



Cite this: *RSC Adv.*, 2023, 13, 24264

Received 4th July 2023

Accepted 31st July 2023

DOI: 10.1039/d3ra04454h

rsc.li/rsc-advances

CuI-catalyzed synthesis of multisubstituted pyrido[1,2-*a*]pyrimidin-4-ones through tandem Ullmann-type C–N cross-coupling and intramolecular amidation reaction†

Baichuan Mo,^a Chunxia Chen^{*ab} and Jinsong Peng^{*a}

Various multi-substituted pyrido[1,2-*a*]pyrimidin-4-ones were synthesized via a one-pot tandem CuI-catalyzed C–N bond formation/intramolecular amidation reaction at 130 °C in DMF. This protocol features simple operation, broad substrate scope, good functional group tolerance and gram scale preparation, thus allowing practical and modular synthesis of pyrido[1,2-*a*]pyrimidin-4-ones from readily available 2-halopyridine and (*Z*)-3-amino-3-arylacrylate ester in good to excellent yields.

1. Introduction

The nitrogen-containing bicyclic heterocycles have attracted considerable attention due to their wide applications in pharmaceuticals, agrochemicals and material sciences.^{1–3} In particular, pyrido[1,2-*a*]pyrimidine scaffolds have emerged as one of the most important N-fused bicyclic heterocycles for drug discovery and development,⁴ and numerous pyrido[1,2-*a*]pyrimidin-4-ones have exhibited a wide range of biological activities such as in antipsychotic agents,⁵ tranquilizers,⁶ antioxidants,⁷ anticancer agents,⁸ antiulcer agents,⁹ antihypertensives,¹⁰ antidepressants,¹¹ antiallergics,¹² antiplasmodial falcipain-2 inhibitors,¹³ and spinal muscular atrophy (SMA) drugs.¹⁴ Several pyrido[1,2-*a*]pyrimidin-4-one derivatives, such as pirenperone, seganserin, lusaperidone, and risdiplam have been applied in clinical trials for decades (Fig. 1).

Given the importance of pyrido[1,2-*a*]pyrimidin-4-ones in drug discovery, much efforts have been focused on the synthetic strategies and methods to construct and diversify these scaffolds efficiently,¹⁵ and the most frequently used approaches are based on acid-catalyzed condensation reaction or thermal cyclization.¹⁶ However, these conventional protocols suffer from some disadvantages such as harsh and corrosive conditions, high reaction temperatures, and limited substrate scope. Therefore, the design and development of more milder and sustainable processes for the synthesis of pyrido[1,2-*a*]pyrimidin-4-ones is imperative. Over the past few years, several transition metal-catalyzed protocols

have been well established for the rapid buildup and modification of pyridopyrimidinone core, enclosing the following approaches (i) Pd-catalyzed regioselective C–H alkenylation at the 3 site,¹⁷ (ii) Pd-catalyzed enolic C–OH activation-arylation pathway,¹⁸ (iii) Pd-catalyzed carbonylative cycloamidation of ketoimines,¹⁹ (iv) Mn-catalyzed carbonylative alkyne annulations of 2-pyridyl hydrazone,²⁰ (v) Ag-catalyzed one-pot cyclization of 2-aminopyridines and alkynoates,²¹ (vi) Pd-catalyzed Ag(I)-promoted C–H arylation at the 3 site with haloarenes as the substrates,²² (vii) Rh-catalyzed three-component coupling of aldehydes, 2-aminopyridines, and diazo esters,²³ (viii) Pd-catalyzed pyridocarbonylation.²⁴

Although significant progress in the palladium-, rhodium- and silver-catalyzed synthetic methodologies have been made, the high cost of these catalysts led to a need to explore the use of more available and cheaper first-row metals for the synthesis of pyrido[1,2-*a*]pyrimidin-4-ones. As an alternative nonprecious metal, copper-mediated catalysis for the synthesis of pyridopyrimidine derivatives has become an important goal.^{25,26}

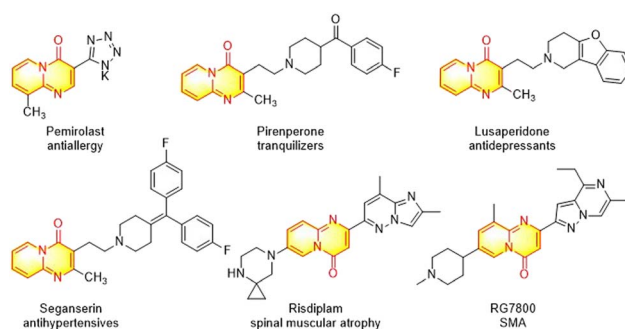


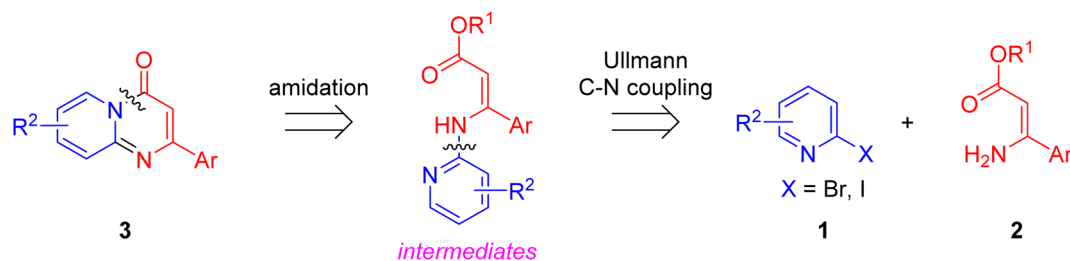
Fig. 1 Selected drugs containing the pyrido[1,2-*a*]pyrimidin-4-one scaffold.

^aCollege of Chemistry, Chemical Engineering and Resource Utilization, Northeast Forestry University, No. 26 Hexing Road, Harbin, 150040, P. R. China. E-mail: ccx1759@163.com; jspeng1998@nefu.edu.cn

^bMaterial Science and Engineering College, Northeast Forestry University, No. 26 Hexing Road, Harbin, 150040, P. R. China

† Electronic supplementary information (ESI) available: Copies of ¹H, ¹³C NMR and HRMS spectra of new compounds. See DOI: <https://doi.org/10.1039/d3ra04454h>





Scheme 1 Retrosynthetic analysis of pyrido[1,2-a]pyrimidin-4-ones.

With our continuous efforts on the Cu-catalyzed synthesis of nitrogen-containing bicyclic heterocycles,²⁷ herein we design an efficient Cu-catalyzed synthesis of pyrido[1,2-a]pyrimidin-4-ones. Retrosynthetically, the formation of substituted pyrido

[1,2-a]pyrimidin-4-ones **3** in a one-pot manner can be perceived through a tandem Cu-catalyzed Ullmann-type aromatic amination and intramolecular amide bond formation (Scheme 1).

Table 1 Optimization of one-pot tandem reaction conditions^a

Entry	Cu/ligand ^b	Base	Solvent	Yield ^c (%)
1	CuO/L1	K ₂ CO ₃	DMA	13
2	Cu(OTf) ₂ /L1	K ₂ CO ₃	DMA	15
3	CuBr ₂ /L1	K ₂ CO ₃	DMA	17
4	CuSO ₄ ·5H ₂ O/L1	K ₂ CO ₃	DMA	9
5	Cu(OAc) ₂ /L1	K ₂ CO ₃	DMA	23
6	CuCl ₂ /L1	K ₂ CO ₃	DMA	21
7	CuCl/L1	K ₂ CO ₃	DMA	12
8	CuBr/L1	K ₂ CO ₃	DMA	14
9	CuI/L1	K ₂ CO ₃	DMA	46
10	CuI/L1	NaHCO ₃	DMA	26
11	CuI/L1	Li ₂ CO ₃	DMA	23
12	CuI/L1	KHCO ₃	DMA	58
13	CuI/L1	K ₃ PO ₄	DMA	28
14	CuI/L1	NaO ^t Bu	DMA	33
15	CuI/L1	KOH	DMA	18
16	CuI/L1	KO ^t Bu	DMA	32
17	CuI/L1	KHCO ₃	DMSO	65
18	CuI/L1	KHCO ₃	DMF	71
19	CuI/L1	KHCO ₃	Toluene	62
20	CuI/L1	KHCO ₃	CH ₃ CN	60
21	CuI/L2	KHCO ₃	DMF	69
22	CuI/L3	KHCO ₃	DMF	61
23	CuI/L4	KHCO ₃	DMF	89
24	CuI/L5	KHCO ₃	DMF	80
25	CuI/L6	KHCO ₃	DMF	78
26	CuI/L7	KHCO ₃	DMF	75
27 ^d	CuI/L4	KHCO ₃	DMF	55 ^e /67 ^f /72 ^g
28 ^h	CuI/L4	KHCO ₃	DMF	58 ⁱ /62 ^j /70 ^k
29 ^l	CuI/L4	KHCO ₃	DMF	36 ^m /68 ⁿ /75 ^o
30 ^p	CuI/L4	KHCO ₃	DMF	53 ^q /60 ^r /67 ^s

^a Reaction conditions: **1a** (0.2 mmol), **2a** (0.24 mmol), base (0.4 mmol), catalyst (20 mol%, 0.04 mmol), ligand (30 mol%, 0.06 mmol), air, 130 °C, 48 h, solvent (1.0 mL). ^b L1 = 2-(dicyclohexylphosphino)biphenyl, L2 = 1,10-phenanthroline, L3 = 2,2'-bipyridine, L4 = Mephos, L5 = Davephos, L6 = triphenylphosphine, L7 = tricyclohexylphosphine. ^c Isolated yield. ^d 48 h. ^e 80 °C. ^f 100 °C. ^g 120 °C. ^h 130 °C. ⁱ 12 h. ^j 24 h. ^k 36 h. ^l L4 = 30 mol%. ^m CuI = 5 mol%. ⁿ CuI = 10 mol%. ^o CuI = 15 mol%. ^p CuI = 20 mol%. ^q L4 = 10 mol%. ^r L4 = 15 mol%. ^s L4 = 20 mol%.

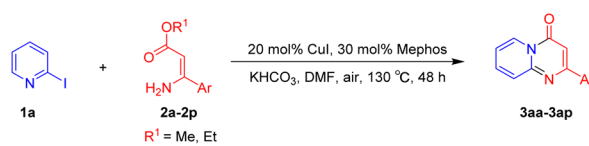
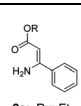
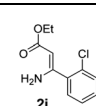
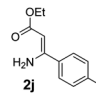
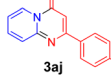
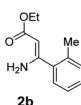
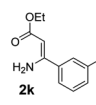
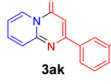
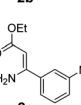
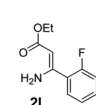
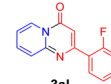
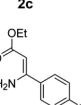
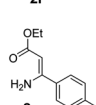
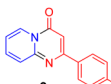
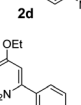
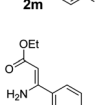
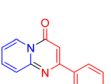
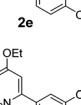
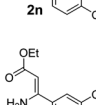
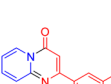
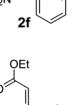
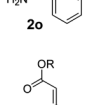
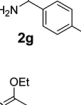


2. Result and discussion

For the initial experiments, 2-iodopyridine **1a** and ethyl (Z)-3-amino-3-phenylacrylate **2a** as model substrates were selected for this copper-catalyzed tandem reaction to construct pyrido[1,2-*a*]pyrimidin-4-one **3aa**. Our previous work revealed that the combined use of copper salts and phosphorus ligands is beneficial to the formation of Ullmann-type C–N bonds.^{27a} Based on the previous result, some new investigations were carried out and reaction optimization results were summarized in Table 1. First, a variety of copper catalysts such as CuO,

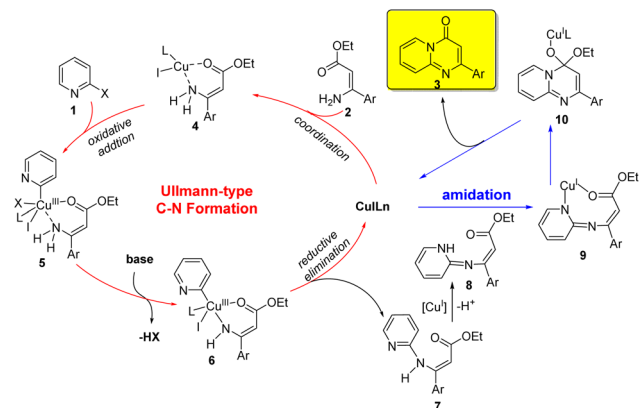
Cu(OTf)₂, CuBr₂, CuSO₄·5H₂O, Cu(OAc)₂, CuCl₂, CuCl, and CuBr was examined with 2-(dicyclohexylphosphino)biphenyl as the ligand in DMA, the desired product **3aa** was obtained from 9% to 23% yield (entries 1–8). However, we were pleased to find that CuI was superior to other copper catalysts and provided **3aa** in 46% yield (entry 9), indicating that the nature of copper sources was essential to the transformation. We continued to optimize the effect of various bases on the reaction, mainly including NaHCO₃, Li₂CO₃, KHCO₃, K₃PO₄, NaO^tBu, KOH and KO^tBu (entries 10–16), potassium bicarbonate is the most suitable base for this transformation, giving **3aa** in 58% yield

Table 2 Variation of the enamine substrates^a

							
Entry	S-2	P-3	Yield ^b	Entry	S-2	P-3	Yield ^b
1			89	10			53
2	2a': R = Me		85	11			77
3			75	12			71
4			82	13			29
5			84	14			67
6			60	15			70
7			62	16			57
8			58	17			70
9			49	18	2p': R = Me		57

^a Reaction conditions: **2** (0.24 mmol), **1a** (0.2 mmol), CuI (20 mol%), Mephos (30 mol%), KHCO₃ (0.4 mmol), DMF (1.0 mL), air, 130 °C, 48 h. ^b Yield of the isolated product.





Scheme 2 Proposed mechanism for the formation of pyrido[1,2-*a*]pyrimidinones.

be converted to the desired products **3ba** in excellent yield, however, the use of bromo or chloro-substituted substrates (**1b'** and **1b''**, entry 1) afforded inferior results than their iodo analogue, providing 64% and 31% yield respectively. The yield for 2-halopyridines follows the order pyridinyl iodide > bromide > chloride. The result was probably attributed to poorer tendency of C–Br and C–Cl to undergo oxidative addition to active copper species. Various 2-halopyridines with substituents such as Me, OMe and Cl on the pyridine moiety were next explored, and the corresponding products **3ba–3ia** can be obtained in 31–64% yields. The electronic nature of these substituents seemed to have little effect on the reaction outcome, the similar yields were obtained for the selected 2-bromopyridine substrates except **1f** with methoxy at the meta position (entries 2–7). The incorporation of the sterically hindering methyl or methoxy group in the *ortho* positions of halogen seemed not to affect the reaction, and the corresponding products can be obtained in the same yield (51%, entries 2 and 6). To our delight, the reaction conditions was also suitable for 2-iodoquinoline, which could smoothly convert into the corresponding product (entry 9). Finally, the reaction can be carried out on a 1.0 mmol scale to provide the target product **3ba** in 73% yield (entry 1), demonstrating its utility in organic synthesis.

Based on the above experimental results, a plausible mechanism for the copper-catalyzed sequential Ullmann-type C–N formation and amidation reaction was outlined in Scheme 2. The initial step involved the coordination of ester (*Z*)-3-amino-3-arylacrylate **2** to the Cu(I) species to give chelate **4**, which underwent an intermolecular oxidative addition of 2-halopyridine **1** to form a Cu(III) species **5**. A base-promoted deprotonation of amino group led to form a Cu–N bond and provide a complex **6**. The resulting complex **6** underwent reductive elimination to give an intermediate **7** and regenerate the active Cu(I) catalyst (Ullmann-type C–N coupling process). Subsequent, **7** was deprotonated to form intermediate **8**, which could afford the product **3** through copper-catalyzed intramolecular amide bond formation. It's noteworthy that CuI played two critical roles in this reaction, which could not only catalyze the intermolecular C–N formation

through Ullmann reaction, but also promote the subsequent intramolecular cyclization *via* amide bond formation.²⁶

3. Conclusions

In summary, a novel and efficient approach to construct biologically relevant pyrido[1,2-*a*]pyrimidin-4-ones have been developed through Cu-catalyzed sequential Ullmann-type C–N formation and intramolecular amidation starting from 2-halopyridines and ester (*Z*)-3-amino-3-arylacrylates. The reaction has some advantages of broad functional group compatibility, facile scalability and easy product derivatization. At the same time, considering their potential biological activities, the method could be further employed to construct more complex biological molecules in synthetic and pharmaceutical chemistry.

4. Experimental section

4.1. General information

Chemicals were all purchased from commercial supplies and used without further purification unless otherwise stated. Solvents were dried and purified according to the standard procedures before use. Reactions were monitored by analytical thin-layer chromatography (TLC). All reactions were conducted in dried glassware. Reaction products was purified by flash chromatography with 230–400 mesh silica gel. Ester (*Z*)-3-aryl-3-aminoacrylate substrates were prepared according to the literature methods.²⁸ Melting points were determined on a melting point apparatus in open capillaries and were uncorrected. Infrared spectra of samples were recorded from 4000 to 500 cm^{−1} in ATR (attenuated total reflectance) mode using an FT-IR instrument. ¹H NMR spectra were recorded on a 400 or 500 MHz spectrometer. ¹³C NMR spectra were recorded at 126 or 151 MHz. Unless otherwise stated, deuteriochloroform (CDCl₃) was used as a solvent. Chemical shifts (δ) are given in parts per million downfield relative to tetramethylsilane (TMS). Chemical shifts for carbon resonances are reported in parts per million and are referenced to the carbon resonance of the solvent CHCl₃ (δ = 77.16 ppm). The splitting patterns are reported as s (singlet), d (doublet), dd (double doublet), td (triplet of doublet), t (triplet), q (quartet), br (broad), and m (multiplet). Coupling constants are given in hertz. High-resolution mass spectra were recorded on a BIO TOF Q mass spectrometer equipped with an electrospray ion source (ESI), operated in the positive mode.

4.2. General procedures for the synthesis of pyridopyrimidinone derivatives

A 10 mL schlenk tube or standard vial equipped with a magnetic stirring bar was charged with 2-halopyridine (0.2 mmol, 1.0 equiv.), enamines (0.24 mmol, 1.2 equiv.), and KHCO₃ (40 mg, 0.4 mmol, 2.0 equiv.), and then CuI (7.6 mg, 0.04 mmol, 0.2 equiv.) and Mephos (21.8 mg, 0.06 mmol, 0.3 equiv.) were added. Finally, DMF (1.0 mL) was added to the mixture *via* syringe at room temperature under air. The tube was sealed and



put into a preheated oil bath at 130 °C for 48 h. The mixture was cooled to room temperature, quenched with water (3 mL), and diluted with ethyl acetate (5 mL). The layers were separated, and the aqueous layer was extracted with 3 × 5 mL of ethyl acetate. The combined organic extracts were dried over anhydrous sodium sulfate, filtered, and concentrated *in vacuo*. The crude product was then purified by flash chromatography on silica gel (H), eluting with the mixture of ethyl acetate and petroleum ether (from 1 : 4 to 1 : 3).

4.2.1 2-Phenyl-4H-pyrido[1,2-*a*]pyrimidin-4-one (3aa).¹⁸ Yield, 89% (39.5 mg); white solid, mp 155–156 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.07 (d, *J* = 7.1 Hz, 1H), 8.09 (dd, *J* = 6.5, 3.1 Hz, 2H), 7.73 (d, *J* = 3.6 Hz, 2H), 7.52–7.47 (m, 3H), 7.12 (dt, *J* = 7.5, 3.9 Hz, 1H), 6.91 (s, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 162.09, 158.66, 151.05, 137.29, 136.20, 130.67, 128.85, 127.44, 127.30, 126.81, 115.23, 100.14.

4.2.2 2-(*o*-Tolyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3ab).¹⁹ Yield, 75% (35.4 mg); white solid, mp 125–126 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.13 (d, *J* = 7.1 Hz, 1H), 7.79–7.70 (m, 2H), 7.49 (d, *J* = 7.4 Hz, 1H), 7.34 (dt, *J* = 14.4, 6.6 Hz, 3H), 7.18 (t, *J* = 6.6 Hz, 1H), 6.59 (s, 1H), 2.47 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 165.56, 158.14, 150.74, 138.68, 136.27, 135.85, 131.04, 129.27, 129.10, 127.25, 126.66, 126.07, 115.46, 104.40, 20.38.

4.2.3 2-(*m*-Tolyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3ac).¹⁹ Yield, 82% (38.7 mg); light yellow solid, mp 120–121 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.04 (d, *J* = 7.1 Hz, 1H), 7.91 (s, 1H), 7.84 (d, *J* = 7.7 Hz, 1H), 7.71 (s, 1H), 7.37 (t, *J* = 7.6 Hz, 1H), 7.28 (d, *J* = 7.5 Hz, 1H), 7.14–7.08 (m, 2H), 6.89 (s, 1H), 2.44 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 162.23, 158.61, 150.99, 138.51, 137.21, 136.14, 131.44, 128.73, 128.06, 127.25, 126.73, 124.57, 115.17, 100.12, 21.57.

4.2.4 2-(*p*-Tolyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3ad).¹⁹ Yield, 84% (39.6 mg); white solid, mp 167–168 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.05 (d, *J* = 7.1 Hz, 1H), 7.99 (d, *J* = 7.7 Hz, 2H), 7.71 (s, 2H), 7.30 (d, *J* = 7.8 Hz, 2H), 7.10 (s, 1H), 6.89 (s, 1H), 2.42 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 162.02, 158.68, 150.99, 141.06, 136.08, 134.40, 129.57, 127.36, 127.27, 126.73, 115.07, 99.60, 21.47.

4.2.5 2-(4-Methoxyphenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3ae).¹⁹ Yield, 60% (30.2 mg); white solid, mp 174–176 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.04 (d, *J* = 7.1 Hz, 1H), 8.07 (d, *J* = 8.8 Hz, 2H), 7.70 (d, *J* = 6.0 Hz, 2H), 7.08 (td, *J* = 6.9, 6.1, 2.2 Hz, 1H), 7.00 (d, *J* = 8.8 Hz, 2H), 6.85 (s, 1H), 3.87 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 161.84, 161.57, 158.65, 150.96, 136.07, 129.57, 129.01, 127.28, 126.62, 114.93, 114.18, 98.84, 55.44.

4.2.6 2-(3-Methoxyphenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3af).⁸ Yield, 62% (31.2 mg); white solid, mp 157–158 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.05 (d, *J* = 7.0 Hz, 1H), 7.72 (s, 2H), 7.63 (d, *J* = 9.8 Hz, 2H), 7.39 (t, *J* = 7.9 Hz, 1H), 7.14–7.08 (m, 1H), 7.02 (d, *J* = 8.1 Hz, 1H), 6.89 (s, 1H), 3.89 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 161.83, 159.98, 158.61, 150.95, 138.71, 136.20, 129.84, 127.25, 126.78, 119.81, 116.60, 115.26, 112.55, 100.25, 55.42.

4.2.7 2-(4-Chlorophenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3ag).¹⁹ Yield, 58% (29.7 mg); white solid, mp 154–155 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.06 (d, *J* = 7.1 Hz, 1H), 8.04 (d, *J* = 7.9 Hz, 2H), 7.78–7.69 (m, 2H), 7.47 (d, *J* = 7.9 Hz, 2H), 7.14 (t, *J* = 6.6 Hz, 1H), 6.87 (s, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 160.76, 158.56, 151.07, 136.91, 136.41, 135.67, 129.05, 128.74, 127.33, 126.75, 115.38, 99.88.

4.2.8 2-(3-Chlorophenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3ah).¹⁹ Yield, 49% (25.1 mg); white solid, mp 165–166 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.05 (d, *J* = 7.1 Hz, 1H), 8.11 (s, 1H), 7.91 (d, *J* = 7.2 Hz, 1H), 7.74 (q, *J* = 8.8 Hz, 2H), 7.42 (q, *J* = 8.3, 7.8 Hz, 2H), 7.14 (t, *J* = 6.6 Hz, 1H), 6.86 (s, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 160.44, 158.49, 151.04, 139.06, 136.47, 134.95, 130.56, 130.03, 127.64, 127.30, 126.78, 125.41, 115.48, 100.22.

4.2.9 2-(2-Chlorophenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3ai).¹⁹ Yield, 53% (27.1 mg); white solid, mp 187–188 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.12 (d, *J* = 7.1 Hz, 1H), 7.80–7.72 (m, 2H), 7.66–7.61 (m, 1H), 7.53–7.47 (m, 1H), 7.41–7.35 (m, 2H), 7.19 (t, *J* = 6.6 Hz, 1H), 6.78 (s, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 161.14, 156.92, 150.01, 136.61, 135.34, 131.15, 129.77, 129.44, 129.42, 126.32, 126.03, 125.72, 114.63, 104.27.

4.2.10 2-(4-Fluorophenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3aj).¹⁹ Yield, 77% (36.9 mg); white solid, mp 214–215 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.07 (d, *J* = 7.1 Hz, 1H), 8.13–8.07 (m, 2H), 7.74 (q, *J* = 8.5 Hz, 2H), 7.16 (dt, *J* = 15.0, 7.4 Hz, 3H), 6.86 (s, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 164.50 (d, ¹*J*_{C-F} = 251.2 Hz), 160.95, 158.58, 151.05, 136.36, 133.36, 129.51 (d, ³*J*_{C-F} = 8.7 Hz), 127.32, 126.71, 115.86 (d, ²*J*_{C-F} = 21.7 Hz), 115.29, 99.69.

4.2.11 2-(3-Fluorophenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3ak). Yield, 71% (34 mg); white solid, mp 159–160 °C; IR (KBr, cm⁻¹): 3434, 1707, 1638, 1526, 1493, 1464, 1446, 1231, 779, 756; ¹H NMR (500 MHz, CDCl₃) δ 9.07 (d, *J* = 7.1 Hz, 1H), 7.86–7.82 (m, 2H), 7.79–7.73 (m, 2H), 7.46 (td, *J* = 8.1, 5.9 Hz, 1H), 7.21–7.13 (m, 2H), 6.88 (s, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 162.13 (d, ¹*J*_{C-F} = 246.1 Hz), 159.53 (d, ⁴*J*_{C-F} = 2.6 Hz), 157.50, 150.01, 138.56 (d, ³*J*_{C-F} = 7.5 Hz), 135.41, 129.31 (d, ³*J*_{C-F} = 8.1 Hz), 126.29, 125.76, 121.92 (d, ⁴*J*_{C-F} = 2.8 Hz), 116.46 (d, ²*J*_{C-F} = 21.4 Hz), 114.43, 113.44 (d, ²*J*_{C-F} = 23.1 Hz), 99.25. HRMS (ESI) *m/z* calcd for C₁₄H₁₀FN₂O⁺ (*M* + *H*)⁺ 241.07717, found 241.07657.

4.2.12 2-(2-Fluorophenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3al). Yield, 29% (13.9 mg); white solid, mp 151–152 °C; IR (KBr, cm⁻¹): 3434, 1672, 1533, 1475, 1217, 1138, 841, 755; ¹H NMR (500 MHz, CDCl₃) δ 9.00 (d, *J* = 7.1 Hz, 1H), 8.07–8.01 (m, 1H), 7.70–7.64 (m, 2H), 7.40–7.35 (m, 1H), 7.24–7.20 (m, 1H), 7.14–7.06 (m, 2H), 6.96 (d, *J* = 1.1 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 159.97 (d, ¹*J*_{C-F} = 253.0 Hz), 157.29, 156.98 (d, ⁴*J*_{C-F} = 2.2 Hz), 149.86, 135.13, 130.83 (d, ³*J*_{C-F} = 8.8 Hz), 129.87 (d, ⁴*J*_{C-F} = 2.2 Hz), 126.22, 125.68, 124.57 (d, ²*J*_{C-F} = 10.7 Hz), 123.47 (d, ⁴*J*_{C-F} = 3.6 Hz), 115.52 (d, ²*J*_{C-F} = 22.9 Hz), 114.33, 103.74 (d, ³*J*_{C-F} = 10.8 Hz). HRMS (ESI) *m/z* calcd for C₁₄H₁₀FN₂O⁺ (*M* + *H*)⁺ 241.07717, found 241.07717.

4.2.13 2-(4-Bromophenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3am).^{19f,20} Yield, 67% (40.2 mg); white solid, mp 175–176 °C; ¹H NMR (400 MHz, CDCl₃) δ 9.06 (d, *J* = 7.1 Hz, 1H), 7.97 (d, *J* = 8.5 Hz, 1H), 7.83 (s, 2H), 7.78–7.70 (m, 2H), 7.63 (d, *J* = 8.5 Hz, 1H), 7.14 (t, *J* = 6.3 Hz, 1H), 6.87 (s, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 160.98, 158.56, 151.01, 138.02, 136.75, 136.43, 136.14, 132.03, 129.05, 127.34, 126.77, 125.38, 115.40, 99.88.

4.2.14 2-(4-(Trifluoromethyl)phenyl)-4H-pyrido[1,2-*a*]pyrimidin-4-one (3an).^{19f,20} Yield, 70% (40.6 mg); light yellow

solid, mp 172–173 °C; ^1H NMR (400 MHz, CDCl_3) δ 9.07 (d, J = 7.1 Hz, 1H), 8.19 (d, J = 8.0 Hz, 2H), 7.75 (t, J = 9.0 Hz, 4H), 7.16 (t, J = 6.6 Hz, 1H), 6.92 (s, 1H). ^{13}C NMR (151 MHz, CDCl_3) δ 159.4, 157.5, 150.1, 139.6, 135.5, 131.2 (q, $^2J_{\text{C-F}}$ = 31.7 Hz, 131.5, 131.3, 131.1, 130.9), 126.7, 126.3, 125.8, 124.7 (q, $^3J_{\text{C-F}}$ = 3.0 Hz, 124.71, 124.69, 124.67, 124.65), 122.9 (q, $^1J_{\text{C-F}}$ = 271.8 Hz, 125.6, 123.8, 122.0, 120.2), 114.6, 99.6.

4.2.15 2-(3-(Trifluoromethyl)phenyl)-4H-pyrido[1,2-a]pyrimidin-4-one (3ao). Yield, 57% (33.1 mg); light yellow solid, mp 150–152 °C; IR (KBr, cm^{-1}): 3434, 1704, 1690, 1636, 1534, 1498, 1457, 1434, 1409, 1319, 1231, 1182, 1168, 1128, 1095, 1080, 802, 765, 689; ^1H NMR (400 MHz, CDCl_3) δ 9.07 (d, J = 7.1 Hz, 1H), 8.41 (s, 1H), 8.22 (d, J = 7.8 Hz, 1H), 7.80–7.71 (m, 3H), 7.61 (t, J = 7.8 Hz, 1H), 7.16 (ddd, J = 7.7, 6.1, 2.1 Hz, 1H), 6.92 (s, 1H). ^{13}C NMR (151 MHz, CDCl_3) δ 159.3, 157.5, 150.1, 137.0, 135.5, 130.3 (q, $^2J_{\text{C-F}}$ = 31.7 Hz, 130.6, 130.4, 130.2, 129.9), 129.4, 128.3, 126.3, 126.1 (q, $^3J_{\text{C-F}}$ = 3.0 Hz, 126.14, 126.12, 126.10, 126.07), 125.8, 123.4 (q, $^3J_{\text{C-F}}$ = 3.0 Hz, 123.43, 123.41, 123.38, 123.36), 123.0 (q, $^1J_{\text{C-F}}$ = 273.3 Hz, 125.7, 123.9, 122.1, 120.3), 114.6, 99.2. HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{10}\text{F}_3\text{N}_2\text{O}^+$ ($\text{M} + \text{H}$) $^+$ 291.07397, found 291.07373.

4.2.16 2-Methyl-4H-pyrido[1,2-a]pyrimidin-4-one (3ap).^{16d} Yield, 70% (22.4 mg); light yellow solid, mp 118–120 °C; ^1H NMR (400 MHz, CDCl_3) δ 9.02 (d, J = 7.1 Hz, 1H), 7.72 (t, J = 7.7 Hz, 1H), 7.60 (s, 1H), 7.10 (t, J = 6.9 Hz, 1H), 6.34 (s, 1H), 2.46 (s, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 165.32, 157.89, 150.74, 136.26, 127.27, 125.84, 115.03, 103.37, 24.72.

4.2.17 2-Phenyl-4H-pyrido[1,2-a]pyrimidin-4-one (3ba).^{19,20} Yield, 64% (28.4 mg); white solid, mp 155–156 °C; ^1H NMR (400 MHz, CDCl_3) δ 9.08 (d, J = 7.0 Hz, 1H), 8.12–8.07 (m, 2H), 7.74 (s, 2H), 7.50 (s, 3H), 7.13 (s, 1H), 6.92 (s, 1H). ^{13}C NMR (151 MHz, CDCl_3) δ 161.07, 157.63, 150.02, 136.25, 135.15, 129.63, 127.81, 126.40, 126.26, 125.76, 114.19, 99.10.

4.2.18 9-Methyl-2-phenyl-4H-pyrido[1,2-a]pyrimidin-4-one (3ca).²⁰ Yield, 51% (24 mg); light yellow solid, mp 176–177 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.94 (s, 1H), 8.18–8.11 (m, 2H), 7.57 (d, J = 6.6 Hz, 1H), 7.49 (d, J = 3.7 Hz, 3H), 7.01 (t, J = 6.9 Hz, 1H), 6.93 (s, 1H), 2.67 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 159.48, 158.18, 149.39, 136.30, 134.29, 133.80, 129.56, 127.69, 126.37, 124.22, 113.67, 98.48, 17.27.

4.2.19 8-Methyl-2-phenyl-4H-pyrido[1,2-a]pyrimidin-4-one (3da).²⁰ Yield, 51% (24 mg); light yellow solid, mp 167–168 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.95 (d, J = 7.2 Hz, 1H), 8.08–8.05 (m, 2H), 7.52–7.47 (m, 4H), 6.95 (dd, J = 7.2, 1.6 Hz, 1H), 6.84 (s, 1H), 2.48 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 161.26, 157.66, 150.00, 147.24, 136.45, 129.49, 127.74, 126.35, 125.52, 123.81, 116.88, 98.21, 20.40.

4.2.20 7-Methyl-2-phenyl-4H-pyrido[1,2-a]pyrimidin-4-one (3ea).^{19,20} Yield, 50% (23.6 mg); light yellow solid, mp 167–168 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.86 (s, 1H), 8.07 (d, J = 2.4 Hz, 2H), 7.64 (d, J = 9.0 Hz, 1H), 7.58 (d, J = 10.2 Hz, 1H), 7.50–7.45 (m, 3H), 6.88 (s, 1H), 2.41 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 160.57, 157.49, 148.92, 138.08, 136.33, 129.46, 127.75, 126.30, 125.16, 124.49, 123.66, 98.79, 17.32.

4.2.21 8-Methoxy-2-phenyl-4H-pyrido[1,2-a]pyrimidin-4-one (3fa). Yield, 25% (12.6 mg); white solid, mp 157–158 °C; IR (KBr, cm^{-1}): 3438, 1673, 1645, 1500, 1470, 1457, 1403, 1354, 1223, 1162, 1019, 837, 779; ^1H NMR (500 MHz, CDCl_3) δ 8.90 (d,

J = 7.8 Hz, 1H), 8.02 (dd, J = 6.5, 3.1 Hz, 2H), 7.46 (dd, J = 5.0, 1.8 Hz, 3H), 6.92 (d, J = 2.7 Hz, 1H), 6.77–6.69 (m, 2H), 3.93 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 164.23, 161.61, 157.62, 152.07, 136.49, 129.44, 127.71, 127.32, 126.26, 109.27, 101.31, 96.98, 55.35. HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{13}\text{N}_2\text{O}_2^+$ ($\text{M} + \text{H}$) $^+$ 253.09715, found 253.09712.

4.2.22 9-Methoxy-2-phenyl-4H-pyrido[1,2-a]pyrimidin-4-one (3ga). Yield, 51% (25.7 mg); light yellow solid, mp 169–170 °C; IR (KBr, cm^{-1}): 3435, 1677, 1634, 1496, 1472, 1415, 1277, 1162, 1042, 753; ^1H NMR (500 MHz, CDCl_3) δ 8.73–8.68 (m, 1H), 8.13–8.08 (m, 2H), 7.48 (d, J = 6.6 Hz, 3H), 7.07–7.00 (m, 2H), 6.94 (s, 1H), 4.06 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 159.97, 157.70, 151.47, 144.38, 136.33, 129.54, 127.78, 126.49, 118.06, 113.17, 110.08, 99.93, 55.82. HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{13}\text{N}_2\text{O}_2^+$ ($\text{M} + \text{H}$) $^+$ 253.09715, found 253.09732.

4.2.23 7-Methoxy-2-phenyl-4H-pyrido[1,2-a]pyrimidin-4-one (3ha). Yield, 43% (21.6 mg); white solid, mp 167–168 °C; IR (KBr, cm^{-1}): 3435, 1678, 1637, 1533, 1504, 1458, 1255, 1016, 822; ^1H NMR (500 MHz, CDCl_3) δ 8.57 (d, J = 2.8 Hz, 1H), 8.09–8.06 (m, 2H), 7.69 (d, J = 9.6 Hz, 1H), 7.53 (dd, J = 9.6, 2.9 Hz, 1H), 7.49 (ddt, J = 6.8, 5.3, 2.6 Hz, 3H), 6.92 (s, 1H), 3.96 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 159.86, 157.52, 149.72, 147.12, 136.33, 130.57, 129.41, 127.79, 126.46, 126.26, 105.74, 98.49, 55.46. HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{13}\text{N}_2\text{O}_2^+$ ($\text{M} + \text{H}$) $^+$ 253.09715, found 253.09709.

4.2.24 7-Chloro-2-phenyl-4H-pyrido[1,2-a]pyrimidin-4-one (3ia).¹⁹ Yield, 62% (31.8 mg); white solid, mp 169–170 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.99 (s, 1H), 8.01–7.96 (m, 2H), 7.59 (s, 2H), 7.45–7.41 (m, 3H), 6.85 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 160.86, 156.62, 148.32, 136.40, 135.75, 129.86, 127.85, 126.68, 126.37, 124.05, 122.86, 99.44.

4.2.25 3-Phenyl-1H-pyrimido[1,2-a]quinolin-1-one (3ja).²⁹ Yield, 48% (26.1 mg); white solid, mp 159–161 °C; ^1H NMR (500 MHz, CDCl_3) δ 9.85 (d, J = 8.8 Hz, 1H), 8.05–8.01 (m, 2H), 7.71 (d, J = 9.1 Hz, 1H), 7.64–7.56 (m, 2H), 7.50–7.41 (m, 4H), 7.36 (dd, J = 10.0, 3.6 Hz, 1H), 6.92–6.90 (m, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 162.99, 157.63, 150.33, 135.42, 135.19, 134.44, 129.61, 128.68, 127.79, 127.07, 126.19, 125.86, 124.00, 123.91, 121.30, 104.62.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We are grateful for the Natural Science Foundation of Heilongjiang Province (ZD2021C001), financial support from the Fundamental Research Funds for the Central Universities (2572020AW45) and the Programme of Introducing Talents of Discipline to Universities (111 Project, B20088).

Notes and references

- (a) A. K. Bagdi, S. Santra, K. Monira and A. Hajra, *Chem. Commun.*, 2015, **51**, 1555; (b) J. Panda, B. P. Raiguru, M. Mishra, S. Mohapatra and S. Nayak, *ChemistrySelect*,



- 2022, 7, e202103987; (c) A. Manaithiya, O. Alam, V. Sharma, M. J. Naim, S. Mittal, F. Azam, A. Husain, A. A. Sheikh, M. Imran and I. A. Khan, *Curr. Top. Med. Chem.*, 2021, **21**, 2292; (d) E. Vitaku, D. T. Smith and J. T. Njardarson, *J. Med. Chem.*, 2014, **57**, 10257; (e) I. Khan, A. Ibrar, W. Ahmed and A. Saeed, *Eur. J. Med. Chem.*, 2015, **90**, 124.
- 2 R. R. Mittapalli, S. J. Coles, W. T. Klooster and A. P. Dobbs, *J. Org. Chem.*, 2021, **86**, 2076.
- 3 J. Wan, C. Zheng, M. Fung, X. Liu, C. Lee and X. Zhang, *J. Mater. Chem.*, 2012, **22**, 4502.
- 4 I. Hermecz and Z. Mészáros, *Med. Res. Rev.*, 1988, **8**, 203.
- 5 C. Fenton and L. J. Scott, *CNS Drugs*, 2005, **19**, 429.
- 6 R. L. Smith, R. J. Barrett and E. Sanders-Bush, *J. Pharmacol. Exp. Ther.*, 1995, **275**, 1050.
- 7 C. L. Motta, S. Sartini, L. Mugnaini, F. Simorini, S. Taliani, S. Salerno, A. M. Marini, F. D. Settimo, A. Lavecchia, E. Novellino, M. Cantore, P. Failli and M. Ciuffi, *J. Med. Chem.*, 2007, **50**, 4917.
- 8 G. Priyadarshani, S. Amrutkar, A. Nayak, U. C. Banerjee, C. N. Kundu and S. K. Guchhait, *Eur. J. Med. Chem.*, 2016, **122**, 43.
- 9 G. C. B. Harriman, S. Chi, M. Zhang, A. Crowe, R. A. Bennett and I. Parsons, *Tetrahedron Lett.*, 2003, **44**, 3659.
- 10 A. Pettersson, K. Gradin, T. Hedner and B. Persson, *Naunyn-Schmiedeberg's Arch. Pharmacol.*, 1985, **329**, 394.
- 11 L. E. J. Kennis, F. P. Bischoff, C. J. Mertens, C. J. Love, F. A. F. Van den Keybus, S. Pieters, M. Braeken, A. A. H. P. Megens and J. E. Leysen, *Bioorg. Med. Chem. Lett.*, 2000, **10**, 71.
- 12 (a) Y. Yanagihara, H. Kasai, T. Kawashima and T. Shida, *Jpn. J. Pharmacol.*, 1988, **48**, 91; (b) A. Sano, M. Ishihara, J. Yoshihara, M. Sumino and H. Nawa, *Chem. Pharm. Bull.*, 1995, **43**, 683.
- 13 U. R. Mane, H. Li, J. Huang, R. C. Gupta, S. S. Nadkarni, R. Giridhar, P. P. Naik and M. R. Yadav, *Bioorg. Med. Chem.*, 2012, **20**, 6296.
- 14 (a) H. Ratni, R. S. Scalco and A. H. Stephan, *ACS Med. Chem. Lett.*, 2021, **12**, 874; (b) H. Ratni, M. Ebeling, J. Baird, S. Bendels, J. Bylund, K. S. Chen, N. Denk, Z. Feng, L. Green, M. Guerard, P. Jablonski, B. Jacobsen, O. Khwaja, H. Kletzl, C.-P. Ko, S. Kustermann, A. Marquet, F. Metzger, B. Mueller, N. A. Naryshkin, S. V. Paushkin, E. Pinard, A. Poirier, M. Reutlinger, M. Weetall, A. Zeller, X. Zhao and L. Mueller, *J. Med. Chem.*, 2018, **61**, 6501.
- 15 (a) K. M. Elattar, R. Rabie and M. M. Hammouda, *Monatsh. Chem.*, 2017, **148**, 601; (b) R. T. Bhawale, A. S. Chillal and U. A. Kshirsagar, *J. Heterocycl. Chem.*, 2023, **60**, 1356.
- 16 (a) W. Liu, S. Wang, H. Zhan and M. Li, *Synlett*, 2014, **25**, 1478; (b) A. Molnár, F. Faigl, B. Podányi, Z. Finta, L. Balázs and I. Hermecz, *Heterocycles*, 2009, **78**, 2477; (c) M. Shur and S. S. Israelstam, *J. Org. Chem.*, 1968, **33**, 3015; (d) S. Mathavan, A. K. D. Raj and R. B. R. D. Yamajala, *ChemistrySelect*, 2019, **4**, 10737; (e) S. N. Basahel, N. S. Ahmed, K. Narasimharao and M. Mokhtar, *RSC Adv.*, 2016, **6**, 11921; (f) H. G. Bonacorso, F. J. Righi, I. R. Rodrigues, C. A. Cechinel, M. B. Costa, A. D. Wastowski, M. A. P. Martins and N. Zanatta, *J. Heterocycl. Chem.*, 2006, **43**, 229; (g) M. Hussaina and J. Liu, *Tetrahedron Lett.*, 2020, **61**, 152269.
- 17 (a) W. Liu, S. Wang, Q. Zhang, J. Yu, J. Li, Z. Xie and H. Cao, *Chem.-Asian J.*, 2014, **9**, 2436; (b) J. Lv, Y. Liang, P. He, Z. Cai, J. Liu and F. Huang, *RSC Adv.*, 2015, **5**, 36171.
- 18 S. K. Guchhait and G. Priyadarshani, *J. Org. Chem.*, 2015, **80**, 6342.
- 19 Y. Xie, T. Chen, S. Fu, H. Jiang and W. Zeng, *Chem. Commun.*, 2015, **51**, 9377.
- 20 Y.-F. Liang, R. Steinbock, A. Münch, D. Stalke and L. Ackermann, *Angew. Chem., Int. Ed.*, 2018, **57**, 5384.
- 21 Z. Chen, Y. Wen, H. Ding, G. Luo, M. Ye, L. Liu and J. Xue, *Tetrahedron Lett.*, 2017, **58**, 13.
- 22 S. K. Guchhait and G. Priyadarshani, *J. Org. Chem.*, 2015, **80**, 8482.
- 23 G. L. Hoang, A. J. Zoll and J. A. Ellman, *Org. Lett.*, 2019, **21**, 3886.
- 24 (a) D. Liang, Y. He and Q. Zhu, *Org. Lett.*, 2014, **16**, 2748; (b) G. Yan and J. E. Golden, *Org. Lett.*, 2018, **20**, 4393.
- 25 K. Tatsumi, T. Fujihara, J. Terao and Y. Tsuji, *Adv. Synth. Catal.*, 2018, **360**, 3245.
- 26 Y. Yang, W.-M. Shu, S.-B. Yu, F. Ni, M. Gao and A.-X. Wu, *Chem. Commun.*, 2013, **49**, 1729.
- 27 (a) Y. Li, J. Peng, X. Chen, B. Mo, X. Li, P. Sun and C. Chen, *J. Org. Chem.*, 2018, **83**, 5288; (b) X. Chen, Y. Bian, B. Mo, P. Sun, C. Chen and J. Peng, *RSC Adv.*, 2020, **10**, 24830; (c) P. Sun, J. Yang, Z. Song, Y. Cai, Y. Liu, C. Chen, X. Chen and J. Peng, *Synthesis*, 2020, **52**, 75.
- 28 G. Hou, W. Li, M. Ma, X. Zhang and X. Zhang, *J. Am. Chem. Soc.*, 2010, **132**, 12844.
- 29 X. Huang and F.-C. Ye, *Chin. J. Org. Chem.*, 1993, **13**, 518.

