Chemical Science



EDGE ARTICLE

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Cite this: Chem. Sci., 2020, 11, 12047

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photosensitizers for metal-organic photodynamic therapy†

Bis[pyrrolyl Ru(11)] triads: a new class of

A new family of ten dinuclear Ru(II) complexes based on the bis[pyrrolyl Ru(III)] triad scaffold, where two Ru(bpy)₂ centers are separated by a variety of organic linkers, was prepared to evaluate the influence of the organic chromophore on the spectroscopic and in vitro photodynamic therapy (PDT) properties of the compounds. The bis[pyrrolyl Ru(ii)] triads absorbed strongly throughout the visible region, with several members having molar extinction coefficients (ε) $\geq 10^4$ at 600-620 nm and longer. Phosphorescence quantum yields (Φ_n) were generally less than 0.1% and in some cases undetectable. The singlet oxygen quantum yields (Φ_{Δ}) ranged from 5% to 77% and generally correlated with their photocytotoxicities toward human leukemia (HL-60) cells regardless of the wavelength of light used. Dark cytotoxicities varied ten-fold, with EC₅₀ values in the range of 10-100 μM and phototherapeutic indices (PIs) as large as 5400 and 260 with broadband visible (28 J cm $^{-2}$, 7.8 mW cm $^{-2}$) and 625 nm red (100 J cm⁻², 42 mW cm⁻²) light, respectively. The bis[pyrrolyl Ru(III)] triad with a pyrenyl linker (5h) was especially potent, with an EC $_{50}$ value of 1 nM and PI > 27 000 with visible light and subnanomolar activity with 625 nm light (100 J cm⁻², 28 mW cm⁻²). The lead compound 5h was also tested in a tumor spheroid assay using the HL60 cell line and exhibited greater photocytotoxicity in this more resistant model (EC₅₀ = 60 nM and PI > 1200 with 625 nm light) despite a lower dark cytotoxicity. The in vitro PDT effects of 5h extended to bacteria, where submicromolar EC $_{50}$ values and PIs >300 against S. mutans and S. aureus were obtained with visible light. This activity was attenuated with 625 nm red light, but PIs were still near 50. The ligand-localized ${}^3\pi\pi^*$ state contributed by the pyrenyl linker of 5h likely plays a key role in its phototoxic effects toward cancer cells and bacteria.

Received 15th August 2020 Accepted 6th October 2020

DOI: 10.1039/d0sc04500d

rsc.li/chemical-science

1. Introduction

Light-responsive prodrugs are the basis for selectively targeting unwanted cells and tissue in photodynamic therapy (PDT). Activation of an otherwise nontoxic photosensitizer (PS) produces cytotoxic singlet oxygen ($^{1}O_{2}$) and other reactive oxygen species (ROS) in regions where the PS, light, and oxygen overlap spatiotemporally, $^{1-3}$ thus confining toxicity to diseased

tissue while sparing healthy tissue. The antitumor effects of PDT result from destruction of primary tumors and tumor vasculature, but can also include a systemic immunological response.⁴⁻¹² Photofrin, a mixture of oligomeric tetrapyrroles, remains arguably the most utilized PS for PDT.¹²⁻¹⁵ However, a variety of second- and third-generation derivatives, including metallated tetrapyrroles, that seek to improve upon the properties of earlier PSs have gained attention and (in some cases) approval in certain countries.^{16,17}

Metal complexes that are not simply metallated tetrapyrroles are particularly intriguing as PSs for PDT, ^{18,19} and there are numerous reports highlighting their rich photophysical and photochemical properties. ²⁰ Their modular architectures can be exploited to produce a variety of energetically accessible excited state configurations: metal-to-ligand charge transfer (MLCT), ²¹ metal centered (MC), ²²⁻²⁴ ligand centered (LC) or intraligand (IL), ²⁵⁻²⁷ intraligand charge transfer (ILCT), ³¹⁻³³ ligand-to-metal charge transfer (LMCT), ³⁴ and metal-to-metal charge transfer (MMCT) in the case of multimetallic systems. ³⁵⁻³⁸ Some of these excited states

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[†] Electronic supplementary information (ESI) available: Additional synthetic procedures for the synthesis of pyrrole **1a** and aryl dibromides **i**, **j** and **k**. Figures giving ¹H and ¹³C NMR spectra and UV/vis absorption spectra for all bis(pyrrole)s (2), ligands (3) and bis(ruthenium) complex salts (4). See DOI: 10.1039/d0sc04500d

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(and combinations thereof) may undergo the type I and II photoprocesses that define PDT or they may exert phototoxic effects *via* alternate mechanisms that do not involve oxygen. The oxygen-independent pathways, which includes stoichiometric photodissociation of ligands,^{22,24,39-45} have been collectively grouped as photochemotherapy (PCT) although no PCT agents have been approved for cancer therapy to date.^{16,46}

Through our search for PSs that produce phototoxic effects in hypoxia via catalytic photosensitization pathways, we have found that the best features of both organic and inorganic PSs can be combined to produce hybrid systems, and the resulting metal-organic dyads exhibit unprecedented photocytotoxicities and phototherapeutic indices (PIs).26,47,48 Organic chromophores, either contiguously fused or tethered to coordinating diimine ligands, serve as excellent collection points for excitation energy from singlet excited states provided their localized ³IL states are energetically accessible through equilibration or relaxation. Organic triplets offer a unique means of slowing T → S intersystem crossing (ISC) in metal complexes, while the metal facilitates efficient formation of these triplet excited states and the possibility of oxygen-independent photoreactivity. Pure ³IL states that are lower in energy than the lowest lying ³MLCT state(s) tend to possess exceptionally long lifetimes (>20 µs) and proved very effective for in vitro PDT. 26,47,49-52

From our extensive work in this area, we have found that organic triplets having charge transfer character (3 ILCT) contributed by α -oligothienyl groups in certain systems are particularly photoreactive and make excellent PDT agents. ${}^{16,48,53-61}$ Our TLD1433 is one example, which is a bisheteroleptic Ru(II) complex based on the α -terthienylappended imidazo[4,5-f][1,10]phenanthroline (IP-3T) ligand that generates 1O_2 with almost unity efficiency. ${}^{16,17,48,62-66}$ TLD1433 is the first Ru(II) complex to enter a human clinical trial and is being evaluated in a Phase 2 clinical trial for treating non-muscle invasive bladder cancer with PDT (Clinicaltrials.gov identifier: NCT03945162). 16,17,66

Our ongoing interest in exploring the photoreactivity of Ru(II) metal-organic systems, including TLD1433, inspired the present study. Herein, we explore the bis[Ru(II)-pyrrolide] scaffold, a metal-organic-metal triad, to push the envelope for achieving unprecedented in vitro PDT potency with 3IL excited states. This construct simultaneously satisfies three criteria: (i) low energy singlet and triplet MLCT states, (ii) utilization of two metal centers to funnel energy to an organic triplet, and (iii) incorporation of an organic chromophore with a triplet excited state of suitable energy and lifetime. Previously, we have shown that 2-formyl and 2-keto pyrroles can replace one of the 2,2'bipyridyl (bpy) ligands in [Ru(bpy)₃]²⁺ to form stable complexes under ambient conditions with MLCT states shifted as much as 1.52 eV relative to the parent complex,67 building on work involving 2-formylpyridyl ligands that convert to their hydrate form upon complexation.68 In our model mononuclear 2-formylpyrrolide complexes, continuous absorption out to 600 nm was achieved without the need for sterically-demanding diimines such as 2,2'-biquinoline (biq) that are known to lower the

energies of both MLCT and MC states, leading to red-shifted absorption, but also photodissociation.

The small, bidentate pyrrolide ligand forms strong N- σ (η^1) bonds to Ru(II), lowering the energy of MLCT states without promoting ligand loss from dissociative 3MC states. Conversion of this 2-formyl pyrrole ligand into its symmetric bis(formyl-pyrrole) counterpart with a central organic chromophore linker and coordination of the termini to Ru(II) diimine units was expected to result in complexes with a larger percentage of accessible 3IL triplets. Herein we report the synthesis and characterization of a family of bis[Ru(II)-pyrrolide] triads that differ in the identity of the organic chromophore used as the central linker. The influence of this unit on the photobiological activities within this class of compounds is examined in detail, and the potent *in vitro* PDT effects discussed.

2. Experimental procedures

2.1 Materials

All chemicals and reagents were purchased from commercial sources and were used as received, unless otherwise noted. Ethyl acetate, hexanes and dichloromethane were obtained crude and purified *via* distillation, under air and at 1 atm pressure, before use. Reagent-grade tetrahydrofuran (THF), ethylene glycol, isopropanol (IPA) and acetone were employed where stated. Anhydrous dichloromethane and dimethylformamide (DMF) were purchased from EMD Chemicals and Sigma Aldrich, respectively. All glassware was oven dried and purged with inert gas before use. Gravity column chromatography was performed using 230–400 mesh Silicycle ultrapure silica gel or 150-mesh Brockman III activated neutral aluminum oxide. TLC was performed on silica gel or aluminum oxide plates and visualized using UV light (254 and/or 365 nm) and/or developed with vanillin stain.

Characterized fetal bovine serum (FBS) and Iscove's Modified Dulbecco's Medium (IMDM) supplemented with 4 mM L-glutamine were purchased from Fisher Scientific. Human promyelocytic leukemia cells (HL-60), *Streptococcus mutans*, and *Streptococcus aureus* were purchased from American Type Culture Collection (ATCC) through Cedarlane (Burlington, ON). Prior to use, FBS was aliquoted in 40 mL volumes, heat inactivated for 30 min at 55 °C, and stored at -20 °C. Water for biological experiments was deionized to a resistivity of 18 M Ω cm using a Barnstead filtration system.

2.2 Instrumentation

NMR spectra were recorded using a 500 MHz spectrometer. All 1 H and 13 C NMR chemical shifts are expressed in parts per million (ppm) using the solvent signal [CDCl₃ (1 H 7.26 ppm; 13 C 77.16 ppm); DMSO- d_6 (1 H 2.50 ppm; 13 C 39.52 ppm); THF- d_8 (1 H 1.73, 3.58 ppm; 13 C 25.4, 67.6 ppm); CD₂Cl₂ (1 H 5.32 ppm; 13 C 53.8 ppm)] as the internal reference. Splitting patterns are indicated as follows: br, broad; s, singlet; d, doublet; t, triplet; at, apparent triplet; q, quartet; m, multiplet; sep, septet. All coupling constants (f) are reported in Hertz (Hz). Ultravioletvisible spectra were recorded using a Varian Cary 100 Bio

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spectrophotometer. Mass spectra were recorded using ion trap (ESI or APCI) instruments. Microwave-promoted reactions were carried out using a Biotage Initiator 8 microwave with 0-400 W power at 2.45 GHz. Melting points are uncorrected.

2.3 Synthesis and characterization

2.3.1 General procedures

General procedure for the synthesis of bis(pyrrole)s (2) by Heck reaction (GP1). Palladium(II) acetate (1 mol%) and 2,4-pentanedione (2 mol%) were added to a solution of aryl dibromide (0.35 mmol, 1 equiv.) in anhydrous DMF (2.0 mL) at room temperature under argon, and stirred for 10 minutes. 2-Vinyl-N-Boc pyrrole (1a) (0.88 mmol, 2.5 equiv.) was then added as an oil, followed by potassium carbonate (0.7 mmol, 2 equiv.) as a solid in one portion, and the flask was sealed with a glass stopper before heating to 130 °C (Caution: always use a blast shield when heating a sealed system), using a sand bath covered with aluminum foil, with stirring for 6 hours. After cooling slightly, the reaction mixture was poured into ice-water (40 mL), neutralized with a few drops of 1 M HCl and refrigerated (4 °C) overnight. The resulting precipitate was collected using a Millipore filtration apparatus and then dried in a vacuum oven to give the crude product, which was subsequently washed with 0-30% diethyl ether/hexanes on a Millipore filter to give the desired bis(pyrrole) without the need for further purification, unless otherwise stated.

General procedure for the synthesis of bis(pyrrole)s (2) by Suzuki reaction (GP2). A solution of aryl dibromide (0.15 mmol, 1 equiv.) and 1-Boc-pyrrole-2-boronic acid (1b) (0.45 mmol, 3 equiv.) in anhydrous DMF (3 mL) was sparged with nitrogen gas for 10 minutes. Tetrakis(triphenylphosphine)palladium(0) (0.015 mmol, 0.1 equiv.) and potassium carbonate (0.60 mmol, 4 equiv.) were then added with stirring, and the solution was sparged with nitrogen for a further 5 minutes before the flask was sealed and heated to 110 °C for 24 hours. The reaction mixture was then cooled to room temperature and separated between dichloromethane (50 mL) and water (50 mL). The aqueous phase was extracted with dichloromethane (2 \times 50 mL) and the combined organic extracts were washed with water (4 \times 100 mL) and brine (100 mL), dried over anhydrous sodium sulfate, and concentrated to give the crude product which was purified using column chromatography on silica gel.

General procedure for the synthesis of bis(formylpyrrole)s (3) using Vilsmeier-Haack reaction (GP3). The desired bis(pyrrole) (2) (0.2 mmol, 1 equiv.) was dissolved in anhydrous DMF (4 mL) with stirring under nitrogen, and the solution was cooled to 0 °C in an ice bath. Phosphorous oxychloride (0.44 mmol, 2.2 equiv.), was then added drop-wise and the reaction mixture was warmed to 60 °C with stirring for 1.5 hours. After cooling to room temperature, 5% (w/v) aqueous potassium carbonate solution $(\sim 3 \text{ mL})$ was added slowly until the solution became basic $(\sim pH)$ 8, pH paper). The reaction mixture was then heated to 80 °C with stirring for 2 hours, before being poured into ice-water to precipitate the product which was collected using a Millipore filtration apparatus. The product was then dried in a vacuum oven and finally washed with 50-100% diethyl ether/hexanes.

General procedure for the synthesis of bis(ruthenium(11))hexafluorophosphate complex salts (4) (GP4). Triethylamine (0.24 mmol, 8 equiv.) was added to a suspension of dipyrrolic ligand (3) (0.031 mmol, 1.03 equiv.) and *cis*-bis-(2,2'-bipyridine) dichlororuthenium(II)dihydrate (0.06 mmol, 2 equiv.) in ethylene glycol (2.0 mL) in a Biotage microwave vial (2-5 mL capacity). The vial was then sealed using a manual cap crimper and placed in the microwave reactor, where it was heated at 125 °C for 80 minutes, at a maximum of 400 W power. After cooling, the reaction mixture was poured into a solution of ammonium hexafluorophosphate (0.45 mmol, 15 equiv.) in water (20 mL) and left to stand at room temperature overnight. The solution was then extracted thoroughly with dichloromethane (4 × 20 mL). The combined organic extracts were washed with brine (50 mL), dried over anhydrous sodium sulfate and concentrated to give the crude product, which was purified using column chromatography on silica gel (0-8% IPA/ dichloromethane) and/or neutral alumina (0-8% methanol/ dichloromethane).

General procedure for the conversion of bis(ruthenium(11))hexafluorophosphate complex salts to chloride salts (5) (GP5). Tetrabutylammonium chloride monohydrate (0.25 mmol, 20 equiv.) was added to a solution of the bis(ruthenium)hexafluorophosphate salt (4) (0.0125 mmol, 1.0 equiv.) in acetone (12 mL, 1 mM) with stirring at room temperature for 15 minutes. The desired chloride salt was generally observed to form as a precipitate during this time (unless otherwise stated), which was collected using Millipore filtration and washed with 30% acetone/hexanes before drying in a vacuum oven.

2.3.2 Experimental data

(E)-2-Styryl-1H-pyrrole (2a).69 Compound 2a was synthesized from 2-vinyl-N-Boc pyrrole (1a, 1.3 equiv.) and bromobenzene (a) using GP1 and a reaction time of 3 h. After cooling to room temperature, the reaction mixture was separated between diethyl ether (30 mL) and water (20 mL). The aqueous phase was extracted with diethyl ether (4 × 20 mL) and the combined organic extracts were washed with water (100 mL) and brine (100 mL), dried over anhydrous magnesium sulfate and concentrated in vacuo. The crude product was purified using column chromatography on silica gel eluting with 15% ethyl acetate/hexanes to give the title compound (34 mg, 64% yield) as a pale yellow solid. Mp 110–115 °C. 1 H NMR (CDCl₃, 500 MHz) δ : 7.43 (d, 2H, J = 7.5 Hz, ArH), 7.33 (at, 2H, J = 7.8 Hz, ArH), 7.21 (t, 1H, J = 7.3 Hz, ArH), 6.98 (d, 1H, J = 16.5 Hz, ArH), 6.83-6.82(m, 1H), 6.67 (d, 2H, J = 16.5 Hz, ArH), 6.35–6.36 (m, 1H), 6.25 (aq, 1H, J = 3.0 Hz) ppm. ¹³C NMR (CDCl₃, 125 MHz) δ : 137.6, 130.9, 128.8, 127.1, 126.0, 123.5, 119.2, 119.1, 110.2, 109.3 ppm. LRMS: 170.1 (M + H) $^{+}$; HRMS calculated for C₁₂H₁₂N: 170.0964; found 170.0964.

1,4-Bis((E)-2-(1H-pyrrol-2-yl)vinyl)benzene (2b). Compound 2b was synthesized from 2-vinyl-N-Boc pyrrole (1a) and 1,4-dibromobenzene (b) using GP1 to give the title compound (95 mg, 86% yield) as a dark yellow solid. Mp/dp > 280 °C. 1 H NMR (THF- d_{8} , 500 MHz) δ : 10.27 (br s, 2H, NH), 7.34 (s, 4H, ArH), 6.97 (d, 2H, J =16.5 Hz, C=CH), 6.72-6.71 (m, 2H, PyH), 6.68 (d, 2H, J = 16.5 Hz, C=CH), 6.20-6.19 (m, 2H, PyH), 6.06-6.05 (m, 2H, PyH) ppm. ¹³C NMR (THF- d_8 , 125 MHz) δ : 137.5, 132.0, 126.6, 122.8, 120.0, 119.9, 109.9, 109.8 ppm. LRMS: 259.1 (M - H) $^-$; HRMS calculated for $C_{18}H_{15}N_2$: 259.1241; found 259.1238. $\varepsilon_{386~nm}=48~000$ (THF).

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4,4'-Bis((E)-2-(1H-pyrrol-2-yl)vinyl)-1,1'-biphenyl (2c). Compound 2c was synthesized from 2-vinyl-N-Boc pyrrole (1a) and 4,4'-dibromobiphenyl (c) using GP1. The crude product was washed with 1:1 diethyl ether: hexanes to give the title compound (95 mg, 86% yield) as a light brown solid. Mp/dp > 250 °C. ¹H NMR (DMSO- d_6 , 500 MHz) δ: 11.20 (br s, 2H, NH), 7.67 (d, 4H, J = 8.3 Hz, ArH), 7.52 (d, 4H, J = 8.3 Hz, ArH), 7.09 (d, 2H, J = 16.5 Hz, C=CH), 6.88 (d, 2H, J = 16.5 Hz, C=CH), 6.84 (br s, 2H, pyH), 6.28 (br s, 2H, PyH), 6.08 (br s, 2H, PyH) ppm. 13 C NMR (DMSO- d_6 , 125 MHz) δ: 137.5, 136.8, 130.4, 126.5, 126.0, 121.6, 119.9, 119.8, 109.3, 109.0 ppm. LRMS: 337.2 (M + H) $^+$; HRMS calculated for C₂₄H₂₁N₂: 337.1699; found 337.1688. $\varepsilon_{380 \text{ nm}}$ = 74 000 (THF).

2,6-Bis((E)-2-(1H-pyrrol-2-yl)vinyl)naphthalene (2d). Compound 2d was synthesized from 2-vinyl-N-Boc pyrrole (1a) and 2,6-dibromonaphthalene (d) using GP1 to give the title compound (95 mg, 97% yield) as a light brown solid. Mp/dp > 250 °C. ¹H NMR (DMSO- d_6 , 500 MHz) δ: 11.23 (br s, 2H, NH), 7.82 (d, 2H, J = 8.5 Hz, ArH), 7.76 (s, 2H, ArH), 7.69 (d, 2H, J = 8.5 Hz, ArH), 7.17 (d, 2H, J = 16.5 Hz, C=CH), 6.99 (d, 2H, J = 16.5 Hz, C=CH), 6.86 (dd, 2H, J = 2.5, 4.0 Hz, PyH), 6.30 (br s, 2H, PyH), 6.09 (dd, 2H, J = 2.5, 5.5 Hz, PyH) ppm. 13 C NMR (DMSO- d_6 , 125 MHz) δ: 135.0, 132.5, 130.5, 128.1, 124.5, 123.6, 122.2, 120.0, 119.9, 109.4, 109.0 ppm. LRMS: 311.2 (M + H) $^+$; HRMS (APCI) calculated for $C_{24}H_{21}N_2$: 311.1543; found 311.1528. $ε_{384}$ nm = 59 000 (THF).

4,7-Bis((E)-2-(1H-pyrrol-2-yl)vinyl)benzo[c][1,2,5]thiadiazole (2e). Compound 2e was synthesized from 2-vinyl-N-Boc pyrrole (1a) and 4,7-dibromobenzo[c]-1,2,5-thiadiazole (e) using GP1. After cooling to room temperature, the reaction mixture was separated between 1:2 THF: diethyl ether (30 mL) and water (20 mL). The aqueous phase was extracted with 1:2 THF: diethyl ether $(4 \times 20 \text{ mL})$ and the combined organic extracts were concentrated in vacuo. The crude product was purified using column chromatography on silica eluting with 30% ethyl acetate/hexanes to give the title compound (99 mg, 91% yield) as a red solid. Mp 200-205 °C. ¹H NMR (CDCl₃, 500 MHz) δ : 8.51 (br s, 2H, NH), 7.84 (d, 2H, J = 16.5 Hz, C=CH), 7.52 (s, 2H, ArH), 7.17 (d, 2H, J = 16.5 Hz, C=CH), 6.88 (dd, 2H, J = 2.5, 4.0 Hz, PyH, 6.51 (br s, 2H, PyH), 6.30 (dd, 2H, J = 2.5, 6.0 Hz, PyH) ppm. 13 C NMR (CDCl₃, 125 MHz) δ : 153.9, 131.5, 128.8, 125.9, 122.8, 120.1, 119.3, 110.6, 110.4 ppm. LRMS: 319.1 $(M + H)^+$; HRMS (APCI) calculated for $C_{18}H_{15}N_4S$: 319.1012; found 319.1000. $\varepsilon_{520 \text{ nm}} = 18\ 000$; $\varepsilon_{360 \text{ nm}} = 27\ 000$; $\varepsilon_{266 \text{ nm}} =$ 14 000 (THF).

9,10-Bis((E)-2-(1H-pyrrol-2-yl)vinyl)anthracene (2f). Compound 2f was synthesized from 2-vinyl-N-Boc pyrrole (1a) and 9,10'-dibromoanthracene (f) using GP1. The crude product was washed with 10% diethyl ether/hexanes to give the title compound (114 mg, 97% yield) as a light brown solid. Mp 215–220 °C. 1 H NMR (THF- d_8 , 500 MHz) δ : 10.60 (br s, 2H, NH), 8.44–8.42 (m, 4H, ArH), 7.56 (d, 2H, J = 16.5 Hz, C=CH), 7.43–7.41 (m, 4H, ArH), 6.84 (br s, 2H, PyH), 6.76 (d, 2H, J = 16.5 Hz, C=CH),

6.33 (br s, 2H, PyH), 6.15 (br s, 2H, PyH) ppm. ¹³C NMR (THF- d_8 , 125 MHz) δ : 133.4, 131.8, 130.6, 129.4, 127.3, 125.6, 120.2, 118.8, 110.1, 109.9 ppm. LRMS: 361.2 (M + H)⁺; HRMS (APCI) calculated for $C_{26}H_{21}N_2$: 361.1699; found 361.1688. $\varepsilon_{424~\rm nm} = 12~600$; $\varepsilon_{259~\rm nm} = 60~000$ (THF).

2,7-Bis((E)-2-(1H-pyrrol-2-yl)vinyl)-9H-fluorene (2g). Compound 2g was synthesized from 2-vinyl-N-Boc pyrrole (1a) and 2,7-dibromofluorene (g) using GP1 to give the title compound (108 mg, quantitative) as a yellow/brown solid. Mp/dp > 250 °C. ¹H NMR (THF- d_8 , 500 MHz) δ: 10.30 (br s, 2H, NH), 7.67 (d, 2H, J = 8.0 Hz, ArH), 7.61 (s, 2H, ArH), 7.39 (d, 2H, J = 8.0 Hz, ArH), 7.05 (d, 2H, J = 16.5 Hz, C=CH), 6.78 (d, 2H, J = 16.5 Hz, C=CH), 6.73-6.72 (m, 2H, PyH), 6.22 (br s, 2H, PyH), 6.07-6.06 (m, 2H, PyH), 3.87 (s, 2H, CH₂) ppm. ¹³C NMR (THF- d_8 , 125 MHz) δ: 144.8, 141.2, 137.8, 132.0, 125.7, 123.4, 122.5, 120.4, 120.0, 110.0, 109.8, 37.3 ppm (one signal missing). LRMS: 349.2 (M + H)⁺; HRMS (APCI) calculated for C₂₅H₂₁N₂: 349.1699; found 349.1694. $ε_{390 \text{ nm}}$ = 55 000 (THF).

1,6-Bis((E)-2-(1H-pyrrol-2-yl)vinyl)pyrene (2h). Compound 2h was synthesized from 2-vinyl-N-Boc pyrrole (1a) and 1,6-dibromopyrene (h) using GP1 to give the title compound (48 mg, quantitative) as a dark yellow/brown solid. Mp/dp > 250 °C. ¹H NMR (THF- d_8 , 500 MHz) δ: 10.56 (br s, 2H, NH), 8.48 (d, 2H, J = 8.5 Hz, ArH), 8.30 (d, 2H, J = 8.5 Hz, ArH), 8.10 (d, 2H, J = 9.0 Hz, ArH), 8.05 (d, 2H, J = 9.0 Hz, ArH), 7.87 (d, 2H, J = 16.0 Hz, C=CH), 7.29 (d, 2H, J = 16.0 Hz, C=CH), 6.83 (br s, 2H, PyH), 6.37 (br s, 2H, PyH), 6.14 (br s, 2H, PyH) ppm. 13 C NMR (THF- d_8 , 125 MHz) δ: 133.7, 132.6, 131.0, 129.2, 127.9, 126.7, 125.7, 123.4, 123.2, 123.1, 120.4, 119.6, 110.8, 110.1 ppm (one signal missing). LRMS: 385.2 (M + H) $^{+}$; HRMS calculated for C₂₈H₂₁N₂: 385.1699; found 385.1686. $\varepsilon_{433 \text{ nm}}$ = 37 000; $\varepsilon_{299 \text{ nm}}$ = 24 000 (THF).

4,7-Bis(4-((E)-2-(1H-pyrrol-2-yl)vinyl)phenyl)benzo[c][1,2,5]thiadiazole (2i). Compound 2i was synthesized from 2-vinyl-N-Boc pyrrole (1a) and 4,7-bis(4-bromophenyl)benzo[c][1,2,5]thiadiazole (i)⁷⁰ using GP1 to give the title compound (127 mg, quantitative) as a dark yellow/brown solid. Mp/dp > 250 °C. ¹H NMR (THF- d_8 , 500 MHz) δ: 10.37 (br s, 2H, NH), 8.07 (d, 4H, J = 8.5 Hz, ArH), 7.91 (s, 2H, ArH), 7.58 (d, 4H, J = 8.5 Hz, ArH), 7.13 (d, 2H, J = 16.5 Hz, C=CH), 6.81 (d, 2H, J = 16.5 Hz, C=CH), 6.77 (br s, 2H, PyH), 6.28 (br s, 2H, PyH), 6.09 (br s, 2H, PyH) ppm. ¹³C NMR (THF- d_8 , 125 MHz) δ: 155.1, 139.3, 136.3, 133.3, 131.9, 130.3, 128.4, 126.4, 122.4, 121.2, 120.3, 110.5, 110.0 ppm. LRMS: 471.2 (M + H)[†]; HRMS calculated for C₃₀H₂₃N₄S: 471.1638; found 471.1624. ε_{447 nm} = 31 000; ε_{354 nm} = 52 000 (THF).

4,7-Bis(1-methyl-1H,1'H-[2,2'-bipyrrol]-5-yl)benzo[c][1,2,5]thia-diazole (2j). Compound 2j was synthesized from N-Boc-pyrrole-2-boronic acid (1b) and 4,7-bis(5-bromo-1-methyl-1H-pyrrol-2-yl) benzo[c][1,2,5]thiadiazole (j)⁷¹ using GP2. After cooling to room temperature the reaction mixture was separated between 2:1 diethyl ether: THF (100 mL) and water (100 mL). The aqueous phase was extracted with 2:1 diethyl ether: THF (2 × 100 mL) and the combined organic extracts were washed with water (200 mL) and brine (200 mL), dried over anhydrous magnesium sulfate, and concentrated *in vacuo*. The crude product was washed with 0–20% diethyl ether/hexanes and then further purified using column chromatography on silica eluting with

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50% diethyl ether/hexanes to give the title compound (140 mg, 85% yield) as a dark red/purple solid. Mp 184–187 °C. ¹H NMR (THF- d_8 , 500 MHz) δ : 10.26 (br s, 2H, NH), 7.64 (s, 2H, ArH), 6.78 (br s, 2H, PyH), 6.54 (d, 2H, J = 3.5 Hz, PyH), 6.29 (d, 2H, J =3.5 Hz, PyH), 6.26 (br s, 2H, PyH), 6.16 (d, 2H, J = 2.5 Hz, PyH), 3.68 (s, 6H, 2 × NCH₃) ppm. ¹³C NMR (THF- d_8 , 125 MHz) δ : 155.2, 132.3, 132.1, 128.9, 126.1, 125.4, 118.8, 112.6, 109.3, 108.2, 107.7, 35.1 ppm. LRMS: 425.2 (M + H)⁺; HRMS calculated for $C_{24}H_{21}N_6S$: 425.1543; found 425.1556. $\varepsilon_{519 \text{ nm}} = 11\ 300$; $\varepsilon_{311 \text{ nm}} =$ 29 000 (THF).

N,N'-Bis(2-ethylhexyl)-6,6'-bis(1H-pyrrol-2-yl)isoindigo (2k). Compound 2k was synthesized from N-Boc-pyrrole-2-boronic acid (1b) and N,N'-bis(2-ethylhexyl)-6,6'-dibromoisoindigo (k)⁷² using GP2 with stirring at 115 °C for 18 h, then 125 °C for an additional 5 h. The crude product was purified using column chromatography on silica eluting with 30-60% diethyl ether in hexanes to give the title compound (203 mg, 53% yield) as a dark blue/black solid. Mp 232-234 °C. ¹H NMR (THF-d₈, 500 MHz) δ : 10.63 (br s, 2H, NH), 9.30 (d, 2H, J = 8.5 Hz, ArH), 7.16 (dd, 2H, J = 8.5, 1.5 Hz, ArH), 7.04 (d, 2H, J = 1.5 Hz, ArH), 6.85(br s, 2H, PyH), 6.63 (br s, 2H, PyH), 6.18 (dd, 2H, J = 5.5, 2.5 Hz, PyH), 3.79-3.71 (m, 4H, $2 \times NCH_2$), 2.00-1.95 (m, 2H, $2 \times CH$), 1.48-1.29 (m, 16H, $8 \times \text{CH}_2$), 0.97 (t, 6H, J = 7.5 Hz, $2 \times \text{CH}_3$), 0.91 (t, 6H, J = 7.0 Hz, $2 \times$ CH₃) ppm. ¹³C NMR (THF- d_8 , 125 MHz) δ : 169.5, 146.6, 137.3, 132.9, 131.4, 131.2, 121.2, 120.4, 116.7, 110.6, 108.5, 103.5, 44.4, 38.6, 31.5, 29.5, 24.8, 24.0, 14.4, 11.0 ppm. LRMS: $617.4 (M + H)^{+}$; HRMS calculated for $C_{40}H_{49}N_4O_2$: 617.3850; found 617.3849. $\varepsilon_{578 \text{ nm}} = 32 \text{ 800}$; $\varepsilon_{470 \text{ nm}}$ = 19 200; $\varepsilon_{310 \text{ nm}}$ = 31 700 (THF).

(E)-5-Styryl-1H-pyrrole-2-carbaldehyde (3a). 73 2-Styryl pyrrole (2a, 53 mg, 0.31 mmol) was dissolved in anhydrous DMF (1.0 mL) with stirring under nitrogen, and the solution was cooled to 0 °C in an ice bath. Phosphorous oxychloride (30 μL, 0.33 mmol), was then added dropwise with continued stirring at 0 °C for 2 hours. 10% (w/v) aqueous potassium carbonate solution (2 mL) was then added, and the reaction mixture was separated between dichloromethane and water. The aqueous phase was extracted with dichloromethane (3 \times 10 mL) and the combined organic extracts were washed with water (2 × 40 mL) and brine (30 mL), dried over anhydrous sodium sulfate and concentrated in vacuo. The crude product was purified using column chromatography on silica eluting with 20-30% ethyl acetate in hexanes to give the title compound (26 mg, 42% yield) as a light yellow solid. Mp 141–144 °C. 1 H NMR (CDCl₃, 500 MHz) δ : 9.65 (brs, 1H, NH), 9.49 (s, 1H, CHO), 7.49 (d, 2H, J = 7.5 Hz, ArH), 7.38 (t, 2H, J = 7.5 Hz, ArH), 7.30 (t, 1H, J = 7.5 Hz, ArH), 7.07 (d, 1H, J = 16.5 Hz, CH = C), 6.99-6.97 (m, 1H, PyH), 6.97 (d, 1H, J = C)16.5 Hz, CH=C), 6.49 (dd, 1H, J = 3.5, 2.5 Hz, PyH) ppm. ¹³C NMR (CDCl₃, 125 MHz) δ: 178.7, 139.1, 136.4, 133.0, 131.0, 129.0, 128.5, 126.7, 123.0, 117.4, 110.9 ppm. LRMS: 220.1 (M + Na)⁺; HRMS calculated for C₁₃H₁₁NONa: 220.0733; found 220.0734.

5,5'-((1E,1'E)-1,4-Phenylenebis(ethene-2,1-diyl))bis(1H-pyrrole-2-carbaldehyde) (3b). Compound 3b was synthesized from 2b (130 mg, 0.50 mmol) using GP3 to give the title compound (135 mg, 85% yield) as a dark yellow solid. Mp/dp > 250 °C. ¹H NMR (DMSO- d_6 , 500 MHz) δ : 12.24 (br s, 2H, NH), 9.44 (s, 2H,

CHO), 7.52 (s, 4H, ArH), 7.37 (d, 2H, J = 16.5 Hz, C=CH), 7.13 (d, 2H, J = 16.5 Hz, C=CH), 7.03 (dd, 2H, J = 2.0, 3.5 Hz, PyH),6.58 (dd, 2H, J = 2.0, 3.5 Hz, PyH) ppm. ¹³C NMR (DMSO- d_6 , 125 MHz) δ : 178.5, 138.8, 136.3, 133.3, 129.4, 126.8, 118.0, 110.5 ppm (one signal missing). LRMS: 315.1 $(M - H)^-$; HRMS calculated for $C_{20}H_{15}N_2O_2$: 315.1139; found 315.1131. $\varepsilon_{437 \text{ nm}} =$ 46 000; $\varepsilon_{413 \text{ nm}} = 59\ 000\ \text{(DMSO)}$.

5.5'-((1E,1'E)-[1,1'-Biphenyl]-4.4'-diylbis(ethene-2,1-diyl))bis(1H-pyrrole-2-carbaldehyde) (3c). Compound 3c was synthesized from 2c (50 mg, 0.15 mmol) using GP3 to give the title compound (50 mg, 85% yield) as a dark yellow solid. Mp/dp > 250 °C. ¹H NMR (DMSO- d_6 , 500 MHz) δ : 12.25 (br s, 2H, NH), 9.44 (s, 2H, CHO), 7.76 (d, 4H, J = 8.0 Hz, ArH), 7.60 (d, 4H, J =8.0 Hz, ArH), 7.42 (d, 2H, J = 16.5 Hz, C=CH), 7.17 (d, 2H, J =16.5 Hz, C=CH), 7.04 (d, 2H, J = 3.3 Hz, PyH), 6.60 (d, 2H, J =3.3 Hz, PyH) ppm. 13 C NMR (DMSO- d_6 , 125 MHz) δ : 178.5, 138.7, 135.9, 133.2, 129.3, 127.1, 126.9, 126.8, 118.1, 110.4 ppm (one signal missing). LRMS: 393.2 (M + H)⁺; HRMS (APCI) calculated for $C_{26}H_{21}N_2O_2$: 393.1598; found 393.1596. $\varepsilon_{401 \text{ nm}} = 81\,000$ (DMSO).

5,5'-((1E,1'E)-Naphthalene-2,6-diylbis(ethene-2,1-diyl))bis(1Hpyrrole-2-carbaldehyde) (3d). Compound 3d was synthesized from 2d (60 mg, 0.19 mmol) using GP3 to give the title compound (54 mg, 76% yield) as a dark yellow solid. Mp/dp > 250 °C. ¹H NMR (DMSO- d_6 , 500 MHz) δ : 12.29 (br s, 2H, NH), 9.46 (s, 2H, CHO), 7.93 (d, 2H, J = 9.3 Hz, ArH), 7.90 (s, 2H, ArH), 7.75 (d, 2H, J = 9.3 Hz, ArH), 7.54 (d, 2H, J = 16.5 Hz, C = CH), 7.25 (d, 2H, I = 16.5 Hz, C=CH), 7.06 (d, 2H, I = 3.9 Hz, PyH), 6.62 (d, 2H, J = 3.9 Hz, PyH) ppm. ¹³C NMR (DMSO- d_6 , 125 MHz) δ : 178.6, 138.8, 134.5, 133.4, 132.9, 129.8, 128.6, 126.2, 123.8, 118.6, 110.6 ppm (one signal missing). LRMS: 367.2 (M + H) $^{+}$; HRMS (APCI) calculated for $C_{24}H_{18}N_2O_2$: 367.1441; found 367.1431. $\varepsilon_{433 \text{ nm}} = 26\ 000; \ \varepsilon_{408 \text{ nm}} = 31\ 000 \ \text{(DMSO)}.$

5,5'-((1E,1'E)-Benzo[c][1,2,5]thiadiazole-4,7-diylbis(ethene-2,1diyl))bis(1H-pyrrole-2-carbaldehyde) (3e). Compound 3e was synthesized from 2e (50 mg, 0.16 mmol) using GP3 to give the title compound (57 mg, 97% yield) as a dark red solid. Mp/dp > 250 °C. ¹H NMR (THF- d_8 , 500 MHz) δ : 11.57 (br s, 2H, NH), 9.48 (s, 2H, CHO), 8.13 (d, 2H, J = 16.3 Hz, C=CH), 7.68 (s, 2H, ArH), 7.64 (d, 2H, J = 16.3 Hz, C=CH), 6.94 (br s, 2H, PyH), 6.62 (br s, 2H, PyH) ppm. 13 C NMR (THF- d_8 , 125 MHz) δ: 178.5, 154.7, 139.6, 135.4, 130.1, 129.0, 126.5, 124.0, 122.0, 111.5 ppm. LRMS: 375.1 $(M + H)^+$; HRMS (APCI) calculated for $C_{20}H_{15}N_4SO_2$: 375.0910; found 375.0892. $\varepsilon_{500 \text{ nm}} = 28\ 000$; $\varepsilon_{377 \text{ nm}} = 29\ 000$ (DMSO).

5,5'-((1E,1'E)-Anthracene-9,10-diylbis(ethene-2,1-diyl))bis(1Hpyrrole-2-carbaldehyde) (3f). Compound 3f was synthesized from 2f (84 mg, 0.23 mmol) using GP3 to give the title compound (84 mg, 87% yield) as a dark yellow solid. Mp/dp > 250 °C. ¹H NMR (DMSO- d_6 , 500 MHz) δ : 12.51 (br s, 2H, NH), 9.53 (s, 2H, CHO), 8.44-8.42 (m, 4H, ArH), 8.31 (d, 2H, J = 16.5 Hz, C=CH), 7.60–7.58 (m, 4H, ArH), 7.13–7.12 (m, 2H, PyH), 6.89 (d, 2H, J =16.5 Hz, C=CH), 6.78-6.77 (m, 2H, PyH) ppm. ¹³C NMR (DMSO d_6 , 125 MHz) δ : 178.9, 138.3, 133.5, 131.9, 128.8, 126.9, 126.1, 126.0, 125.8, 111.1 ppm (one signal missing). LRMS: 417.2 (M + H)⁺; HRMS (APCI) calculated for $C_{28}H_{21}N_2O_2$: 417.1598; found

417.1581. $\varepsilon_{431~\rm nm}=19~000;\ \varepsilon_{334~\rm nm}=22~000;\ \varepsilon_{306~\rm nm}=20~000;\ \varepsilon_{264~\rm nm}=62~000$ (DMSO).

5,5'-((1E,1'E)-(9H-Fluorene-2,7-diyl)bis(ethene-2,1-diyl))bis(1H-pyrrole-2-carbaldehyde) (3g). Compound 3g was synthesized from 2g (97 mg, 0.28 mmol) using GP3 to give the title compound (106 mg, 94% yield) as a brown solid. Mp/dp > 250 °C. ¹H NMR (DMSO- d_6 , 500 MHz) δ: 12.25 (br s, 2H, NH), 9.44 (s, 2H, CHO), 7.90 (d, 2H, J = 8.0 Hz, ArH), 7.74 (s, 2H, ArH), 7.53 (d, 2H, J = 8.0 Hz, ArH), 7.46 (d, 2H, J = 16.5 Hz, C=CH), 7.17 (d, 2H, J = 16.5 Hz, C=CH), 7.04 (d, 2H, J = 3.5 Hz, PyH), 6.59 (d, 2H, J = 3.5 Hz, PyH), 4.00 (s, 2H, CH₂) ppm. 13 C NMR (DMSO- d_6 , 125 MHz) δ: 178.4, 144.2, 140.8, 139.0, 135.5, 133.2, 130.3, 125.8, 122.6, 120.5, 117.5, 110.3, 36.3 ppm (one signal missing). LRMS: 405.2 (M + H)⁺; HRMS (APCI) calculated for C₂₇H₂₁N₂O₂: 405.1598; found 405.1580. ε _{436 nm} = 55 000; ε _{412 nm} = 70 000 (DMSO).

5,5'-((1E,1'E)-Pyrene-1,6-diylbis(ethene-2,1-diyl))bis(1H-pyrrole-2-carbaldehyde) (3h). Compound 3h was synthesized from 2h (60 mg, 0.16 mmol) using GP3 to give the title compound (65 mg, 95% yield) as a brown solid. Mp/dp > 250 °C. ¹H NMR (DMSO- d_6 , 500 MHz) δ: 12.56 (br s, 2H, NH), 9.51 (s, 2H, CHO), 8.80 (d, 2H, J = 9.3 Hz, ArH), 8.60 (d, 2H, J = 16.5 Hz, C=CH), 8.48 (d, 2H, J = 8.0 Hz, ArH), 8.32 (d, 2H, J = 8.0 Hz, ArH), 8.27 (d, 2H, J = 9.3 Hz, ArH), 7.48 (d, 2H, J = 16.5 Hz, C=CH), 7.11 (br s, 2H, PyH), 6.70 (br s, 2H, PyH) ppm. ¹³C NMR (DMSO- d_6 , 125 MHz) δ: 178.8, 139.1, 133.8, 131.3, 130.3, 128.5, 127.6, 125.6, 125.5, 124.7, 123.2, 122.9, 120.5, 112.5 ppm (one signal missing). LRMS: 441.2 (M + H)†; HRMS (APCI) calculated for $C_{30}H_{21}N_2O_2$: 441.1598; found 441.1588. $ε_{453 \text{ nm}}$ = 55 000; $ε_{332 \text{ nm}}$ = 38 000; $ε_{257 \text{ nm}}$ = 31 000 (DMSO).

5,5'-((1E,1'E)-(Benzo[c][1,2,5]thiadiazole-4,7-diylbis(4,1-phenylene))bis(ethene-2,1-diyl))bis(1H-pyrrole-2-carbaldehyde) (3i). Compound 3i was synthesized from 2i (44 mg, 0.11 mmol) using GP3 to give the title compound (113 mg, 92% yield) as a brown solid. Mp/dp > 250 °C. ¹H NMR (DMSO- d_6 , 500 MHz) δ : 12.30 (br s, 2H, NH), 9.46 (s, 2H, CHO), 8.11 (d, 2H, J = 7.5 Hz, ArH), 8.03 (s, 2H, ArH), 7.70 (d, 2H, J = 7.5 Hz, ArH), 7.48 (d, 2H, J = 16.0 Hz, C=CH), 7.23 (d, 2H, J = 16.0 Hz, C=CH), 7.06 (br s, 2H, PyH), 6.63 (br s, 2H, PyH) ppm. ¹³C NMR (DMSO- d_6 , 125 MHz) δ : 178.6, 153.4, 138.7, 136.7, 136.1, 133.4, 131.7, 129.6, 129.3, 128.1, 126.5, 118.7, 110.6 ppm (one signal missing). LRMS: 527.1 (M + H)†; HRMS (APCI) calculated for $C_{32}H_{23}N_4SO_2$: 527.1536; found 527.1512. $\varepsilon_{431 \text{ nm}}$ = 42 000; $\varepsilon_{373 \text{ nm}}$ = 53 000 (DMSO).

5,5'-(Benzo[c][1,2,5]thiadiazole-4,7-diyl)bis(1'-methyl-1H,1'H-[2,2'-bipyrrole]-5-carbaldehyde) (3j). Compound 3j was synthesized from 2j (50 mg, 0.12 mmol) using GP3 and purified using column chromatography over silica eluting with 2 : 1 : 2 diethyl ether : THF : hexane to give the title compound (47 mg, 84% yield) as a dark red solid. Mp/dp > 250 °C. ¹H NMR (DMSO-d₆, 500 MHz) δ: 12.26 (br s, 2H, NH), 9.49 (s, 2H, CHO), 7.78 (s, 2H, ArH), 7.14 (d, 2H, J = 3.5 Hz, PyH), 6.82 (d, 2H, J = 3.5 Hz, PyH), 6.62 (d, 2H, J = 3.5 Hz, PyH), 6.60 (d, 2H, J = 3.5 Hz, PyH), 3.71 (s, 6H, NMe) ppm. 13 C NMR (DMSO-d₆, 125 MHz) δ: 178.4, 153.5, 133.0, 132.8, 132.5, 128.9, 128.1, 124.2, 112.3, 112.2, 110.1, 109.7, 35.1 ppm. LRMS: 481.1 (M + H) $^+$; HRMS calculated for C₂₆H₂₁N₆SO₂: 481.1441; found 481.1422. ε _{496 nm} = 32 700; ε _{365 nm} = 72 400 (DMSO).

(E)-5,5'-(1,1'-Bis(2-ethylhexyl)-2,2'-dioxo-[3,3'-biindolinylidene]-6,6'-diyl)bis(1H-pyrrole-2-carbaldehyde) (3k). Compound 3k was synthesized from 2k (60 mg, 0.10 mmol) using GP3 to give the title compound (59 mg, 90% yield) as a dark purple/black solid. Mp/dp > 250 °C. ¹H NMR (DMSO-d₆, 500 MHz) δ: 12.54 (br s, 2H, NH), 9.56 (s, 2H, CHO), 9.04 (d, 2H, J = 8.0 Hz, ArH), 7.56 (d, 2H, J = 8.0 Hz, ArH), 7.48 (s, 2H, ArH), 7.16 (br s, 2H, PyH), 6.99 (br s, 2H, PyH), 3.62–3.55 (m, 4H, NCH₂), 1.91–1.83 (m, 2H, CH), 1.35–1.25 (m, 10H, CH₂), 1.25–1.17 (m, 6H, CH₂), 0.85 (t, 6H, J = 6.8 Hz, CH₃), 0.84–0.78 (m, 6H, CH₃) ppm. ¹³C NMR (DMSO-d₆, 125 MHz) δ: 179.3, 167.6, 145.5, 139.0, 134.5, 134.4, 131.1, 129.4, 122.4, 120.4, 118.8, 110.7, 104.9, 43.5, 36.7, 29.8, 27.8, 23.3, 22.6, 13.9, 10.4 ppm. LRMS: 673.4 (M + H)⁺; HRMS calculated for C₄₂H₄₉N₄O₄: 673.3748; found 673.3737. ε_{579 nm} = 26 800; ε_{466 nm} = 24 200; ε_{331 nm} = 29 200 (DMSO).

 $[Ru(3a)(bpy)_2]PF_6$ complex salt (4a). Complex salt 4a was synthesized from ligand 3a using GP4 and 1 equiv. cis-bis-(2,2'bipyridine)dichlororuthenium(II) dihydrate for 1 h to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4b (50 mg, 96% yield) as a black glittery solid following isolation by Millipore filtration. Mp 170–175 °C. ¹H NMR (CDCl₃, 500 MHz) δ : 8.55 (s, 1H, CHO), 8.53 (d, 1H, J = 6.0 Hz, ArH), 8.39 (d, 1H, J =8.0 Hz, ArH), 8.36 (t, 2H, J = 7.0 Hz, ArH), 8.30 (d, 1H, J = 8.0 Hz, ArH), 7.99 (t, 1H, J = 8.5 Hz, ArH), 7.94 (t, 1H, J = 7.0 Hz, ArH), 7.91–7.87 (m, 2H, ArH), 7.85 (t, 1H, J = 7.5 Hz, ArH), 7.79 (t, 1H, J= 7.5 Hz, ArH, 7.53-7.50 (m, 2H, ArH), 7.40 (t, 1H, J = 6.5 Hz,ArH), 7.28-7.21 (m, 3H, ArH), 7.18-7.15 (m, 3H, ArH), 6.82 (d, 1H, J = 16.5 Hz, CH=C), 6.73 (ad, 2H, J = 7.5 Hz, ArH), 6.70 (d, 1H, J = 4.5 Hz, ArH), 5.50 (d, 1H, J = 16.5 Hz, CH=C) ppm. ¹³C NMR (CDCl₃, 125 MHz) δ: 179.6, 159.4, 158.3, 158.1, 157.2, 155.0, 153.0, 151.9, 151.7, 150.6, 144.8, 136.8, 136.6, 135.9, 135.1, 132.0, 128.6, 128.1, 127.0, 126.9, 126.8, 126.7, 126.3, 125.8, 123.9, 123.5, 123.4, 120.6, 114.6 ppm (two signals missing). LRMS: 610.1 (M)⁺; HRMS calculated for C₃₃H₂₆N₅ORu: 610.1175; found 610.1156. $\varepsilon_{473 \text{ nm}} = 10\ 900$; $\varepsilon_{346 \text{ nm}} = 27\ 300$; $\varepsilon_{295 \text{ nm}} = 57 \text{ } 100 \text{ (CH}_2\text{Cl}_2\text{)}$. The corresponding chloride salt 5a was obtained following GP5, after which the reaction mixture was concentrated in vacuo and the residue was purified over basic alumina eluting with 10-40% methanol in ethyl acetate to give 5a (13 mg, 83%) as a red/brown solid. Mp/dp > 250 °C. LRMS: $610.1 \, (M)^+$; PF_6^- ion not observed in negative mode.

[$Ru_2(3b)(bpy)_4$](PF_6)₂ complex salt (4b). Complex salt 4b was synthesized from ligand 3b using GP4 to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4b (56 mg, 86% yield) as a black glittery solid. Mp/dp > 250 °C. ¹H NMR (CD₂Cl₂, 500 MHz) δ : 8.56 (s, 2H, 2× CHO), 8.56–8.55 (m, 2H, ArH), 8.40–8.37 (m, 4H, ArH), 8.32 (d, 2H, J = 8.5 Hz, ArH), 8.24 (d, 2H, J = 8.0 Hz, ArH), 8.00–7.92 (m, 8H, ArH), 7.89–7.82 (m, 4H, ArH), 7.55–7.53 (m, 4H, ArH), 7.44 (t, 2H, J = 6.5 Hz, ArH), 7.28–7.20 (m, 6H, ArH), 6.79 (d, 2H, J = 16.0 Hz, ArH), 6.70 (d, 2H, J = 4.0 Hz, ArH), 6.61 (s, 4H, ArH), 5.45 (d, 2H, J = 16.0 Hz, ArH) ppm. 13 C NMR (CD₂Cl₂, 125 MHz) δ : 180.2, 159.7, 158.8, 158.1, 157.6, 155.1, 153.4, 152.5, 152.0, 151.0, 145.3, 136.8, 136.5, 136.0, 135.3, 131.5, 127.4, 126.94, 126.86, 126.8, 126.6, 125.8, 124.0, 123.5, 123.4, 123.3, 120.94, 120.92, 114.9, 70.8 ppm. LRMS: 571.1 (M/2)⁺ and 145.0 (PF₆)⁻; HRMS calculated

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for $C_{60}H_{46}N_{10}O_2Ru_2$: 571.0941; found 571.0917. $\varepsilon_{489\ nm} =$ 41 000; $\varepsilon_{377~\rm nm}=54~000$; $\varepsilon_{294~\rm nm}=112~000$ (CH₂Cl₂). The corresponding dichloride salt 5b was obtained following GP5 and isolated via Millipore filtration (13 mg, 73%) as a red/brown solid. Mp/dp > 250 °C. LRMS: 571.1 $(M/2)^+$; PF₆ ion not observed in negative mode.

 $[Ru_2(3c)(bpy)_4](PF_6)_2$ complex salt (4c). Complex salt 4c was synthesized from ligand 3c using GP4 to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4c (22 mg, 61% yield) as a deep red solid. Mp/dp > 250 °C; ¹H NMR (CD₂Cl₂, 500 MHz) δ: 8.59–8.56 (m, 2H, ArH), 8.56 (s, 2H, CHO), 8.45 (t, 2H, J = 7.3 Hz, ArH, 8.38-8.36 (m, 2H, ArH), 8.32 (d, 2H, J = 8.0 Hz, ArH), 8.25 (d, 2H, J = 8.0 Hz, ArH), 8.10 (t, 2H, J = 7.5 Hz, ArH), 7.99-7.95 (m, 6H, ArH), 7.91 (d, 2H, J = 5.5 Hz, ArH), 7.86-7.83 (m, 2H, ArH), 7.57-7.53 (m, 4H, ArH), 7.49 (d, 4H, <math>J = 8.3 HzArH), 7.44 (t, 2H, J = 6.0 Hz, ArH), 7.40–7.36 (m, 2H, ArH), 7.26 (d, 2H, J = 4.5 Hz, PyH), 7.22 (t, 2H, J = 6.5 Hz, ArH), 6.87 (d, 2H, J = 6.J = 16.0 Hz, C = CH), 6.81 (d, 4H, J = 8.3 Hz, ArH), 6.73 (d, 2H, J)= 4.5 Hz, PyH), 5.55 (d, 2H, J = 16.0 Hz, C=CH), 1.53 (br s, 8H, H_2O) ppm. ¹³C NMR (CD₂Cl₂, 125 MHz) δ : 180.2, 159.6, 158.8, 158.2, 157.7, 155.1, 153.3, 152.6, 152.0, 151.1, 145.2, 139.7, 136.7, 136.5, 136.2, 136.1, 136.0, 131.4, 127.3 ($2 \times C$) 127.1 ($2 \times C$) C), 126.9 (2× C), 125.9, 124.4, 123.4 (2× C), 123.3, 121.0, 114.7 ppm. LRMS: 609.1 (M/2)^+ and $145.0 \text{ (PF}_6)^-$; HRMS calculated for $C_{66}H_{50}N_{10}O_2Ru_2$: 609.1097; found 609.1101. $\varepsilon_{472 \text{ nm}} =$ 36 000; $\varepsilon_{430~\rm nm}=42$ 000; $\varepsilon_{374~\rm nm}=66$ 000; $\varepsilon_{294~\rm nm}=106$ 000 (CH₂Cl₂). The corresponding dichloride salt 5c was obtained following **GP5** and isolated *via* Millipore filtration (11 mg, 72%) as a red/brown solid. Mp/dp > 250 °C. LRMS: 609.1 (M/2)^+ ; PF₆ ion not observed in negative mode.

 $[Ru_2(3d)(bpy)_4](PF_6)_2$ complex salt (4d). Complex salt 4d was synthesized from ligand 3d using GP4 to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4d (18 mg, 42% yield) as a deep red solid. Mp/dp > 250 °C. ¹H NMR (CD₂Cl₂, 500 MHz) δ: 8.57 (s, 2H, CHO), 8.57-8.56 (m, 2H, ArH), 8.51 (dd, 2H, J = 4.5, 8.0 Hz, ArH, 8.42 (dd, 2H, J = 4.0, 8.0 Hz, ArH), 8.31 (d, 2H, J = 8.5 Hz, ArH, 8.25 (d, 2H, J = 8.0 Hz, ArH), 8.03-7.93 (m, 3.03-7.93 (m,10H, ArH), 7.86-7.83 (m, 2H, ArH), 7.57-7.52 (m, 6H, ArH), 7.45-7.42 (m, 2H, ArH), 7.37-7.33 (m, 2H, ArH), 7.27 (d, 2H, J =4.0 Hz, ArH), 7.23-7.21 (m, 4H, ArH), 6.98 (dd, 2H, J = 2.0, 16.0 Hz, C=CH), 6.78-6.76 (m, 4H, ArH), 5.59 (dd, 2H, J = 5.5, 16.0 Hz, C=CH), 1.53 (br s, 8H, H₂O) ppm. ¹³C NMR (CD₂Cl₂, 125 MHz) δ: 180.3, 159.7, 158.8, 158.2, 157.7, 155.1, 153.4, 152.5, 152.0, 151.0, 145.3, 136.7, 136.5, 136.0, 135.8, 134.9, 133.3, 131.9, 128.2, 127.4, 127.2, 126.9 ($2 \times C$), 125.84, 125.79, 124.8, 124.4, 123.5, 123.4, 123.3, 121.5, 114.9 ppm. LRMS: 596.1 (M/2)⁺ and 145.0 (PF₆) $^-$; HRMS calculated for $C_{64}H_{48}N_{10}O_2Ru_2$: 596.1019; found 596.1005. $\varepsilon_{481 \text{ nm}} = 42\ 000$; $\varepsilon_{437\ \text{nm}} = 42\ 000$; $\varepsilon_{380~\mathrm{nm}}$ = 58 000; $\varepsilon_{294~\mathrm{nm}}$ = 116 000 (CH₂Cl₂). The corresponding dichloride salt 5d was obtained following GP5 and isolated via Millipore filtration (7 mg, 78%) as a red/brown solid. Mp/dp > 250 °C. LRMS: 596.1 $(M/2)^+$; PF₆ ion not observed in negative

 $[Ru_2(3e)(bpy)_4](PF_6)_2$ complex salt (4e). Complex salt 4e was synthesized from ligand 3e using GP4 to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4e (24 mg, 61% yield) as a deep purple solid. Mp/dp > 250 °C. ¹H NMR (CD₂Cl₂,

500 MHz) δ: 8.62 (s, 2H, CHO), 8.52-8.50 (m, 2H, ArH), 8.46-8.39 (m, 4H, ArH), 8.32-8.29 (m, 2H, ArH), 8.24-8.22 (m, 2H, ArH), 8.00-7.95 (m, 6H, ArH), 7.91-7.86 (m, 4H, ArH), 7.85-7.81 (m, 2H, ArH), 7.54-7.51 (m, 2H, ArH), 7.49 (d, 2H, J = 7.0 Hz, ArH), 7.45–7.41 (m, 2H, ArH), 7.34 (dd, 2H, J = 2.0, 16.0, C=CH), 7.30 (d, 2H, I = 5.0 Hz, ArH), 7.24–7.17 (m, 4H, ArH), 6.93 (s, 2H, ArH), 6.86 (d, 2H, J = 4.5 Hz, ArH), 6.39 (dd, 2H, J = 2.0, 16.0, C=CH), 1.54 (br s, 8H, H₂O) ppm. ¹³C NMR (CD₂Cl₂, 125 MHz) δ: 180.7, 159.5, 158.8, 158.2, 157.8, 155.4, 153.7, 153.3, 152.6, 151.8, 150.9, 145.9, 136.8, 136.5, 136.1, 135.6, 129.1, 128.0, 127.4, 127.3, 127.0, 126.9, 126.8, 125.8, 125.4, 124.2, 123.5, 123.31, 123.26, 115.4 ppm. LRMS: $600.1 \, (M/2)^+$ and $145.0 \, (PF_6)^-$; HRMS calculated for C₆₀H₄₄N₁₂SO₂Ru₂: 600.0753; found 600.0733. $\varepsilon_{525 \text{ nm}} = 42\ 000;\ \varepsilon_{358\ \text{nm}} = 40\ 000;\ \varepsilon_{295\ \text{nm}} = 114\ 000$ (CH₂Cl₂). The corresponding dichloride salt 5e was obtained following GP5 and isolated via Millipore filtration (11 mg, 71%) as a brown solid. Mp/dp > 250 °C. LRMS: $600.1 (M/2)^{+}$; PF₆ ion not observed in negative mode.

 $[Ru_2(3f)(bpy)_4](PF_6)_2$ complex salt (4f). Salt 4f was synthesized from ligand 3f using GP4 to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4f (24 mg, 45% yield) as a deep red solid. Mp/dp > 250 °C. 1 H NMR (CD₂Cl₂, 500 MHz) δ : 8.64 (s, 2H, CHO), 8.59 (d, 2H, J = 5.5 Hz, ArH), 8.36 (d, 2H, J = 8.0 Hz, ArH), 8.31-8.25 (m, 4H, ArH), 8.19 (d, 2H, J = 8.0 Hz, ArH), 8.12-8.08 (m, 2H, ArH), 8.04-8.01 (m, 4H, ArH), 8.00-7.96 (m, 2H, ArH), 7.93-7.89 (m, 4H, ArH), 7.81–7.77 (m, 2H, ArH), 7.68 (d, 2H, J = 16.0 Hz, C=CH), 7.62-7.59 (m, 2H, ArH), 7.52-7.48 (m, 4H, ArH), 7.44-7.42 (m, 4H, ArH), 7.37–7.35 (m, 2H, ArH), 7.18–7.15 (m, 2H, ArH), 6.99– 6.95 (m, 2H, ArH), 6.54-6.49 (m, 2H, ArH), 6.10-6.04 (m, 2H, ArH), 5.04 (d, 2H, J = 16.0 Hz, C=CH), 1.54 (br s, 8H, H₂O) ppm. ¹³C NMR (CD₂Cl₂, 125 MHz) δ: 181.0, 158.7, 158.2, 157.6, 154.4, 152.5, 152.4, 152.3, 152.00, 151.96, 151.0, 145.1, 136.7, 136.4, 136.3, 133.7, 132.4, 131.0, 129.2, 127.5, 127.3, 127.0, 126.8, 126.5, 125.8, 125.7, 125.6, 123.9, 123.5, 123.3, 114.5 ppm. LRMS: 621.1 (M/2)⁺ and 145.0 (PF₆)⁻; HRMS calculated for $C_{68}H_{50}N_{10}O_2Ru_2$: 621.1097; found 621.1074. $\varepsilon_{508 \text{ nm}} = 30\ 000;\ \varepsilon_{345\ \text{nm}} = 34\ 000;\ \varepsilon_{295\ \text{nm}} =$ 110 000 (CH₂Cl₂). The corresponding dichloride salt 5f was obtained following GP5 and isolated via Millipore filtration (9 mg, 52%) as a red/brown solid. Mp/dp > 250 °C. LRMS: 621.1 $(M/2)^+$; PF₆⁻ ion not observed in negative mode.

 $[Ru_2(3g)(bpy)_4](PF_6)_2$ complex salt (4g). Complex salt 4g was synthesized from ligand 3g using GP4 to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4g (39 mg, 66% yield) as a deep red solid. Mp/dp > 250 °C. ¹H NMR (CD₂Cl₂, 500 MHz) δ: 8.57 (d, 2H, J = 5.5 Hz, ArH), 8.55 (s, 2H, CHO), 8.51 (d, 2H, J = 8.0 Hz, ArH), 8.41 (d, 2H, J = 8.0 Hz, ArH), 8.31 (d, 2H, J =8.0 Hz, ArH), 8.25 (d, 2H, J = 8.0 Hz, ArH), 8.07 (t, 2H, J = 8.0 Hz, ArH), 7.99–7.94 (m, 8H, ArH), 7.86–7.83 (m, 2H, ArH), 7.59–7.53 (m, 6H, ArH), 7.44 (t, 2H, J = 7.0 Hz, ArH), 7.40–7.37 (m, 2H, ArH), 7.26 (d, 2H, J = 4.5 Hz, ArH), 7.22 (t, 2H, J = 6.0 Hz, ArH), 6.93 (d, 2H, J =16.5 Hz, C=CH), 6.88 (s, 2H, ArH), 6.85 (d, 2H, J = 8.0 Hz, ArH), 6.74 (d, 2H, J = 4.5 Hz, ArH), 5.57 (dd, 2H, J = 6.0, 16.5 Hz, C=CH),3.81 (s, 2H, CH₂), 1.54 (br s, 8H, H₂O) ppm. ¹³C NMR (CD₂Cl₂, 125 MHz) δ : 179.9, 159.7, 158.8, 158.2, 157.7, 155.3, 153.4, 152.5, 152.0, 151.0, 145.2, 144.4, 141.5, 136.7, 136.4, 136.0, 135.9, 135.7, 132.3, 127.3, 127.2, 126.9, 126.7, 125.9, 124.4, 123.5, 123.34, 123.26, 122.1, 120.5, 120.4, 120.2, 114.7, 36.6 ppm. LRMS: 615.1 (M/2)⁺ and 145.0 (PF₆)⁻; HRMS calculated for $C_{67}H_{50}N_{10}O_2Ru_2$: 615.1097; found 615.1084. $\varepsilon_{477~nm}=49~000$; $\varepsilon_{435~nm}=54~000$; $\varepsilon_{381~nm}=72~000$; $\varepsilon_{294~nm}=124~000$ (CH₂Cl₂). The corresponding dichloride salt **5g** was obtained following **GP5** and isolated *via* Millipore filtration (13 mg, 75%) as a red/brown solid. Mp/dp > 250 °C. LRMS: 615.1 (M/2)⁺; PF₆⁻ ion not observed in negative mode.

 $[Ru_2(3h)(bpy)_4](PF_6)_2$ complex salt (4h). Complex salt 4h was synthesized from ligand 3h using GP4 to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4h (39 mg, 69% yield) as a deep red solid. Mp/dp > 250 °C. 1 H NMR (CD₂Cl₂, 500 MHz) δ : 8.62 (s, 2H, CHO), 8.60 (d, 2H, I = 5.5 Hz, ArH), 8.38-8.30 (m, 8H, ArH), 8.27-8.24 (m, 4H, ArH), 8.04-8.03 (m, 2H, ArH), 8.02-7.95 (m, 6H, ArH), 7.92-7.91 (m, 2H, ArH), 7.88-7.84 (m, 4H, ArH), 7.77 (t, 2H, J = 7.5 Hz, ArH), 7.59–7.56 (m, 4H, ArH), 7.46 (t, 2H, I = 6.8 Hz, ArH), 7.33 (d, 2H, I = 4.5 Hz, ArH), 7.23 (t, 2H, J = 6.3 Hz, ArH), 7.17–7.12 (m, 4H, ArH), 6.94 (d, 2H, J = 4.5, ArH), 5.75 (d, 2H, J = 16.0 Hz, C=CH), 1.53 (br s, 8H, H_2O) ppm. ¹³C NMR (CD₂Cl₂, 125 MHz) δ: 180.6, 159.7, 158.8, 158.2, 157.6, 155.3, 153.3, 152.5, 152.0, 151.1, 145.4, 136.8, 136.6, 136.1, 135.3, 131.8, 131.0, 128.6, 128.3, 128.0, 127.4, 127.0, 126.9 (2 \times C), 126.0, 125.5, 125.2, 125.1, 124.2, 124.1, 123.8, 123.4, 123.3, 123.0, 114.9 ppm. LRMS: 633.1 (M/2)⁺ and 145.0 (PF₆)⁻; HRMS calculated for $C_{70}H_{50}N_{10}O_2Ru_2$: 633.1097; found 633.1119. $\varepsilon_{511 \text{ nm}} = 64\ 000;\ \varepsilon_{401 \text{ nm}} = 40\ 000;\ \varepsilon_{294 \text{ nm}} =$ 132 000 (CH₂Cl₂). The corresponding dichloride salt 5h was obtained following GP5 and isolated via Millipore filtration (15 mg, 83%) as a red/brown solid. Mp/dp > 250 °C. LRMS: 633.1 $(M/2)^+$; PF₆ ion not observed in negative mode.

 $[Ru_2(3i)(bpy)_4](PF_6)_2$ complex salt (4i). Complex salt 4i was synthesized from ligand 3i using GP4 to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4i (46 mg, 72% yield) as a deep red solid. Mp/dp > 250 °C. ¹H NMR (CD₂Cl₂, 500 MHz) δ : 8.59 (s, 2H, CHO), 8.60–8.57 (m, 2H, ArH), 8.40–8.34 (m, 6H, ArH), 8.28 (d, 2H, J = 8.0 Hz, ArH), 8.01–7.96 (m, 10H, ArH), 7.86 (at, 8H, J = 8.3 Hz, ArH), 7.58-7.55 (m, 4H, ArH), 7.45 (t, 2H, J = 6.5 Hz, ArH), 7.34 (t, 2H, J = 6.8 Hz, ArH), 7.28 (d, 2H, J =4.5 Hz, ArH), 7.23 (t, 2H, J = 6.5 Hz, ArH), 6.96-6.91 (m, 6H, ArH), 6.77 (d, 2H, J = 4.0 Hz, ArH), 5.62 (d, 2H, J = 16.5 Hz, C= CH), 1.55 (br s, 4H, H_2O) ppm. ¹³C NMR (CD_2Cl_2 , 125 MHz) δ : 180.4, 159.8, 158.7, 158.2, 157.5, 155.0, 154.3, 153.4, 152.5, 152.0, 151.1, 145.3, 137.1, 137.0, 136.8, 136.6, 136.0, 135.4, 132.7, 131.3, 129.6 (2× C), 128.4, 127.4, 127.1, 126.9, 126.6 (2× C), 125.9, 124.1, 123.44, 123.37 (2× C), 121.6, 114.8 ppm (one signal missing). LRMS: $676.1 \, (\text{M/2})^+$ and $145.0 \, (\text{PF}_6)^-$; HRMS calculated for $C_{72}H_{52}N_{12}SO_2Ru_2$: 676.1066; found 676.1039. $\varepsilon_{475\ nm}=$ 47 000; $\varepsilon_{358~\mathrm{nm}}=57~000$; $\varepsilon_{295~\mathrm{nm}}=122~000$ (CH₂Cl₂). The corresponding dichloride salt 5i was obtained following GP5 and isolated via Millipore filtration (13 mg, 75%) as a red/brown solid. Mp/dp > 250 °C. LRMS: 676.1 $(M/2)^+$; PF₆ ion not observed in negative mode.

[$Ru_2(3j)(bpy)_4$](PF_6)₂ complex salt (4j). Complex salt 4j was synthesized from ligand 3j using GP4 to give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4j (30 mg, 62% yield) as a deep red/black solid. Mp/dp > 250 °C. ¹H NMR (CD₂Cl₂, 500 MHz) δ : 8.73 (s, 2H, CHO), 8.54 (d, 2H, J = 5.5 Hz, ArH), 8.36–8.32 (m, 4H, ArH), 8.25–8.22 (m, 4H, ArH), 8.10 (t, 2H, J = 7.5 Hz, ArH), 8.03 (t, 2H, J = 7.3 Hz, ArH), 7.97 (t, 2H, J =

7.8 Hz, ArH), 7.80 (t, 2H, J = 7.8 Hz, ArH), 7.63-7.59 (m, 2H, ArH), 7.59-7.56 (m, 2H, ArH), 7.55-7.53 (m, 4H, ArH), 7.50 (t, 2H, J = 6.5 Hz, ArH), 7.42 (s, 2H, ArH), 7.35 (d, 2H, J = 4.0 Hz,ArH), 7.16 (t, 2H, J = 6.5 Hz, ArH), 6.97-6.93 (m, 2H, ArH), 6.43(dd, 2H, J = 4.5, 1.0 Hz, ArH), 5.88 (dd, 2H, J = 3.5, 5.5 Hz, ArH),5.46 (t, 2H, I = 3.0 Hz, ArH), 2.95 (s, 6H, NMe) ppm. ¹³C NMR $(CD_2Cl_2, 125 \text{ MHz}) \delta$: 182.4, 158.9, 158.5, 158.4, 158.3, 154.4, 152.8, 152.6, 152.2, 150.9, 148.1, 144.7, 136.7, 136.13, 136.07, 135.4, 130.6, 130.4, 129.2, 127.2, 126.7, 126.5, 126.2, 125.0, 124.6, 123.30, 123.26, 123.2, 122.9, 119.8, 111.9, 110.1, 33.4 ppm. LRMS: 653.1 (M/2)^+ and $145.0 \text{ (PF}_6)^-$; HRMS calculated for $C_{66}H_{50}N_{14}SO_2Ru_2$: 653.1019; found 653.1011. $\varepsilon_{509 \text{ nm}} =$ 35 000; $\varepsilon_{356~\rm nm} = 35~000$; $\varepsilon_{294~\rm nm} = 127~000$ (CH₂Cl₂). The corresponding dichloride salt 5j was obtained following GP5 with 10:1 acetone: hexanes, and isolated via Millipore filtration (8 mg, 93%) as a red/brown solid. Mp/dp > 250 °C. LRMS: 653.1 $(M/2)^+$; PF₆ ion not observed in negative mode.

 $[Ru_2(3k)(bpy)_4](PF_6)_2$ complex salt (4k). Complex salt 4k was synthesized from ligand 3k using GP4 in 9:1 methanol: water for give the corresponding bis(ruthenium(II))hexafluorophosphate salt 4k (18 mg, 70% yield) as a dark brown/black solid. Mp/dp 208-213 °C. 1 H NMR (CD₂Cl₂, 500 MHz) δ : 8.76 (s, 2H, CHO), 8.60-8.58 (m, 2H, ArH), 8.51-8.48 (m, 2H, ArH), 8.35-8.29 (m, 4H, ArH), 8.22 (d, 2H, J = 7.0 Hz, ArH), 8.06–7.98 (m, 8H, ArH), 7.80 (t, 2H, J = 7.8 Hz, ArH), 7.58–7.55 (m, 2H, ArH), 7.51 (t, 2H, I = 6.3 Hz, ArH), 7.45-7.39 (m, 4H, ArH), 7.35-7.32 (m, 4H, ArH), 7.17 (t, 2H, J = 6.8 Hz, ArH), 6.79–6.71 (m, 2H, ArH), 6.46 (d, 2H, I = 4.0 Hz, ArH), 6.29–6.18 (m, 4H, ArH), 3.70–3.58 (m, 2H, NCH₂), 3.47-3.33 (m, 2H, NCH₂), 1.69 (br s, 2H, CHEt), 1.35-1.17 (m, 16H, CH₂), 0.92–0.81 (m, 12H, CH₃) ppm. ¹³C NMR (CD₂Cl₂, 125 MHz) δ: 182.9, 168.3, 158.9, 158.8, 158.5, 158.2, 157.0, 152.7, 152.3, 152.1, 151.3, 145.7, 144.7, 140.2, 136.8, 136.5, 136.1, 135.2, 132.4, 129.2, 127.3, 127.0, 126.6, 126.4, 126.2, 125.6, 123.5, 123.4, 123.0, 121.6, 120.8, 118.0, 107.6, 44.3, 37.9, 30.9, 29.0, 24.2, 23.4, 14.2, 10.7 ppm (some peaks were observed in duplicate suggesting diastereomeric effects). LRMS: 749.2 (M/2)^+ and $144.9 \text{ (PF}_6)^-$; HRMS calculated for C₈₂H₇₈N₁₂O₄Ru₂: 749.2173; found 749.2190. $\varepsilon_{516 \text{ nm}} = 30\ 100; \ \varepsilon_{377 \text{ nm}} = 32\ 400; \ \varepsilon_{295 \text{ nm}} = 118\ 400 \ \text{(CH}_2\text{Cl}_2\text{)}.$ The corresponding dichloride salt 5k was obtained following GP5 with 10:1 acetone: hexanes, stirring at room temperature for 30 min. The reaction mixture was then concentrated in vacuo and the residue purified over neutral alumina, eluting with 3-8% methanol in dichloromethane to give 5k (14 mg, 84%) as a dark brown/black solid. Mp/dp > 250 °C. LRMS: 749.2 (M/2)⁺; PF₆⁻ ion not observed in negative mode.

2.4 Methods

2.4.1 Photophysical measurements. Absorption and emission spectra were collected from dilute solutions (5 μ M) in spectroscopic-grade MeCN. Oxygen-free samples were prepared by sparging 4 mL solutions of PSs in long-neck quartz cuvettes (Luzchem SC-10L) with argon (30 min, 50 \pm 10 mmHg) prior to spectroscopic measurements. Luminescence quantum yields ($\Phi_{\rm em}$) were calculated according to eqn (1) (s = sample, r = reference) using [Ru(bpy)₃](PF₆)₂ as the reference ($\Phi_{\rm em}$ = 0.012

in aerated MeCN,⁷⁴ 0.062 in deoxygenated MeCN,²¹ and 0.38 at 77 K in 4:1 v/v ethanol-methanol glass²¹):

$$\Phi_{\rm s} = \Phi_{\rm r} \left(\frac{I_{\rm s}}{A_{\rm s}} \right) \left(\frac{A_{\rm r}}{I_{\rm r}} \right) \left(\frac{\eta_{\rm s}^2}{\eta_{\rm r}^2} \right) \tag{1}$$

Singlet oxygen quantum yields (Φ_{Δ}) were also estimated using eqn (1) with $[Ru(bpy)_3](PF_6)_2$ as the standard $(\Phi_{\Delta}=0.57$ in aerated MeCN). Absorption spectra were recorded using a Jasco V-530 spectrophotometer, and luminescence spectra were collected using a PTI Quantamaster equipped with a standard photomultiplier tube (K170B) and a Hamamatsu R5509-42 photomultiplier tube for NIR detection (<1400 nm). Luminescence lifetimes were measured using a PTI LaserStrobe system incorporating a nitrogen-dye laser (GL-3300/GL-301) integrated with an R928 stroboscopic detector. Emission was also probed by gated methods using a pulsed xenon flash lamp and gated detector. Exponential curve fitting and corrections to the wavelength-dependence of lamp output and detector response were done with PTI Felix32 software.

2.4.2 HL-60 cell culture. HL-60 cells (ATCC CCL-240) were cultured at 37 °C under 5% CO_2 in RPMI 1640 media (Mediatech Media MT-10-040-CV) supplemented with 20% FBS (PAA Laboratories, A15-701) and were passaged 3–4 times per week using standard aseptic technique. Cultures were started at 200 000 cells per mL in 25 cm² tissue culture flasks and were subcultured when growth reached approximately 1×10^6 cells per mL. Cytotoxicity and photocytotoxicity assays were performed on cells of mid-passage number (8–25 passages).

2.4.3 HL-60 cytotoxicity and photocytotoxicity assays. Cell viability experiments were performed in 96-well microtiter plates (Corning Costar, Acton, MA) with each PS dose tested in triplicate. Microtiter plates were prepared in duplicate as follows for dark and light treatments, respectively. Phosphate buffered saline (PBS) (200 µL) supplemented with 2.68 mM potassium chloride, 1.47 mM potassium phosphate monobasic, 137 mM sodium chloride, and 8.10 mM sodium phosphate dibasic was added to non-sample wells along the periphery of the plate to minimize evaporation from the inner sample wells. HL-60 cells growing in log phase (approximately 8×10^5 cells) were transferred in 50 µL aliquots to inner wells containing 25 μL of warm complete culture medium and placed in a 37 °C, 5% CO2 water-jacketed incubator (Thermo Electron Corp., Forma Series II, Model 3110, HEPA Class 100) for 1 h to equilibrate. Prewarmed aliquots (25 µL) of serially diluted ruthenium compounds (in supplemented PBS solution) were added to the microplate sample wells, and the microplates were incubated at 37 °C under 5% CO₂. A light treatment was delivered to one of the microplates at 1 or 16 h (drug-to-light interval (t_{hv})) with unfiltered light (400-700 nm, 27.8 mW cm⁻²) from a 190 W BenQ MS510 overhead projector, visible light from a Luzchem LZC-4X photoreactor equipped with 14 LES-Vis-01 bulbs (7.8 $\mathrm{mW~cm}^{-2}$), or with red light (625 nm, 28.7 $\mathrm{mW~cm}^{-2}$) from an LED array (Photodynamic, Inc.). The irradiation time was varied to yield energy densities ranging from 5 to 100 J cm⁻². Both dark and PDT-treated microplates were incubated for a further 48 h at which point prewarmed, 10 µL aliquots of Alamar Blue

reagent (Life Technologies DAL 1025) were added to all sample wells. Both microplates were incubated for 15–16 h at 37 $^{\circ}C$ under 5% CO_2 after addition of the indicator dye. Cell viability was determined based on the ability of the Alamar Blue redox indicator to be metabolically converted to a fluorescent dye by live cells. Fluorescence was quantified with a Cytofluor 4000 fluorescence microplate reader with the excitation filter set at 530 ± 25 nm and emission filter set at 620 ± 40 nm. EC_{50} values (effective concentration for reducing cell viability to 50%) for cytotoxicity (dark microplates) and photocytotoxicity (light microplates) were calculated from sigmoidal fits of the doseresponse curves using Graph Pad Prism 6.0 according to eqn (2), where γ_i and γ_f are the initial and final fluorescence signal intensities, respectively.

$$\gamma = \gamma_i + \frac{\gamma_i - \gamma_f}{1 + 10^{(\log EC_{50} - x)\alpha \text{(Hill slope)}}} \tag{2}$$

For cells growing in log phase and of similar passage number, EC_{50} values were reproducible to within $\pm 25\%$ in the submicromolar regime; $\pm 10\%$ below 10 μ M; and $\pm 5\%$ above 10 μ M. Photocytotoxicity indices (PIs), a measure of the therapeutic window, were calculated from the ratio of dark to light EC_{50} values obtained from the dose–response curves.

2.4.4 HL-60 multicellular tumor spheroid cytotoxicity and photocytotoxicity assays. Multicellular 3D spheroids of HL-60 human promyelocytic leukemia cells (ATCC CCL-240) were grown using a modified liquid overlay technique. ⁷⁶ Briefly, 5 × 10⁴ cells in 200 μL RPMI 1640 (Mediatech Media MT-10-040-CV) supplemented with 20% FBS (PAA Laboratories, A15-701) were delivered to the inner wells of 96-well microtiter plates (Corning Costar, Acton, MA) coated with 1.5% agarose (Fisher Bioreagents, BP1356-100). The outer wells along the periphery contained 200 µL Dulbecco's phosphate buffered saline (VWR International, CA45000-434) to minimize evaporation from sample wells. One dark plate and a light plate for each irradiation condition were prepared and maintained at 37 °C under 5% CO₂ incubation (Thermo Electron Corp., Forma Series II, Model 3110, HEPA Class 100). The morphological structures and sizes of HL-60 spheroids were confirmed at 40× total magnification using a Nikon inverted microscope (Eclipse TE2000U). When the diameter of the spheroids reached approximately 600 µm (72-96 h), they were dosed with serially diluted PSs in 25 µL aliquots to yield final PS concentrations of 1 nM to 300 μM in the assay. Light plates were irradiated with visible light (7.8 mW cm⁻², 28 J cm⁻²) from a photoreactor (Luzchem LZC-4X), or with 625 nm light (32 mW cm⁻², 100 J cm⁻²) from an LED array made in-house at a PS-to-light interval of 16 h. Dark assay plates were maintained at 37 °C under 5% CO₂ incubator while light plates were irradiated. All plates were incubated for an additional 48 h prior to adding 10 μL aliquots of Alamar blue reagent (Life Technologies DAL 1025) to each well to assess cell viability. Fluorescence from the sample wells was quantified 16 h post Alamar Blue addition using methods described for planktonic cultures (below).

2.4.5 Bacterial culture. *S. mutans* (ATCC 25175) and *S. aureus* (ATCC 25923) cultures were started by suspending half of

the commercially-obtained freeze-dried pellets in 2 mL of tryptic soy broth (TSB) and incubating for 24 h at 37 °C. The bacterial cultures were pelleted, suspended in 5 mL of fresh TSB, and aliquoted (0.5 mL) to 1.5 mL microfuge tubes containing 0.5 mL 70% glycerol in water. These cultures were mixed thoroughly and stored at -80 °C.

2.4.6 Bacterial survival assays. Photodynamic inactivation (PDI) of S. mutans and S. aureus growing as planktonic cultures was probed using a standard broth microdilution method.77 A standard curve of McFarland barium sulfate standards 0.5, 1, 2, 3, 4, and 5 was made, according to a standard method, 77,78 representing approximately 1.5, 3, 6, 9, 12, 15 \times 10⁸ bacterial concentration (CFU mL⁻¹). The absorbance values of the barium sulfate standards (562 nm) was measured, the equation of the trendline was extrapolated, and this was used to quantify the approximate bacterial concentration. On experimental days, a bacterial stock solution was prepared by transferring several bacterial colonies to 2-3 mL sterile water, vortexing well to mix, then reading the absorbance at 562 nm in order to determine the approximate bacterial concentration. An inoculum dilution was then made from the stock at 1×10^6 CFU mL⁻¹ (relative to the established trendline of barium sulfate standards) in fresh TSB. Dark and light experiments were each performed in duplicate in 96-well microplates (Corning Costar 3595), where outer wells along the periphery contained 200 µL of sterile distilled water to prevent evaporation. Cell-free control wells received 100 µL TSB, while control cell wells and sample wells received 100 μ L stock bacterial solution (\sim 1 \times 10⁶ CFU mL⁻¹). The plates were then placed in a 37 °C incubator for at least 30 min to equilibrate.

Serial dilutions of aqueous stock solutions of the Ru compounds were prepared in microcentrifuge tubes in TSB at $2\times$ the concentration needed (final concentrations in the wells were 0.1 nM, 1 nM, 10 nM, 100 nM, 0.1 μM, 1 μM, 10 μM, and 50 μM). Prewarmed 100 μL aliquots of compounds were added to the sample wells (prewarmed TSB to the controls) and final assay volumes were 200 μL (final bacterial concentration \sim 5 \times 10⁵ CFU mL⁻¹). The PS-to-light interval was 1 h. Dark treatment microplates were wrapped in foil and placed in a dark drawer, while PDI-treated microplates were irradiated with visible light (400-700 nm, $40 \pm 0.8 \text{ mW cm}^{-2}$) using a 190 W BenQ MS510 overhead projector or with red light (625 nm, 35 \pm 1.3 mW cm⁻²) from an LED array (Photodynamic Inc.). The irradiation time was 42 min and 48 min respectively, to yield light doses of approximately 100 J cm⁻². Both dark and PDTtreated microplates were incubated overnight. The sample wells were carefully pipetted up and down to mix well and the absorbance at 562 nm was measured for all microplates with a BioTek EL800 plate reader. MIC₅₀ values (the minimum inhibitory concentration at which $\geq 50\%$ of the bacteria is inhibited) for antibiotic (dark) and antimicrobial PDI (light) activity were calculated from sigmoidal fits of the dose response curves using Graph Pad Prism 6.0 according to eqn (2) (above), where γ_i and γ_f are the initial and final absorbance intensities.

Results and discussion

3.1 Synthesis and characterization

We have previously reported the first synthesis of heteroleptic pyrrolyl/2,2'-bypyridyl complexes of ruthenium(II).67 Considering the high stability and unusual UV/vis properties of these mono-ruthenium complexes, we now explore the synthesis and properties of symmetric bis(ruthenium) complexes of this type, with the goal of determining the effect of varying the extent of conjugation in these bis[Ru(II)-pyrrolide] triads. Initial studies concerned the design and synthesis of a mono-pyrrolic ligand bearing extended conjugation, with intent to optimize the synthetic protocol. 67,79 As such, N-Boc-2-vinyl pyrrole (1a)80 was synthesized in a two-step procedure from 2-formyl pyrrole and, following a modified procedure,81 was successfully employed as a Heck substrate with bromobenzene, providing the in situdeprotected styryl-pyrrole 2a in good yield (64%, Scheme 1). Employing 1,4-dibromobenzene as the aryl halide along with 2 equivalents of vinyl-pyrrole 1a resulted in the conjugated, symmetric bis(pyrrole) 2b in high yield (86%).

We then examined the scope of dibromoarene substrates in the double Heck reaction with vinyl pyrrole **1a** (Table 1). A variety of linkers were selected for study, including bicyclic (entries 3 and 4), heterocyclic (entry 5), polycyclic compounds (entries 6–8), and linkers featuring extended conjugation (entries 9–11). The majority of substrates examined were well tolerated, giving bis(pyrrole)s **2b-i** in excellent isolated yields (86–100%). Bithiophene, pyrazine and binaphthyl linkers were unsuccessful in this synthetic screen, as were extended linkers **j** and **k**. A double Suzuki reaction with *N*-Boc-pyrrole-2-boronic acid (**1b**) was subsequently investigated for linkers **j** and **k**, whereupon conditions were developed to generate the corresponding bis(pyrrole)s **2j** and **2k** in yields of 85 and 53%, respectively (entries 10 and 11).

Using mono-pyrrole 2a as a model substrate, Vilsmeier–Haack formylation was found to be successful in installing an α -formyl group, 82,83 providing bidentate ligand 3a in high yield (86%, Table 1, entry 1). Bis(pyrrole)s 2b–2k were subsequently subjected to Vilsmeier–Haack formylation conditions, 84 employing 2 equivalents of phosphoryl chloride, whereby the corresponding bis(bidentate) ligands 3b–3k were isolated in good to excellent yields (76–97%, entries 2–11) following isolation by precipitation in water.

Mono-pyrrolic ligand **3a** was again used as a model substrate for ruthenium complexation, using a previously reported microwave-promoted procedure, ^{67,85} whereupon heteroleptic [Ru(**3a**)(bpy)₂]PF₆ complex salt **4a** was isolated following treatment with aqueous ammonium hexafluorophosphate (96%, Table 2, entry 1). Complexation of bis(bidentate) ligands **3b–3j**, using 2 equivalents of [Ru(bpy)₂Cl₂]·2H₂O and slightly modified reaction conditions, was successful in generating the corresponding bis(ruthenium) complex salts **4b–4j**, (42–86%, entries 2–10), which were purified using column chromatography on neutral alumina. Difficulties were encountered with ligand **3k**, which underwent complexation and concomitant reduction of the central double bond of isoindigo linker **k**. This was thought

Scheme 1 Synthesis of conjugated pyrrole 2a and bis(pyrrole) 2b via Heck reaction.

to be an effect of the ethylene glycol solvent, which is known to oxidize during heating in air to generate the reductant glycolaldehyde. Altering the reaction solvent to 9:1 methanol: water overcame this problem and allowed for isolation of the desired complex salt 4k (70%, entry 11). For the purpose of assessing the photobiological activity of each bis[Ru(II)-pyrrolide] triad, salt conversion of the hexafluorophosphate salts (4a–4k) to the water-soluble chloride salts (5a–5k) was carried out by treatment with tetrabutylammonium chloride (TBAC) in acetone. The salts of the salts (5a–5k) was carried out by treatment with tetrabutylammonium chloride (TBAC) in acetone.

3.2 Spectroscopic properties

The MeCN-soluble PF $_6$ ⁻ salts of the complexes (4a-k) were used for all spectroscopic measurements, while the water-soluble Cl $^-$ salts of the complexes (5a-k) were used for biological studies. The reason MeCN was used as the solvent of choice for spectroscopy (instead of water or other aqueous solution) is that water quenches the $^1{\rm O}_2$ emission, precluding accurate determination of the upper limit for $^1{\rm O}_2$ quantum yields⁸⁸ and because MeCN is the solvent used in many published spectroscopic studies.

3.2.1 Absorption. The electronic absorption properties of bis[Ru(II)-pyrrolide] triads 4b-k (and their corresponding ligands) and mononuclear 4a were investigated in MeCN (Fig. 1a-c and Table 1) and analyzed in the context of the wellstudied Ru(II) polypyridyl complexes.21 Ru(II) polypyridyl complexes such as [Ru(bpy)₃]²⁺ typically display absorption spectra that are characterized by two distinct regions in the UV and visible, respectively: (i) intense and sharp bands corresponding to singlet intraligand ${}^{1}\pi\pi^{*}$ transitions below 300 nm that are localized to the polypyridyl ligands, and (ii) much broader, lower-intensity bands corresponding to singlet metal-to-ligand charge transfer (¹MLCT) transitions between 400 and 500 nm that involve charge transfer from the Ru($d\pi$) orbitals to the π^* orbitals of the ligand(s). While the Ru(II) complexes in our study contain two polypyridyl ligands, the third ligand is an extremely π -delocalized system that in some cases could have significant intraligand charge transfer (ILCT) character due to highly polarizable groups (e.g., 3e, 3j-k). In addition, with respect to each Ru(II) center in the bis[Ru(II)-pyrrolide] triad, this symmetric third ligand is chelated to the second Ru(II) center which could further impact the character of these transitions. It was expected that the absorption spectra of the target complexes would show contributions from these novel ligands that would be influenced by their proximities to the two Ru(II) metal centers.

The absorption spectra of the free ligands are shown in Fig. 1a. For those ligands derived from (poly)cyclic aromatic hydrocarbon linkers (3b-d, 3f-h), the longest wavelength absorption maxima mirrored the ${}^1\pi\pi^*$ transitions characteristic of the linker, but with bathochromic shifts and contributions arising from extended π -conjugation with the vinylappended 2-formyl pyrrolides. For example, free pyrene has longest-wavelength absorption maximum just below 350 nm, 89 whereas 3h, with pyrene as the linker, had its longestwavelength absorption maximum near 448 nm, with a shoulder at 489 nm (≥100 nm red-shift relative to free pyrene). Notably, this significant bathochromic shift places the spectral window of the ${}^1\pi\pi^*$ transition of ligand 3h in a similar position as the ¹MLCT transition of $[Ru(bpy)_3]^{2+}$ ($\lambda_{max} = 448$ nm). The longestwavelength absorption maxima of 3e and 3j-k, with predicted ligand-localized contributions, are even more red-shifted, appearing at wavelengths ≥ 500 nm ($\lambda_{\text{max}} = 593$ nm for 3k). It was anticipated that chelation of these unique π -expanded ligands to Ru(II) to form the bis[Ru(II)-pyrrolide] triads would further widen the visible spectral window and lead to enhanced molar extinction coefficients, especially at the longer wavelengths.

The UV/Vis absorption spectrum of our previously reported 2-formyl pyrrolide Ru(II) complex 6,67 representative of the core mononuclear N,O-coordinated system used in the triads but without extended conjugation, is compared to $[Ru(bpy)_3]^{2+}$, mononuclear **4a**, and bis[Ru(II)-pyrrolide] **4b** in Fig. 1b. Complex 6 was the first published example of a heteroleptic pyrrolide/2,2'-bipyridyl Ru(II) complex. This simple mononuclear construct displays continuous absorption between 200 and 600 nm, with a longest-wavelength absorption maximum near 528 nm for the ¹MLCT transition, which is approximately 80 nm longer than that for $[Ru(bpy)_3]^{2+}$. Redshifts of almost 100 nm for the lowest-energy ¹MLCT transitions (relative to the corresponding Ru(II) systems containing neutral diimine ligands) agrees with what we have previously observed for Ru(II) complexes bearing anionic cyclometalating ligands, such as thionoester-substituted pyrrolides and deprotonated phenylpyridines. 55,57,67,79 Presumably, this shift of the ¹MLCT absorption band is a direct result of a concomitant increase in the energy of the $Ru(d\pi)$ orbitals arising from the strong N- $\sigma(\eta^1)$ donation of the pyrrolide nitrogen.

The styryl substituted pyrrolide complex (4a) led to significant absorption past 500 nm ($\varepsilon_{510} = 1.1 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$) and doubled the extinction coefficients in this region compared to 6 (Fig. 1b). The slight blue-shift of about 13 nm for the longest-

Table 1 Synthesis of a novel series of bis(pyrrolic) ligands (3a-3k)

R: CH=CH ₂ 1a or B(OH) ₂ 1b	izuki /n Linker	1. POCl ₃ , DMF, Ar, 0 - 60 °C, 2 h 2. 5% K ₂ CO ₃ (aq.), 80 °C, 2 h	N Linker	H O
Durrole I	inker	11	Viold of 2 ^b (0%)	Viel

	0. 5(0.17)2					
Entry	Pyrrole	Linker		n	Yield of 2^b (%)	Yield of 3 ^b (%
1	1 a	\mathbf{a}^a	§ — ()	1	64 (2a) ^{<i>a,c</i>}	86 (3 a) ^{a,g}
2	1 a	b	}	1	86 (2b) ^d	85 $(3b)^h$
3	1a	c	}	1	94 (2c) ^e	85 $(3\mathbf{c})^h$
4	1a	d	\$	1	97 (2 d) ^d	76 (3 d) ^h
5	1a	e	N.S.N \$	1	91 (2e) ^d	97 (3e) ^h
6	1a	f		1	97 (2f) ^d	87 (3f) ^h
7	1a	g		1	100 (2g) ^d	94 (3g) ^h
8	1a	h	profes	1	100 (2 h) ^d	95 (3h) ^h
9	1a	i	N.S.N	1	100 $(2\mathbf{i})^d$	92 (3 i) ^h
10	1b	j	N.S.N.N.Y.	0	85 (2 j) ^e	84 $(3\mathbf{j})^h$
11	1b	k	R = 2-Et-hexyl	0	53 (2k) ^{e,f}	90 $(3\mathbf{k})^h$

^a Compounds **2a** (see Scheme 1) and **3a** are mono-pyrroles (pyrrole-CH = CHPh). ^b Isolated yield. ^c Heck reaction conditions: 1 equiv. **1a**, Pd(OAc)₂, 2,4-pentanedione, K_2CO_3 , DMF, Ar, 130 °C, 3 h. ^d Heck reaction, 2 equiv. **1a**, 6 h. ^e Suzuki reaction conditions: Pd(PPh₃)₄, K_2CO_3 , DMF, 110 °C, 24 h. ^f Suzuki reaction, 115 °C for 18 h then 125 °C for 5 h. ^g Vilsmeier reaction, 1 equiv. POCl₃. ^h Vilsmeier reaction, 2 equiv. POCl₃.

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wavelength absorption maximum for **4a** could reflect the enhanced conjugation of the pyrrolide ligand and weaker N- σ (η^1) bonding to the Ru(π) center. Nevertheless, the extended conjugation provided by the styryl group in combination with the relatively strong N- σ donation of the N,O pyrrolide resulted in a Ru(π) complex that absorbs green light ten times more strongly than the related [Ru(bpy)₃]²⁺ complex. In support of our hypothesis that these properties could be improved further, incorporation of two metal chromophores into a triad *via* two terminal 2-formylpyrrolyl ligands tethered to a central benzene linker through alkenyl groups (**4b**) resulted in a four-fold increase in the longest wavelength absorption maximum in comparison to its mononuclear counterpart **4a**, and 40-fold relative to the parent [Ru(bpy)₃]²⁺.

The absorption spectrum of the bis[Ru(II)-pyrrolide] complex 4b appeared to be more than a simple linear combination of two mononuclear fragments and the free organic ligand, thereby suggesting that the two metal centers are in conjugative communication mediated by the shared organic linker. This notion is supported by the observation that the longestwavelength absorption maximum measured for the corresponding complex with a biphenyl linker (4c), which most likely adopts a nonplanar dihedral angle and decouples the two metal centers, is blue-shifted and of reduced intensity relative to both 4a and 4b. The other explored linkers can be structurally grouped as follows: polycyclic aromatics (4d, 4f-h), heterocycles based on benzothiadiazole (4e, 4i-j), or isoindigo (4k). Of all of the complexes, the pyrenyl linker (4h) exhibited the most intense transitions at its longest-wavelength absorption maximum, while the benzothiadiazole (4e) and isoindigo (4k) linkers yielded the longest-wavelength absorption maxima

overall (albeit of reduced intensity relative to **4h**). The absorption spectra of mononuclear **4a** and the ten bis[Ru(11)-pyrrolide] complexes are compared in Fig. 1c.

Generally, complexation of the respective novel ligand 3 to two Ru(II) centers to produce the bis[Ru(II)-pyrrolide] triads 4 resulted in both a widening of the visible absorption window as well as a noticeable hyperchromic shift at these wavelengths for all bis [Ru(II)-pyrrolide] triads except for 4i and 4j. The longest-wavelength absorption bands in 4i were very similar to 3i, and in 4j, the free ligand was more absorptive at the longer wavelengths despite what appeared to be a longer wavelength absorption maximum for its complex. Notably, for the benzothiadiazoles (4e, 4i-j), the groups on either side of the benzothiadiazole had a marked impact on the longest wavelength transitions. For example, vinyl groups directly attached to the central benzothiadiazole group (4e) led to a longest wavelength absorption maximum near 615 nm, which was among the longest in the entire series. Adding phenyl groups between the benzothiadiazole and the vinyl groups (5i) or replacing the vinyl groups with N-methyl pyrrole groups shifted these bands hypsochromically by ≥100 nm. Clearly, there is much to be learned from these SARs and what they suggest in terms of the polarizabilities and CT characters of the ligands and their resulting bis [Ru(II)-pyrrolide] complexes, but the purpose of the present investigation was to provide a very general outline of these observations.

3.2.2 Emission. Mononuclear **4a** and the bis[Ru(π)-pyrrolide] complexes **4b-4k** did not phosphoresce at room temperature under ambient oxygen conditions and very little phosphorescence was observed at room temperature in an argon atmosphere (Fig. 1d and Table 3). The largest phosphorescence quantum yields (Φ_p) were only about 0.1%, but the signal for eight of the eleven complexes was sufficient to identify

Table 2 Bis(ruthenium) complexation of ligands 3b-3k

Entry	Linker	n	Yield of 4^{b} (%)	Yield of 5 ^b (%)
1	\mathbf{a}^a	1	96 (4a) ^{<i>a,c</i>}	83 (5a) ^a
2	b	1	$86 \left(\mathbf{4b}\right)^d$	73 (5b)
3	c	1	$61 \left(\mathbf{4c}\right)^d$	72 (5c)
4	d	1	$42 \left(\mathbf{4d}\right)^d$	78 (5d)
5	e	1	$61 (4e)^d$	71 (5e)
6	f	1	$45 \ (\mathbf{4f})^d$	52 (5f)
7	g	1	$66 \left(\mathbf{4g}\right)^d$	75 (5g)
8	h	1	69 $(4h)^d$	83 (5 h)
9	i	1	$72 \left(\mathbf{4i}\right)^d$	75 (5i)
10	j	0	$62 \ (4j)^d$	93 (5j) ^f
11	k	0	70 $(4k)^{d,e}$	$84 (5k)^f$

 $[^]a$ Compounds 4a and 5a feature mono-pyrrolide ligands and one Ru centre (pyrrole-CH = CHPh). b Isolated yield. c 1 equiv. Ru(bpy) $_2$ Cl $_2 \cdot 2$ H $_2$ O with reaction time of 60 min. d 2 equiv. Ru(bpy) $_2$ Cl $_2 \cdot 2$ H $_2$ O. e Reaction solvent 9:1 methanol: water. f Reaction solvent 10:1 acetone: hexanes.

discernable maxima for the 3 MLCT emission near 743 nm with a longer wavelength shoulder near 800 nm (using the excitation maxima, which occurred near 465–485 nm). For the phosphorescence that was detectable, the various ligands and linkers had little influence on the energy of the emitting 3 MLCT state, which likely involves π^* acceptor orbitals of the bpy ligands, except for 4e and 4h. Complexes 4e and 4h did not yield any phosphorescence, although the tail of their shorter wavelength ligand-centered fluorescence could be discerned in the spectral observation window. While 4k exhibited very weak phosphorescence, a value for Φ_p was not determined due to the lack of a discrete peak. Collectively, the low phosphorescence quantum yields (or absence of phosphorescence) for all of the compounds point toward other efficient relaxation pathways that facilitate excited state decay even in the absence of oxygen.

3.2.3 Singlet oxygen quantum yields. In the presence of oxygen, mononuclear 4a and the bis[Ru(II)-pyrrolide] complexes 4b-4k generated $^{1}O_{2}$ to varying degrees. The $^{1}O_{2}$ quantum yields (Φ_{Δ}) ranged from as low as 5-7% for 4k and 4f, respectively, to as high as 77% for 4i (Table 3). According to their Φ_{Δ} values, the compounds clustered into three groups: (i) 5-13% (4a > 4f > 4k), (ii) 30-40% (4g = 4j > 4e), and (iii) >50% (4i > 4b \approx 4h > 4d > 4c). With the exception of 4i (benzothiadiazole flanked by two

phenyl groups), the compounds with the largest $^{1}O_{2}$ quantum yields were those with phenyl, biphenyl, or polycyclic aromatic hydrocarbon (pyrenyl and naphthalene) linkers. Anthracene as the central linker (4f) was among the poorest $^{1}O_{2}$ generators of the group ($\Phi_{\Delta}=7\%$), and fluorene (4g) was near the middle ($\Phi_{\Delta}=37\%$). Whether the 3 MLCT state(s), observed in the emission measurements, contributed to $^{1}O_{2}$ production remains unknown but it is anticipated that non-emissive 3 IL or 3 ILCT states may play a role with regard to the more highly photosensitizing systems. It was anticipated that compounds with the higher $^{1}O_{2}$ quantum yields might act as PDT agents so we next investigated their cytotoxicities toward cancer cells with light activation, and compared to their dark cytotoxicities.

3.3 Photobiological activity

3.3.1 HL-60 cytotoxicity and photocytotoxicity assays for the series

Cellular assays. The water-soluble Cl⁻ salts (5a-k) were used for the biological experiments. The dark cytotoxicities of the mononuclear reference compound 5a and the bis[Ru(II)-pyrrolide] triads 5b-5k were determined using a human leukemia (HL-60) cell line. This cell line was chosen because it grows as a suspension rather than an adherent monolayer, thus

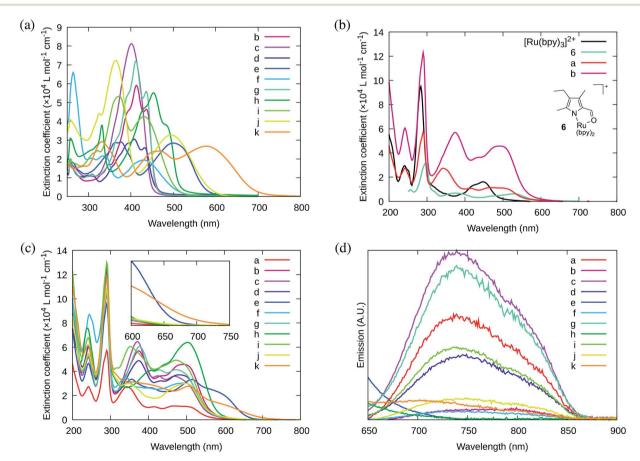


Fig. 1 UV/Vis absorption spectra of (a) ligands 3b-k; (b) previously reported 6 and $[Ru(bpy)_3]^{2+}$ as reference complexes for mononuclear 4a and bis $[Ru(\shortparallel)$ -pyrrolide] triad 4b; and (c) mononuclear 4a and bis $[Ru(\shortparallel)$ -pyrrolide] triads 4b-k. (d) Phosphorescence emission spectra for mononuclear 4a and bis $[Ru(\shortparallel)$ -pyrrolide] triads 4b-k (collected in Ar using $\lambda_{ex\ max}$). Absorption and emission spectra were collected on the PF $_6$ salts of the complexes (5 μ M) in MeCN.

Table 3 Spectroscopic properties

Cmpd	$\mathrm{Abs_{max}}/\mathrm{nm} \; (\mathrm{log} \; arepsilon)$	$\lambda_{\rm em\ max}^{a}(\lambda_{\rm ex})/nm$	$\Phi_{\mathrm{p}}^{}a}\left(1 imes10^{-3} ight)$	Φ_{Δ}
5a	244 (4.43), 284 (4.70), 290 (4.76), 340 (4.44), 416 (4.02), 464 (4.04), 514 (3.99)	743 (466)	1.07	0.13
5 b	242 (4.78), 284 (5.04), 290 (5.09), 378 (4.75), 434 (4.56), 494 (4.66), 515 (4.60)	760 (500)	0.10	0.69
5 c	244 (4.70), 284 (4.96), 290 (5.02), 372 (4.81), 428 (4.56), 470 (4.56), 504 (4.47)	743 (470)	1.20	0.57
5d	244 (4.77), 284 (4.99), 288 (5.02), 376 (4.69), 436 (4.50), 484 (4.57)	750 (484)	0.52	0.61
5e	244 (4.66), 282 (4.90), 290 (4.98), 360 (4.52), 414 (4.41), 518 (4.52), 602 (4.29)	b	<u></u> b	0.32
5f	248 (4.93), 252 (4.91), 284 (5.02), 290 (5.07), 340 (4.55), 404 (4.42), 472 (4.47), 514 (4.44)	765 (495)	0.067	0.07
5g	206 (4.91), 244 (4.72), 284 (4.97), 290 (5.02), 378 (4.78), 430 (4.61), 474 (4.62), 502 (4.56)	743 (475)	0.69	0.37
5h	240 (4.88), 290 (5.11), 406 (4.61), 442 (4.66), 508 (4.80)	b	<u></u> b	0.68
5i	242 (4.83), 292 (5.11), 318 (4.63), 354 (4.78), 476 (4.69), 510 (4.60)	738 (474)	0.68	0.77
5j	244 (4.73), 290 (5.03), 316 (4.53), 352 (4.47), 438 (4.30), 504 (4.47)	746 (500)	0.28	0.33
5k	242 (4.79), 292 (5.06), 398 (4.47), 510 (4.44), 618 (4.02)	715 (507)	<u></u> c	0.05

^a 298 K, Ar. ^b Emission from the ³MLCT state at 298 K was not observed (the tail of ¹LC emission was observed). ^c Very weak ³MLCT emission that was continuous over the observation window.

eliminating some additional variability in the cellular assay that arises when treating differentially formed monolayers. Briefly, cells growing in log phase were dosed with the compounds at concentrations between 1 nM and 300 µM and assessed for viability after approximately 64 h using the Alamar Blue reagent. The photocytotoxicities were determined in an analogous manner except that a light treatment was delivered approximately 16 h after the cells were dosed with compound. The cell viability was quantified from dose-response curve fits to yield the effective concentration required to reduce cell viability by 50% (EC₅₀) in the dark (dark EC₅₀) and with the light treatment (light EC_{50}). The phototherapeutic indices (PIs) were calculated as the ratios of the dark EC50 and light EC50 values, and represent the amplification of the cytotoxic effect with the light trigger. All cellular assays were carried out in triplicate under normoxic conditions, with representative data compiled in Table 4. For reference, the well-known cytotoxic chemotherapy agent cisplatin yields an EC50 value of approximately 25 µM with no difference between the dark and light condition (PI = 1).

Dark cytotoxicity. The dark cytotoxicities of the compounds investigated varied over two orders of magnitude from approximately 1.7 μM for the mononuclear $\bf 5a$ to just over 170 μM for the bis-Ru(II) triad $\bf 5e$ (Table 4, Fig. 2a). Notably, the mononuclear compound $\bf 5a$ was distinctly more cytotoxic than its triad counterparts, being seven-fold more cytotoxic than the most dark cytotoxic triad $\bf 5k$ (dark $EC_{50} = 11.5$ μM). There was a ten-fold variation among the Ru(II) triads that clustered into roughly three groups: least cytotoxic with dark EC_{50} values near 100 or more ($\bf 5b$, $\bf 5d$ -e), moderately cytotoxic with values near $\bf 30$ – $\bf 50$ μM ($\bf 5c$, $\bf 5f$ -h, $\bf 5j$), and cytotoxic with values between $\bf 10$ – $\bf 15$ μM ($\bf 5i$, $\bf 5k$).

Structurally, the bis[Ru(II)-pyrrolide] systems can be divided into three classes: (i) those with aromatic hydrocarbon linkers that vary in the extent π -conjugation (5**b**-**d**, 5**f**-**h**), (ii) those with benzothiadiazole linkers with or without conjugated groups (5**e**, 5**i**-**j**), and (iii) one with an isoindigo linker (5**k**). The dark cytotoxicities of class (i) varied from 32 to 103 μ M, while those for class (ii) varied from 14 to 173 μ M. Complex 5**k** with the isoindigo linker was the most cytotoxic at 11.5 μ M, and 5**e** with the benzothiadiazole linker was the least at 173 μ M. Interestingly, incorporation of phenyl rings (5**i**) or N-Me pyrrole rings (5**j**) on

either side of the benzothiadiazole group led to increased cytotoxicity relative to the parent **5e**. Likewise, there was a substantial difference between incorporation of one phenyl ring (**5b**) as the linker and two (**5c**), with the latter resulting in elevated cytotoxicity. The incorporation of two *fused* rings, as in naphthalene (**5d**), resulted in a slightly reduced cytotoxicity relative to **5b**.

Parameters such as lipophilicity and cellular uptake and distribution were not investigated as part of this study so it would be premature to speculate on reasons behind the observed differences in cytotoxicity. Rather, our intention here is to highlight the breadth of cytotoxic activity that can be obtained in a relatively small structural family of a new compound class and to also use the dark EC_{50} values as a reference point for assessing phototoxic effects and corresponding PIs. This significant variation within and between the classes underscores that the linker unit is an important point of variation for manipulating the inherent cytotoxicity of bis[Ru(π)-pyrrolide] triads, which could prove advantageous for optimization of PI values.

Photocytotoxicity. The photocytotoxicities of mononuclear **5a** along with the bis[Ru(II)-pyrrolide] triads were determined with broadband visible light (28 J cm $^{-2}$, 7.8 mW cm $^{-2}$) and with 625 nm red light (100 J cm $^{-2}$, 42 mW cm $^{-2}$) (Fig. 2a and Table 4). Their visible light EC $_{50}$ values under this condition varied by just over three orders of magnitude, ranging from approximately 3–11 μM for the least phototoxic systems (**5f**, **5j–k**) to 10–70 nM for the most potent phototoxic compounds (**5g**, **5h**). Other family members clustered near 150–270 nM (**5a**, **5c–d**, **5i**), with **5b** and **5e** much closer to 1 μM.

Because the light EC₅₀ values contain contributions from the baseline dark cytotoxicity, the true phototoxic effects were assessed as PI values, or fold-amplification between the dark and light condition (Fig. 2b and Table 4). According to their PIs, the compounds could be grouped by having (i) very little phototherapeutic effect with PIs <<100 (5a, 5f, 5j-k), (ii) marginal effects with PIs near 100–200 (5b-c, 5e, 5i), or (iii) very good effects with PIs >>200 (5d, 5g, 5h). Bis[Ru(II)-pyrrolide] 5h, exhibiting one of the larger 1 O₂ quantum yields, stood out from the rest with its visible PI exceeding 5000 using this relatively soft light dose. The PIs generally correlated with 1 O₂ quantum

261

39

2

5h

5i

5i

5k

Red PI^b Complex Dark EC50 (µM) Vis light^a EC₅₀ (µM) Vis PIb Red light^c EC₅₀ (µM) 1.69 ± 0.06 0.20 ± 0.01 8 0.29 ± 0.07 6 5a 89.1 ± 0.8 5b 0.55 ± 0.02 161 1.20 ± 0.03 74 5c 31.6 ± 1.7 $\textbf{0.27} \pm \textbf{0.04}$ 0.79 ± 0.04 40 115 5d 103 ± 0.6 0.19 ± 0.01 534 0.84 ± 0.02 123 $173\,\pm\,6$ $\textbf{0.84} \pm \textbf{0.01}$ 206 $\textbf{0.73}\,\pm\,\textbf{0.02}$ 237 5f 48.1 ± 0.4 $3.05\,\pm\,0.21$ 16 4.06 ± 0.09 12 5g 54.4 ± 0.9 0.07 ± 0.01 734 0.35 ± 0.02 157

Table 4 Compilation of the dark cytotoxicities and photocytotoxicities of 5a-5k toward HL-60 cancer cells

 0.01 ± 0.01

 0.15 ± 0.01

 10.8 ± 0.3

 6.36 ± 0.14

5439

95

yields across the series (Fig. 3a), but the correlation was not strict when comparing individual compounds. For example, 5h had a much larger PI than the other family members (best emphasized in Fig. 3b), yet it did not have the largest ${}^{1}O_{2}$ yield of the series. Certainly, other ROS and other phototoxic mechanisms could be at play, the cell-free ${}^{1}O_{2}$ quantum yields may not reflect the cellular ${}^{1}O_{2}$ quantum yields, and/or the subcellular targets may have a larger impact on the PI than the precise ${}^{1}O_{2}$ quantum yield. Nevertheless, this compound class can be considered a new source of PSs for PDT.

 36.8 ± 2.9

 14.3 ± 0.4

 39.8 ± 0.9

 11.5 ± 0.3

Structurally, the largest PIs were observed for the bis[Ru(π)-pyrrolide] systems with conjugated aromatic hydrocarbon linkers in the order: pyrene (5h) > fluorene (5g) > naphthalene (5d). The smallest PIs were obtained for the mononuclear 5a, which had very high dark cytotoxicity, and the bis[Ru(π)-pyrrolide] triads with anthracene (5f), isoindigo (5k), and bis(NMePy) benzothiadiazole (5j) as central linkers. The family members with intermediate and similar PIs contained phenyl and biphenyl linkers (5b), and (5c), respectively, as well as benzothiadiazole and diphenylbenzothiadiazole linkers (5e), and (5i), respectively. It is tempting to speculate that linkers with the requisite triplet state energies to act as excited state reservoirs might lead to increased sensitivity to oxygen (and other excited state quenchers) in these triads and thus larger PIs. However, as

triplet state energies of the free ligands (and the corresponding ³IL or ³ILCT energies of the complexes) form part of a future extensive spectroscopic study we will not speculate at this time.

 0.14 ± 0.01

 0.37 ± 0.05

 6.48 ± 0.16

 10.2 ± 0.1

The photocytotoxicities and PIs for the bis[Ru(II)-pyrrolide] triads were also measured using a slightly stronger broadband visible light dose (100 J cm⁻², 28 mW cm⁻²) from a different light source to cross-confirm the phototoxic effects across the series (Fig. 2c and 3b). The difference in light fluence or irradiance between the two experiments was almost four-fold, and the resulting PIs did not scale linearly with this change. However, the compounds clustered in the same groups based on their PIs and ¹O₂ quantum yields (Fig. 3b). The PI differences between the two visible light conditions were compounddependent, ranging from two-fold (5j) to sixteen-fold (5i). Differences near ten-fold (5b-d and 5f) or five-fold (5e, 5g-h, 5k) were measured for the rest of the family. Notably, 5h had a visible EC₅₀ value near 1 nM and PI > 27 000, while the PI values for 5d and 5g were >6000 and >3500, respectively. 5h has one of the larger PI reported to date (Fig. 4).

Since mononuclear 5a and the bis[Ru(π)-pyrrolide] systems display longest-wavelength absorption maxima that are redshifted compared to many well-studied Ru(π) polypyridyl complexes,²¹ their photocytotoxicities and PIs were also investigated using 625 nm red LEDs (100 J cm⁻², 42 mW cm⁻²). As

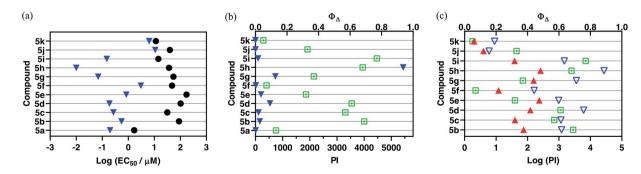


Fig. 2 (a) Activity plot for 5a-5k showing cytotoxicities in the dark (\blacksquare) and with light activation using broadband visible light (\blacktriangledown , 28 J cm⁻², 7.8 mW cm⁻²); (b) activity plot for 5a-5k highlighting phototherapeutic indices (PIs) under the same light conditions as in (a), as well as ${}^{1}O_{2}$ quantum yields (\blacksquare); and (c) activity plot for the bis[Ru(\parallel)-pyrrolide] triads 5b-5k showing their log PI values with visible (\blacktriangledown , 100 J cm⁻², 28 mW cm⁻²) or 625 nm red (\blacktriangle , 100 J cm⁻², 42 mW cm⁻²) light.

 $[^]a$ Vis condition: 16 h DLI followed by broadband visible light irradiation (28 J cm $^{-2}$, 7.8 mW cm $^{-2}$). b PI = phototherapeutic index (ratio of dark EC $_{50}$ to visible-light EC $_{50}$). c Red condition: 16 h DLI followed by light irradiation with 625 nm LEDs (100 J cm $^{-2}$, 42 mW cm $^{-2}$).

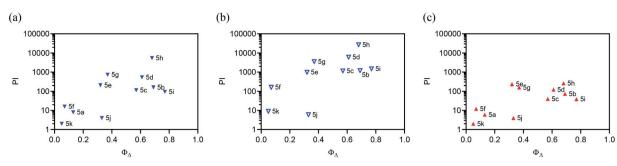


Fig. 3 Plots correlating Pl value with the $^{1}O_{2}$ quantum yield for each complex under three different light conditions: (a) broadband visible (\bigvee , 28 J cm $^{-2}$, 7.8 mW cm $^{-2}$) for 5a–k, (b) broadband visible (\bigvee , 100 J cm $^{-2}$, 28 mW cm $^{-2}$) for 5b–k, and 625 nm red (\triangle , 100 J cm $^{-2}$, 42 mW cm $^{-2}$) for 5a–k.

observed for the two different visible light treatments, the compounds clustered in the same groups based on their PIs and ¹O₂ quantum yields (Fig. 3c), but their PIs were attenuated. The red PIs ranged from 2 for the least photoactive compound (5k) to 260 for the most photoactive system (5h) (Table 4), with four of the triads maintaining PIs > 100 (5d-e, 5g-5h). The visibleand red-light treatments with a fluence of 100 J cm⁻² (but different irradiances) are compared in Fig. 2c. The PIs for the bis[Ru(II)-pyrrolide] triads were attenuated to different extents using lower-energy red light, from 100-fold for 5h to two-fold with 5j. The order of attenuation appeared to parallel the magnitudes of the PIs with visible light rather than the molar extinction coefficients at 625 nm, with the more photoactive compounds being the most affected. Of the compounds considered most active under all three illumination conditions investigated, only **5e** absorbs red light significantly (log $\varepsilon_{625 \text{ nm}}$ = 4.08) yet 5h (log $\epsilon_{\rm 625~nm}$ = 2.93) had a larger PI. The only other compound that absorbs light substantially at 625 nm is 5k (log $\varepsilon_{625 \text{ nm}} = 3.97$), which was dark cytotoxic and considered relatively non-phototoxic under all light conditions explored. These variances present intriguing launch points for future investigation.

3.3.2 Selected assays to investigate the scope of activity for bis[Ru(II)-pyrrolide] 5h

Wide concentration range photocytotoxicity assay. The visible light condition with a fluence of 100 J cm⁻² described above yielded an EC_{50} value for $5\mathbf{h}$ near 1 nM, which was the lowest concentration tested in that assay. To gain more insight regarding the visible light EC_{50} value with 100 J cm⁻², we rescreened $5\mathbf{h}$ starting at 100 pM and reduced the drug-to-light (DLI) interval from 16 h to 1 h (Fig. 5a). This new condition yielded a visible-light EC_{50} value for $5\mathbf{h}$ of 1.33 nM (PI = 24 100).

The PI was slightly reduced in this assay due to a higher dark cytotoxicity of 30.8 μ M (*versus* 36.8 μ M in the narrower range screen). In parallel, we also used red light (625 nm, 100 J cm⁻², 29 mW cm⁻²) and obtained a red light EC₅₀ value of 129 nM (PI = 239), which was similar to what was determined in the narrower concentration range assay.

Optimization of the red-light PI. Given that bis[Ru(II)-pyrrolide] 5h clearly emerged as a compound of interest for further investigation due to its unprecedented visible PI with both the high and low light fluences tested, we wondered whether the attenuated red-light PIs of ~240-260 obtained with a fluence of 100 J cm⁻² could be improved. The light parameter offers a unique opportunity to optimize the PI as the wavelength, fluence, irradiance, DLI, and dosing regimen can be manipulated. While the optimal light dosimetry parameters are not absolute and most certainly are compound-dependent, simple changes such as increasing the fluence and dosing interval are straightforward. We optimized the PI for 625 nm red light (100 J cm^{-2} , 29 mW cm^{-2}) with a 16 h DLI, where the red EC₅₀ value in this assay was 161 nM and the PI was 195 (Fig. 5b). These unoptimized values differ slightly between assays16 so the reference condition was always run in parallel for comparison. Delivering the same total fluence but in four 25 J cm $^{-2}$ intervals separated by 15 min increased the potency by almost four-fold (red $EC_{50} = 45.7$ nM, PI = 690). Increasing the light fluence to 200 J cm⁻² delivered in two intervals of 100 J cm⁻² separated by 1 h led to subnanomolar potency: red $EC_{50} = 630$ pM and PI = 50 000 (Fig. 5b). The superior potency with this light regimen exceeded even that of the visible light condition that yielded a PI > 27 000. PIs of these magnitudes have not been reported. This very limited optimization study underscores how the light regimen can compensate for marginal extinction coefficients at

Fig. 4 Molecular structures of the bis[Ru(II)-pyrrolides] with the largest PIs.

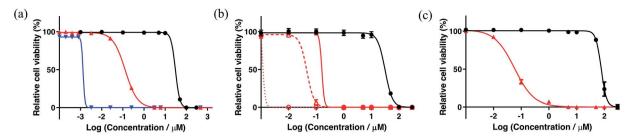


Fig. 5 (a) Wide concentration range dark/light cytotoxicity assay performed with 5h using the HL-60 cell line. Cells dosed with 5h received a dark (black) or light treatment with red (625 nm LEDs, red) or broadband visible (blue) light (100 J cm⁻², 29 mW cm⁻²) with a DLI of 1h. (b) Cytotoxicity (black) and photocytotoxicity (red) using the three 625 nm red light conditions: the red light dose used in (a) but with different concentrations of 5h (—); 100 J cm^{-2} (29 mW cm⁻²) delivered in four 25 J cm⁻² fractions separated by 1h (••). (c) HL60 multicellular 3D spheroid cytotoxicity (black) and photocytotoxicity (red) assay with 1h using the red light condition described for (a).

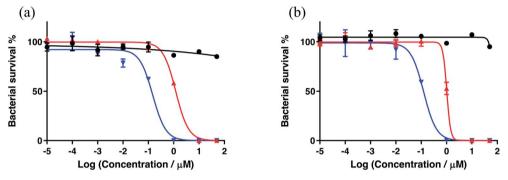


Fig. 6 In vitro cytotoxic effects of 5h against S. mutans (a) and S. aureus (b) growing as planktonic cultures in the dark (black) or with a light treatment. The light treatments were broadband visible (blue) or 625 nm red (red) light (100 J cm $^{-2}$, 28 mW cm $^{-2}$) with a DLI of 1 h.

the activation wavelength. In this preliminary investigation, we did not investigate the mechanism behind this improved response as part of this study, but it is known that fractionated dosing can (in some cases) improve response.¹²

Multicellular 3D tumor spheroid assay. The 3D multicellular tumor spheroid model can be exploited to mimic the highly plastic migratory/invasive tumor phenotypes that characterize some of the most aggressive conditions in vivo.90 For instance, they have hard-to-reach hypoxic regions that impart drug resistance. To test whether 5h could maintain potency against tumor spheroids of the same cell line used for the 2D suspension assays (HL-60), spheroids were grown to sizes of about 600 μm in diameter and treated with **5h** in the concentration range of 1 nM to 300 μ M. The spheroids were either kept in the dark or treated with 625 nm red light (100 J cm⁻², 29 mW cm⁻²) with a 16 h DLI. As expected the HL-60 tumor spheroids were greater than two-fold more resistant to 5h in the dark (compared to 2D HL-60 cultures), with a dark EC_{50} of approximately 77 μ M. Surprisingly, however, the photocytotoxicity was greater against the 3D tumor spheroids, with a red-light EC₅₀ value of 60 nM and PI > 1200. We did not examine the source of this enhanced photocytotoxicity in the 3D tumor spheroid model, which should be scrutinized more closely across spheroids of different sizes and of different cell lines to assess whether this is a general property of 5h.

Bacterial survival assays. The ability of 5h to act as a photocytotoxic compound toward bacteria was briefly explored. Two bacterial species were grown as planktonic cultures and treated with 5h in the concentration range of 10 pM to 50 μM, where no dark cytotoxicity was apparent. Further treatment with either broadband visible or 625 nm red light (100 J cm⁻², 28 mW cm⁻²) using a DLI of 1 h resulted in phototoxic effects toward both S. mutans and S. aureus (Fig. 6). There was no selectivity for either bacterial species, with visible EC_{50} values on the order of 130 to 160 nM and PIs >300 (PIs not determined because there was no dark cytotoxicity at the concentrations investigated). As observed with the HL-60 cells, the photocytotoxicity was attenuated upon moving to the use of red light, rather than visible light of the same fluence and irradiance. The reduction was approximately eight-fold, yielding PIs > 40-50. This result confirms that the phototoxic effect exhibited by 5h extends to other types of cells and that this new class of bis[Ru(II)-pyrrolide] triad shows potential for use as photoactive antimicrobials.

4. Concluding remarks

In summary, the ten new bis[Ru(11)pyrrolide] triads reported herein demonstrated unequivocally that the central organic linker plays a pivotal role in determining the spectroscopic, biological, and photobiological properties of the metal-organic

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systems and that these properties in many cases are improved relative to the mononuclear counterpart 5a. The compounds demonstrated a large breadth of activity as exemplified by a very wide range for $^1\mathrm{O}_2$ quantum yields, dark cytotoxicities, and PIs. Simple variation of the central organic chromophore resulted in some compounds being excellent *in vitro* phototoxic agents, while others exhibited almost no photoactivity and could be considered traditional cytotoxic agents. The source of higher dark cytotoxicity for certain compounds is not known but could be related to their lipophilicities and resulting cellular uptake and/or localization.

Although the generation of ¹O₂ under the cell-free condition does not establish ROS as the definitive mediator of photocytotoxicity, we presume PDT effects are responsible. As such, the excited state dynamics and redox characteristics of the complexes must be explored in order to propose a mechanism(s). However, the fact that 5h with the central pyrenyl group emerged as an extremely potent photosensitizer for in vitro PDT and that the triplet state energy of the isolated pyrenyl group is in energetic proximity to that of many well-studied ³MLCT states suggests at least a possible role for ³IL states in producing the larger ¹O₂ quantum yield and greater in vitro PDT potency toward cancer cells. At the time 5h was evaluated, PIs of such magnitude had not been reported and the opportunity to use interval dosing to achieve PIs >27 000 had not been explored by groups developing new PSs. Compound 5h was also highly active toward the more resistant tumor spheroid model, which is characterized by multicellular resistance and regions of hypoxia, and also toward bacteria. The versatility of this new photosensitizer for both light-mediated anticancer and antimicrobial applications highlights the potential utility of the bis [Ru(II)-pyrrolide] scaffold for photobiological applications and introduces a new platform for further optimization of these important light-responsive agents.

Author contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

Conflicts of interest

S. A. M. has a potential research conflict of interest due to a financial interest with Theralase Technologies, Inc. and PhotoDynamic, Inc. A management plan has been created to preserve objectivity in research in accordance with UTA policy.

Acknowledgements

S. A. M. and A. T. thank the Natural Sciences and Engineering Research Council of Canada (NSERC) for financial support. S. A. M. and C. G. C. thank the National Cancer Institute (NCI) of the National Institutes of Health (NIH) (Award R01CA222227) for partial support of this work. The content in this article is solely the responsibility of the authors and does not necessarily

represent the official views of the National Institutes of Health. The TOC graphic was made using https://Biorender.com/.

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