



Cite this: *Org. Biomol. Chem.*, 2015, **13**, 1690

Preparation of luminescent chemosensors by post-functionalization of vesicle surfaces†

Andreas Müller and Burkhard König*

Surface-reactive luminescent vesicles were prepared by self-assembly of phospholipids, amphiphilic maleimides and fluorophors in aqueous solution. Those preformed liposomes were functionalized with various thiolated receptor units using a thiol-click reaction. As recognition elements, a bis-Zn²⁺-cyclen derivative for the detection of phosphate moieties or a DNA aptamer for the specific binding of the antibiotic ampicillin were utilized. A FRET-based assay revealed the close spatial proximity of the membrane-embedded dansyl molecules with the subsequently immobilized thiols, which is the origin for the signaling mechanism of the obtained vesicular sensors. Those receptor-functionalized liposomes indicate the binding of the targets to their surface by changes of the fluorescence emission properties of the membrane co-embedded carboxyfluorescein dyes. The post-functionalization concept can also be used for molecular imprinting on vesicle surfaces. The template-guided patterning of receptors based on bis-Zn²⁺-cyclen resulted in fluorescent sensors suitable for the specific recognition of a bivalent peptide.

Received 1st November 2014,
Accepted 26th November 2014

DOI: 10.1039/c4ob02327g

www.rsc.org/obc

1. Introduction

Molecular recognition at biomembranes is of fundamental importance for biological processes like cell recognition, adhesion, fusion or membrane transport phenomena. Since phospholipid vesicles can be considered as simple cell mimics, non-covalent interactions at the surface of receptor-functionalized synthetic liposomes resemble recognition events at membranes as they appear in nature.¹ Hence, the knowledge obtained from the investigation of artificial vesicular systems can help to gain improved insights into the correspondent processes in biology. Conversely, a better understanding of the natural prototypes can be exploited for the development of powerful devices for analytical applications. Various strategies have been established to transform the binding event at the vesicle membrane into an observable signal.² Fluorescent assays provide high sensitivities allowing the detection of analytes present in trace amounts, whereas colorimetric formats enable the analytic readout by the naked eye.³ Typically, the signaling unit is in direct contact with the recognition element by either covalent attachment^{4,5} or by non-covalent coordination as in indicator-displacement assays.^{4,6} However, membranes are highly dynamic due to their supramolecular composition.¹ Based on this, our group

recently reported on a novel strategy for the modular construction of chemosensors by means of unilamellar vesicular membranes, which serve as self-assembled supporting matrix for amphiphilic metal complex receptors and fluorescent reporter dyes.⁷ Due to lateral phase separation, the co-embedded amphiphilic fluorophors and binding sites are expected to cluster on the surface resulting in fluorescence quenching of the amphiphilic dyes (Fig. 1). Binding of analyte molecules to the surface-accessible receptors alters the physico-chemical properties in their microenvironment and thus induces a re-organization of the membrane components. These processes influence the optical properties of the co-embedded dyes, which in turn respond with a change of their emission intensity.

This co-embedding strategy allows an effortless preparation of sensor devices by simple mixing of different functional amphiphiles in aqueous solution. By careful variation of the binding and signaling units, their ratios and concentrations on the surface and by the choice of suitable phospholipids determining the physical properties of the membrane, vesicular sensors with specific characteristics can be designed. However, as the synthesis of the amphiphilic receptors can be quite tedious, the availability of suitable binding sites and hence the variety of addressable analytes is limited. Moreover, control over the two-dimensional receptor assembly for multi-point interactions on the surface is still a challenge.⁸ Its accomplishment is an essential requirement for the highly selective recognition of multivalent analytes. In order to approach these issues, we developed a more universal method, which allows the facile preparation of vesicular chemosensors

Institut für Organische Chemie, Universität Regensburg, 93040 Regensburg,

Germany. E-mail: burkhard.koenig@ur.de; Fax: +49 943 1717; Tel: +49 943 4575

† Electronic supplementary information (ESI) available: Experimental details, synthesis of compounds, preparation of functionalized vesicles, emission spectra, binding curves and optimization studies. See DOI: 10.1039/c4ob02327g



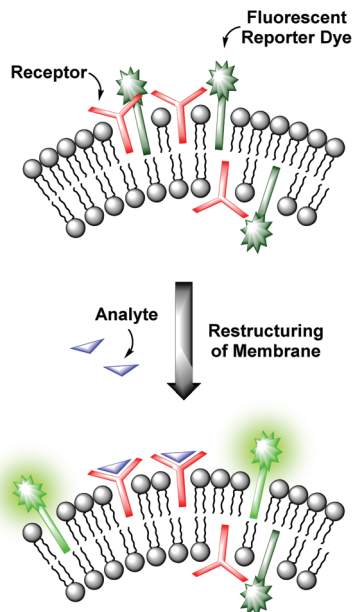


Fig. 1 Sensing mechanism of receptor-functionalized fluorescent vesicles.

in a modular fashion by means of surface “post-functionalization” *via* click chemistry. The surface modification of preformed liposomes is an established method for the conjugation of various ligands ranging from sugars to peptides, proteins and antibodies and has found applications in, *e.g.*, the investigation of drug delivery systems.⁹ We now combine this concept of vesicle post-modification with our co-embedding strategy of functional amphiphiles into phospholipid membranes. Only groups of the vesicle membranes which are exposed to the outer side are functionalized. The resulting asymmetric membrane composition represents a much better mimic of the bilayers appearing in nature. In view of more complex receptors of biological origin, we utilized a thiol-ene click reaction¹⁰ due to the frequent occurrence of thiol groups in biomolecules. The reaction is fully biocompatible and does not, in contrast to the copper(I)-catalyzed azide-alkyne cycloaddition,¹¹ require a metal catalyst.

2. Results and discussion

We first tested the applicability of the UV light-induced radical addition of thiols to electron-rich alkene moieties.¹² This type of reaction would implicate the possibility to externally trigger the functionalization process by controlled exposure to UV light. However, the envisaged photoreaction is at micromolar concentrations in aqueous solution not efficient. Thus, we focused on the widely used Michael addition of thiols to maleimides.¹³ This reaction performs well over a large concentration range in aqueous media under benign conditions without the formation of byproducts and is commonly applied in bioconjugation.¹⁰ We recently employed this method for the construction of an aptamer-based biosensor by attachment of thiol-

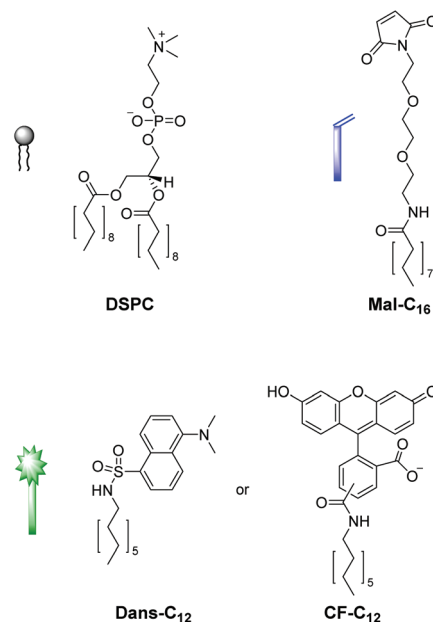


Fig. 2 Structures of the functional amphiphiles used for vesicle preparation.

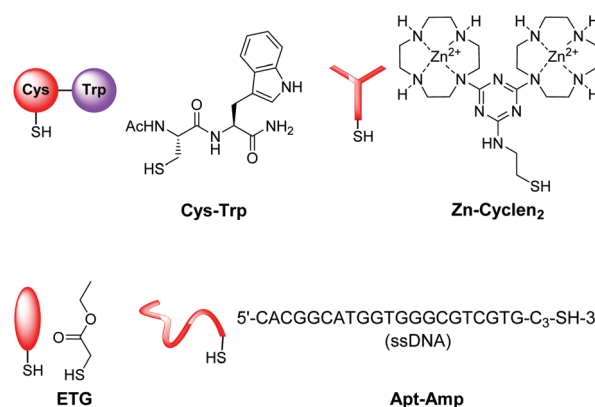


Fig. 3 Thiols used for the post-functionalization of vesicle surfaces.

modified aptamers to membrane-embedded amphiphilic maleimide **Mal-C₁₆** (Fig. 2).¹⁴

Here, we report on the development of the post-functionalization strategy of luminescent vesicle surfaces with receptor sites and demonstrate its facile and versatile applicability for the preparation of various sensors. As signaling units, either amphiphilic dansyl dye **Dans-C₁₂**¹⁵ or carboxyfluorescein **CF-C₁₂** were incorporated into membranes of small unilamellar DSPC vesicles. Fig. 3 depicts the different thiols which were used for our investigations. The syntheses of compounds **Mal-C₁₆**¹⁴ and **CF-C₁₂**^{7a} were previously reported by us, the preparation of **Cys-Trp** and **Zn-Cyclen₂** is described in the ESI.†

2.1 Optimization of the post-functionalization of preformed vesicles

The sensing mechanism of our vesicular sensors is based on the close spatial proximity of the binding sites to the



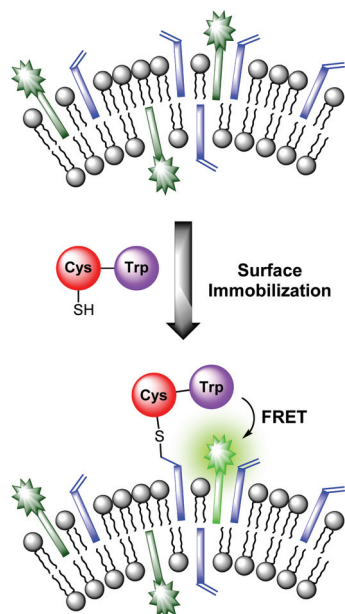


Fig. 4 FRET assay proving the close spatial proximity of immobilized thiols to the membrane-embedded fluorophores.

membrane-embedded dyes. To test whether the subsequently immobilized receptor molecules fulfill this condition, we developed an assay based on the distance-dependent Förster resonance energy transfer (FRET, Fig. 4). As simple model system, we synthesized dipeptide **Cys-Trp** (Fig. 3). The FRET partner of tryptophan, the amphiphilic dansyl dye **Dans-C₁₂** (Förster distance: 2.1 nm),¹⁶ was co-embedded (5.0 mol%) into the membrane of DSPC vesicles together with the attachment site for thiols **Mal-C₁₆** (5.0 mol%). The preparation of the vesicles was performed in 25 mM HEPES buffer at a pH of 7.4 according to previously reported procedures (see ESI† for details). Extrusion through 100 nm polycarbonate membranes yielded vesicles **V-Dans** with a size distribution of around 130–150 nm (see Fig. S1, ESI†). The non-functionalized vesicles were stable and did not show significant aggregation after one day (see Fig. S2, ESI†).

Addition of **Cys-Trp** to **V-Dans** showed a time-dependent decrease of the tryptophan emission evidencing the fluorescence energy transfer and a considerable increase of the dansyl signal (Fig. 5).

Apart from the contribution of the resonance energy transfer from the tryptophan moiety, the extraordinarily strong increase of the dansyl signal must be attributed to two additional independent effects: (i) It is well documented that the fluorescence emission of dansyl dyes is quenched by adjacent maleimides *via* photoinduced electron transfer (PET), which proceeds from the excited state of the fluorophore to the maleimide.¹⁷ After conjugation of thiols to the electron-deficient double bond of the maleimide π -system of **Mal-C₁₆**, this PET process is interrupted resulting in a dansyl fluorescence quantum yield enhancement. (ii) Furthermore, by the altered physico-chemical properties of the local environment

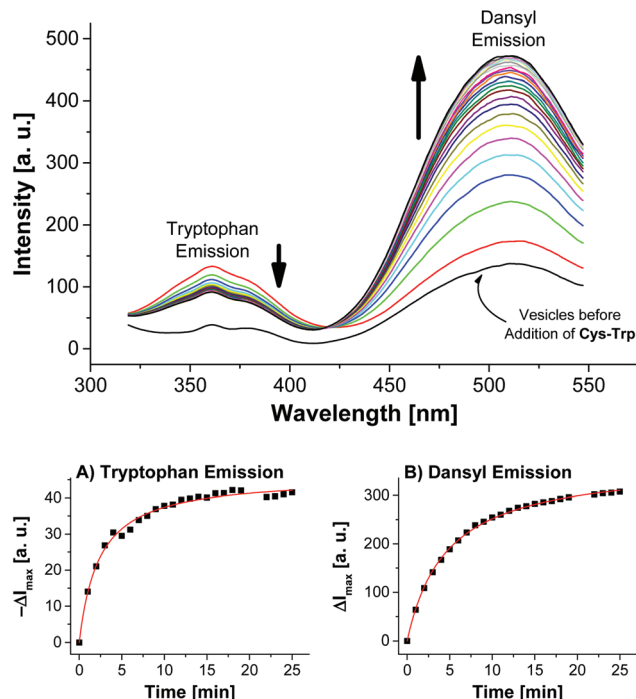


Fig. 5 Top: Time-dependent fluorescence spectra of vesicles **V-Dans** after addition of 3.0 mol% of peptide **Cys-Trp** (λ_{ex} = 280 nm). Bottom: course of surface reaction as evidenced by the emission signals of either the tryptophan or dansyl dye.

upon surface functionalization, the optical properties of the membrane-embedded fluorophores change causing an increase in emission intensity. A fluorescence increase of **V-Dans** during the surface reaction with a non-tryptophan-containing thiol like **ETG** (ethyl thioglycolate, Fig. 3) confirms this interpretation (see Fig. S3, ESI†). The observations clearly show that the thiol is covalently attached to the membrane-embedded maleimide **Mal-C₁₆** and the membrane-bound thiols in turn are in close proximity to the co-embedded fluorophores. Membranes without **Mal-C₁₆** show a much higher initial dansyl emission intensity and no time-dependent change of the emission signal of either fluorophores, the tryptophan or dansyl dye, occurred after addition of **Cys-Trp** (see Fig. S5, ESI†). The same observation was made for vesicles **V-Dans** whose surface-accessible maleimides were saturated with **ETG** prior to the addition of **Cys-Trp** (see Fig. S4, ESI†). The experiments demonstrate that non-functionalized, maleimide-containing vesicles **V-Dans** can also be regarded as simple self-assembled sensors for thiols.¹⁸

In contrast to dansyl dyes, carboxyfluorescein derivatives do not show PET processes in presence of maleimides in the membrane. To further study the effect (ii), we prepared vesicles **V-CF** with embedded carboxyfluorescein **CF-C₁₂**, which showed a time-dependent increase of the fluorescence signal after addition of thiol **ETG** indicating membrane restructuring processes upon surface functionalization (see Fig. S6, ESI†). By non-linear curve fitting, the second-order rate constants of thiol–maleimide conjugations on the vesicle membranes were



Table 1 Rate constants for vesicle functionalization reactions based on the signal changes of the co-embedded fluorophores

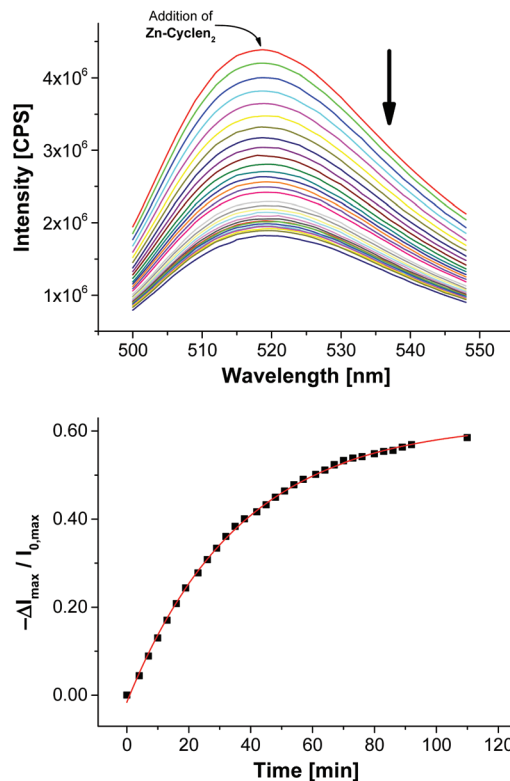
Entry	Vesicles	Thiol	k_{obs} [$\text{M}^{-1} \text{s}^{-1}$]
1	V-Dans	Cys-Trp	315
2	V-Dans	ETG	108
3	V-CF	ETG	455

determined (Table 1). The rate constants are all in the same order of magnitude. Small deviations along with different combinations of employed thiol and membrane composition can be explained by the course of the thiol–maleimide addition *via* thiolate anions. Their concentration is directly related to the pH of the reaction medium and to the $\text{p}K_{\text{a}}$ value of the corresponding thiol. These $\text{p}K_{\text{a}}$ values in turn are influenced by both the chemical nature of the thiol and the physico-chemical properties in their environment, *i.e.*, by the individual properties of the particular vesicle–water interface.¹⁹ Noteworthy, considerably basic conditions drastically enhance the nucleophilicity, however, also accelerate the oxidation of thiols to disulfides. Therefore, the pH value of 7.4 in our vesicular systems poses a good compromise between reaction rate and thiol stability. Compared to literature known rate constants from homogeneous systems ($600\text{--}8850 \text{ M}^{-1} \text{s}^{-1}$ at $\text{pH} = 7.0$),²⁰ the lower rate constants of about one order of magnitude at the liposomal surfaces are probably due to the lower diffusion rates of vesicles.²¹

2.2. Preparation of chemosensors for phosphates by post-functionalization of vesicles with Zn-Cyclen₂

Next, we utilized the established conditions for the membrane attachment of thiolated species for the preparation of a simple chemosensor. As phosphates are ubiquitous in nature, they pose important targets for analytical devices. Recently, our group reported on the preparation of vesicular chemosensors for phosphates based on amphiphilic derivatives of bis- Zn^{2+} -cyclen complexes.^{4,7a} Those receptors are known to bind to phosphates with nanomolar affinities.²² To demonstrate the proof of principle of our post-functionalization approach and to show its applicability for sensor preparation in analogy to our previous investigations, we synthesized the thiolated bis- Zn^{2+} -cyclen receptor **Zn-Cyclen₂** (Fig. 3) and immobilized it on maleimide-modified vesicles **V-CF**. For that purpose, we added a sub-stoichiometric amount of the thiol (0.8 equivalents)²³ with respect to the surface-accessible concentration of **Mal-C₁₆** in order to avoid the separation of excess **Zn-Cyclen₂** after vesicle functionalization. Fluorescence measurements revealed a remarkable initial drop of the emission signal (see Fig. S7, ESI†) upon addition of **Zn-Cyclen₂** followed by a further time-dependent decrease of the fluorescence intensity indicating the progress of the surface reaction (Fig. 6).

Compared to the surface reaction of vesicles **V-CF** with ETG, the functionalization with **Zn-Cyclen₂** was considerably slower (Table 2, entry 1). We explain this with the occurrence of multiple equilibria diminishing the concentration of free

**Fig. 6** Time-dependent fluorescence response showing the functionalization progress of vesicles **V-CF** with 2.0 mol% of **Zn-Cyclen₂**.**Table 2** Rate constants for the functionalization of vesicles **V-CF** with different amounts of **Zn-Cyclen₂** in absence or presence of PP_i (for fluorescence emission data, see Fig. S7 and S8, ESI)

Entry	Zn-Cyclen₂ ^a	PP_i ^b	k_{obs} [$\text{M}^{-1} \text{s}^{-1}$]
1	0.8 eq. (2.0 mol%)	—	82
2	1.0 eq. (2.5 mol%)	—	78
3	1.0 eq. (2.5 mol%)	2.0 eq.	322
4	2.0 eq. (5.0 mol%)	—	31
5	2.0 eq. (5.0 mol%)	2.0 eq.	202

^a Equivalents with respect to surface-accessible amount of **Mal-C₁₆**.

^b Equivalents with respect to total amount of **Zn-Cyclen₂**.

Zn-Cyclen₂: (i) The strong complexation abilities of thiol groups towards metal centers like zinc cations are supposed to result in intermolecular Zn^{2+} –SH coordination.²⁴ (ii) Furthermore, the considerable instant quenching of the fluorescence signal after addition of the receptor indicates electrostatic interactions of the bis- Zn^{2+} -cyclen head groups with the negatively charged membrane-embedded carboxyfluorescein derivatives. To further verify these assumptions, we varied the amount of receptor **Zn-Cyclen₂** since the proposed equilibria are expected to be shifted towards the coordinated species at higher concentrations of **Zn-Cyclen₂** (entries 2 and 4). Additionally, we performed the surface reaction in the presence of pyrophosphate (PP_i , entries 3 and 5) due to its ability to compete for the Zn^{2+} -centered coordination site of the



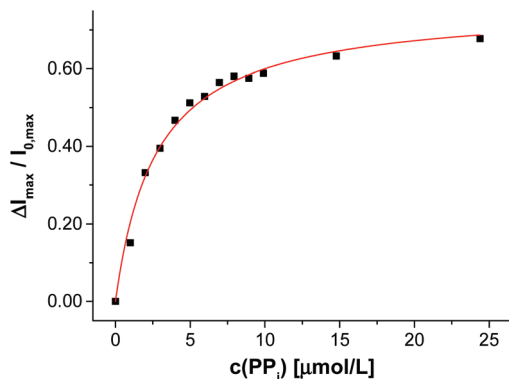


Fig. 7 Fluorescence response upon titration of PP_i to functionalized vesicles **V-CF-Zn**.

receptor and thus to increase the concentration of free **Zn-Cyclen**₂ thiol functions. The results presented in Table 2 show two obvious trends which confirm our hypothesis: An enhancement of the reaction rate in the presence of PP_i and a deceleration of the reaction along with higher thiol concentrations.

The addition of disulfide precursor of **Zn-Cyclen**₂ (compound 7 in Scheme S2, ESI†) to **V-CF** leads to a strong decrease of the emission signal supporting the proposed coordinative interaction of the Zn^{2+} -complex with **CF-C**₁₂. However, as expected no further time-dependent decrease was observed due to the absence of free thiol groups required for the surface attachment (see Fig. S9, ESI†).

After completion of the functionalization process of **V-CF** with **Zn-Cyclen**₂, the obtained vesicles **V-CF-Zn** were ready for use as luminescent sensors for phosphate species. Titration of the analyte PP_i resulted in a strong increase of their fluorescence emission signal (Fig. 7).

For control experiments, non-functionalized vesicles were prepared by either omitting **Zn-Cyclen**₂ or by addition of **Zn-Cyclen**₂ to vesicles lacking the membrane-embedded attachment site **Mal-C**₁₆. Titration of PP_i did not show a significant response of the fluorescence signal in those cases (for titration data, see Fig. S10, ESI†). These results proved that the membrane attachment of the receptor molecules is an essential requirement for the sensing mechanism of the PP_i analyte molecules.

We examined different phosphate-containing analytes in their binding affinities to vesicles **V-CF-Zn** and found binding constants which are in good agreement with literature values demonstrating the proof of concept (Table 3). The absence of a fluorescence emission change upon titration of sulfate as a dianionic control analyte confirmed the specificity of **V-CF-Zn** towards phosphate anions.

Next, we investigated the dependency of the fluorescence response on the concentration of membrane-embedded fluorophores and immobilized receptor molecules (see Fig. S12, ESI†). Best signal to noise ratios were obtained with increasing amounts of both dye and receptor. These results are consistent with our previous observations and support our mechanistic

Table 3 Binding affinities of **Zn-Cyclen**₂-functionalized vesicles **V-CF-Zn** to different analytes (for titration data, see Fig. S11, ESI)

Entry	Analyte	$\lg K_a$	Reference ^a
1	PP_i	6.3	6.2 ^{7a}
2	D-Fructose 1,6 bisphosphate	6.0	6.1 ⁴
3	Phosphorylated hexapeptide	5.6	5.9 ^{26a}
4	Pep-P-His (Fig. 8) SO_4^{2-}	No response	—

^a Refers to vesicular sensor systems obtained by co-embedding of amphiphilic bis- Zn^{2+} -cyclen receptors into the membrane.

hypothesis of receptor-dye patches on the vesicle surface, which reorganize upon formation of receptor-analyte complexes.^{7a}

2.3 Molecular imprinting of vesicle surfaces

Our investigations with the **Zn-Cyclen**₂ receptor revealed that the post-functionalization technique is well suited for the preparation of luminescent chemosensors in a modular fashion. Table 3 shows that the obtained binding constants of the different phosphates are almost equal. However, for the development of powerful analytical devices being able to discriminate between structurally related analytes, well-defined multipoint interactions between hosts and guests are essential. That means, the arrangement of the recognition elements on the vesicle surface must be complementary to the position of the binding units at the envisaged targets. Molecular imprinting is the classic technique in polymer science to achieve this aim.²⁵ In this connection, a molecule is utilized as a stamp to create its negative print in a suitable matrix. Recently, our group reported on two different strategies towards the preparation of highly specific, multi-receptor chemosensors. Both approaches are based on dynamic, template-guided recruitment of amphiphilic receptors on fluid vesicle membranes followed by fixation of the assemblies either by cooling the membrane down below the vesicular phase transition temperature^{26a,b} or by photopolymerization of the membrane-embedded receptor arrays.^{26c} A post-functionalization strategy can also be used for the molecular imprinting of defined two-dimensional receptor assemblies since the maleimide-decorated vesicles can be regarded as reactive matrices for template-assisted covalent functionalization. A fundamental prerequisite for that approach is the rigidity of the vesicle membrane preventing the lateral redistribution of the receptors after surface attachment. Hence, DSPC membranes with a phase transition temperature of 54 °C²⁷ represent an ideal scaffold for that purpose. Fig. 8 illustrates the principle of molecular imprinting by vesicle post-functionalization. In detail, membrane-attached receptor-template complexes direct the surface fixation of additional receptor molecules by coordinative interactions in spatially defined arrangements. Subsequent separation of the templates leads to an imprinted vesicle surface with patterns of binding sites.

Complex **Zn-Cyclen**₂ is known to bind besides phosphates also to histidine moieties.²⁸ As template for our imprinting



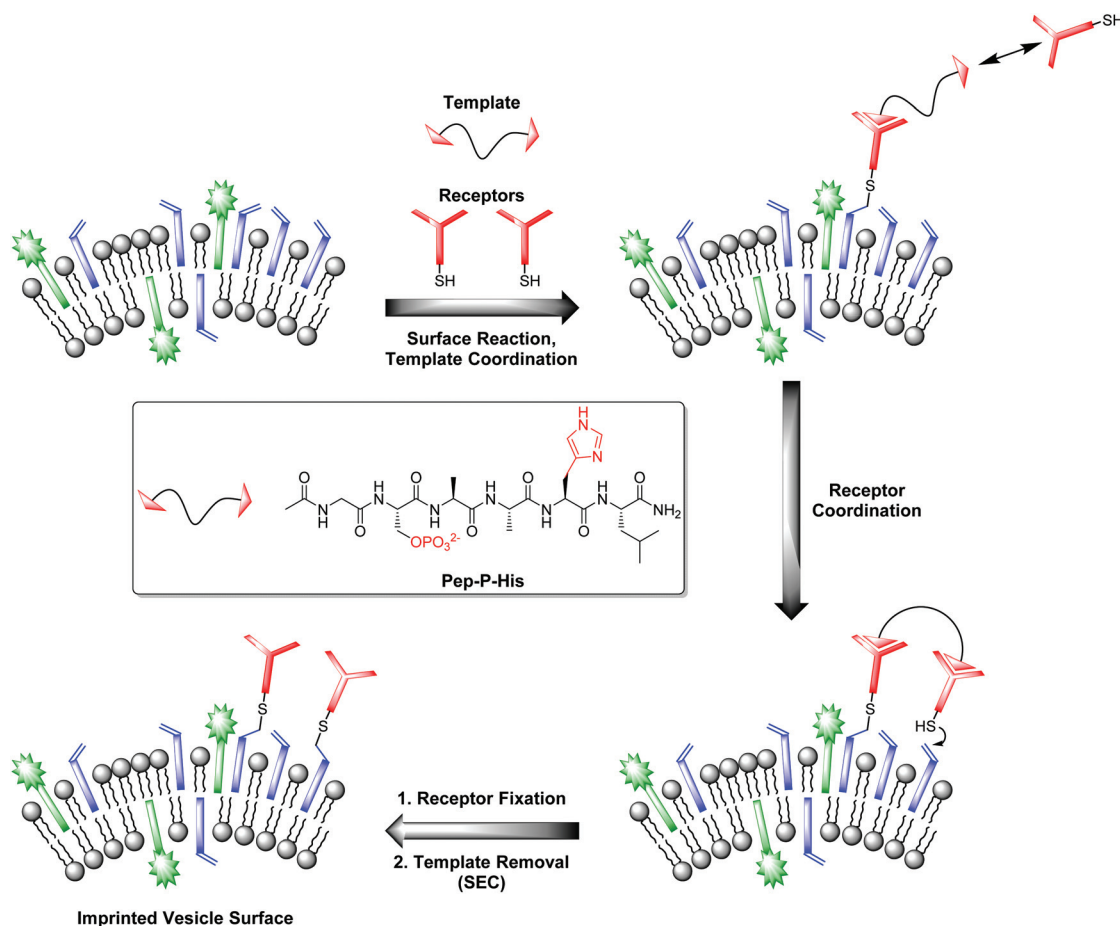


Fig. 8 Principle of vesicle surface imprinting by template-assisted post-functionalization and bivalent peptide **Pep-P-His** used as model template.

studies, we therefore utilized bivalent hexapeptide **Pep-P-His** (Fig. 8) being able to form a ternary complex with two receptor molecules. We incubated a 1 : 2 mixture of the peptide and the thiolated receptor **Zn-Cyclen**₂ with vesicles **V-CF** (0.4 equivalents of receptor with respect to surface-accessible maleimide) and removed the template molecules by size-exclusion chromatography (SEC) after completion of the surface functionalization. For comparison, vesicle surfaces were post-functionalized with **Zn-Cyclen**₂ in absence of the bivalent peptide, but treated identically apart from that. Rebinding studies of the two obtained vesicle samples revealed a significantly different course of the fluorescence emission responses (Fig. 9). While the imprinted vesicles displayed a fluorescence decrease upon titration of up to 0.5 equivalents of peptide **Pep-P-His**, the non-imprinted samples showed a steady increase. After further peptide addition, the sign of the fluorescence signal change reversed in case of the imprinted vesicles and the curve progression approximated that of the non-imprinted ones.

This divergent behavior can be rationalized by different binding modes in the two cases. We hypothesize that the geometric confinement of the functional amphiphiles on the vesicle surface upon the formation of 2 : 1 complexes results in

a further quenching of the co-embedded fluorophores. Gradual transition into 1 : 1 binding mode after addition of more than 0.5 equivalents of the peptide is assumed to provide a higher translational freedom of the membrane-embedded compounds leading to an emission intensity increase of the dye molecules.²⁹ Due to the superposition of two opposing effects in the case of imprinted vesicles, the overall fluorescence response is much lower compared to that of the non-imprinted samples. The macroscopic binding constants in the two cases obtained by Hill fitting of the titration curves are $\lg K_a = 7.1$ for the imprinted vesicles (based on the addition of up to 0.8 equivalents of **Pep-P-His**) versus 5.1 for the control experiment.³⁰ However, due to the superposition of two diverging signal responses, the apparent affinity constant for the multi-point binding is likely to be underestimated.

Variation of the receptor concentration indicated that the success of vesicle surface patterning is dependent on the proportion of added receptor molecules and membrane-embedded maleimides. At an almost equimolar ratio (≈ 0.8 equivalents of receptor with respect to surface-accessible maleimide), the different binding behavior of imprinted and non-imprinted samples is much less pronounced (see Fig. S13, ESI†). We explain this result with the fact that here, approxi-



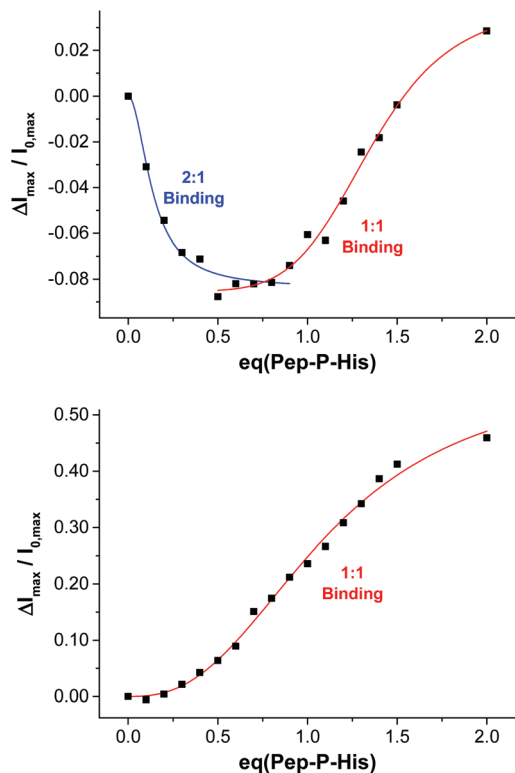


Fig. 9 Rebinding isotherms of imprinted (top) and non-imprinted (bottom) vesicles V-CF-Zn upon addition of Pep-P-His.

mately all surface-accessible maleimide molecules bear a receptor unit, *i.e.*, the receptor assembly simply reflects the random assembly of the membrane-embedded **Mal-C₁₆**. Thus, no external control over their spatial distribution is possible and the receptors cannot attain an appropriate geometric arrangement for multipoint binding to the analyte. Conversely, when immobilizing the receptor in sub-stoichiometric amounts, only membrane-embedded maleimides whose mutual distances are compatible to the formation of ternary receptor-template complexes are functionalized. The other maleimides remain unmodified and thereby do not impede the 2 : 1 binding on the vesicle surface.

2.4 Vesicle functionalization with an aptamer for ampicillin

In living systems, multipoint interactions dominate the molecular recognition phenomena and provide for remarkably high specificities. Thus, to explore the full potential of our technique, we next extended the vesicular post-functionalization strategy to more complex binding units of biological origin. Among those, oligonucleotide-based aptamers represent one of the most exciting and promising class of receptors. Since they are developed by an evolutionary process of repeated selection and amplification steps, aptamers exhibit very high binding affinities and specificities to their targets ranging from small organics to proteins and pathogens.³¹ In many cases, structurally related analytes can be discriminated on

basis of subtle differences such as the presence or absence of methyl or hydroxyl groups. Compared to other biological recognition elements like antibodies, aptamers display some remarkable advantages including low immunogenicity, small size, easy production and modification by established solid-phase synthesis methods and therefore inexpensive commercial availability. Moreover, they are characterized by a high stability and convenient handling. Fluorescence-based methods for the detection of aptamer–target binding typically rely on covalent modification of the oligonucleotides with luminescent reporter moieties.³² Another established technique involves hybridization of the aptamers with accordingly labeled complementary oligonucleotide strands and their subsequent displacement by the analyte molecules. This requirement of covalent attachment of fluorophores can make such assays laborious and costly. Label-free strategies on the other hand, *e.g.*, based on intercalation of a fluorescent dye into a double helix domain, are often limited in their applicability as they depend on particular properties of the aptamers such as specific folding modes upon complex formation with the target. In contrast to these analytical methods, the signaling mechanism of our vesicular sensors is generally supposed to originate rather from an analyte-induced redistribution of charges at the liposome–water interface than on defined conformational changes of the recognition sites. Therefore, aptamer-functionalized vesicles with co-embedded reporter dyes potentially represent versatile systems for the detection of aptamer–target interactions. The modular nature of such aptasensors based on self-assembly and subsequent membrane modification allows for their easy preparation and facile optimization of their composition. Recently, we reported on the post-functionalization of luminescent vesicles with the thrombin-binding aptamer resulting in liposomal sensors, which signal the binding of the protein analyte to their surface.¹⁴ To demonstrate the universal applicability of this approach for the preparation of various aptasensors, we utilized a DNA-based aptamer developed by Song *et al.* which specifically binds to the antibiotic ampicillin (Fig. 10) with an affinity constant of $\lg K_a = 8.0$.³³

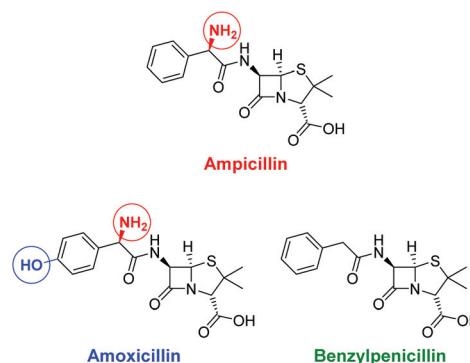


Fig. 10 Structures of the antibiotic ampicillin and of related control antibiotics amoxicillin and benzylpenicillin; structural differences are marked with colored circles.



For the selection of the ampicillin aptamer, the authors used TRIS buffer with added salts (50 mM NaCl, 5 mM KCl, 5 mM MgCl₂). The presence of cations such as K⁺ or Mg²⁺ is important to stabilize the folded analyte binding conformations of some aptamers. On the other hand, it is well explored that in particular bivalent cations can induce aggregation or even fusion of negatively charged vesicles (as it is the case for **V-CF**) by neutralizing their surface charge and thus by decreasing their electrostatic repulsion barrier.³⁴ In our case, it turned out that non-functionalized vesicles **V-CF** are indeed unstable and aggregate under the original buffer conditions. For that reason, we screened different buffer systems with respect to both vesicle stability and emission response of aptamer-functionalized vesicles after addition of ampicillin (see ESI† for details). Buffer systems without added salts did not result in significant fluorescence responses, whereas very high salt concentrations induced vesicle aggregation as determined by dynamic light scattering. HEPES buffer with added KCl (5 mM) and MgCl₂ (5 mM) represented the best compromise. The surface functionalization of maleimide-containing vesicles **V-CF** was performed with thiol-modified oligonucleotide **Apt-Amp** (Fig. 3). A time-dependent change of the emission signal again displayed the progress of the aptamer-immobilization on the vesicular membrane (Fig. 11).

In contrast to the post-functionalization of **V-CF** with receptor **Zn-Cyclen**₂, the fluorescence signal increased during the surface reaction. We believe that the presence of the highly negatively charged oligonucleotides at the vesicle surface partially displaces the likewise negatively charged carboxyfluorescein molecules from the clusters of the functional amphiphiles. As a consequence, a reduction of the fluorophore self-quenching occurs. Binding of ampicillin to the surface-immobilized aptamers is thought to induce a more compact conformation of the aptamer strands. Therefore, a higher negative charge density at the vesicle membrane is generated, which in turn enforces more dye molecules to be squeezed out of their membrane domains. This process gives rise to a significant further increase of the emission signal (Fig. 12).³⁵

The time-dependent fluorescence spectra depicted in Fig. 12 show a rather slow emission response of **V-CF-Amp**

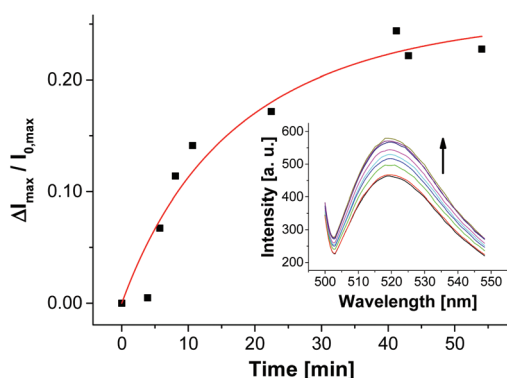


Fig. 11 Fluorescence response showing the immobilization progress of vesicles **V-CF** with aptamer **Apt-Amp** (1.0 mol%).

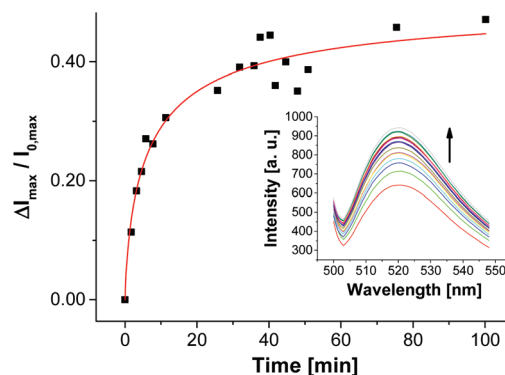


Fig. 12 Time-dependent change of fluorescence emission of aptamer-functionalized vesicles **V-CF-Amp** (2.0 mol% of **Apt-Amp**) after addition of ampicillin (0.40 μM).

upon addition of ampicillin. We assume that the aptamer-functionalized vesicles are more prone to salt-induced aggregation than the non-functionalized ones due to their higher surface charge.³⁴ This assumption is supported by investigations of the vesicular size distribution, which revealed a significant cross linking of the vesicles after functionalization with the oligonucleotide (see Fig. S15, ESI†).³⁶ This process in turn reduces the accessibility of the surface-bound aptamers for ampicillin. Binding of the analyte molecules to their membrane receptors, however, seems to partially reverse the aggregation as indicated by a subsequent decrease of the size distribution maximum of the vesicles.³⁷

Variation of the aptamer concentration and addition of ampicillin to the functionalized vesicles gave best emission responses of almost 50% with 2.0 mol% of oligonucleotides immobilized on the surface (Fig. 13). These conditions having approximately equimolar amounts of the receptor molecules with respect to the membrane-embedded fluorophores on the outer vesicle provide highest sensitivities, which is in accordance with our results from **Zn-Cyclen**₂-functionalized vesicles.

The specificity of the aptasensors **V-CF-Amp** towards ampicillin was illustrated by addition of the structurally related antibiotics benzylpenicillin or amoxicillin, which showed no significant fluorescence responses of the membrane-embedded carboxyfluorescein dyes (Fig. 13). Vesicles lacking the amphiphilic attachment site for thiols **Mal-C**₁₆ were prepared and incubated with the aptamer. Addition of ampicillin to that sample did not lead to a change of the emission. This observation proves that the analyte signaling depends on the interactions with the membrane-conjugated aptamers.

3. Conclusion

In summary, we have demonstrated that preformed, maleimide-decorated luminescent vesicles can be easily functionalized at the surface with different receptors by nucleophilic thiol addition. It was shown that the immobilized compounds may range from small organic molecules to peptides and



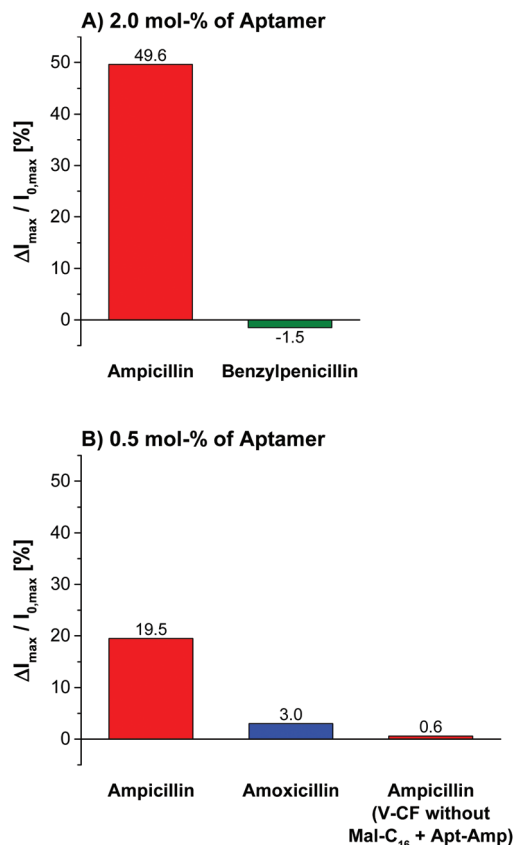


Fig. 13 Comparison of relative fluorescence emission responses of V-CF-Amp, functionalized with different amounts of the aptamer, and of a mixture of maleimide-lacking vesicles V-CF and Apt-Amp after addition of ampicillin, benzylpenicillin or amoxicillin (0.40 μM in each case).

metal complexes, such as bis- Zn^{2+} -cyclen, and to even more complex structures like oligonucleotides. The membrane-embedded fluorophors respond to the presence of the receptor units and to the formation of host-guest complexes on the vesicle surface with changing emission properties. We demonstrated potential applications of the concept including the detection of simple phosphates with surface-attached **Zn-Cyclen**₂ or the specific sensing of ampicillin by an aptamer. Moreover, we showed that the post-functionalization strategy allows external control over the two-dimensional distribution of the receptors by assistance of multivalent templates. Here, significant differences between imprinted and non-imprinted samples were obtained in the fluorescence output of rebinding titrations. Due to the highly modular approach, our presented strategy allows an easy, fast and inexpensive access to a broad variety of supramolecular indicators with tailor-made properties. The non-functionalized vesicles can be prepared in advance. Since many heterobifunctional reagents for the *in situ* introduction of thiol groups to amines are commercially available (e.g., Traut's reagent^{38a} or SATA^{38b}), our method is not only limited to thiol-containing host molecules, but can be readily extended to recognition elements based on other peptides, oligonucleotides or complex macromolecules.

Hence, the development of novel customized bioanalytical devices may be facilitated.

Acknowledgements

A. M. thanks the Bayerische Eliteförderung for a research fellowship.

Notes and references

- (a) J. Voskuhl and B. J. Ravoo, *Chem. Soc. Rev.*, 2009, **38**, 495–505; (b) K. Ariga, H. Ito, J. P. Hill and H. Tsukube, *Chem. Soc. Rev.*, 2012, **41**, 5800–5835.
- (a) K. A. Edwards and A. J. Baemner, *Talanta*, 2006, **68**, 1421–1431; (b) Q. Liu and B. J. Boyd, *Analyst*, 2013, **138**, 391–409.
- R. Martínez-Máñez and F. Sancenón, *Chem. Rev.*, 2003, **103**, 4419–4476.
- B. Gruber, S. Stadlbauer, K. Woinaroschy and B. König, *Org. Biomol. Chem.*, 2010, **8**, 3704–3714.
- T. W. Bell and N. M. Hext, *Chem. Soc. Rev.*, 2004, **33**, 589–598.
- B. T. Nguyen and E. V. Anslyn, *Coord. Chem. Rev.*, 2006, **250**, 3118–3127.
- (a) B. Gruber, S. Stadlbauer, A. Späth, S. Weiss, M. Kalinina and B. König, *Angew. Chem., Int. Ed.*, 2010, **49**, 7125–7128, (*Angew. Chem.*, 2010, **122**, 7280–7284); (b) S. Banerjee, M. Bhuyan and B. König, *Chem. Commun.*, 2013, **49**, 5681–5683.
- (a) M. Mammen, S.-K. Choi and G. M. Whitesides, *Angew. Chem., Int. Ed.*, 1998, **37**, 2754–2794, (*Angew. Chem.*, 1998, **110**, 2908–2953); (b) H. Jung, A. D. Robison and P. S. Cremer, *J. Struct. Biol.*, 2009, **168**, 90–94; (c) A. Grochmal, E. Ferrero, L. Milanesi and S. Tomas, *J. Am. Chem. Soc.*, 2013, **135**, 10172–10177.
- (a) L. Nobs, F. Buchegger, R. Gurny and E. Allémann, *J. Pharm. Sci.*, 2004, **93**, 1980–1992; (b) V. P. Torchilin, *Nat. Rev. Drug Discovery*, 2005, **4**, 145–160; (c) F. S. Hassane, B. Frisch and F. Schuber, *Bioconjugate Chem.*, 2006, **17**, 849–854; (d) R. I. Jøelck, L. N. Feldborg, S. Andersen, S. M. Moghimi and T. L. Andresen, *Adv. Biochem. Eng./Biotechnol.*, 2011, **125**, 251–280.
- (a) C. E. Hoyle and C. N. Bowman, *Angew. Chem., Int. Ed.*, 2010, **49**, 1540–1573, (*Angew. Chem.*, 2010, **122**, 1584–1617); (b) C. E. Hoyle, A. B. Lowe and C. N. Bowman, *Chem. Soc. Rev.*, 2010, **39**, 1355–1387.
- V. D. Bock, H. Hiemstra and J. H. van Maarseveen, *Eur. J. Org. Chem.*, 2006, 51–68.
- C. E. Hoyle, T. Y. Lee and T. Roper, *J. Polym. Sci., Part A: Polym. Chem.*, 2004, **42**, 5301–5338.
- (a) B. Frisch, C. Boeckler and F. Schuber, *Bioconjugate Chem.*, 1996, **7**, 180–186; (b) P. Schelte, C. Boeckler, B. Frisch and F. Schuber, *Bioconjugate Chem.*, 2000, **11**, 118–123; (c) R. J. Pounder, M. J. Stanford, P. Brooks,



- S. P. Richards and A. P. Dove, *Chem. Commun.*, 2008, 5158–5160.
- 14 A. Müller and B. König, *Chem. Commun.*, 2014, **50**, 12665–12668.
 - 15 Y. Kitano, Y. Nogata, K. Matsumura, E. Yoshimura, K. Chiba, M. Tada and I. Sakaguchi, *Tetrahedron*, 2005, **61**, 9969–9973.
 - 16 P. Wu and L. Brand, *Anal. Biochem.*, 1994, **218**, 1–13.
 - 17 J. Guy, K. Caron, S. Dufresne, S. W. Michnick, W. G. Skene and J. W. Keillor, *J. Am. Chem. Soc.*, 2007, **129**, 11969–11977.
 - 18 For examples of optical probes for thiol detection, see: X. Chen, Y. Zhou, X. Peng and J. Yoon, *Chem. Soc. Rev.*, 2010, **39**, 2120–2135.
 - 19 M. P. Lutolf, N. Tirelli, S. Cerritelli, L. Cavalli and J. A. Hubbell, *Bioconjugate Chem.*, 2001, **12**, 1051–1056.
 - 20 J. Li, Q. Xu, D. M. Cortes, E. Perozo, A. Laskey and A. Karlin, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 11605–11610.
 - 21 P. Schelté, C. Boeckler, B. Frisch and F. Schuber, *Bioconjugate Chem.*, 2000, **11**, 118–123.
 - 22 (a) T. Koike, S. Kajitani, I. Nakamura, E. Kimura and M. Shiro, *J. Am. Chem. Soc.*, 1995, **117**, 1210–1219; (b) E. Kimura, S. Aoki, T. Koike and M. Shiro, *J. Am. Chem. Soc.*, 1997, **119**, 3068–3076; (c) S. Aoki, M. Zulkefeli, M. Shiro, M. Kohsako, K. Takeda and E. Kimura, *J. Am. Chem. Soc.*, 2005, **127**, 9129–9139.
 - 23 With respect to a total concentration of **Mal-C₁₆** of 5.0 mol% and therefore a surface-accessible concentration of approximately 2.5 mol%, 0.8 equivalents of **Zn-Cyclen₂** equate to 2.0 mol% relating to the total amphiphile concentration.
 - 24 S. Lacerda, M. P. Campello, I. C. Santos, I. Santos and R. Delgado, *Polyhedron*, 2007, **26**, 3763–3773.
 - 25 G. Wulff, *Angew. Chem., Int. Ed. Engl.*, 1995, **34**, 1812–1832, (*Angew. Chem.*, 1995, **107**, 1958–1979).
 - 26 (a) B. Gruber, S. Balk, S. Stadlbauer and B. König, *Angew. Chem., Int. Ed.*, 2012, **51**, 10060–10063, (*Angew. Chem.*, 2012, **124**, 10207–10210); (b) S. Balk and B. König, *J. Inclusion Phenom. Macrocyclic Chem.*, 2014, DOI: 10.1007/s10847-014-0442-2; (c) S. Banerjee and B. König, *J. Am. Chem. Soc.*, 2013, **135**, 2967–2970.
 - 27 M. Ueno, S. Katoh, S. Kobayashi, E. Tomoyama, S. Ohsawa, N. Koyama and Y. Morita, *J. Colloid Interface Sci.*, 1990, **134**, 589–592.
 - 28 A. Grauer, A. Riechers, S. Ritter and B. König, *Chem. – Eur. J.*, 2008, **14**, 8922–8927.
 - 29 In our previously reported dynamic imprinting approaches,²⁶ we observed a steady increase of the signal in both cases, however, with the different binding affinities being reflected in different slopes of the binding isotherms. We explain this with the dynamic nature of the receptor recruitment process in that case providing a diverse arrangement of the fluorophore molecules relative to the receptor assemblies.
 - 30 The lower value of lg K_a in case of non-imprinted vesicles compared to that from Table 3 might be attributed to an incomplete separation of the template peptide by the SEC in that case. However, as the exactly identical conditions for SEC were applied for both the imprinted vesicles and the non-imprinted control solutions, the relative change of the binding affinities has to be considered.
 - 31 (a) S. D. Jayasena, *Clin. Chem.*, 1999, **45**, 1628–1650; (b) C. K. O'Sullivan, *Anal. Bioanal. Chem.*, 2002, **372**, 44–48.
 - 32 (a) E. J. Cho, J.-W. Lee and A. D. Ellington, *Annu. Rev. Anal. Chem.*, 2009, **2**, 241–264; (b) R. E. Wang, Y. Zhang, J. Cai, W. Cai and T. Gao, *Curr. Med. Chem.*, 2011, **18**, 4175–4184.
 - 33 K.-M. Song, E. Jeong, W. Jeon, M. Cho and C. Ban, *Anal. Bioanal. Chem.*, 2012, **402**, 2153–2161.
 - 34 S. Nir, J. Bentz, J. Wilschut and N. Düzgünes, *Prog. Surf. Sci.*, 1983, **13**, 1–124.
 - 35 The surface-immobilized **Zn-Cyclen₂** complexes, on the other hand, are supposed to electrostatically interact with the membrane-embedded carboxyfluorescein molecules resulting in a partial quenching of their fluorescence emission.
 - 36 Vesicle aggregation based on specific intermolecular interactions between surface-attached oligonucleotides is unlikely due to the absence of complementary sequences in the immobilized aptamer required for hybridization.
 - 37 We did not face that aggregation behavior in our previously reported studies regarding the vesicular thrombin-aptasensor.¹⁴ There, the presence of Mg²⁺ was not required for the conformational stabilization of the oligonucleotides. As a result, a dynamic and quick fluorescence response over a wide concentration range was observed.
 - 38 (a) R. Jue, J. M. Lambert, L. R. Pierce and R. R. Traut, *Biochemistry*, 1978, **17**, 5399–5406; (b) R. Julian, S. Duncan, P. D. Weston and R. Wigglesworth, *Anal. Biochem.*, 1983, **132**, 68–73.

