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Synthesis and bioactivity of psilocybin analogues containing a stable carbon–phosphorus bond†

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Psilocybin analogues have been synthesized comprising a non-hydrolysable P–C bond to evaluate the biological activity and the selectivity towards 5-HT_{2A}R, 5-HT_{2B}R and the TNAP receptor. No activity was observed towards the phosphatase, however all compounds showed good binding affinity for 5-HT_{2A}R and 5-HT_{2B}R and one compound showed a higher selectivity towards 5-HT_{2A}R than psilocin.

The precipitous escalation, quantified at 25%, in the global prevalence of anxiety and depression in the wake of the COVID-19 pandemic serves as an alarming indicator of a profound and pervasive mental health crisis.^{1,2} Despite the diverse array of treatments available, the well-known first-line antidepressants exhibit limited efficacy. The treatment of major depressive disorder (MDD) with conventional drugs does not only grapple with a therapeutic delay of at least two weeks, thereby increasing the vulnerability to self-harm and suicide, but also with adverse effects such as sexual dysfunction.^{3,4} Another factor that amplifies the socio-economic burden of MDD is treatment resistant depression (TRD) in which remission subsequent to treatment has failed. Approximately 30% of adults suffering from MDD experience inadequate response to the administration of at least two different antidepressants of adequate dose and duration.^{5,6} The aforementioned deficiencies emphasize the urgency with which the scientific community must redirect its focus towards alternatives to the commonly used, yet suboptimal first-line antidepressants.

In this context, psychedelic drugs such as psilocybin, LSD, ketamine and MDMA have gained prominence as potential candidates for the treatment of psychiatric disorders.^{7,8} Notably, the regulatory approvals in March and December 2019 for intranasal administration of esketamine in conjunction with a conventional antidepressant for adults suffering from TRD are evidence of the evolving landscape.⁹ Completed phase 2 clinical trials show that psilocybin-assisted therapies produce large, rapid and long-lasting therapeutic effects among patients with MDD due to

increased spine density and brain network integration.^{10–12} Despite all promising insights, the hallucinogenic nature of psychedelic drugs necessitates supervised administration, thereby hampering its widespread use.¹³ Four recent studies exploring the design and synthesis of non-hallucinogenic, non-toxic compounds offer potential solutions to this constraint.^{14–17} Another challenge in the search for new leads is the selectivity over off-targets such as 5-HT_{2B}R where its activation is associated with valvular heart disease.¹⁸

Recent investigations identified a second binding mode of psilocin in the orthosteric site of 5-HT_{2A}R which triggers only a modest activation of β -arrestin signaling. This nuanced mechanism results in antidepressive and anxiolytic effects devoid of hallucinogenic manifestations.^{15,19} Moreover, psilocybin's lower addiction potential and reduced toxicity compared to ketamine underscore its compelling potential as alternative antidepressant.^{20–22}

Psilocybin (*O*-phosphoryl-4-hydroxy-*N,N*-dimethyltryptamine) (Chart 1), a tryptamine alkaloid found in psilocybe mushrooms species,²³ undergoes dephosphorylation in the human body primarily facilitated by an alkaline phosphatase. This process yields its active metabolite psilocin, which is active on 5-HT_{2A}R and 5-HT_{2B}R in our brain, among others.²⁴

Considering the significance of the alkaline phosphatase in drug design and delivery,^{25,26} coupled with recent insights into the application of phosphonates in medicinal chemistry,²⁷ our present work endeavors to ascertain the activity of phosphonate analogues of psilocybin on the 5-HT_{2A}

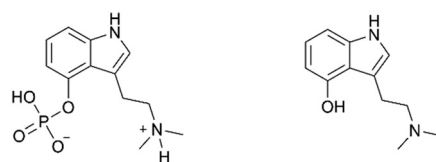


Chart 1 Structures of psilocybin (left) and psilocin (right).

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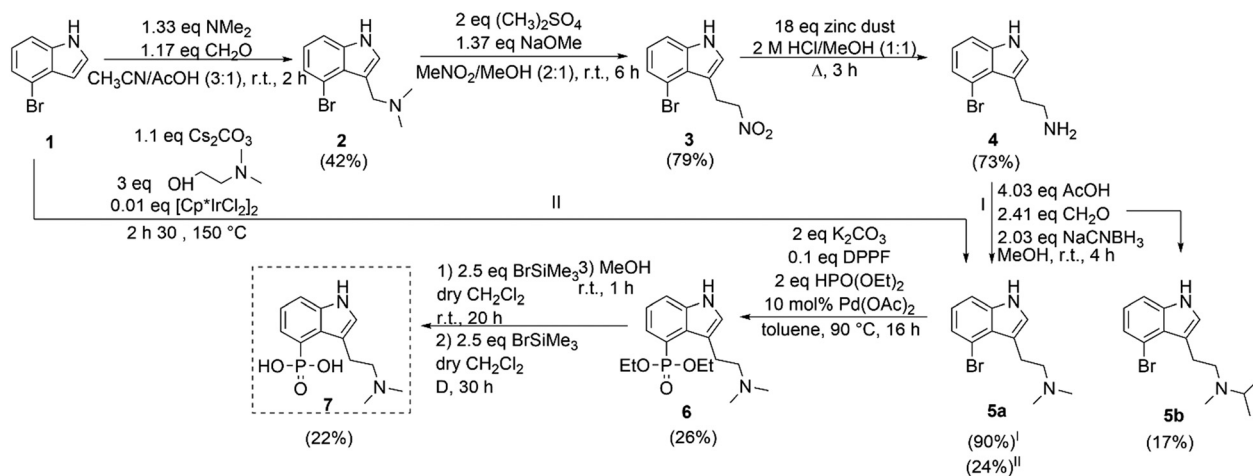
and 5-HT_{2B} receptors. Notable, investigations into the inhibition of alkaline phosphatase by phosphonates remain absent in the existing literature. Hence, we aim to elucidate their interactions with and potential inhibition of the alkaline phosphatase, thereby contributing to the understanding and development of novel therapeutic strategies for mental health disorders. In alignment with this objective, we have innovatively developed a new synthesis pathway and optimized reaction conditions for phosphonoindoles and phosphonotryptamines.

The introduction of a CH₂ group adjacent the aromatic ring results in a more isosteric analogue of psilocybin. However, this modification increases the count of rotational bonds. Conversely, the phosphonate and phosphonic acid analogues devised in this study feature few rotational bonds allowing for a conformation in the receptors of interests which more closely mirrors that of psilocin. Moreover, the synthesis of the variant at the 4-position would prove challenging owing to the lack of stereoselectivity inherent in the Fischer-indole reaction.

To obtain the target structure 7, a new pathway starting from 4-bromo indole 1 (Scheme 1) was investigated. The first step included a Mannich transformation into compound 2, which was used in the subsequent introduction of a nitromethane anion to obtain compound 3. The latter was further reduced using zinc dust in a sulfuric acid/methanol mixture. ¹H-NMR analysis revealed low purity of the product 4 which necessitated the use of less harsh reaction conditions such as a hydrochloric acid/methanol (1:1) mixture. An initial yield of 34% was obtained under these conditions, which could be further increased to 73% by minimizing the use of water in the work-up. Next, a variant of the Eschweiler–Clarke reaction which entails the methylation of a primary or secondary amine, gives access to compound 5a. In this particular case, sodium cyanoborohydride was used for reductive amination. Due to a contamination by acetone of the solvent used in the reaction, side product 5b with an

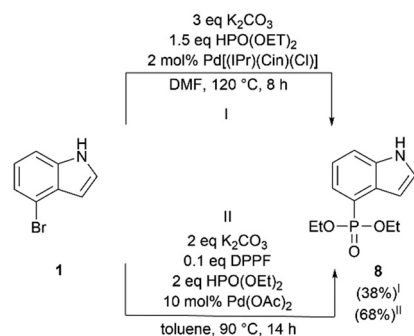
isopropyl group instead of a methyl group was formed. Optimization of the reaction led to the formation of compound 5a, which was obtained with an average yield of 90%. Method I, characterized by its simplicity, employs readily accessible, cheaper as well as less hazardous reagents and gives higher yields compared to preceding synthesis pathways of 5a.^{28,29} Recently, we developed a second pathway (Scheme 1) towards 5a, which consists of only one step and makes use of the borrowing hydrogen strategy. Method II reduces the time required to synthesize compound 5a but requires the use of a more expensive [Cp*IrCl₂]₂ catalyst.

During pathway optimization, different reaction conditions were tested in parallel on the more accessible 5-bromo-*N,N*-dimethyltryptamine (Table S1†). First, the Michaelis-Arbuzov reaction was evaluated under mild reaction conditions using BF₃·Et₂O. After 4 h, ³¹P-NMR revealed the presence of triethyl phosphite, the product of its hydrolysis and its oxidation (triethyl phosphate). However, no evidence of product formation was observed. Rasheed *et al.* demonstrated that the phosphonylation of 5-bromo indole is feasible with a Michaelis-Arbuzov reaction in an ionic liquid (bbim[Br]).³⁰ Reactions on 4-bromo indole and 5-bromo tryptamine in the ionic liquid failed to form the product, leaving only triethyl phosphite, diethyl phosphite and triethyl phosphate according to ³¹P-NMR. Additionally, we evaluated a nickel-catalyzed Tavares reaction according to the procedure of Shearan *et al.*³¹ Again, no evidence of product formation was found after 54 h of stirring at 160 °C. Another option to introduce a phosphonate group is the Hirao coupling.³² Literature research revealed the lack of Hirao couplings performed on indole containing compounds. To evaluate the feasibility of phosphonotryptamine synthesis, we initially investigated the synthesis of compound 8 (Table S2,† Scheme 2). Starting from 4-bromo indole, the desired compound was synthesized using a non-commercial NHC catalyst (Pd[(IPr)(Cin)(Cl)])³³ under the reaction conditions described by Xu *et al.* with an isolated yield of 38%.



Scheme 1 Optimized pathway for the synthesis of the target compound (7) via I or II.





Scheme 2 Synthesis of diethyl (1*H*-indol-4-yl)phosphonate via I or II.

These reaction conditions were then evaluated for 5-bromo-*N,N*-dimethyltryptamine, however only 29% conversion was achieved according to ^{31}P -NMR. In experiment 5 and 6 (Table S1†), $\text{Pd}(\text{OAc})_2$ and K_2CO_3 were evaluated. The reaction mixture was stirred under a continuous nitrogen flow, but no full conversion was observed. Changing the base to Et_3N in a THF solution, as described by Francesconi *et al.*, resulted in an even lower conversion (12%).³⁴ In experiment 8, similar reaction conditions as in experiment 6 were applied but the flask was carefully flushed with nitrogen and sealed, resulting in full conversion and isolation of compound **9** in a high yield (95%, Table S1†). It is worth mentioning that the same approach is applicable to the synthesis of compound **8**. Moreover, these reaction conditions provided a cleaner reaction mixture and a higher isolated yield of 68%. A Mannich reaction was also evaluated on compound **8** but this resulted in a complex reaction mixture. Hence, this path was not further investigated.

With the optimized reaction conditions established, we expanded the library from indoles to tryptamines. First, compound **6**, which was only one step away from the target structure, was synthesized following the same procedure as mentioned before. To evaluate the effect of the length and bulkiness of the alkyl side chain, compounds **2** and **5b** were phosphonylated as well to obtain compounds **10** and **11** in isolated yields of 9% and 10%, respectively.

Subsequently, phosphonates were dealkylated to their corresponding phosphonic acids. The mild McKenna method was favored for the compounds of interest. The hydrolysis of compound **9** was achieved after 20 h of stirring at room temperature, followed by stirring in methanol for 1 h. No further purification was required, and **12** was isolated in 82% yield. Hydrolysis of compound **6** went less smoothly as 50% of the start product was partially silylated and 50% fully silylated after 20 h of stirring at room temperature according to ^{31}P -NMR. After five days, no further changes were detected. Hence, the reaction mixture was refluxed at 55 °C. The solvent and bromotrimethylsilane were then periodically reintroduced into the mixture. After an additional 30 h, the conversion was complete and methanol was added.

After the chemical synthesis of the target compound **6** and five other compounds (Chart 2), we wanted to verify our two hypotheses. To evaluate the affinity of the phosphonate

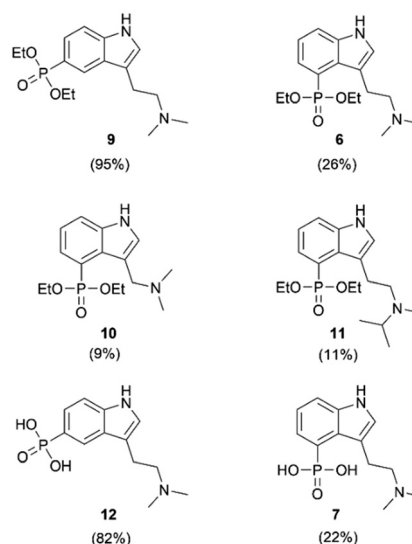


Chart 2 Overview of synthesized and *in vitro* tested compounds with corresponding yields.

analogues towards 5-HT_{2A} and 5-HT_{2B}R, a competitive radioligand binding assay was conducted. Additionally, we wanted to determine whether these compounds could influence the metabolism, possibly as inhibitors of alkaline phosphatase. To answer this question, a tissue-nonspecific alkaline phosphatase (TNAP) assay was performed. The results of the initial screening are presented in Table S3.†

Surprisingly, the results of the *in vitro* assays revealed that all synthesized compounds lacked affinity for TNAP. This refutes our initial hypothesis regarding the inhibitory effects of psilocybin phosphonate analogs on the enzyme, but gives us more information regarding the metabolic stability of the compounds.

However, the results for 5-HT_{2A}R and 5-HT_{2B}R are promising and show a clear structure–activity relationship. For 5-HT_{2B}R, the phosphonate esters of *N,N*-dimethyltryptamine (DMT) **9** and **6** exhibited the highest inhibitory activity. The phosphonic acids show a significantly lower affinity for 5-HT_{2B}R and the phosphonate ester of gramine **10** displays the lowest affinity of all compounds. Thus, the synthesized compounds can be ranked in terms of increasing affinity for 5-HT_{2B}R as follows: phosphonate esters of DMT > phosphonate ester of *N*-methyl-*N*-isopropyltryptamine (MIPT) > phosphonic acids of DMT > phosphonate esters of gramine (**9** > **6** > **11** > **7** > **12** > **10**).

A similar trend is observed for the activity at 5-HT_{2A}R: **9** > **6** > **12** > **11** > **7** > **10**. The phosphonate esters of DMT continue to exhibit the highest affinity and the phosphonate ester of gramine has the lowest affinity for this receptor. Interestingly, unlike the 5-HT_{2B} receptor, the phosphonic acid at the 5-position demonstrates a higher affinity compared to the phosphonic acid at the 4-position. In general, the inhibition percentages of all compounds are lower for 5-HT_{2A}R compared to 5-HT_{2B}R. Chadeayne *et al.* reported that psilocybin (with a phosphoryloxy group)



Table 1 Overview of inhibition constants (K_i) for compound **9** on 5-HT_{2A}R and 5-HT_{2B}R

Compound	5-HT _{2A} R (nM)	5-HT _{2B} R (nM)
9	620	140
Psilocin ³⁵	107.2	4.6
Psilocybin ³⁵	>10 000	98.7

displays a higher selectivity towards 5-HT_{2B}R compared to psilocin.³⁵ While the synthesized compounds contain a phosphorus group similar to psilocybin, their selectivity for 5-HT_{2B}R is relatively less prominent. Furthermore, the observed lower affinity of compound **11** for both 5-HT_{2A}R and 5-HT_{2B}R is consistent with the findings by McKenna *et al.* elaborating the comparison between 4-OH-DMT and 4-OH-MIPT.³⁶ For compound **9**, further analysis was performed to determine IC₅₀ and inhibition constants (K_i) on 5-HT_{2A}R and 5-HT_{2B}R. The dose–response curves and a comparison of the inhibition constants of compound **9** with values for psilocin and psilocybin found in literature, are represented in Fig. S10† and Table 1.³⁵

Compound **9** shows a higher binding affinity than psilocybin for 5-HT_{2A}R, but a lower although comparable binding affinity for 5-HT_{2B}R. Psilocin is two orders of magnitude more potent than compound **9** for 5-HT_{2B}R, but the difference is smaller for 5-HT_{2A}R. Consequently, the selectivity of compound **9** for 5-HT_{2B}R is *circa* 5 times lower than is the case for psilocin which addresses the previously mentioned concerns regarding valvular diseases.¹⁸

In the present work, we describe the successful and innovative synthesis of seven phosphonate analogs of indoles *via* a Hirao coupling. Six analogs were subjected to activity testing on three receptors, 5-HT_{2A}R, 5-HT_{2B}R, and TNAP, to investigate the influence of the compounds on metabolism and serotonergic receptors in the brain. Interestingly, none of the compounds showed affinity for TNAP, thereby disproving a possible theory of inhibition of the alkaline phosphatase enzyme by the compounds. Results at the serotonergic receptors showed a clear structure–activity relationship, which provides insights for future selective drug design. Compound **9** showed the highest potency of all the tested compounds. Despite its lower potency compared to psilocin, the selectivity for 5-HT_{2B}R is significantly lower which is favorable with regards to valvular heart diseases.

Leveraging the information from the abovementioned results, the search for candidates with a higher selectivity regarding G protein and β -arrestin signaling and other off-targets can become more oriented. Further *in vivo* tests can provide us with more insights into the effects of the synthesized compounds.

Author contributions

Marthe Vandeveld: data curation, formal analysis, investigation, methodology, visualization, writing – original draft, writing – review & editing, conceptualization. Andreas

Simoens: data curation, formal analysis, conceptualization, project administration, writing – original draft, writing – review & editing, supervision. Bavo Vandekerckhove: writing – review & editing, investigation, methodology. Christian Stevens: conceptualization, project administration, supervision, validation, resources, funding acquisition, writing – review & editing.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- M. Daly and E. Robinson, *Lancet*, 2022, **399**, 518.
- Z. Li, M. Ruan, J. Chen and Y. Fang, *Neurosci. Bull.*, 2021, **37**, 863–880.
- E. Jing and K. Straw-Wilson, *Ment. Health Clin.*, 2016, **6**, 191–196.
- J. M. Witkin, A. E. Martin, L. K. Golani, N. Z. Xu and J. L. Smith, in *Advances in Pharmacology*, ed. J. M. Witkin, Academic Press, 2019, vol. 86, pp. 47–96.
- S. Karkare, M. Zhdanava, D. Pilon, A. I. Nash, L. Morrison, A. Shah, P. Lefebvre and K. Joshi, *Clin. Ther.*, 2022, **44**, 1432–1448.
- M. Zhdanava, D. Pilon, I. Ghelerter, W. Chow, K. Joshi, P. Lefebvre and J. J. Sheehan, *J. Clin. Psychiatry*, 2021, **82**(2), DOI: [10.4088/JCP.20m13699](https://doi.org/10.4088/JCP.20m13699).
- J. M. Mitchell, G. M. O'talora, B. van der Kolk, S. Shannon, M. Bogenschutz, Y. Gelfand, C. Paleos, C. R. Nicholas, S. Quevedo, B. Balliett, S. Hamilton, M. Mithoefer, S. Kleiman, K. Parker-Guilbert, K. Tzarfaty, C. Harrison, A. de Boer, R. Doblin, B. Yazar-Klosinski and M. S. C. Group, *Nat. Med.*, 2023, **29**, 2473–2480.
- W. Duan, D. Cao, S. Wang and J. Cheng, *Chem. Rev.*, 2024, **124**(1), 124–163.
- R. S. McIntyre, J. D. Rosenblat, C. B. Nemeroff, G. Sanacora, J. W. Murrough, M. Berk, E. Brietzke, S. Dodd, P. Gorwood, R. Ho, D. V. Iosifescu, C. Lopez Jaramillo, S. Kasper, K. Kratiuk, J. G. Lee, Y. Lee, L. M. W. Lui, R. B. Mansur, G. I. Papakostas, M. Subramaniapillai, M. Thase, E. Vieta, A. H. Young, C. A. Zarate, Jr. and S. Stahl, *Am. J. Psychiatry*, 2021, **178**, 383–399.
- A. K. Davis, F. S. Barrett, D. G. May, M. P. Cosimano, N. D. Sepeda, M. W. Johnson, P. H. Finan and R. R. Griffiths, *JAMA Psychiatry*, 2021, **78**, 481–489.
- R. E. Daws, C. Timmermann, B. Giribaldi, J. D. Sexton, M. B. Wall, D. Erritzoe, L. Roseman, D. Nutt and R. Carhart-Harris, *Nat. Med.*, 2022, **28**, 844–851.
- L. X. Shao, C. Liao, I. Gregg, P. A. Davoudian, N. K. Savalia, K. Delagarza and A. C. Kwan, *Neuron*, 2021, **109**, 2535–2544e2534.
- R. F. Service, *Science*, 2022, **375**, 370.



- 14 L. P. Cameron, R. J. Tombari, J. Lu, A. J. Pell, Z. Q. Hurley, Y. Ehinger, M. V. Vargas, M. N. McCarroll, J. C. Taylor, D. Myers-Turnbull, T. Liu, B. Yaghoobi, L. J. Laskowski, E. I. Anderson, G. Zhang, J. Viswanathan, B. M. Brown, M. Tjia, L. E. Dunlap, Z. T. Rabow, O. Fiehn, H. Wulff, J. D. McCorvy, P. J. Lein, D. Kokel, D. Ron, J. Peters, Y. Zuo and D. E. Olson, *Nature*, 2021, **589**, 474–479.
- 15 D. Cao, J. Yu, H. Wang, Z. Luo, X. Liu, L. He, J. Qi, L. Fan, L. Tang, Z. Chen, J. Li, J. Cheng and S. Wang, *Science*, 2022, **375**, 403–411.
- 16 A. L. Kaplan, D. N. Confair, K. Kim, X. Barros-Álvarez, R. M. Rodriguiz, Y. Yang, O. S. Kweon, T. Che, J. D. McCorvy, D. N. Kamber, J. P. Phelan, L. C. Martins, V. M. Pogorelov, J. F. DiBerto, S. T. Slocum, X.-P. Huang, J. M. Kumar, M. J. Robertson, O. Panova, A. B. Seven, A. Q. Wetsel, W. C. Wetsel, J. J. Irwin, G. Skiniotis, B. K. Shoichet, B. L. Roth and J. A. Ellman, *Nature*, 2022, **610**, 582–591.
- 17 C. Dong, C. Ly, L. E. Dunlap, M. V. Vargas, J. Sun, I.-W. Hwang, A. Azinfar, W. C. Oh, W. C. Wetsel, D. E. Olson and L. Tian, *Cell*, 2021, **184**, 2779–2792.e2718.
- 18 B. L. Roth, *N. Engl. J. Med.*, 2007, **356**, 6–9.
- 19 Y. N. Yin and T. M. Gao, *Neurosci. Bull.*, 2023, **39**, 170–172.
- 20 M. W. Johnson, R. R. Griffiths, P. S. Hendricks and J. E. Henningfield, *Neuropharmacology*, 2018, **142**, 143–166.
- 21 R. S. Gable, *Am. J. Drug Alcohol Abuse*, 1993, **19**, 263–281.
- 22 L. Orsolini, V. Salvi and U. Volpe, *Expert Opin. Drug Saf.*, 2022, **21**, 803–812.
- 23 M. Coppola, F. Bevione and R. Mondola, *J. Xenobiot.*, 2022, **12**, 41–52.
- 24 H. A. Geiger, M. G. Wurst and R. N. Daniels, *ACS Chem. Neurosci.*, 2018, **9**, 2438–2447.
- 25 K. Moonen, I. Laureyn and C. V. Stevens, *Chem. Rev.*, 2004, **104**, 6177–6216.
- 26 B. Le-Vinh, Z. B. Akkuş-Dağdeviren, N. M. N. Le, I. Nazir and A. Bernkop-Schnürch, *Adv. Ther.*, 2022, **5**, 2100219.
- 27 M. Krecmerova, P. Majer, R. Rais and B. S. Slusher, *Front. Chem.*, 2022, **10**, 889737.
- 28 M. Alzweiri, A. S. Alsegiani, E. Karaj, D. A. Almarghalani, Y. Tabaza, Z. A. Shah and L. M. V. Tillekeratne, *Bioorg. Med. Chem. Lett.*, 2021, **47**, 128205.
- 29 E. K. Olsen, E. Hansen, L. W. K. Moodie, J. Isaksson, K. Sepčić, M. Cergolj, J. Svenson and J. H. Andersen, *Org. Biomol. Chem.*, 2016, **14**, 1629–1640.
- 30 S. Rasheed, D. D. Subba Rao, S. Chennamsetty, T. Basha and C. Raju, *ChemInform*, 2015, **46**, no-no.
- 31 S. J. I. Shearan, E. Andreoli and M. Taddei, *Beilstein J. Org. Chem.*, 2022, **18**, 1518–1523.
- 32 S. Van der Jeught and C. V. Stevens, *Chem. Rev.*, 2009, **109**, 2672–2702.
- 33 A. Simoens, T. Scattolin, T. Cauwenbergh, G. Pisano, C. S. J. Cazin, C. V. Stevens and S. P. Nolan, *Chem. – Eur. J.*, 2021, **27**, 5653–5657.
- 34 O. Francesconi, M. Martinucci, L. Badii, C. Nativi and S. Roelens, *Chem. – Eur. J.*, 2018, **24**, 6828–6836.
- 35 A. R. Chadeayne, D. N. K. Pham, B. G. Reid, J. A. Golen and D. R. Manke, *ACS Omega*, 2020, **5**, 16940–16943.
- 36 D. J. McKenna, D. B. Repke, L. Lo and S. J. Peroutka, *Neuropharmacology*, 1990, **29**, 193–198.

