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The influence of fluorine spin-diffusion on ^{13}C solid-state NMR line shapes of CF_3 groups†

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Indirect spin–spin couplings (“J-couplings”) lead to well-known multiplet patterns in nuclear magnetic resonance (NMR) spectra that are also observable in non-decoupled solid-state NMR spectra, if the J-coupling constant exceeds the linewidth. Such J-multiplet line shapes in the solid state might however be affected by spin diffusion (SD) on the passive nuclei. When the SD rate constant is fast compared to the J-coupling constant, the multiplet resolution can be lost due to a so-called “self-decoupling” mechanism as has been already reported in the context of decoupling and for proton SD in solid adamantane. We herein report on the influence of ^{19}F SD on ^{13}C -detected solid-state NMR spectra of a small organic molecule bearing a trifluoromethyl group. The target compound is the chiral α -(trifluoromethyl)lactic acid (TFLA). Enantiopure phases ((R) or (S), respectively) of TFLA are composed of homochiral dimers whereas the racemic phase consists of heterochiral dimers in the solid state. Despite their structural similarity, the ^{13}C line shapes of the CF_3 group in cross-polarization spectra recorded at slow to medium magic-angle spinning (MAS) frequencies – in the range between 14.0 kHz and 60.0 kHz – differ substantially. By combining experimental observations, analytical calculations based on the Bloch–McConnell equations, and numerical spin-dynamics simulations, we demonstrate that differences in the ^{19}F SD rate constant between racemic and enantiopure TFLA-phases significantly affect the respective solid-state ^{13}C NMR spectral line shapes. Slowing down SD by increasing the MAS frequency restores the quartet line shape for both phases of TFLA.

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Introduction

The presence of fluorine has significant effects on the physico-chemical properties of molecules (for some selective reviews see ref. 1–9). From a chemical point-of-view, the high electronegativity of the fluorine atoms leads to compounds with

polarized carbon-fluorine bonds and, therefore, partially negatively charged fluorine atoms,¹⁰ which might cause electrostatic repulsion. In medicinal chemistry, the trifluoromethyl ($-\text{CF}_3$) group is often used as a bioisostere for a methyl ($-\text{CH}_3$) group.^{2,5,11–13} As a consequence, the molecule becomes more lipophilic, which often improves the desired bioactivity,¹⁴ the latter being, however, still a matter of debate.¹⁵ The presence of a CF_3 group also changes other physicochemical properties, such as volatility, boiling and melting point, as well as solubility.^{16,17} Furthermore, CF_3 groups have been reported to be engaged in amphiphilic noncovalent bonding, acting as both electrophiles and nucleophiles.¹⁸ In addition, the role of CF_3 groups as hydrogen-bond acceptors has recently been discussed (for a recent example see ref. 19).

Fluorine also plays an important role in sublimations of chiral compounds, where self-disproportionations of enantiomers have been observed.^{16,20} In such processes, the sublimation rates of enantiopure solid entities and their racemic counterparts differ.²¹ Soloshonok and co-workers have observed such phenomena for various compounds bearing CF_3 groups.¹⁶ For α -(trifluoromethyl)lactic acid (TFLA) discussed here, this

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behaviour leads to an enantiomeric enrichment over time for the remaining solid.²⁰ Because the crystal structure of the enantiopure compound contains homochiral dimers, whereas heterochiral dimers are found in the racemic form,²⁰ it was suggested that the solid-state molecular packing might explain the differences in the sublimation behaviour, for instance caused by electrostatic repulsions of CF₃ groups.^{16,20,22,23}

We have recently employed NMR spectroscopy in solution using an *in operando* setup²⁴ and in the solid state to investigate the influence of mechanochemistry, *e.g.*, ball milling and resonant-acoustic mixing, on the self-disproportionation process and on molecular-recognition events involving TFLA in general.^{24,25} In the course of such studies, we noticed that the ¹³C magic-angle spinning (MAS) NMR line shapes for the CF₃ groups differ significantly between the enantiopure and racemic phases of TFLA.²⁵ We herein report ¹³C-detected solid-state NMR MAS experiments to probe how differences in the ¹⁹F spin-diffusion (SD) rate constants for enantiopure and racemic TFLA affect the ¹³C NMR resonances of the corresponding CF₃ groups. Spin diffusion, which describes the energy-conserving transfer of polarization among dipolar-coupled nuclear spins,²⁶ might lead to the so-called “self-decoupling” mechanism^{27–31} affecting the ¹⁹F–¹³C J-multiplet line shape. Similar effects on ¹³C line shapes have for instance already been reported for fluorinated single-walled carbon nanotubes³² or flurbiprofen.²⁸

Theory

As reported previously, the effect of SD on J-coupled multiplet lines in solid-state NMR spectra can conveniently be described as an exchange process between the multiplet components using the Bloch–McConnell equations.^{27,33} For this purpose, a symmetry-adapted basis set for the three magnetically equivalent fluorine atoms in the CF₃ group needs to be constructed (which is obviously identical to the ones of protons in a methyl group). In CF₃, the three fluorine nuclei with C_{3v} symmetry generate eight spin states, which can be grouped by their symmetry into the three irreducible representations A, E_a and E_b, for which the symmetry-adapted basis sets are listed below.^{34–36}

The A manifold (symmetric to spin exchange), with four states:

$$\begin{aligned} &|\alpha\alpha\alpha\rangle \\ &(|\alpha\alpha\beta\rangle + |\alpha\beta\alpha\rangle + |\beta\alpha\alpha\rangle)3^{-\frac{1}{2}} \\ &(|\alpha\beta\beta\rangle + |\beta\alpha\beta\rangle + |\beta\beta\alpha\rangle)3^{-\frac{1}{2}} \\ &|\beta\beta\beta\rangle, \end{aligned} \quad (1)$$

the E_a manifold (non-symmetric to spin exchange), with two states:

$$\begin{aligned} &\left(|\alpha\alpha\beta\rangle + e^{\frac{i2\pi}{3}}|\alpha\beta\alpha\rangle + e^{-\frac{i2\pi}{3}}|\beta\alpha\alpha\rangle\right)3^{-\frac{1}{2}} \\ &\left(e^{-\frac{i2\pi}{3}}|\alpha\beta\beta\rangle + e^{\frac{i2\pi}{3}}|\beta\alpha\beta\rangle + |\beta\beta\alpha\rangle\right)\left(-3^{-\frac{1}{2}}\right) \end{aligned} \quad (2)$$

and the E_b manifold (non-symmetric to spin exchange), with two states:

$$\begin{aligned} &\left(|\alpha\alpha\beta\rangle + e^{-\frac{i2\pi}{3}}|\alpha\beta\alpha\rangle + e^{\frac{i2\pi}{3}}|\beta\alpha\alpha\rangle\right)3^{-\frac{1}{2}} \\ &\left(e^{\frac{i2\pi}{3}}|\alpha\beta\beta\rangle + e^{-\frac{i2\pi}{3}}|\beta\alpha\beta\rangle + |\beta\beta\alpha\rangle\right)\left(-3^{-\frac{1}{2}}\right) \end{aligned} \quad (3)$$

The four A states have a total group spin $\hat{F} = \frac{3}{2}$, while the four E_a and E_b states have a total group spin of $\hat{F} = \frac{1}{2}$. The only observable transitions are single-quantum transitions within states of the same irreducible representation, and transitions between different manifolds are forbidden. This leads to the expected quartet line shape in the ¹³C spectra of molecules bearing CF₃ groups with a 1:3:3:1 integral ratio (1:1:1:1 quartet caused by the A states and two 1:1 doublets caused by the two E-states). A schematic representation of the spin states for a CF₃ group is shown in Scheme 1 and the transition probabilities between these states under a perturbation of strength ω_i (dipolar coupling) is given by:²⁷

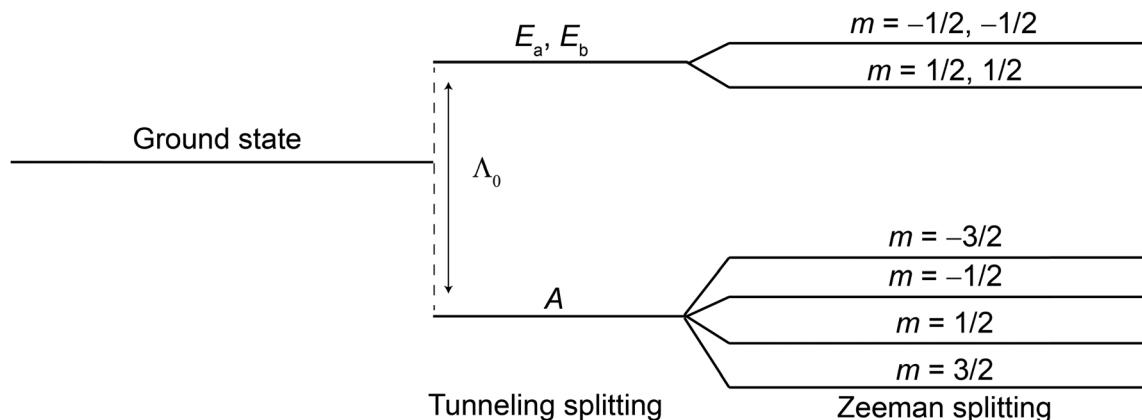
$$W_{m,m-1} = \frac{\pi}{2}\omega_i^2 f(0)(I+m)(I-m+1) = k_{\text{ex}}(I+m)(I-m+1), \quad (4)$$

wherein $f(0)$ is the zero-quantum line intensity at frequency zero,³⁷ while I represents the nuclear spin and m the magnetic quantum numbers.²⁷

Therefore, the Bloch–McConnell equations to describe SD for the $\hat{F} = \frac{1}{2}$ group spin are given by:

$$\begin{aligned} &\frac{d}{dt} \begin{pmatrix} M_x \left(+\frac{1}{2} \right) \\ M_x \left(-\frac{1}{2} \right) \end{pmatrix} \\ &= \begin{pmatrix} -k_{\text{ex}} - \frac{\pi}{T_2} - i\pi J_{\text{CF}} & k_{\text{ex}} \\ k_{\text{ex}} & -k_{\text{ex}} - \frac{\pi}{T_2} + i\pi J_{\text{CF}} \end{pmatrix} \begin{pmatrix} M_x \left(+\frac{1}{2} \right) \\ M_x \left(-\frac{1}{2} \right) \end{pmatrix} \end{aligned} \quad (5)$$





Scheme 1 Spin-states ground state energy level scheme for an isolated CF_3 group (without including the coupling to the ^{13}C spin, adapted from ref. 38). The energy difference between the A and E_a, E_b manifolds is the tunneling splitting Λ_0 .

and for the $\hat{F} = \frac{3}{2}$ group spin by:

$$\frac{d}{dt} \begin{pmatrix} M_x \left(+\frac{3}{2} \right) \\ M_x \left(+\frac{1}{2} \right) \\ M_x \left(-\frac{1}{2} \right) \\ M_x \left(-\frac{3}{2} \right) \end{pmatrix} = \begin{pmatrix} -3k_{\text{ex}} - \frac{\pi}{T_2} - 3i\pi J_{\text{CF}} & 3k_{\text{ex}} & 0 & 0 \\ 3k_{\text{ex}} & -7k_{\text{ex}} - \frac{\pi}{T_2} - i\pi J_{\text{CF}} & 4k_{\text{ex}} & 0 \\ 0 & 4k_{\text{ex}} & -7k_{\text{ex}} - \frac{\pi}{T_2} + i\pi J_{\text{CF}} & 3k_{\text{ex}} \\ 0 & 0 & 3k_{\text{ex}} & -3k_{\text{ex}} - \frac{\pi}{T_2} + 3i\pi J_{\text{CF}} \end{pmatrix} \begin{pmatrix} M_x \left(+\frac{3}{2} \right) \\ M_x \left(+\frac{1}{2} \right) \\ M_x \left(-\frac{1}{2} \right) \\ M_x \left(-\frac{3}{2} \right) \end{pmatrix} \quad (6)$$

Here, T_2 represents the inverse of the ^{13}C line width, which contains the coherent (incomplete MAS averaging of anisotropic interactions) and incoherent (stochastic relaxation) contributions to the residual line width of the ^{13}C line under MAS. k_{ex} stands for the ^{19}F SD rate constant and J_{CF} the ^{19}F – ^{13}C scalar spin–spin-coupling constant.

Results and discussion

The line shapes of the CF_3 group in ^{13}C -detected spectra of enantiopure and racemic TFLA differ

Fig. 1 shows the chemical structure of TFLA together with the experimental ^1H – ^{13}C cross-polarization (CP) solid-state NMR spectra of the racemic and enantiopure (*S*)-TFLA crystalline phases recorded at 17.5 kHz MAS frequency and at the same probe temperature. Note that the spectra were recorded under ^1H SPINAL-64 decoupling,³⁹ but no ^{19}F decoupling was applied. Fig. 1a clearly shows that the ^{13}C line shape for the CF_3 resonances at ~ 124 ppm differs significantly between the two samples. While the expected 1:3:3:1 quartet caused by the three one bond ^{19}F – ^{13}C scalar spin–spin couplings for the three magnetically equivalent fluorine nuclei is observed for the CF_3 group of *rac*-TFLA, the enantiopure sample shows strong line broadening with a “quartet-like” line shape buried

underneath a broad feature. Fig. S1 (ESI[†]) shows the comparison of the spectra of (*S*)-TFLA and (*R*)-TFLA, illustrating that

this line shape is observed for both enantiomeric compounds whose spectral properties are, as expected, identical.

To determine whether this broadening originates from carbon–fluorine indirect spin–spin interactions (J-couplings), ^{13}C -detected ^{19}F – ^{13}C CP spectra of (*S*)-TFLA with and without ^{19}F high-power decoupling during data acquisition have been recorded and are shown in Fig. 1b (note that in these experiments no ^1H decoupling was applied). Indeed, a similar line shape as in Fig. 1a can be observed in case of non- ^{19}F decoupled spectra, while the expected significantly sharper singlet is detected in the ^{19}F -decoupled spectra, pointing to ^{19}F – ^{13}C spin–spin couplings contributing to the broad ^{13}C resonance. This motivated us to further investigate the reasons behind the observed line shape differences between the enantiopure and racemic compounds.

The effect of ^{19}F SD on the ^{13}C NMR multiplet line shape of CF_3 groups

To address this observation, we simulated how the ^{19}F SD affects the CF_3 multiplet line. The role of ^1H SD on ^{13}C NMR line shapes has been reported in detail for adamantane, for which the influence of “self-decoupling” of the ^1H – ^{13}C J-coupling by proton SD has been discussed.²⁷ Also in context of ^{19}F – ^{13}C spin pairs this effect has been reported for the monofluorinated molecule flurbiprofen.²⁸





Fig. 1 (a) ^1H - ^{13}C CP spectra of (S)- and *rac*-TFLA recorded at 17.5 kHz and 16.4 T static magnetic-field strength. The chemical structure of TFLA is shown. (b) ^{19}F - ^{13}C CP spectra of (S)-TFLA recorded at 15.0 kHz and 11.7 T with and without ^{19}F high-power decoupling during data acquisition. The homochiral and heterochiral dimeric units taken from the crystal structures of (S)- and *rac*-TFLA (CSD numbers: (S)-TFLA refined from 666327 – for more details see Experimental section – and *rac*-TFLA 666328), respectively, are additionally shown.²⁰ * denotes MAS spinning sidebands. Signal truncation in a results from too short acquisition times.

Fig. 2 shows simulated ^{13}C spectra for the CF₃ multiplet line as a function of the SD rate constant k_{ex} (fixing the transverse ^{13}C relaxation time, T_2 , to a constant value) using Bloch-McConnell equations as introduced above.²⁷ Depending on the ratio of k_{ex} and the magnitude of the ^{19}F - ^{13}C J-coupling

constant, different line shapes are observed, consistent with three exchange-like regimes. For the case of $k_{\text{ex}} \ll 2\pi J_{\text{CF}}$ (similar to the slow exchange regime), the result is a perfectly resolved quartet multiplet. In case of coalescence $k_{\text{ex}} \approx 2\pi J_{\text{CF}}$, only a single line is observed whose linewidth decreases for



Fig. 2 Simulation of the ^{13}C NMR spectrum of a CF_3 group affected by ^{19}F SD assuming a ^{19}F – ^{13}C J-coupling constant of 280 Hz and a linewidth of 10 Hz ($\text{FWHM} = 1/(\pi T_2)$). The spectra have been simulated as a function of the SD rate constant k_{ex} .

$k_{\text{ex}} \gg 2\pi J_{\text{CF}}$ (fast exchange regime). In the fast exchange regime, the sharp resonance is denoted as “self-decoupled”.

The SD rate constant scales down with increasing MAS frequency, since second-order terms in the effective Hamiltonian, which dominate the SD process, scale with the inverse of the spinning frequency.⁴⁰ Therefore, we turned to MAS-dependent ^{13}C CP-spectra of (*S*)-TFLA to investigate whether slowing down ^{19}F SD has an effect on the CF_3 multiplet line. As a matter of fact, ^{13}C NMR spectra recorded at faster MAS frequencies would result in the expected CF_3 line shape featuring the 1:3:3:1 quartet also for the enantiopure sample, assuming the main contribution to the observed line broadening comes from ^{19}F SD. Fig. 3 shows the ^{13}C -detected CP spectra of (*S*)-(a) and *rac*-TFLA (b) recorded at MAS frequencies ranging from 14.0 kHz to 60.0 kHz (spectra recorded in 3.2 mm and 1.3 mm rotors under high-power ^1H SPINAL-64 decoupling³⁹). For both substances, a narrowing of the multiplet lines and an increase in resolution of the quartet line shape is

observed with increasing MAS frequency. In (*S*)-TFLA (Fig. 3a), the quartet becomes resolved at around 30.0 kHz MAS and above, while decreasing the MAS to 14.0 kHz produces a significantly broadened resonance as reported before. Similar observations can be drawn for the racemic sample (Fig. 3b), although the quartet becomes resolved at slightly lower MAS frequencies already. Fig. 3 also shows the simulations of the spectra (orange curves) based on a nonlinear least-square fit of the SD rate constant, k_{ex} , and the line width ($\text{FWHM} = 1/(\pi T_2)$) according to the Bloch–McConnell equations (for more details see the Experimental Section). The comparison of the two sets of spectra and the SD rate constants taken from the simulations indicates that the SD rate is approximately a factor of three to four larger for (*S*)-TFLA compared to the racemic case (*vide infra*).

Plotting the resulting fit of k_{ex} against the inverse of the MAS frequency in Fig. 4 reveals the expected linear decrease in k_{ex} for faster MAS frequencies as well as the mentioned factor of three to four difference in magnitude of k_{ex} between enantiopure and racemic TFLA. The fitted k_{ex} -values obtained from a nonlinear



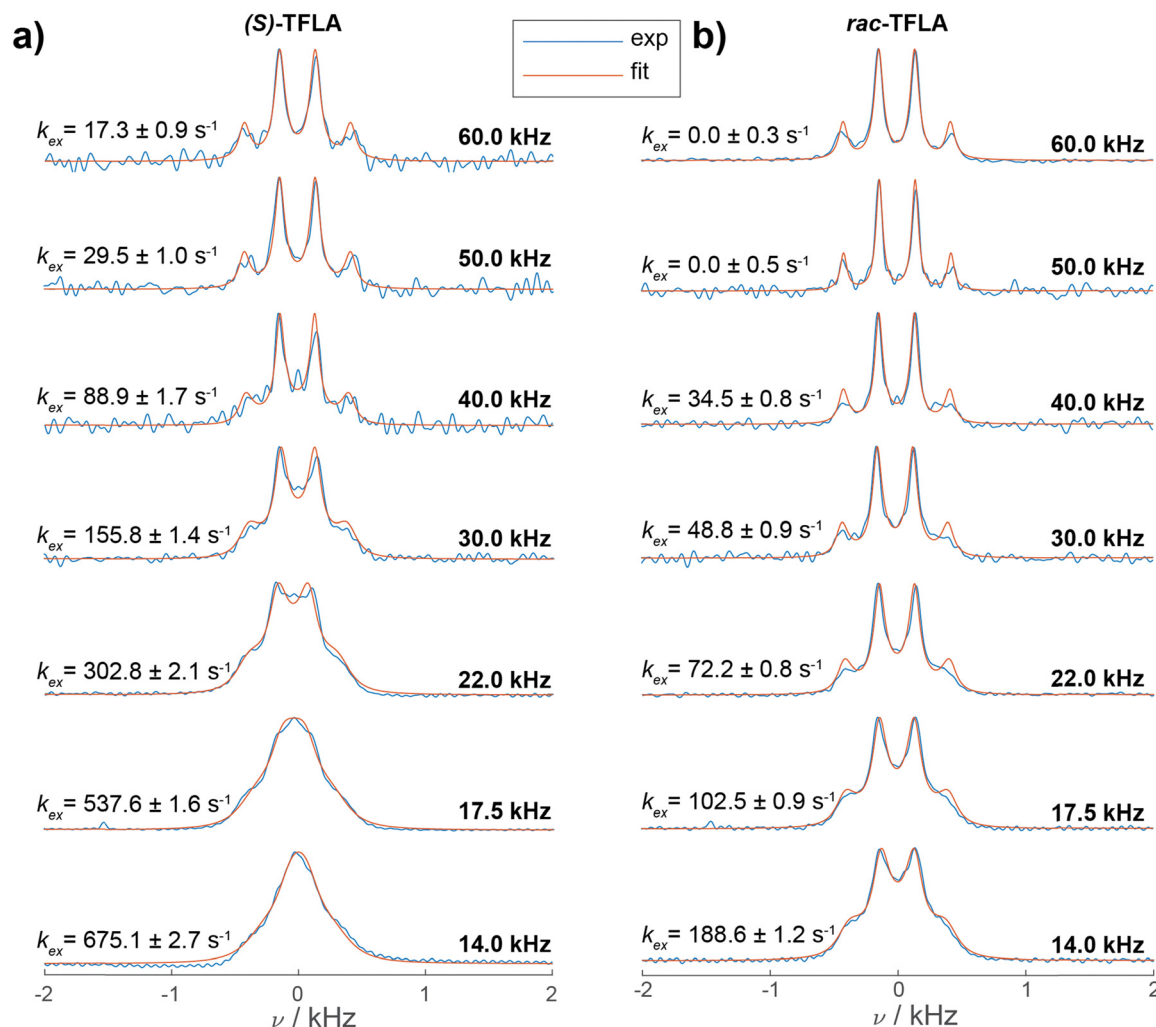


Fig. 3 ^1H - ^{13}C CP spectra of (a) (S)- and (b) *rac*-TFLA recorded at various MAS frequencies and at 16.4 T (blue curves). The spectra measured at 14.0, 17.5 and 22.0 kHz MAS were recorded in a 3.2 mm rotor, the spectra at faster MAS in a 1.3 mm rotor. Displayed is a zoom in the ^{13}C CF_3 spectral region for the two compounds. For each spectrum the simulated spectrum resulting from Bloch–McConnell equations using as variable parameters the optimal values of T_2 and k_{ex} obtained from a nonlinear least-square fit are shown (orange curves, for details regarding the error analysis see ESI†).

least-square minimization fit of the spectra at various MAS frequencies correspond to local minima, for which however we expect relatively large errors in the obtained rate constants. To determine the goodness-of-fit, we compared the 1D fit results to a 2D minimum χ^2 -estimation with variable T_2 - and k_{ex} -values (Fig. S2, ESI†) indeed showing that the minima reported in Fig. 3 are a good estimate for the rate constant.

The ^{19}F SD can alternatively be slowed down by applying off-resonance radio frequency (rf) irradiation on the ^{19}F nuclei, thereby scaling the homonuclear ^{19}F dipolar interaction and, thus, the SD rate constants as $(P_2(\cos\theta))^2$. Note that this will also scale the heteronuclear J-coupling, but with a different scaling factor, namely with $\cos\theta$. Such experiments have been performed for solid adamantane before.²⁷ Fig. 5a shows a series of ^{19}F - ^{13}C CP spectra of (S)- and *rac*-TFLA recorded at 15.0 kHz MAS and with variation of the angle of the fluorine effective field during data acquisition with respect to the static magnetic field direction, θ , as shown schematically in Fig. 5b. The angle θ

was varied between 20° and 54.7° (magic angle), with the 90° case representing the control experiment. At the magic angle, $\theta = \arccos(1/\sqrt{3})$, the homonuclear ^{19}F flip-flop term in the effective NMR-Hamiltonian vanishes and the contributions of ^{19}F SD to the line shape should, therefore, be eliminated. In such a scenario, the expected quartet line shape should be observed. As can be concluded from the spectra, there is a clear deviation from both a symmetric quartet line shape when setting θ to the magic angle and a single decoupled line for $\theta = 90^\circ$, which is caused by second-order contributions between a rather large ^{19}F chemical-shielding anisotropy (CSA) and the heteronuclear dipolar coupling under CW irradiation.^{41,42}

To examine whether the experimentally observed line shapes under off-resonance CW irradiation are compatible with the determined parameters, numerical simulations of the CF_3 group including SD have been performed. The rf-field amplitude and the irradiation offsets were set to the same values as in the experimental measurements of Fig. 5a using an unscaled



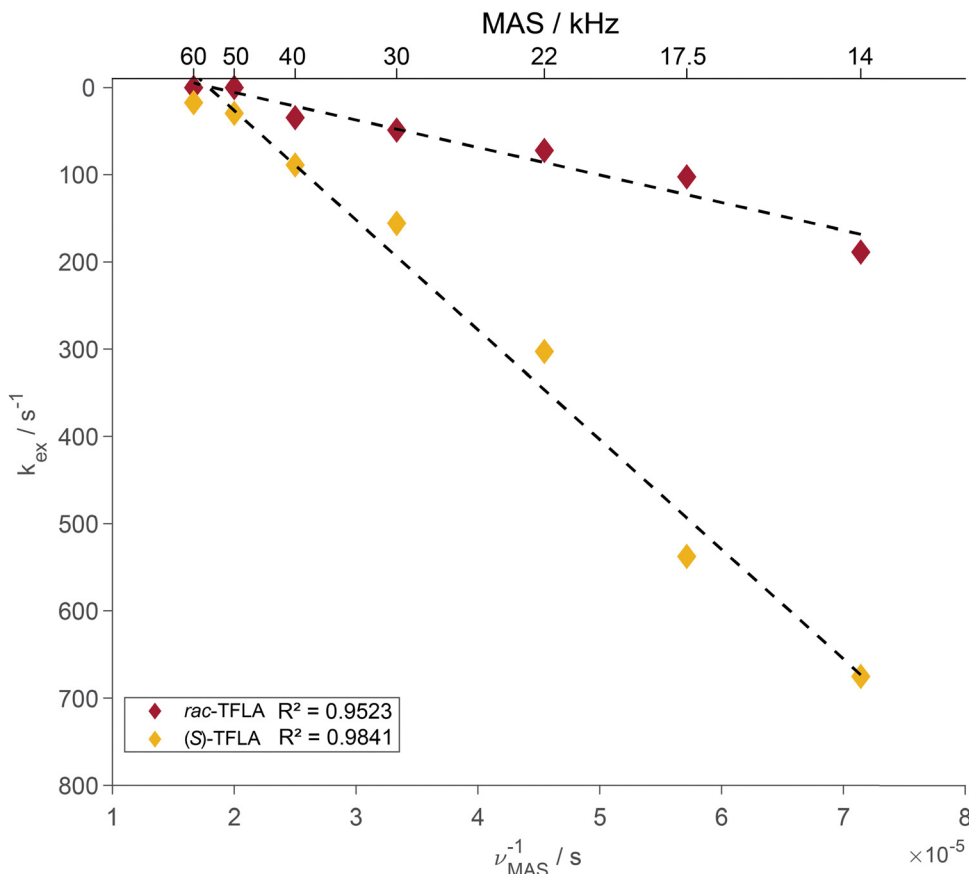


Fig. 4 Plot of the dependence of the fitted k_{ex} -values (obtained from 1D nonlinear least-square fits of the experimental and simulated spectra) on the inverse of the MAS frequency, $1/\nu_{\text{MAS}}$. The corresponding value of MAS frequency is reported on the x-axis on top for each datapoint. Dashed lines are the linear regressions of the datapoints for both enantiopure and racemic TFLA samples. The R^2 -values resulting from the linear regressions are reported in the legend.

exchange-rate constant of $k_{\text{ex}} = 160 \text{ s}^{-1}$ for *rac*-TFLA and a value of 670 s^{-1} for (S)-TFLA as determined above. The agreement between the simulations and the experimental data is not perfect, but the general features of the line shapes are reproduced quite well. Furthermore, as can be observed by comparing the *rac*- and (S)-TFLA spectra in Fig. 5a and c, the CF_3 line shapes for the two compounds become very similar if the SD is quenched near an effective field along the magic angle indeed supporting the assumption of different SD rate constants for the two compounds.

We initially assumed that the different SD rate constants are caused by differences in the homonuclear ^{19}F - ^{19}F dipolar-coupling network in the two TFLA-phases, which might also explain the differences in their ^{19}F static spin echo lineshapes (Fig. S3, see Fig. S4 for the corresponding MAS spectra, ESI†). We thus calculated the square root of the sum of squared dipolar couplings,⁴³ d_i^{RSS} , using the published and refined crystal structures[§] of *rac*- and (S)-TFLA, respectively²⁰ (for more

details see Experimental section). The resulting d_i^{RSS} taking only intermolecular CF_3 group contacts into account and calculated between the centers of mass of the three fluorine nuclei in the CF_3 groups are reported in Table S1 (ESI†) and amount to 11.4 kHz for *rac*-TFLA and 11.6 kHz for (S)-TFLA. The d_i^{RSS} -values have also been calculated taking an explicit averaging over all possible combinations on the circle into account and assuming “uncorrelated” rotations of the involved CF_3 groups (10.6 kHz for *rac*- and 9.7 kHz for (S)-TFLA). In both cases, the d_i^{RSS} -values do not reflect the expected difference by a factor of $\sqrt{3}$ as predicted from the experimentally observed ratio of three of the SD rate constants. We also note that reducing particle size by means of milling the sample does not affect the line shape, e.g., due to reduced longitudinal relaxation times upon milling samples.⁴⁴

Although the detailed reason for the different SD rate constants remains currently unclear, we note that the spatial orientations of the two CF_3 groups in the dimers differ (Fig. 1b). While the CF_3 groups in (S)-TFLA are oriented face-to-face with an inversion centre between them (e.g., the ^{19}F CSA- and the ^{19}F - ^{19}F dipolar-coupling tensor are co-linear), they are stacked in *rac*-TFLA (non-linear tensor orientations). The importance of such geometric effects for instance

§ We note here that the (S)-TFLA crystal structure has been refined based on the deposited structure (CSD number: 666327, ref. 20) (see ESI† Fig. S5), since the experimentally observed PXRD pattern does not match with the predicted one (for details see ref. 25).



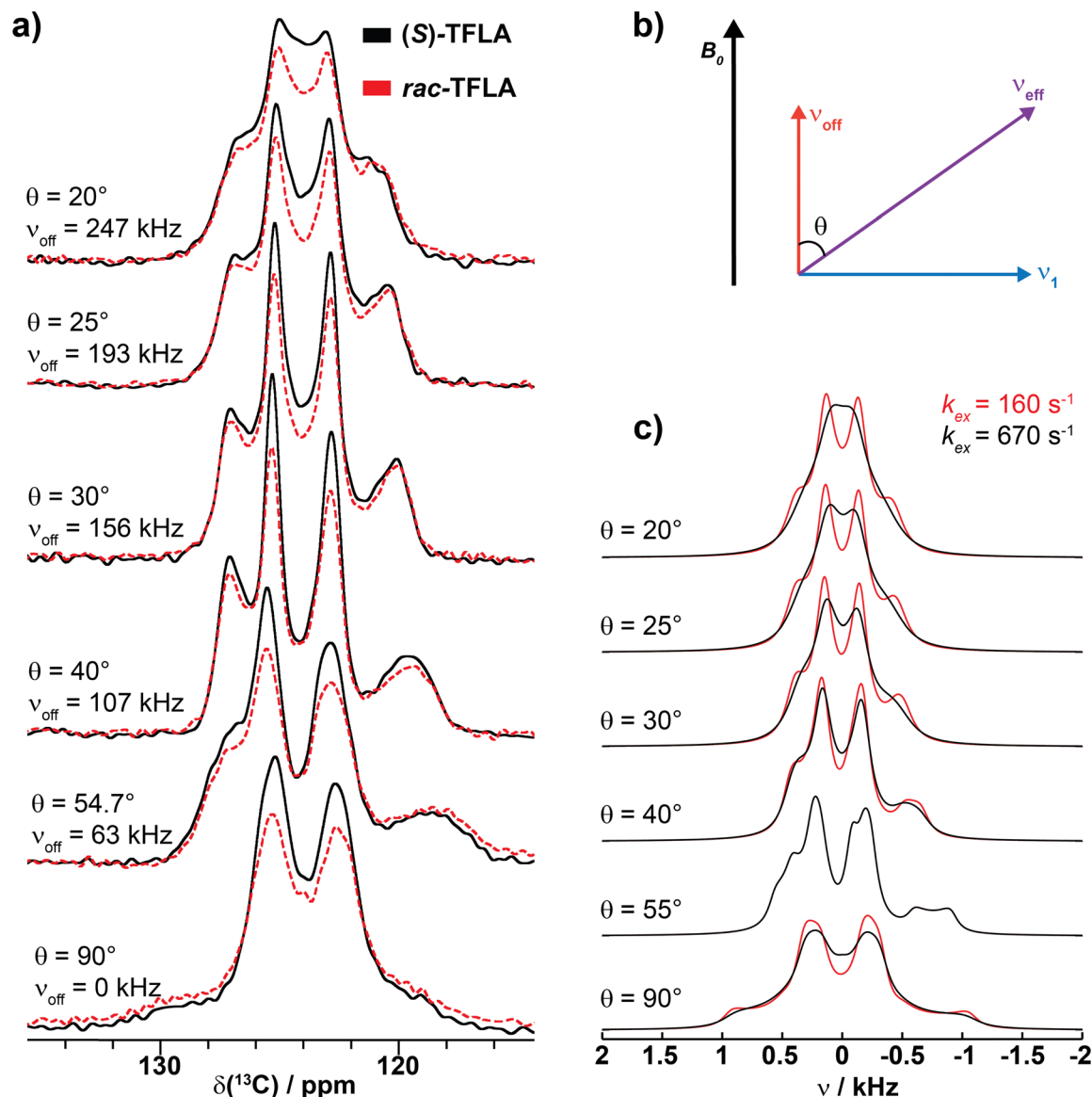


Fig. 5 (a) ^{19}F - ^{13}C CP spectra of the CF_3 spectral region for racemic (dashed red curve) and (*S*)-TFLA (straight black curve) recorded at 15.0 kHz MAS under off-resonance ^{19}F decoupling. The angle θ of the effective ^{19}F field with respect to B_0 during CW decoupling varies as shown in the figure from 90° to 20° . (b) schematic representation for the calculations of the effective field during CW decoupling. (c) Numerical simulations of off-resonance ^{19}F decoupled spectra as a function of θ and with $k_{\text{ex}} = 160 \text{ s}^{-1}$ representing *rac*-TFLA and $k_{\text{ex}} = 670 \text{ s}^{-1}$ representing (*S*)-TFLA. The simulations are based on the following parameters: $J(^{19}\text{F}-^{13}\text{C}) = -280 \text{ Hz}$, $J(^{19}\text{F}-^{19}\text{F}) = 100 \text{ Hz}$, $\delta(^{19}\text{F}-^{13}\text{C})/2\pi = 7870 \text{ Hz}$, $\delta(^{19}\text{F}-^{19}\text{F})/2\pi = 10700 \text{ Hz}$, $\delta_\sigma(^{19}\text{F})/2\pi = 26352 \text{ Hz}$ with all tensors oriented along the rotation axis of the CF_3 group. The MAS frequency was set to 15.0 kHz, B_1 to 90 kHz, and the offsets values as indicated in the figure.

on $^{31}\text{P}\{^{19}\text{F}\}$ REDOR dephasing curves involving CF_3 groups has been reported.⁴⁵ The differences in the SD rate constants might be caused by dynamics. Similar ^{13}C and ^{19}F T_1 relaxation times for the CF_3 groups in *rac*- and (*S*)-TFLA point to fast rotations of these groups with correlation times in the order of 10–100 ps for both samples. The above-mentioned face-to-face orientation of the CF_3 -

groups in (*S*)-TFLA might however lead to faster SD if a correlated (cooperative) rotation of such groups exist, as for instance reported for the CF_3 -group in 3-(trifluoromethyl)phenanthrene.^{46,47} We have currently no direct experimental evidence for this hypothesis and will further explore this effect in our laboratories.

Conclusions

We show that ^{13}C -detected solid-state NMR spectra of molecules containing CF_3 groups can be affected by ^{19}F spin diffusion leading to line broadening and sometimes even precluding the observation of the expected quartet multiplet at slow to

† As a final remark, we point out that we investigated two additional samples containing a CF_3 group, namely a supported-ionic liquid phase (SILP) containing bistriflimide as an anion, and the organic molecule 4-(trifluoromethyl)benzene-1-carboximidamide hydrochloride hydrate (data discussed in ESI,[†] Fig. S6). For both cases, the CF_3 groups spectra show a resolved quartet, in agreement with the slow exchange-regime scenario of Fig. 2.

moderate MAS frequencies in spectra without ^{19}F decoupling. While in the case of small ^{19}F spin-diffusion rate constants, the expected quartet line shape can be observed, this is not the case for faster rate constants. Our study on the small organic molecule TFLA crystallizing as enantiopure and racemic phases illustrates the high sensitivity of solid-state NMR to small differences in the ^{19}F spin-diffusion rate constants by using the CF_3 group as a highly sensitive marker. We hypothesize that these differences could be the result of correlated or uncorrelated motion of the two face-to-face oriented CF_3 groups.

Author contributions

E. B. and I. D. A. S. recorded the solid-state NMR experiments. C. Q. prepared and provided the TFLA samples. F. M. performed the PXRD structure refinement for (S)-TFLA. E. B. and M. E. performed the Bloch-McConnell and the numerical simulations. E. B., M. E. and T. W. carried out the analysis and interpretation of the data and wrote an initial version of the manuscript. All co-authors contributed to the writing of the final version of the manuscript. M. E. and T. W. designed the research, which was supervised by M. Z., C. B., M. E. and T. W.

Data availability

The scripts and data used in this manuscript are available through a public Github repository hosted at https://github.com/ebartalucci/CF3_self_decoupling.git.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 T. Liang, C. N. Neumann and T. Ritter, *Angew. Chem., Int. Ed.*, 2013, **52**, 8214–8264.
- 2 E. P. Gillis, K. J. Eastman, M. D. Hill, D. J. Donnelly and N. A. Meanwell, *J. Med. Chem.*, 2015, **58**, 8315–8359.
- 3 S. Purser, P. R. Moore, S. Swallow and V. Gouverneur, *Chem. Soc. Rev.*, 2008, **37**, 320–330.
- 4 G. Shabir, A. Saeed, W. Zahid, F. Naseer, Z. Riaz, N. Khalil, M. Muneeba and F. Albericio, *Pharmaceuticals*, 2023, **16**, 1162.
- 5 A. S. Nair, A. K. Singh, A. Kumar, S. Kumar, S. Sukumaran, V. P. Koyiparambath, L. K. Pappachen, T. M. Rangarajan, H. Kim and B. Mathew, *Processes*, 2022, **10**, 2054.
- 6 E. A. Ilardi, E. Vitaku and J. T. Njardarson, *J. Med. Chem.*, 2014, **57**, 2832–2842.
- 7 D. O'Hagan, *J. Fluorine Chem.*, 2010, **131**, 1071–1081.
- 8 M. Inoue, Y. Sumii and N. Shibata, *ACS Omega*, 2020, **5**, 10633–10640.
- 9 Q. Wang, Y. Bian, G. Dhawan, W. Zhang, A. E. Sorochinsky, A. Makarem, V. A. Soloshonok and J. Han, *Chin. Chem. Lett.*, 2024, **35**, 109780.
- 10 D. O'Hagan, *Chem. Soc. Rev.*, 2008, **37**, 308–319.
- 11 H. L. Yale, *J. Med. Pharm. Chem.*, 1959, **1**, 121–133.
- 12 N. A. Meanwell, *J. Med. Chem.*, 2018, **61**, 5822–5880.
- 13 C.-C. Tseng, G. Baillie, G. Donvito, M. A. Mustafa, S. E. Juola, C. Zanato, C. Massarenti, S. Dall'Angelo, W. T. A. Harrison, A. H. Lichtman, R. A. Ross, M. Zanda and I. R. Greig, *J. Med. Chem.*, 2019, **62**, 5049–5062.
- 14 B. E. Smart, *J. Fluorine Chem.*, 2001, **109**, 3–11.
- 15 A. Abula, Z. Xu, Z. Zhu, C. Peng, Z. Chen, W. Zhu and H. A. Aisa, *J. Chem. Inf. Model.*, 2020, **60**, 6242–6250.
- 16 J. Han, A. Wzorek, K. D. Klika and V. A. Soloshonok, in *Frontiers of Organofluorine Chemistry*, ed. I. Ojima, World Scientific, 2020, pp. 283–340.
- 17 J. Wang, M. Sánchez-Roselló, J. L. Aceña, C. del Pozo, A. E. Sorochinsky, S. Fustero, V. A. Soloshonok and H. Liu, *Chem. Rev.*, 2014, **114**, 2432–2506.
- 18 C. Esterhuysen, A. Heßelmann and T. Clark, *ChemPhysChem*, 2017, **18**, 772–784.
- 19 M. Wang, N. Garrison, P. M. Nguyen, A. Prasad, Y. Wang, H.-K. Kwon, G. Kim, M. A. Siegler and T. Lectka, *J. Org. Chem.*, 2024, **89**, 9681–9685.
- 20 V. A. Soloshonok, H. Ueki, M. Yasumoto, S. Mekala, J. S. Hirschi and D. A. Singleton, *J. Am. Chem. Soc.*, 2007, **129**, 12112–12113.
- 21 J. Han, O. Kitagawa, A. Wzorek, K. D. Klika and V. A. Soloshonok, *Chem. Sci.*, 2018, **9**, 1718–1739.
- 22 R. Tonner, V. A. Soloshonok and P. Schwerdtfeger, *Phys. Chem. Chem. Phys.*, 2011, **13**, 811–817.
- 23 M. A. Suhm and M. Albrecht, *Phys. Chem. Chem. Phys.*, 2011, **13**, 4159–4160.
- 24 F. Puccetti, T. Rinesch, S. Suljić, K. Rahimi, A. Herrmann and C. Bolm, *Chemistry*, 2023, **9**, 1318–1332.
- 25 C. Quaranta, I. D. A. A. Silva, S. Moos, E. Bartalucci, L. Hendrickx, B. M. D. Fahl, C. Pasqualini, F. Puccetti,



- M. Zobel, C. Bolm and T. Wiegand, *Angew. Chem., Int. Ed.*, 2024, **63**, e202410801.
- 26 N. Bloembergen, *Physica*, 1949, **15**, 386–426.
- 27 M. Ernst, A. Verhoeven and B. H. Meier, *J. Magn. Reson.*, 1998, **130**, 176–185.
- 28 G. Antonioli and P. Hodgkinson, *J. Magn. Reson.*, 2004, **168**, 124–131.
- 29 G. Sinning, M. Mehring and A. Pines, *Chem. Phys. Lett.*, 1976, **43**, 382–386.
- 30 M. Mehring and G. Sinning, *Phys. Rev. B: Solid State*, 1977, **15**, 2519–2532.
- 31 M. Mehring, G. Sinning and A. Pines, *J. Phys. B*, 1976, **24**, 73–76.
- 32 L. B. Alemany, L. Zhang, L. Zeng, C. L. Edwards and A. R. Barron, *Chem. Mater.*, 2007, **19**, 735–744.
- 33 H. M. McConnell, *J. Chem. Phys.*, 1958, **28**, 430–431.
- 34 J. Tang and A. Pines, *J. Chem. Phys.*, 1980, **72**, 3290–3297.
- 35 J.-N. Dumez, P. Håkansson, S. Mamone, B. Meier, G. Stevanato, J. T. Hill-Cousins, S. S. Roy, R. C. D. Brown, G. Pileio and M. H. Levitt, *J. Chem. Phys.*, 2015, **142**, 044506.
- 36 M. Negroni, D. Guarin, K. Che, L. M. Epasto, E. Turhan, A. Selimović, F. Kozak, S. Cousin, D. Abergel, G. Bodenhausen and D. Kurzbach, *J. Phys. Chem. B*, 2022, **126**, 4599–4610.
- 37 D. Suter and R. R. Ernst, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 1985, **32**, 5608–5627.
- 38 K. Morimoto, *J. Phys. Soc. Jpn.*, 1981, **50**, 2404–2412.
- 39 B. M. Fung, A. K. Khitrin and K. Ermolaev, *J. Magn. Reson.*, 2000, **142**, 97–101.
- 40 M. Chávez, T. Wiegand, A. A. Malär, B. H. Meier and M. Ernst, *Magn. Reson.*, 2021, **2**, 499–509.
- 41 M. Ernst, S. Bush, A. C. Kolbert and A. Pines, *J. Chem. Phys.*, 1996, **105**, 3387–3397.
- 42 M. Ernst, *J. Magn. Reson.*, 2003, **162**, 1–34.
- 43 V. E. Zorin, S. P. Brown and P. Hodgkinson, *J. Chem. Phys.*, 2006, **125**, 144508.
- 44 K. E. Dempah, J. W. Lubach and E. J. Munson, *Mol. Pharmaceutics*, 2017, **14**, 856–865.
- 45 E. A. Louie, P. Chirakul, V. Raghunathan, S. T. Sigurdsson and G. P. Drobny, *J. Magn. Reson.*, 2006, **178**, 11–24.
- 46 X. Wang, F. B. Mallory, C. W. Mallory, P. A. Beckmann, A. L. Rheingold and M. M. Francl, *J. Phys. Chem. A*, 2006, **110**, 3954–3960.
- 47 P. A. Beckmann, J. Rosenberg, K. Nordstrom, C. W. Mallory and F. B. Mallory, *J. Phys. Chem. A*, 2006, **110**, 3947–3953.

