsequent gelation are fast steps, the transient accumulation of the assembled structures will be determined by the rate limiting oxidation of the cation by the environment. In order to find out the suitable rate of deactivation, the acid and base were mixed in a 1 : 1 ratio (245 mM each) in the presence of varying concentrations of **Fe(ClO<sub>4</sub>)<sub>3</sub>**, a common oxidizing agent for ferrocene (Fig. S4, ESI†). The pH of the system was measured to be in the range of 8.1 to 8.3. From 2 mM to 12.5 mM, gels were formed but interestingly showed autonomous disassembly (Fig. 1e–j). Due to faster oxidation in the presence of a higher amount of the oxidant, the lifetimes of these transient gels showed a gradual decrease from *ca.* 24 h to 2 h (Fig. S4, ESI†). Also, the time required for gel formation increased with the increase in the concentration of the oxidant, from about a minute to up to *ca.* 16 min (Fig. S4, ESI†). With the increase of the oxidant concentration, the oxidation of the fuel and thus the deactivation kinetics, *i.e.* the disassembly process, are expedited as waste is generated before the formation of the organic salt. This eventually retards the gelation process as reflected from slow gelation. Finally, when the concentration of the oxidant reaches 15 mM or beyond, no gelation was observed. Rheology done in the presence of different oxidant concentrations showed a decrease of the storage modulus ( $G'$ ) with an increase in the **Fe(ClO<sub>4</sub>)<sub>3</sub>** concentration (Fig. S5, ESI†).

We were intrigued to investigate the morphology of the transient fatty acid assemblies directed in the presence of an aromatic counteranion. Transmission electron microscopy (TEM) performed at different time intervals revealed distinct



**Fig. 2** TEM images of (a) **C18A-Fc-NMe<sub>2</sub>** for  $t \approx 0$  h gel (inset: enlarged view), (b)  $t = 3$  h sol, and (c) refueled gel (4 mM **Fc-NMe<sub>2</sub>**). (d) Histogram of hexagon dimensions from the TEM results at  $t \approx 0$  h. SEM images (e) at  $t \approx 0$  h with blue traces showing the hexagonal structures, (f)  $t = 3$  h sol, (g) refueled gel (5 min), and (h) AFM image at  $t \approx 0$  h and (i) the corresponding height profiles.



hexagonal morphologies at  $t \approx 0$  h with an average diameter of *ca.* 153 nm (Fig. 2a, histograms of distance between the apexes in Fig. 2d). Interestingly, TEM done at  $t = 3$  h showed deformed hexagons along with few intact hexagonal structures, underpinning the transient nature of the assemblies (Fig. 2b). Scanning electron microscopy (SEM) also showed similar morphologies of stacks of hexagonal nanostructures at  $t = 0$  h (Fig. 2g and S6, ESI†) and autonomous deformation at  $t = 3$  h (Fig. 2e and f). Atomic force microscopy (AFM, Fig. 2h and i) at  $t \approx 0$  h showed uniform height profiles in the range of  $4.5 \pm 0.2$  nm, indicating the interdigitation of two fatty acid chains (Fig. 1b). These hexagons formed from the mixing of **Fc-NMe<sub>2</sub>** and **C18A** acid underwent further higher order assembly leading to the formation of fibre like intertwined structures (Fig. 1c and S6, ESI†).

We monitored the rate of conversion of **Fc-NMe<sub>2</sub>** to the oxidized waste **Fc(+)-NMe<sub>2</sub>** by HPLC (Fig. 3a and b, with 25 times diluted samples, see the ESI† for details). A gradual decrease in the **Fc-NMe<sub>2</sub>** area was observed with time. At  $t = 3$  h, when the gel melts autonomously, the extent of decrease corresponded to 6 mM **Fc-NMe<sub>2</sub>** (calculated from the standard plot, Fig. S7, ESI†), thus suggesting the presence of 239 mM **Fc-NMe<sub>2</sub>**. Despite the presence of a substantially larger amount (239 mM) of residual **Fc-NMe<sub>2</sub>**, the collapse of the gel is in contrast to the finding that 210 mM was sufficient for gelation in the absence of an oxidising environment. Hence, these results underpin the role of oxidation from the assembled state. As expected, when we added a 1 : 1 mixture of **C18A-Fc(+)-NMe<sub>2</sub>**, no gel formation was observed. To find out the role of self-assembled state, HPLC

of the **C3A** salt of **Fc-NMe<sub>2</sub>** was done. **C3A-Fc-NMe<sub>2</sub>** did not show any viscous behavior and did not assemble (TEM, Fig. S8, ESI†). Intriguingly, the consumption after 60 min in the case of **C3A-Fc-NMe<sub>2</sub>** salt was *ca.* 9.2 mM while it was only 2.9 mM for **C18A-Fc-NMe<sub>2</sub>** (Fig. 3a). These data suggested that the formation of the assembled hexagons reduces the kinetics of oxidation and increases the lifetime of the gel. To further understand the role of the transient assembled state, we simulated the disassembly by premixing different ratios of waste **Fc(+)-NMe<sub>2</sub>** with the fuel **Fc-NMe<sub>2</sub>**, keeping the total concentration of **Fc(+)-NMe<sub>2</sub>** and **Fc-NMe<sub>2</sub>** fixed at 245 mM (Fig. 3c and S9, ESI†). These were added to **C18A** in the absence of the oxidizing agent. The concentration of **Fc(+)-NMe<sub>2</sub>** was kept from 0–15 mM as suggested from HPLC. As expected, only viscous liquids (VLs) or free flowing solutions were observed. This result indicated that the instability of the assembled structures arises due to the presence of oxidized cations as the ion pair of **C18A**. Interestingly, when the same experiment was carried out by adding the oxidized waste **Fc(+)-NMe<sub>2</sub>** after the formation of the gel of **C18A** and **Fc-NMe<sub>2</sub>**, the gels remained intact in the entire concentration range (Fig. 3c). Hence, when the assembled structures are already formed, the oxidized waste is unable to exchange with the fuel to be an ion pair of **C18A**. This result along with the slower oxidation kinetics for the assembled state (Fig. 3a), underpinned the fact that the oxidative conversion of the fuel to waste occurs from the assembled state in the **C18A-Fc-NMe<sub>2</sub>** system which subsequently results in instability.<sup>7–9</sup> This was further supported from SEM images where ordered structures were seen when 5 mM of **Fc(+)-NMe<sub>2</sub>** (Fig. 3d) was added after



Fig. 3 (a) Comparison of the consumption kinetics of **Fc-NMe<sub>2</sub>** in salts with **C18A** and **C3A**, (b) representative HPLC chromatogram of the consumption of **Fc-NMe<sub>2</sub>** with **C18A** at  $t \approx 0$  and  $t = 4$  h, and (c) effect of the addition of **Fc(+)-NMe<sub>2</sub>** in varying ratios to the pre-assembled and post-assembled mixture of **C18A** and **Fc-NMe<sub>2</sub>** (total concentration of **Fc-NMe<sub>2</sub>** and **Fc(+)-NMe<sub>2</sub>** kept constant at 245 mM). Representative SEM images of (d) gel state when oxidized waste (5 mM) was added after assembly and (e) when oxidized waste (5 mM) was added before assembly. (f) Time resolved rheology of gel-sol-gel cycles. The red arrow shows the refueling step with **Fc-NMe<sub>2</sub>** (4 mM). The error bars are the result of triplicate measurements.





assembly. In contrast, when the same amount of **Fc(+)-NMe<sub>2</sub>** was added before assembly, less ordered structures were seen (Fig. 3e). Since the fatty acid exhibited fidelity towards the fuel over the oxidized waste for assembly (Fig. 3c and S9, ESI†), we expected the system to display multiple sol to gel to sol cycles upon refuelling. Indeed, when **Fc-NMe<sub>2</sub>** (4 mM) was added again to a representative system (245 mM, 1 : 1 with 10 mM oxidant), reformation of the gel was observed (Fig. 1g–j). Remarkably, TEM and SEM showed the reappearance of hexagonal nanostructures (Fig. 2c and g, histogram in Fig. S10, ESI†). The pH remained in the range of pH  $8.3 \pm 0.1$ . The sol–gel–sol transition could be performed three times before the accumulation of waste and consumption of the oxidant resulted in no further gelation. This autonomous sol–gel–sol transition could also be seen from the viscoelastic behaviour. The samples with different ages and with refuelling batches were subjected to rheology which displayed temporal changes in the storage modulus,  $G'$  (Fig. 3f).

The temporal existence of the hexagonal fatty acid assemblies prompted us to investigate their capability of entrapping small molecules and hosting guests. Fluorophore pyrene suffered rapid quenching when added to the gel of **C18A-Fc-NMe<sub>2</sub>** (Fig. S11, ESI†) due to the known capability of ferrocene to quench pyrene's fluorescence.<sup>32,33</sup> Interestingly, the extent of quenching was substantially less for the **C3A-Fc-NMe<sub>2</sub>** system which does not assemble, suggesting closer packing and entrapment in the case of the assembled structures (Fig. S11, ESI†). We asked whether these hexagonal compartments can entrap hemin, the prosthetic group of extant peroxidases and metalloproteins, to mimic and possibly regulate temporally the artificial peroxidase activity.<sup>32–34</sup>

Temporal enzyme regulation through compartmentalization is a feature seen in extant biology.<sup>30,34–36</sup> To investigate the interaction of hemin in transient systems, UV-vis spectroscopy was employed (Fig. 4a). In 10% DMSO/water, free hemin showed a Soret band at 392 nm and a shoulder at 365 nm along with a low intensity band at around 610 nm, indicating the presence hemin dimers connected through  $\mu$ -oxo bridges.<sup>35,37</sup> In the **C18A-Fc-NMe<sub>2</sub>** gel at  $t = 0$  h, the Soret band red shifted to 401 nm indicating the existence of monomeric hemin and thus

supporting the entrapment of hemin.<sup>35,37</sup> In the sol state at  $t = 3$  h, hemin still retained the Soret peak at 401 nm (Fig. 4a).

For the investigation of hemin activity, pyrogallol was used as a standard substrate in the presence of 1 mM histidine and  $\text{H}_2\text{O}_2$  as the oxidant.<sup>35</sup> Intriguingly, hemin mixed with the **C18A-Fc-NMe<sub>2</sub>** gel showed temporal changes in the oxidation rates, and displayed autonomous upregulation and down-regulation of peroxidase like activity (Fig. 4b and c and the ESI† for details). The upregulated activity at  $t \approx 0$  h ( $k_{\text{cat}} = 10.6 \pm 1 \text{ s}^{-1}$ , Fig. S12, Table S1, ESI†) was significantly higher than the activity of free hemin.<sup>35</sup> This activity showed autonomous decrease and dropped to  $k_{\text{cat}} = 0.84 \pm 0.2$  in 3 h (Fig. S13, Table S1, ESI†). Specific constants were found to decrease from  $0.35 \pm 0.03$  in the gel state to  $0.19 \pm 0.02 \text{ mM}^{-1} \text{ s}^{-1}$  in the sol state (Table S1, ESI†). The reason behind the temporal increase of the  $k_{\text{cat}}$  values can be attributed to the transient formation of amphiphilic self-assembled nanostructures. Nanostructured gels have been previously shown to activate hemin and proteins such as peroxidases and hydrolases.<sup>34,35,38–40</sup> On autonomous gel to sol transformation, the density of amphiphilic nanostructures decreased, resulting in less activity in the sol state (Fig. 2b and f).<sup>34,35,38–40</sup> Although, the temporal upregulation and downregulation of activity showed a modest range of an order of magnitude in terms of  $k_{\text{cat}}$  values, this result underpins the subtle but significant role of the transiently assembled compartments in the artificial peroxidase activity. Furthermore, after each refueling, the  $k_{\text{cat}}$  values showed two spikes of  $8.3 \pm 0.8$  and  $8.06 \pm 0.7 \text{ s}^{-1}$  respectively (Fig. 4c and S12, ESI†). Interestingly, the specific constants after refueling also registered a recovery up to  $0.32 \pm 0.04 \text{ mM}^{-1} \text{ s}^{-1}$  (Table S1, ESI†). The  $k_{\text{cat}}$  values subsequently decrease again to  $0.67 \pm 0.1$  and  $0.65 \pm 0.1 \text{ s}^{-1}$  within 1.5 and 1 h respectively (Fig. 4c). Controls studied with transient gels in the absence of  $\text{H}_2\text{O}_2$  but in the presence of  $\text{Fe}(\text{ClO}_4)_3$  showed formation of miniscule amounts of oxidation products under the experimental conditions. This suggested that  $\text{H}_2\text{O}_2$  is the only oxidant under the experimental conditions. Furthermore, the control system of **C3A-Fc-NMe<sub>2</sub>** studied in this range did not show such temporal variation of activity (Fig. 4c and Table S1, ESI†). The pH increase of  $\sim 0.1$  units after each refueling step (4 mM **Fc-NMe<sub>2</sub>**) did not have an impact on the activity of hemin in the assembled systems.

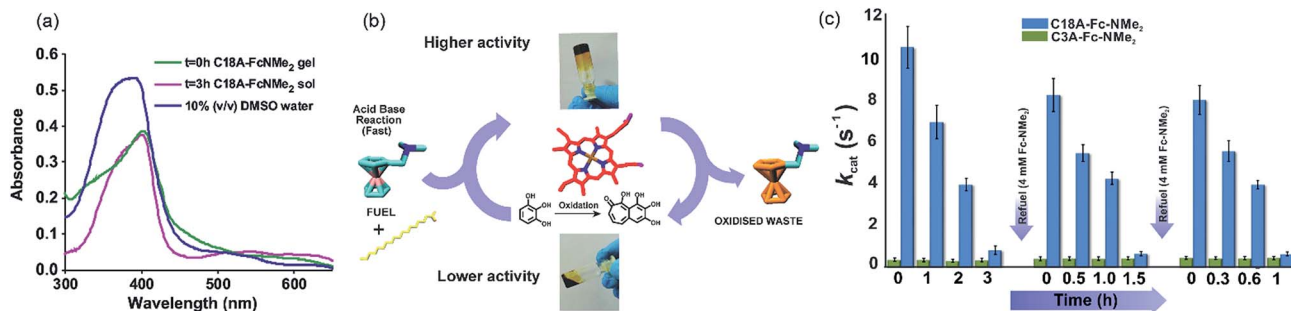


Fig. 4 (a) UV-vis spectra of free hemin and hemin entrapped **C18A-Fc-NMe<sub>2</sub>** gel at  $\approx 0$  h and 3 h. All samples had histidine (1 mM). (b) Schematic representation of the temporal regulation of artificial peroxidase activity of hemin entrapped in the transient system. (c)  $k_{\text{cat}}$  values of hemin (0.1 mM) catalyzed oxidation of pyrogallol (varying concentration) by  $\text{H}_2\text{O}_2$  (30 mM) in the presence of 1 mM histidine for **C18A-Fc-NMe<sub>2</sub>** and **C3A-Fc-NMe<sub>2</sub>** systems. The error bars are the result of triplicate measurements.



## Conclusions

In conclusion, we have developed fatty acid based transient hexagonal nanostructured assemblies which dissipate energy from the conversion of chemical fuels to oxidized waste. In an oxidizing environment, the high-energy self-supporting gel state is stable only when the fuel is supplied. Furthermore, these hexagonal nanostructures with transient lifetimes were able to host hemin and temporally upregulate and down-regulate the peroxidase mimicking activity up to an order of magnitude, thus foreshadowing the compartmentalized biochemical processes seen in extant biology.

## Conflicts of interest

There are no conflicts to declare.

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## Notes and references

- 1 J. H. van Esch, R. Klajn and S. Otto, *Chem. Soc. Rev.*, 2017, **46**, 5474–5475.
- 2 B. A. Grzybowski and W. T. S. Huck, *Nat. Nanotechnol.*, 2016, **11**, 585–592.
- 3 E. Mattia and S. Otto, *Nat. Nanotechnol.*, 2015, **10**, 111–119.
- 4 G. M. Whitesides and B. Grzybowski, *Science*, 2002, **295**, 2418–2421.
- 5 H. Hess and J. L. Ross, *Chem. Soc. Rev.*, 2017, **46**, 5570–5587.
- 6 M. Caplow and J. Shanks, *J. Biol. Chem.*, 1990, **265**, 8935–8941.
- 7 R. D. Astumian, *Nat. Nanotechnol.*, 2012, **7**, 684–688.
- 8 C. Pezzato, C. Cheng, J. F. Stoddart and R. D. Astumian, *Chem. Soc. Rev.*, 2017, **46**, 5491–5507.
- 9 C. Cheng, P. R. McGonigal, J. F. Stoddart and R. D. Astumian, *ACS Nano*, 2015, **9**, 8672–8688.
- 10 G. Ragazzon and L. J. Prins, *Nat. Nanotechnol.*, 2018, **13**, 882–889.
- 11 S. De and R. Klajn, *Adv. Mater.*, 2018, **30**, 1706750.
- 12 H. Zhao, S. Sen, T. Udayabhaskararao, M. Sawczyk, K. Kučanda, D. Manna, P. K. Kundu, J.-W. Lee, P. Král and R. Klajn, *Nat. Nanotechnol.*, 2016, **11**, 82–88.
- 13 S. Maiti, I. Fortunati, C. Ferrante, P. Scrimin and L. J. Prins, *Nat. Chem.*, 2016, **8**, 725–731.
- 14 A. Sorrenti, J. Leira-Iglesias, A. J. Markvoort, T. F. A. de Greef and T. M. Hermans, *Chem. Soc. Rev.*, 2017, **46**, 5476–5490.
- 15 J. K. Sahoo, C. G. Pappas, I. R. Sasselli, Y. M. Abul-Haija and R. V. Ulijn, *Angew. Chem., Int. Ed.*, 2017, **56**, 6828–6832.
- 16 J. Boekhoven, A. Brizard, K. Kowli, G. Koper, R. Eelkema and J. H. van Esch, *Angew. Chem., Int. Ed.*, 2010, **49**, 4825–4828.
- 17 N. Singh, M. P. Conte, R. V. Ulijn, J. F. Miravet and B. Escuder, *Chem. Commun.*, 2015, **51**, 13213–13216.
- 18 J. Boekhoven, W. Hendriksen, G. Koper, R. Eelkema and J. H. van Esch, *Science*, 2015, **349**, 1075–1079.
- 19 P. S. Muñana, G. Ragazzon, J. Dupont, C. Z.-J. Ren, L. J. Prins and J. L.-Y. Chen, *Angew. Chem., Int. Ed.*, 2018, **57**, 16469–16474.
- 20 S. Bal, K. Das, S. Ahmed and D. Das, *Angew. Chem., Int. Ed.*, 2019, **58**, 244–247.
- 21 S. Debnath, S. Roy and R. V. Ulijn, *J. Am. Chem. Soc.*, 2013, **135**, 16789–16792.
- 22 M. Kumar, N. L. Ing, V. Narang, N. K. Wijerathne, A. L. Hochbaum and R. V. Ulijn, *Nat. Chem.*, 2018, **10**, 696–703.
- 23 J. P. Wojciechowski, A. D. Martin and P. Thordarson, *J. Am. Chem. Soc.*, 2018, **140**, 2869–2874.
- 24 S. Dhiman, A. Jain and S. J. George, *Angew. Chem., Int. Ed.*, 2017, **56**, 1329–1333.
- 25 S. Dhiman, A. Jain, M. Kumar and S. J. George, *J. Am. Chem. Soc.*, 2017, **139**, 16568–16575.
- 26 A. Sorrenti, J. Leira-Iglesias, A. Sato and T. M. Hermans, *Nat. Commun.*, 2017, **8**, 15899.
- 27 D. Spitzer, L. L. Rodrigues, D. Straßburger, M. Mezger and P. Besenius, *Angew. Chem., Int. Ed.*, 2017, **56**, 15461–15465.
- 28 J. Leira-Iglesias, A. Tassoni, T. Adachi, M. Stich and T. M. Hermans, *Nat. Nanotechnol.*, 2018, **13**, 1021–1027.
- 29 F. Hinzpeter, U. Gerland and F. Tostevin, *Biophys. J.*, 2017, **112**, 767–779.
- 30 R. P. Goodman, S. E. Calvo and V. K. Mootha, *J. Biol. Chem.*, 2018, **293**, 7508–7516.
- 31 M. S. Cuesta, S. A. Rahman, N. Furnham and J. M. Thornton, *Biophys. J.*, 2015, **109**, 1082–1086.
- 32 S. Matsumura, Á. Kun, M. Ryckelynck, F. Coldren, A. Szilágyi, F. Jossinet, C. Rick, P. Nghe, E. Szathmáry and A. D. Griffiths, *Science*, 2016, **354**, 1293–1296.
- 33 S. Basak, I. Singh, A. Ferranco, J. Syed and H. B. Kraatz, *Angew. Chem., Int. Ed.*, 2017, **56**, 13288–13292.
- 34 Q. Wang, Z. Yang, L. Wang, M. Ma and B. Xu, *Chem. Commun.*, 2007, 1032–1034.
- 35 Q. Wang, Z. Yang, X. Zhang, X. Xiao, C. K. Chang and B. Xu, *Angew. Chem., Int. Ed.*, 2007, **46**, 4285–4289.
- 36 P. T. T. Nguyen, A. J. Conley, J. Sneyd, R. S. F. Lee, T. K. Soboleva and P. R. Shorten, *J. Theor. Biol.*, 2013, **332**, 52–64.
- 37 P. K. Shantha, G. S. S. Saini, H. H. Thanga and A. L. Verma, *J. Raman Spectrosc.*, 2001, **32**, 159.
- 38 Q. Liu, H. Wang, X. H. Shi, Z. G. Wang and B. Q. Ding, *ACS Nano*, 2017, **11**, 7251–7258.
- 39 T. Bhattacharyya, Y. P. Kumar and J. Dash, *ACS Biomater. Sci. Eng.*, 2017, **3**, 2358–2365.
- 40 N. Kapil, A. Singh and D. Das, *Angew. Chem., Int. Ed.*, 2015, **54**, 6492.

