


 Cite this: *RSC Adv.*, 2017, 7, 38463

Received 21st June 2017

Accepted 28th July 2017

DOI: 10.1039/c7ra06946d

rsc.li/rsc-advances

Dual modal imaging agents based on chromophore-bearing DTPA analogues†

 CuhaWijay Sathiyajith,^{id} Andrew J. Hallett, Angelo J. Amoroso* and Peter G. Edwards*

Two new DTPA analogues, centrally (L^1) and terminally (L^2) functionalised with a 1,8-naphthalimide chromophore, have been successfully prepared and fully characterized. Their Gd(III) complexes have also been prepared and evaluated for their ability to act as dual modal contrast agents (MRI/OI). The highly reproducible R_1 relaxivity of L^1 ($8.10 \pm 0.21 \text{ mM}^{-1} \text{ s}^{-1}$, 25 °C and 30 MHz) is markedly higher than other DTPA based contrast agents. The Gd(III) complexes of both L^1 and L^2 have been evaluated as luminescence probes; the ligand based fluorescence is not quenched upon complexation.

Introduction

Diethylenetriaminepentaacetic acid (DTPA) is one of the most well-known ligands and yields highly stable complexes with hard metal ions. DTPA and its derivatives have been used in a vast array of applications of medical interest, such as contrast agents for medical imaging,^{1–5} radiotherapy, radiolabeling of antibodies and peptides,^{6–9} radiopharmaceuticals and chelation therapy.^{10–12} In particular, the gadolinium complex of DTPA has long been successful as a contrast agent, ever since it was approved by the FDA in 1988.¹³

However, the perceived disadvantage of Gd-DTPA is that the chelate has been prepared in hypertonic solutions, where issues related to high osmolarity become significant. The investigations have been intensified on the synthesis and characteristics of novel Gd(III) complexes to minimize toxicity issues while ensuring relaxation rate enhancement. The biocompatibility of the multidentate DTPA derivatives makes it an ideal choice. DTPA also offers the advantage of incorporating a well-defined structural molecule, to a chosen biological molecule (which could be proteins in the blood, antibodies or similar biological macromolecules) due to their multidentate feature.

Several strategies have been utilised in the synthesis of substituted DTPA frameworks. These approaches include (i) the attachment of substituents *via* an amide linkage to a carboxylate group of the preformed DTPA ligand or (ii) the direct bonding of the substituent to the diethyltriamine core.

The attachment of substituents to DTPA *via* an amide link is a useful strategy as it is a simple and convenient method that can give compounds in high yields and good purity. There are

numerous reported methods to carry this reaction out.^{14–25} Some methods involve the use of coupling reagents to couple the amine containing substituent to the DTPA,²⁶ though the simplest method involves the conversion of DTPA to the bisanhydride analogue, which may then be directly reacted with a suitable amine.^{27–29} While this is an extremely effective approach, the method has some drawbacks in that, despite reports of the synthesis on mono substituted analogues,³⁰ such reactions are difficult and often an impure product is obtained which requires extensive efforts to purify. Secondly, the substituent is bound by an amide group which results in a lower thermodynamic stability and potentially increased toxicity.

For the direct attachment of a moiety to the diethyltriamine framework, typically, (i) a single moiety may be attached to the central nitrogen,³¹ (ii) two identical moieties may be attached to the terminal nitrogens^{32,33} or (iii) the ethylene spacers of the framework may be substituted.^{34,35}

The dialkylation of a primary amine using di-*tert*-butyl 2,2'-((2-bromoethyl)azanediyl)diacetate (2) will result in the formation of a DTPA framework. Amino acids, such as L-cysteine,³⁶ L-phenylalanine,³⁷ L-glutamic acid and lysine,³⁸ were previously utilised as primary amines in the construction of such DTPA analogues. In addition, there is a further report of an aliphatic alkyl amine³¹ being used as the central moiety of a constructed DTPA-like ligand.

Further, bi-substituted DTPA analogues have been constructed from diethylene triamine starting material using two equivalents of 2,2'-bipyridine-5-carboxaldehyde,³² while Artali *et al.*³³ reported an analogous approach to incorporate the 2-hydroxymethyl pyridine group. However, it should be noted, that reports of mono substituted terminal nitrogens of a DTPA framework are scarce.^{39–42}

Furthermore, reproducible relaxivity measurements reported for L^1 in this report is on the higher side. Most of the polyamino polycarboxylate scaffolds used as commercial

School of Chemistry, Cardiff University, Main Building, Park Place, CF10 3AT, Cardiff, UK. E-mail: AmorosoAJ@cardiff.ac.uk

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c7ra06946d



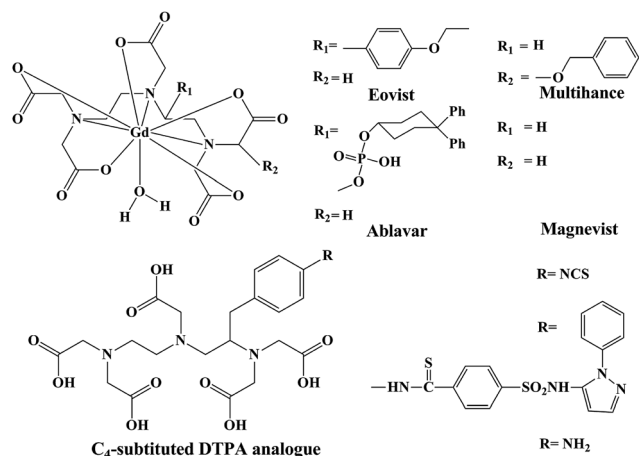


Fig. 1 Schematic representation of commercial contrast agents and some selected C4-substituted DTPA analogues.

contrast agents have relaxivities in the range of $4\text{--}5\text{ mM}^{-1}\text{ s}^{-1}$ (at 30 MHz, 25 °C). Acyclic DTPA analogues reported to date also have comparable relaxivities to those of commercially available contrast agents based on polyamino polycarboxylates (Fig. 1). Exceptionally higher relaxivities were reported by Raymond *et al.*⁴³ for their HOPO based contrast agents ($7\text{--}13\text{ mM}^{-1}\text{ s}^{-1}$). Very recently, piperidine based rigidified DTPA analogues have been reported to exhibit relaxivity in the range of $5.2\text{--}6.0\text{ mM}^{-1}\text{ s}^{-1}$ (at 20 MHz, 25 °C).

DTPA based amide derivatives bearing chromophores: a novel approach

Herein we report the synthesis of two potential dual mode imaging agents. The synthesised ligand will contain a single organic luminescent moiety which has been directly alkylated to the nitrogen atom of a diethylene triamine. Two possibilities exist, with the lumophore being alkylated to the central nitrogen or to one of the two terminal nitrogens. The first approach utilises **2** for the dialkylation of the appropriate amine. This approach will produce a symmetric ligand (Fig. 2, type A). The appropriate choice of amine will yield a ligand with a lumophore bonded to the central amine. The second approach utilises a monosubstituted diethylene triamine species which may be alkylated to yield an

asymmetrically monosubstituted DTPA ligand (type B). The lumophore is incorporated into these ligand frameworks as the naphthalimide moiety (Fig. 2).

Experimental

General experimental

Reagents were obtained from commercial sources and used as received unless otherwise noted. Solvents were dried and purified under N_2 by using standard methods and were distilled immediately before use, specifically dimethylformamide, was distilled using anhydrous calcium hydride. All compounds were prepared under N_2 unless otherwise mentioned. The NMR spectra were obtained using a Bruker ARX 400 at 20 °C in CDCl_3 unless otherwise noted. Mass spectra were performed on a micromass platform II system, operating in flow injection analysis mode, with the electrospray method. Infrared spectra were recorded with a JASCO FTIR-410 spectrometer, between 4000 and 500 cm^{-1} as KBr pellets. UV/vis spectra and measurements were recorded with a JASCO V-570 spectrophotometer.

The synthesis of 2-(2-aminoethyl)-1*H*-benzo[*de*]isoquinoline-1,3(2*H*)-dione (**3**) was carried out following the reported literature procedure.^{36b}

Luminescence measurements

The photophysical data for luminescence were obtained on a JobinYvon-Horiba Fluorolog spectrometer fitted with a JYTBX picosecond photo detection module. Water insoluble DTPA analogues were dissolved in chloroform and as required water-soluble lanthanide complexes of DTPA analogues were dissolved in water. Although all of the new ligands and complexes prepared were air stable both in the solid state and solution, some of the ligands and complexes were found to be hygroscopic, when left in the open atmosphere for prolonged periods of time.

^1H NMRD acquisition

The $1/T_1$ NMRD profiles were obtained on a Stellar Spin master FFC-2000 relaxometer, typically covering a continuum of magnetic fields from 2.4×10^{-4} to 0.72 T (corresponding to a proton Larmor frequency range 0.01–30 MHz). A Spin master Variable Temperature Controller (VCT) allowed the setting and monitoring of the sample temperature at two different temperatures, 25 °C and 37 °C with a resolution up to 0.1 °C. Each sample was allowed to acclimatise to the desired temperature for 15 minutes prior to data collection. The reproducibility in T_1 measurements was within $\pm 1\%$.

Synthesis of di-*tert*-butyl 2,2'-((2-hydroxyethyl)azanediyl) diacetate (1**).**³⁶ The procedure as reported by Choi *et al.*³⁶ was used, except for slight modifications. To *tert*-butyl bromoacetate (17.02 ml, 116 mmol) dissolved in DMF (30 ml), potassium carbonate (12.51 g, 90 mmol) was added. The reaction mixture was cooled down to 0 °C and ethanolamine (2.98 ml, 49.4 mmol) added dropwise for 5 min. The reaction mixture was stirred at 0 °C for 30 min and then allowed to stir overnight at room

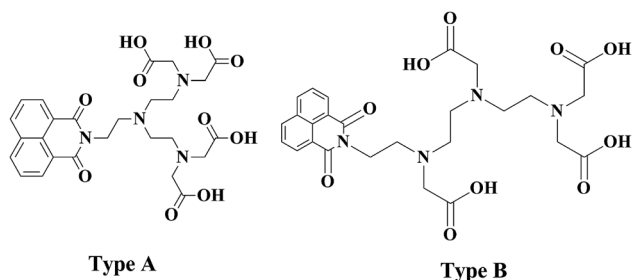


Fig. 2 Depicting the novel chromophore bearing centrally substituted and terminally substituted DTPA analogues.



temperature. After an addition of concentrated NaHCO_3 (100 ml) and diethyl ether (150 ml), the organic layer was separated and washed with concentrated NaHCO_3 (50 ml) and subsequently with brine (200 ml). The solvent was removed under reduced pressure to yield the product as pale yellow solid. Yield 14.2 g, 42%; characterisation data was similar to that previously reported.³⁶

Synthesis of di-*tert*-butyl 2,2'-((2-bromoethyl)azanediyl) diacetate (2).³⁸ PPh_3 (4.34 g, 16.55 mmol) was dissolved in dichloromethane (25 ml), and NBS (2.99 g, 16.80 mmol) was added portion wise. The mixture was allowed to stir at 0 °C (until the colour of the solution changes from yellow to light brown). The mixture was added to a dichloromethane (25 ml) solution of **1** (6.00 g, 20.74 mmol), dropwise at 0 °C. The reaction mixture was allowed to stir overnight. The solvent was removed and to the crude product was added diethyl ether (30 ml). The sticky solid was triturated and filtered. The filtrate was evaporated and the crude product was flash chromatographed (hexane : diethyl ether, 5 : 1). The product was obtained as a colourless oil. Yield 2.0 g, 28%; ^1H NMR (400 MHz; CDCl_3) δ_{H} (ppm) 3.45 (s, 4H, $\text{NCH}_2\text{COOC}(\text{CH}_3)_3$), 3.41 (t, 2H, $J = 7.4$ Hz, BrCH_2), 3.11 (t, 2H, $J = 7.4$ Hz, $\text{BrCH}_2\text{CH}_2\text{N}$), 1.44 (s, 18H, $\text{COOC}(\text{CH}_3)_3$).

Synthesis of tetra-*tert*-butyl 2,2',2'',2'''-(((2-(1,3-dioxo-1H-benzo[de]isoquinolin-2(3H)-yl)ethyl)azanediyl)bis(ethane-2,1-diyl))bis(azanetriyl))tetraacetate (4). To bromide **2** (1.00 g, 2.84 mmol) was added K_2CO_3 (3.40 g, 25 mmol) and previously synthesized **3** (0.34 g, 1.42 mmol). DMF was added and the reaction mixture was gently heated to 65 °C and stirred overnight. The solvent was removed. The crude product was purified by flash chromatography (5% MeOH : DCM mixture). The pure product was obtained as a yellow oil. Yield 0.62 g, 54%; ^1H NMR (400 MHz, CDCl_3): δ_{H} (ppm) 8.50 (d, 2H, $J = 7.27$ Hz, ArH), 8.10 (d, 2H, $J = 8.13$ Hz, ArH), 7.70 (m, 2H, ArH), 4.30 (m, 4H), 3.40 (m, 8H), 2.85 (br s, 8H), 1.40 (s, 36H); ^{13}C NMR (400 MHz; D_2O): δ_{C} 170.7, 163.4, 133.2, 130.6, 127.2, 121.5, 80.9, 45.4, 52.4, 55.1, 27.1; IR (KBr disc) (cm^{-1}): 3440(br), 2977(s), 2931(w), 1736(s), 1702(w), 1661(vs), 1628(w), 1591(s); ESI-MS (+ion): found m/z 783.4578, calc. 783.4544 for $[(4) \text{H}]^+$.

2,2',2'',2'''-(((2-(1,3-dioxo-1H-benzo[de]isoquinolin-2(3H)-yl)ethyl)azanediyl)bis(ethane-2,1-diyl))bis(azanetriyl))tetraacetic acid, (L**¹).** To compound **4** (0.12 g, 0.153 mmol) was added 2 M HCl (10 ml, 323 mmol) and the reaction mixture was refluxed for 2 h. The solvent was removed under high vacuum to give the product as pale yellow solid. Yield 0.072 g, 84%; ^1H NMR (400 MHz, D_2O) δ_{H} (ppm) 8.00 (d, 2H, $J = 6.79$ Hz), 7.92 (d, 2H, $J = 8.41$ Hz), 7.40 (t, 2H, $J = 6.94$ Hz), 3.7 (m, 4H), 3.40 (s, 8H), 2.90 (m, 4H), 2.80 (m, 4H); ^{13}C NMR (400 Hz, D_2O) δ_{C} 172.2, 168.7, 135.4, 135.6, 131.8, 131.6, 130.8, 127.1, 126.6, 119.9, 55.8, 55.3, 52.3, 50.8, 37.0; IR (KBr disc) (cm^{-1}): 3430(br), 2963(w), 1695(s), 1655(vs), 1625(w), 1588(s); ESI-MS (–ion): found m/z 557.16, calc. 557.00 for $[(\text{L}^1) \text{H}]^-$.

Complexation of **L**¹ with Gd(III) chloride

L¹ (28 mg, 0.05 mmol) and $\text{GdCl}_3 \cdot 6\text{H}_2\text{O}$ (18.0 mg 0.05 mmol) were added to two different vials. The ligand was dissolved in

ethanol 10 ml, and heated slightly to ensure complete dissolution. Then GdCl_3 was dissolved in distilled H_2O , and again heated to ensure complete dissolution. Thereafter the vial containing the metal salt solution was continuously stirred, and the ligand solution was added dropwise. The instantaneous formation of the precipitate was observed. After the complete addition of the ligand to the metal, the mixture was stirred for two days in the dark. The solvent was evaporated to give a yellow precipitate of the complex. Yield 84%; IR (KBr disc, cm^{-1}): 3425(br), 1729(w), 1625(s), 1408(s); ESI-MS (–ion): found m/z 712.0898, calc. 712.0890 for $[(\text{L}^1) \text{Gd}]$. UV/vis [λ_{max} , nm (ϵ , $\text{M}^{-1} \text{cm}^{-1}$)] in H_2O : 235(17 658), 274(4128), 344(6279). The complexation reaction of **L**¹ with Eu(III) chloride, Yb(III) chloride and Nd(III) chloride were carried out as described for complexation with Gadolinium chloride. However in the case of Nd(III) chloride it had to be dissolved in DMF, instead of water.

Synthesis of 2-(2-((2-(2-aminoethyl)amino)ethyl)amino)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (5). To 1,8-naphthalic anhydride (1.00 g, 5.05 mmol) was added triethylenetetramine (4.50 ml, 30 mmol) and allowed it to reflux overnight. Solvent was removed by distillation. The crude product was dissolved in dichloromethane and filtered. The filtrate was evaporated. To purify further, the product was extracted into toluene; the toluene washings (3 × 30 ml) were evaporated to dryness and the product was obtained as a dark brown sticky solid. Yield 0.7 g, 42%; ^1H NMR (D_2O , 400 Hz) δ_{H} (ppm) 8.50 (d, 2H, $J = 8.53$ Hz, ArH), 8.10 (d, 2H, $J = 8.14$ Hz, ArH), 7.70 (t, 2H, $J = 8.00$ Hz, ArH), 4.35 (t, 2H, $J = 5.8$ Hz, $\text{NCH}_2\text{CH}_2\text{NH}$), 2.95 (t, 4H, $J = 12$ Hz, $\text{NCH}_2\text{CH}_2\text{NH}$), 2.80 (t, 2H, $J = 8.00$ Hz, $\text{NHCH}_2\text{CH}_2\text{NH}$), 2.7 (t, 2H, $J = 10$ Hz, $\text{NHCH}_2\text{CH}_2\text{NH}_2$), 2.6 (m, 4H); IR (KBr disc) (cm^{-1}): 3416(br), 2964(s), 2822(w), 1699(s), 1660(vs), 1625(w), 1590(s), 1439(s); ESI-MS (+ion): found m/z 349.15, calc. 349.17 for $[(5) \text{H}]^+$.

Synthesis of di-*tert*-butyl 2,2'-((2-((2-(*tert*-butoxy)-2-oxoethyl)(2-((2-(*tert*-butoxy)-2-oxoethyl)(2-(1,3-dioxo-1H-benzo[de]isoquinolin-2(3H)-yl)ethyl)amino)ethyl)amino)ethyl)azanediyldiacetate (6). To *tert*-butyl bromoacetate (2.27 ml, 15.58 mmol) was added K_2CO_3 (10.00 g, 72.46 mmol) and previously synthesized **5** (1.01 g, 3.00 mmol). After the addition of DMF (20 ml), the reaction mixture was gently heated up to 65 °C and allowed to stir overnight. Then the solvent was removed. The crude product was purified by flash chromatography the material (90 : 10 dichloromethane/methanol); the product was obtained from the eluents by evaporation of the solvent mixture. Yield 0.25 g, 20%; ^1H NMR (CDCl_3 , 400 MHz) δ_{H} 8.50 (d, 2H, $J = 7.13$, ArH_a), 8.10 (d, 2H, $J = 7.98$, ArH_c), 7.70 (t, 2H, $J = 7.73$, ArH_b), 4.2 (m, 2H), 3.4 (s, 8H), 2.9 (m, 2H), 2.8 (m, 2H), 2.7 (m, 2H), 2.6 (m, 4H), 1.4 (s, 36H); ^{13}C NMR (400 MHz, CDCl_3): δ_{C} 169.0, 163.1, 132.8, 130.6, 130.2, 125.9, 121.7, 79.8, 55.0, 54.8, 51.9, 51.8, 50.9, 37.2, 27.1, 27.0, 27.1, 26.9; IR (KBr disc) (cm^{-1}): 3467(br), 2973(s), 2360(s), 1733(vs), 1648(s), 1595(s); ESI-MS (+ion): found m/z 783.45, calc. 783.45 for $[(6) \text{H}]^+$.

Synthesis of 2,2'-((2-((carboxymethyl)(2-((carboxymethyl)(2-(1,3-dioxo-1H-benzo[de]isoquinolin-2(3H)-yl)ethyl)amino)ethyl)amino)ethyl)azanediyl)diacetic acid (L**²).** The compound **6** (0.12 g, 0.15 mmol) was added to a round bottom flask and 2 M



HCl (10 ml) was added and the reaction mixture was refluxed for 2 h. After 2 h, the solvent was removed *in vacuo*. Yield 0.07 g, 58%; ^1H NMR (400 MHz, D_2O) δ_{H} (ppm) 8.00 (dd, 4H, $J = 8.8$ Hz, other constant not determined, $\text{ArH}_{\text{a,c}}$) 7.45 (t, 2H, $J = 26$ Hz, ArH_{b}), 3.00–4.25 (m, 20H); IR (KBr disc) (cm^{-1}): 3428(br), 29649(w), 2532(w), 1738(s), 1702(w), 1654(s), 1590(w); ESI-MS (+ion): found m/z 559.2057, calc. 559.2040 for $[(\text{L}^2)\text{H}]^+$.

Complexation of L^2 with Gd(III) chloride

Complexation was carried out in a similar manner described for L^1 . Yield 85%; IR (KBr disc) (cm^{-1}): 3404(br), 1728(w), 1644(s), 1408(s); ESI-MS (–ion): found m/z 714.23, calc. 714.11 for $[(\text{L}^2)\text{Gd}]$. UV/vis [λ_{max} , nm (ϵ_{M} , $\text{M}^{-1}\text{cm}^{-1}$)] in H_2O : 235(6752), 267(1679), 345(2309).

Results and discussion

General synthesis

Ligand design and synthesis of DTPA analogues

Symmetric ligand, L^1

(a) *Synthesis of 2*. The synthesis of **2** following the reported procedure gave low yields in our hands. However a small modification to the procedure allowed us to synthesise **2** in improved yield and purity. The phosphonium bromide was first prepared by the portionwise addition of NBS to the dissolved solution of triphenyl phosphine in dichloromethane (successful preparation of the phosphonium bromide is indicated by the formation of sticky, semi solid-triphenylphosphine oxide).⁴⁴ The prepared phosphonium bromide was then added dropwise to the solution of **1** in dichloromethane. The rest of the procedure was followed as reported by Anelli *et al.*³⁸

(b) *Alkylation of 3*. Initially, compound **3** was added together with bromide **2** in the stoichiometric ratio of 1 : 2 in anhydrous DMF (Fig. 3). The reaction mixture was heated to 65 °C and allowed to reflux for 48 h. Purification by thin layer chromatography (TLC) was carried out with 5% methanol/dichloromethane used as the eluent. The purified protected form of the ligand thus obtained, subsequently underwent deprotection by refluxing the protected ligand in 2 M HCl for two hours according to the procedure reported by Vogt *et al.*³⁰ The ligand L^1 was isolated in good yield (90%).

Asymmetric ligand, L^2 . Triethylenetetramine (TRIEN) was reacted with 1,8-naphthalic anhydride to produce the new ligand **5**. The ^1H NMR spectrum of the crude mixture confirmed the reaction had occurred with a significant change to the aliphatic protons consistent with a lowering of the symmetry of the TRIEN core.

The alkylation of this product was carried out using the same conditions as for the symmetric ligand, using stoichiometric amounts of alkylation agent (Fig. 4). With resulting low yields, a 1 : 5 molar ratio was used to try to improve the amount of product and its purity, however ^1H NMR still indicated an excess of compound **5**. The reaction time was varied from 2 to 7 days, with little effect on the yield. Similarly, increasing the reaction temperature did not appreciably improve the yield. Purification of the reaction mixture was carried out by

chromatography (95 : 5 dichloromethane/methanol) giving a yield of <20% isolated product.

Characterisation

^1H and ^{13}C NMR. The purified, protected and deprotected forms of L^1 and L^2 were characterized by NMR spectroscopy. Each compound gave a spectrum consistent with the proposed structure. Chemical shifts were assigned by comparison to previously published data.^{15,35,45}

In **4**, the four equivalent pendant carboxylate CH_2 -protons appear as a broad singlet at 2.85 ppm indicating a strong upfield shift due to alkylation and the CH_2 -protons are subsequently shifted to 3.40 ppm upon deprotection, in L^1 . Curiously, a similar singlet was visible in the protected form of L^2 (**6**) at 3.40 ppm, suggesting a less intensive inductive effect by adjacent *t*-butyl groups.

IR spectroscopy. The infrared spectrum of both L^1 and L^2 exhibited strong absorption bands in the region of 3300–3400 cm^{-1} and 1650 cm^{-1} , corresponding to NH and CO stretching vibrations, respectively (Table 1). Upon complexation with gadolinium, the CO stretching vibrations of the amide

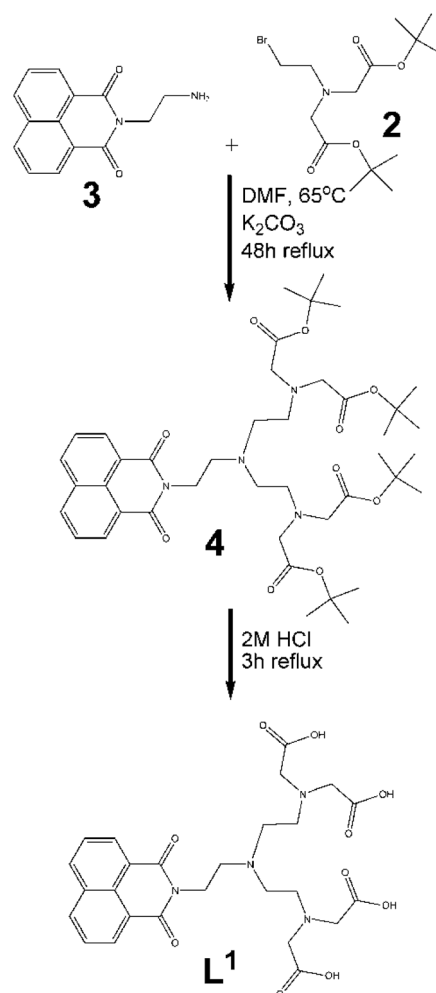


Fig. 3 Schematic representation of the synthesis of symmetric ligand (L^1).



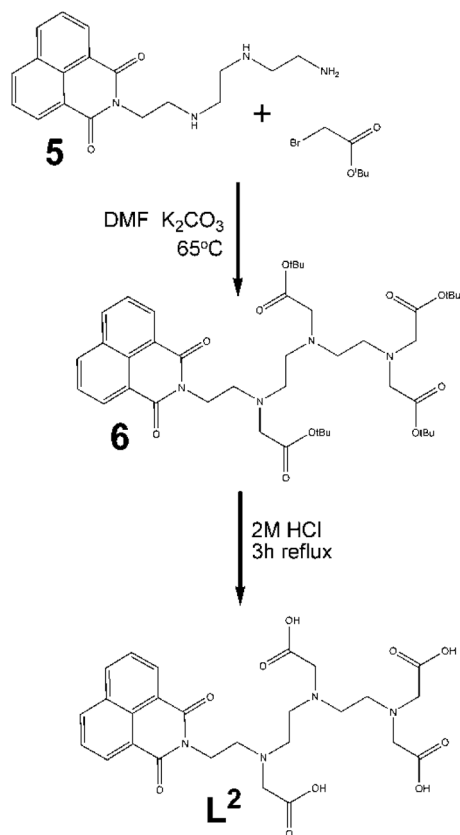


Fig. 4 Schematic representation of the synthesis of asymmetric ligand (L^2).

attracts a red shift for both L^1 and L^2 . However, while L^1 exhibited a significant red shift (*ca.* 30 cm^{-1}), L^2 exhibited a smaller shift (*ca.* 10 cm^{-1}). This may be attributed to weak coordination of the amide oxygen to the metal ion, in contrast to L^1 .

Mass spectrometry. The molecular ions for the Gd(III) complexes of both L^1 and L^2 were visible in the mass spectra of these respective ligands. In the ESI mass spectra acquired in the negative mode indicated molecular ion peak of L^1 at 557.16. However, the most intense peak for L^2 at 559.20 was found in the positive mode.

Evaluation of DTPA analogues

Both centrally and terminally substituted DTPA analogues have been exploited for their use in MRI applications.⁴⁷ As of today, most of the DTPA analogues tested and recognized as contrast agents belong to the terminally-substituted category,⁴⁸ but investigations related to direct substitution of a chromophoric moiety in the central nitrogen, remains scarce.^{13,44,46}

Similarly, terminally substituted and C-substituted (on ethylenic carbon) analogues comprise the majority of DTPA analogues (including the commercially available contrast agents, namely Eovist® and Multihance®). The relaxivity of the new contrast agent based on L^1 at relevant magnetic field strengths (8.10 $\text{mM}^{-1} \text{s}^{-1}$ at 30 MHz and 25 °C) is considerably higher than that of the commercially available contrast agents

Table 1 A comparison of frequencies (cm^{-1}) obtained for different vibrational modes of IR for the symmetric and asymmetric ligands along with their starting materials and Gd(III) complexed forms^a

Compound	$\nu(\text{C=O})$ amide	$\nu(\text{C=O})$ ester	$\nu(\text{C=O})$ acid	$\nu(\text{O-H})$	$\nu(\text{N-H})$
2		1737			
3	1662				3346
4	1654	1739			3314
L^1	1655		1695	3429	
Gd- L^1	1623		1729	3424	
1 ^a		1731			
5	1659				3416
6	1648	1732			3414
L^2	1654		1702	3427	
Gd- L^2	1644		1728	3403	

^a 1^a-*tert*-butyl bromoacetate.

Eovist®, Multihance® and Ablavar™ (previously known as Vasovist® or MS-325) (Table 2).

In contrast to L^1 , the poor yield of L^2 limited analogous investigations of relaxivity.

DTPA analogues have also been exploited for luminescence based applications. Examples include DTPA functionalised with Carbostyryl-124 (CS-124),⁵² CS-124 covalently bound to trimethoprim,⁵³ amine functionalised reactive groups such as isothiocyanates and thiol-reactive groups such as pridylthiol, maleimide, methanethiosulfonate and haloacetyl acetamide.⁵⁴ Further, DTPA analogues have been investigated for their usage in dissociation enhanced fluoroimmunoassay (DELFI).⁵⁵

Fluorescence on binding with lanthanides

In the following experiments we have not concerned ourselves with the lanthanide phosphorescence spectrum. Instead, we are interested by changes in the organic fluorescence spectrum on lanthanide binding. Primarily, we wish to show that the organic lumophore is not totally quenched upon lanthanide binding and any shift in the spectrum is small.

The emission profiles of the protected form of the symmetric ligand, 4, compared to its deprotected form, L^1 , exhibited only slight differences, with only a slight red shift occurring in the excitation wavelength. The formation of the gadolinium

Table 2 A comparison of relaxivities of commercially available MRI contrast agents with L^1

Ligand	Temp. (°C)	¹ H freq. (MHz)	r_1 ($\text{s}^{-1} \text{mM}^{-1}$)	Ref.
GdCl ₃ 1 mM (free)	25	30	12.64	
Gd(III)- L^1 (complex)	25	30	8.10	
Magnevist®	37	20	3.80	49
Omniscan® ^a	37	20	3.80	50
Eovist®	37	64	7.14	49
Multihance®	39	20	5.35	51
Ablavar™	37	64	5.20	49

^a Gd-DTPA-BMA.



Table 3 Spectroscopic properties of the symmetric ligand in its protected, deprotected and complexed forms, measured in $\text{CHCl}_3 \cdot \text{H}_2\text{O}$ and λ_{ex} , λ_{em} , are excitation maxima, emission maxima and fluorescent yield respectively

Compound	λ_{ex} (nm)	λ_{em} (nm)	A	ϵ
4^a	336	383		
L^1	345	396		
Gd-L^1	345	395	235	6752
6^a	335	383		
L^2	345	396		
Gd-L^2	345	396	235	17 658

^a Water insoluble ligand, dissolved in chloroform.

complex, confirmed through the use of xylene orange, results in little change to the luminescence spectrum, with no difference noted in the excitation wavelength and no change observed in the emission spectrum, compared to L^1 (Table 3 and Fig. 5). Similarly, the formation of the gadolinium complex of L^2 did not result in a significant quenching of the luminescence signal of the free ligand (Fig. 6), nor did it shift the excitation and emission wavelengths to a significant degree.

Clearly, the luminescence spectroscopy indicates that it may be possible to utilize the gadolinium complexes of L^1 and L^2 as luminescent probes, though their efficiency in such an application remains to be determined.

Relaxivity measurements

2,2',2'',2'''-(((2-(1,3-dioxo-1H-benzo[de]isoquinolin-2(3H)-yl)ethyl)azanediyl)bis(ethane-2,1 diyl))bis(azanetriyl))tetra acetic acid (L^1). Fig. 7 depicts the ^1H NMRD profile of the $\text{Gd}(\text{III})$ complex of L^1 . Reproducibility of the results was ensured by repeating the whole synthesis and repeating the measurements under the same conditions. Long term reproducibility of relaxivity of the same sample, under identical experimental conditions over an extended period of time (4 months) was also established. Long term stability of the same sample in solution was also corroborated by testing with xylene orange indicator indicating the absence of free $\text{Gd}(\text{III})$ in the sample. The relaxivity of a 1 mM concentration solution of the complex is high compared to other standard DTPA based complexes ($8.10 \pm 0.21 \text{ mM}^{-1} \text{ s}^{-1}$ at 30 MHz, 25 °C). The comparable relaxivity data is given in Table 2.

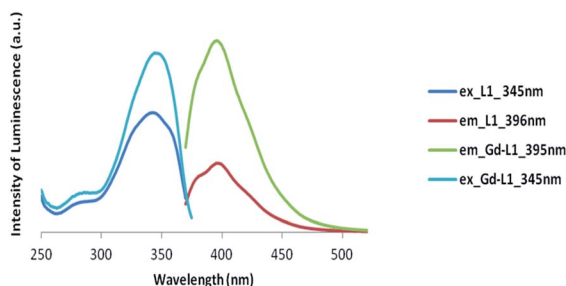


Fig. 5 Luminescence spectra of Gd complex of symmetric ligand (Gd-L^1) (excitation at 345 nm and emission at 395 nm).

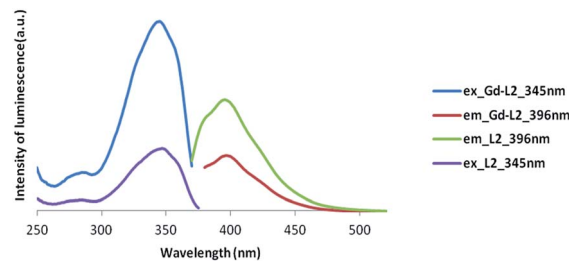


Fig. 6 Luminescence spectra of Gd complex of asymmetric ligand (Gd-L^2) (excitation at 345 nm and emission at 396 nm).

The enhancement relaxivity of water protons results from dipolar interaction of the electron magnetic moment of the paramagnetic metal ion with that of the nuclear magnetic moment of solvent nuclei. This interaction comprises contributions from water molecules that belong to the inner, outer and secondary coordination spheres. It has been reported, that for low molecular weight $\text{Gd}(\text{III})$ based mono aqua hydrophilic complexes, the inner-sphere and outer-sphere contributions are comparable, giving rise to relaxivity values which lie in the range of $4\text{--}5 \text{ mM}^{-1} \text{ s}^{-1}$ at 25 °C and 20 MHz,¹ while the contribution from secondary-sphere water molecules has not been as extensively studied.⁵⁶

However, the observed relaxivity was very much on the higher side for $\text{Gd}(\text{III})\text{-L}^1$ ($8.65 \text{ mM}^{-1} \text{ s}^{-1}$) could be related to the concentration of the contrast agent by the following equation.

$$R_{\text{obs}} = R_{\text{IW}} + [\text{Gd}(\text{III})\text{-L}^1]r_{1p}$$

where R_{IW} represents the relaxivity of pure water ($0.55 \text{ mM}^{-1} \text{ s}^{-1}$ in 30 MHz, 25 °C). Therefore the true relaxivity of $\text{Gd}(\text{III})\text{-L}^1$ is $8.10 \text{ mM}^{-1} \text{ s}^{-1}$, which is still relatively higher than the relaxivity of the commercial contrast agent, 'Eovist'. As per the equation below, relaxivity is highly dependent on the number of water molecules bound to the metal centre (q) and their rate of exchange with the bulk water (τ_M).

$$(1/T_1) = qP_m[1/(T_{1m} + \tau_M)]$$

where $1/T_1$ is the longitudinal relaxation rate and P_m is the mole fraction of water coordinated to the metal centre. All

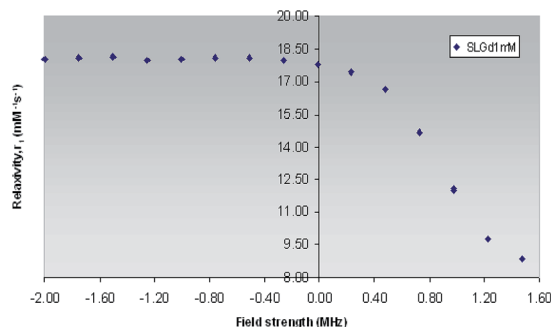


Fig. 7 ^1H NMRD Profile of $\text{Gd}(\text{III})\text{-L}^1$ complex obtained across field strengths ranging from 0.01–30 MHz (expressed in logarithmic scale for purpose of clarity) at 25 °C.



commercial contrast agents possess $q = 1$, which results in comparatively low relaxivity. Gd(III) complexes possessing high relaxivity with high q values ($q = 2$) have been reported in the past for acyclic aminocarboxylate compounds as well.⁵⁷ However, such an approach is problematic due to decreased complex stability and hence increased agent toxicity. In addition, higher q values may lead to the ready formation of ternary complexes in biological media and ultimately lead to lower relaxivity *in vivo*.

Conclusions

Two new DTPA analogues have been synthesized. A chromophore bearing, central nitrogen substituted, DTPA analogue has been synthesized distinguishing these new ligands from commercial contrast agents and incorporating the naphthalimide moiety into the DTPA backbone. The Gd(III) complexes exhibit ligand-based luminescence and Gd-L¹ has a relatively high relaxivity. Further investigations are necessary to determine if these reagents are toxic and the relaxivity is maintained *in vivo*.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

We would like to thank CITER (Cardiff Institute of Tissue Engineering and Repair) for infrastructure support (JobinYvon-Horiba Fluorolog spectrometer and Stellar Spin master FFC-2000 relaxometer).

References

- 1 P. Caravan, J. J. Ellison, T. J. McMurphy and R. B. Lauffer, *Chem. Rev.*, 1999, **99**, 2293–2352.
- 2 R. B. Lauffer, *Chem. Rev.*, 1987, **87**, 901–927.
- 3 C. Paul-Roth and K. N. Raymond, *Inorg. Chem.*, 1995, **34**, 1408–1412.
- 4 H. S. Chong, K. Garmestani, L. H. Bryant and M. W. Brechbiel, *J. Org. Chem.*, 2001, **66**, 7745–7750.
- 5 P. Verwilt, S. Park, B. Yoon and J. S. Kim, *Chem. Soc. Rev.*, 2015, **44**, 1791–1806.
- 6 M. W. Brechbiel, *Q. J. Nucl. Med. Mol. Imag.*, 2008, **52**, 166–173.
- 7 J. D. G. Correia, A. Paulo, P. D. Raposinho and I. Santos, *Dalton Trans.*, 2011, **40**, 6144–6167.
- 8 Y. F. Liu and C. C. Wu, *Pure Appl. Chem.*, 1991, **63**, 427–463.
- 9 G. L. Griffiths, D. M. Goldenberg, A. L. Jones and H. J. Hansen, *Bioconjugate Chem.*, 1992, **3**, 91–99.
- 10 Y. Liu, G. Liu and D. J. Hnatowich, *Materials*, 2010, **3**, 3204–3217.
- 11 R. E. Weiner and M. L. Thakur, *Appl. Radiat. Isot.*, 2002, **57**, 749–763.
- 12 T. Storr, K. H. Thompson and C. Orvig, *Chem. Soc. Rev.*, 2006, **35**, 534–544.
- 13 V. C. Pierre, M. J. Allen and P. Caravan, *J. Biol. Inorg. Chem.*, 2014, **19**, 127–131.
- 14 D. W. Zhang, Z. Y. Yang, B. D. Wang, S. P. Zhang and R. D. Yang, *Chem. Pharm. Bull.*, 2006, **54**, 1203–1206.
- 15 Z. Jászberényi, A. Sour, É. Tóth, M. Benmelouka and A. E. Merbach, *Dalton Trans.*, 2005, 2713–2719.
- 16 J. Feng, G. Sun, F. Pei and M. Liu, *Bioorg. Med. Chem.*, 2003, **11**, 3359–3366.
- 17 L. V. Elst, A. Roch, P. Gillis, S. Laurent, F. Botteman, J. W. M. Bulte and R. N. Muller, *Magn. Reson. Med.*, 2002, **47**, 1121–1130.
- 18 Y. M. Wang, S. T. Lin, Y. J. Wang and R. S. Sheu, *Polyhedron*, 1998, **17**, 2021–2028.
- 19 H. Imura, G. R. Choppin, W. P. Cacheris, L. A. de Learie, T. J. Dunn and D. H. White, *Inorg. Chim. Acta*, 1997, **258**, 227–236.
- 20 C. F. G. C. Geraldes, A. M. Urbano, M. C. Alpoim, A. D. Sherry, K. T. Kuan, R. Rajagopalan, F. Maton and R. N. Muller, *Magn. Reson. Imaging*, 1995, **13**, 401–420.
- 21 S. Aime, M. Botta, W. Dastru, M. Fasano, M. Panero and A. Arnesi, *Inorg. Chem.*, 1993, **32**, 2068–2071.
- 22 C. F. G. C. Geraldes, A. M. Urbano, M. C. Alpoim, M. A. Hoefnagel and J. A. Peters, *J. Chem. Soc., Chem. Commun.*, 1991, 656–658.
- 23 W. P. Cacheris, S. C. Quay and S. M. Rocklage, *Magn. Reson. Imaging*, 1990, **8**, 467–481.
- 24 A. D. Sherry, W. P. Cacheris and K.-T. Kuan, *Magn. Reson. Med.*, 1988, **8**, 180–190.
- 25 L. Frullano and P. Caravan, *Curr. Org. Synth.*, 2011, **8**, 535–565.
- 26 V. Darras, M. Nelea, F. M. Winnik and M. D. Buschmann, *Carbohydr. Polym.*, 2010, **80**, 1137–1146.
- 27 S. W. A. Bligh, A. H. M. S. Chowdhury, D. Kennedy, C. Luchinat and G. Parigi, *Magn. Reson. Med.*, 1999, **41**, 767–773.
- 28 Y. M. Wang, T. H. Cheng, G. C. Liu and R. S. Sheu, *J. Chem. Soc., Dalton Trans.*, 1997, 833–838.
- 29 M. S. Konings, W. C. Dow, D. B. Love, K. N. Raymond, S. C. Quay and S. M. Rocklage, *Inorg. Chem.*, 1990, **29**, 1488–1491.
- 30 T. N. Parac-Vogt, K. Kimpe, S. Laurent, C. Piérart, L. V. Elst, R. N. Muller and K. Binnemans, *Eur. J. Inorg. Chem.*, 2004, **2004**, 3538–3543.
- 31 T. Storr, B. R. Cameron, R. A. Gossage, H. Yee, R. T. Skerlj, M. C. Darkes, S. P. Fricker, G. J. Bridger, N. A. Davies, M. T. Wilson, K. P. Maresca and J. Zubieta, *Eur. J. Inorg. Chem.*, 2005, **2005**, 2685–2697.
- 32 J. M. Couchet, J. Azéma, P. Tisnès and C. Picard, *Inorg. Chem. Commun.*, 2003, **6**, 978–981.
- 33 R. Artali, M. Botta, C. Cavallotti, G. B. Giovenzana, G. Palmisano and M. Sisti, *Org. Biomol. Chem.*, 2007, **5**, 2441–2447.
- 34 M. W. Brechbiel and O. A. Gansow, *Bioconjugate Chem.*, 1991, **2**, 187–194.
- 35 C. H. Cummins, E. W. Rutter and W. A. Fordyce, *Bioconjugate Chem.*, 1991, **2**, 180–186.



- 36 (a) K. Choi, Y. D. Hong, M. S. Pyun and S. J. Choi, *Bull. Korean Chem. Soc.*, 2006, **27**, 1194–1198; (b) M. Licchelli, A. O. Biroli, A. Poggi, D. Sacchi, C. Sangermani and M. Zema, *Dalton Trans.*, 2003, 4537–4545.
- 37 M. A. Williams and H. Rapoport, *J. Org. Chem.*, 1993, **58**, 1151–1158.
- 38 P. L. Anelli, F. Fedeli, O. Gazzotti, L. Lattuada, G. Lux and F. Rebasti, *Bioconjugate Chem.*, 1999, **10**, 137–140.
- 39 Y. Arano, T. Uezono, H. Akizawa, M. Ono, K. Wakisaka, M. Nakayama, H. Sakahara, J. Konishi and A. Yokoyama, *J. Med. Chem.*, 1996, **39**, 3451–3460.
- 40 S. Laurent, F. Botteman, L. V. Elst and R. N. Muller, *Helv. Chim. Acta*, 2004, **87**, 1077–1089.
- 41 J. F. W. Keana and J. S. Mann, *J. Org. Chem.*, 1990, **55**, 2868–2871.
- 42 D. A. Westerberg, P. L. Carney, P. E. Rogers, S. J. Kline and D. K. Johnson, *J. Med. Chem.*, 1989, **32**, 236–243.
- 43 A. Datta and K. N. Raymond, *Acc. Chem. Res.*, 2009, **42**, 938–947.
- 44 C. A. G. N. Montalbetti and V. Falque, *Tetrahedron*, 2005, **61**, 10827–10852.
- 45 (a) M. S. Pyun, K. H. Choi, Y. D. Hong and S. J. Choi, *Bull. Korean Chem. Soc.*, 2009, **30**, 1187–1189; (b) H. R. Maecke, A. Riesen and W. Ritter, *J. Nucl. Med.*, 1989, **30**, 1235–1239.
- 46 R. Krüger, K. Braun, R. Pipkorn and W. D. Lehmann, *J. Anal. At. Spectrom.*, 2004, **19**, 852–857.
- 47 S. Laurent, C. Henoumont, L. V. Elst and R. N. Muller, *Eur. J. Inorg. Chem.*, 2012, **2012**, 1889–1915.
- 48 P. Lebdušková, A. Sour, L. Helm, É. Tóth, J. Kotek, I. Lukeš and A. E. Merbach, *Dalton Trans.*, 2006, 3399–3406.
- 49 K.-H. Jung, H.-K. Kim, G. H. Lee, D.-S. Kang, J.-A. Park, K. M. Kim, Y. Chang and T.-J. Kim, *J. Med. Chem.*, 2011, **54**, 5385–5394.
- 50 S. Laurent, L. V. Elst and R. N. Muller, *Contrast Media Mol. Imaging*, 2006, **1**, 128–137.
- 51 F. Uggeri, S. Aime, P. L. Anelli, M. Botta, M. Brocchetta, C. de Haeen, G. Ermondi, M. Grandi and P. Paoli, *Inorg. Chem.*, 1995, **34**, 633–643.
- 52 P. R. Selvin, *Annu. Rev. Biophys. Biomol. Struct.*, 2002, **31**, 275–302.
- 53 D. R. Reddy, L. E. Pedró Rosa and L. W. Miller, *Bioconjugate Chem.*, 2011, **22**, 1402–1409.
- 54 P. Ge and P. R. Selvin, *Bioconjugate Chem.*, 2004, **15**, 1088–1094.
- 55 J. Peuralahti, L. Meriö, V.-M. Mikkala, K. Blomberg and J. Hovinen, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 4760–4762.
- 56 S. Aime, M. Botta and E. Terreno, *Adv. Inorg. Chem.*, 2005, **57**, 173–237.
- 57 J. Costa, É. Tóth, L. Helm and A. E. Merbach, *Inorg. Chem.*, 2005, **44**, 4747–4755.

