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## The P-type ATPase inhibiting potential of polyoxotungstates†

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Polyoxometalates (POMs) are transition metal complexes that exhibit a broad diversity of structures and properties rendering them promising for biological purposes. POMs are able to inhibit a series of biologically important enzymes, including phosphatases, and thus are able to affect many biochemical processes. In the present study, we analyzed and compared the inhibitory effects of nine different polyoxotungstates (POTs) on two P-type ATPases,  $\text{Ca}^{2+}$ -ATPase from skeletal muscle and  $\text{Na}^+/\text{K}^+$ -ATPase from basal membrane of skin epithelia. For  $\text{Ca}^{2+}$ -ATPase inhibition, an *in vitro* study was performed and the strongest inhibitors were determined to be the large heteropolytungstate  $\text{K}_9(\text{C}_2\text{H}_8\text{N})_5[\text{H}_{10}\text{Se}_2\text{W}_{29}\text{O}_{103}]$  ( $\text{Se}_2\text{W}_{29}$ ) and the Dawson-type POT  $\text{K}_6[\alpha\text{-P}_2\text{W}_{18}\text{O}_{62}]$  ( $\text{P}_2\text{W}_{18}$ ) exhibiting  $\text{IC}_{50}$  values of 0.3 and 0.6  $\mu\text{M}$ , respectively. Promising results were also shown for the Keggin-based POTs  $\text{K}_6\text{H}_2[\text{CoW}_{11}\text{TiO}_{40}]$  ( $\text{CoW}_{11}\text{Ti}$ ,  $\text{IC}_{50} = 4 \mu\text{M}$ ) and  $\text{Na}_{10}[\alpha\text{-SiW}_9\text{O}_{34}]$  ( $\text{SiW}_9$ ,  $\text{IC}_{50} = 16 \mu\text{M}$ ),  $\text{K}_{14}[\text{As}_2\text{W}_{19}\text{O}_{67}(\text{H}_2\text{O})]$  ( $\text{As}_2\text{W}_{19}$ ,  $\text{IC}_{50} = 28 \mu\text{M}$ ) and the lacunary Dawson  $\text{K}_{12}[\alpha\text{-H}_2\text{P}_2\text{W}_{12}\text{O}_{48}]$  ( $\text{P}_2\text{W}_{12}$ ,  $\text{IC}_{50} = 11 \mu\text{M}$ ), whereas low inhibitory potencies were observed for the isopolytungstate  $\text{Na}_{12}[\text{H}_4\text{W}_{22}\text{O}_{74}]$  ( $\text{W}_{22}$ ,  $\text{IC}_{50} = 68 \mu\text{M}$ ) and the Anderson-type  $\text{Na}_6[\text{TeW}_6\text{O}_{24}]$  ( $\text{TeW}_6$ ,  $\text{IC}_{50} = 200 \mu\text{M}$ ). Regarding the inhibition of  $\text{Na}^+/\text{K}^+$ -ATPase activity, for the first time an *ex vivo* study was conducted using the opercular epithelium of killifish in order to investigate the effects of POTs on the epithelial chloride secretion. Interestingly, 1  $\mu\text{M}$  of the most potent  $\text{Ca}^{2+}$ -ATPase inhibitor,  $\text{Se}_2\text{W}_{29}$ , showed only a minor inhibitory effect (14% inhibition) on  $\text{Na}^+/\text{K}^+$ -ATPase activity, whereas almost total inhibition (99% inhibition) was achieved using  $\text{P}_2\text{W}_{18}$ . The remaining POTs exhibited similar inhibition rates on both ATPases. These results reveal the high potential of some POTs to act as P-type ATPase inhibitors, with  $\text{Se}_2\text{W}_{29}$  showing high selectivity towards  $\text{Ca}^{2+}$ -ATPase.

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### Significance to metallomics

We studied the inhibitory effects of nine different polyoxotungstates (POTs) on P-type ATPases *in vitro* ( $\text{Ca}^{2+}$ -ATPase) and *ex vivo* ( $\text{Na}^+/\text{K}^+$ -ATPase). The study reveals that some POTs like the Dawson anion  $[\text{P}_2\text{W}_{18}\text{O}_{62}]^{6-}$ , which was highly active *in vitro* and *ex vivo*, are potent ATPase inhibitors. Furthermore, there is a charge density-activity correlation for the most potent POTs ( $\text{IC}_{50} < 16 \mu\text{M}$ ), namely  $\text{Se}_2\text{W}_{29}$ ,  $\text{P}_2\text{W}_{18}$ ,  $\text{CoW}_{11}\text{Ti}$ ,  $\text{SiW}_9$  and  $\text{P}_2\text{W}_{12}$ . As P-type ATPases represent pharmacologically important targets due to their important role in health and disease, the here reported bioactive POTs should be considered as possible future metallodrugs.

## Introduction

Polyoxometalates (POMs) are metal clusters<sup>1</sup> that exhibit a broad diversity of structures and outstanding properties leading to their application in various fields such as catalysis,<sup>2,3</sup> photochemistry,<sup>4</sup> material science,<sup>5,6</sup> macromolecular crystallography<sup>7–15</sup> and medicine.<sup>16–22</sup> POMs can be divided into isopolyanions (IPAs), which consist only of one type of metal atom ( $\text{M} = \text{addenda atom}$ ),  $[\text{M}_m\text{O}_y]^{q-}$ , and heteropolyanions (HPAs), which contain one or more additional elements ( $\text{X} = \text{heteroatom}$ ),  $[\text{X}_m\text{M}_m\text{O}_y]^{q-}$ . The most common representative of IPAs is the Lindqvist structure,

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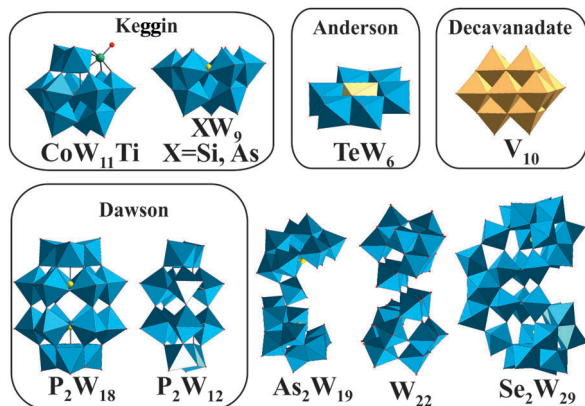
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**Fig. 1** Structures of the investigated POTs (Table 1) and  $[V_{10}O_{28}]^{6-}$ . Color code:  $WO_6$ , blue polyhedra;  $VO_6$ , dark yellow polyhedra; heteroatom, light yellow sphere or polyhedra; Ti as substituted atom, green sphere.

whereas the well-known Dawson, Keggin and Anderson archetypes belong to the HPAs (Fig. 1). POM research represents an emerging field and especially bioactive POMs are getting more and more attractive due to their ability to interact with important enzymes like alkaline phosphatases, ecto-nucleotidases and ATPases and their potential to interfere with specific cellular processes, such as mitochondria respiration.<sup>21–24</sup> POMs like decavanadate or Keggin-type polyoxotungstates (POTs) and polyoxomolybdates are currently the focus of biological and biomedical research as they show promising antibacterial and antidiabetic activities,<sup>21,22,24–28</sup> whereas only few biological studies exist for other POM archetypes such as the Anderson structure.<sup>29</sup>

The main role of the sarcoplasmic reticulum (SR)  $\text{Ca}^{2+}$ -ATPase is translocation of cellular  $\text{Ca}^{2+}$  from the cytoplasm to the SR, which is involved in muscle relaxation.<sup>30,31</sup> However,  $\text{Ca}^{2+}$ -ATPase is globally associated with cellular calcium homeostasis, a process of ion transport that is coupled with ATP hydrolysis. ATP hydrolysis follows a well-known mechanism traversing at least four intermediate steps and two protein conformations, namely E1 and E2, with E1 being the conformation with high affinity for the exported substrate and E2 the form with high affinity for the imported substrate.<sup>30,31</sup> As SR vesicles from skeletal muscle contain a large amount of  $\text{Ca}^{2+}$ -ATPase, they represent a useful *in vitro* model to study the effects of drugs and POMs on calcium homeostasis.<sup>32,33</sup> To our knowledge, only a few POMs, such as decavanadate ( $\text{V}_{10}$ ) and decaniobate ( $\text{Nb}_{10}$ ), were described to be

potent non-competitive inhibitors ( $IC_{50} = 15$  and  $35 \mu M$ , respectively) of the hydrolytic activity of SR  $Ca^{2+}$ -ATPase.<sup>33</sup>  $Na^+/K^+$ -ATPase transports  $Na^+$  out of the cell while pumping  $K^+$  into cells and is thus responsible for the ionic and osmotic balance in cells and an important transducer of signals. As all P-type ATPases, the  $Na^+/K^+$  pump derives energy from ATP hydrolysis.

Herein, we report and compare the effects of nine different POTs (Fig. 1 and Table 1) on the *in vitro* activity of  $\text{Ca}^{2+}$ -ATPase from SR. For the first time, we investigate the effects of POTs on the process of epithelial chloride secretion, energized by the activity of basolateral  $\text{Na}^+/\text{K}^+$ -ATPase, using an *ex vivo* model obtained from basal membrane of epithelial skin (killifish). Putative correlations between the inhibitory activity of POTs ( $\text{IC}_{50}$  values), their charge density and size were derived. The results reveal that some POTs are potent inhibitors of P-type ATPases even under almost physiological conditions (*ex vivo* study) and therefore should be taken into consideration as P-type ATPase targeting drugs. One POT, namely  $\text{K}_9(\text{C}_2\text{H}_8\text{N})_5[\text{H}_{10}\text{Se}_2\text{W}_{29}\text{O}_{103}]$  ( $\text{Se}_2\text{W}_{29}$ ) showed clear selectivity towards one pump ( $\text{Ca}^{2+}$ -ATPase), whereas other POTs like the Anderson archetype  $\text{Na}_6[\text{TeW}_6\text{O}_{24}]$  showed very low inhibition on both ion pumps.

## Experimental section

## Polyoxometalates

The POTs used in this study,  $\text{K}_6[\alpha\text{-P}_2\text{W}_{18}\text{O}_{62}]\cdot 14\text{H}_2\text{O}$ ,<sup>35</sup>  $\text{Na}_6\text{-}[\text{TeW}_6\text{O}_{24}]\cdot 22\text{H}_2\text{O}$ ,<sup>36</sup>  $\text{K}_6\text{H}_2[\text{TiW}_{11}\text{CoO}_{40}]\cdot 13\text{H}_2\text{O}$ ,<sup>37</sup>  $\text{Na}_{10}[\alpha\text{-SiW}_9\text{O}_{34}]\cdot 16\text{H}_2\text{O}$ ,<sup>35</sup>  $\text{Na}_9[\alpha\text{-AsW}_9\text{O}_{33}]\cdot 27\text{H}_2\text{O}$ ,<sup>38</sup>  $\text{K}_{12}[\alpha\text{-H}_2\text{P}_2\text{W}_{12}\text{O}_{48}]\cdot 16\text{H}_2\text{O}$ ,<sup>35</sup>  $\text{K}_{14}[\text{As}_2\text{W}_{19}\text{O}_{67}(\text{H}_2\text{O})]\cdot 23\text{H}_2\text{O}$ ,<sup>39</sup>  $\text{K}_9(\text{C}_2\text{H}_8\text{N})_5[\text{H}_{10}\text{Se}_2\text{W}_{29}\text{O}_{103}]\cdot 30\text{H}_2\text{O}$ <sup>40</sup> and  $\text{Na}_{12}[\text{H}_4\text{W}_{22}\text{O}_{74}]\cdot 50\text{H}_2\text{O}$ <sup>41</sup> (Table 1 and Fig. 1), were synthesized according to published procedures (see references in Table 1) and their identity was confirmed by infrared spectroscopy. Stock solutions of POTs were freshly prepared by dissolving the solid compound in water and keeping the solution on ice to avoid POT decomposition. The concentrations of the stock solutions were 10 mM and 1 mM for all POTs except for  $\text{Se}_2\text{W}_{29}$  (1 mM and 0.1 mM).

### Preparation of sarcoplasmic reticulum $\text{Ca}^{2+}$ -ATPase vesicles

All reagents used for the preparation of the calcium pump vesicles were purchased from Sigma-Aldrich (Portugal). Isolated sarcoplasmic reticulum vesicles (SRVs), prepared from rabbit skeletal muscles as described elsewhere,<sup>33</sup> were suspended in

**Table 1** Structural/molecular features of the POTs used in this study

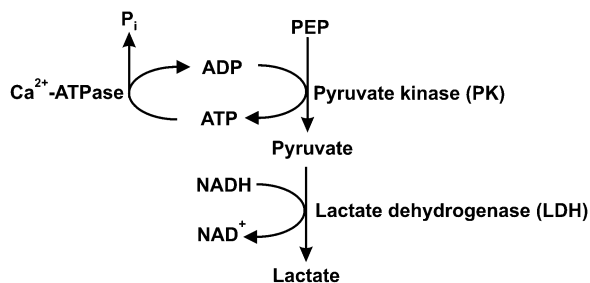
POTs (abbreviated)	Sum formula	$M_r$	Charge	POT archetype	Ref.
P <sub>2</sub> W <sub>18</sub>	K <sub>6</sub> [ $\alpha$ -P <sub>2</sub> W <sub>18</sub> O <sub>62</sub> ] $\cdot$ 14H <sub>2</sub> O	4849.83	6 $-$	Dawson	35
TeW <sub>6</sub>	Na <sub>6</sub> [TeW <sub>6</sub> O <sub>24</sub> ] $\cdot$ 22H <sub>2</sub> O	2148.56	6 $-$	Anderson-Evans	36
CoW <sub>11</sub> Ti	K <sub>6</sub> H <sub>2</sub> [TiW <sub>11</sub> CoO <sub>40</sub> ] $\cdot$ 13H <sub>2</sub> O	3239.62	8 $-$	Mono-substituted Keggin	37
AsW <sub>9</sub>	Na <sub>9</sub> [B- $\alpha$ -AsW <sub>9</sub> O <sub>33</sub> ] $\cdot$ 27H <sub>2</sub> O	2950.37	9 $-$	Tri-lacunary Keggin	38
SiW <sub>9</sub>	Na <sub>10</sub> [A- $\alpha$ -SiW <sub>9</sub> O <sub>34</sub> ] $\cdot$ 16H <sub>2</sub> O	2744.52	10 $-$	Tri-lacunary Keggin	35
P <sub>2</sub> W <sub>12</sub>	K <sub>12</sub> [ $\alpha$ -H <sub>2</sub> P <sub>2</sub> W <sub>12</sub> O <sub>48</sub> ] $\cdot$ 16H <sub>2</sub> O	3795.19	12 $-$	Lacunary Dawson	35
As <sub>2</sub> W <sub>19</sub>	K <sub>14</sub> [As <sub>2</sub> W <sub>19</sub> O <sub>67</sub> (H <sub>2</sub> O)] $\cdot$ 23H <sub>2</sub> O	5694.15	14 $-$	Doubled anion based on tri-lacunary Keggin anions	39
Se <sub>2</sub> W <sub>29</sub>	K <sub>9</sub> (C <sub>2</sub> H <sub>8</sub> N) <sub>5</sub> [H <sub>10</sub> Se <sub>2</sub> W <sub>29</sub> O <sub>103</sub> ] $\cdot$ 30H <sub>2</sub> O	8270.09	14 $-$	Lacunary anion based on two tri-lacunary Keggin anions containing {(WO <sub>7</sub> )W <sub>4</sub> } pentagonal unit	40
W <sub>22</sub>	Na <sub>12</sub> [H <sub>4</sub> W <sub>22</sub> O <sub>74</sub> ] $\cdot$ 50H <sub>2</sub> O	6409.10	12 $-$	Dimeric isopolyanion based on two {W <sub>11</sub> } units	41

The preparations were left to stand for at least 60 min or until a steady basal measurement of bioelectrical variables was achieved. Measurement of the short circuit current ( $I_{sc}$ ,  $\mu A\ cm^{-2}$ ) was performed at symmetric conditions under voltage clamp

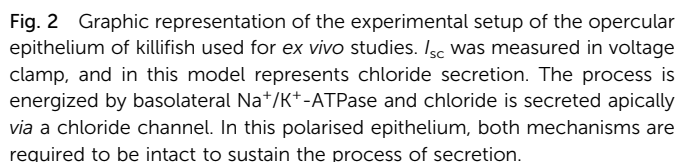
### Effects of POTs on ATP hydrolysis of SR Ca<sup>2+</sup>-ATPase

Steady-state assays of the SR  $\text{Ca}^{2+}$ -ATPase were measured spectrophotometrically at 25 °C using the coupled enzyme pyruvate kinase/lactate dehydrogenase assay (Scheme 1) as described elsewhere.<sup>42</sup> Briefly, after the addition of the enzymes (pyruvate kinase and lactate dehydrogenase) and the substrate phosphoenolpyruvate to the medium, the experiment was initiated by adding NADH (0.25 mM) and the vesicles containing  $\text{Ca}^{2+}$ -ATPase (10  $\mu\text{g mL}^{-1}$ ).

ATP (2.5 mM) was added and the absorbance was recorded for about 1 minute (basal activity). Afterwards, the calcium ionophore calcimycin 4% (w/w), which releases again the  $\text{Ca}^{2+}$  ions, that were pumped in by the ATPase, was added and the decreasing NADH absorbance at 340 nm was measured for about 2 minutes (uncoupled ATPase activity). This was done to increase the ATPase activity (due to the ionophore-mediated impairment of the  $\text{Ca}^{2+}$  gradient) in order to better study the effect of the inhibitors and to ensure that the SR  $\text{Ca}^{2+}$ -ATPase vesicles are not leaky. For the experiments including POTs, freshly prepared POT solutions (10 mM and 1 mM for all POTs except for  $\text{Se}_2\text{W}_{29}$  1 mM and 0.1 mM) were added to the medium prior to the addition of SR  $\text{Ca}^{2+}$ -ATPase. The ATPase activity and its inhibition was measured taking into account the decrease of the OD (optical density) per minute in the absence (100%) and in the presence of the investigated POTs.<sup>33</sup> The detection system was not affected by the POTs themselves (not even at their highest concentrations), which was confirmed by a rapid decrease in absorbance at 340 nm upon addition of 40  $\mu\text{M}$  ADP after the assay. All experiments were performed at least in triplicates. The inhibitory power of the investigated



**Scheme 1** Coupled enzymatic assay for  $\text{Ca}^{2+}$ -ATPase activity. PEP – phosphoenolpyruvate;  $\text{P}_i$  – inorganic phosphate.



**Fig. 2** Graphic representation of the experimental setup of the opercular epithelium of killifish used for *ex vivo* studies.  $I_{sc}$  was measured in voltage clamp, and in this model represents chloride secretion. The process is energized by basolateral  $\text{Na}^+/\text{K}^+$ -ATPase and chloride is secreted apically *via* a chloride channel. In this polarised epithelium, both mechanisms are required to be intact to sustain the process of secretion.

## Results and discussion

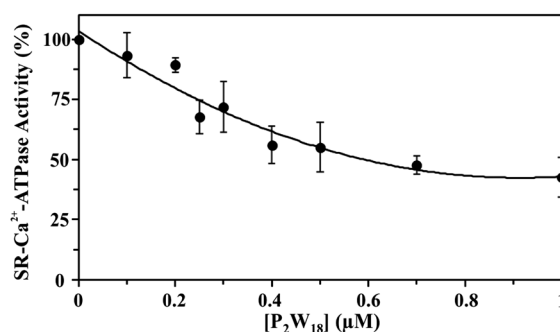
### Inhibition of Ca<sup>2+</sup>-ATPase by POTs: *in vitro* study

The effect of nine different POTs (Fig. 1 and Table 1) on the activity of SR  $\text{Ca}^{2+}$ -ATPase from skeletal muscle was investigated for the first time. All of the investigated POTs inhibited  $\text{Ca}^{2+}$ -ATPase activity in a concentration dependent manner. The inhibitory power

POTs	Ca <sup>2+</sup> -ATPase	Na <sup>+</sup> /K <sup>+</sup> -ATPase	
Compound name	IC <sub>50</sub> , (μM)	ET <sub>50</sub> , (min (depending on concentration of compound))	Maximum inhibition, (%)
P <sub>2</sub> W <sub>18</sub>	0.6	8.2 (0.5 μM)	86
		6.5 (1 μM)	99
		4.3 (10 μM)	100
TeW <sub>6</sub>	200	60 (10 μM)	10
CoW <sub>11</sub> Ti	4	10 (10 μM)	75
SiW <sub>9</sub>	16	nd	nd
P <sub>2</sub> W <sub>12</sub>	11	nd	nd
As <sub>2</sub> W <sub>19</sub>	28	nd	nd
Se <sub>2</sub> W <sub>29</sub>	0.3	6.5 (1 μM)	14
W <sub>22</sub>	68	nd	nd
AsW <sub>9</sub>	20	8.5 (10 μM)	66
Ouabain	—	3.2 (10 μM)	100

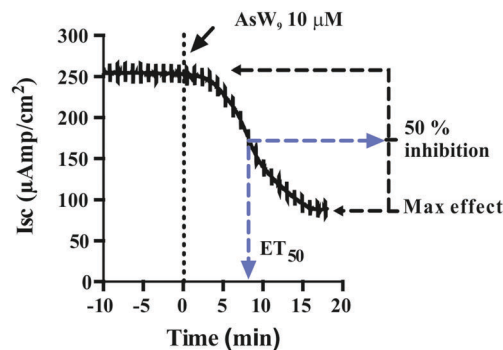
of the investigated POTs was finally evaluated using  $IC_{50}$  values (Table 2). As shown in Table 2, different  $IC_{50}$  values in the  $\mu M$  range were determined for the various POT archetypes exhibiting different negative charges (Fig. 1 and Table 1).  $IC_{50}$  values of  $< 1 \mu M$  were determined for the Dawson anion  $P_{2}W_{18}$  ( $0.6 \mu M$ , Fig. 3) and the larger POT  $Se_{2}W_{29}$  ( $IC_{50} = 0.3 \mu M$ ), whereas the lowest inhibition values were observed for the isopolyanion  $W_{22}$  ( $IC_{50} = 68 \mu M$ ) and the Anderson type  $TeW_6$  ( $IC_{50} = 200 \mu M$ ). The remaining POTs exhibited  $IC_{50}$  values in the range of 1 up to  $28 \mu M$  (Table 2).

Similar moderate IC<sub>50</sub> values for SR Ca<sup>2+</sup>-ATPase activity were previously reported for two isostructural polyanions, decaniobate [Nb<sub>10</sub>O<sub>28</sub>]<sup>6-</sup> (IC<sub>50</sub> = 35 μM) and decavanadate [V<sub>10</sub>O<sub>28</sub>]<sup>6-</sup> (IC<sub>50</sub> = 15 μM).<sup>33,46</sup> Both decaniobate and decavanadate showed a non-competitive inhibition for Ca<sup>2+</sup>-ATPase activity



**Fig. 3** Inhibition of  $\text{Ca}^{2+}$ -ATPase activity by  $\text{P}_2\text{W}_{18}$ .  $\text{Ca}^{2+}$ -ATPase was determined spectrophotometrically at 340 nm and 25 °C, using the coupled enzyme pyruvate kinase/lactate dehydrogenase assay. The experiments were initiated by the addition of  $10 \mu\text{g mL}^{-1}$   $\text{Ca}^{2+}$ -ATPase, in the presence or absence of 4% (w/w) of the calcium ionophore calcimycin. Data are plotted as means  $\pm$  SD and fit to the equation  $y = 73.126x^2 - 133.62x + 103.45$  ( $R^2 = 0.9424$ ).





**Fig. 5** The effect of the Keggin type  $\text{AsW}_9$  applied in basolateral membranes at a concentration of  $10 \mu\text{M}$  is shown. Original trace of the effect of short circuit current ( $I_{\text{sc}}$ ,  $\mu\text{A cm}^{-2}$ ) in the opercular epithelium of killifish mounted in Ussing chambers and kept under voltage clamp ( $V_t = 0 \text{ mV}$ ). Effective time  $50$  ( $\text{ET}_{50}$ ) and maximum inhibitory effects are calculated as the % of basal values. Both parameters were calculated for three individual independent experiments and used to generate Table 2. An arrow indicates the time of POT application and consequently the time point zero. Time with negative values represents stable basal control periods.

(providing information about inhibitor efficacy) and  $ET_{50}$  (providing information about inhibition velocity) are necessary to define the biological effects of POTs (Fig. 5 and Table 2). For the *ex vivo* studies, a positive control experiment was performed with the conventional  $Na^+/K^+$ -ATPase inhibitor ouabain.<sup>43,49</sup> Ouabain (at 10  $\mu$ M) showed a maximum inhibition value of 100% and an  $ET_{50}$  of 3.2 minutes (Fig. S1, ESI†). By inhibiting the basolateral  $Na^+/K^+$ -ATPase activity, ouabain concomitantly prevents apical chloride secretion in the studied epithelia model as this process is energized by  $Na^+/K^+$ -ATPase.<sup>43,49</sup>

The addition of POTs to apical saline had no effect on  $I_{sc}$  and therefore ruling out chloride channels as putative POT targets, at least at POT concentrations up to 10  $\mu\text{M}$ . The largest POT (in terms of volume and number of addenda atoms) under investigation,  $\text{Se}_2\text{W}_{29}$ , exhibited the highest inhibition ( $\text{IC}_{50} = 0.3 \mu\text{M}$ ) of SR  $\text{Ca}^{2+}$ -ATPase activity during the *in vitro* study. However, used at the same concentration (1  $\mu\text{M}$ ), it was one of the weakest inhibitors (14% inhibition; Table 2 and Fig. 6) for the  $\text{Na}^+/\text{K}^+$ -ATPase activity during the *ex vivo* study. In contrast,  $\text{P}_2\text{W}_{18}$  efficiently inhibited both the SR  $\text{Ca}^{2+}$ -ATPase *in vitro* ( $\text{IC}_{50} = 0.6 \mu\text{M}$ ) and the  $\text{Na}^+/\text{K}^+$ -ATPase *ex vivo* (99% inhibition) (Table 2 and Fig. 6). In fact,  $\text{P}_2\text{W}_{18}$  was demonstrated to be as potent as ouabain in inhibiting the  $\text{Na}^+/\text{K}^+$ -ATPase activity (Table 2). The remaining studied POTs showed similar inhibitory effects *in vitro* and *ex vivo*. For example, the potential of  $\text{TeW}_6$  to inhibit  $\text{Ca}^{2+}$ -ATPase ( $\text{IC}_{50}$  value of 200  $\mu\text{M}$ ) was as low as its effect against  $\text{Na}^+/\text{K}^+$ -ATPase (inhibition of 10%; Table 2).

Both experiments (*in vitro* and *ex vivo*) clearly demonstrated the high selectivity of  $\text{Se}_2\text{W}_{29}$  for inhibiting the  $\text{Ca}^{2+}$  pump due to its rather sobering *ex vivo* results rendering this POT not the best choice to target the  $\text{Na}^+/\text{K}^+$ -pump *in vivo*. The size (in terms of volume and number of addenda atoms) of this large POT could be one aspect affecting the kinetics of its cellular uptake, thus preventing the POT from targeting the enzyme. The mechanisms of POT uptake and their permeation through



Fig. 6 Inhibition (%) of the  $\text{Na}^+/\text{K}^+$ -ATPase from basal membrane of the skin epithelia by two POTs  $\text{P}_2\text{W}_{18}$  and  $\text{Se}_2\text{W}_{29}$ . The inhibition rate of  $\text{P}_2\text{W}_{18}$  ( $1 \mu\text{M}$ ) was 82% after 30 minutes (red), whereas for the most potent  $\text{Ca}^{2+}$ -ATPase inhibitor so far described,  $\text{Se}_2\text{W}_{29}$  ( $\text{IC}_{50} = 0.3 \mu\text{M}$ ), a minor effect (green) was observed (14% inhibition, after 30 minutes). Perpendicular lines were used to calculate tissue resistance. As it can be observed, the *ex vivo* epithelia preparations retained integrity and selectivity after POT exposure.

epithelia still need to be clarified. The same selectivity pattern for inhibition of  $\text{Ca}^{2+}$ -ATPase activity ( $\text{IC}_{50} = 400 \mu\text{M}$ )<sup>33</sup> over  $\text{Na}^+/\text{K}^+$ -ATPase ( $\text{IC}_{50} = 1.5 \text{ mM}$ )<sup>34</sup> was shown in previous studies for orthotungstate ( $\text{HWO}_4^-$ ). Therefore, it seems that POT-mediated inhibition is pump-specific and there is no POT structure that is perfectly suited for all ion pumps in general. Moreover, the *ex vivo* results show that not only the affinity of the inhibitory compound is relevant, but also how the POT gains access to the inhibition site within an intracellular compartment, rendering the POT- $\text{Na}^+/\text{K}^+$ -ATPase interaction a complex one. The presented combination

of *in vitro* and *ex vivo* studies using two different models to study the effects of POTs on the activity of ATPases indicates the importance of establishing experimental conditions to be as close to the physiological environment as possible.

The majority of P-type ATPase inhibitors in therapy target the  $\text{Na}^+/\text{K}^+$ -ATPase.<sup>32,48</sup> These compounds, which are used for the treatment of several diseases such as heart failure, psychosis, malaria and bacterial infection, show inhibitory capacities resembling those of the here investigated POTs.<sup>48</sup> Only a few kinetic studies have been described so far testing POTs as P-type ATPase inhibitors.<sup>24,32,42,46</sup> The *in vitro* inhibition of  $\text{Na}^+/\text{K}^+$ -ATPase by the Keggin POTs  $\text{H}_3\text{PW}_{12}\text{O}_{40}$  and  $\text{H}_4\text{SiW}_{12}\text{O}_{40}$  was previously described reporting  $\text{IC}_{50}$  values between 3 to  $4 \mu\text{M}$ <sup>32</sup> although information about the type of inhibition and the mechanism of action are still lacking. In this study the comparable  $\text{IC}_{50}$  value for the isostructural Keggin POT  $\text{CoW}_{11}\text{Ti}$  ( $\text{IC}_{50} = 4 \mu\text{M}$ ) was observed.

### Protein interactions and structure/function features of POTs

In order to decipher specific features of the nine POTs that are responsible for the inhibition of  $\text{Ca}^{2+}$ -ATPase, we correlated the POT parameters like size and charge density with their  $\text{IC}_{50}$  values of inhibition (Fig. 7A and B). No correlation was found when considering all nine POTs and thus all determined  $\text{IC}_{50}$  values. However, when taking only into account the high affinity POTs, exhibiting  $\text{IC}_{50}$  values lower than  $16 \mu\text{M}$ , we observed a correlation between their activity ( $\text{IC}_{50}$  value) and their charge density, which was defined as charge of the POT divided by its number of W atoms (Fig. 7A) as well as by the volume of POT anion (Fig. 7B). As can be deduced from this data, POTs such as  $\text{Se}_2\text{W}_{29}$  and  $\text{P}_2\text{W}_{18}$  with a low charge density (Fig. 7A and B) favored the inhibition of  $\text{Ca}^{2+}$ -ATPase activity indicating that besides electrostatic interactions also steric interactions (depending on the shape complementarity between the POT and the inhibition site) might play important roles in the successful inhibition of  $\text{Ca}^{2+}$ -ATPase.

For the *ex vivo* results ( $\text{Na}^+/\text{K}^+$ -ATPase) no correlation between the  $\text{ET}_{50}$  values and POT charge density was observed.

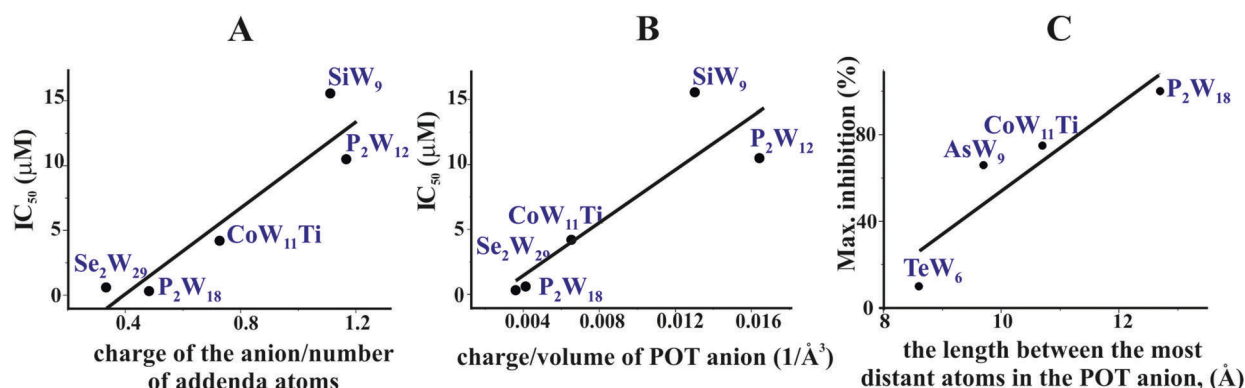


Fig. 7 Structure–activity correlations of different POTs for ATPases inhibition. (A) Correlation between the  $\text{IC}_{50}$  values of five POTs ( $\text{IC}_{50}$  lower than  $16 \mu\text{M}$ ) of  $\text{Ca}^{2+}$ -ATPase inhibition and their charge density expressed as charge of the POT divided by its number of W atoms. (B) Correlation between the  $\text{IC}_{50}$  values of five POTs ( $\text{IC}_{50}$  lower than  $16 \mu\text{M}$ ) against  $\text{Ca}^{2+}$ -ATPase and the charge density expressed as charge of the POT divided by its volume (in  $\text{\AA}^{-3}$ ). (C) Correlation between the percentage of maximum inhibition of four POTs (applied with the same concentration of  $10 \mu\text{M}$ ) against  $\text{Na}^+/\text{K}^+$ -ATPase and POT size expressed as the length between the most distant atoms in the POT anion (in  $\text{\AA}$ ).



**Fig. 8** Electrostatic (Coulomb) potential surface presentation (scale in kcal mol<sup>-1</sup> e<sup>-1</sup>, TM – transmembrane area) of two P-type ATPases: (A) Ca<sup>2+</sup>-pump in E2 conformation<sup>31,32</sup> (PDB entry: 1KJU) showing the proposed binding site of decavanadate V<sub>10</sub>,<sup>47</sup> (B) electronic (coulombic) surface representation of the Na<sup>+</sup>/K<sup>+</sup>-pump (green circles indicate potential binding sites for POMs); (C) structure of the Na<sup>+</sup>/K<sup>+</sup>-ATPase-ouabain complex<sup>31</sup> (PDB entry: 3A3Y). Na<sup>+</sup>/K<sup>+</sup>-ATPase is illustrated as beige cartoon, whereas bound ouabain is depicted in sphere mode.

However, a dependency of the maximum inhibition of four POTs on their size, defined as length between the most distant atoms in the POT anion was found (Fig. 7C).

POTs are also known to be strong kinase and phosphatase inhibitors by acting through noncovalent interactions, which is indispensable for the usage of POMs in the therapy of various diseases.<sup>17–19,26,27</sup> It was demonstrated that decavanadate V<sub>10</sub> exhibits specific interactions with SR Ca<sup>2+</sup>-ATPase, which is supposed to be non-competitive with respect to ATP and induces protein cysteine oxidation with concomitant vanadium reduction explaining the high inhibitory capacity of V<sub>10</sub> (IC<sub>50</sub> = 15 μM).<sup>24,33,47,48</sup> The V<sub>10</sub> binding site, which is formed by three protein domains,<sup>47</sup> is located at the cell cytoplasmic side (Fig. 8A). V<sub>10</sub> can interact with proteins by electrostatic interactions or by hydrogen bonding and the specific residues involved in V<sub>10</sub>-SR Ca<sup>2+</sup>-ATPase interaction still need to be established, but might include the oxidized cysteine.<sup>24</sup> In contrast to monomeric vanadate, which only binds to the E2 conformation, V<sub>10</sub> binds to all protein conformations, E1, E1P, E2 and E2P,<sup>33</sup> indicating the possibility of V<sub>10</sub>-ATPase interactions at the extracellular side of the enzyme.<sup>48</sup> The region where V<sub>10</sub> is expected to bind exhibits a positively charged surface and could therefore also be addressed by other negatively charged POMs (Fig. 8A).

Many drugs are known to act as ionic pumps inhibitors, such as ouabain, omeprazole or thapsigargin, but only for some of these compounds like ouabain the mechanisms and protein binding sites were clearly established.<sup>32</sup> According to structural analysis, ouabain inhibits the Na<sup>+</sup>/K<sup>+</sup>-ATPase through binding to a cavity formed by transmembrane helices (Fig. 8C, PDB entry: 3A3Y).<sup>32</sup> The binding sites for POMs are not known yet, however, considering the structures of POMs and ouabain, it is very unlikely that they share the same binding site (within the neutral transmembrane area). Analysis of the electrostatic (coulombic) surface of Na<sup>+</sup>/K<sup>+</sup>-ATPase reveals that both the cytoplasmic and extracellular region of the enzyme possess areas exhibiting a positive surface potential (Fig. 8B),<sup>31</sup> which

could be addressed by the negatively charged POMs, the binding sites of which need to be identified.

## Conclusions

In general, polyoxometalates are able to inhibit phosphatases, ecto-nucleotidases and P-type ATPases. Here, we demonstrated that the Ca<sup>2+</sup>-ATPase activity from sarcoplasmic reticulum is inhibited by several POTs. P<sub>2</sub>W<sub>18</sub> was the most potent ATPase inhibitor in this study as it exhibited the highest inhibitory activity for the Na<sup>+</sup>/K<sup>+</sup>-ATPase (100% inhibition at 10 μM) and the second highest for the Ca<sup>2+</sup>-ATPase (IC<sub>50</sub> = 0.6 μM). A mixed type of inhibition was observed for P<sub>2</sub>W<sub>18</sub> and TeW<sub>6</sub> suggesting a different mode of protein interaction with Ca<sup>2+</sup>-ATPase activity than those observed for decavanadate and decaniobate (non-competitive inhibitors). The most potent Ca<sup>2+</sup>-ATPase inhibitor Se<sub>2</sub>W<sub>29</sub> showed only limited effects on the Na<sup>+</sup>/K<sup>+</sup>-ATPase from basal membrane of the skin epithelia demonstrating that some POTs exhibit selectivity against certain ion pumps. The here reported *ex vivo* model of Na<sup>+</sup>/K<sup>+</sup>-ATPase was used for the first time to study the effects of POTs on the processes of epithelial chloride secretion, energized by the activity of the basolateral Na<sup>+</sup>/K<sup>+</sup>-ATPase. Finally, we were able to derive structure–activity relationships for high affinity POTs (IC<sub>50</sub> < 16 μM) indicating that the inhibition potential of the POTs is correlated with their charge density, which will help to clarify their different inhibitory activity. Polyoxotungstates are promising inorganic inhibitors of P-type ATPases although their potential *in vivo* applications require more studies and toxicological information.

## Conflicts of interest

There are no conflicts to declare.

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