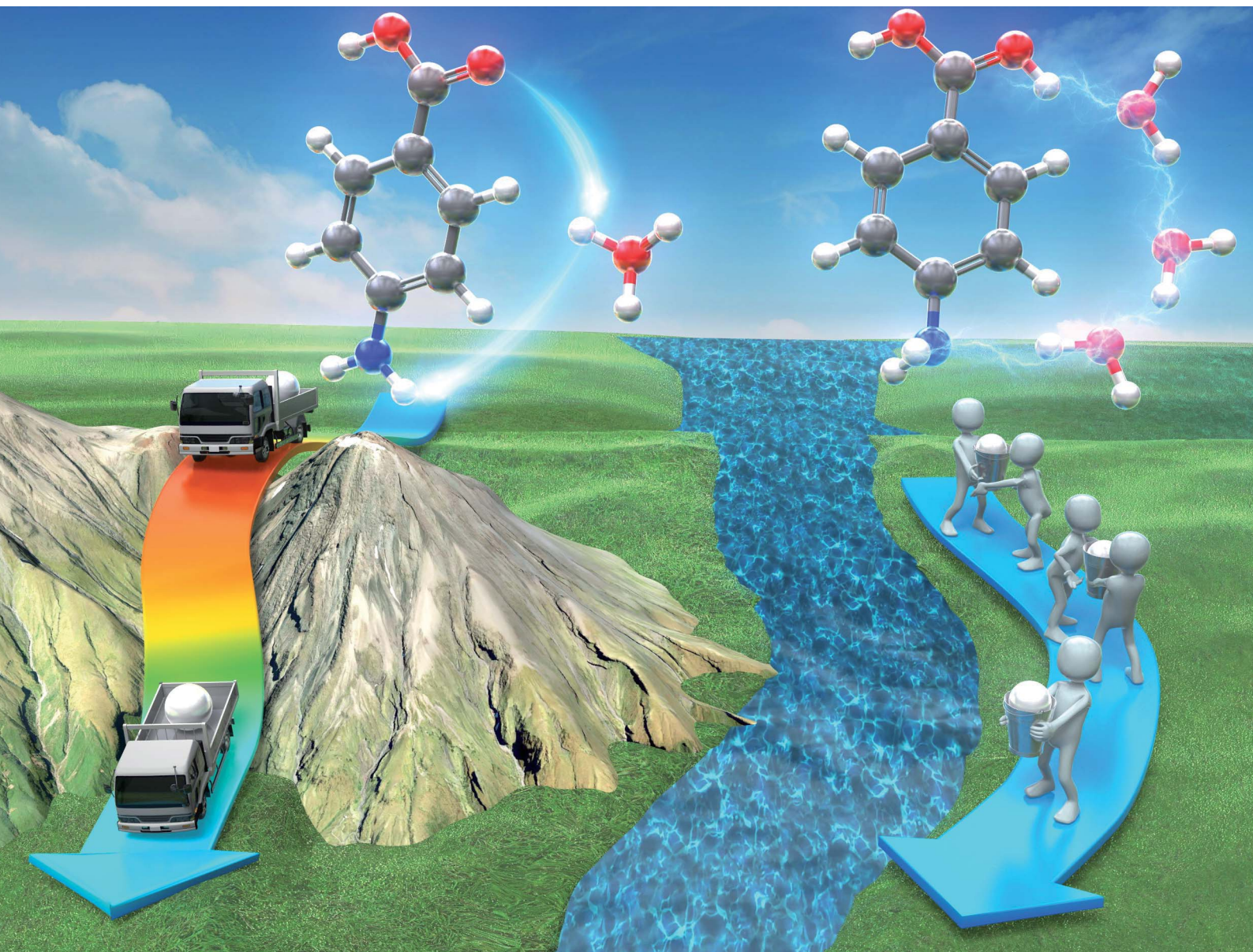


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Transition from vehicle to Grotthuss proton transfer in a nanosized flask: cryogenic ion spectroscopy of protonated *p*-aminobenzoic acid solvated with D₂O†

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Proton transfer (PT) is one of the most ubiquitous reactions in chemistry and life science. The unique nature of PT has been rationalized not by the transport of a solvated proton (vehicle mechanism) but by the Grotthuss mechanism in which a proton is transported to the nearest proton acceptor along a hydrogen-bonded network. However, clear experimental evidence of the Grotthuss mechanism has not been reported yet. Herein we show by infrared spectroscopy that a vehicle-type PT occurs in the penta- and hexahydrated clusters of protonated *p*-aminobenzoic acid, while Grotthuss-type PT is observed in heptahydrated clusters, indicating a change in the PT mechanism depending on the degree of hydration. These findings emphasize the importance of the usually ignored vehicle mechanism as well as the degree of hydration. It highlights the possibility of controlling the PT mechanism by the number of water molecules in chemical and biological environments.

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Introduction

Proton transfer (PT) is one of the most ubiquitous reactions prevailing in aqueous solutions,^{1,2} proton-conducting oxides/polymers,^{3,4} metal-organic frameworks (MOFs),^{5,6} and biological macromolecules.⁷⁻⁹ Much experimental and computational effort has been devoted to unravelling the nature of PT. An elemental reaction mechanism is the direct transport of a solvated proton, such as H₃O⁺ in water, which is called the vehicle mechanism.¹⁰ However, this mechanism cannot rationalize several important properties often observed for PT. For example, it predicts almost identical diffusion coefficients (*D*) for the hydrated proton (or H₃O⁺) and H₂O in aqueous solution because of their similar molecular size. However, *D*_{proton} is around five times higher than *D*_{H₂O} at room temperature.² The anomalously high *D*_{proton} has been ascribed to another

mechanism, named Grotthuss mechanism,¹¹ first proposed more than 200 years ago.^{12,13} It involves sequential proton shuttling to an adjacent proton acceptor site along a hydrogen-bonded (H-bonded) water network. Calculations^{14,15} using classical and quantum molecular dynamics together with chemical insight^{2,11} predict that the proton relay experiences a lower reaction barrier than the vehicle mechanism, suggesting that the high *D*_{proton} value shall indeed be ascribed to the Grotthuss mechanism. This mechanism is now considered to be the “golden” standard for PT in aqueous solutions and various other molecular systems mentioned above,¹⁻⁹ although there is surprisingly little experimental validation at the molecular level.

Experimental evidence for the Grotthuss mechanism has been obtained by the photo-induced PT dynamics of the rhodopsin and yellow proteins.^{7,16} However, it is limited only to photo-initiated PT processes in the excited electronic state. Significantly, it is not straightforward to transfer this conclusion to ground-state PT phenomena, like those in aqueous solutions, because of its stochastic character. Indeed, many studies on ground-state PT in the condensed phase suggest the Grotthuss mechanism simply because of the low barrier for ion conductivity without presenting direct evidence.¹⁷⁻¹⁹ One of the approaches to characterize the PT mechanism is the assignment of intermediates. In the theoretical framework of the Grotthuss mechanism in aqueous solution, a proton of an H₃O⁺ ion hydrated by three water molecules (Eigen ion, H₉O₄⁺)¹ is transferred to its nearest H₂O to form another Eigen structure

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via the transient Zundel ion (H_5O_2^+).^{20–22} Two-dimensional IR spectroscopy revealed that an asymmetric Zundel structure can be formed in aqueous solution.²¹ However, the presence of such intermediates is not always directly connected to the Grotthuss mechanism. The general difficulty in determining the PT mechanisms in various molecular systems prevents a deeper understanding of the mechanism–property correlations at the molecular level.

PT in hydrated clusters consisting of a finite number of water molecules^{23–28} and other solvent molecules^{29–31} provides a well-defined benchmark to explore the so far elusive PT mechanism. For example, *p*-aminobenzoic acid (PABA, Fig. 1) is a prototypical molecule for microhydration-induced PT.^{23,24,32–34} PABA has two protonation sites, *i.e.*, the O- and N-protomers resulting from protonation of the carboxy and amino groups, respectively (Fig. 1). Their relative stability varies with the surrounding environment. The O-protomer is more stable in the gas phase while the N-protomer is more stable in a polar solvent.^{35,36} Studies using ion mobility mass spectrometry³⁷ and infrared photodissociation (IRPD) spectroscopy²³ claim that for the hydrated clusters of protonated PABA, $\text{PABA}^+(\text{H}_2\text{O})_n$, generated by electrospray ionization (ESI), the N-protomer becomes detectable at $n = 6$, despite the spectral ambiguity in both the arrival time distributions and the IRPD spectra. In subsequent studies, we re-determined the threshold size (n) for the O \rightarrow N protomer switching as $n = 5$ using cryogenic double ion trap IR spectroscopy.²⁴ For the PT mechanism, none of previous studies (including ours) have provided any experimental evidence whether the vehicle or the Grotthuss mechanism takes place.^{23,24,37} The previous studies using ion mobility and IRPD claim the Grotthuss mechanism because their density functional theory (DFT) calculations show a stable structure for $n = 6$, in which the O- and N-protonation sites are bridged by a H-bonded water chain. However, no experimental evaluation for this structure was presented. We also applied DFT calculations for $n = 5$ and found that a structure locally hydrated

around the protonated amino group is significantly more stable than the ones in which the O- and N-sites are bridged by an H-bonded water chain, and thus the only observed hydration motif. Based on this exclusively observed hydration motif, we suggested the vehicle rather than the Grotthuss mechanism, although again we had no direct experimental evidence regarding which PT mechanism is in operation for $n \geq 5$.

To determine the PT mechanisms operating in hydrated PABA^+ at the molecular level, we combine herein cryogenic double ion trap IR spectroscopy^{38–42} with isotopic labelling. Deuterated hydrated clusters of PABA^+ can be produced by introducing D_2O vapour into the reaction trap. One can readily expect that, starting from the O-protomer, the Grotthuss-type PT gives a singly deuterated N-protomer (NH_2D_1) in the deuterated hydrated clusters because the proton source is D_2O (in Fig. 1). The vehicle-type PT yields both deuterated (NH_2D_1) and nondeuterated (NH_3) N-protomers via the transfer of HD_2O^+ (Fig. 1). Clearly, the detection of NH_3 provides direct evidence for the vehicle mechanism. However, the appearance of NH_3 can hardly be probed by conventional IRPD spectroscopy because its vibrational signatures are largely red-shifted and broadened by H-bonding with water molecules, causing spectral congestion.^{23,43}

This problem is overcome herein by applying the novel collision-assisted stripping (CAS) method, developed recently by our group.^{24,27,43} In CAS experiments, one measures IRPD spectra of the PABA^+ monomer core after removing all hydrated water molecules of the cluster by soft collisions in the ion trap, allowing for the clear spectroscopic assignment of deuterated isotopologues of PABA^+ , as the IR spectra of the solvent-free ions are not congested. Herein, we perform CAS-IRPD of $\text{PABA}^+(\text{D}_2\text{O})_n$ to determine the PT mechanisms as a function of the degree of hydration (see Experimental in the ESI[†]). We obtain clear evidence of the vehicle mechanism for $n = 5/6$ and suggestions of the Grotthuss mechanism for $n = 7$, respectively, suggesting a switch in the PT mechanism between

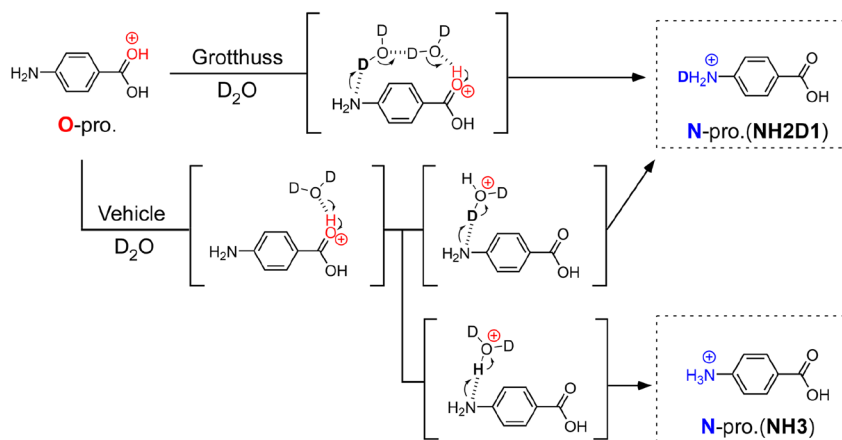


Fig. 1 Schematic illustration of hydration-induced O \rightarrow N proton transfer of PABA^+ by vehicle and Grotthuss mechanisms. The parentheses show a possible intermediate structure for the proton transfer. In the Grotthuss mechanism, proton transfer is mediated by a water bridge between CO and NH sites, resulting in a singly deuterated N-protomer denoted as NH_2D_1 . On the other hand, vehicle-type proton transfer affords both NH_2D_1 and nondeuterated NH_3 . Note that the reaction intermediate in the scheme is simplified for clarity. For example, we do not exclude the possibility of bridged structures with more than two water molecules for the Grotthuss mechanism.



$n = 5$ and $n = 7$. The consistency with the IRPD spectra of the hydrated clusters is also discussed.

Results and discussion

Fig. 2 shows the CAS-IRPD spectra of $\text{PABA}^+(\text{D}_2\text{O})_n$ with $n = 0$ and 4–7. All bands observed for $\text{PABA}^+(n = 0)$ are ascribed to vibrational modes of the O-protomer according to previous assignments: symmetric NH_2 stretch $\nu_{\text{NH}_2}^{\text{S}}(\text{O}) = 3435 \text{ cm}^{-1}$; outward-facing OH stretch $\nu_{\text{z-OH}}(\text{O}) = 3496 \text{ cm}^{-1}$; antisymmetric NH_2 stretch $\nu_{\text{NH}_2}^{\text{AS}}(\text{O}) = 3535 \text{ cm}^{-1}$; benzene-facing OH stretch $\nu_{\text{E-OH}}(\text{O}) = 3558 \text{ cm}^{-1}$.²⁴ The lack of any transitions in the 3200–3400 cm^{-1} range indicates the complete absence of the N-protomer of bare PABA^+ under the chosen experimental conditions.²⁴ The ESI ion beam of bare PABA^+ with pure O-protomer population was injected into the reaction trap and exposed to deuterated water vapor and He buffer gas. Under such conditions, the hydration-induced O \rightarrow N PT reaction can be directly probed by the generation of the N-protomer in the hydrated clusters.

The CAS-IRPD spectrum of $\text{PABA}^+(\text{D}_2\text{O})_5$ shows new bands in the 3220–3320 cm^{-1} range, which are absent for $n \leq 4$ (Fig. 2). The spectrum of these bands measured with a twice higher laser intensity is also shown as the inset. These bands are assigned to NH stretches of the N-protomer,^{24,44} indicating the appearance of the N-protomer core upon O \rightarrow N PT. A further new band at 2416 cm^{-1} is attributed to an ND stretch of the N-protomer based on its frequency (see Fig. S1 in the ESI† for detailed band assignments), confirming the appearance of the N-protomer core at $n = 5$. The threshold size ($n = 5$)²⁴ for O \rightarrow N protomer

switching does not change upon $\text{H}_2\text{O} \rightarrow \text{D}_2\text{O}$ substitution. Based on the given assignment, the other new band at 2620 cm^{-1} can be assigned to the OD stretching of the N-protomer core ($\nu_{\text{OD}}(\text{N})$, Fig. S1†). The remaining two bands at 2585 and 2631 cm^{-1} are then attributed to $\nu_{\text{z-OD}}(\text{O})$ and $\nu_{\text{E-OD}}(\text{O})$, respectively (Fig. S1†). Note that the OD stretches also appear for $n = 4$, indicating H/D exchange at the carboxylic group without protomer switching. One may wonder why H/D exchange happens at such low temperatures ($T = 80 \text{ K}$), at which protonated glycine is not deuterated by clustering with D_2O .⁴⁵ This observation is rationalized here by the high acidity of the protonated carboxy group. Recently, the Johnson group reported IRPD spectra of hydrated deuterated isotopologues of the N-protomer ($\text{ND}_3^+\text{C}_6\text{H}_4\text{COOD}(\text{H}_2\text{O})_n$) and found that the acidic COOD group is more susceptible to H/D exchange than the charged ND_3^+ group.⁴⁶ The protonated $\text{C}(\text{OH})_2^+$ group of the O-protomer may more readily replace H by D.

The CAS-IRPD spectra of $\text{PABA}^+(\text{D}_2\text{O})_{6,7}$ also exhibit the ND/NH stretches of the N-protomer core in the 2380–2470/3220–3320 cm^{-1} ranges, but their band patterns are more complicated than that of $n = 5$ (Fig. 3a). To understand the PT mechanism, one must assign the vibrational bands of deuterated (**NH2D1**) and nondeuterated (**NH3**) N-protomers without ambiguity. For this purpose, we measured the IR spectra of nondeuterated (m/z 138), monodeuterated (m/z 139, **NH3OD1** and **NH2D1**), and dideuterated (m/z 140, **NH2D1OD1** and **NH1D2**) forms of the bare N-protomer in the NH stretch range, which are shown in Fig. 3b along with molecular structures. Details of the assignments are summarized in the ESI.† Briefly, three prominent bands in the spectrum of m/z 138 are assigned

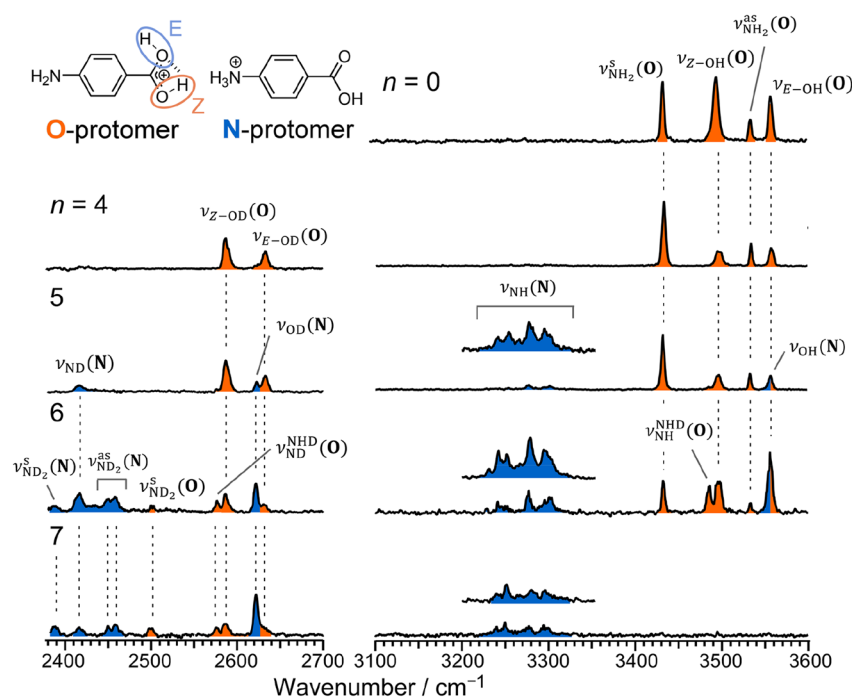


Fig. 2 IRPD spectrum of PABA^+ and CAS-IRPD spectra of $\text{PABA}^+(\text{D}_2\text{O})_n$ ($n = 4-7$). The orange- and blue-colored bands are assigned to the vibrational modes of O- and N-protomers, respectively. The inset CAS-IRPD spectra in 3200–3360 cm^{-1} for $n = 5-7$ were re-measured with ~two times higher IR-laser fluence. The indices N and O in the notation of vibrational modes indicate N- and O-protomers, respectively.



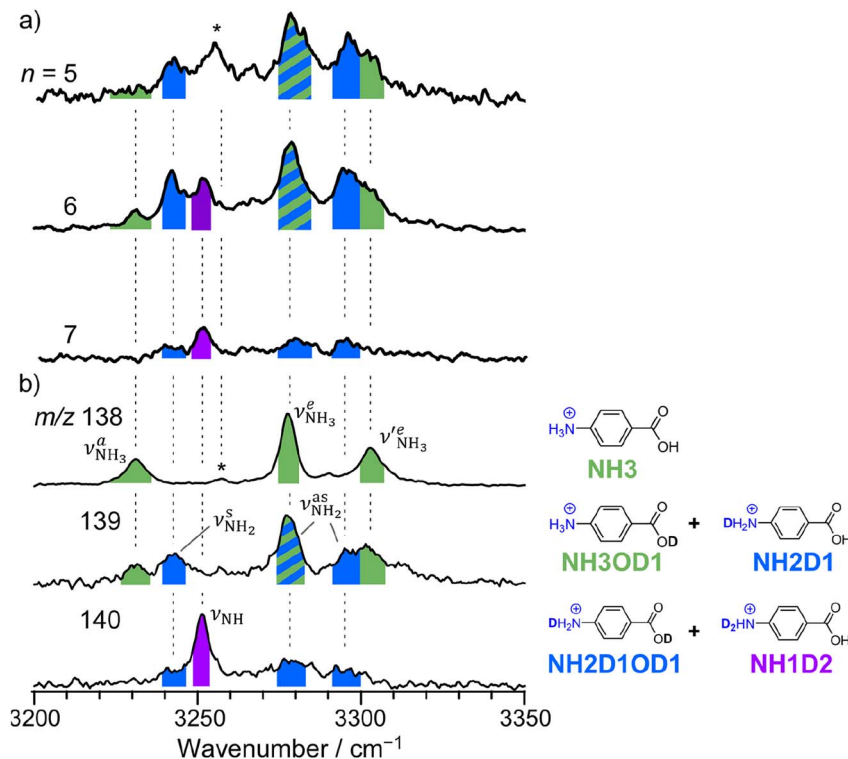


Fig. 3 (a) CAS-IRPD spectra of PABAH⁺(D₂O)_n ($n = 5-7$) in the 3200–3350 cm⁻¹ range (reprinted from the inset spectra in Fig. 2). (b) IRPD spectra of PABAH⁺ (m/z 138), singly deuterated protonated PABA (m/z 139), and doubly deuterated protonated PABA (m/z 140). These spectra are necessary to assign the CAS-IRPD spectra because the monomer core after the CAS event includes several deuterated isotopomers. The green-, blue-, and purple-colored bands are assigned to NH stretches of NH₃, NH₂D1, and NH₁D₂, respectively. The band marked by an asterisk is assigned to a combination band of the O-protomer (in-plane NH₂ bend^{24,44} at ~1660 cm⁻¹ + aryl C–C stretch^{24,44} at ~1600 cm⁻¹). This type of combination band is also observed for an analog of PABA, ethyl *p*-aminobenzoate.⁴³

to NH₃ stretching bands (coloured in green, ν^a_{NH₃}(N), ν^e_{NH₃}(N), and ν^e_{NH₃}(N)) of the N-protomer of the fully hydrogenated species (NH₃). The new bands indicated by blue colour in the spectrum of m/z 139 are NH₂ stretching bands (ν^s_{NH₂}(N) and ν^{as}_{NH₂}(N)) of the amino-deuterated N-protomer (NH₂D1). The bands due to the NH₃ stretching (green) are overlapped with transitions of the coexisting N-protomer deuterated at the carboxylate group (NH₃OD1). The newly observed band (purple) in the spectrum of m/z 140 is an NH stretch mode of the dideuterated amino group of the N-protomer (NH₁D₂). Among these bands, the vibrational transitions of NH₃ (ν^a_{NH₃}(N) and ν^e_{NH₃}(N)) are the most important ones because they indicate the presence of the N-protomer (NH₃) which can be generated only by the vehicle mechanism (Fig. 1).

Let us check the presence of ν^a_{NH₃}(N) and ν^e_{NH₃}(N) (green), the marker bands of the vehicle mechanism, in the CAS-IRPD spectra of PABAH⁺(D₂O)_n with $n = 5-7$. Fig. 3a reproduces expanded spectra in the 3200–3350 cm⁻¹ range. The green marker bands obviously appear in the spectra of $n = 5$ and 6, but are clearly absent in the $n = 7$ spectrum. This result directly indicates that the vehicle mechanism operates in the $n = 5$ and 6 clusters but not for $n = 7$. One cannot directly conclude from the CAS-IRPD spectra whether the Grothuss mechanism operates in $n = 5$ and 6. The observed bands in the $n = 7$ spectrum

correspond to N-protomers of NH₂D1 (blue) and NH₁D₂ (purple). The former, NH₂D1, can be generated by both the vehicle and Grothuss mechanisms. As the absence of the vehicle mechanism for $n = 7$ is clear, the blue bands can only arise from the Grothuss PT. The latter, NH₁D₂, cannot be generated by a single O → N PT reaction *via* either the Grothuss or the vehicle mechanism. The observed multiple deuteration thus is ascribed to H/D exchange by D₂O or the interconversion between N- and O-protomers by O → N and N → O back reactions (Fig. S5 in the ESI[†]). Here, we cannot fully exclude the possibility that the absence of ν^a_{NH₃}(N), the signature of the vehicle mechanism, may arise from the H/D exchange reaction because such an H/D exchange is observed for $n \geq 4$ (see above). However, if this scenario were the case, H/D exchange also modulates the intensities of ν^a_{NH₃}(N) in the spectra of $n = 5$ and 6 where the band is clearly observed. This observation means that the relative intensity of ν^a_{NH₃}(N) for $n = 7$ is rather weak in comparison to those for $n = 5$ and 6, and the vehicle PT mechanism is negligibly minor for $n = 7$.

The size-dependent PT mechanisms proposed by the experiment are consistent with the molecular structures of microhydrated PABAH⁺. The bridged structures in which the NH₃⁺ and COOH groups are connected by an H-bonded water chain are more abundant than the unbridged structures for the N-protomer at $n = 7$, whereas the bridged structures become



minor for $n = 6$ and are not observed for $n = 5$ (Fig. S6 in the ESI†).²⁴ This strongly suggests that the water-bridged intermediates for the Grotthuss-type PT are stabilized at $n = 7$ (and partly stabilized at $n = 6$). In this sense, the Grotthuss mechanism may be involved in part in the PT at $n = 6$. These observations show that the reaction barriers of the Grotthuss mechanism can be altered by the number of water molecules. The Grotthuss mechanism with a lowered reaction barrier would account for the minor contribution of the vehicle mechanism at $n = 7$. This finding gives suggestions for the poorly understood properties of PT in various systems. For example, it is known that proton conductivity in porous materials such as MOFs is sensitive to humidity.^{5,6} Some MOFs show a drastic decrease in the conductivity below a critical humidity,^{47,48} however, the reason remains unclear. The poor conductivity at low humidity has been hampering the application of MOFs to proton-conducting membranes in fuel cells. Our conclusion that the PT mechanisms would be changed from Grotthuss to vehicle with decreasing number of water molecules would rationalize such a drastic change in PT by humidity.

Conclusion

In summary, we spectroscopically determined the mechanisms for hydration-induced PT of PABAH⁺ by cryogenic double ion trap IR spectroscopy combined with deuterium labelling. The presence of NH₃ for PABAH⁺(D₂O)_{5,6} unambiguously indicates O → N PT *via* the vehicle mechanism, in sharp contrast to the computational assumption that the Grotthuss mechanism is dominant in this molecular system^{49,50} with inference from experimental results. The vehicle mechanism for solvent-mediated PT is poorly experimentally validated for water,⁵¹ including methanol²⁹ and ammonia.^{30,31} We have spectroscopically validated the vehicle mechanism for water-containing molecular systems at the molecular level. This study experimentally demonstrates the significance of the vehicle mechanism in water as well, which has often been ignored in previous work. On the other hand, the absence of NH₃ and the presence of NH₂D₁ for PABAH⁺(D₂O)₇ strongly suggest the Grotthuss mechanism, which is correlated with the stability of the water-bridged structures at this cluster size. The calculated bridged structure has three/four water molecules in the chain connecting the O and N sites. The remaining water molecules attach to this chain and the amino NH₃⁺ group. This result highlights the importance of the second-shell water molecules in stabilizing the water bridge. Although the system we studied is not the same as solution in terms of the number of water molecules and temperature, the findings afford a clue to understanding the nature of PT in molecular systems consisting of a finite number of water molecules. In particular, PT in functional materials such as proton-conducting polymers⁴ and MOFs^{6,47,48} and biological macromolecules⁹ could be facilitated by residual water in their nanosized pores. The number of water molecules in the pores is pivotal to stabilize the water chain and control the PT mechanism.

Data availability

Essential data are fully provided in the main text and ESI.† Further data in this study are available from the corresponding authors upon a reasonable request.

Author contributions

K. H., O. D., S. I., and M. F. conceived and designed the project. K. A. and K. H. carried out the experiments and analyzed the data. All the authors contributed to writing the manuscript and approved the final version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

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References

- M. Eigen, *Angew. Chem., Int. Ed.*, 1964, **3**, 1–19.
- K.-D. Kreuer, *Chem. Mater.*, 1996, **8**, 610–641.
- K.-D. Kreuer, *Annu. Rev. Mater. Res.*, 2003, **33**, 333–359.
- K.-D. Kreuer, *J. Membr. Sci.*, 2001, **185**, 29–39.
- P. Ramaswamy, N. E. Wong and G. K. H. Shimizu, *Chem. Soc. Rev.*, 2014, **43**, 5913–5932.
- D.-W. Lim, M. Sadakiyo and H. Kitagawa, *Chem. Sci.*, 2019, **10**, 16–33.
- F. Garczarek and K. Gerwert, *Nature*, 2006, **439**, 109–112.
- J. R. Schnell and J. J. Chou, *Nature*, 2008, **451**, 591–595.
- O. P. Ernst, D. T. Lodowski, M. Elstner, P. Hegemann, L. S. Brown and H. Kandori, *Chem. Rev.*, 2014, **114**, 126–163.
- K.-D. Kreuer, A. Rabenau and W. Weppner, *Angew. Chem., Int. Ed.*, 1982, **21**, 208–209.
- N. Agmon, *Chem. Phys. Lett.*, 1995, **244**, 456–462.
- C. J. T. d. Grotthuss, *Ann. Chim.*, 1806, **LVIII**, 54–74.
- D. Marx, *ChemPhysChem*, 2006, **7**, 1848–1870.
- D. Marx, M. E. Tuckerman, J. Hutter and M. Parrinello, *Nature*, 1999, **397**, 601–604.
- M. Tuckerman, K. Laasonen, M. Sprik and M. Parrinello, *J. Chem. Phys.*, 1995, **103**, 150–161.



- 16 H. Kuramochi, S. Takeuchi, K. Yonezawa, H. Kamikubo, M. Kataoka and T. Tahara, *Nat. Chem.*, 2017, **9**, 660–666.
- 17 I. Popov, Z. Zhu, A. R. Young-Gonzales, R. L. Sacchi, E. Mamontov, C. Gainaru, S. J. Paddison and A. P. Sokolov, *Commun. Chem.*, 2023, **6**, 77.
- 18 X. Wu, J. J. Hong, W. Shin, L. Ma, T. Liu, X. Bi, Y. Yuan, Y. Qi, T. W. Surta, W. Huang, *et al.*, *Nat. Energy*, 2019, **4**, 123–130.
- 19 Y. Yang, P. Zhang, L. Hao, P. Cheng, Y. Chen and Z. Zhang, *Angew. Chem., Int. Ed.*, 2021, **60**, 21838–21845.
- 20 R. Janoschek, E. G. Weidemann, H. Pfeiffer and G. Zundel, *J. Am. Chem. Soc.*, 1972, **94**, 2387–2396.
- 21 J. A. Fournier, W. B. Carpenter, N. H. C. Lewis and A. Tokmakoff, *Nat. Chem.*, 2018, **10**, 932–937.
- 22 C. T. Wolke, J. A. Fournier, L. C. Dzugan, M. R. Fagiani, T. T. Odbadrakh, H. Knorke, K. D. Jordan, A. B. McCoy, K. R. Asmis and M. A. Johnson, *Science*, 2016, **354**, 1131–1135.
- 23 T. M. Chang, J. S. Prell, E. R. Warrick and E. R. Williams, *J. Am. Chem. Soc.*, 2012, **134**, 15805–15813.
- 24 K. Akasaka, K. Hirata, F. Haddad, O. Dopfer, S. Ishiuchi and M. Fujii, *Phys. Chem. Chem. Phys.*, 2023, **25**, 4481–4488.
- 25 K. Chatterjee and O. Dopfer, *J. Phys. Chem. A*, 2020, **124**, 1134–1151.
- 26 N. Takeda, K. Hirata, K. Tsuruta, G. D. Santis, S. S. Xantheas, S. Ishiuchi and M. Fujii, *Phys. Chem. Chem. Phys.*, 2022, **24**, 5786–5793.
- 27 G. D. Santis, N. Takeda, K. Hirata, K. Tsuruta, S. Ishiuchi, S. S. Xantheas and M. Fujii, *J. Am. Chem. Soc.*, 2022, **144**, 16698–16702.
- 28 M.-P. Gaigeot, A. Cimas, M. Seydou, J.-Y. Kim, S. Lee and J.-P. Schermann, *J. Am. Chem. Soc.*, 2010, **132**, 18067–18077.
- 29 B. Ucur, O. J. Shiels, S. J. Blanksby and A. J. Trevitt, *J. Am. Soc. Mass Spectrom.*, 2023, **34**, 1428–1435.
- 30 K. Ohshimo, S. Miyazaki, K. Hattori and F. Misaizu, *Phys. Chem. Chem. Phys.*, 2020, **22**, 8164–8170.
- 31 K. Ohshimo, R. Sato, Y. Takasaki, K. Tsunoda, R. Ito, M. Kanno and F. Misaizu, *J. Phys. Chem. Lett.*, 2023, 8281–8288.
- 32 Z. Tian and S. R. Kass, *Angew. Chem., Int. Ed.*, 2009, **48**, 1321–1323.
- 33 J. L. Campbell, J. C. Y. Le Blanc and B. B. Schneider, *Anal. Chem.*, 2012, **84**, 7857–7864.
- 34 P. R. Batista, T. C. Penna, L. C. Ducati and T. C. Correra, *Phys. Chem. Chem. Phys.*, 2021, **23**, 19659–19672.
- 35 J. Seo, S. Warnke, S. Gewinner, W. Schöllkopf, M. T. Bowers, K. Pagel and G. von Helden, *Phys. Chem. Chem. Phys.*, 2016, **18**, 25474–25482.
- 36 S. Warnke, J. Seo, J. Boschmans, F. Sobott, J. H. Scrivens, C. Bleiholder, M. T. Bowers, S. Gewinner, W. Schöllkopf, K. Pagel, *et al.*, *J. Am. Chem. Soc.*, 2015, **137**, 4236–4242.
- 37 M. J. Hebert and D. H. Russell, *J. Phys. Chem. B*, 2020, **124**, 2081–2087.
- 38 S. Ishiuchi, H. Wako, D. Kato and M. Fujii, *J. Mol. Spectrosc.*, 2017, **332**, 45–51.
- 39 B. M. Marsh, J. M. Voss and E. Garand, *J. Chem. Phys.*, 2015, **143**, 204201.
- 40 E. Sato, K. Hirata, J. M. Lisy, S. Ishiuchi and M. Fujii, *J. Phys. Chem. Lett.*, 2021, **12**, 1754–1758.
- 41 O. V. Boyarkin, S. R. Mercier, A. Kamariotis and T. R. Rizzo, *J. Am. Chem. Soc.*, 2006, **128**, 2816–2817.
- 42 J. G. Redwine, Z. A. Davis, N. L. Burke, R. A. Oglesbee, S. A. McLuckey and T. S. Zwier, *Int. J. Mass Spectrom.*, 2013, **348**, 9–14.
- 43 K. Hirata, F. Haddad, O. Dopfer, S. Ishiuchi and M. Fujii, *Phys. Chem. Chem. Phys.*, 2022, **24**, 5774–5779.
- 44 T. Khuu, N. Yang and M. A. Johnson, *Int. J. Mass Spectrom.*, 2020, **457**, 116427.
- 45 J. M. Voss, K. C. Fischer and E. Garand, *J. Phys. Chem. Lett.*, 2018, **9**, 2246–2250.
- 46 T. Khuu, S. J. Stropoli, K. Greis, N. Yang and M. A. Johnson, *J. Chem. Phys.*, 2022, **157**, 131102.
- 47 H. Okawa, A. Shigematsu, M. Sadakiyo, T. Miyagawa, K. Yoneda, M. Ohba and H. Kitagawa, *J. Am. Chem. Soc.*, 2009, **131**, 13516–13522.
- 48 M. Sadakiyo, H. Okawa, A. Shigematsu, M. Ohba, T. Yamada and H. Kitagawa, *J. Am. Chem. Soc.*, 2012, **134**, 5472–5475.
- 49 H. Xia and A. B. Attygalle, *J. Mass Spectrom.*, 2018, **53**, 353–360.
- 50 J. L. Campbell, A. M.-C. Yang, L. R. Melo and W. S. Hopkins, *J. Am. Soc. Mass Spectrom.*, 2016, **27**, 1277–1284.
- 51 Y. Matsuda, A. Yamada, K. Hanaue, N. Mikami and A. Fujii, *Angew. Chem., Int. Ed.*, 2010, **49**, 4898–4901.

