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Flexible, Transparent and Robust SERS Tapes through a Two-Step Block Copolymer Self-Assembly Process

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Surface enhanced Raman scattering (SERS) is an analytical technique that offers the capability of remote sensing, single molecule detection, and detection of trace contaminants (in parts per million) with high sensitivity and accuracy. Here, we demonstrate a simple and economical method for fabricating large area SERS-active tapes that are flexible, transparent and robust using a two-step process. The first is the fabrication of the gold nanoclusters on a flat chip using block copolymer self-assembly followed by directed electrostatic self-organization of gold nanoparticles (AuNPs). The second step involves the transfer of the resulting metal nanoclusters onto a thermal tape by a simple 'stick and peel' technique. Such substrates facilitate the detection and quantification of contaminates on irregular surfaces such as fruit skin, fabrics and other non-planar surfaces without the need to extract the analyte. Furthermore, the SERS measurements are highly quantitative, reproducible and the two-step fabrication process is unprecedented and has potential towards realizing large scale manufacturing of low-cost, flexible SERS-tape for on-field applications.

1. Introduction

Surface-enhanced Raman spectroscopy (SERS) has been an attractive analytical technique for applications that include remote sensing, single molecule detection, label-free detection, bio-sensing, forensics, pesticides and trace chemical detection.¹⁻⁶ It offers an excellent platform to build low cost, versatile and large area SERS-active substrates due to its potential for high sensitivity and accuracy. Raman signals being extremely sensitive to the shape, size and arrangement of the metal structures on the substrate, the fabrication technique involved⁵⁻⁷ demands the need for precise control over feature dimensions and geometry. Furthermore, reproducibility in SERS signal enhancement is crucial to sensing and largely depends on the uniformity in substrate features that are achieved through various nanofabrication processes such as electron beam lithography, UV photolithography and Anodic Aluminium Oxide (AAO) arrays.⁸⁻¹³ Although, these techniques provide highly uniform pattern arrangement that in turn offer high and uniform signal enhancement, they compromise not only on the cost, simplicity in approach, and throughput but also in the adaptability of the techniques to fabricate on flexible substrates for direct sensing of analytes on an irregular surface.

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Flexible SERS substrates are highly versatile in their utility and can be put in use onto irregular surfaces such as fabrics, fruit skin and in forensics^{8, 14-16}. On the other hand, the flat, rigid chips fail to adapt to such conditions, since they demand the extraction of the analyte (using solvent) from non-planar surfaces prior to analysis which is not effective especially in detecting chemicals of trace quantities where the extraction is bound to result in material loss. To overcome these limitations, there are some reports on the fabrication of flexible SERS substrates made of cellulose paper, nanofiber mat and polymer support.¹⁶⁻²⁰ The direct deposition of metal NPs on flexible substrates using various coating methods result in random distribution and varied density of NPs on the surface resulting in variation in SERS signal and thereby making the quantification of analyte difficult.²¹ Moreover, many of these supporting substrates either are not transparent^{22, 23} or the transparency suffers after the addition of the SERS-active layer. The stability of these flexible materials towards harsh solvent and high energy laser also demands further investigation. Fiber-optic based SERS is an alternate, however, they involve high cost and at times not convenient to perform on-field measurements.^{2, 3, 24, 25}

To meet the needs for sensitive, disposable sensor, in this report, we demonstrate the fabrication of flexible SERS-active tapes. They are obtained by pattern transfer of the SERS-active metal nano cluster arrays fabricated through self-assembly processes from flat chip on to a flexible tape through a 'stick and peel' technique. With portable SERS techniques gaining popularity, large area uniform metal nano cluster arrays on the tape that mirror the pattern-arrangement defined on the

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⁺Electronic Supplementary Information (ESI) available: [AFM of the chip after pattern-transfer, histogram of nanoparticles, SEM of the SERS tape and background Raman signal of the SERS-tape]. See DOI: 10.1039/x0xx00000x

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planar substrate becomes the ideal substrate that can adapt to the on-field environment.

Self-assembly of block copolymer (BCP) is often regarded as the technique to produce macroscopic, functional arrays of polymers with ultra-high density in the scale of tens of terabit in⁻².^{26, 27} The BCP self-assembly not only allows for the ordered arrangement of the polymer features but also offers the handle to tune the geometry of the features, viz., the size, pitch and feature-to-feature spacing.^{26, 28} Furthermore, the inherently functionalized features of the blocks provide the leverage to attach (either electrostatically or in-situ) different material components and thereby extends the range of applications starting from memory devices to sensing through SERS.²⁹⁻³² This simplicity offered by BCP self-assembly is exploited to generate gold nanocluster arrays that eliminates the high cost associated with the conventional nanofabrication techniques while at the same time maintain a high level of sensitivity. Together with the ease in pattern transferring onto the flexible tape, the new design represents a significant advancement in on-field analytics making flexible-SERS much more accessible in terms of cost and performance. In addition, the flexible substrates are transparent and allow the direct positioning of the SERS-tape onto the object under investigation directly for efficient analyte collection and detection. The technique demonstrated possibility of high detection limits, is quantitative and has fairly low signal variability across the substrate surface (< 10%) and has potential implications towards commercialization.

2. Experimental

2.1. Materials

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Quartz wafers were purchased from SYST Integration Pte. Ltd., Singapore. Kapton® tape was purchased from S-Chem International (S) Pte. Ltd. Polystyrene-block-poly (2vinylpyridine) (PS-b-PVP) (57000-b-57000 g/mol, PDI = 1.1) was purchased from Polymer Source Inc., (Montreal, Canada). 2propanol and *m*-xylene were obtained as anhydrous solvents with purity > 99 % from Sigma-Aldrich Pte Ltd. Crystal violet citrate (Sigma), Sodium (Sigma) and Hydrogen tetrachloroaurate (III) trihydrate (HAuCl₄.3H₂O) (99.9%, Aldrich) were used as received. Point Probe Plus silicon tips for imaging in tapping mode with atomic force microscopy were purchased from Nanosensors (Neuchatel, Switzerland).

2.2. Methods

The quartz wafers were diced into chips of size 10 mm x 10 mm. They were cleaned in acetone and 2-propanol for 10 min and finally exposed to UV/Ozone to remove organic contaminants (SAMCO UV-1, SAMCO Inc., Kyoto, Japan) for 10 minutes. The array of Au nanoparticles (AuNPs) was fabricated as described elsewhere.²⁶ In short, reverse micelle of polystyrene-*block*-poly(2-vinylpyridine) (PS-*b*-PVP) of 0.5 % (w/w) concentration in *m*-xylene were self-assembled on the

guartz surface by spin-coating where PVP forms the core and PS forms the continuous layer. This resulted in the regular arrangement of the polymer features with pitch of ~75 nm. This was followed by immersion of the polymer coated substrates in aqueous suspension of citrate ion stabilized presynthesized AuNPs for 2 hours. This initiates the directed electrostatic self-assembly of the negatively charged AuNPs on to the positively charged pyridyl units of the PVP core reverse micelles. The polymer template was then removed by exposing them to O₂ plasma reactive ion etch (Oxford Plasmalab 100, Oxfordshire, UK) for 10 min (30 W, 65 mTorr, 20 sccm O₂) to complete the nanocluster array fabrication. The AuNP colloidal solution was synthesized according to a procedure reported in the literature.³³ The pattern geometry such as the mean heights, width and pitch of the arrays were characterized using tapping mode AFM (Dimension Icon, Bruker Corporation, CA, USA). The plan view of the gold nanoparticles dispensed on a Cu grid was obtained using transmission electron microscopy performed using Philips CM300 TEM operating at 300 kV. The gold nano cluster arrays were then transferred onto the adhesive coated Kapton tape by sticking the latter to the quartz chip followed by gentle pressing and peeling off. The SERS spectra were recorded using a uRaman system (Technospex, Singapore) equipped with a Nikon microscope (Nikon Ci-L), a TE cooled CCD detector, and an excitation laser of 785 nm. The laser was coupled through a 20x objective with numerical aperture of 0.7 for exciting the sample as well as collecting the Raman signals. Prior to each Raman experiment, the instrument was calibrated using the Raman signal from a silicon standard centered at 521 cm⁻¹. The analyte solution was dispensed onto the SERS substrate and a glass coverslip is placed over it. In order to confirm the consistency of SERStape, measurements were performed at 8-10 random locations on each sample. The laser power was set at 38.7 mW and acquisition time of 5 s was kept constant for all experiments.

3. Results and discussion

The block-copolymer self-assembly process is a versatile bottom-up approach capable of achieving patterned arrays at the wafer-level. A detailed illustration of the self-assembly process towards the fabrication of flexible substrates starting from gold nanoclusters on flat chips is shown in the schematic (Figure 1). The PVP forms the core and is responsible for pattern formation while the PS block of the BCP forms a thin continuous layer. The latter prevents any random deposition when the self-assembled patterns are introduced to a solution of negatively charges pre-formed gold nanoparticles. The PVP core that contains the pyridyl units becomes positively charged upon protonation and selectively guides the electrostatic selfassembly of the AuNPs (pH 5.8) onto its surface thus forming islets of AuNP clusters.¹⁷ After the selective deposition, the BCP template is removed by exposing the arrays to oxygen plasma etch for 10 min resulting in gold nanocluster arrays. Figure 2 shows the as spin-coated BCP arrays of PS-b-PVP with a height of ~23 nm and diameter of ~55 nm that



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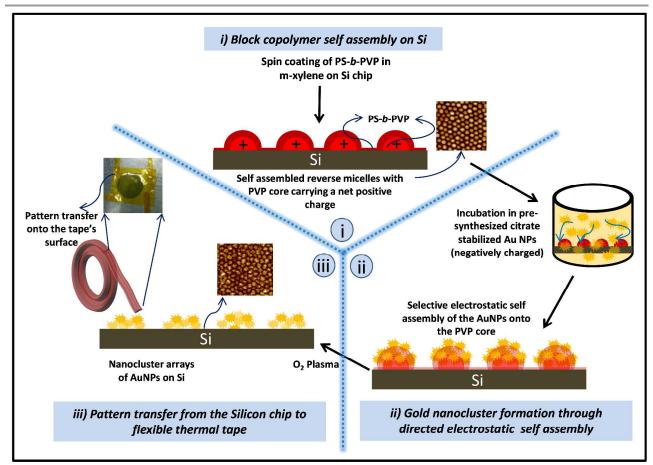


Figure 1. Schematic depicting the process route for the fabrication of flexible SERS-tapes starting from the flat chips. i) The self-assembled PS-*b*-PVP on quartz chips enabled through spin coating offers the template for subsequent ii) selective deposition of gold nanoparticles from the pre-formed negatively charged citrate stabilized aqueous colloid. Upon exposure to oxygen plasma reactive etch, iii) the polymer template is removed and thereby resulting in gold nanocluster arrays which was then transferred to the tape through a 'stick and peel' technique.

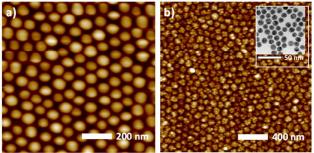
acted as the template to guide the electrostatic self-assembly of pre-formed AuNPs of size 11.5 nm (inset in **Figure 2b**) in order to form the gold nanocluster arrays. A histogram showing the distribution in the size of the nanoparticles can be found in **Figure S1** of the supporting info.

We have reported the SERS performances of the resulting arrays on conventional rigid substrates for concentrations up to picoMolar.^{17, 34} Nevertheless, the need to have SERS substrates that allow more versatility for practical applications and on curved non-planar surfaces where flat chips fail to establish their utility calls for the need to have flexible substrates. The latter remain highly useful for specific tasks that involve forensic analysis, picking chemicals from surfaces

inaccessible to flat SERS chips and in food processing industry to check for contamination. A Kapton tape is then attached to the flat chip on which the AuNP clusters were formed and pressed down gently. The tape was eventually peeled that results in the transfer of the entire SERS-active assembly from the chip to the tape. Quartz, being transparent, had been the support on which the SERS-sensitive layers were built in order to demonstrate the complete transfer of the SERS-active components onto the tape without leaving behind any residue, although, Si can also be employed. An AFM performed on the quartz chip after the pattern transformation shows no signs of residual patterns of gold and with the surface exhibiting a roughness of < 1 nm (**Figure S2** of the supporting info).

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75 nm and diameter 55 nm and b) the resulting gold nanocluster arrays after template removal. The inset shows the TEM image of the pre-formed AuNPs used in the nanocluster fabrication.

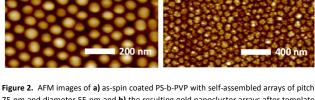
The 'stick and peel' technique as depicted in Figure 3A to transfer the gold nanocluster patterns from the chip to the tape overcome several issues that are associated with direct fabrication of metal cluster patterns on tape. The tape being not flat cannot be used as a substrate for spin coating. Furthermore, the micelles from the solution did not selfassemble, owing to the incompatibility in the surface energy of the tape to the apolar solvent. On the other hand, the 'stick and peel' technique was ensured complete pattern-transfer as evident from the clean quartz chip after transfer (Figure S2). More importantly, the gold nanoclusters was preserved after transferring to the tape. This was confirmed by characterizing the optical properties of the nano cluster arrays using transmittance measurements performed with a micro spectrophotometer (CRAIC Technologies, CA, US) that enabled measurement areas of 23 µm x 23 µm. This provides more qualitative information on the pattern integrity at a highly localized space on the substrate than a scanning electron microscopy (SEM) which often burns the polymer. Moreover, a coating of the SERS-active tape with a thin metal layer to counter the charging in SEM affects the observation of the actual pattern. Nevertheless, an attempt to image the gold nanoclusters (without additional coating) on the tape was made using low energy SEM that revealed the nanoclusters were indeed intact after transfer (Figure S3). It is reported earlier, that the resonance wavelength is known to be sensitive towards the arrangement and packing density of the AuNPs.¹⁷ The transmittance spectra for both the SERS-chip and tape exhibit a single broad peak around 730 nm (Figure 3B), with no evidence of shift associated with any plausible change in the pattern-arrangement. The 'stick and peel' method provides a route to precisely control AuNPs arrangement on flexible substrates to yield high SERS enhancement; this is not easily achievable by direct fabrication.

While flexible SERS substrates are advantageous in terms of its utility in real world applications, they usually suffer in terms of their robustness, compared to traditional support like silicon and glass. On the other hand, Kapton® polyimide tape with silicon adhesive as the support material is known to offer good chemical, mechanical and thermal stability. Chemically stability

is vital as SERS substrates are exposed to harsh and corrosive solvent that contains the analyte of interest. A support that is susceptible to solvent exposure will result in destruction of plasmonic hot spots and losing its sensitivity for detection. The chemical stability of our SERS-tape was evaluated by immersion of the entire substrate into various solvents, i.e. water, hexane (apolar) and ethanol (polar) for 15 min. There was no observable change in the plasmonic resonance wavelength of the gold nano clusters after immersion, suggesting that the 1) the tape is able to withstand the treatment in solvents, and 2) the adhesive layer on the tape is stable and maintained the arrangement of the gold nano clusters (Figure 3C).

Temperature stability of the support is necessary as the SERS substrates can be exposed to high temperature which arise when the laser is focused at the same spot for prolong duration. Paper based SERS substrates tend to burn easily and result in fluctuation in Raman intensity, making quantification studies difficult. This issue was not observed on our SERS-tape as the Kapton support is stable up to 400°C. In addition, the SERS-tape has low background signal, which facilitates ultralow concentration detection (Figure S4).

An effective SERS substrate needs to have shelf time stability for its high SERS enhancement. Degradation in SERS enhancement may arise due to oxidation or change in the physical arrangement of the metallic nanostructures or deposition of contaminants from the ambient environment onto the substrate which prevents absorption of the analyte molecules. Any of these occurrences will result in change in the optical properties of the gold nano clusters, which will be readily picked up by micro spectrophotometer measurements. The prepared SERS-tapes were kept under normal laboratory storage conditions for 1 month and the transmission spectra were collected at various time points. There was no observable change in the spectra on all occasions over a period of one month suggesting that there was no change in the adhesive characteristics and variation in the charges on the tape surface, which kept the gold nano clusters intact (Figure 3D).



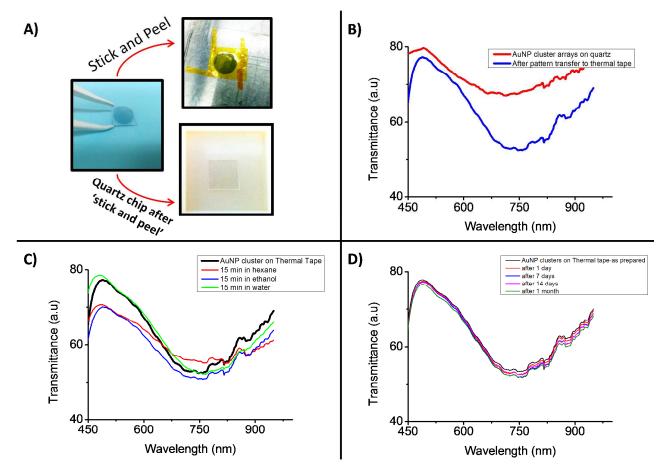


Figure 3. A) Photographs of the SERS-quartz chip and the SERS-tape. The picture of the quartz chip after 'stick and peel' presents no residue after transfer. B) Transmission spectra of the quartz chip and after transfer to the tape demonstrating that the arrangement of gold nano clusters is preserved. C) Transmission spectra to demonstrate the structural integrity and stability of the SERS-tape after immersion in various solvents. D) Transmission spectra to demonstrate stability in shelf life of gold nano clusters on SERS-tape

The SERS-tapes were assessed for its detection sensitivity using, crystal violet (CV), a test analyte that physisorbed to the substrate via non-covalent interaction. CV is non-resonant with the excitation wavelength of 785 nm. The Raman characteristics bands of CV were clearly shown for all concentrations from 1 mM down to 1 µM (Figure 4A). A control experiment with bare thermal tape (no gold nanoclusters) showed no enhancement. As the polyimide based Kapton tape has excellent optical transparency, we hypothesis that Raman signal can be collected by illuminating the laser from the backside of the tape. The SERS intensities collected through the backside for the same concentration of CV is shown in Figure 4B, the intensities is very similar to those obtain by excitation from the front side. This suggest that the SERS-tape is indeed highly transparent to light around 785 nm and can be used for direct collection and analysis of analyte on irregular surface by pasting the SERS-tape over it. This is especially useful for detection and quantification of contaminants on the surface of a arbitrary shaped object. In addition, SERS-tape has a secondary function of serving as an

adhesive, which facilitates easy attachment to the object of interest.

To demonstrate this, a curved (non-planar) object, viz., a rod of diameter ~8mm was locally contaminated with 50 µM CV in water (Figure 4C). The SERS tape was then wrapped around the object, with the plasmonic gold structures in contact with the analyte and Raman signal was collected by excitation of plasmons from the back side mode. The most intense peak of CV at 1172 cm⁻¹ was used for comparison of the signal obtained at different concentration. The enhanced intensity fell in the region between the intensities observed for 100 μ M and 10 µM concentrations of CV (Figure 4D). Although, many reports have demonstrated the ability to detect trace quantities of analytes, they suffer from substrate incompatibility towards harsh solvents that corrode the supporting substrates (paper based substrates), and thereby limiting the substrate's range of applications.²³ It is thus clear that the SERS-tape is versatile in its utility, stable and promises huge potential for large scale fabrication. Since, the

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methodology starts with the BCP self-assembly, the process is feasible at a wafer level with assured plausibility in the pattern-transfer of the plasmonic features to a tape of similar dimension. As an extension, the flexible SERS-tape was tested for its performance using 1-thionapthol in n-hexane and potassium dichromate in water and still can be extended to a

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range of other chemicals such as melamine and carbaryl (pesticide) for detecting adulteration or contamination in food products.

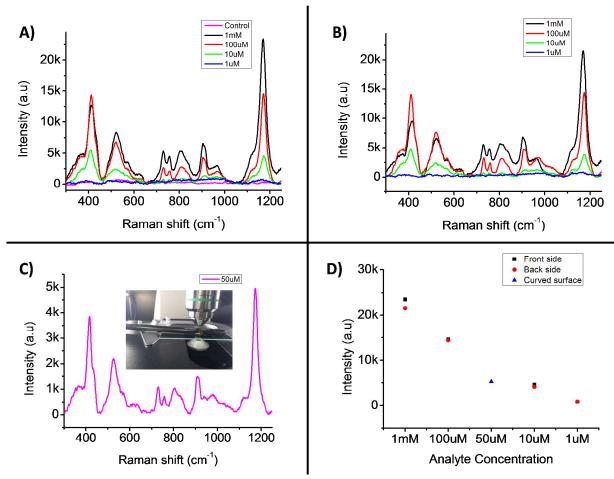


Figure 4. Detection sensitivity of the SERS-tape with **A**) the laser shining directly on the front side of SERS active surface using crystal violet (CV) as analyte, **B**) laser shining from the back side of the tape. **C**) A 50 μM solution of CV was stained around a 8 mm diameter rod which was picked by the SERS-tape that wrapped around it and raman signal was collected by back side mode. **D**) Plot showing trend in the intensity of the SERS signals corresponding to the most intense peak of CV as a function of concentrations of CV collected via the front side and back side mode.

4. Conclusions

Recent advances in portable Raman instrumentation have dramatically increased their application for on-field analysis. With this development, we can no longer be tied to flat and rigid SERS substrates. What is required are flexible and transparent SERS substrates which can conform onto irregular shaped sample and one that facilitates direct in-situ measurement of the analyte without an additional extraction step. In this paper, we report the fabrication of gold nanoclusters arrays on flexible tapes for SERS applications. The SERS-tape does not succumb easily to harsh environments by virtue of the substrate property and nanoclusters integrity and thereby enabling its usage to a wide range of operating conditions. The gold nanoclusters created on Kapton tape extend their adaptability to wide range of analytes in various polar and apolar solvents. Moreover, with wafer-level processing, large area SERS-tapes are easy to fabricate with the simple 'stick and peel' technique (yield dependent on the wafer size) to generate large quantities of flexible SERS-tapes within short period of time. This offers excellent opportunities to pursue further on the development of innovative SERS substrates for laboratory and industrial applications on a large scale.

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References

- 1 B. Liu, P. Zhou, X. Liu, X. Sun, H. Li and M. Lin, *Food Bioprocess Technol*, 2013, **6**, 710-718.
- 2 F. Deiss, N. Sojic, D. J. White and P. R. Stoddart, *Analytical and bioanalytical chemistry*, 2010, **396**, 53-71.
- 3 G. F. S. Andrade, M. Fan and A. G. Brolo, *Biosensors and Bioelectronics*, 2010, **25**, 2270-2275.
- 4 M. Fan, G. F. S. Andrade and A. G. Brolo, *Analytica Chimica Acta*, 2011, **693**, 7-25.
- 5 L. Scarabelli, M. Coronado-Puchau, J. J. Giner-Casares, J. Langer and L. M. Liz-Marzán, ACS Nano, 2014, 8, 5833-5842.
- 6 P. Guo, D. Sikdar, X. Huang, K. J. Si, W. Xiong, S. Gong, L. W. Yap, M. Premaratne and W. Cheng, *Nanoscale*, 2015, 7, 2862-2868.
- 7 M. R. Langille, M. L. Personick, J. Zhang and C. A. Mirkin, Journal of the American Chemical Society, 2012, 134, 14542-14554.
- 8 A. J. Chung, Y. S. Huh and D. Erickson, *Nanoscale*, 2011, **3**, 2903-2908.
- 9 X.-M. Lin, Y. Cui, Y.-H. Xu, B. Ren and Z.-Q. Tian, Analytical and Bioanalytical Chemistry, 2009, **394**, 1729-1745.
- 10 Y. Weisheng, W. Zhihong, Y. Yang, C. Longqing, S. Ahad, W. Kimchong and W. Xianbin, *Journal of Micromechanics and Microengineering*, 2012, **22**, 125007.
- 11 W.-Y. Zhang, X.-Z. Xiao, C. Lv, J. Zhao, G. Wang, X. Gu, R. Zhang, B.-B. Xu, D.-D. Zhang, A.-W. Li, Y.-L. Zhang and H.-B. Sun, *Macromol. Res.*, 2013, **21**, 306-310.
- 12 T. Siegfried, M. Kind, A. Terfort, O. J. F. Martin, M. Zharnikov, N. Ballav and H. Sigg, *Journal of Raman Spectroscopy*, 2013, 44, 170-175.
- J. Li, C. Chen, H. Jans, X. Xu, N. Verellen, I. Vos, Y. Okumura,
 V. V. Moshchalkov, L. Lagae and P. Van Dorpe, *Nanoscale*, 2014, 6, 12391-12396.
- 14 A. Martin, J. J. Wang and D. Iacopino, *RSC Advances*, 2014, 4, 20038-20043.
- 15 L.-B. Zhong, J. Yin, Y.-M. Zheng, Q. Liu, X.-X. Cheng and F.-H. Luo, *Analytical Chemistry*, 2014, **86**, 6262-6267.
- 16 L. Polavarapu and L. M. Liz-Marzan, *Physical Chemistry Chemical Physics*, 2013, **15**, 5288-5300.
- 17 F. L. Yap, P. Thoniyot, S. Krishnan and S. Krishnamoorthy, ACS Nano, 2012, 6, 2056-2070.
- 18 J. F. Betz, W. W. Yu, Y. Cheng, I. M. White and G. W. Rubloff, *Physical Chemistry Chemical Physics*, 2014, 16, 2224-2239.
- 19 D. He, B. Hu, Q.-F. Yao, K. Wang and S.-H. Yu, ACS Nano, 2009, 3, 3993-4002.
- 20 P. M. Fierro-Mercado, Hern, #xe1 and S. P. ndez-Rivera, International Journal of Spectroscopy, 2012, 2012, 7.
- 21 M. Fan, Z. Zhang, J. Hu, F. Cheng, C. Wang, C. Tang, J. Lin, A. G. Brolo and H. Zhan, *Materials Letters*, 2014, **133**, 57-59.
- 22 C. H. Lee, L. Tian and S. Singamaneni, ACS Applied Materials & Interfaces, 2010, 2, 3429-3435.
- 23 W. W. Yu and I. M. White, Analyst, 2013, 138, 1020-1025.
- 24 C. Zhenyi, D. Zhangmin, C. Na, L. Shupeng, P. Fufei, L. Bo and W. Tingyun, *Photonics Technology Letters, IEEE*, 2014, 26, 777-780.

- 25 H. Chen, F. Tian, J. Chi, J. Kanka and H. Du, *Opt. Lett.*, 2014, **39**, 5822-5825.
- 26 V. Suresh, S. Madapusi and S. Krishnamoorthy, ACS Nano, 2013, 7, 7513-7523.
- 27 S. Park, D. H. Lee, J. Xu, B. Kim, S. W. Hong, U. Jeong, T. Xu and T. P. Russell, *Science*, 2009, **323**, 1030-1033.
- 28 V. Suresh, M. S. Huang, M. P. Srinivasan and S. Krishnamoorthy, *J. Mater. Chem.*, 2012, **22**, 21871-21877.
- 29 V. Suresh, M. S. Huang, M. P. Srinivasan and S. Krishnamoorthy, *ACS Appl. Mater. Interfaces*, 2013, **5**, 5727-5732.
- 30 V. Suresh, D. Y. Kusuma, P. S. Lee, F. L. Yap, M. P. Srinivasan and S. Krishnamoorthy, ACS Applied Materials & Interfaces, 2015, 7, 279-286.
- 31 S. Krishnamoorthy, K. K. Manipaddy and F. L. Yap, *Adv. Funct. Mater.*, 2011, **21**, 1102-1112.
- 32 F. L. Yap and S. Krishnamoorthy, J. Mater. Chem., 2010, 20, 10211-10216.
- 33 K. C. Grabar, R. G. Freeman, M. B. Hommer and M. J. Natan, Anal. Chem., 1995, 67, 735-743.
- 34 S. Dinda, F. L. Yap, V. Suresh, R. K. Gupta, D. Das and S. Krishnamoorthy, Aust. J. Chem., 2013, 66, 1034-1038.