

Cite this: *Chem. Sci.*, 2025, 16, 4328 All publication charges for this article have been paid for by the Royal Society of Chemistry

From two-component enzyme complex to nanobiohybrid for energy-efficient water–gas shift reaction†

Thomas Pichon,^a Claudio Righetti,^b Julien Pérard,^a Alan Le Goff^{*b} and Christine Cavazza^{*a}

The water–gas shift reaction (WGSR, $\text{CO} + \text{H}_2\text{O} \rightleftharpoons \text{CO}_2 + \text{H}_2$) is widely used for the upgrading of syngas, a key substrate for various chemical processes. However, the industrial WGSR requires high pressure and temperature, and has low selectivity. Here, we have designed a biohybrid catalyst by combining CODH from *Rhodospirillum rubrum*, which catalyzes CO-to- CO_2 conversion and a bioinspired nickel bisdiphosphine complex, which catalyzes the hydrogen evolution reaction, immobilized on carbon nanotubes. Carbon nanotubes enable the dual functioning of both catalysts providing efficient electrical conductivity and allowing electroless CO-to- CO_2 conversion and H_2 evolution. Owing to CO tolerance of the Ni complex, this bioinspired nanohybrid catalyst shows high performance by reaching 100% conversion yield and maximum TOF of 30 s^{-1} towards WGSR at ambient temperature and pressure in the presence of either pure CO or syngas.

Received 20th September 2024

Accepted 2nd January 2025

DOI: 10.1039/d4sc06394e

rsc.li/chemical-science

1 Introduction

The syngas market has continuously grown in the last few years and is projected to reach 406 860 MWth by 2025. Syngas is a cheap and versatile substrate composed mainly of N_2 , CO_2 , H_2 and CO, used for the synthesis of a wide range of products (liquid fuels, methanol, *etc.*).¹ However, the current challenge is that this gas is mainly produced from fossil fuels. Concomitantly, the generation of syngas from a diverse range of biomass materials is increasing thanks to the development of biomass or waste feedstock gasification. However, while gasification and purification of gas from fossil raw materials is a well-established technology, relevant differences still exist when using biomass as feedstock, and specific challenges still must be addressed. It is noteworthy that achieving the precise H_2/CO ratio is a significant prerequisite for the synthesis of fuels and chemicals. For instance, a H_2 -to-CO ratio of 2 is required for the synthesis of methanol. Given that the H_2/CO ratio in syngas from slagging gasifiers typically ranges from 0.3 to 1, substantial H_2 -enrichment processes are imperative. Furthermore, the impurity levels are higher in derived biomass gas.² The key reaction for the upgrading of syngas is the water–gas shift reaction or WGSR ($\text{CO} + \text{H}_2\text{O} \rightleftharpoons \text{CO}_2 + \text{H}_2$). However, the current industrial processes occur at high pressure and temperature.

Furthermore, the presence of contaminant species in the inlet gas can result in catalyst poisoning and a reduction in the overall process efficiency. Another significant limitation of the reaction is the initial H_2/CO ratio, which must be relatively high to prevent unwanted side reactions.³ Optimizing performances and energy efficiency of WGSR is therefore of great interest. In nature, several microorganisms offer great potential as sustainable alternatives to fossil-based synthesis of chemicals and fuels. In acetogens, hydrogen-dependent CO_2 reductases (HDCR) directly use H_2 to reduced CO_2 to formate: this is the first reaction in the Wood–Ljungdhal pathway, an ancient pathway for CO_2 fixation, but also for energy conservation. The discovery of hydrogenogenic carboxydophilic bacteria (*e.g. Rhodospirillum rubrum*) capable of using CO as their sole energy source, thanks to a biological analogue of WGSR (bio-WGSR) has the potential to offer a promising avenue for the design of green alternatives to the industrial catalysts.⁴ This is because the thermodynamically favorable bio-WGSR is less sensitive to syngas impurities and operates at room temperature and atmospheric pressure. In these microorganisms, the WGSR is catalyzed by a multiprotein complex using two redox nickel enzymes: a carbon monoxide dehydrogenase (CODH), which oxidizes CO to CO_2 and an energy-conserving [NiFe] hydrogenase, which catalyzes the proton reduction to H_2 .⁴ In addition, a ferredoxin transfers the electron from the CODH to the hydrogenase.

For several decades now, these families of enzymes have been interfaced with electrodes, and have demonstrated respective reversible CO oxidation and proton reduction at near zero-overpotential requirement.^{5–13} This is the reason why Armstrong and Reisner were able to achieve enzymatic WGSR by mixing these enzymes with graphite pellets in a test tube.¹⁴ Owing to the

^aUniv. Grenoble Alpes, CEA, CNRS, IRIG, CBM, F-38000 Grenoble, France. E-mail: christine.cavazza@cea.fr^bUniv. Grenoble Alpes, CNRS, DCM, F-38000 Grenoble, France. E-mail: alan.le-goff@univ-grenoble-alpes.fr† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4sc06394e>

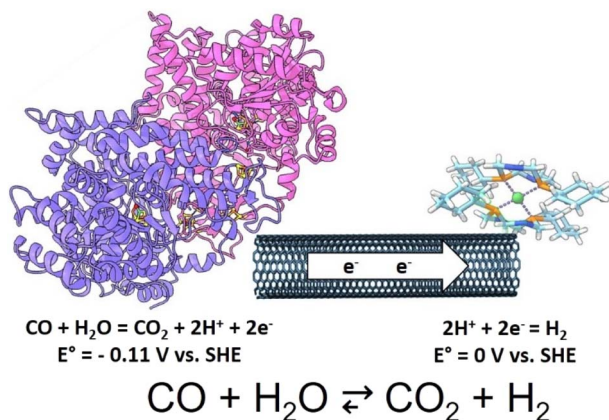


Fig. 1 Bioinspired WGS principle.

conductivity of graphite pellets and the addition of CO, both wired enzymes were able to drive the WGS at room temperature and pressure, without any external energy output. However, this first attempt was limited by the low specific surface area of graphite pellets and the well-known inhibition of hydrogenases by CO. We and others have recently developed the use of carbon nanotubes (CNTs) as high-performance electrode material for the wiring of such enzymes.^{15–19} We have also demonstrated that specific molecular functionalization can improve their immobilization and electrical interfacing with carbon nanotubes. In the case of hydrogenases, a series of mononuclear nickel complexes have been first designed by D. L. DuBois and coworkers and further refined to approach the exceptional performances of hydrogenases towards both hydrogen evolution and uptake.^{20–24} Again, carbon nanotubes have demonstrated their ability to immobilize Ni complex and integrate these bio-inspired catalysts in high performance devices.^{25–27} In addition, these Ni complexes have demonstrated a much better tolerance toward CO than hydrogenases, making them great candidates for WGS.²⁸ While many nanobiohybrids have been designed to combine enzymes and/or molecular complexes on surfaces to perform catalytic reactions *via* a cascade strategy,^{29–33} only few examples have employed the nano-object support as the electronic bridge between the two catalysts operating in tandem.

In this work, we investigate the use of carbon nanotubes for the co-immobilization of two oxygen-sensitive catalysts: the recombinant CODH from *Rhodospirillum rubrum* overproduced in *Escherichia coli* (Rec-RrCODH) and a bio-inspired Ni complex to achieve WGS. Their ability to act as nanowires for the intimate connection between CODH and Ni complex is aimed at achieving bio-inspired electroless WGS under CO without external driving force. The influence of pH, catalyst ratio and type of organic functionalization of CNTs on the performances of bioinspired WGS was investigated and tested with pure CO or syngas (Fig. 1).

2 Results and discussion

2.1 Functionalization and electrochemistry of MWCNT films

First, multi-walled carbon nanotubes (MWCNT) electrodes were fabricated by filtration of a homogenous dispersion of 2 mg

MWCNTs in 50 : 50 water and ethanol (2 mL) either on a carbon-cloth electrode (for electrochemical analysis) or on a microporous polytetrafluoroethylene (PTFE) membrane (for WGS experiments) (Fig. S1†). This technique has been previously developed to afford highly homogenous CNT films with controlled thickness and morphology.^{25,34}

The accessible carbon-cloth surface was reduced to 0.07 cm² for electrochemical analysis. Fig. 2 shows the optimized response of **Rec-RrCODH** immobilized at a MWCNT electrode. As previously described, the functionalization of MWCNTs has a significant impact on the surface concentration of immobilized enzymes and bioinspired catalysts. MWCNTs were previously modified with pyrene-adamantane (^{ADA}MWCNTs), demonstrating the superior attachment of **Rec-RrCODH** on these functionalized MWCNTs owing to hydrophobic interactions.¹⁷ On the other hand, diazoniumnaphtoate tetrafluoroborate was employed to promote the functionalization of MWCNTs (^{NA}MWCNTs) with the [Ni^{II}(P^{Cy}N^{Arg})₂]⁷⁺ complex (**NiPNP^{Arg}**) owing to ionic interactions.^{26,27} CV of the **Rec-RrCODH**-modified electrode in the presence of CO exhibits an irreversible electrocatalytic wave corresponding to the CO-to-CO₂ electroenzymatic conversion (blue curve, Fig. 2A) with maximum current density of 2.2 mA cm⁻² in CO-saturated solution at pH 7.0.

The **NiPNP^{Arg}**-modified ^{NA}MWCNT electrode exhibits a reversible system at $E_{1/2}^{\text{red}} = -0.32$ V vs. SHE corresponding to the electroactivity of the immobilized **NiPNP^{Arg}** accompanied with an electrocatalytic wave corresponding to the hydrogen evolution reaction (HER) activity of the complex (curve red, Fig. 2A). Despite the fact that **NiPNP^{Arg}** is known to be more efficient for H₂ oxidation (10 s⁻¹ vs. 0.5 s⁻¹ for hydrogen

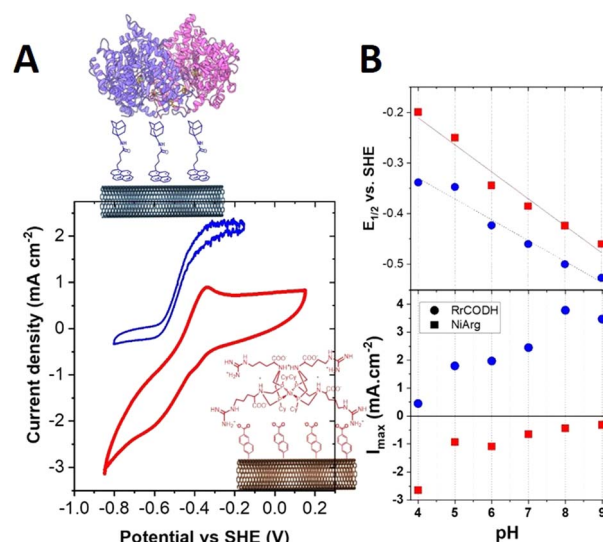


Fig. 2 (A) Cyclic voltammetry (CV) of **NiPNP^{Arg}**-modified ^{NA}MWCNT electrode (red) under Ar, and **Rec-RrCODH** modified ^{ADA}MWCNT electrode (blue) under CO in 50 mM Bis-Tris propane buffer pH 7.0 ($v = 5$ mV s⁻¹); (B) half-wave potential and I_{max} versus pH for **NiPNP^{Arg}**-modified MWCNT electrode (red) under Ar, and **Rec-RrCODH**-modified MWCNT electrode (blue) under CO in 50 mM Bis-Tris propane buffer pH 7.0.



evolution),^{22,27,35} this is still a high-performance catalyst for HER over a wide range of pH. Maximum current densities and half-wave potentials for both electrocatalytic response were investigated as a function of pH (Fig. 2B). As expected, these two electrocatalytic reactions follow a classic one-proton/one electron ratio potential-pH dependence. These measurements unambiguously demonstrate that the utilization of these two catalysts for CO oxidation and proton reduction provides a thermodynamic driving force for the WGS reaction within a pH range of 4.0 to 9.0, with an average ΔE of 80 mV.

MWCNT films were then modified with both **Rec-RrCODH** and **NiPNP^{Arg}** by two successive soaking steps. CV under CO is shown in Fig. 3 for this bifunctionalized MWCNT electrode.

When comparing the CV of the bifunctionalized electrode under CO (red curve, Fig. 3A) and CV of the **NiPNP^{Arg}**-modified MWCNT electrode under CO (red curve, Fig. 3B), a decrease of the HER catalytic current is observed corresponding to about 58% of the initial current and attributed to the partial inhibition of the HER of **NiPNP^{Arg}** in high CO concentration. This inhibition has already been observed for the complex in solution under high CO concentration and might be caused by weak coordination of CO to the **NiPNP^{Arg}**.^{22,36} Interesting, the CV response of the bifunctionalized electrodes corresponds to the sum of the responses of both catalysts, when tested independently (corresponding black curve from Fig. 3A and B). This electrocatalytic response shows that both catalysts can be effectively and stably immobilized at the surface of MWCNTs while keeping their electrocatalytic performances.

2.2 Bioinspired WGS

MWCNT films deposited on PTFE membranes were then modified with both **Rec-RrCODH** and **NiPNP^{Arg}** for WGS experiments under anaerobic conditions in a glove box ($[O_2] < 2$ ppm). It is noteworthy that the deposition of the enzyme prior to the complex or the opposite, as well as mixing both catalysts in the soaking solution, has no noticeable effect on the WGS performances. Surface concentrations, 0.9 nmol cm⁻² and 110

nmol cm⁻² for **Rec-RrCODH** and **NiPNP^{Arg}** respectively, were calculated from the total amount of each catalyst deposited on the MWCNT. This bifunctionalized MWCNT film was then introduced into a gas-tight vial with a minimum amount of buffer solution to immerse the MWCNT film (0.6 mL) (Fig. S1†). The active surface of the film was set to 1.77 cm². WGS was started by adding CO using a gastight syringe. The vial was then placed outside the glove box in a temperature-controlled chamber at 150 rpm at 25 °C. The gas phase in the vial was analyzed at regular intervals by sampling 50 μ L and injecting into gas chromatography to evaluate the CO₂ and H₂ formation and CO consumption over time. No traces of O₂ were detected at any stage of the experiment. Fig. 4A shows the representative gas measurement (CO, CO₂, H₂) over time for a MWCNT film modified with **RrCODH** and **NiPNP^{Arg}** after injection of 5 mL of CO in the cell.

CO consumption over time, accompanied by the evolution of both CO₂ and H₂, confirms the WGS activity of the bifunctionalized nanohybrid film (Fig. 4A). As illustrated in Fig. 4B, increasing the quantity of CO injected resulted in corresponding increase in H₂ production with near 100% conversion yield at low CO concentration and 75% at 400 μ mol CO. Under these conditions, no evidence of catalyst inhibition by CO was observed. The unmodified MWCNT films and MWCNT films modified only with the Ni complex do not demonstrate any CO

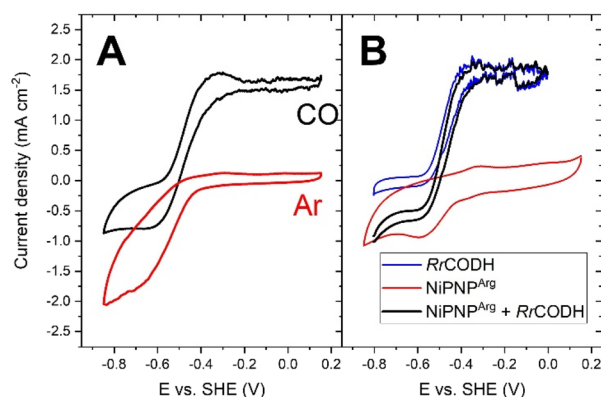


Fig. 3 (A) CV of the bifunctionalized MWCNT electrode under Ar and CO in 50 mM Bis-Tris propane buffer pH 7.0 ($v = 5$ mV s⁻¹); (B) CV of the **Rec-RrCODH**-modified MWCNT electrode (blue) and **NiPNP^{Arg}**-modified MWCNT electrode (red) under CO in 50 mM Bis-Tris propane buffer pH 7.0 ($v = 5$ mV s⁻¹) and the corresponding sum of both CV (black, capacitive contribution of one CV was removed).

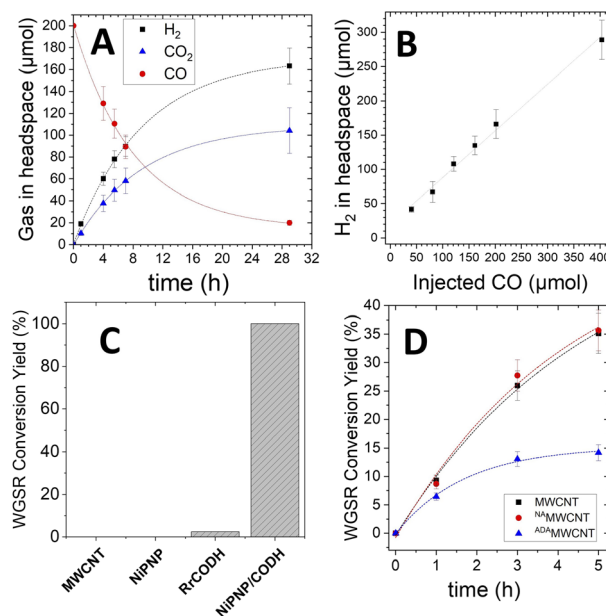


Fig. 4 (A) Quantity of gas in headspace during WGS experiment determined by GC over time for H₂, CO and CO₂ (200 μ mol CO injected at t_0). (B) Quantity of H₂ measured after 24 h at various CO concentration in 50 mM Bis-Tris propane buffer, pH 7.0 at 25 °C (pristine MWCNT film, 0.9 nmol per cm² **Rec-RrCODH** and 110 nmol per cm² **NiPNP^{Arg}**). (C) WGS conversion yield for a nonmodified film, film modified with **NiPNP^{Arg}**, film modified with **Rec-RrCODH** and film modified with both **NiPNP^{Arg}** and **Rec-RrCODH**; (D) quantity of H₂ in headspace of WGS experiment determined by GC over time for pristine MWCNT, ^{NA}MWCNT and ^{ADA}MWCNT in 50 mM Bis-Tris propane buffer pH 7.0 at 25 °C (2 nmol **Rec-RrCODH** and 240 nmol **NiPNP^{Arg}**, 280 μ mol injected CO).

consumption or H_2/CO_2 evolution (Fig. 4C). Conversely, a MWCNT film modified with only **Rec-RrCODH** displays slight amounts of H_2 and CO_2 evolution corresponding to a 2.5% conversion yield. The low activity of **Rec-RrCODH**-modified MWCNT films may be attributed to an inherent low HER activity when the enzyme is immobilized on MWCNTs, as previously described.³⁷

The influence of surface modification of MWCNTs was also investigated to evaluate the effect of immobilization promoters such as pyreneadamantylamide or naphthoate groups. In the case of **NiPNP^{Arg}**, surface loadings were estimated from the integration of the charge under the **NiPNP^{Arg}** redox system obtained by CV in nonturnover conditions at pH 7. This pH was taken in order to partially avoid contribution of the catalytic activity of the complex towards both hydrogen evolution and H_2 oxidation at this pH. It is noteworthy that surface loadings measured by CV corresponds to 6%, 18% and 63% of the amount of **NiPNP^{Arg}** in the soaking solution for pristine MWCNT, ^{ADA}MWCNT and ^{NA}MWCNT, respectively. This result underlines the efficiency of ^{NA}MWCNT to immobilize the catalyst. While MWCNT and ^{ADA}MWCNTs exhibits lower surface loadings, the CV of the Ni complex at ^{ADA}MWCNT electrodes shows that the adamantane functionalization inhibits HER current density (Fig. S2†). This inhibition also correlates with WGSR performance in which the use of pyreneadamantylamide reduces the WGSR activity resulting in a WGSR conversion yield of 40% (Fig. 4D). WGSR experiments show that both pristine MWCNT and naphthoate groups led to a 100% conversion yield, indicating ^{NA}MWCNTs do not provide a significant WGSR improvement. Despite the fact that ^{NA}MWCNT promotes **NiPNP^{Arg}** immobilization efficiency, it does not increase HER significantly as compared to pristine MWCNTs.

Optimization of the WGSR performance was also investigated over a range of pH and upon catalyst loadings. The optimum pH range is between 7.5 and 9 (Fig. 5A). As previously described, lower pHs have a drastic effect on the activity of the enzyme.¹⁷ On the contrary, at high pH, the HER efficiency of **NiPNP^{Arg}** is reduced^{22,38} and limits the overall WGSR efficiency.

Optimum surface coverage was also investigated for both catalysts in terms of H_2 production and overall turnover number (TON) by changing the soaking concentration of each catalyst (Fig. S3†). Surface concentrations for **Rec-RrCODH** is difficult to measure by CV considering the low amount of enzyme as compared to the Ni complex (almost two order of magnitude), the high background current arising from MWCNT capacitance contribution and the overlapping catalytic enzyme activity from the reduction of residual dissolved CO_2 . In order to estimate the amount of immobilized CODH, the UV-visible spectra of the remaining enzyme solution before and after the incubation step was performed (Fig. S4†). Following the absorbance spectra at 280 nm, between 70 and 80% of CODHs in solution are immobilized on CNT electrodes from high to low concentration range respectively.

WGSR performances were measured at difference concentrations of both catalysts. Performances level off for **Rec-RrCODH** concentrations above 0.1 nmol cm^{-2} and **NiPNP^{Arg}** concentrations above 14 nmol cm^{-2} (Fig. S3†). Optimum

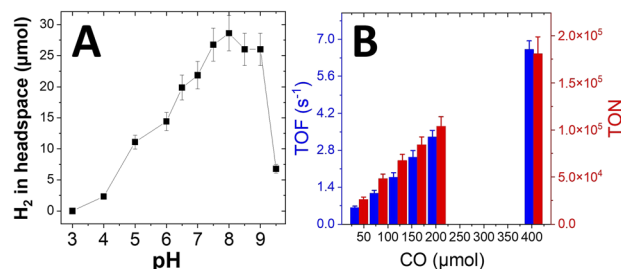


Fig. 5 (A) Quantity of H_2 in headspace of WGSR experiment determined by GC after 24 h versus pH (1 mL CO injected at t_0), $0.7 \text{ nmol per cm}^2$ **Rec-RrCODH** and 14 nmol per cm^2 **NiPNP^{Arg}**, (buffer used: 50 mM citric acid pH 3, 50 mM sodium acetate pH 4 to 5.5, 50 mM Bis-Tris propane buffer pH 6 to 9.5); (B) turnover frequency (TOF) measured after 1 h WGSR, according to the total amount of **Rec-RrCODH**, and turnover number (TON) measured after 24 h versus starting amount of injected CO (50 mM Bis-Tris propane buffer pH 7, $0.9 \text{ nmol per cm}^2$ **Rec-RrCODH** and $110 \text{ nmol per cm}^2$ **NiPNP^{Arg}**).

concentrations underline the fact that more than 100 times less enzyme is required as compared to **NiPNP^{Arg}** to maximize WGSR performances. This mostly arises from the fact that there are several orders of magnitude between the catalytic rate of CO oxidation by **Rec-RrCODH** and the HER catalytic rate by **NiPNP^{Arg}**. Interestingly, when performing WGSR at low **NiPNP^{Arg}** concentration, CO inhibition is observed at 100 μmol CO (Fig. S5†). This might arise from partial CO inhibition of HER which correlates with what is observed on CV from Fig. 3. The optimal compromise for 24 hours experiments was determined to be a concentration of 0.7 nmol cm^{-2} of **Rec-RrCODH** and 14 nmol cm^{-2} of **NiPNP^{Arg}** at which no instability of the catalyst is observed. In addition, this concentration is sufficiently high for preventing CO inhibition to limit WGSR performances. More Ni catalysts is therefore required to counterbalance its lower catalytic activity and its CO sensitivity. In optimized conditions, the more CO is injected the more H_2 is produced with maximum TON of 180 000 after 24 hours and maximum TOF of 6.6 s^{-1} (Fig. 5B). It is noteworthy that a higher TOF value of 30 s^{-1} can be reached by using less enzyme (Fig. S6†). Stability of the WGSR was also investigated over time: after three injections of CO at 0, 25 and 50 h, WGSR performance decreased by 30% after 150 h. After injection at 50 h, the biohybrid catalyst is still active but operates at a slower rate of 38% of the initial apparent TOF (*i.e.* 0.48 s^{-1}) (Fig. S7A†). This is expected from the stability of CODH over time observed in solution and on electrode.^{15,17} Furthermore, oxygen injection was also performed. The injection of 0.5% of oxygen has little effect on the WGSR performance (Fig. S7B†). When 2.5% of oxygen were added during CO injection or added after 3 h, a drastic decrease of the WGSR performance (42% and 34% respectively) is observed. This instability over time or towards oxygen injection mostly relates to the instability of CODHs investigated in solution or on electrode.^{9,15,17,39}

This places our bio-inspired WGSR device as a top performing catalyst at room temperature, in agreement with the fact that CODHs catalyze the CO oxidation reaction in a thermodynamically efficient way, *i.e.* by a direct electronic pathway



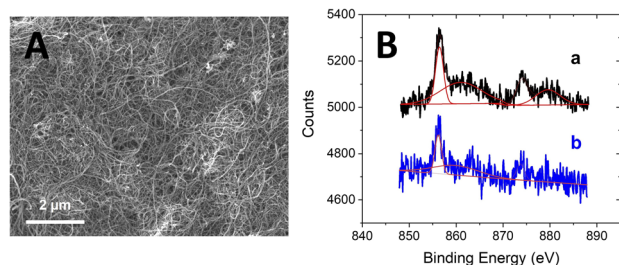


Fig. 6 (A) SEM micrograph and (B) XPS spectra of Ni 2p core energy levels for MWCNT film modified with both NiPNP^{Arg}- and Rec-RrCODH before (a) and after (b) 24 h WGS accompanied with XPS simulation (red line).

between both redox catalysts (without the need of a natural or synthetic redox partner acting as an electron relay). Industrial iron oxide based catalysts and other types of metal-oxide-based catalysts reaches TOF values between 10^{-3} and 4 s^{-1} above 300°C .^{40–42} In addition, the only example of the combination of a CODH and a hydrogenase at graphite pellets reaches TOF of 2.5 s^{-1} at 30°C . This strategy is also limited by the low H_2 production. Only low amounts of CO can be used as it inhibits hydrogenase activity and impact WGS performance.

Surface characterization was performed by Scanning Electron Microscopy (SEM) and X-ray photoelectron spectroscopy (XPS) (Fig. 6). SEM underlines the large surface and porosity of the functionalized MWCNT film (Fig. 6A). XPS analysis was performed before and after 24 h WGS run (Fig. 6B).

The presence of the intact NiPNP^{Arg} complex is confirmed at the Ni 2p core level by the sharp peaks at 856.4 and 874.1 eV accompanied with broad satellite peaks at 863.1 and 880.9 eV characteristics of Ni^{II} species.²⁵ A 32% decrease in the intensity of the Ni signal after 24 h WGS indicates that partial demetallation of the NiPNP^{Arg} complex is observed. This is supported by the fact that N 1s and P 1s core energy levels, respectively observed at 400.9 and 133.2 eV are not impacted as compared to Ni 2p levels (Fig. S8†). Unfortunately, the small amount of Rec-RrCODH compared to that of NiPNP^{Arg} precludes its characterization by XPS in either the Fe 2p or Ni 2p region.

2.3 Optimized bio-inspired WGS for syngas upgrading

Finally, a WGS set-up was performed by using a representative syngas mixture containing 30% CO, 30% H_2 , 20% CO_2 , 10% CH_4 and 10% N_2 with the Rec-RrCODH/NiPNP^{Arg}-modified MWCNT system using previously optimized concentrations. The proportions of H_2 and CO in the mixture were subsequently determined by GC. Over time, the 7 mL of injected syngas was upgraded to a H_2 -rich and CO-poor mixture, with a final H_2 to CO ratio of 4.4 after 24 h, and only 10% of CO remaining. The presence of methane, a typical component of syngas, had no discernible effect on the catalytic performance of the system, with a maximum TOF of 6.6 s^{-1} towards H_2 production. Notably, a H_2 to CO ratio of 2.1 was reached after 5 h, under mild conditions and without the necessity of process gas-catalyst interface optimization. In upgrading syngas systems, this ratio corresponds to that required for future applications

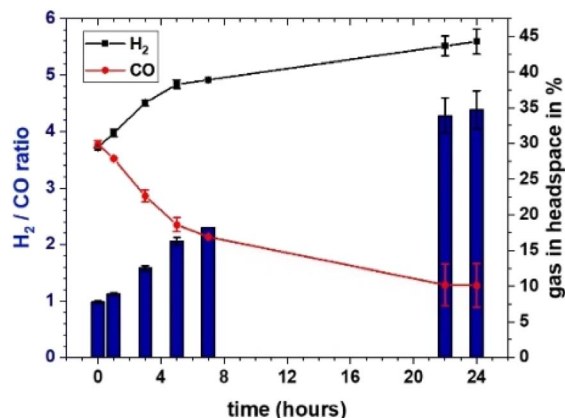


Fig. 7 Evolution of the proportions of H_2 and CO detected by GC in the headspace of a WGS experiment over time and its corresponding H_2/CO ratio (vial flushed with syngas mix (30% CO, 30% H_2 , 20% CO_2 , 10% CH_4 , 10% N_2 at t_0 , corresponding to 7 mL gas mix at ambient pressure and 25°C , $0.9 \text{ nmol per cm}^2$ Rec-RrCODH and $110 \text{ nmol per cm}^2$ NiPNP^{Arg})).

such as the synthesis of methanol or the synthesis of liquid fuels by the Fischer-Tropsch process (Fig. 7).

3 Experimental

3.1 Electrode preparation

Gas diffusion electrodes were modified with MWCNT. A dispersion of MWCNT in 50% H_2O and 50% ethanol solution was prepared by 4 h sonication of 1 mg mL^{-1} MWCNT in solution. 2 mL of dispersion were drop casted on a 15 mm diameter GDE, and the solvent was removed under vacuum. The GDE was rinsed with ethanol, and reduced to 3 mm diameter before using.

3.1.1 Functionalization of MWCNT with 1-pyrenebutyric adamantyl amide acid, naphthoate function, NiPNP^{Arg} complex and Rec-RrCODH. MWCNT-modified electrodes were soaked for 30 minutes in DMF solution containing 5 mM 1-pyrenebutyric acid adamantyl amide and after rinsed in DMF solution and two times in MilliQ water. MWCNT-modified electrodes were soaked for 30 minutes in DMF solution containing 1 mM 4-carboxylatonaphtyldiazonium tetrafluoroborate and after rinsed in DMF solution and two times in MilliQ water. The Rec-RrCODH modified electrodes were prepared by incubating MWCNT electrodes for 1 hour with 20 μL of enzyme solution in 50 mM Tris-HCl buffer pH 8.5 (2 mg mL^{-1}). The latter step was carried out inside an anaerobic glove box ($\text{O}_2 < 2 \text{ ppm}$, Jacomex). The electrodes were finally washed with 50 mM Tris-HCl buffer pH 8.5. When not used the electrodes were kept in buffer 50 mM Tris-HCl, pH 8.5. The NiPNP^{Arg} modified electrodes were prepared by incubation with an aqueous 2 mM NiPNP^{Arg} solution for 1 h. The electrode was rinsed with water, and kept in water until use.

3.1.2 WGS experiments. MWCNT were deposited on a hydrophilic PTFE filter (Sigma, 65 μm thickness, 0.45 μm pore size, 80% porosity). A dispersion of MWCNT in 50% H_2O and 50% ethanol solution was prepared by 4 h sonication of 1 mg



mL⁻¹ MWCNT in solution. 2 mL of dispersion were drop casted on a 15 mm diameter filter, and the solvent was removed under vacuum. The deposits were rinsed with ethanol, dried under vacuum, and stored in water until use. Deposits were functionalized with 1-pyrenebutyric acid adamantyl amide and 4-carboxylatonaphthyl diazonium tetrafluoroborate in the same way as the gas diffusion electrode (GDE). In an anaerobic glove box, a deposit was placed on the bottom of an 8 mL vial. **NiPNP^{Arg}** modification was carried out by incubating the deposit with 60 µL of aqueous **NiPNP^{Arg}** solution for 12 h at 4 °C. The solution was then removed, and the deposit was rinsed with water. 60 µL of **Rec-RrCODH** solution in 50 mM Tris-HCl buffer pH 8.5 were then added for incubation. After 1 h, the solution was removed and the deposit was rinsed with 50 mM Bis-Tris propane buffer pH 7.6. 600 µL of 50 mM Bis-Tris propane buffer pH 7 were added to the vial, which was then closed using a rubber septa.

WGSr was started by adding CO or syngas mix using a gastight syringe. The vial was then put in a temperature-controlled chamber under agitation at 25 °C. The gas phase in the vial was analyzed at regular interval by sampling 50 µL and injecting in gas chromatography.

4 Conclusions

This work demonstrates the coupling of a high-performance enzyme for CO-to-CO₂ oxidation with a Ni complex catalyzing efficiently the hydrogen evolution reaction, at MWCNTs. This dual catalytic nanohybrid system achieves efficient WGSr with CO or syngas as substrate, working over a period of 24 hours, with TOF reaching a maximum of 30 s⁻¹ under mild conditions. This makes it a proof of concept for the development of a bio-inspired WGSr catalyst. Notably, the isolated CODH: Ferredoxin: Hydrogenase complex from *Carboxydotherrmus hydrogenoformans*, reaches a WGSr activity of 1700 s⁻¹ at 70 °C.⁴⁰ This underlines the fact that there is still room for improvement if such TOF values could be achieved in a nano-structured nanohybrid system. For example, the development of WGSr with more robust catalysts towards oxygen or temperature *via* protein engineering or enzyme selection is one possible strategy. It is noteworthy that the two catalysts used in this study are bidirectional, allowing their potential use in reverse water-gas shift reaction (rWGSr, CO₂ + H₂ ⇌ CO + H₂O) with applications in power-to-syngas processes.

In the future, the system developed here could therefore be optimized for WGSr in its direct or reverse way for its integration in operational devices.

Data availability

The data supporting this article have been included as part of the ESI.†

Author contributions

T. Pichon: investigation, formal analysis, C. Righetti: investigation, formal analysis, J. Pérard: investigation, formal analysis,

A. Le Goff: conceptualization, funding acquisition, supervision, writing original draft, writing review and editing, C. Cavazza: conceptualization, funding acquisition, supervision, writing original draft, writing review and editing.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the Agence Nationale de la Recherche through the LabEx ARCANÉ program (ANR-11-LABX-0003-01) and the Graduate School on Chemistry, Biology and Health of Univ Grenoble Alpes CBH-EUR-GS (ANR-17-EURE-0003). This work was supported by “the NTE PhD program, CEA” for TP’s PhD funding. This work has benefited from French State aid managed by the Agence Nationale de la Recherche under France 2030 plan, bearing the reference code ANR-22-PESP-0010: Projet ciblé “POWERCO2” within the PEPR project SPLEEN. We kindly thank Dr Bertrand Reuillard for his help for the synthesis of the **NiPNP^{Arg}** complex. Valérie Flaud and Eddy Petit from Institut Charles Gerhardt (University of Montpellier 2) are gratefully acknowledged for XPS analysis.

Notes and references

- 1 A. Galadima and O. Muraza, *J. Nat. Gas Sci. Eng.*, 2015, **25**, 303–316.
- 2 Ö. Tezer, N. Karabağ, A. Öngen, C. Ö. Çolpan and A. Ayol, *Int. J. Hydrogen Energy*, 2022, **47**, 15419–15433.
- 3 E. Baraj, K. Ciahotný and T. Hlinčík, *Fuel*, 2021, **288**, 119817.
- 4 B. Soboh, D. Linder and R. Hedderich, *Eur. J. Biochem.*, 2002, **269**, 5712–5721.
- 5 V. C.-C. Wang, S. W. Ragsdale and F. A. Armstrong, *ChemBioChem*, 2013, **14**, 1845–1851.
- 6 M. Can, F. A. Armstrong and S. W. Ragsdale, *Chem. Rev.*, 2014, **114**, 4149–4174.
- 7 A. Parkin, J. Seravalli, K. A. Vincent, S. W. Ragsdale and F. A. Armstrong, *J. Am. Chem. Soc.*, 2007, **129**, 10328–10329.
- 8 T. W. Woolerton, S. Sheard, E. Reisner, E. Pierce, S. W. Ragsdale and F. A. Armstrong, *J. Am. Chem. Soc.*, 2010, 2132–2133.
- 9 E. C. Wittenborn, C. Guendon, M. Merrouch, M. Benvenuti, V. Fourmond, C. Léger, C. L. Drennan and S. Dementin, *ACS Catal.*, 2020, **10**, 7328–7335.
- 10 W. Lubitz, H. Ogata, O. Rüdiger and E. Reijerse, *Chem. Rev.*, 2014, **114**, 4081–4148.
- 11 M. G. Allan, T. Pichon, J. A. McCune, C. Cavazza, A. Le Goff and M. F. Kühnel, *Angew. Chem., Int. Ed.*, 2023, **62**, e202219176.
- 12 N. Lalaoui, A. de Poulpique, R. Haddad, A. Le Goff, M. Holzinger, S. Gounel, M. Mermoux, P. Infossi, N. Mano, E. Lojou and S. Cosnier, *Chem. Commun.*, 2015, **51**, 7447–7450.



- 13 L. Domnik, M. Merrouch, S. Goetzl, J.-H. Jeoung, C. Leger, S. Dementin, V. Fourmond and H. Dobbek, *Angew. Chem., Int. Ed.*, 2017, **56**, 15466–15469.
- 14 O. Lazarus, T. W. Woolerton, A. Parkin, M. J. Lukey, E. Reisner, J. Seravalli, E. Pierce, S. W. Ragsdale, F. Sargent and F. A. Armstrong, *J. Am. Chem. Soc.*, 2009, **131**, 14154–14155.
- 15 U. Contaldo, M. Curtil, J. Pérard, C. Cavazza and A. Le Goff, *Angew. Chem., Int. Ed.*, 2022, **134**, e202117212.
- 16 S. Gentil, S. M. Che Mansor, H. Jamet, S. Cosnier, C. Cavazza and A. Le Goff, *ACS Catal.*, 2018, 3957–3964.
- 17 U. Contaldo, B. Guigliarelli, J. Perard, C. Rinaldi, A. Le Goff and C. Cavazza, *ACS Catal.*, 2021, 5808–5817.
- 18 M. A. Alonso-Lomillo, O. Rüdiger, A. Maroto-Valiente, M. Velez, I. Rodríguez-Ramos, F. J. Muñoz, V. M. Fernández and A. L. De Lacey, *Nano Lett.*, 2007, **7**, 1603–1608.
- 19 A. Le Goff and M. Holzinger, *Sustain. Energy Fuels*, 2018, **2**, 2555–2566.
- 20 A. D. Wilson, R. H. Newell, M. J. McNevin, J. T. Muckerman, M. Rakowski DuBois and D. L. DuBois, *J. Am. Chem. Soc.*, 2006, **128**, 358–366.
- 21 S. Raugei, S. Chen, M.-H. Ho, B. Ginovska-Pangovska, R. J. Rousseau, M. Dupuis, D. L. DuBois and R. M. Bullock, *Chem. Commun.*, 2012, 6493–6506.
- 22 A. Dutta, J. A. S. Roberts and W. J. Shaw, *Angew. Chem., Int. Ed.*, 2014, **53**, 6487–6491.
- 23 A. Dutta, D. L. DuBois, J. A. S. Roberts and W. J. Shaw, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, 16286–16291.
- 24 A. A. Oughli, A. Ruff, N. P. Boralugodage, P. Rodríguez-Maciá, N. Plumeré, W. Lubitz, W. J. Shaw, W. Schuhmann and O. Rüdiger, *Nat. Commun.*, 2018, **9**, 864.
- 25 A. Le Goff, V. Artero, B. Jousselme, P. D. Tran, N. Guillet, R. Metaye, A. Fihri, S. Palacin and M. Fontecave, *Science*, 2009, **326**, 1384–1387.
- 26 N. Coutard, B. Reuillard, T. N. Huan, F. Valentino, R. T. Jane, S. Gentil, E. Andreiadis, A. Le Goff, T. Asset, F. Maillard, B. Jousselme, A. Morozan, S. Lyonnard, V. Artero and P. Chenevier, *Chem Catal.*, 2021, **1**(1), 88–105.
- 27 S. Gentil, N. Lalaoui, A. Dutta, Y. Nedellec, S. Cosnier, W. J. Shaw, V. Artero and A. Le Goff, *Angew. Chem., Int. Ed.*, 2017, **56**, 1845–1849.
- 28 P. D. Tran, A. Le Goff, J. Heidkamp, B. Jousselme, N. Guillet, S. Palacin, H. Dau, M. Fontecave and V. Artero, *Angew. Chem., Int. Ed.*, 2011, **50**, 1371–1374.
- 29 F. C. Macazo and S. D. Minter, *Curr. Opin. Electrochem.*, 2017, **5**, 114–120.
- 30 C. Agnès, B. Reuillard, A. Le Goff, M. Holzinger and S. Cosnier, *Electrochem. commun.*, 2013, 105–108.
- 31 B. Reuillard, A. Le Goff and S. Cosnier, *Anal. Chem.*, 2014, **86**, 4409–4415.
- 32 B. Reuillard, A. Le Goff and S. Cosnier, *Chem. Commun.*, 2014, **50**, 11731–11734.
- 33 M. Yuan, M. J. Kummer and S. D. Minter, *Chem.–Eur. J.*, 2019, **25**, 14258–14266.
- 34 Z. Wu, Z. Chen, X. Du, J. M. Logan, J. Sippel, M. Nikolou, K. Kamaras, J. R. Reynolds, D. B. Tanner, A. F. Hebard and A. G. Rinzler, *Science*, 2004, **305**, 1273–1276.
- 35 B. Reuillard, C. Costentin and V. Artero, *Angew. Chem., Int. Ed.*, 2023, **62**, e202302779.
- 36 A. D. Wilson, K. Frazee, B. Twamley, S. M. Miller, D. L. DuBois and M. Rakowski DuBois, *J. Am. Chem. Soc.*, 2008, **130**, 1061–1068.
- 37 S. Menon and S. W. Ragsdale, *Biochemistry*, 1996, **35**, 15814–15821.
- 38 S. Gentil, N. Lalaoui, A. Dutta, Y. Nedellec, S. Cosnier, W. J. Shaw, V. Artero and A. Le Goff, *Angew. Chem., Int. Ed.*, 2017, **56**, 1845–1849.
- 39 M. Merrouch, J. Hadj-Saïd, L. Domnik, H. Dobbek, C. Léger, S. Dementin and V. Fourmond, *Chem.–Eur. J.*, 2015, **21**, 18934–18938.
- 40 B. Soboh, D. Linder and R. Hedderich, *Eur. J. Biochem.*, 2002, **269**, 5712–5721.
- 41 R. B. King, *J. Organomet. Chem.*, 1999, **586**, 2–17.
- 42 J. A. Rodriguez, S. Ma, P. Liu, J. Hrbek, J. Evans and M. Pérez, *Science*, 2007, **318**, 1757–1760.

