


 Cite this: *RSC Adv.*, 2020, 10, 39808

ACE2 as a potential therapeutic target for pandemic COVID-19

 Bhaswati Chatterjee ^{*a} and Suman S. Thakur ^{*b}

SARS-CoV-2 virus invades the host through angiotensin-converting enzyme 2 (ACE2) receptors by decreasing the ACE2 expression of the host. This disturbs the dynamic equilibrium between the ACE/Ang II/AT1R axis and ACE2/Ang (1–7)/Mas receptor axis. Therefore, the clinically approved drugs belonging to (i) angiotensin converting enzyme (ACE) inhibitors such as captopril, and enalaprilat, (ii) angiotensin-receptor blockers (ARBs) such as losartan, candesartan, olmesartan, azilsartan, irbesartan, and telmisartan and (iii) the combination of ACE inhibitors and ARBs such as losartan with lisinopril and captopril with losartan, and (iv) recombinant ACE2, were studied for their ability to activate ACE2 in different medical conditions including hypertension, inflammation, cardiovascular, renal and lung diseases. These clinically approved drugs were found to activate ACE2 that had been downregulated in different medical conditions including hypertension, inflammation, cardiovascular, renal and lung diseases. Therefore, these drugs may be repurposed to re-activate the downregulated ACE2 of COVID-19 patients. These drugs either alone or in combination may be repurposed as prophylactics and therapeutics against SARS-CoV-2 virus.

Received 25th September 2020

Accepted 26th October 2020

DOI: 10.1039/d0ra08228g

rsc.li/rsc-advances

Introduction

Coronavirus disease (COVID-19) is a life-threatening infectious viral disease that emerged in Wuhan, China. It is caused by SARS-CoV-2 infection¹ which has infected more than 41 104 946 humans worldwide and has caused more than 1 128 325 deaths as reported by the World Health Organization (WHO). SARS-CoV-2 infects the respiratory tract and causes symptoms associated with cardiovascular, digestive and neurological diseases.^{2,3} Elderly people with cardiovascular conditions including hypertension are more prone to develop COVID-19.⁴ COVID-19 patients have higher levels of angiotensin II as compared to healthy individuals.⁵ It has been found that this disease infects humans and is transmitted from human to human and also from humans to animals.^{6,7} This disease is pandemic causing a more serious global health threat than severe acute respiratory syndrome (SARS) and the Middle East Respiratory Syndrome (MERS) epidemics, therefore there is an urgent requirement for therapeutic and prophylactic interventions for COVID-19.

The pathogen, SARS-CoV-2 is a single stranded RNA virus belonging to β -coronavirus genus with lineage B.⁸ Further, S1 subunits of spike (S) proteins of SARS-CoV-2 and SARS-CoV have

affinity towards human angiotensin-converting enzyme 2 (hACE2) receptors.⁸ ACE2 is involved in conversion of angiotensin II (Ang II) to angiotensin-(1–7) [Ang (1–7)] and angiotensin I (Ang I) to angiotensin-(1–9) [Ang (1–9)].⁹ Angiotensin-converting enzyme 2 (ACE2) is homolog of the angiotensin converting enzyme (ACE) that is expressed in cell membranes of heart, kidney, lungs, arteries and intestines.¹⁰ Interestingly, SARS-CoV-2 binds to ACE2 receptor, 10–20 folds higher compared to SARS-CoV resulting in higher infection and transmissibility.¹¹

Interestingly, ACE2 protects lungs from severe acute lung injury (Imai *et al.*, 2005)¹⁷ and kidneys from damage.^{12,13} The deficiency of ACE2 is related with advanced glomerulosclerosis, oxidative stress in kidneys and diabetic nephropathy.^{14–16} ACE2 is a member of renin–angiotensin–aldosterone system (RAAS) that help in regulating hypertension, electrolyte balance and contributes to pathophysiology of cardiovascular/renal/lung diseases. ACE2 expression is decreased during SARS-CoV infections resulting in disturbance of dynamic equilibrium state between ACE/Ang II/AT1R axis and ACE2/Ang (1–7)/Mas receptor axis¹⁷ and similarly this may happen during SARS-CoV-2 infection.

Interestingly, upregulation of ACE2 has also been found to be species specific upon interferon stimulation. Notably, interferon upregulate the human ACE2 in nasal epithelia and lung tissue while it is not able to upregulate the mouse ACE2 thereby helping in generating more targets for SARS-COV-2 and exploring more areas for invasion of virus.¹⁸

^aNational Institute of Pharmaceutical Education and Research (NIPER), Dept. of Pharmaceuticals, Ministry of Chemicals and Fertilizers, Govt. of India, Balanagar, Hyderabad-500 037, Telangana, India. E-mail: bhaswaticatterjee@yahoo.com; bhaswati@niperhyd.ac.in

^bCentre for Cellular and Molecular Biology, Uppal Road, Hyderabad-500007, India. E-mail: sst@cmb.res.in; Tel: +91-40-27192865, +91-40-27192505



There is mixed findings on ACE2 levels by ACE inhibitors and angiotensin-receptor blockers (ARBs)^{19–34} Notably, the effect of angiotensin converting enzyme (ACE) inhibitors and angiotensin-receptor blockers (ARBs) on ACE2 is expected to be different as they have dissimilar effects on angiotensin II.³⁵

As SARS-CoV-2 sneaks into humans through hACE2 receptors and then downregulates its expression such that ACE2 forgoes its protective effect in different organs, therefore activating ACE2 may lead to the treatment for COVID-19 (Fig. 1). Till date, there is no effective therapeutic drug to cure COVID-19. Considering the complexity of the pathogenesis of COVID-19, we are highlighting the studies on clinically approved drugs which are activating ACE2 and may act as potential COVID-19 therapeutics (Fig. 1). The drugs activating ACE2 belongs to (i) angiotensin converting enzyme (ACE) inhibitors (Fig. 2), (ii) angiotensin-receptor blockers (ARBs) (Fig. 2), (iii) the combinations of ACE inhibitor and ARB, (iv) recombinant ACE2.

(i) Angiotensin converting enzyme (ACE) inhibitors

Since the active sites of ACE and ACE2 are different, therefore ACE inhibitors does not directly affect ACE2 activity³⁶ but can cause activation of ACE2. Captopril, an ACE inhibitor activates ACE2/angiotensin-(1–7) level/Mas receptor axis,³⁷ lowering mean pulmonary arterial pressure (PAP) and right ventricular index.³⁸ Furthermore, pre-treatment with captopril prevents lipopolysaccharide-induced injury in pulmonary microvascular endothelial cells of rat by bringing back the ACE/ACE2 ratio to

the normal levels and lowers the ratio of angiotensin II to angiotensin-(1–7).³⁷ Captopril prevents lung apoptosis by maintaining the normal level of ACE2/ACE axis during acute pulmonary embolism demonstrated in porcine model.³⁹

Treatment with enalapril at in neurogenic pulmonary edema expressed in rabbit models reduces the injury of lungs by reducing angiotensin II concentration and ratio of ACE mRNA to ACE2 mRNA in lungs.⁴⁰ Further, enalapril has the ability to inhibit the inflammation in airway caused by acrolein.⁴¹

As there is a well-established association between obesity, hypertension and diabetes, enalapril treatment gives protection to pancreatic islets and controls body mass, glucose intolerance and insulin resistance with the increase of ACE2/Ang (1–7)/Mas receptor axis and level of adiponectin.⁴²

(ii) Angiotensin-receptor blockers (ARBs)

The treatment with ARBs causes an upregulation of ACE2 as seen from animal studies^{20,25,43} The angiotensin II type 1 receptor (AT1R) is the main receptor mediating angiotensin II effects. Losartan is angiotensin II type 1 receptor antagonists that demonstrate lung protectiveness. Its treatment in pulmonary arterial hypertension decreases the level of angiotensin II, ACE expression and increases ACE2 expression in lung tissue.⁴⁴ Interestingly, losartan prevents lipopolysaccharide induced lung injury by lowering cytokines and high mobility group box 1 (HMGB1) protein levels and controls ACE2 activity in rat model system.⁴⁵ In smoke-induced pulmonary arterial remodeling and



Fig. 1 Schematic mechanism for ACE inhibitor, ARBs and recombinant ACE2 as repurposed drugs for COVID-19.





Fig. 2 Chemical structures of the angiotensin converting enzyme (ACE) inhibitors and angiotensin-receptor blockers (ARBs) as repurposed drugs for COVID-19.

pulmonary arterial hypertension model system, losartan plays by lowering the angiotensin II level in lungs and increasing ACE2 levels.⁴⁶

In spontaneously hypertensive rats, losartan administration resulted in high level of renal ACE2 and angiotensin-(1-7) generation suggesting the renal protection.⁴⁷ Candesartan is renoprotective in intermediate doses in diabetic mice by upregulating ACE2/AT2R/Mas axis protective components.⁴⁸ In addition, olmesartan treatment in hypertensive patients resulted in increase of ACE2 expression in urine thus providing renoprotective effects.³⁴ Iwanami⁴⁹ *et al.* studied the impact of treatment with azilsartan and olmesartan which are angiotensin II type 1 (AT1) receptor blockers in transgenic mice with the human renin and angiotensinogen genes (hRN/hANG-Tg) and found that azilsartan decreases blood pressure and increases sodium concentration in urine more than that of olmesartan with activation of ACE2/Ang-(1-7)/Mas axis.

Losartan or telmisartan is also used for treatment of non-alcoholic fatty liver disease (NAFLD) in obese and/or hypertensive model system by lowering hepatic triacylglycerol levels, PLIN 2 and improve glycemic control by using ACE2/rMAS axis.⁵⁰ Additionally, ARBs have anti-inflammatory properties as seen from irbesartan which decreases the intestinal inflammation induced by stress *via* activation of ACE2 pathway.⁵¹

Increase in ACE2 expression in heart is observed in rat after administration of losartan and olmesartan.²⁹ In the patients having essential hypertension, olmesartan usage for long

period decreases left ventricular mass index.⁵² Interestingly, cardiac hypertrophy not dependent on blood pressure is inhibited by olmesartan through the original AT1R blockade and also with the increase of ACE2/Ang (1-7)/Mas axis with decrease of Nox4 expression.⁵³ Losartan or captopril combined with angiotensin-(1-7) resulted in cardioprotection for diabetes-induced cardiac dysfunction by ACE2/Ang-(1-7)/Mas receptor axis.⁵⁴ The treatment with candesartan controlled the progression of dilated cardiomyopathy with the upregulation of Ang (1-7), ACE2 and mas1 along with the modulation of endoplasmic reticulum stress and cardiac apoptosis.⁵⁵ Furthermore, telmisartan treatment provided cardioprotective effects in dilated cardiomyopathy of mice model through stimulation of ACE2/ANG 1-7/Mas receptor.⁵⁶ Similarly, olmesartan treatment also ameliorates dilated cardiomyopathy through modulating ACE2/Ang (1-7)/Mas receptor axis.⁵⁷

(iii) Combinations of angiotensin converting enzyme (ACE) inhibitors and angiotensin-receptor blockers (ARBs)

Interesting, lungs are protected from pneumonitis and fibrosis induced by radiation with the help of ACE inhibitors and ARBs.⁵⁸ Further, increase of cardio protective biomarkers is associated with combined use of ACE inhibitor, ARBs and beta blocker.⁵⁹ Notably, increase of cardiac ACE2 activity is associated with combined effect of losartan and lisinopril.²⁰ The inhibition of ACE may result from the total effect of lower angiotensin II formation and metabolism of angiotensin-(1-7)



whereas angiotensin II type 1 receptor antagonists may work by raising angiotensin II metabolism by ACE2 in cardiac system.²⁰

Furthermore, under hypoxia conditions, when mouse Lewis lung carcinoma cells are treated with captopril and losartan, an increase in ACE2 and angiotensin II type 2 receptors expression and a decrease in ACE and angiotensin II type 1 receptor expression is observed.⁶⁰

(iv) Recombinant ACE2

Recombinant human ACE2 is known as therapeutic RAS modulator. Supplementation with recombinant human ACE2 results in lowering of Ang II and increase of Ang-(1-7)/Ang II ratio.⁶¹ It has been found to reverse lung injury and lower angiotensin II levels in viral infections.⁶²⁻⁶⁴ Additionally, it has been found to play cardioprotective role in heart failure⁶⁵ and ameliorate kidney fibrosis.⁶⁶ Further, the diabetic kidney injury is reduced when treated with recombinant human ACE2.⁶⁷

GSK2586881, a recombinant human ACE2 was found to decrease the level of Ang II with increase of Ang (1-7) in acute respiratory distress syndrome patients.⁶⁴ Notably, in early stages of SARS-CoV-2 infection, human recombinant soluble ACE2 was found to be active against SARS-CoV-2 virus and blocks this viral infection as found from *in vitro* studies.^{68,69}

Conclusions and outlook

On the basis of the available evidences, these drugs have the ability to activate the ACE2 expression that had been down-regulated in different diseases including hypertension, cardiovascular, renal and lung diseases. As SARS-CoV-2 binds to host ACE2 receptor and downregulates its expression, therefore these drugs either alone or in combination may be repurposed to re-activate the host ACE2 thereby serving as probable SARS-CoV-2 therapeutics. Notably, these drugs may enhance the immune responses; reduce inflammations and organ damage due to COVID-19 complications. Further, these drugs may also reduce the death rate associated with COVID-19. Therefore, repurposing these clinically approved drugs might be a good option. The advantage of repurposing clinically approved drugs are their side effects is known and they might serve as the good candidates for SARS-CoV-2 therapeutics and prophylactics.

Author contributions

BC and SST have conceived the idea and wrote the manuscript.

Funding

There is no funding for this work.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

BC has received the fellowship from Department of Science and Technology (DST) in area of Women Scientist Scheme A (WOS-A).

References

- 1 A. E. Gorbalenya, S. C. Baker, R. S. Baric, R. J. de Groot, C. Drosten, A. A. Gulyaeva, B. L. Haagmans, C. Lauber, A. M. Leontovich, B. W. Neuman, D. Penzar, S. Perlman, L. L. M. Poon, D. V. Samborskiy, I. A. Sidorov, I. Sola and J. Ziebuhr, *Nat. Microbiol.*, 2020, 5, 536–544.
- 2 C. Huang, Y. Wang, X. Li, L. Ren, J. Zhao, Y. Hu, L. Zhang, G. Fan, J. Xu, X. Gu, Z. Cheng, T. Yu, J. Xia, Y. Wei, W. Wu, X. Xie, W. Yin, H. Li, M. Liu, Y. Xiao, H. Gao, L. Guo, J. Xie, G. Wang, R. Jiang, Z. Gao, Q. Jin, J. Wang and B. Cao, *Lancet*, 2020, 395, 497–506.
- 3 N. Chen, M. Zhou, X. Dong, J. Qu, F. Gong, Y. Han, Y. Qiu, J. Wang, Y. Liu, Y. Wei, J. Xia, T. Yu, X. Zhang and L. Zhang, *Lancet*, 2020, 395, 507–513.
- 4 Z. Wu and J. M. McGoogan, *JAMA, J. Am. Med. Assoc.*, 2020, 323, 1239–1242.
- 5 Y. Liu, Y. Yang, C. Zhang, F. Huang, F. Wang, J. Yuan, Z. Wang, J. Li, J. Li, C. Feng, Z. Zhang, L. Wang, L. Peng, L. Chen, Y. Qin, D. Zhao, S. Tan, L. Yin, J. Xu, C. Zhou, C. Jiang and L. Liu, *Sci. China: Life Sci.*, 2020, 63, 364–374.
- 6 P. J. Halfmann, M. Hatta, S. Chiba, T. Maemura, S. Fan, M. Takeda, N. Kinoshita, S.-I. Hattori, Y. S. Tagawa, K. I. Horimoto, M. Imai and Y. Kawaoka, *N. Engl. J. Med.*, 2020, 383, 592–594.
- 7 T. H. C. Sit, C. J. Brackman, S. Ming Ip, K. W. S. Tam, P. Y. T. Law, E. M. W. To, V. Y. T. Yu, L. D. Sims, D. N. C. Tsang, D. K. W. Chu, R. A. P. M. Perera, L. L. M. Poon and M. Peiris, *Nature*, 2020, DOI: 10.1038/s41586-020-2334-5.
- 8 P. Zhou, X.-L. Yang, X.-G. Wang, B. Hu, L. Zhang, W. Zhang, H.-R. Si, Y. Zhu, B. Li, C.-L. Huang, H.-D. Chen, J. Chen, Y. Luo, H. Guo, R.-D. Jiang, M.-Q. Liu, Y. Chen, X.-R. Shen, X. Wang, X.-S. Zheng, K. Zhao, Q.-J. Chen, F. Deng, L.-L. Liu, B. Yan, F.-X. Zhan, Y.-Y. Wang, G.-F. Xiao and Z.-L. Shi, *Nature*, 2020, 579, 270–273.
- 9 C. Vickers, P. Hales, V. Kaushik, L. Dick, J. Gavin, J. Tang, K. Godbout, T. Parsons, E. Baronas, F. Hsieh, S. Acton, M. Patane, A. Nichols and P. Tummino, *J. Biol. Chem.*, 2002, 277, 14838–14843.
- 10 I. Hamming, W. Timens, M. L. C. Bulthuis, A. T. Lely, G. J. Navis and H. van Goor, *J. Pathol.*, 2004, 203, 631–637.
- 11 D. Wrapp, N. Wang, K. S. Corbett, J. A. Goldsmith, C.-L. Hsieh, O. Abiona, B. S. Graham and J. S. McLellan, *Science*, 2020, 367, 1260–1263.
- 12 M. Ye, J. Wysocki, P. Naaz, M. R. Salabat, M. S. LaPointe and D. Battle, *Hypertension*, 2004, 43, 1120–1125.
- 13 G. Wang, F. M.-M. Lai, K.-B. Lai, K.-M. Chow, C.-H. B. Kwan, K.-T. P. Li and C.-C. Szeto, *Diabetologia*, 2008, 51, 1062–1067.



- 14 G. Y. Oudit, A. M. Herzenberg, Z. Kassiri, D. Wong, H. Reich, R. Khokha, M. A. Crackower, P. H. Backx, J. M. Penninger and J. W. Scholey, *Am. J. Pathol.*, 2006, **168**, 1808–1820.
- 15 D. W. Wong, G. Y. Oudit, H. Reich, Z. Kassiri, J. Zhou, Q. C. Liu, P. H. Backx, J. M. Penninger, A. M. Herzenberg and J. W. Scholey, *Am. J. Pathol.*, 2007, **171**, 438–451.
- 16 J. Wysocki, D. I. Ortiz-Melo, N. K. Mattocks, K. Xu, J. Prescott, K. Evora, M. Ye, M. A. Sparks, S. K. Haque, D. Battle and S. B. Gurley, *Physiol. Rep.*, 2014, **2**, e00264.
- 17 K. Kuba, Y. Imai, S. Rao, H. Gao, F. Guo, B. Guan, Y. Huan, P. Yang, Y. Zhang, W. Deng, L. Bao, B. Zhang, G. Liu, Z. Wang, M. Chappell, Y. Liu, D. Zheng, A. Leibbrandt, T. Wada, A. S. Slutsky, D. Liu, C. Qin, C. Jiang and J. M. Penninger, *Nat. Med.*, 2005, **11**, 875–879.
- 18 C. G. K. Ziegler, S. J. Allon, S. K. Nyquist, I. M. Mbanjo, V. N. Miao, C. N. Tzouanas, Y. Cao, A. S. Yousif, J. Bals, B. M. Hauser, J. Feldman, C. Muus, M. H. Wadsworth 2nd, S. W. Kazer, T. K. Hughes, B. Doran, G. J. Gatter, M. Vukovic, F. Taliaferro, B. E. Mead, Z. Guo, J. P. Wang, D. Gras, M. Plaisant, M. Ansari, I. Angelidis, H. Adler, J. M. S. Sucre, C. J. Taylor, B. Lin, A. Waghray, V. Mitsialis, D. F. Dwyer, K. M. Buchheit, J. A. Boyce, N. A. Barrett, T. M. Laidlaw, S. L. Carroll, L. Colonna, V. Tkachev, C. W. Peterson, A. Yu, H. B. Zheng, H. P. Gideon, C. G. Winchell, P. L. Lin, C. D. Bingle, S. B. Snapper, J. A. Kropski, F. J. Theis, H. B. Schiller, L.-E. Zaragosi, P. Barbry, A. Leslie, H.-P. Kiem, J. L. Flynn, S. M. Fortune, B. Berger, R. W. Finberg, L. S. Kean, M. Garber, A. G. Schmidt, D. Lingwood, A. K. Shalek and J. Ordoval-Montanes, HCA Lung Biological Network. Electronic address: lung-network@humancellatlas.org; HCA Lung Biological Network, *Cell*, 2020, **181**, 1016–1035.e19.
- 19 I. Hamming, H. van Goor, A. J. Turner, C. A. Rushworth, A. A. Michaud, P. Corvol and G. Navis, *Exp. Physiol.*, 2008, **93**, 631–638.
- 20 C. M. Ferrario, J. Jessup, M. C. Chappell, D. B. Averill, K. B. Brosnihan, E. A. Tallant, D. I. Diz and P. E. Gallagher, *Circulation*, 2005, **111**, 2605–2610.
- 21 M. P. Ocaranza, I. Godoy, J. E. Jalil, M. Varas, P. Collantes, M. Pinto, M. Roman, C. Ramirez, M. Copaja, G. Diaz-Araya, P. Castro and S. Lavandero, *Hypertension*, 2006, **48**, 572–578.
- 22 L. J. Burchill, E. Velkoska, R. G. Dean, K. Griggs, S. K. Patel and L. M. Burrell, *Clin. Sci.*, 2012, **123**, 649–658.
- 23 L. M. Burrell, J. Risvanis, E. Kubota, R. G. Dean, P. S. MacDonald, S. Lu, C. Tikellis, S. L. Grant, R. A. Lew, A. I. Smith, M. E. Cooper and C. I. Johnston, *Eur. Heart J.*, 2005, **26**, 369–375.
- 24 R. N. Vuille-dit-Bille, S. M. Camargo, L. Emmenegger, T. Sasse, E. Kummer, J. Jando, Q. M. Hamie, C. F. Meier, S. Hunziker, Z. Forras-Kaufmann, S. Kuyumcu, M. Fox, W. Schwizer, M. Fried, M. Lindenmeyer, O. Götze and F. Verrey, *Amino Acids*, 2015, **47**, 693–705.
- 25 M. J. Soler, M. Ye, J. Wysocki, J. William, J. Lloveras and D. Battle, *Am. J. Physiol. Renal Physiol.*, 2009, **296**, F398–F405.
- 26 V. Sukumaran, P. T. Veeraveedu, N. Gurusamy, A. P. Lakshmanan, K. Yamaguchi, M. Ma, K. Suzuki, M. Nagata, R. Takagi, M. Kodama and K. Watanabe, *Mol. Cell. Endocrinol.*, 2012, **351**, 208–219.
- 27 A. P. Lakshmanan, R. A. Thandavarayan, K. Watanabe, F. R. Sari, H. Meilei, V. V. Giridharan, V. Sukumaran, V. Soetikno, S. Arumugam, K. Suzuki and M. Kodama, *Mol. Cell. Endocrinol.*, 2012, **348**, 104–111.
- 28 C. M. Ferrario, S. Ahmad and L. Groban, *Nat. Rev. Cardiol.*, 2020, **17**, 378.
- 29 Y. Ishiyama, P. E. Gallagher, D. B. Averill, E. A. Tallant, K. B. Brosnihan and C. M. Ferrario, *Hypertension*, 2004, **43**, 970–976.
- 30 M. Igase, W. B. Strawn, P. E. Gallagher, R. L. Geary and C. M. Ferrario, *Am. J. Physiol.: Heart Circ. Physiol.*, 2005, **289**, H1013–H1019.
- 31 J.-C. Zhong, J.-Y. Ye, H.-Y. Jin, X. Yu, H.-M. Yu, D.-L. Zhu, P.-J. Gao, D.-Y. Huang, M. Shuster, H. Loibner, J.-M. Guo, X.-Y. Yu, B.-X. Xiao, Z.-H. Gong, J. M. Penninger and G. Y. Oudit, *Regul. Pept.*, 2011, **166**, 90–97.
- 32 V. Sukumaran, P. T. Veeraveedu, N. Gurusamy, K. Yamaguchi, A. P. Lakshmanan, M. Ma, K. Suzuki, M. Kodama and K. Watanabe, *Int. J. Biol. Sci.*, 2011, **7**, 1077–1092.
- 33 V. Sukumaran, H. Tsuchimochi, E. Tatsumi, M. Shirai and J. T. Pearson, *Biochem. Pharmacol.*, 2017, **144**, 90–99.
- 34 M. Furuhashi, N. Moniwa, T. Mita, T. Fuseya, S. Ishimura, K. Ohno, S. Shibata, M. Tanaka, Y. Watanabe, H. Akasaka, H. Ohnishi, H. Yoshida, H. Takizawa, S. Saitoh, N. Ura, K. Shimamoto and T. Miura, *Am. J. Hypertens.*, 2015, **28**, 15–21.
- 35 M. Vaduganathan, O. Vardeny, T. Michel, J. J. V. McMurray, M. A. Pfeffer and S. D. Solomon, *N. Engl. J. Med.*, 2020, **382**, 1653–1659.
- 36 G. I. Rice, D. A. Thomas, P. J. Grant, A. J. Turner and N. M. Hooper, *Biochem. J.*, 2004, **383**, 45–51.
- 37 Y. Li, Z. Zeng, Y. Li, W. Huang, M. Zhou, X. Zhang and W. Jiang, *Shock*, 2015, **43**, 395–404.
- 38 X. M. Wang, K. Shi, J. J. Li, T. T. Chen, Y. H. Guo, Y. L. Liu, Y. F. Yang and S. Yang, *Genet. Mol. Res.*, 2015, **14**, 1707–1717.
- 39 H.-L. Xiao, L.-X. Zhao, J. Yang, N. Tong, L. An, Q.-T. Liu, M.-R. Xie and C.-S. Li, *Mol. Med. Rep.*, 2018, **17**, 4221–4228.
- 40 Y. Chen, F. Song, G. Lu and Z. Lu, *Zhonghua Erke Zazhi*, 2014, **52**, 602–606.
- 41 Z.-b. Xu, Y.-l. Feng, J. Xiao, Y.-j. Tang and T. Qiu, *Sichuan Daxue Xuebao, Yixueban*, 2008, **39**, 583–587.
- 42 E. D. C. Frantz, C. Crespo-Mascarenhas, A. R. C. Barreto-Vianna, M. B. Aguila and C. A. Mandarim-de-Lacerda, *PLoS One*, 2013, **8**, e67192.
- 43 X. Wang, Y. Ye, H. Gong, J. Wu, J. Yuan, S. Wang, P. Yin, Z. Ding, L. Kang, Q. Jiang, W. Zhang, Y. Li, J. Ge and Y. Zou, *J. Mol. Cell. Cardiol.*, 2016, **97**, 180–190.
- 44 Y.-M. Yuan, L. Luo, Z. Guo, M. Yang, R.-S. Ye and C. Luo, *J. Renin-Angiotensin-Aldosterone Syst.*, 2015, **16**, 249–253.
- 45 S. Hagiwara, H. Iwasaka, S. Hidaka, A. Hasegawa, H. Koga and T. Noguchi, *Intensive Care Med.*, 2009, **35**, 1471–1478.
- 46 S.-X. Han, G.-M. He, T. Wang, L. Chen, Y.-Y. Ning, F. Luo, J. An, T. Yang, J.-J. Dong, Z.-L. Liao, D. Xu and F.-Q. Wen, *Toxicol. Appl. Pharmacol.*, 2010, **245**, 100–107.



- 47 J. Klimas, M. Olvedy, K. Ochodnicka-Mackovicova, P. Kruzliak, S. Cacanyiova, F. Kristek, P. Krenek and P. Ochodnický, *J. Cell. Mol. Med.*, 2015, **19**, 1965–1974.
- 48 G. E. Callera, T. T. Antunes, J. W. Correa, D. Moorman, A. Gutsol, Y. He, A. N. D. Cat, A. M. Briones, A. C. Montezano, K. D. Burns and R. M. Touyz, *Biosci. Rep.*, 2016, **36**, e00398.
- 49 J. Iwanami, M. Mogi, K. Tsukuda, X.-L. Wang, H. Nakaoka, K. Ohshima, T. Chisaka, H.-Y. Bai, H. Kanno, L.-J. Min and M. Horiuchi, *Hypertens. Res.*, 2014, **37**, 616–620.
- 50 F. Graus-Nunes, F. de Oliveira Santos, T. de Souza Marinho, C. S. Miranda, S. Barbosa-da-Silva and V. Souza-Mello, *World J. Hepatol.*, 2019, **11**, 359–369.
- 51 M. Yisireyli, Y. Uchida, K. Yamamoto, T. Nakayama, X. Wu Cheng, T. Matsushita, S. Nakamura, T. Murohara and K. Takeshita, *Brain Behav. Immun.*, 2018, **69**, 167–179.
- 52 T. Tsutamoto, K. Nishiyama, M. Yamaji, C. Kawahara, M. Fujii, T. Yamamoto and M. Horie, *Hypertens. Res.*, 2010, **33**, 118–122.
- 53 T. Tanno, H. Tomita, I. Narita, T. Kinjo, K. Nishizaki, H. Ichikawa, Y. Kimura, M. Tanaka, T. Osanai and K. Okumura, *J. Cardiovasc. Pharmacol.*, 2016, **67**, 503–509.
- 54 M. H. M. Yousif, G. S. Dhaunsi, B. M. Makki, B. A. Qabazard, S. Akhtar and I. F. Benter, *Pharmacol. Res.*, 2012, **66**, 269–275.
- 55 S. Arumugam, R. A. Thandavarayan, S. S. Palaniyandi, V. V. Giridharan, W. Arozal, F. R. Sari, V. Soetikno, M. Harima, K. Suzuki, M. Kodama and K. Watanabe, *Toxicology*, 2012, **291**, 139–145.
- 56 V. Sukumaran, P. T. Veeraveedu, N. Gurusamy, A. P. Lakshmanan, K. Yamaguchi, M. Ma, K. Suzuki, M. Kodama and K. Watanabe, *Life Sci.*, 2012, **90**, 289–300.
- 57 V. Sukumaran, P. T. Veeraveedu, A. P. Lakshmanan, N. Gurusamy, K. Yamaguchi, M. Ma, K. Suzuki, M. Kodama and K. Watanabe, *Free Radic. Res.*, 2012b, **46**, 850–860.
- 58 A. Molteni, L. F. Wolfe, W. F. Ward, C. H. Ts'ao, L. B. Molteni, P. Veno, B. L. Fish, J. M. Taylor, N. Quintanilla, B. Herndon and J. E. Moulder, *Curr. Pharm. Des.*, 2007, **13**, 1307–1316.
- 59 D. Lezama-Martinez, J. Flores-Monroy, S. Fonseca-Coronado, M. E. Hernandez-Campos, I. Valencia-Hernandez and L. Martinez-Aguilar, *J. Cardiovasc. Pharmacol.*, 2018, **72**, 291–295.
- 60 L. Fan, Y. Feng, H. Y. Wan, L. Ni, Y. R. Qian, Y. Guo, Y. Xiang and Q. Y. Li, *Genet. Mol. Res.*, 2014, **13**, 10562–10573.
- 61 R. Basu, M. Poglitsch, H. Yogasundaram, J. Thomas, B. H. Rowe and G. Y. Oudit, *J. Am. Coll. Cardiol.*, 2017, **69**, 805–819.
- 62 Z. Zou, Y. Yan, Y. Shu, R. Gao, Y. Sun, X. Li, X. Ju, Z. Liang, Q. Liu, Y. Zhao, F. Guo, T. Bai, Z. Han, J. Zhu, H. Zhou, F. Huang, C. Li, H. Lu, N. Li, D. Li, N. Jin, J. M. Penninger and C. Jiang, *Nat. Commun.*, 2014, **5**, 3594.
- 63 H. Gu, Z. Xie, T. Li, S. Zhang, C. Lai, P. Zhu, K. Wang, L. Han, Y. Duan, Z. Zhao, X. Yang, L. Xing, P. Zhang, Z. Wang, R. Li, J. J. Yu, X. Wang and P. Yang, *Sci. Rep.*, 2016, **6**, 19840.
- 64 A. Khan, C. Benthin, B. Zeno, T. E. Albertson, J. Boyd, J. D. Christie, R. Hall, G. Poirier, J. J. Ronco, M. Tidswell, K. Hardes, W. M. Powley, T. J. Wright, S. K. Siederer, D. A. Fairman, D. A. Lipson, A. I. Bayliffe and A. L. Lazaar, *Crit. Care*, 2017, **21**, 234.
- 65 V. B. Patel, J. N. Lezutekong, X. Chen and G. Y. Oudit, *Can. J. Cardiol.*, 2017, **33**, 943–946.
- 66 E. H. Bae, F. Fang, V. R. Williams, A. Konvalinka, X. Zhou, V. B. Patel, X. Song, R. John, G. Y. Oudit, Y. Pei and J. W. Scholey, *Kidney Int.*, 2017, **91**, 1347–1361.
- 67 G. Y. Oudit, G. C. Liu, J. Zhong, R. Basu, F. L. Chow, J. Zhou, H. Loibner, E. Janzek, M. Schuster, J. M. Penninger, A. M. Herzenberg, Z. Kassiri and J. W. Scholey, *Diabetes*, 2010, **59**, 529–538.
- 68 V. Monteil, H. Kwon, P. Prado, A. Hagekkrüys, R. A. Wimmer, M. Stahl, A. Leopoldi, E. Garreta, C. H. D. Pozo, F. Prosper, J. P. Romero, G. Wirnsberger, H. Zhang, A. S. Slutsky, R. Conder, N. Montserrat, A. Mirazimi and J. M. Penninger, *Cell*, 2020, **181**, 905–913.e7.
- 69 F. Alhenc-Gelas and T. B. Drueke, *Kidney Int.*, 2020, **97**, 1091–1093.

