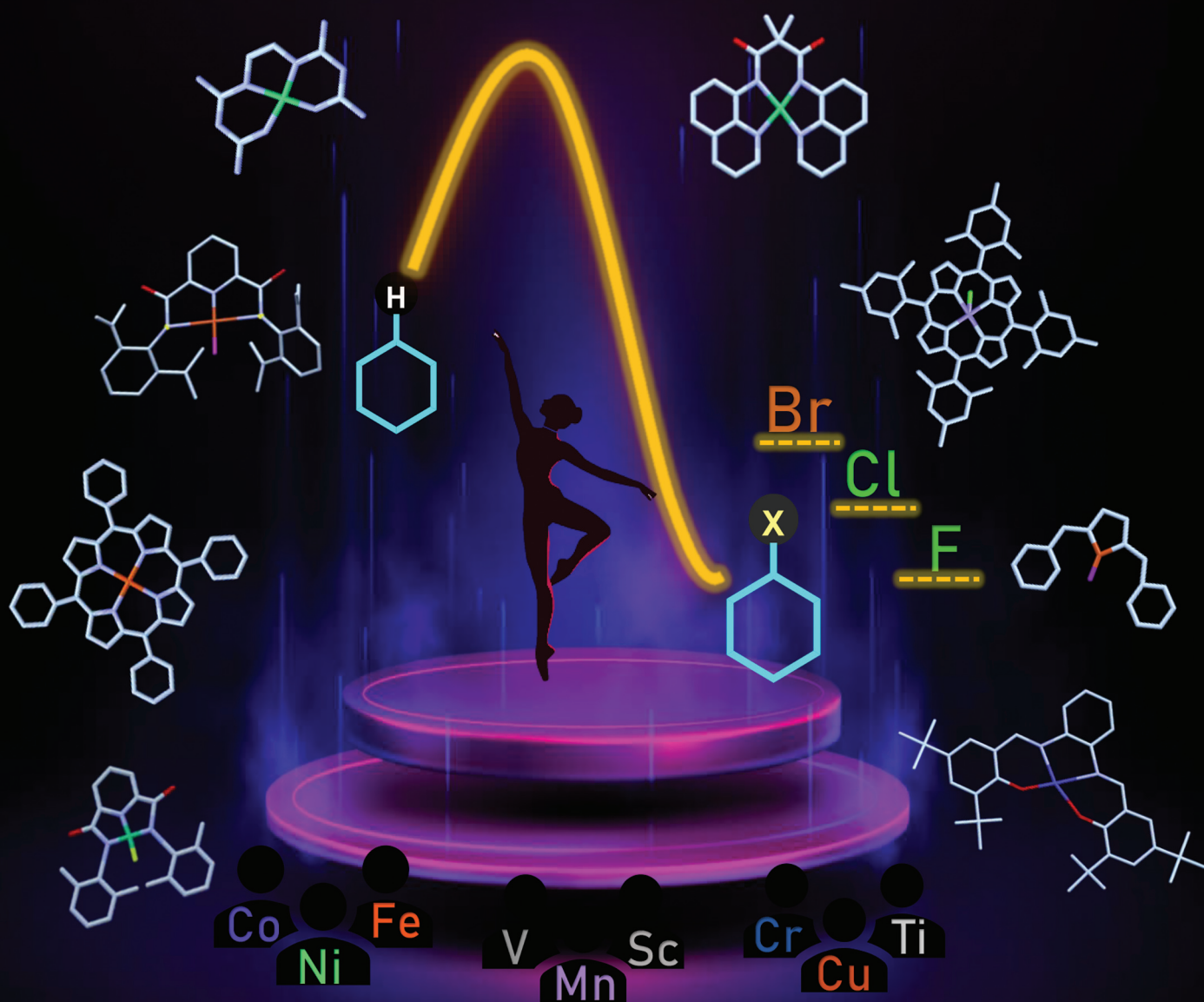


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C(sp³)-H halogenation using first-row transition metal catalysts under homogeneous conditions

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The direct functionalization of unactivated C–H bonds has emerged as a promising strategy to achieve step-economical, atom-efficient, and environmentally benign chemical processes. In this context, ongoing research is focused on the design and development of biomimetic homogeneous catalysts based on first-row transition metals for C(sp³)-H halogenation under mild conditions. In essence, this review highlights key advances in catalyst design based on first-row transition metals, showcasing their growing utility in the development of new synthetic methods.

1. Introduction

Halogenated organic compounds are essential in modern society (Fig. 1). Approximately 85% of pharmaceutical drugs involve chlorine or chlorinated compounds in their synthesis.^{1a-c} The presence of carbon-halogen bonds in both synthetic active pharmaceutical ingredients and natural compounds enhances their stability, resistance to biodegradation and oxidation, biological activity, and membrane permeability.

In the agrochemical sector, more than 90% of herbicides, fungicides, insecticides, acaricides, and nematicides produced since 2010 contain halogen atoms.^{1a,d} For example, imidacloprid, a neonicotinoid used to control pests like sucking insects, soil insects, termites and fleas, is the most widely used insecticide globally.² To date, over 5000 naturally occurring organic halogenated compounds have been identified and characterized.³ An example is vancomycin, a chlorine-containing antibiotic from *Streptomyces orientalis*, used to treat methicillin-resistant *Staphylococcus aureus* (MRSA) infections.⁴ In 2021, marking the 50th anniversary of the National Cancer Act, the FDA approved six halogenated compounds, including tivozanib, sotorasib, melphalan flufenamide, asciminib, infirgratinib, and umbralisib.⁵ Fluorine is also widely used in

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frameworks for organic transformations such as C–H hydroxylation, halogenation, and asymmetric halogenation, along with late-stage functionalization reactions, and electrochemical oxidation.



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Jaipriya Khatri joined the Chemistry Department at SNIOE, Delhi NCR, as a Ph.D. student in 2021. She joined the Catalysis Lab under the supervision of Dr Basab Bijayi Dhar after receiving her bachelor's and master's degrees from the University of Rajasthan. Her doctoral research focuses on the development of first-row transition metal-based homogeneous and heterogeneous catalysts for C–H activation and the electrochemical CO₂ reduction reaction (ECCO₂RR) in aqueous media.



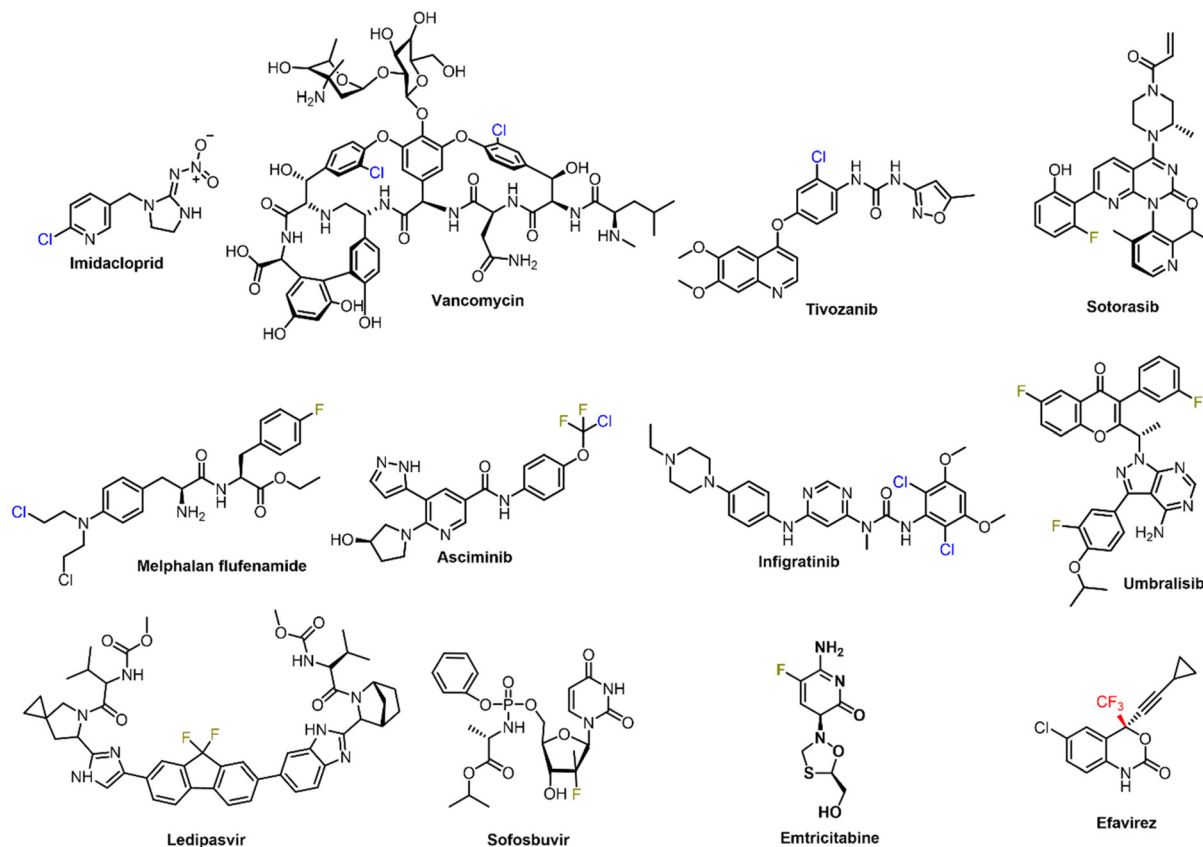


Fig. 1 Biologically important halogenated compounds.

modern medicinal chemistry.⁶ Ledipasvir/sofosbuvir, a fluorinated drug marketed as Harvoni, is used to treat chronic hepatitis C.⁷ In 2021, eight out of 14 newly approved drugs by the FDA contained fluorine.⁸ Triple combination therapy is now the standard treatment for HIV infection, with one of the most successful combinations being Atripla, which contains two fluorinated drugs, such as emtricitabine and efavirenz, along with tenofovir.⁹ Emtricitabine¹⁰ and efavirenz¹¹ are nucleoside and non-nucleoside inhibitors of HIV-1 reverse transcriptase,

the enzyme responsible for viral replication. In addition to these uses, various fluorinated and iodinated compounds are employed in radiotherapy and biomedical imaging.

From a synthetic chemistry standpoint, organohalides are typically produced by adding molecular halogens, hydrohalogenation, or using reagents that promote nucleophilic substitution and electrophilic aromatic substitution.¹² These methods can present environmental and selectivity challenges due to the reactive, toxic, and corrosive nature of the reagents



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involved, as well as the potential for creating complex mixtures of products. However, in nature, thousands of halogenated compounds are exclusively produced by specialised metalloenzymes, such as the Fe(II)/ α KG-dependent enzyme SyrB2¹³ and heme- and vanadium-dependent haloperoxidases.¹⁴ Flavin-dependent halogenases also show promise as biocatalysts for the regioselective halogenation of aromatic compounds.¹⁵ Inspired by these natural enzymes, many research groups have explored various first-row transition metal complexes to activate inert C–H bonds. First-row transition metals are highly effective single-electron transfer (SET) agents, making them particularly well-suited for promoting radical-based C(sp³)–H functionalization. In this review, we will report on the development of first-row transition metal catalysts for halogenation under homogeneous conditions, exploring their promising potential for enabling C–H activation and diversification in synthetic chemistry.

2. Enzymatic C–H halogenation

In nature, a range of enzymes are known to insert halogens into unactivated starting materials. Halohydrin dehalogenases (HHDHs), which have no transition metals, catalyse the reversible interconversion of epoxides into halohydrins.¹⁶ Industrially, HHDHs are used in the production of chiral epichlorohydrin,¹⁷ which serves as an intermediate in the manufacturing of synthetic rubbers, epoxy resins, paper chemicals, surfactants, adhesives, insecticides, agricultural chemicals, coatings, ion-exchange resins, solvents, plasticizers, and pharmaceuticals.¹⁸ Additionally, it is a vital building block for biologically active compounds such as the β -adrenergic blocker (S)-atenolol and the amino acid derivative (R)-carnitine.¹⁸ The most notable industrial use of HHDHs is in the synthesis of ethyl (R)-4-cyano-3-hydroxybutanoate, a critical precursor in

the production of atorvastatin, the active ingredient in Lipitor®, which became the first drug to achieve annual sales exceeding 10 billion USD.¹⁹

The halogenation of an aliphatic methyl group on a protein-bound threonyl thioester is catalyzed by syringomycin halogenase (SyrB2), a non-heme Fe(II)/ α -ketoglutarate (α KG)-dependent enzyme.²⁰ SyrB2 can also halogenate a non-native substrate, α -aminobutyric acid, to yield monochlorinated, dichlorinated, and trichlorinated products. It can also be used to broaden its reactivity scope to include multibromination.²¹ The crystal structure reveals that the Fe centre is coordinated by two histidines, a bidentate α KG, water, and chloride (Fig. 1, species 1).²² Binding of L-Thr-S-SyrB1 would exclude water from the active site, enabling dioxygen binding (Fig. 2, species 2 and 3). Decarboxylation of α KG generates a high-energy ferryl-oxo intermediate (Fig. 2, species 4), which then abstracts a hydrogen atom from the substrate. The substrate radical subsequently captures the chlorine atom (Fig. 2, species 5), producing chlorinated L-Thr-S-SyrB1 and regenerating the reduced Fe(II) center. Once the substrate carbon radical is formed, competition between chlorine (Cl[•]) and hydroxyl (OH[•]) transfer is expected. However, no threonine hydroxylation has been observed, indicating a strong preference for Cl[•] rebound. It is important to note that SyrB2 requires substrates bound to a peptidyl carrier protein (PCP), which complicates its biotechnological application.

Haloperoxidases, which contain either heme iron or vanadium as a cofactor, halogenate electron-rich double bonds in substrates through the use of electrophilic halonium ions (Cl⁺, Br⁺ and I⁺).²³ In both types of haloperoxidases, hypohalites formed in the enzyme active site diffuse out. As a result, any oxidative transformation is not influenced by the (chiral) enzyme environment, and the reaction's selectivity is determined by the reactivity of the starting material, not by the enzyme itself. The heme-dependent haloperoxidases generate



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Currently he is designing various new ligands for asymmetric C–H halogenation.

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Basab Bijayi Dhar

focuses on developing catalysts based on first-row transition metals for C–H activation, water oxidation, and CO₂ reduction, aiming to reduce dependence on rare metals in synthetic chemistry.

Dr Basab Bijayi Dhar earned his M.Sc. in Chemistry from the University of Calcutta and completed his Ph.D. at Jadavpur University under Prof. Rupendranath Banerjee on electron transfer reactions of manganese(IV). He did postdoctoral research with Prof. E. S. Gould (Kent State University) and Prof. R. Stan Brown (Queen's University). Currently he is an Associate Professor at SNIoE, Delhi NCR, and his research



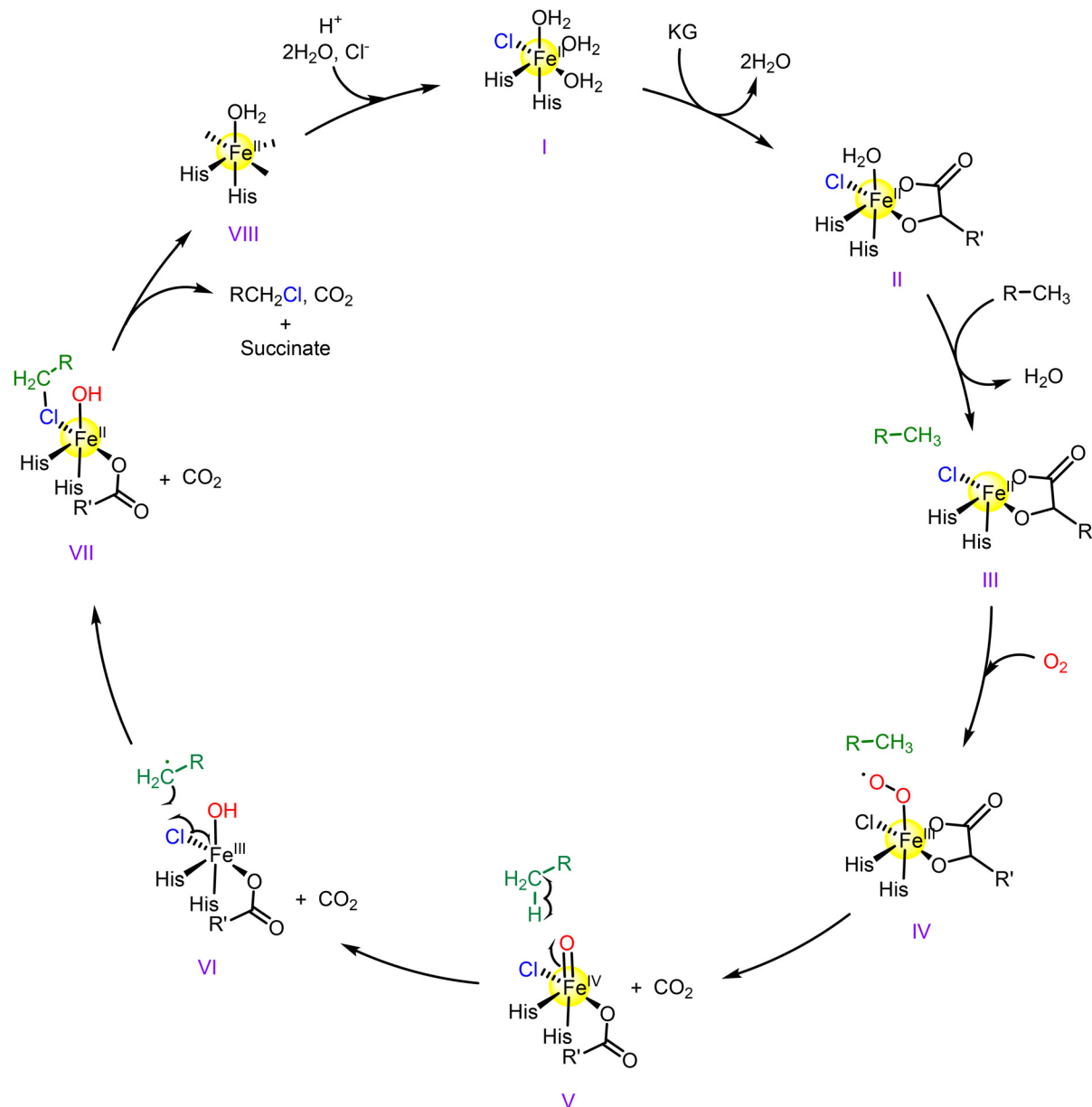


Fig. 2 Mechanistic pathway of SyrB2.

an Fe^{IV}-(oxo)-porphyrin⁺ complex in the presence of hydrogen peroxide (H₂O₂), which then oxidizes halides (Fig. 3).^{24a} The research group of Holtmann utilized the heme-dependent LfCPO for halogenating the terpene thymol to enhance its antimicrobial properties.²⁵ Itoh *et al.* studied the halogenation of various electron-rich arenes using CpVBPO from *Corallina pilulifera*^{26a} and halohydroxylation of alkenes (Fig. 4) using LfCPO62.^{26b} When these reactions were carried out in aqueous media in the presence of OH⁻ ions, the formation of halohydrins took place.

The vanadium-dependent haloperoxidases use a peroxovanadate species formed by the reaction with H₂O₂ (Fig. 4).^{24b} Schrader and his team explored the halohydroxylation reactions of terpenes using chloroperoxidase from *Caldariomyces*

fumago.^{24c,d} Chemoenzymatic production of halogenated phenolic monoterpenes thymol and carvacrol using chloroperoxidase extracted from *Caldariomyces fumago* was reported by Getrey *et al.*²⁷ In a similar manner, Hartung and co-workers investigated the bromination of various phenols using the vanadium-dependent bromoperoxidase extracted from *Ascophyllum nodosum*.²⁸ Using vanadium-dependent chloroperoxidase from *Curvularia inaequalis*, the efficient bromination of thymol (TON ~2 000 000) was carried out by Fernández-Fueyo *et al.*²⁹

Recently, flavin-dependent non-metal tryptophan halogenases have become popular for synthetic applications.³⁰ These enzymes regioselectively chlorinate electron-rich carbon centres using electrophilic halonium ions (Fig. 5). Members of



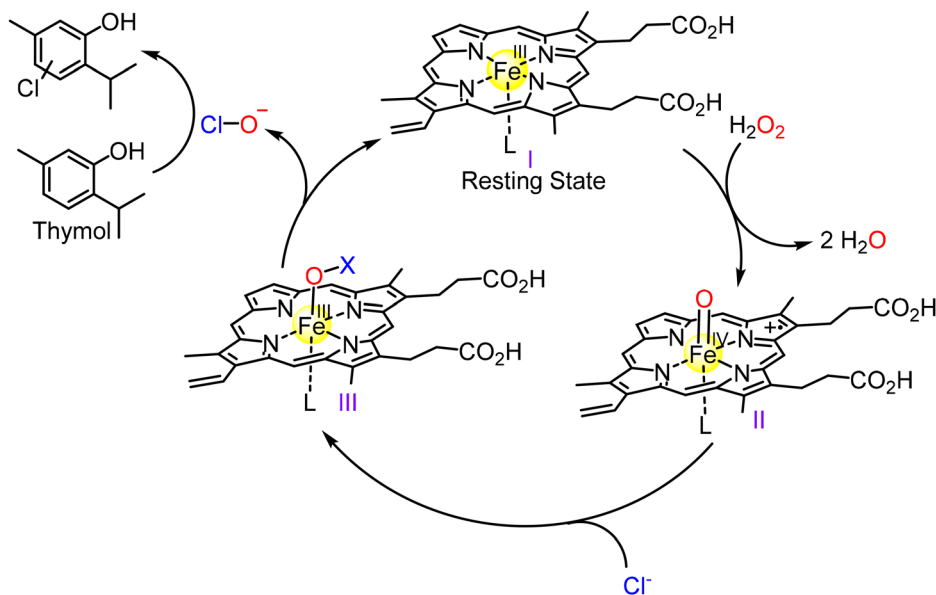


Fig. 3 Mechanistic pathway of heme-dependent haloperoxidase and chemoenzymatic halogenation of thymol by Holtmann and his research group.

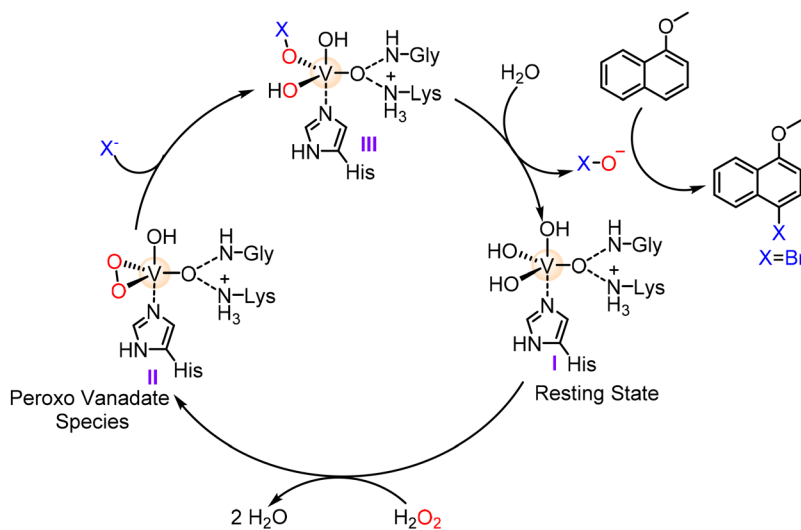


Fig. 4 Bromination of activated arenes using the vanadium-dependent bromoperoxidase.

this halogenase superfamily have been identified in the biosynthetic pathways of the antibiotics balhimycin and vancomycin, the antitumor agent rebeccamycin, and other halometabolites. Likely, regiospecific halogenation reactions carried out by bacteria are largely catalysed by flavin-dependent halogenases. To account for the regioselectivity, two mechanisms have been proposed. On one hand, the substrate is oxidized to an epoxide, which is decomposed by a nucleophilic attack of Cl⁻. On the other hand, direct chlorination by a high-energy flavin hypochlorite intermediate occurs. Van Pèe and co-workers revealed regioselective halogenation of the indole moiety using various flavin-dependent tryptophan halogenases under benign conditions (X: Cl, Br).³¹

3. C–H chlorination and bromination

3.1 Non-heme iron-based catalysts

Gif chemistry, originally developed by Sir Derek Barton for the selective functionalization of saturated hydrocarbons, has evolved beyond its initial focus on oxidation to encompass halogenation reactions.^{32,33} The GoAgg^{III} system, an advanced variant of Gif chemistry, has demonstrated effective bromination of alkanes using polyhaloalkanes such as CBrCl₃ as halogen sources.^{33a} Reaction rates vary with the structure of the polyhaloalkane used, and the selectivity toward various saturated hydrocarbons differs markedly from that seen in radical halogenation. Furthermore, product distribution



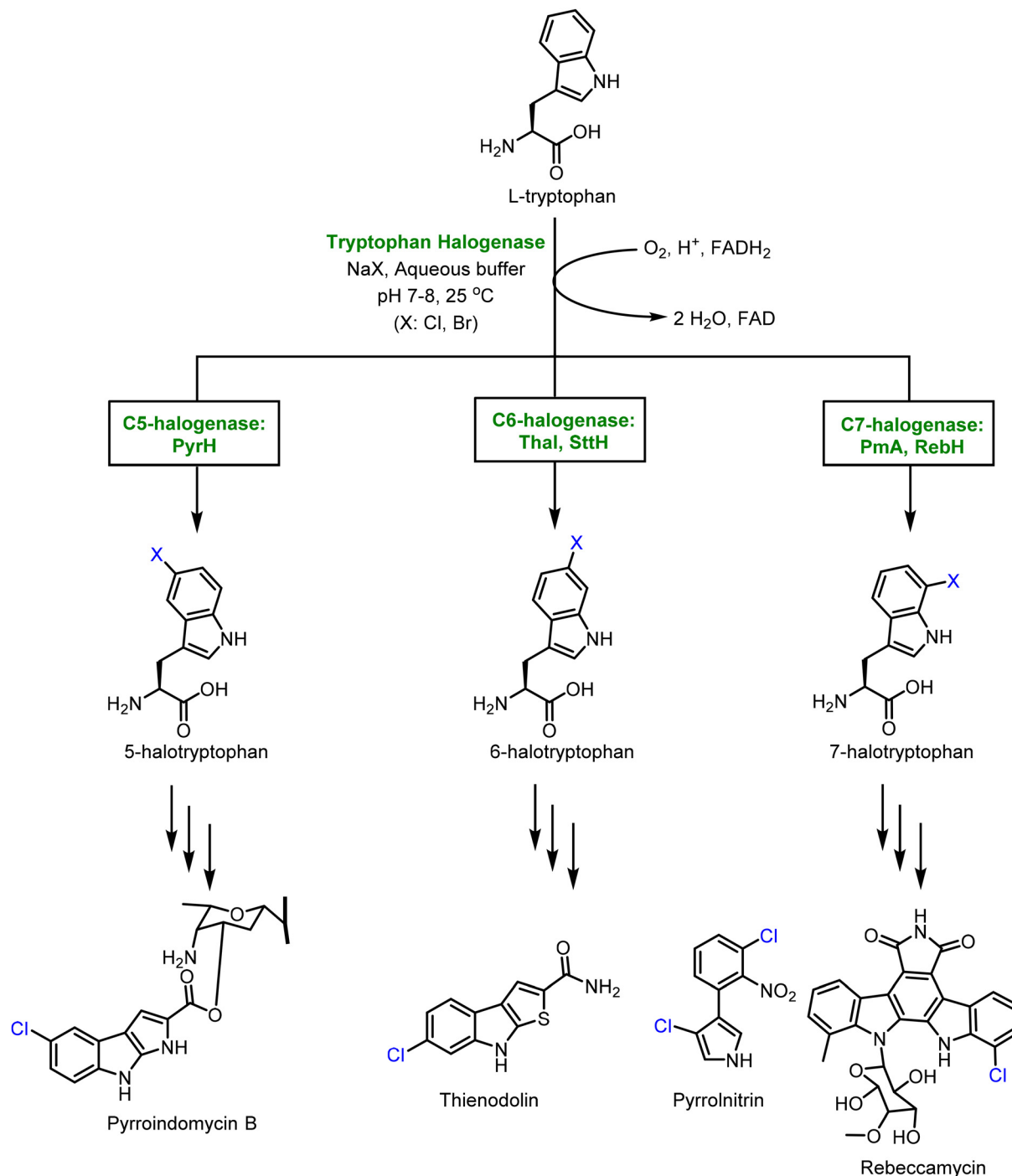


Fig. 5 Flavin-dependent non-metal tryptophan halogenases and biosynthetic pathways of the antibiotics balhimycin and vancomycin, the antitumor agent rebeccamycin, and other halometabolites using tryptophan halogenases.

studies, such as those involving the bromination of cyclohexyl bromide, highlight a distinct regioselectivity profile under Gif-type conditions. This selectivity is attributed to the involvement of a high-valent metal-oxo intermediate rather than a simple, unselective free radical. The metal center directs the hydrogen abstraction, preferring the less sterically hindered secondary position.³² However, isolating the highly reactive,

short-lived metal intermediates in Gif chemistry has been a significant challenge. Therefore, the nature of the reaction mechanism, whether radical or non-radical, was highly debated and described as the “Gif paradox”.^{33b} Chlorination reactions using similar systems (*e.g.*, GoAgg^{III}) and chlorine-containing reagents have also been explored, further underscoring the generality of Gif-type halogenation for forming



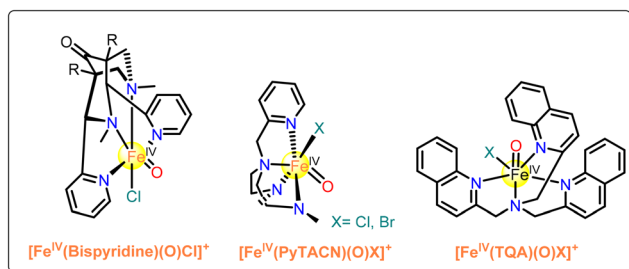


Fig. 6 Biomimetic iron-oxo-halide frameworks for enzyme-inspired halogenation.

both C–Br and C–Cl bonds.^{33a} In summary, while Barton's Gif chemistry opened a novel pathway for C–H functionalization under non-radical conditions, its practical limitations include environmental concerns, selectivity issues, and poor scalability, highlighting the need for more refined, catalytic, and sustainable halogenation methods.

In 1993, Que Jr. and co-workers reported a biomimetic system that models the activity of non-heme halogenases.^{34a} They synthesized a series of $[\text{Fe}(\text{III})\text{X}_2]^+$ complexes (X = Cl, Br, N_3), where L is tris(2-pyridylmethyl)amine (TPA), capable of functionalizing alkanes under ambient conditions. Upon treatment with a stoichiometric amount of alkyl hydroperoxide in the presence of cyclohexane, these non-heme iron complexes generated halocyclohexane in good yields. A significant kinetic

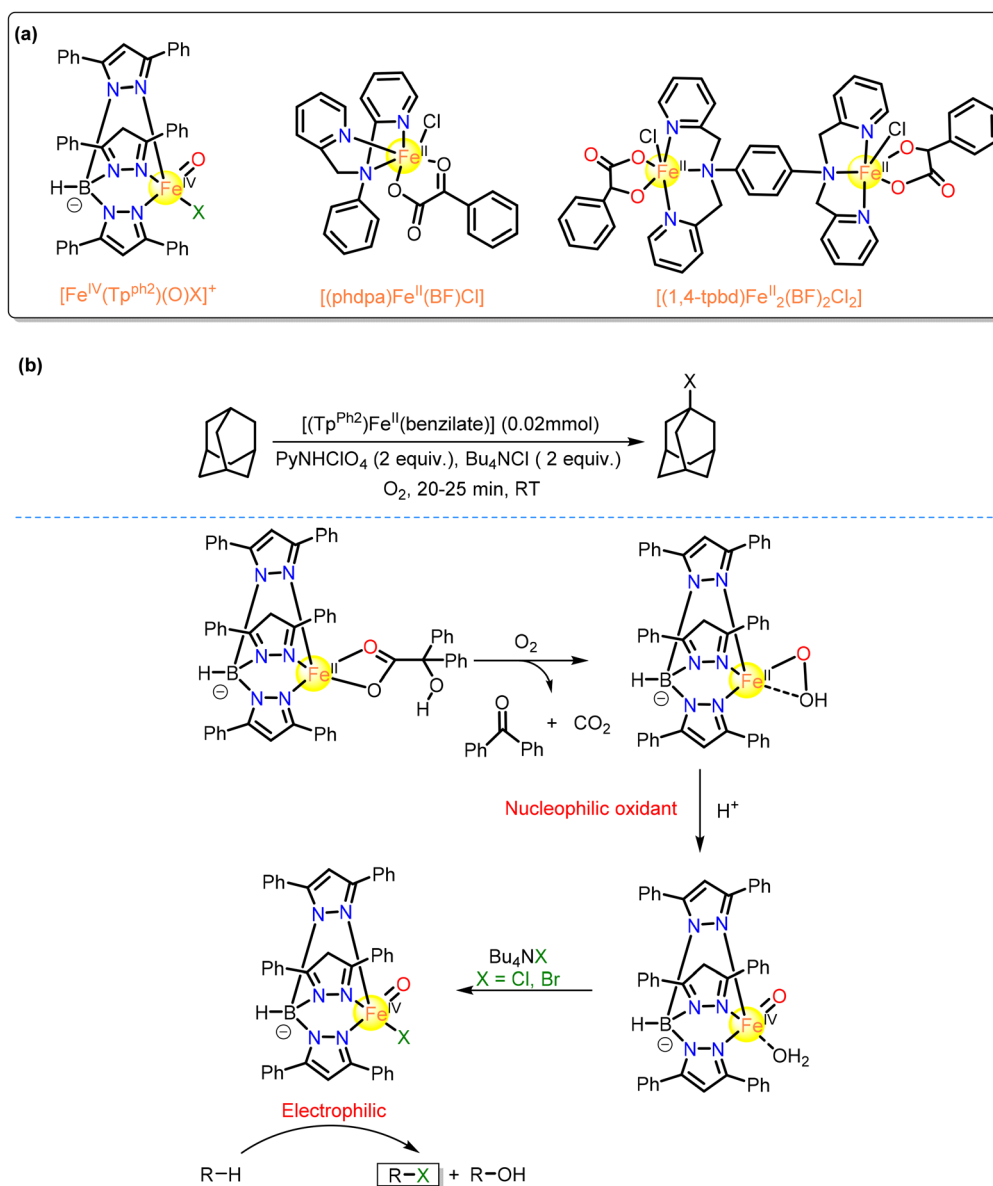


Fig. 7 (a) Various Fe-based non-heme complexes utilized in C–H halogenation. (b) Proposed mechanistic pathway for the generation of an electrophilic iron-oxo species and its involvement in aliphatic C–H halogenation.



isotope effect (KIE) was observed when comparing reactions using cyclohexane and $[D_{12}]$ cyclohexane ($K_H/K_D = 7-10$), indicating that hydrogen atom abstraction (HAA) constitutes the rate-determining step of the transformation. Notably, the magnitude of the KIE was found to vary depending on both the halide ligand and the structure of the tripodal ligand frame-

Table 1 Comparison of different reported homogeneous catalysts for C–H bond chlorination of cyclohexane

S. no.	Catalyst	TON (chlorination of cyclohexane)
1	Mn(TPP)Cl/NaOCl ⁴³	87
2	[Ni ^{II} (Pytacn)(CF ₃ SO ₃) ₂] (Pytacn = 1-(2-pyridylmethyl)-4,7-dimethyl-1,4,7-triazacyclononane)/NaOCl ⁴⁴	24
3	[Ni ^{II} (L)] (L = bisamidate ligand)/NaOCl ⁴⁵	44
4	[Fe ^{IV} (2 PyN2Q)(O)] ²⁺ /NaOCl ^{40b}	52
5	[Ni ^{II} -amidoquinoline]/NaOCl ^{46a}	212
6	[Ni ^{II} -bisbiguanide]/NaOCl ⁴⁷	680

work. In contrast, changing the nature of the peracid oxidant, by altering the R group (*e.g.*, *tert*-butyl or cumyl), had no measurable impact on the reaction outcome. The authors proposed the involvement of an iron(III)-alkyl peroxide intermediate, which undergoes O–O bond cleavage to generate a high-valent iron(V)-oxo-halide species. This highly reactive intermediate is responsible for the hydrogen atom abstraction (HAA) from the C–H bonds of cycloalkanes. The alkyl radical then undergoes a halide rebound pathway, predominantly yielding the halogenated product, with only a minor amount of the hydroxylated product observed. However, Arends *et al.* proposed that in the $[Fe(TPA)X_2]^+/t\text{-BuOOH}$ system ($X = \text{Cl}, \text{Br}$ *etc.*), *t*-BuO[•] radicals, rather than metal-based oxidants, serve as the sole agents responsible for alkyl radical generation.^{34b} The preferential formation of the halogenated compound is attributed to the difference in redox potentials between the chloride and hydroxide radicals during the radical rebound step, with $E^\circ(\text{Cl}^\bullet/\text{Cl}^-) = 0.36 \text{ V}$ vs. NHE and $E^\circ(\text{OH}^\bullet/\text{OH}^-) = 2.20 \text{ V}$ vs. NHE. However, in the presence of excess alkyl hydroperoxide (150 equivalents with respect to the Fe catalyst), cyclohexanol was the predominant product, which is the major drawback of the $[Fe(TPA)X_2]^+/t\text{-BuOOH}$ system. Noack and Siegbahn investigated the mechanism of oxidative chlorination catalyzed by the non-heme iron complex $[(\text{TPA})Fe^{\text{III}}\text{Cl}_2]^+$ in the presence of

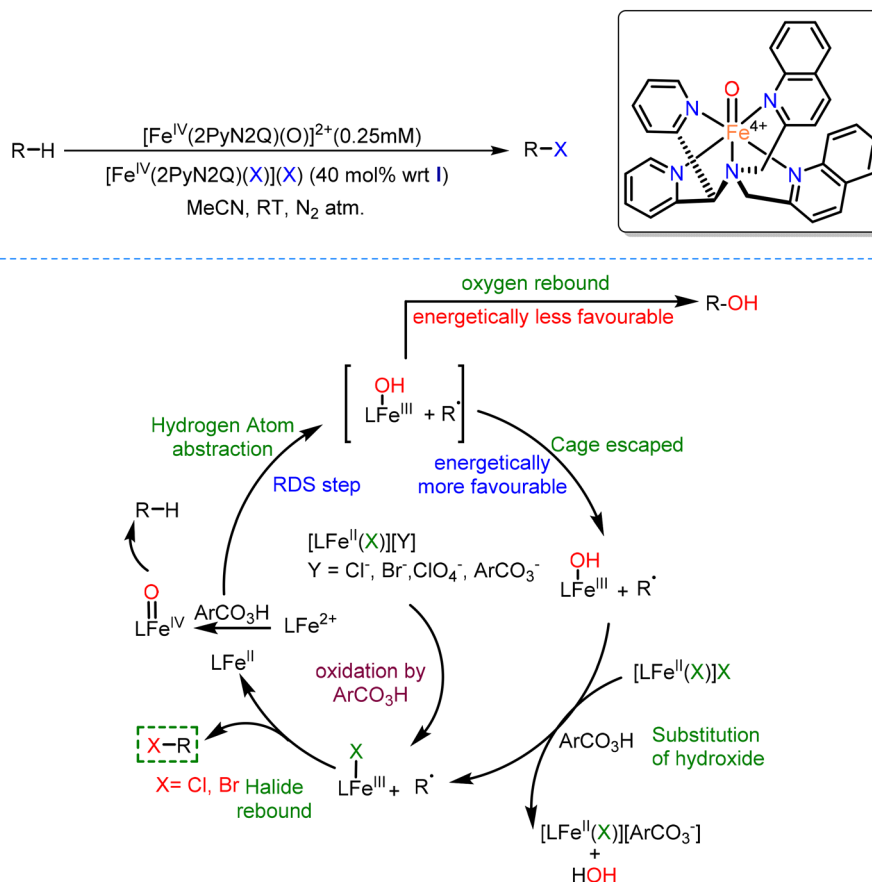


Fig. 8 Plausible mechanistic pathway for C–H halogenation mediated by iron(IV)-oxo in the presence of iron(II)-halide complexes.



a peroxide oxidant using density functional theory (DFT) calculations.³⁵ They reported that $\text{Fe}^{\text{V}}=\text{O}$ or $\text{Fe}^{\text{IV}}=\text{O}$ species serve as the key catalytic intermediate.

In 2010, Comba and co-workers reported a dichloroiron(II) complex featuring a tetradentate bispidine ligand {L = 3,7-

dimethyl-9-oxo-2,4-bis(2-pyridyl)-3,7-diazabicyclo[3.3.1]nonane-1,5-dicarboxylate methyl ester} (Fig. 6), which demonstrated catalytic activity for C–H halogenation of alkanes using various oxidants, including PhIO, TBHP, and H_2O_2 .³⁶ The selectivity between halogenation and hydroxylation was found to be

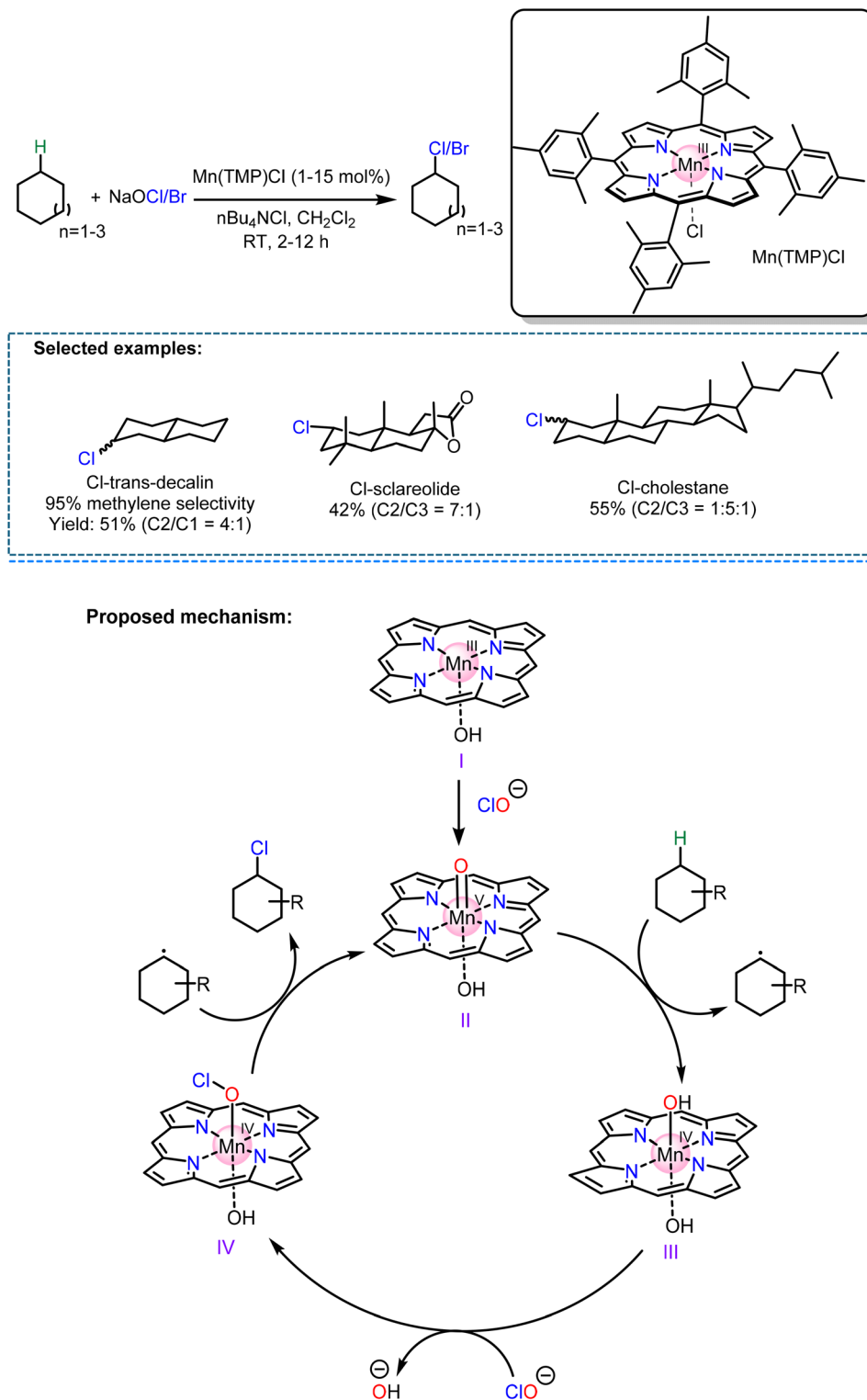


Fig. 9 Groves' catalyst for C–H chlorination and bromination.



oxidant-dependent. Kinetic isotope effect (KIE) studies using cyclohexane and $[D_{12}]$ cyclohexane revealed K_H/K_D values of 14 (PhIO), 9 (TBHP), and 3 (H_2O_2), indicating that hydrogen atom abstraction (HAA) is the rate-determining step and suggesting the involvement of different high-valent iron-oxo intermediates, either $Fe^V=O$ or $Fe^{IV}=O$, depending on the oxidant. Subsequently, Costas and co-workers explored the reactivity of $[Fe^{IV}(PyTACN)(O)Cl]^+$ (Fig. 6) for C–H halogenation.³⁷ However, no halogenated products were observed due to the strong coordination of the halide to the metal center, which likely inhibits halogen rebound. Later, Que and co-workers reported the $[Fe^{IV}(O)(TQA)X]^+$ ($X = Cl, Br$) complex (Fig. 6), where TQA = tris(quinolin-2-ylmethyl)amine, as an effective system for C–H bond halogenation.³⁸ This complex represents the first synthetic example that reproduces the Mössbauer parameters characteristic of an $S = 2$ oxoiron(IV) species.

Paine and co-workers reported the first example of a biomimetic iron(II) complex supported by the hydro[tris(3,5-diphenyl-pyrazol-1-yl)]borate ligand (Fig. 7), which undergoes aerobic oxidation under acidic conditions.^{39a} This system enables aliphatic C–H bond halogenation of substrates such as

adamantane, toluene, and cyclohexane *via* the formation of an Fe^{IV} -oxoaquo intermediate (Fig. 7) in the presence of tetra-*n*-butylammonium halide and pyridinium perchlorate. The same group also developed two non-heme iron(II)- α -keto acid complexes: $[(phdpa)Fe(BF)Cl]$ and $[(1,4-tpbd)Fe_2(BF)_2Cl_2]$ (Fig. 7), incorporating phdpa (*N,N*-bis(2-pyridylmethyl)benzene-1,4-diamine), BF (benzoylformate), and 1,4-tpbd (*N,N,N',N'*-tetrakis(2-pyridylmethyl)benzene-1,4-diamine) as ligands.^{39b} These systems catalyze the oxidative halogenation of aliphatic C–H bonds using molecular oxygen as the terminal oxidant. Alongside halogenated products, hydroxylated and overoxidized species were also observed.

Maiti and co-workers developed a new synthetic scheme for selective halogenation utilizing a pentadentate ligand [ligand (1,1-di-(pyridin-2-yl)-*N,N*-bis(quinolin-2-yl-methyl)methanamine), 2PyNQ]-derived nonheme iron complex.^{40a} They used a combination of $[Fe^{IV}(2PyN_2Q)(O)]^{2+}$ and $[Fe^{II}(2PyN_2Q)(OTf)_2]$, where $Fe^{IV}=O$ abstracts the hydrogen from the aliphatic substrate *via* hydrogen atom abstraction (HAA) to form the radical, and iron(II)-halide used as the halide source to generate the halogenated product (Table 1 and Fig. 8).^{40b} Maiti's research

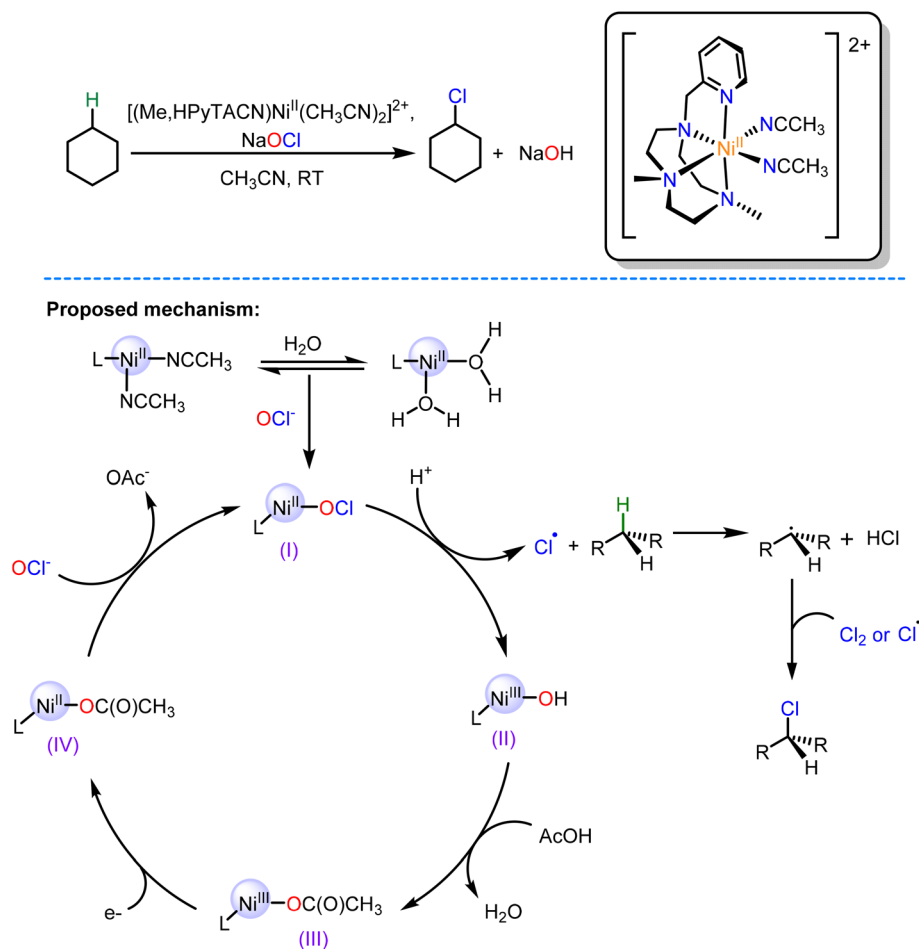
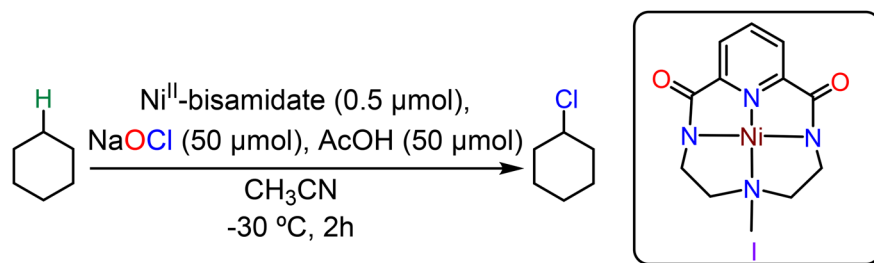


Fig. 10 Costa's catalyst for the chlorination of alkanes.





Proposed mechanism:

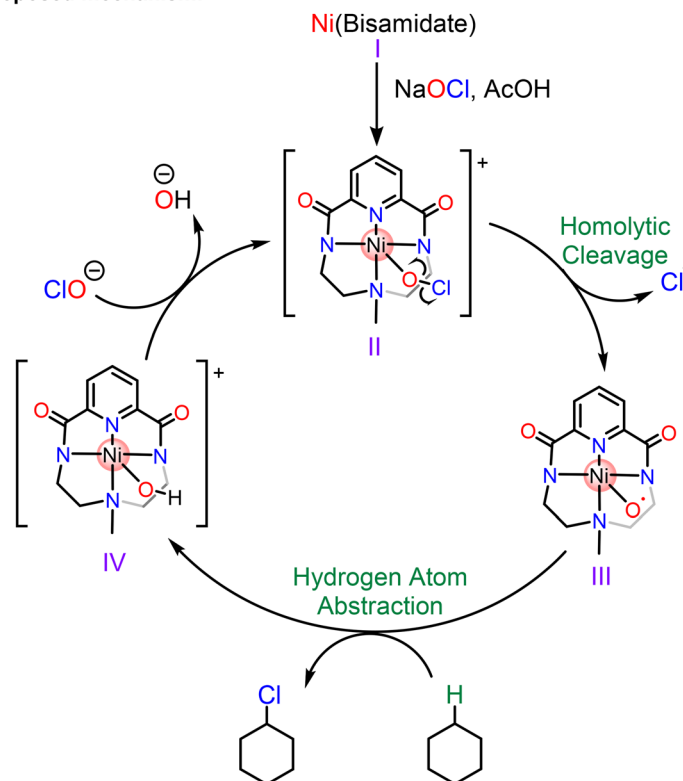


Fig. 11 A. Company's catalyst for the chlorination of alkanes.

Table 2 Comparison of catalytic C–H chlorination and bromination of different substrates^{46a,47}

S. no.	Substrate	Product	TON (Ni ^{II} -amidoquinoline) ^a	TON (Ni ^{II} -bisbiguanide) ^b
1	Cyclohexane	Chlorocyclohexane	212	680 ± 60
2	Methylbenzene	(Chloromethyl)benzene	202	200 ± 20
3	Ethylbenzene	(1-Chloroethyl)benzene	208	370 ± 30
4	Adamantane	1-Chloroadamantane (3°)	173	1600 ± 200 (3°)
		2-Chloroadamantane (2°)	5	320 ± 30 (2°)
5	Methylbiphenyl	4-Chloromethylbiphenyl	196	260 ± 25
6	2,3-Dimethylbutane	2-Chloro-2,3-dimethylbutane	92	130 ± 20
7	<i>n</i> -Hexane	2°-Chlorohexane	—	108 (2°)
		1°-Chlorohexane	—	27 (1°)
8 ^c	Cyclohexane	Bromocyclohexane	—	125
9 ^c	Methylbenzene	(Bromomethyl)benzene	—	82
10 ^c	Ethylbenzene	(1-Bromoethyl)benzene	—	115 (2°); 6 (1°)

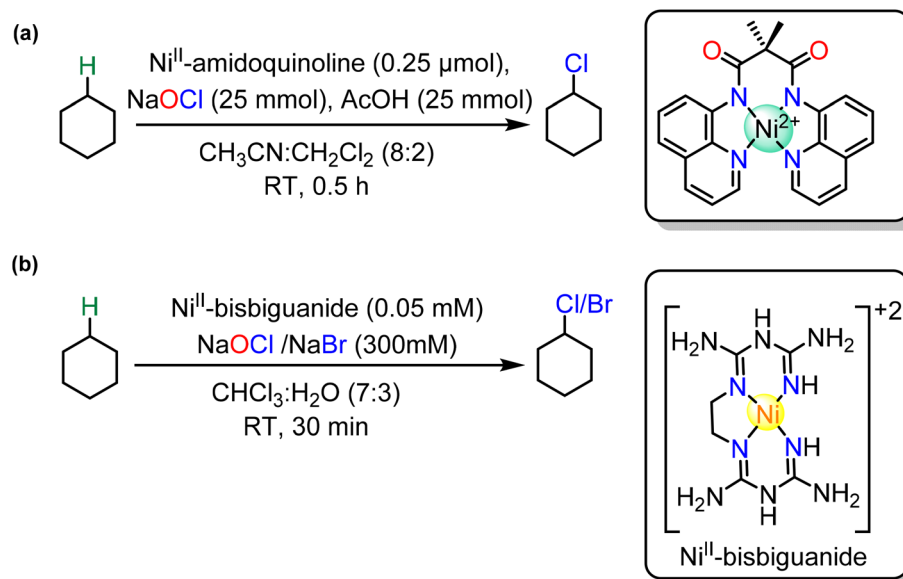
^a Reaction conditions: Ni catalyst (0.25 mmol), substrate (0.25 mmol), NaOCl (0.25 mmol), and AcOH (0.25 mmol) in 1 mL of CH₂Cl₂/CH₃CN (2 : 8 v/v) at RT for 30 minutes under N₂. ^b Ni catalyst (0.05 μmol), substrate (0.25 mmol), NaOCl (0.30 mmol), and AcOH (0.15 mmol) in 1 mL of H₂O:CHCl₃ (7 : 3 v/v) for 30 minutes at RT. ^c Ni catalyst (0.05 μmol), substrate (0.25 mmol), NaOCl (0.30 mmol), NaBr (0.30 mmol), and AcOH (0.15 mmol) in 1 mL of H₂O/CHCl₃ (7 : 3 v/v) for 30 minutes at RT. The products were identified using GC-MS.



group has also demonstrated a method for benzylic C–H chlorination using an easily prepared Mn(salen) complex as the catalyst in the presence of NaOCl.^{40c}

Han and co-workers have developed a bioinspired pincer iron(III) complex that catalyzes both the oxidation and chlorination of aliphatic C(sp³)–H bonds. The use of *cis*-decalin as a mechanistic probe suggests the involvement of a long-lived carbon-centered radical intermediate, indicative of a stepwise reaction pathway. Detailed studies point to an iron-oxo species as the likely oxidant responsible for the rate-determining C–H

activation step.⁴¹ Recent investigations by the research groups of Company and Costas have explored the reactivity of a series of well-defined iron(IV)-oxo complexes supported by variants of the tetradentate 1-(2-pyridylmethyl)-1,4,7-triazacyclononane (Pytacn) ligand.⁴² These studies revealed a remarkable switch in chemoselectivity during C–H bond functionalization, modulated by the reaction conditions. In the presence of halide anions, the complexes promote exclusive C–H oxygenation, whereas the addition of a superacid diverts the reaction pathway towards C–H halogenation. Mechanistic investigations



Proposed mechanism:

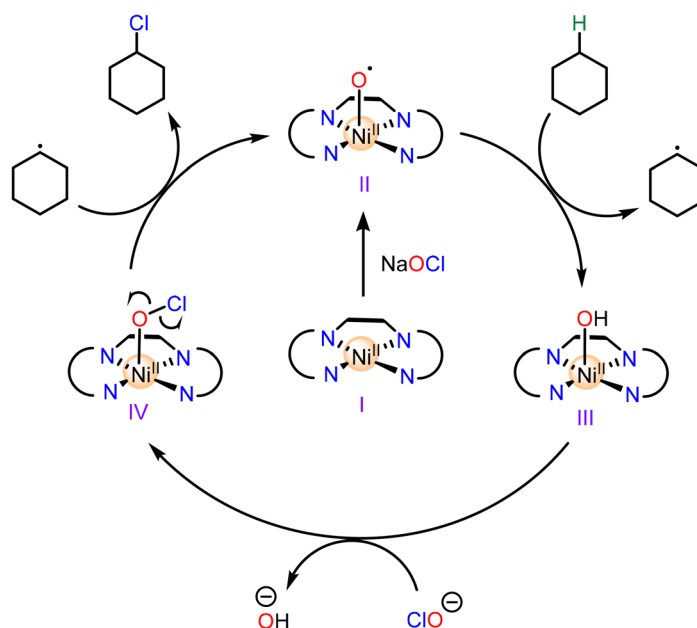


Fig. 12 (a) Ni(II)-(amidoquinoline) as a catalyst for the chlorination of alkanes. (b) Ni(II) complexes supported by the bisbiguanide ligand and its activity towards halogenation.



suggest that both pathways share a common rate-determining hydrogen atom transfer (HAT) step. The nature of the reaction environment, specifically, the presence of acid, plays a critical role in dictating the fate of the resulting carbon-centered radical, favouring halogenation over oxygenation under acidic conditions.

3.2 Porphyrin-based catalysts

In 2010, Groves and his co-workers demonstrated that a highly electron-deficient manganese porphyrin, Mn(TPPFP)Cl, efficiently and selectively catalyzes C–H chlorination and bromination of various simple alkanes in dichloromethane

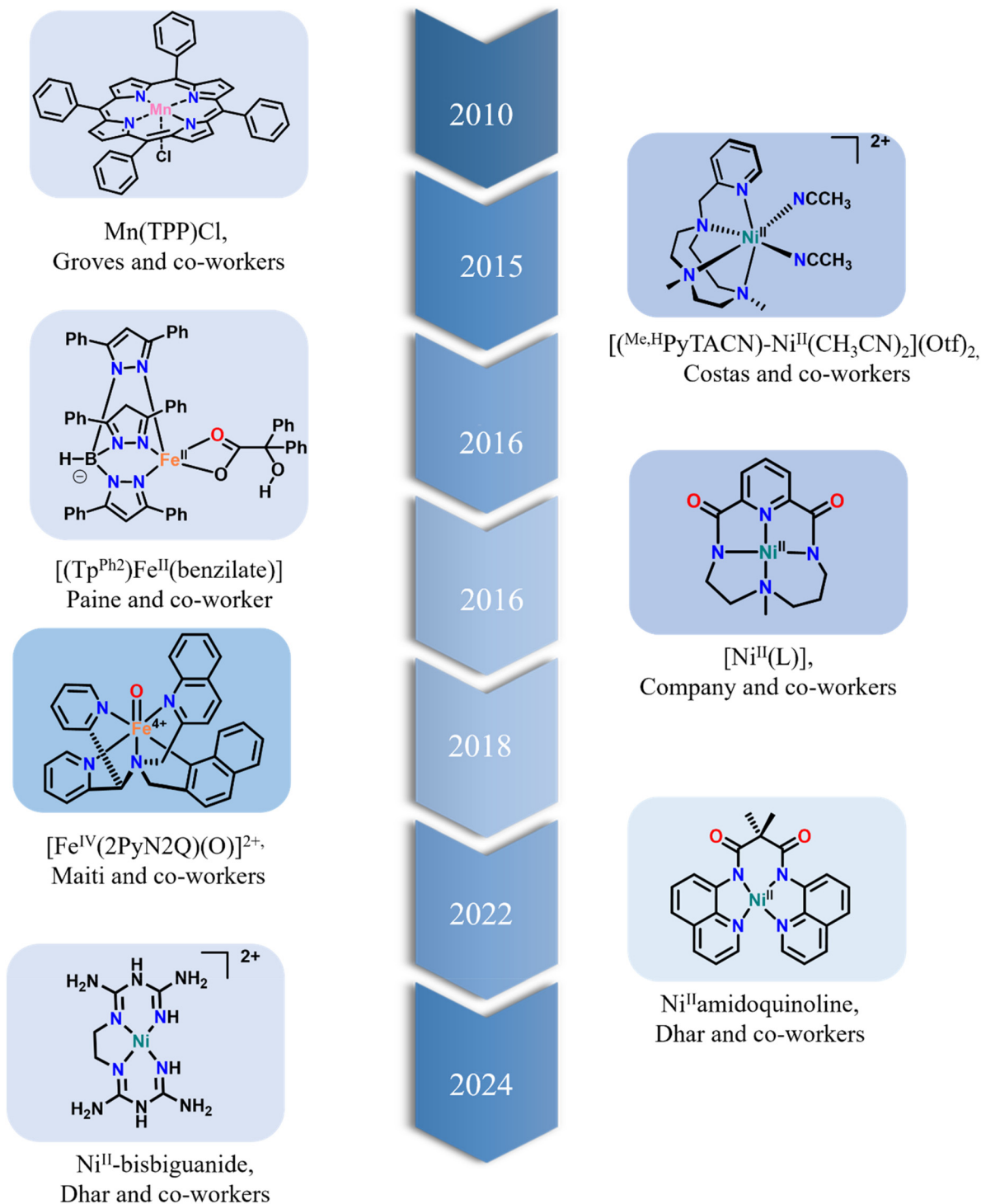


Fig. 13 First-row transition metal-based catalysts for C–H chlorination and bromination from 2010 onwards.



(DCM) at room temperature (RT) (Table 1 and Fig. 9).⁴³ Mechanistic studies showed that the reactive oxomanganese(v) species readily abstracts a hydrogen atom from a substrate C–H bond (supported by the kinetic isotope effect (KIE); a

large magnitude of kinetic isotopic effect value of 8.7 ± 0.7 is observed for Mn(TPP)Cl, indicating that C–H abstraction is the rate-limiting step as the radicals like ClO[•] and Cl[•] show lower kinetic isotope effect values), forming a substrate-derived

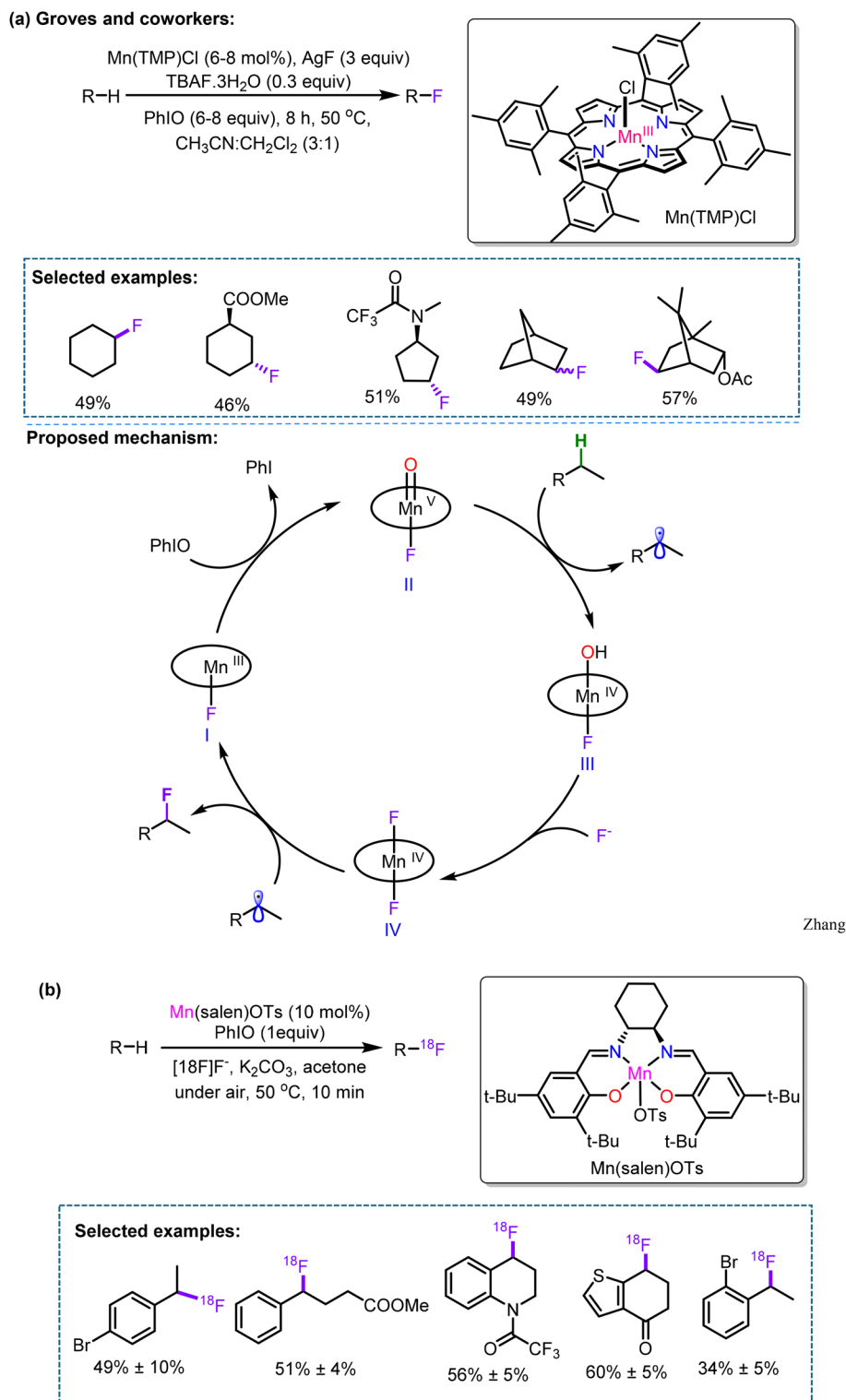


Fig. 14 Fluorination using (a) Mn(TMP)Cl catalyst, (b) Mn(Salen)Cl catalyst, both the catalysts facilitated the transfer of fluoride to a broad range of hydrocarbons when used alongside a hypervalent iodine-based oxidant and AgF and TBAF·3H₂O as fluoride sources.



radical and an HO-Mn^{IV}(por) intermediate. The HO-Mn^{IV} intermediate then exchanges the hydroxyl ligand with excess hypochlorite anions, transferring chlorine to the alkyl radical.

3.3. Nickel-based catalysts

The Costas group used a Ni(II) catalyst [(^{Me,H}PyTACN)-Ni^{II}(CH₃CN)₂](OTf)₂, with a robust triazacyclononane ligand (L = ^{Me,H}PyTACN) for the chlorination of alkanes with NaOCl as the terminal oxidant and chloride source (Table 1 and Fig. 10).⁴⁴ Reaction of NaOCl with [(^{Me,H}PyTACN)-Ni^{II}(CH₃CN)₂](OTf)₂ in acetonitrile generated a transient nickel(II)-hypochlorite species, [(L)Ni^{II}-OCl(S)]⁺ (A), where S = solvent. The intermediate was identified by electrospray ionization mass spectrometry (ESI-MS). Spectroscopic studies such as UV-Vis absorption, electron paramagnetic resonance (EPR), and resonance Raman spectroscopy, *etc.*, indicate that intermediate **I** undergoes decay to yield the corresponding nickel(III)-hydroxo complex, [(L)Ni^{III}-OH(S)]²⁺ (**II**), presumably *via* homolytic cleavage of the O-Cl bond with concomitant release of a Cl[•] radical. Under the reaction conditions, hydrolysis of acetonitrile to acetic acid leads to the formation of the nickel(III)-acetate species, [(L)Ni^{III}-OOCCH₃(S)]²⁺ (**III**), which is subsequently reduced to the nickel(II) analogue, [(L)Ni^{II}-OOCCH₃(S)]²⁺ (**IV**), likely *via* a reaction with OCl⁻ or ClO₂⁻. Upon reintroduction of NaOCl to complex **IV**, regeneration of **II** occurs more rapidly and in higher yield. According to the report, the addition of acids such as acetic acid or triflic acid significantly enhances both the rate and extent of formation of intermediate (**II**), suggesting that protonation facilitates O-Cl bond homolysis and promotes the generation of reactive high-valent nickel-oxo species. Later, the Company group used Ni(II) complexes with tetradentate macrocyclic bis(amidate) ligands (Table 1 and Fig. 11), showing promising reactivity toward alkanes at -30 °C in the presence of NaOCl.⁴⁵ DFT calculations suggested the formation of a nickel-hypochlorite species. Experimental and computational analyses indicate that the intermediate is best formulated as a Ni^{III} complex with one unpaired electron delocalized in the ligands surrounding the metal centre.

In 2022, our research group reported that Ni(II) complexes supported by an amido-quinoline ligand (Tables 1, 2 and Fig. 12a) effectively catalysed C-H chlorination of a series of hydrocarbons in the presence of NaOCl and acetic acid at RT in CH₂Cl₂/CH₃CN (2 : 8 v/v).^{46a} Bond dissociation energy of the C(sp³)-H bond of the substrates varies from 99.3 kcal mol⁻¹ (cyclohexane) to 87 kcal mol⁻¹ (ethyl benzene). Exclusively chlorinated products (TON: 220 for cyclohexane) were obtained without any hydroxylated products and thus mimicking the activity of the halogenase enzyme (Table 2 and Fig. 12).^{46a} Inspired by our work, Thomas R. Ward's research group, biotinylated Ni(II)-(amido-quinoline) complexes within streptavidin.^{46b} The resulting artificial nickel chlorinase was also efficient for the chlorination of diverse C(sp³)-H bonds in an acetonitrile-phosphate buffer mixture. However, the solubility and stability of first-row transition metal catalysts in an aqueous medium are the major challenges. Therefore, synthetic chemists are trying to develop a new catalyst capable of achieving selective halogenation in an aqueous medium.

Recently, Khatri *et al.* described that water-soluble guanidine complexes of Ni(II) (Fig. 12b) successfully chlorinate various hydrocarbons in the presence of NaOCl and acetic acid in a water and chloroform mixture (7 : 3) at RT (TON ~680 for cyclohexane) (Table 2).⁴⁷ The KIE (*k_H/k_D*) value was found to be

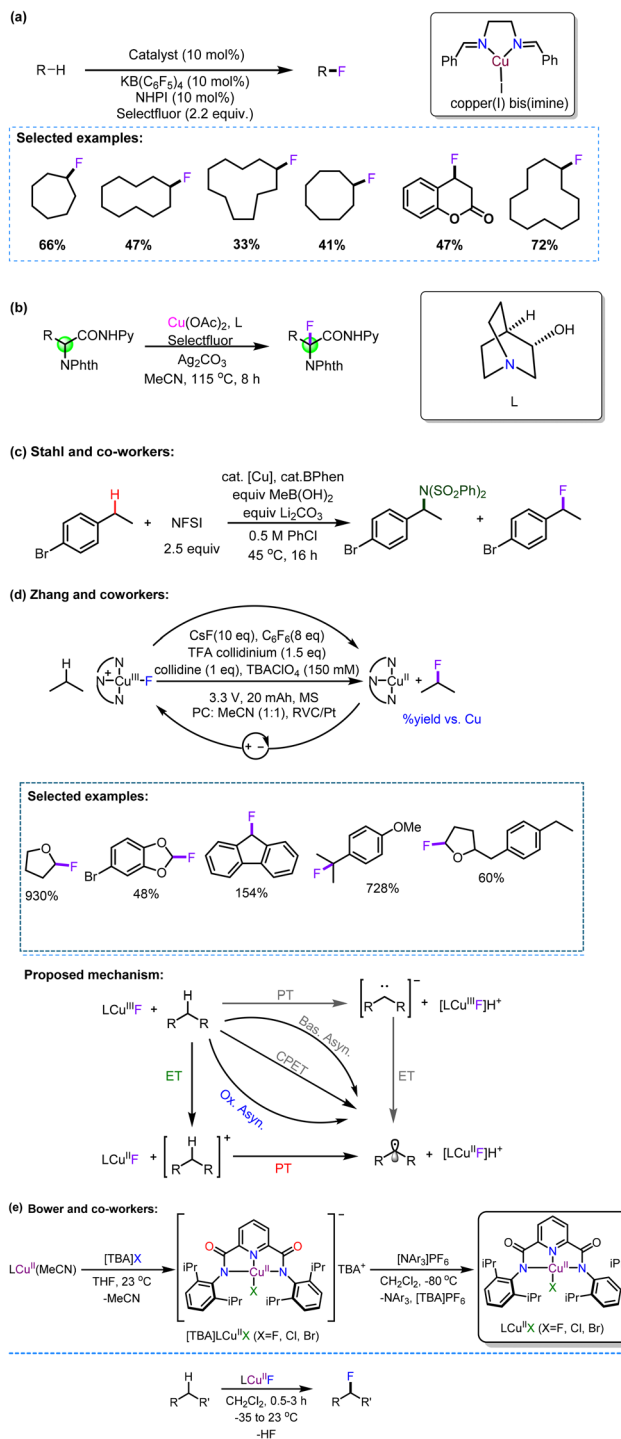


Fig. 15 (a-c) Cu-catalysed C(sp³)-H fluorination; (d) organic electrochemical strategy for C(sp³)-H fluorination; 1 TON = 100% yield vs. Cu. (e) C(sp³)-H fluorination by a copper(III) fluoride complex.



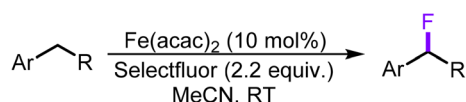
12.2. It is suggested that C–H abstraction is the rate-determining step. Moreover, bromination was also carried out by *in situ* generation of NaOBr using an equimolar mixture of NaOCl and NaBr. A minimal amount of chlorinated product was found with the brominated product (Table 2).⁴⁷ Fig. 13 illustrates the progression of first-row transition metal-based catalytic systems reported since 2010 for the direct halogenation of C–H bonds, specifically targeting chlorination and bromination transformations.

4. Fluorination

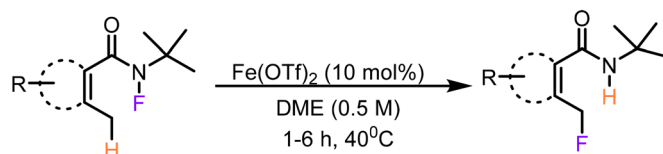
C–H fluorination is exceedingly rare in nature, and living organisms have scarcely evolved a dedicated biochemistry for fluorine. A few tropical and subtropical plants are known to

produce fluoroacetate, although its biosynthetic pathway remains poorly understood.⁴⁸ Moreover, reports of other naturally occurring fluorinated compounds are often viewed with caution, as they may result from contamination by anthropogenic or industrial sources rather than true biological synthesis. Nowadays, carbon–fluorine bonds are becoming increasingly crucial components in pharmaceuticals, agrochemicals, and positron emission tomography (PET) tracers.⁴⁹ Among the top 200 pharmaceutical products by retail sales in 2016, 36 contain fluorine.⁵⁰ In recent years, numerous methods have been developed for incorporating fluorine into small molecules. However, relatively few studies focus on direct fluorination.⁵¹ This approach is particularly appealing due to its efficiency and potential for late-stage functionalization, enabling rapid access to complex, fluorinated analogues of high value. Despite these advantages, the inherent

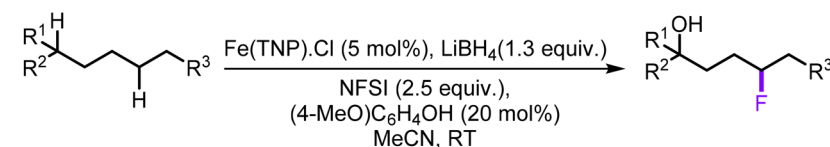
(a) Lectka and co-workers



(b) Cook and co-workers



(c) Cook and co-workers



proposed mechanism:

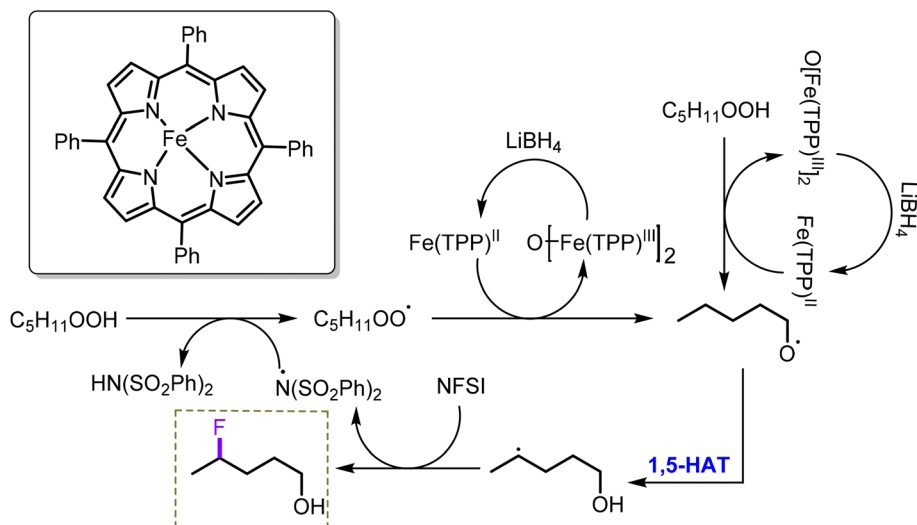


Fig. 16 (a–c) Iron-catalysed C(sp³)–H fluorination.



challenges of C–H fluorination have limited the number of successful examples, especially when compared to other halogenation strategies. In the context of C–F bond formation, several additional obstacles complicate direct fluorination. Notably, the strong metal–fluorine interactions and the high hydration energy of fluoride ions often result in substantial reaction barriers. Although C–F bonds are thermodynamically stable, with bond dissociation energies exceeding 100 kcal mol⁻¹,⁵² the use of fluoride salts as fluorine sources can be problematic due to their high lattice energies, which limit their reactivity. Electrophilic fluorinating reagents present a more reactive alternative, circumventing some of these issues, but they are typically more costly and less atom-economical than fluoride-based reagents. Considering both the intrinsic difficulties of C–H functionalization and the unique challenges associated with C–F bond formation, it is perhaps unsurprising that direct C–H fluorination remains a relatively underdeveloped area in synthetic chemistry.

C–H fluorination could be classified as directed C–H fluorination, aromatic C–H fluorination, and aliphatic C–H fluorination. Directed C–H fluorination and aromatic C–H fluorination are mostly carried out using second- and third-row transition metal catalysts. In this review, we are focusing on the application of first-row transition metal-based catalysts in C(sp³)–H fluorination. These synthetic methods require extreme and non-biological conditions to overcome the high C–H bond

dissociation energy and the low nucleophilicity of the fluoride ion.

4.1 Aliphatic C–H fluorination

Groves and co-workers demonstrated that various manganese catalysts, such as Mn(TMP)Cl, Mn(Salen)Cl, and Mn(TPFPP)Cl, can facilitate the transfer of fluoride to a broad range of hydrocarbons when used alongside a hypervalent iodine-based oxidant and AgF and TBAF·3H₂O as fluoride sources (Fig. 14a).⁵³ Mechanistic studies indicate that the regioselectivity of C–H bond activation is governed by an oxomanganese(v) catalytic intermediate. However, the competing hydroxide rebound reaction is a key challenge in these processes. Later, the Groves group further achieved the Mn(salen)OT-catalyzed ¹⁸F-radiofluorination of benzylic C(sp³)–H bonds with a no-carrier added [¹⁸F]-fluoride ion (Fig. 14b).⁵⁴ This approach employs Mn(salen)OTs as the fluoride transfer catalyst, enabling efficient labelling of various bioactive molecules and building blocks with radiochemical yields between 20% and 72% within 10 minutes.

Lectka and collaborators reported that combining Selectfluor (SF) and CuI in the presence of diamine ligands enables C(sp³)–H bond fluorination, yielding fluorinated products in modest to good yields (up to 75%) (Fig. 15a).⁵⁵ It was found that the addition of *N*-hydroxyphthalimide and potassium iodide could improve the yield of this reaction and facili-

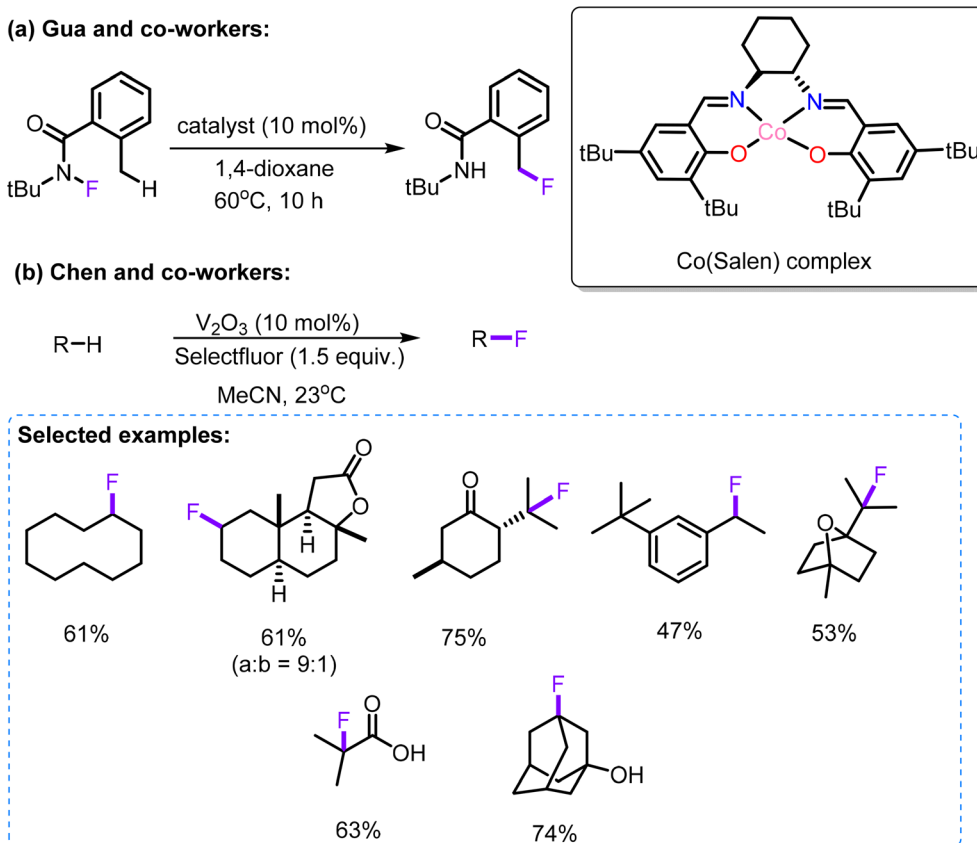


Fig. 17 C(sp³)–H fluorination using (a) Cobalt-catalyst (b) Vanadium-catalyst.



tate the regeneration of the Cu(I) catalyst as well. In 2018, Liu and co-workers reported a novel Cu(II)-catalyzed α -C(sp³)-H fluorination of amino acid derivatives using Selectfluor as the fluorine source (Fig. 15b).⁵⁶ Notably, the addition of (*R*)-3-hydroxyquinclidine was found to significantly enhance the reaction yield. The proposed mechanism involves initial pre-coordination of Cu(OAc)₂ with (*R*)-3-hydroxyquinclidine, which is believed to facilitate the catalytic process. In 2020, Stahl and co-workers reported a Cu(I)-catalyzed benzylic C(sp³)-H fluorination reaction using *N*-fluorobenzenesulfonimide (NFSI) as the fluorinating agent (Fig. 15c).⁵⁷ In 2023, the Zhang group developed an organic electrochemical strategy for C(sp³)-H fluorination, enabling the synthesis of a broad range of alkyl fluorides *via* a Cu (BCAP)(MeCN)-catalyzed process (Fig. 15d).⁵⁸ This work built upon their earlier studies involving a stoichiometric copper(III) reagent, now advancing to a catalytic system. To prevent the undesired reduction of Cu(II) at the cathode, hexafluorobenzene (C₆F₆) was employed as a sacrificial electron acceptor. Substrate scope investigations revealed a clear preference for fluorination at C-H bonds adjacent to oxygen atoms over benzylic C-H sites, highlighting the method's selectivity and synthetic utility. In 2025, Díaz *et al.* reported a copper(I)-initiated, site-selective β - ζ -C(sp³)-H bond fluorination of ketones, carboxylic esters, and amides using Selectfluor as the fluorine source.⁵⁹ This methodology demonstrates high site selectivity in C(sp³)-F bond formation and was successfully applied to the late-stage functionalization of complex natural products and pharmaceutical derivatives, highlighting its potential utility in medicinal chemistry and molecular diversification.

In 2020, Bower and colleagues reported the C(sp³)-H fluorination by a copper(III) fluoride complex (Fig. 15e).^{58b} Mechanistic studies using a triphenylmethyl radical probe revealed that the complex, LCuF, engages in radical capture through a concerted pathway. Notably, LCuF demonstrated the ability to mediate both hydrogen atom abstraction and radical capture, enabling efficient fluorination of allylic and benzylic C-H bonds, as well as α -C-H bonds of ethers, under mild, room-temperature conditions.

In 2013, the Lectka group reported an Fe(II)-catalyzed C(sp³)-H fluorination of alkylbenzenes using Selectfluor as the fluorine source (Fig. 16a).⁶⁰ The reaction goes through a radical pathway. The Cook group reported a mild, iron-mediated, amide-directed C-H fluorination of benzylic, allylic, and unactivated positions (Fig. 16b).⁶¹ In this method, *N*-fluoro-2-methylbenzamides undergo chemoselective fluorine transfer upon treatment with a catalytic amount of iron(II) triflate (Fe(OTf)₂), affording the corresponding fluorinated products in high yields. The reaction exhibits broad substrate scope and excellent functional group tolerance, all without the need for noble metal catalysts. Mechanistic and computational studies support a pathway involving short-lived radical intermediates, with fluorine transfer occurring directly through an iron-mediated process. An alkoxyl radical-guided strategy for site-selective functionalization of unactivated methylene and

methine C-H bonds, enabled by an Fe^{II}-catalyzed redox process, is described by the Liu group (Fig. 16c).⁶²

Guo and team developed a redox-neutral, radical-relay cobalt-catalyzed intramolecular C-H fluorination of

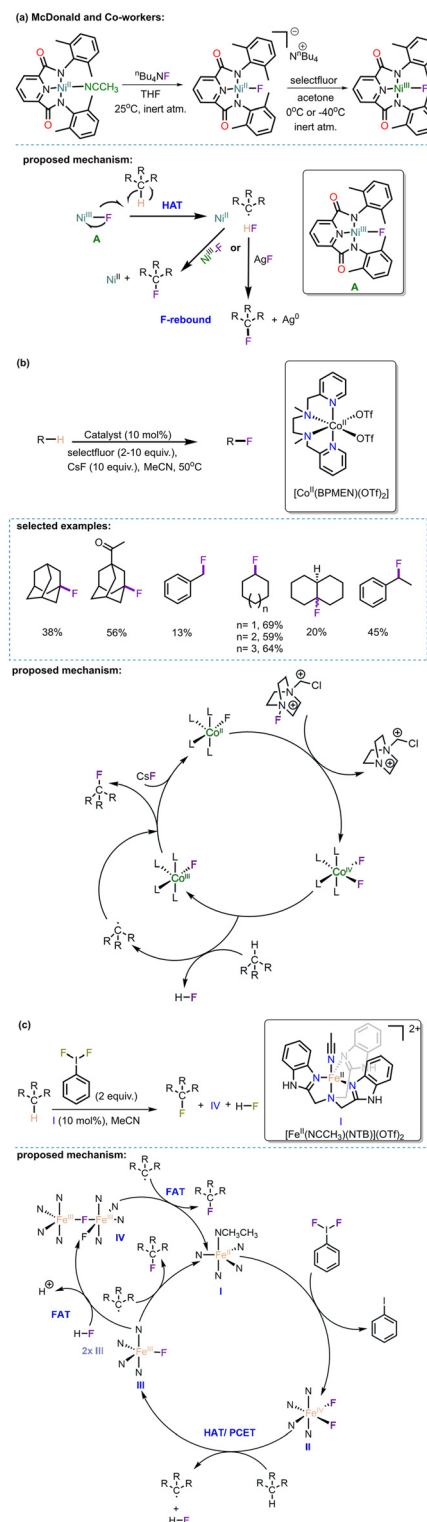


Fig. 18 (a) Nickel-catalyzed, (b) Cobalt-catalyzed and (c) Iron-catalyzed C(sp³)-H fluorination developed by McDonald and his research group.



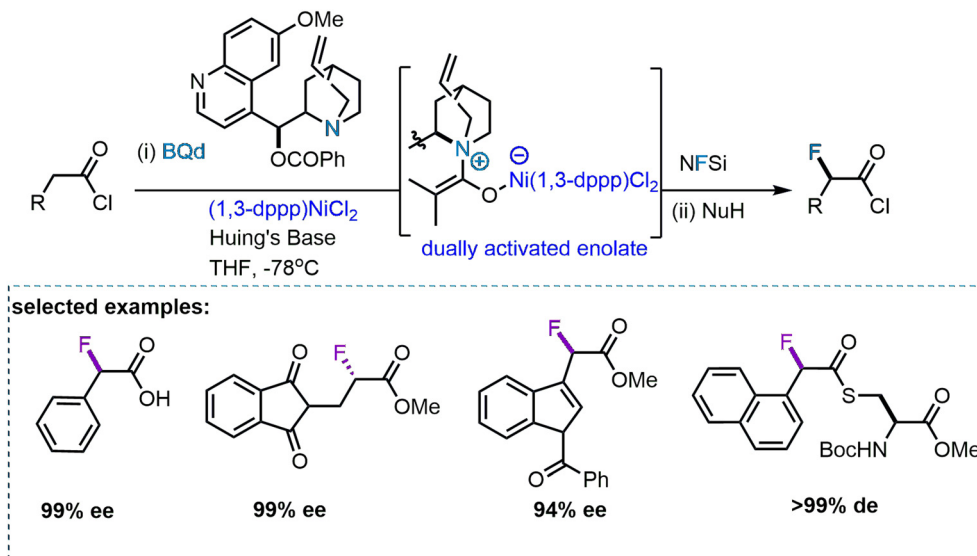


Fig. 20 α -Fluorination of acid chlorides via cooperative action of two discrete catalysts: a chiral nucleophile and an achiral Ni(II) complex.

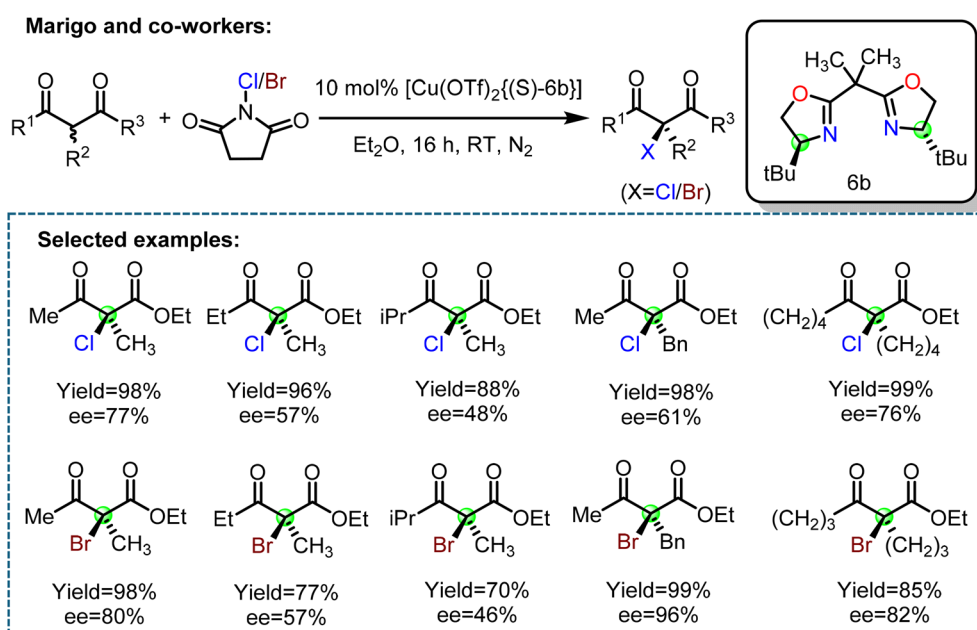


Fig. 21 Enantioselective halogenation of both acyclic and cyclic β -ketoesters, and β -diketone using NCS and NBS as halogen sources in combination with chiral bisoxazoline copper(II) complexes.

they obtained the corresponding optically active α -halogenated products in excellent yields and with moderate to good enantioselectivities (Fig. 21).⁷¹ Frings *et al.* developed an asymmetric halogenation of cyclic and acyclic β -oxoesters using a sulfoximine-copper complex, producing α -bromo-, α -chloro-, and α -fluoro-compounds in high yields and with moderate to good enantioselectivity.⁷² Shibatomi *et al.* further advanced enantioselective halogenation by employing a chiral Lewis acid

catalyst formed from $\text{Cu}(\text{OTf})_2$ and a newly designed spirooxazoline ligand. This system enabled the α -chlorination of β -ketoesters and malonates with excellent enantioselectivity, reaching up to 98% ee.⁷³ Reddy *et al.* achieved highly enantioselective fluorination of aryl acetyl- and 3-butenoyl-substituted thiazolidinones using a combinatorial catalytic system composed of DBFOX-Ph/Ni(II)/HFIP/2,6-lutidine, providing an efficient route to chiral α -fluorinated compounds.⁷⁴



6. Conclusions

Halogenating enzymes have the potential to play a significant role in future “white” biotechnology. However, several aspects must be optimized before these enzymes gain widespread use in industrial biocatalysis. In contrast, transition metal-catalyzed C–H activation has opened new avenues for synthesizing a broader range of halogenated compounds by enabling more selective halogenation of difficult substrates. As summarized in this review, significant progress has been achieved in the C(sp³)-H halogenation field through the use of manganese, iron, cobalt, nickel, and copper catalysts, greatly expanding the synthetic toolbox for accessing halogenated compounds. However, to date, a major challenge in C(sp³)-H functionalization lies in achieving precise site selectivity amid the multitude of similar C–H bonds within a molecule.⁷⁵ Future research is expected to address this issue through the development of innovative strategies aimed at enhancing selectivity and control. The design of catalysts and ligands will play a pivotal role in overcoming the inherent reactivity differences among primary, secondary, and tertiary C–H bonds, enabling selective and remote C–H functionalization. Integrating computational modeling to predict and guide the development of highly selective catalyst–ligand systems is expected to accelerate advances in this area.

The development of general and high-yielding methods for asymmetric C(sp³)-H halogenation remains a significant challenge. Success in this area would represent a breakthrough, offering powerful new tools for the synthesis of chiral pharmaceuticals and fine chemicals. Recent advances have demonstrated that earth-abundant metals such as nickel and cobalt can enable enantioconvergent radical transformations.⁷⁵ This emerging strategy holds significant promise for the synthesis of chiral molecules and is poised for further expansion.

Furthermore, to facilitate the translation of these diverse methodologies from academic research to industrial practice, the implementation of flow technology could be a promising approach. Flow systems offer superior heat and mass transfer, enhanced safety over conventional batch processes, and are especially well-suited for managing highly reactive intermediates. Furthermore, the intrinsic scalability of flow chemistry makes it an attractive platform for large-scale applications.

Conflicts of interest

There are no conflicts to declare.

Data availability

This review article does not include the generation or analysis of original data; therefore, a data availability statement is not applicable.

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