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## 1 Introduction

The Paris agreement was created to ensure that the global average temperature does not raise more than 2 °C above preindustrial levels.<sup>1</sup> Another facet of the agreement was focused on pursuing new technologies and methods to limit the temperature increase to 1.5 °C above pre-industrial levels.<sup>1</sup> The report of the Intergovernmental Panel on Climate Change (IPCC) concluded that in order to limit warming to 1.5 °C negative emissions technologies (NETs) must be implemented by the middle of this century.<sup>2</sup> A large portfolio of NETs are available; including coastal blue carbon, terrestrial carbon

# Enhanced precipitation of magnesium carbonates using carbonic anhydrase<sup>†</sup>‡

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Carbonate precipitation, as part of the carbon dioxide (CO<sub>2</sub>) mineralization process, is generally regarded as a high-temperature, high-pressure, and high-purity CO<sub>2</sub> process. Typical conditions consist of temperatures around 120 °C and a pressure of 100 bar of pure CO<sub>2</sub>, making the process costly. A major challenge facing carbonate precipitation is performing the reaction at low temperatures and low partial pressures of CO<sub>2</sub> (p<sub>CO2</sub>) such as 25 °C and CO<sub>2</sub> flue gas concentration. In this work, we investigated the effect of carbonic anhydrase (CA) to favor magnesium (Mg) carbonate precipitation at low temperatures and low  $p_{CO_2}$ . CA is an enzyme that accelerates CO<sub>2</sub> hydration promoting its conversion into HCO<sub>3</sub><sup>-</sup> and then CO3<sup>2-</sup>. This increases supersaturation with respect to Mg-carbonates. A geochemical model was implemented and used to identify supersaturated conditions with respect to Mg-carbonates. Tests were run at 25, 40, and 50 °C and at 1 bar of either pure CO<sub>2</sub> or 10 vol% CO<sub>2</sub> and 90 vol% N<sub>2</sub>. The concentration of 10 vol% CO2 was chosen to resemble CO2 concentration in flue gas. In selected tests, the CA enzyme was added directly as bovine CA or through microalgae (Scenedesmus obliguus). Experiments were run for 48 hours; 24 hours to reach equilibrium, then another 24 hours until the supersaturated conditions were established. After 48 hours the experiments were interrupted and the solids were characterized. Results show that the addition of CA, either directly or through Scenedesmus obliquus, enhances Mg-carbonate precipitation. Regardless of the temperature, the precipitates were made entirely of nesquehonite (MgCO<sub>3</sub>-3H<sub>2</sub>O) when pure CO<sub>2</sub> was used. Otherwise, a solid solution containing brucite (Mg (OH)<sub>2</sub>) and MgCO<sub>3</sub>-3H<sub>2</sub>O was formed. Overall, these findings suggest that CA can promote carbonate precipitation at low temperatures, pressures, and CO<sub>2</sub> purity. The enzyme is effective when added directly or supplied through microalgae, opening up the possibility for a CO<sub>2</sub> mineralization process to be implemented directly at a combustion plant as a  $CO_2$  storage option without preliminary  $CO_2$  capture.

removal and sequestration, bioenergy with carbon capture and sequestration, direct air capture, geologic sequestration, and carbon dioxide  $(CO_2)$  mineralization (aka mineral carbonation).

 $CO_2$  mineralization is an attractive NET solution because it has the potential to permanently sequester  $CO_2$  into stable minerals without long-term monitoring and liability issues. Conventional  $CO_2$  mineralization is an *ex situ* technology that relies on the dissolution of silicates (*e.g.*, olivine, serpentinite, and wollastonite) within mafic and ultra-mafic rocks. This technology also relies on the precipitation of carbonates to fix carbon into stable carbonate crystals which can be reused in construction.<sup>3–5</sup> In contrast to the emerging alternative option of *in situ*  $CO_2$  mineralization,<sup>6,7</sup> *ex situ*  $CO_2$  mineralization has the advantage of being an above-ground technology. Unlike *in situ*  $CO_2$  mineralization, above-ground technology like *ex situ*  $CO_2$  mineralization is beneficial because it can be monitored and controlled. However, the process typically relies on material pre-processing (grinding and milling), high tempera-



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<sup>&</sup>lt;sup>‡</sup>Unclassified – distribution a: approved for public release; distribution is unlimited.

tures, and high partial pressures of  $CO_2$  ( $p_{CO_2}$ ).<sup>8–11</sup> These conditions are strenuous, making the process costly.<sup>12</sup>

To offset this cost several authors have proposed alternative sources of alkalinity, which are more reactive, such as industrial waste, mine tailing, and brines (including salt lake brines, mine drainage, seawater, and desalination brines).<sup>13-18</sup> With these alternative sources CO2 mineralization can be carried out at low temperatures and low p<sub>CO2</sub>. Brines are particularly interesting because alkaline earth elements (e.g., Ca, Mg, and Na) are readily accessible for precipitation. The accessibility of these elements means that neither rock pre-processing nor rock dissolution are required. Additionally, the concentrations of these elements can be significant, making lower temperatures and  $p_{CO_2}$  possible. However, the precipitation of stable carbonates (such as magnesite and calcite) at low temperatures still remains a challenge,<sup>19,20</sup> and new approaches to catalyze the reaction are being investigated. For instance, the addition of carbonate seeds,<sup>21</sup> organic additives (e.g., ferric EDTA),<sup>22</sup> polystyrene microspheres,<sup>23</sup> microorganisms (such as microalgae),<sup>24,25</sup> and enzymes(such as carbonic anhydrase (CA))<sup>25–27</sup> have been proposed. Microalgae excrete extracellular CA which accelerates CO<sub>2</sub> hydration, promoting its conversion into HCO3<sup>-</sup> and CO3<sup>2-</sup> ions and favoring carbonate precipitation. Microalgae and cyanobacteria (e.g., Scenedesmus obliquus, Chlorella vulgaris, and Anabaena variabilis) produce CA naturally<sup>28,29</sup> through a carbon concentrating mechanism that allows them to grow fast at low  $p_{CO_2}$  concentrations. In living organisms CA has been shown to be an integral component of biogenic Ca-carbonate formation.30 As of yet, the effect of microalgae on carbonate precipitation from brines at low temperatures and low p<sub>CO<sub>2</sub></sub> has not been studied.

In this paper, we present an experimental work on the precipitation of Mg-carbonates at low temperatures and  $p_{CO_2}$  in the presence of both bovine CA and *Scenedesmus obliquus*. Geochemical simulations were carried out to determine supersaturated conditions with respect to Mg-carbonates. Through systematic experiments with consistently varied conditions, we illustrate the mineralization of solid products characterized using X-ray diffraction and electron microscopy.

## 2 Materials and methods

#### 2.1 Set-up

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Precipitation experiments were carried out with a continuously stirred batch reactor (Series 5100 Glass Reactor, Parr Instrument, U.S.A.), which we employed in an earlier paper<sup>31</sup> and upgraded for this work. A schematic of the set-up is shown in Fig. 1 and a photo is reported in Fig. 1 of the ESI† document. The reactor consists of a double-jacket glass vessel connected to a cylinder of  $CO_2$  (99.99% purity, Welding Supply Company, U.S.A.) and a temperature controlling system (HE Heating Circulator, Julabo, U.S.A.). During each experiment the temperature and pressure were continuously measured and digitally recorded with online sensors (Parr 4848 Reactor Controller, Parr Instrument Company, U.S.A.) placed inside



Fig. 1 Schematic of the set-up used for the experiments in this work. The light was used in the presence of microalgae.

the reactor. A high-pressure pipette (Parr Liquid Charging Pipette, Parr Instrument Company, U.S.A.) was used to inject a solution into the vessel after stabilization (explained more in detail in section 2.2). The pipette was filled with a solution of given composition and pressurized with nitrogen gas (N<sub>2</sub>, 99.99% purity, Welding Supply. U.S.A.). The pipette was pressurized at a slightly higher pressure than that in the reactor in order to overcome the resistance of the  $CO_2$  rich atmosphere (approximately 1 bar).

#### 2.2 Experiments

Precipitation experiments were carried out using the set-up described in section 2.1. First, a 400 mL solution was prepared using ultrapurified water (Milli-Q, U.S.A.) and sodium hydroxide (NaOH, 98.9% purity, Sigma-Aldrich) at a concentration of 1.5 molar. Then, the reactor was stabilized for 24 hours at a temperature and CO<sub>2</sub> partial pressure chosen on the basis of the operating conditions to be tested while stirring at a medium speed. The total pressure inside the reactor was always set equal to  $1 \pm 0.2$  bar, but the partial pressure of CO<sub>2</sub> changed between 1 and 0.1 bar. This was established by using a cylinder of simulated flue gas or pure CO<sub>2</sub>. In the cylinder with simulated flue gas, the chemical make-up was 10 vol% CO2 and 90 vol% N2. Additives were used in some experiments and they were placed into the initial NaOH solution. The additives were either pure bovine CA or microalgae (Scenedesmus obliquus). The concentrations of the additives were varied throughout the experiments. The activity of the CA was measured before and after the stabilization phase. After the initial solution was in the reactor for 24 hours, a 30 mL solution prepared using ultrapurified water and magnesium chloride (MgCl<sub>2</sub>-6H<sub>2</sub>O, Sigma-Aldrich) at a concentration between 0.5 and 3 molar MgCl<sub>2</sub>-6H<sub>2</sub>O was injected into the reactor

using the high-pressure pipette. The composition of the solution (concentration of Na and Mg) was selected to represent a deep surface brine as an alternative to the conventional approach in  $CO_2$  mineralization based on mafic/ultramafic rock dissolution.

After this injection, the mixture was maintained at the selected temperature and pressure conditions for 24 hours. Upon completion, the stirring was stopped while the reactor depressurized. The solids were allowed to settle, and then were collected via filtration. Before characterization the solids were dried at room temperature and pressure conditions. In two experiments, namely exp. 26 and 27 (reported in Table 1 below), the time was altered to observe how the time affected the results. During the experiments where freshwater microalgae was added to the system, the reactor was continuously illuminated by 4 LED lights (40 W, 36 LED, 25 cm in length and 2 cm in width). These lights were equally distributed around the vessel (Fig. 1) to achieve 98  $\mu$ mol (m<sup>2</sup> s)<sup>-1</sup> of LED light intensity. The rationale behind the lighting is to maintain microalgae activity and promote the excretion of CA from the cells into the solution.

#### 2.3 Selected enzyme and microorganism

The additives consist of pure lyophilized powder enzyme CA from bovine erythrocytes (Sigma-Aldrich. U.S.A.) and freshwater green microalga (*Scenedesmus obliquus* ATCC® 11477) (Fig. 2). Pure CA was added directly to the initial solution. A microalga culture was grown for 7 days in a growth chamber at 25 °C and under continuous shaking at 120 rpm. The growth medium used was BG-11, whose recipe is described in an



Fig. 2 *Scenedesmus obliquus* images performed using: (a) fluorescence microscopy; and (b) electroscanning microscopy.

 Table 1
 Operating conditions applied during the experiments and solid products

Exp.	<i>T</i> , °C	p <sub>CO2</sub> , bar	$Na^+$ , mol $L^{-1}$	$Mg^{2+}$ , mol $L^{-1}$	Type of additive	$c_{ m add}, { m g \ L^{-1}}$	$a_{\rm CA},\mu { m L}^{-1}$	Type of product	Product mass, g
1	25	1	1.5	1.5	_	_	_	_	_
2	25	1	1.5	2.0	_	_	_	_	_
3	25	1	1.5	2.5	_	_	_	_	_
4	25	1	1.5	3.0	_	_	_	_	_
16	27	1	1.5	2.0	Algae	0.20	171.9	Nesquehonite	3.87
15	28	1	1.5	2.5	Algae	0.20	165.4	Nesquehonite	2.09
14	29	1	1.5	3.0	Algae	0.21	170.3	Nesquehonite	8.91
14R	28	1	1.5	3.0	Algae	0.23	184.2	Nesquehonite	7.20
20	26	0.1	1.5	2.5	Algae	0.21	173.5	Brucite	19.7
19	27	0.1	1.5	3	Algae	0.14	26.4	Nesquehonite	22.6
19R1	25	0.1	1.5	3	Algae	0.14	145.3	Nesquehonite	20.3
19R2	25	0.1	1.5	3	Algae	0.22	149.7	Nesquehonite	21.7
19R3	25	0.1	1.5	3	Algae	0.21	153.5	Nesquehonite	22.4
19R4	25	0.1	1.5	3	Algae	0.40	354.1	Nesquehonite	20.6
23	25	0.1	1.5	3	Bovine CA	0.04	36.1	Nesquehonite	27.9
29	25	0.1	1.5	3	Bovine CA	0.10	112.7	Nesquehonite	25.3
18	40	1	1.5	0.5	_	—	_		
5	40	1	1.5	1.0	_	—	_	—	
5R	40	1	1.5	1.0	_	—	_	—	
6	40	1	1.5	1.5	_	—	_	Nesquehonite	1.1
7	40	1	1.5	2.0	_	—	_	Nesquehonite	4.3
8	40	1	1.5	2.5	_	_	_	Nesquehonite	7.1
17	40	1	1.5	1.5	Algae	0.20	160.4	Nesquehonite	2.6
9	40	0.1	1.5	0.5	_		_	Brucite	1.2
9R	40	0.1	1.5	0.5	—	—	—	Brucite	1.6
10R	40	0.1	1.5	1.0	—	—	—	Brucite	6.5
11	40	0.1	1.5	1.5	—	—	—	Brucite	16.7
11R	40	0.1	1.5	1.5	—	—	—	Brucite	16.7
13R	40	0.1	1.5	2.5	—		—	Brucite	14.8
21	40	0.1	1.5	3.0	—	—		Brucite	15.4
$26^a$	40	0.1	1.5	2.5	—	—	—	Brucite	12.1
27 <sup>b</sup>	40	0.1	1.5	2.5	—	—	—	Brucite	18.2
24	40	0.1	1.5	2.5	Bovine CA	0.04	36.1	Brucite	14.3
25	40	0.1	1.5	2.5	Algae	0.35	287.3	Nesquehonite	10.0
25R1	40	0.1	1.5	2.5	Algae	0.20	147.9	Nesquehonite	11.0
25R2	40	0.1	1.5	2.5	Algae	0.40	362.8	Nesquehonite	12.4

Symbols indicate: R#, repetition. <sup>*a*</sup> 2 days of stabilization and 4 days of precipitation for a total of 144 hours. <sup>*b*</sup> 1 day of stabilization and 8 days of precipitation for a total of 216 hours.

earlier paper.<sup>31</sup> The culture was centrifuged and the pellet washed with ultrapurified water 5 times before the cells were added into the NaOH solution. This washing process removes all nutrients from the supernatant growth medium and avoid possible interference in the mineralization process.

#### 2.4 Characterization of the additives

The tested concentrations for these additives  $(c_{add})$  ranged from 0.14 to 0.40 g  $L^{-1}$  for Scenedesmus obliquus and 0.035 to  $0.2 \text{ g L}^{-1}$  for CA. The pure CA values were chosen according to Power et al.<sup>27</sup> and Fuchs et al.<sup>32</sup> while Scenedesmus obliquus values were increased incrementally. The total suspended solids and the enzymatic activity of the culture was calculated for each experiments using microorganisms. Enzymatic activity of pure CA as well as from microalgae was assayed spectrophotometrically using a method adapted for determining p-nitrophenyl acetate (p-NPA) activity.<sup>33</sup> Briefly, this assay is based on the enzyme mediated hydrolysis of p-NPA. CA has the same active site for the hydration reaction of CO<sub>2</sub> as that for the hydrolysis reaction of p-NPA. The latter reaction is much slower than the former, making it suitable for CA activity determination. The component p-NPA is hydrolyzed by the esterase activity of CA and the product of bright-yellow color can be spectrophotometrically determined by analyzing the absorbance at 400 nm.

#### 2.5 Characterization of the solid products

The solids were characterized by scanning electron microscope (SEM, Auriga 40, Zeiss) and X-ray diffraction analysis (XRD, Ultima IV, Rigaku). The XRD measurements of our products were compared with reference spectra of carbonates.<sup>34</sup>

## 3 Geochemical model

The open-source software PHREEQC.5  $^{35}$  with the Pitzer database was used to determine the chemical composition of the system and design the experiments. The chemical reactions for the precipitation of the magnesium carbonate minerals such as artinite (Mg<sub>2</sub>(CO<sub>3</sub>)(H<sub>2</sub>O)-3H<sub>2</sub>O), nesquehonite (MgCO<sub>3</sub>-3H<sub>2</sub>O), hydromagnesite (Mg<sub>5</sub>(CO<sub>3</sub>)4(OH)<sub>2</sub>-4H<sub>2</sub>O), and magnesite (MgCO<sub>3</sub>) are, respectively,

$$Mg_2(CO_3)(OH)_2-3H_2O_s + 3H^+ \leftrightarrow 2Mg^{2+} + HCO_3^- + 5H_2O,$$
 (1)

$$MgCO_3-3H_2O_s + H^+ \leftrightarrow Mg^{2+} + HCO_3^- + 3H_2O, \quad (2)$$

$$Mg_{5}(CO_{3})_{4}(OH)_{2}-4H_{2}O_{8}+6H^{+}$$
  

$$\leftrightarrow 5Mg^{2+}+4HCO_{3}^{-}+6H_{2}O,$$
(3)

$$MgCO_{3,s} + H^+ \ \leftrightarrow \ Mg^{2+} + HCO_3^{-}, \eqno(4)$$

and the dominant reactions (*i.e.*, the compounds have a concentration larger than  $10^{-6}$  m) of the aqueous speciation are,

$$Mg(OH)_{2a} \leftrightarrow Mg^{2+} + 2OH^{-},$$
 (5)

$$CO_{2,g} \leftrightarrow CO_{2,a},$$
 (6)

$$CO_{2,a} + H_2O \leftrightarrow H_2CO_3,$$
 (7)

$$H_2CO_3 \leftrightarrow HCO_3^- + H^+,$$
 (8)

$$\text{HCO}_3^- \leftrightarrow \text{CO}_3^{2-} + \text{H}^+.$$
 (9)

Composition diagrams were created at 25 and 40 °C and  $p_{CO_2}$  of 1 and 0.1 bar by running PHREEQC from MATLAB<sup>36</sup> through the IPhreeqc module. The diagrams are reported as a function of Na<sup>+</sup> and Mg<sup>2+</sup> concentrations in Fig. 3 and 4. They show the contours of pH and saturation indexes (SI) with respect to magnesium carbonate minerals, whose reactions are given in eqn (1) through (4),

$$\mathrm{SI}_{\mathrm{art}} = \left(\frac{a_{\mathrm{Mg}^{2+}}^{2}a_{\mathrm{HCO}_{3}^{-}}a_{\mathrm{H}_{2}\mathrm{O}^{5}}}{a_{\mathrm{H}^{+}}^{3}K_{\mathrm{sp,art}}}\right), \tag{10}$$

$$SI_{nes} = \left(\frac{a_{Mg^{2+}}a_{HCO_{3}}a_{H_{2}O}^{3}}{a_{H^{+}}K_{sp,nes}}\right),$$
(11)

$$\mathrm{SI}_{\mathrm{hyd}} = \left(\frac{a_{\mathrm{Mg}^{2+}} \, {}^{5}a_{\mathrm{HCO}_{3-}} \, {}^{4}a_{\mathrm{H}_{2}\mathrm{O}}^{6}}{a_{\mathrm{H}^{+}} \, {}^{6}K_{\mathrm{sp,hyd}}}\right),\tag{12}$$

$$\mathrm{SI}_{\mathrm{mag}} = \left(\frac{a_{\mathrm{Mg}^{2+}}a_{\mathrm{HCO}_{3}^{-}}}{a_{\mathrm{H}^{+}}K_{\mathrm{sp,mag}}}\right),\tag{13}$$

where *a* and  $K_{sp}$  are the activities and the solubility products of the subscript species and solid phases, respectively. In all cases, the system is under saturated with respect to MgOH<sub>2</sub> as it is possible to see in Fig. 2 in the ESI<sup>†</sup> document.

## 4 Results and discussion

In this section, the results of this work are presented and discussed. Table 1 lists the operating conditions applied during the experiments together with the type of solid products obtained and the total mass of the solids determined after washing and drying at room conditions. In Fig. 5, the temperature and pressure profiles recorded during two representative experiments are presented. As it is clear to see in the graph, the pressure is temporarily affected by solution injection with the high-pressure pipette but it re-establishes around the set value of 1 bar soon afterwards. The temperature is not affected by the injection or the precipitation reaction. This is the case for all the experiments.

#### 4.1 Tests run at 25 °C

In this section, the mineralogical phases and morphologies of the products obtained at the end of the experiments performed at 25 °C are reported and discussed. In these tests the partial pressure of  $CO_2$  was set equal to either 1 bar of pure  $CO_2$  or 0.1 bar of  $CO_2$  flue gas concentration. The tests were run with no additives, by adding microalgae, or by adding bovine CA to the system. No precipitation was observed in exp. 1 through 4 (no additives, 1 bar pure  $CO_2$ ) despite the supersaturated conditions (see Fig. 3 panels a through d) with respect to the hydrated metastable phase of MgCO<sub>3</sub>-3H<sub>2</sub>O.



Fig. 3 Contours of pH (red) and SI (blue) calculated at 25 °C at p<sub>CO2</sub> equal to (a-d) 1 and (e-h) 0.1 bar. The dots indicate the applied experimental conditions.

Previous works show precipitation at similar conditions.<sup>31,37</sup> However, in those cases the reactor was slightly depressurized in order to inject the solution containing Mg<sup>2+</sup>. Depressurization,

even for a few seconds, temporarily reduces the  $CO_2$  concentration in the atmosphere leading to an increase in pH that may favor precipitation. Degassing of  $CO_2$  has even been used in



Fig. 4 Contours of pH (red) and SI (blue) calculated at 40 °C at p<sub>CO2</sub> equal to (a–d) 1 and (e–h) 0.1 bar. The dots indicate the applied experimental conditions.

earlier works to favor  $MgCO_3$ - $3H_2O$  precipitation.<sup>25</sup> In our work, we used a high-pressure pipette that allowed us to avoid depressurization of the reactor and disturbance to the aqueous system.

No experiments were run at 25  $^{\circ}$ C and 0.1 bar without additives because the system was undersaturated with respect to MgCO<sub>3</sub>-3H<sub>2</sub>O at those conditions (see Fig. 3 panels e through



Fig. 5 Temperature and pressure measurements recorded during (a) exp. 20 and (b) exp. 13.

h). When an additive was used at  $p_{CO_2}$  of 1 bar the experiments (exp. 16 to 14 in Table 1) consistently produced seemingly pure MgCO<sub>3</sub>-3H<sub>2</sub>O, as shown in Fig. 6. The temperatures of exp. 16 through 14 were slightly higher than 25 °C due to the lights used during the algae experiments. This increase was unintentional and small enough to be considered negligible. Since the only condition that significantly changed between exp. 1 through 4 and exp. 14 through 16 was the addition of algae, we can conclude that algae does aid in the formation of carbonate crystals and in the mineralization of CO<sub>2</sub>.

Similar results were obtained when an additive was added at p<sub>CO2</sub> of 0.1 bar in exp. 19, 23, and 29 in Table 1. In the SEM-EDS and XRD analyses reported in Fig. 7 it can be observed that a solid solution made of Mg(OH)<sub>2</sub> and Mg-carbonates formed. In all of the XRD analyses, an amorphous phase dominates in the product as indicated by the smooth bump between 20 and 40  $2\Theta$ . The peaks in the XRD analyses are characteristic of solids such as Mg(OH)<sub>2</sub>, MgCO<sub>3</sub>-3H<sub>2</sub>O, and MgCO<sub>3</sub>. The geochemical simulations (Fig. 2 in the ESI<sup>†</sup> document) indicate that the system is undersaturated with respect to  $Mg(OH)_2$ , however the addition of additives may change the solution composition, which is not considered in the simulation. This change favors the precipitation of Mg (OH)<sub>2</sub> as well as Mg-carbonate minerals. The precipitation of  $Mg(OH)_2$  should be avoided as it competes with the desired products, the Mg-carbonate minerals. Though algae do help



**Fig. 6** Characterization of the solid products obtained from experiments (a and b) 16; (c and d) 15; and (e and f) 14 run at ~25 °C and 1 bar of  $p_{CO_{\circ}}$  in the presence of microalgae *S. obliquus*.



**Fig. 7** Characterization of the solid products obtained from experiments (a–c) 19; (d–f) 19R1; (g–i) 19R4; and (j–l) 23 run at 25 °C and 0.1 bar of  $p_{CO_2}$  and in the presence of either microalgae *S. obliquus* or bovine CA as reported in Table 1.

the experiment create a pure carbonate crystal at 1 bar, they lead to solid mixtures at 0.1 bar, as shown in Fig. 7. It is important to point out that the EDS analysis of the solid product does not show any nitrogen on the surface, suggesting the additives are not in the precipitates.

Bovine CA was not tested at 1 bar of  $CO_2$ , it was only tested at 0.1 bar. It is however expected that bovine CA would have aided Mg-carbonate precipitation similarly to microalgae. As mentioned above, CA is an enzyme produced by microorganisms such as algae which favors the hydration of  $CO_2$  increasing the concentration of  $HCO_3^-$  ions.

Inhibiting factors in these experiments could have been either temperature or  $p_{CO_2}$ . Higher temperature or  $p_{CO_2}$  may have limited Mg(OH)<sub>2</sub> precipitation and favor Mg-carbonate precipitation, instead. With this in mind, we ran tests at a higher temperature and the results are shown in section 4.2.

#### 4.2 Tests run at 40 and 50 °C

In this section the experiments performed at temperatures of 40 and 50 °C are reported and discussed. Similarly to the section above, the partial pressure of  $CO_2$  was set equal to either 1 bar or 0.1 bar. The tests were run with no additives, by adding microalgae, or by adding bovine CA to the system. In experiments run without additives, at 1 bar, and 40 °C (exp. 5 through 8, and 18 in Table 1) MgCO<sub>3</sub>-3H<sub>2</sub>O only precipitated in exp. 6 where the concentration of Mg<sup>2+</sup> was 1.5 molar. In experts the SEM images and the XRD measurements of these experiments showing that the applied conditions were able to produce seemingly pure nesquehonite.

In exp. 17 the same conditions as exp. 6 were applied, except algae were added. The solid product that formed was  $MgCO_3$ - $3H_2O$ , as shown in Fig. 9. The amount of product collected in exp. 17 was twice that collected at the end of exp. 6. This indicates that algae affect the rate at which the solid formed leading to double the amount of the solid in the same reaction time, namely 24 hours.



Fig. 8 Characterization of the solid products obtained from experiments (a and b) 6; (c and d) 7; and (e and f) 8 run at 40 °C and 1 bar of  $P_{CO_2}$ .



Fig. 9 Characterization of the solid product obtained from experiments 17 run at 40 °C and 1 bar of  $p_{CO_2}$  in the presence of algae. (a) SEM image; (b) EDS analysis; and (c) XRD measurements.



Fig. 10 Characterization of the solid products obtained from experiments (a-c) 9; (d-g) 11; and (h and i) 13R run at 40 °C and 0.1 bar of  $P_{CO_2}$ .

With the aim to investigate the effect of  $CO_2$  partial pressure,  $p_{CO_2}$  was reduced to 0.1 bar. At 0.1 bar, the results show an amorphous phase within the solid products which may resemble  $Mg(OH)_2$  (Fig. 10). Even extending the precipitation reaction up to 8 days (exp. 27 In Table 1) no Mg-carbonate formed. In these tests, the solution was undersaturated with respect to  $MgCO_3$ - $3H_2O$  (Fig. 4 part f) suggesting that supersaturation with respect to  $MgCO_3$ - $3H_2O$  is critical for the formation of Mg-carbonates at these conditions.

Continuing tests at  $p_{CO_2}$  of 0.1 bar, additives were added to the solution (exp. 24 though 25R2). Exp. 24 did not yield carbonates, possibly due to the small concentration of the enzyme added. However, in the presence of algae, solids formed and they resembled a solid made of Mg(OH)<sub>2</sub>, MgCO<sub>3</sub>-3H<sub>2</sub>O, and MgCO<sub>3</sub> (Fig. 11). Similar to the results at 25 °C, the



**Fig. 11** Characterization of the solid products obtained from experiments (a-c) 25; (d-g) 25R1; and (h and i) 25R2 run at 40 °C and 0.1 bar of  $p_{CO_2}$  and in the presence of microalgae *S. obliquus* and bovine CA as reported in Table 1.

EDS analyses of the solid products do not show any nitrogen, indicating no inclusions of additives in the precipitates. As the enzyme activity of microalgae was increased from 287.3 to  $362.8 \ \mu \ L^{-1}$ , neither the amount nor the characteristics of the solid products changed.

The results from the experiments at 40 °C show that the precipitation reaction of Mg-carbonates is dependent on the  $p_{CO_2}$  and concentration of the enzyme. Increasing  $p_{CO_2}$  or enzyme concentration favor the precipitation of Mg-carbonates. But raising the temperature to 50 °C (exp. 28 in Table 1) did not favor precipitation. This is because solubility of  $CO_2$  decreases with temperature lowering the supersaturation ratio values with respect to Mg-carbonates. The enzyme was not added at this temperature because of more rapid decay.

#### 4.3 Stability of carbonic anhydrase during the experiments

We analyzed the stability of the enzyme for all experiments in which additives were used. The enzyme activity was measured at the beginning of the experiment, after the 24 h stabilization period, and at the end of the experiment. The development of the activity over time is shown in Fig. 12. In these diagrams, it is possible to see that during the stabilization period there is not significant change of the activity, although a slight decrease can be observed. This confirmed that the enzyme is stable during the 24 h stabilization period, confirming that the determination of the enzyme activity at the beginning of a test is representative of that when the supersaturated conditions are established. However, after the precipitation reaction occurs, the activity decreases of 60-80%. The largest drop in CA activity corresponds to the largest initial CA in the system. There might be several reasons that explain this. Pure CA requires cold temperatures (around 4 °C) for storage in order to maintain its integrity. Since the experiments were run at higher temperatures (either 25 or 40 °C) CA might have undergone thermal degradation. If the temperature were the cause, then the CA should have degraded more at 40 °C than 25 °C. However, the trends of the activity in the two parts of Fig. 12 are very similar. Alternatively, precipitation may have removed CA either through inclusion in the solid structure or sweeping. However, EDS analyses do not show any nitrogen element on the surface of the precipitates. A more likely explanation is that CA degrades over



**Fig. 12** Evolution of the activity of CA within the microalgae at (a) 25 °C and (b) 40 °C. CA was added to the system through different amounts of microalgae expressed as total suspended solids (TSS).

time. We ran preliminary tests to determine the correlation between total solid concentration and CA activity (Fig. 3 ESI<sup>†</sup> document). In these tests, we noticed that the same sample of pure CA 24 hours apart showed a significant reduction in CA activity. Given that the precipitation experiments were ran for 48 hours, the enzyme activity might have dropped. The combination of temperature, hydration, and decrease of pH upon precipitation may have accelerated this decay.

#### 5 Conclusions

In this paper, we present a study on the effect of temperature, partial pressure of CO<sub>2</sub> (p<sub>CO2</sub>), and carbonic anhydrase enzyme on Mg-carbonate precipitation. The enzyme was added directly as bovine CA or through microalgae (Scenedesmus obliquus). A geochemical model was implemented and used to select the operating laboratory conditions. Systematic tests at 25 and 40 °C and total pressure of 1 bar were run. The values of  $p_{CO_2}$ were varied between 1 and 0.1 bar to simulate pure CO2 and flue gas. The results show that the addition of CA through microalgae is effective in promoting the precipitation of a hydrated Mg-carbonate (nesquehonite,MgCO<sub>3</sub>-3H<sub>2</sub>O). This is due to the effect of the enzyme in promoting the conversion of CO2 into HCO3<sup>-</sup> and CO3<sup>2-</sup> which increases the supersaturation ratio with respect to carbonate solid phases. However, the activity of the enzyme seems to decrease during the experiment and further investigation on its decay is needed.

Overall, this study demonstrates that the microalgae *Scenedesmus obliquus* can be used to enhance the precipitation of a hydrated Mg-carbonate mineral from a brine at low temperature and flue gas partial pressure  $(p_{CO_2})$ . This opens up the possibility to implement the direct conversion of  $CO_2$  into carbonate minerals as a negative emission technology (NET) at a combustion plant, without prior capture.

## Author contributions

Brian Caulfield: Data curation, investigation, methodology, and writing – original draft. Juliana Abraham: Data curation, investigation, methodology, writing – original draft, and supervision. Christos Christodoulatos: Funding acquisition and writing – original draft. Valentina Prigiobbe: Conceptualization, data curation, formal analysis, funding acquisition, methodology, resources software, supervision, visualization, and writing – original draft.

### Conflicts of interest

There are no conflicts to declare.

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## References

- 1 UNFCCC, 2015.
- 2 H. de Coninck, A. Revi, M. Babiker, P. Bertoldi, M. Buckeridge, A. Cartwright, W. Dong, J. Ford, S. Fuss, J.-C. Hourcade, D. Ley, R. Mechler, P. Newman, A. Revokatova, S. Schultz, L. Steg and T. Sugiyama, Strengthening and Implementing the Global Response. In: Global Warming of 1.5 °C. An IPCC Special Report on the impacts of global warming of 1.5 °C above pre-industrial levels and related global greenhouse gas emission pathways, in *The context of strengthening the global response to the threat of climate change, sustainable development, and efforts to eradicate poverty Masson-Delmotte*, ed. V. P. Zhai, H.-O. Pörtner, D. Roberts, J. Skea, P. R. Shukla, A. Pirani, W. Moufouma-Okia, C. Péan, R. Pidcock, S. Connors, J. B. R. Matthews, Y. Chen, X. Zhou, M. I. Gomis, E. Lonnoy, T. Maycock, M. Tignor and T. Waterfield, 2018.
- 3 B. Metz, O. Davidson, H. de Coninck, M. Loos and L. Meyer, *IPCC Special Report on Carbon Dioxide Capture and Storage; Intergovernmental Panel on Climate Change*, 2005.
- 4 IPCC, IPCC Special Report on Carbon Dioxide Capture and Storage, in *Prepared by Working Group III of the Intergovernmental Panel on Climate Change*, ed. B. Metz, O. Davidson, H. C. de Coninck, M. Loos and L. A. Meyer, Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 2005, p. 442.
- 5 S. Yadav and A. Mehra, *Environ. Sci. Pollut. Res.*, 2021, 28, 12202–12231.
- 6 J. M. Matter and P. B. Kelemen, Nat. Geosci., 2009, 2, 837-841.
- 7 S. K. White, F. A. Spane, H. T. Schaef, Q. R. S. Miller,
   M. D. White, J. A. Horner and B. P. McGrail, *Environ. Sci. Technol.*, 2020, 54, 14609–14616.
- 8 A. Sanna, M. Uibu, G. Caramanna, R. Kuusik and M. M. Maroto-Valer, *Chem. Soc. Rev.*, 2014, 43, 8049–8080.
- 9 M. I. Rashid, E. Benhelal, F. Farhang, T. K. Oliver, M. S. Rayson, G. F. Brent, M. Stockenhuber and E. M. Kennedy, *J. Cleaner Prod.*, 2019, 212, 151–161.
- 10 S. Hariharan and M. Mazzotti, Chem. Eng. J., 2017, 324, 397-413.
- 11 G. Gadikota, J. Matter, P. Kelemen and A.-h. A. Park, *Phys. Chem. Chem. Phys.*, 2014, **16**, 4679–4693.
- 12 A. Kirchofer, A. Brandt, S. Krevor, V. Prigiobbe and J. Wilcox, *Energy Environ. Sci.*, 2012, 5, 8631–8641.

- 13 J.-H. Bang, Y. Yoo, S.-W. Lee, K. Song and S. Chae, *Minerals*, 2017, 7, 11.
- 14 J.-H. Bang, S. C. Chae, S.-W. Lee, J.-W. Kim, K. Song, J. Kim and W. Kim, *J. CO2 Util.*, 2019, **33**, 427–433.
- 15 C. M. Woodall, X. Lu, G. Dipple and J. Wilcox, *Minerals*, 2021, **11**, 8.
- I. M. Power, C. Paulo, H. Long, J. A. Lockhart, A. R. Stubbs, D. French and R. Caldwell, *Environ. Sci. Technol.*, 2021, 55, 10056–10066.
- 17 Y. E. Chai, S. Chalouati, H. Fantucci and R. M. Santos, *Crystals*, 2021, **12**, 1584.
- 18 W. Liu, L. Teng, S. Rohani, Z. Qin, B. Zhao, C. C. Xu, S. Ren, Q. Liu and B. Liang, *Chem. Eng. J.*, 2021, 416, 129093.
- 19 V. Prigiobbe and M. Mazzotti, *Chem. Eng. J.*, 2013, 223, 755–763.
- 20 V. Prigiobbe, J. Environ. Chem. Eng., 2018, 6, 930-936.
- 21 S. Hariharan and M. Mazzotti, *Cryst. Growth Des.*, 2016, **17**, 317–327.
- 22 G. Montes-Hernandez, M. Bah and F. Renard, *J. CO2 Util.*, 2020, **35**, 272–276.
- 23 I. M. Power, P. A. Kenward, G. M. Dipple and M. Raudsepp, *Cryst. Growth Des.*, 2017, 17, 5652–5659.
- 24 I. M. Power, S. A. Wilson, D. P. Small, G. M. Dipple, W. Wan and G. Southam, *Environ. Sci. Technol.*, 2011, 45, 9061–9068.
- 25 T. K. Oliver, B. Z. Dlugogorski and E. M. Kennedy, *Miner. Eng.*, 2014, **61**, 113–120.
- 26 M. Zhang, L. Zhao, G. K. Li, C. Zhu, S. Dong, Z. Li, C. Tang, J. Ji and J. Chen, *Geomicrobiol. J.*, 2021, 38, 549– 560.
- 27 I. M. Power, A. L. Harrison, G. M. Dipple and G. Southam, Int. J. Greenhouse Gas Control, 2013, 16, 145–155.
- 28 M. Madhumanti, S. Khanra, O. N. Tiwari, K. Gayen and G. N. Halder, *Environ. Prog. Sustainable Energy*, 2016, 35, 1605–1615.
- 29 J. da Costa Ores, M. C. A. D. Amarante, S. S. Fernandes and S. J. Kalil, *Biocatal. Biotransform.*, 2016, 34, 57–65.
- 30 W. E. Müller, H. C. Schröder, U. Schlossmacher, M. Neufurth, W. Geurtsen, M. Korzhev and X. Wang, *FEBS Open Bio*, 2013, 3, 357–362.
- 31 Z. Ye, J. Abraham, C. Christodoulatos and V. Prigiobbe, *Energy Fuels*, 2019, **33**, 8843–8851.
- 32 W. Fuchs, F. Steger, J. Reich, D. Ribitsch, S. K.-M. Rittmann and G. Bochmann, *Catalysts*, 2021, **11**, 819.
- 33 Y. Pocker and J. T. Stone, *Biochemistry*, 1967, **6**, 668–678.
- 34 Highlights in Mineralogical Crystallography, ed.
   T. Armbruster and R. M. Danisi, W. De Gruyter, Berlin, Germany, 2016, pp. 213.
- 35 USGS, PHREEQC (Version 3)–A Computer Program for Speciation, Batch-Reaction, One-Dimensional Transport, and Inverse Geochemical Calculations, 2016.
- 36 *MATLAB, version (R2021a)*, The MathWorks Inc., Natick, Massachusetts, 2021.
- 37 M. Hänchen, V. Prigiobbe, R. Baciocchi and M. Mazzotti, *Chem. Eng. Sci.*, 2008, **63**, 1012–1028.