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A facile electrochemical intercalation and microwave assisted exfoliation methodology applied to screen-printed electrochemical-based sensing platforms to impart improved electroanalytical outputs†

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Screen-printed electrodes (SPEs) are ubiquitous with the field of electrochemistry allowing researchers to translate sensors from the laboratory to the field. In this paper, we report an electrochemically driven intercalation process where an electrochemical reaction uses an electrolyte as a conductive medium as well as the intercalation source, which is followed by exfoliation and heating/drying via microwave irradiation, and applied to the working electrode of screen-printed electrodes/sensors (termed EDI-SPEs) for the first time. This novel methodology results in an increase of up to 85% of the sensor area (electrochemically active surface area, as evaluated using an outer-sphere redox probe). Upon further investigation, it is found that an increase in the electroactive area of the EDI-screen-printed based electrochemical sensing platforms is critically dependent upon the analyte and its associated electrochemical mechanism (*i.e.* adsorption vs. diffusion). Proof-of-concept for the electrochemical sensing of capsaicin, a measure of the hotness of chillies and chilli sauce, within both model aqueous solutions and a real sample (Tabasco sauce) is demonstrated in which the electroanalytical sensitivity (a plot of signal vs. concentration) is doubled when utilising EDI-SPEs over that of SPEs.

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Introduction

Over recent decades there has been a focus and acceleration within the research community towards the miniaturisation/simplification of analytical protocols.¹ As such, the utilisation of screen-printing technology for the mass production of screen-printed electrodes (SPEs) has allowed low cost bespoke electrochemical sensors to be realised,^{2,3} allowing researchers to translate sensor technology from the laboratories to the field. To impart improvements in the electroanalytical sensing abilities of screen-printed electrochemical platforms, researchers readily modify SPEs with common nanomaterials such as graphene,⁴ metallic nanoparticles⁵ and metal phthalocyanines,^{6,7}

to name just a few. The approach for the surface modification of SPEs involves drop-casting the desired nanomaterials upon the SPEs but can in some instances produce unstable/irreproducible surfaces.⁶ Another approach to overcome this is to mix the nanomaterial into the bulk of the screen-printing ink for the development of bespoke nanomaterial modified inks.^{8,9} An alternative approach is to make structural/compositional changes to the SPEs, which could possibly provide increased electroanalytical signals. For example, Washe *et al.*¹⁰ reported a solvent-based modification, which is postulated to alter/remove the binder within the screen-printing ink thus creating increased access to the graphitic flakes within the ink and hence an associated increase in the electrochemically active area of up to 57 times. However, recent work has questioned such elaborate increments.¹¹ Other studies have sought to explore the polishing of SPEs upon polishing pads, which is generally undertaken only for solid electrode substrates (such as glassy carbon) which gives rise to changes in the SPE's surface morphology and carbon-oxygen functionalities^{11,12} resulting in significant improvements in sensor performance.¹¹

An approach used in the academic literature to increase the area of carbon materials involves electrochemical techniques,

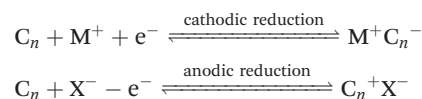
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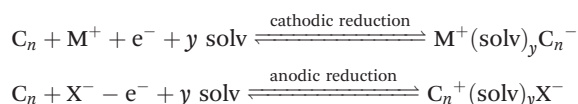
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which offer intercalation with acidic intercalants such as sulphuric, perchloric, nitric and trifluoroacetic acids.^{13–16} Electrochemical Interaction Reactions (EIRs) are utilised to exfoliate graphite. In these approaches, guest ions (cations, M^+ or anions, X^-) intercalate under electrochemical perturbation into the Van der Waal's gaps between carbon layers and enlarge the inter-layer distance.¹⁷ These EIRs can be summarised as:



In such cases, the C_n needs to be electrochemically active and the host carbon structure should be capable of accepting, not only sterically but also electronically, the guest ions *via* a suitable energy band structure.¹⁷ Interaction reactions identified above are reversible and topotactic.¹⁷ Another interesting scenario is the creation of ternary phases, *via*:



Some examples reported in the academic literature utilising the above approaches include the work of Morales *et al.*¹⁸ who reported the production of few-layer graphene *via* electrochemically produced graphite intercalation in aqueous perchloric acid solutions and expansion by microwave radiation, which contributes to the removal of the trapped solvent causing further expansion.¹⁹ While the approaches above are generally utilised to produce expanded graphite nanoplatelets¹⁹ or few-layer graphene,¹⁸ Zhang *et al.*²⁰ have utilised this graphite expansion method to produce a potential anode within a Li-ion battery, where the expansion of the graphitic structure allows for an increased number of reaction sites with the electrolyte, leading to improved energy capabilities.²⁰ The utilisation of the above approaches to screen-printed electrodes has not yet been, to the best of our knowledge, reported which can potentially be utilised to increase the electrochemical active surface area to impart improvements in electroanalytical performance.

Herein, we explore the feasibility of an anodically driven electrochemically intercalation process, where an electrochemical reaction uses an electrolyte as a conductive medium and as an intercalation source, which is followed by exfoliation and heating/drying *via* microwave irradiation, applied to the working electrode of screen-printed electrodes/sensors (EDI-SPEs) for the first time. In this approach, perchloric and formic acids are utilised as both the electrolyte and the intercalation compounds and the EDI-SPEs are evaluated towards redox probes and commonly benchmarked analytically useful analytes: sodium nitrite, dopamine, uric acid and capsaicin.

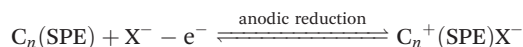
Experimental section

All chemicals used were obtained from Sigma-Aldrich at analytical grade and were used without any further purification. All solutions were prepared with deionised water of resistivity not less than 18.2 MΩ cm. Phosphate buffer solution (PBS) of pH 7 was prepared using disodium hydrogen phosphate (0.025 M) and potassium dihydrogen phosphate (0.025 M). Perchloric acid and phosphoric acid were diluted with water to obtain the desired concentration. Voltammetric measurements were carried out using an Emstat (Palmsens Instruments BV, The Netherlands) potentiostat and controlled by PS Trace 4.4 software. When it was necessary an external electrode Ag/AgCl, 3 M KCl (BAS, RE-5B) and a platinum wire were used as reference and counter electrodes respectively. SEM images were obtained with a JEOL JSM-5600LV model. The X-ray photoelectron spectroscopy (XPS) data was acquired using a AXIS Supra with a Al X-ray source (1486.6 eV) operating at 300 W for survey scans and 450 W for narrow scans. Al X-rays were monochromated using a 500 mm Rowland circle quartz crystal X-ray mirror. The angle between X-ray source and analyser was 54.7°.

Procedure to fabricate the EDI-SPEs

The SPEs fabricated herein have been extensively characterised *via* Raman and XPS analysis and have been published within recent literature;²¹ this information is re-reported here for the convenience of the reader. Screen-printed graphite electrodes were fabricated in-house with appropriate stencil designs using a microDEK 1760RS screen-printing machine (DEK, Weymouth, UK). For each of the screen-printed electrodes, a carbon-graphite ink formulation was first screen-printed onto a polyester flexible film (Autostat, 250 μm thickness). This layer was cured in a box fan oven with extraction at 60 °C for 30 min. Next, a silver/silver chloride (60:40) reference electrode was applied by screen-printing Ag/AgCl paste (Product Code: C2040308P3; Gwent Electronic Materials Ltd, UK) onto the plastic substrate. This layer was once more cured in an oven at 60 °C for 30 min. Finally, an insulating dielectric paste ink (Product Code: D2070423D5; Gwent Electronic Materials Ltd, UK) was printed to cover the connections and define the 3 mm diameter graphite working electrode. After curing at 60 °C for 30 min the screen-printed electrodes are ready to use and have been thoroughly benchmarked previously.^{22,23}

The procedure for the expansion of the electrochemical active area consists of two facile steps: Step 1, electrochemical intercalation step:



where two different supporting electrolytes were explored namely, perchloric acid (HClO_4) and formic acid (CH_2O_2), which have been reported within the literature to be the most beneficial towards the intercalation of graphite.^{17,24,25} In this approach, the perchloric and formic acid act as both the electrolyte and intercalation compound used in the EIR.



Step 2, expansion step: where the electrochemically driven intercalation compounds are removed *via* heating, in this case, microwave irradiation (Argos Stores, UK Simple Value, UK, Model: MM7 17CNF (F)-PM), which will also remove the trapped species, but assist with further expansion of the graphite. In this work, we also considered heating *via* a fan oven (see later).

Electrochemical characterisation (see the ESI†) of the electrodes utilised throughout this work was *via* the effective electroactive area (A_{eff})/electrode area. To evaluate this parameter (A_{eff}) we utilised the redox probe 1 mM hexaammineruthenium(III) chloride in 0.1 M KCl, and the Randles–Ševčík equation (at 298 K):²⁶ $I_p^{\text{quasi}} = 2.65 \times 10^5 n^{3/2} D^{1/2} \nu^{1/2} C A_{\text{eff}}$ where n is the number of electrons involved in the electrochemical process, D is the diffusion coefficient ($9.1 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$) of the electrochemical redox probe used,²⁷ ν is the applied voltammetric scan rate, C is the concentration of the electroactive substance, and A_{eff} is the effective electrode area.

To fully understand and benchmark these sensing platforms an array of scan rates was utilised (0.05 to 0.5 V s^{-1}) to estimate A_{eff} . To determine if the expansion causes an improvement in the surface's electron transfer behaviour, we evaluated the effect of the standard heterogeneous rate constant (k^0), using the Nicholson method:^{23,28} $k^0 = [2.18(DanF\nu/RT)0.5]^{\exp[-(\alpha 2nF/RT) \times \Delta E_p]}$, where the constants are the same as described previously, apart from the transfer coefficient α , which is assumed to correspond to 0.5, F is the Faraday constant, R the universal gas constant, and the experiments are performed at a temperature T . ΔE_p is peak-to-peak separation.

In the case of the determination of capsaicin in hot chilli sauce, ~2 g of Tabasco hot sauce was accurately weighed in a small conical flask and diluted with methanol (typically mL). The samples were ultra-sonicated for 20 minutes in order to extract the capsaicin, *via* liquid–liquid extraction from the chilli sauce. After this step, the samples were centrifuged for 10 min. The real sample was then filtered *via* gravitation into a 20 mL volumetric flask and diluted with methanol. Finally, 1.0 mL of the final solution was placed within an electrochemical cell and diluted further with 9 mL of 0.1 M H_3PO_4 , which acts as the supporting electrolyte.

intercalation process as detailed in the Experimental section, where the electrolyte serves as a conductive medium to undertake electrochemical experiments while also acting as the intercalation source. This is followed by a further exfoliation and heating/drying step *via* microwave irradiation, all applied to the working electrode of screen-printed electrodes/sensors, for the first time (see Fig. 1). The microwave irradiation power, time and electrolytes have been explored and are summarized in the ESI.† The electroanalytical performance of the EDI-SPEs is considered and compared with that of the unmodified SPEs, towards the electroanalytical sensing of various electrochemical redox probes and electroanalytical targets, to allow direct comparison with prior published literature work. Herein, the redox probes hexaammineruthenium(III) chloride/0.1 M KCl and ammonium iron(II) sulfate/0.2 M HClO_4 are used to electrochemically benchmark the EDI-SPEs with the academic literature as well as to explore electroanalytically useful probes. This data is presented in the ESI† and demonstrate that both the intercalation and expansion steps do not induce any significant oxygenated species upon the electrode surface and additionally that there has been no significant electronic change in the electrode/material structure, such results have been corroborated with comparative XPS analysis of unmodified and EDI-SPEs, presented in ESI Fig. 5.†

First, as depicted in Fig. 2 and Table 1, the electroanalytical performance of the EDI-SPEs is benchmarked towards sodium nitrite, dopamine, uric acid and capsaicin. In all cases, utilising the unmodified and EDI-SPE electrochemical analysis over the concentration range is linear, however it is apparent that the EDI-SPE does not significantly improve the sensitivity of the EDI-SPEs/sensor towards the detection of dopamine, sodium nitrite and uric acid. Nonetheless, there are slight analytical improvements (when utilising the EDI-SPEs) within the limits of detection (LOD, 3σ) of each analyte (sodium nitrite, dopamine and uric acid), but the response is hindered due to the increased capacitive current. The electrochemical detection of capsaicin (presented in Fig. 2G and H) reveals that the EDI-SPEs result in an improved sensitivity (0.126 and $0.239 \mu\text{A } \mu\text{M}^{-1}$ for the SPEs and the EDI-SPE respectively) and LOD (1.76 and $0.36 \mu\text{M}$ for the SPEs and the EDI-SPE respectively) when utilising the EDI-SPEs over that of SPEs.

Thus, we can conclude that this modification technique/process does not improve the electroanalytical performance of analytes that undergo diffusional or mixed diffusional-adsorp-

Results and discussion

We explore the use of screen-printed electrodes (SPEs) that have been subjected to the anodic electrochemically driven

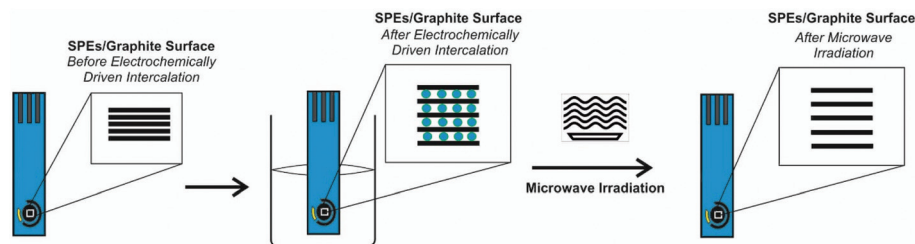


Fig. 1 Schematic overview of the modification protocol of the microwave assisted electrochemically driven intercalated screen-printed electrodes (EDI-SPEs).



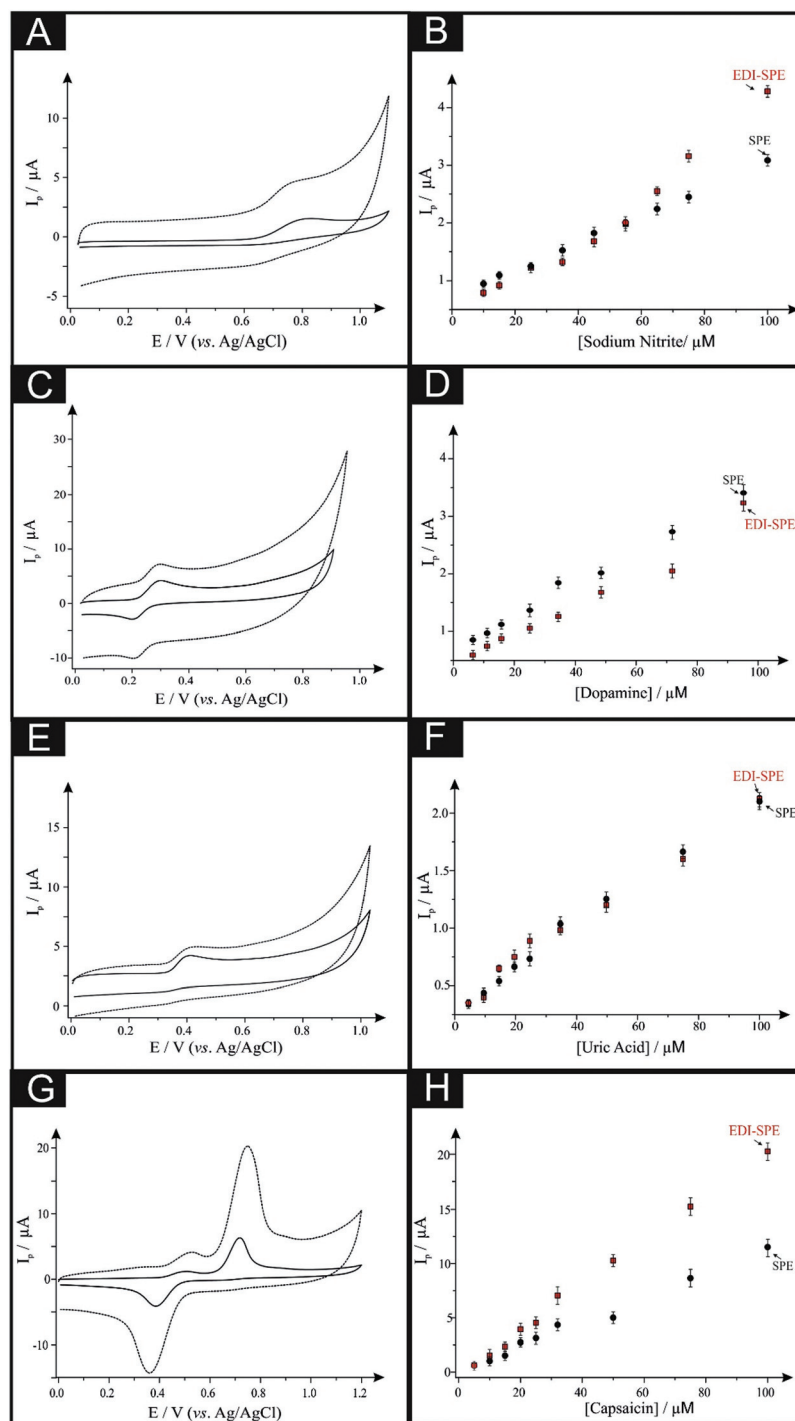


Fig. 2 Cyclic voltammograms utilising EDI-SPEs explored towards the electroanalytical detection of: (A) 50 μM nitrite, (C) 50 μM dopamine, (E) 50 μM uric acid and (G) 50 μM capsaicin obtained for SPEs (solid line) and EDI-SPEs (checked lines). Resulting calibration plots for (B) nitrite, (D) dopamine, (F) uric acid and (H) capsaicin, obtained for SPEs (circles) and EDI-SPEs (squares). In all cases, scan rate: 0.1 V s⁻¹.

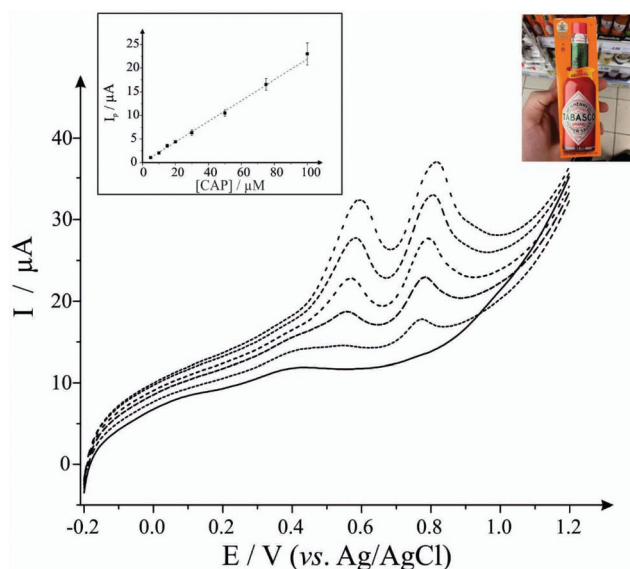
tive reaction mechanisms using the EDI-SPEs. However, it is clear that analytes that require a strong adsorptive mechanism, as is the case with capsaicin, demonstrate an improved electroanalytical performance when utilising EDI-SPEs. The observed electrochemical behaviour agrees with independent work by Compton *et al.*²⁹ who demonstrate that the electrochemical

mechanism of capsaicin is *via* an adsorption mechanism (rather than diffusion). Hence, they utilise this adsorption (*via* adsorption stripping voltammetry) using multi-walled carbon nanotubes (MWCNT) modified electrodes where the adsorption is favoured strongly by pi-pi interactions between the aromatic moieties in capsaicin and the surface of the carbon



Table 1 Figures of merit for different analytes using SPEs (unmodified) and EDI-SPEs

Analyte	Methodology	Sensitivity ($\mu\text{A } \mu\text{M}^{-1}$)	LOD (μM)	Linear range (μM)
Nitrite	SPEs	0.024	1.66	10–100
	EDI-SPEs	0.038	6.32	10–100
Dopamine	SPEs	0.032	0.92	5–100
	EDI-SPEs	0.031	4.91	5–100
Uric acid	SPEs	0.024	1.70	5–100
	EDI-SPEs	0.020	2.47	5–100
Capsaicin	SPEs	0.111	4.76	5–100
	EDI-SPEs	0.209	2.77	5–100

**Fig. 3** Calibration curves for the electroanalytical detection of capsaicin using EDI-SPEs. The insert shows the analysis of addition of capsaicin to pH 1.2 phosphate buffer solution using linear sweep voltammetry (plot of peak current vs. capsaicin concentration with the average and standard deviation presented). The main figure shows the linear sweep voltammograms obtained for different additions of capsaicin made into the chilli sauce (dotted lines) and the blank response (black solid line) prior to capsaicin addition. See experimental section for more details.**Table 2** Recovery assays (five analyses, S1–S5) for capsaicin determination in Tabasco sauce samples

Samples	Added/ μM	Recovery/ μM	Recovery/ %	Relative error/%
S1	10	9.41	94.1	−5.9
S2	20	18.64	93.2	−6.8
S3	30	30.35	101.2	+1.2
S4	40	40.06	100.1	+0.1
S5	50	45.55	91.1	−8.9

Recovery analysis

Mean recovery (R_{exp})	95.94
Standard deviation of recoveries (S_R)	4.45
Number of samples (N)	5
t_{exp}	2.0
$t_{(0.025, 4)}$	2.8

nanotubes (CNTs). Thus in our work, the exfoliation increases the surface area of the basal sheets of graphite providing more aromatic surfaces for capsaicin to adsorb onto; hence the observed increase in the sensitivity ($\sim \times 2$).²⁹

Next, we consider the applicability of the EDI-SPEs towards the detection of capsaicin within a real sample, Tabasco sauce. Fig. 3 demonstrates linear sweep voltammograms directly from five spiked Tabasco sauce samples. However, the content of capsaicin in the real sample was found to be under the detection limit of this analytical procedure, therefore recovery assays were employed in order to validate the methodology, with an average recovery of 95% utilising the EDI-SPEs (see Table 2); in summary we can conclude that the EDI-SPEs are useful for the electroanalytical determination of capsaicin within real chilli sauce.

Conclusions

We have explored an electrochemically driven intercalation process where the electrochemical reaction uses an electrolyte as a conductive medium as well as the intercalation source, which is followed by exfoliation and heating/drying *via* microwave irradiation, applied to the working electrode of screen-printed electrodes/sensors for the first time. We demonstrate proof-of-concept that the protocol can be used beneficially to increase the surface area of electrochemical sensors of adsorption controlled electrochemically active analytes. We demonstrate that the electroanalytical detection of capsaicin using the EDI-SPEs exhibits a double increase in the electroanalytical sensitivity with a lower limit of detection in comparison with unmodified SPEs. We extended this exciting work to the determination of capsaicin in real hot chilli sauce demonstrating that EDI-SPEs can be employed as an interesting alternative for the determination of capsaicin in chilli hot sauce with improved analytical sensitivity. This approach has generic advantages for electrochemically active probes that occur *via* an adsorptive mechanism and should be considered further allowing improvements in the electroanalytical sensitivity.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 S. A. Wring and J. P. Hart, *Analyst*, 1992, **117**, 1215–1229.
- 2 A. P. Ruas de Souza, M. Bertotti, C. W. Foster and C. E. Banks, *Electroanalysis*, 2015, **27**, 2295–2301.
- 3 K. Senthilkumar, T. N. Sithini, N. Thiyagarajan, S. Baskar and J.-M. Zen, *Electrochem. Commun.*, 2015, **60**, 113–116.
- 4 F. Arduini, L. Micheli, D. Moscone, G. Palleschi, S. Piermarini, F. Ricci and G. Volpe, *TrAC, Trends Anal. Chem.*, 2016, **79**, 114–126.
- 5 R. A. Farghali and R. A. Ahmed, *Int. J. Electrochem. Sci.*, 2015, **10**, 11.
- 6 C. Foster, J. Pillay, J. Metters and C. Banks, *Sensors*, 2014, **14**, 21905.
- 7 M. A. T. Gilmartin, J. P. Hart and B. J. Birch, *Analyst*, 1994, **119**, 243–252.
- 8 S. J. Rowley-Neale, C. W. Foster, G. C. Smith, D. A. C. Brownson and C. E. Banks, *Sustainable Energy Fuels*, 2017, **1**, 74–83.
- 9 E. P. Randviir, D. A. C. Brownson, J. P. Metters, R. O. Kadara and C. E. Banks, *Phys. Chem. Chem. Phys.*, 2014, **16**, 4598–4611.
- 10 A. P. Washe, P. Lozano-Sánchez, D. Bejarano-Nosas and I. Katakis, *Electrochim. Acta*, 2013, **91**, 166–172.
- 11 E. Blanco, C. W. Foster, L. R. Cumba, D. R. do Carmo and C. E. Banks, *Analyst*, 2016, **141**, 2783–2790.
- 12 J. Lee, D. W. M. Arrigan and D. S. Silvester, *Sens. Bio-Sensing Res.*, 2016, **9**, 38–44.
- 13 M. S. Dresselhaus and G. Dresselhaus, *Adv. Phys.*, 2002, **51**, 1–186.
- 14 M. Noel and R. Santhanam, *J. Power Sources*, 1998, **72**, 53–65.
- 15 A. Yoshida, Y. Hishiyama and M. Inagaki, *Carbon*, 1991, **29**, 1227–1231.
- 16 M. Inagaki, *J. Mater. Res.*, 2011, **4**, 1560–1568.
- 17 J. O. Besenhard and H. P. Fritz, *Angew. Chem., Int. Ed. Engl.*, 1983, **22**, 950–975.
- 18 G. M. Morales, P. Schifani, G. Ellis, C. Ballesteros, G. Martínez, C. Barbero and H. J. Salavagione, *Carbon*, 2011, **49**, 2809–2816.
- 19 L. M. Viculis, J. J. Mack, O. M. Mayer, H. T. Hahn and R. B. Kaner, *J. Mater. Chem.*, 2005, **15**, 974–978.
- 20 H. Zhang and M. M. Lerner, *Inorg. Chem.*, 2016, **55**, 8281–8284.
- 21 A. Gomis-Berenguer, M. Gomez-Mingot, V. Montiel, A. Canals, T. Thiemann, R. O. Kadara, C. E. Banks and J. Iniesta, *RSC Adv.*, 2012, **2**, 7735–7742.
- 22 F. E. Galdino, C. W. Foster, J. A. Bonacin and C. E. Banks, *Anal. Methods*, 2015, **7**, 1208–1214.
- 23 C. W. Foster, A. P. de Souza, J. P. Metters, M. Bertotti and C. E. Banks, *Analyst*, 2015, **140**, 7598–7612.
- 24 J. Jiang, *Carbon*, 1993, **31**, 663–666.
- 25 F. Kang, Y. Leng and T. Y. Zhang, *Carbon*, 1997, **35**, 1089–1096.
- 26 D. A. C. Brownson, C. W. Foster and C. E. Banks, *Analyst*, 2012, **137**, 1815–1823.
- 27 C. E. Banks, R. G. Compton, A. C. Fisher and I. E. Henley, *Phys. Chem. Chem. Phys.*, 2004, **6**, 3147–3152.
- 28 I. Lavagnini, R. Antiochia and F. Magno, *Electroanalysis*, 2004, **16**, 505–506.
- 29 R. T. Kachoosangi, G. G. Wildgoose and R. G. Compton, *Analyst*, 2008, **133**, 888–895.

