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Surface hydration for antifouling and bio-adhesion†

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Antifouling properties of materials play crucial roles in many important applications such as biomedical implants, marine antifouling coatings, biosensing, and membranes for separation. Poly(ethylene glycol) (or PEG) containing polymers and zwitterionic polymers have been shown to be excellent antifouling materials. It is believed that their outstanding antifouling activity comes from their strong surface hydration. On the other hand, it is difficult to develop underwater glues, although adhesives with strong adhesion in a dry environment are widely available. This is related to dehydration, which is important for adhesion for many cases while water is the enemy of adhesion. In this research, we applied sum frequency generation (SFG) vibrational spectroscopy to investigate buried interfaces between mussel adhesive plaques and a variety of materials including antifouling polymers and control samples, supplemented by studies on marine animal (mussel) behavior and adhesion measurements. It was found that PEG containing polymers and zwitterionic polymers have very strong surface hydration in an aqueous environment, which is the key for their excellent antifouling performance. Because of the strong surface hydration, mussels do not settle on these surfaces even after binding to the surfaces with rubber bands. For control samples, SFG results indicate that their surface hydration is much weaker, and therefore mussels can generate adhesives to displace water to cause dehydration at the interface. Because of the dehydration, mussels can foul on the surfaces of these control materials. Our experiments also showed that if mussels were forced to deposit adhesives onto the PEG containing polymers and zwitterionic polymers, interfacial dehydration did not occur. However, even with the strong interfacial hydration, strong adhesion between mussel adhesives and antifouling polymer surfaces was detected, showing that under certain circumstances, interfacial water could enhance the interfacial bio-adhesion.

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Introduction

Antifouling properties are important for many extensively used materials in various applications and devices such as biomedical implants, drug delivery vehicles, marine antifouling coatings, sensors, and membranes.^{1–4} Extensive research demonstrated that zwitterionic polymers and polymers

containing poly(ethylene glycol) components (referred to as “PEG materials” in this article) exhibit excellent antifouling activities.^{5–9} It is believed that the strong surface hydration of such polymers mediates their excellent antifouling performance.^{10–13} With strong surface hydration, it is difficult for biological media (*e.g.*, protein molecules, bacteria, marine organisms, *etc.*) to displace strongly bonded surface water molecules to stick to (or to foul on) the surfaces of zwitterionic polymers and PEG materials. In contrast, for non-antifouling materials, surface hydration is not as strong.^{12,13} Then biological media can displace or repel surface water molecules to dehydrate the surface to stick to the surface. It is reasonable to believe that strong interfacial hydration leads to antifouling, while interfacial dehydration results in fouling.

Similarly, almost all the man-made adhesives and glues work on a dry surface but fail to stick to a wet hydrated surface, because such adhesives cannot dehydrate the surface. The hard, dry adhesives of insects and geckos also stick well when dry and less well or not at all when substrates are underwater.^{14,15} Different from insects and geckos, a trip to the beach will find

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mussels, barnacles, tube worms, sea stars, oysters, and many more animals bonding to rocks that are quite wet. Presumably, marine biology has devised clever ways clearing water from surfaces to dehydrate the surface in order to create adhesive-substrate bonds.^{16–18} In addition to the understanding on the molecular mechanisms of antifouling, understanding on bio-adhesion in wet environments is equally important, which will greatly aid in the design and development of underwater glues for many important medical and other applications. We believe that bio-adhesion is also mediated by interfacial hydration/dehydration.

However, it is difficult to test the above “beliefs” or “hypothesis” on the relationships between “interfacial hydration/dehydration” and “antifouling/fouling” experimentally because it is challenging to study buried solid/liquid interfaces *in situ* nondestructively to probe interfacial hydration or examine buried solid/solid interfaces to test interfacial dehydration. Traditional surface sensitive techniques require high vacuum to operate. It is therefore impossible to use them to test surface structures in a liquid environment. Recently, sum frequency generation (SFG) vibrational spectroscopy has been developed into a powerful tool to probe buried solid/liquid and solid/solid interfaces *in situ* in real time at the molecular level.^{19–22} We have applied SFG to study the interfacial molecular behavior of proteins including adhesive proteins.^{23–26} Here in order to elucidate the effects of interfacial hydration/dehydration on antifouling and bio-adhesion, we probed buried interfaces between mussel adhesive plaques and a variety of antifouling and fouling materials using SFG, in combination with mussel behavior study and adhesion measurements. Our results demonstrated that strong hydration is the key to achieve excellent antifouling performance of a polymer material, and dehydration leads to fouling. Surprisingly, we also found that hydration can enhance bio-adhesion under certain circumstances.

Experimental

Surface preparation for mussel attachment and adhesion measurements

Fused silica and glass slides, 3 inch \times 1 inch \times 1 mm, were purchased from VWR. Aluminum T6061 sheets of 1/8 inch thickness were ordered from Farmers Copper (Galveston, TX). Poly(methyl methacrylate) (PMMA) and polystyrene (PS) sheets of 1/8 inch thickness were bought from United States Plastic Corporation (Lima, Ohio). Aluminum, PMMA and PS were cut into 3 inch \times 1 inch pieces. All substrates were cleaned with soap and rinsed with deionized water and ethanol. Aluminum, fused silica, and glass received an additional rinse of acetone. These substrates were suitable for mussel deposition and adhesion testing.

For each coating, two substrates (3 inch \times 1 inch) were held together with binder clips with one aluminum piece underneath for support (see ESI, Fig. S1†). Two rubber bands were used to hold each animal in place. Combining two substrates together allowed for more stability and a larger area for animals to deposit adhesives. When substrates are listed in the order of

surface energy, water contact angles from our own measurements and the literature were used for aluminum,²⁷ fused silica,²⁸ polysulfobetaine methacrylate (SBMA), oligo(ethylene glycol) methacrylate (OEGMA),¹⁰ PMMA,²⁹ and PS.³⁰

Surface preparation for SFG spectroscopy

When making thin films suitable for SFG spectroscopy, PMMA ($M_w = 75\,000$ g per mole), PS ($M_w = 280\,000$ g per mole), and toluene were purchased from Sigma-Aldrich. Thin films of PMMA and PS were prepared by spin coating a 1% (w/w) toluene solution on clean fused silica prisms at 3000 rpm for 40 s using a P-6000 spin coater (Speedline Technologies) followed by annealing at 95 °C for 24 h. The brushes of OEGMA and SBMA were prepared on fused silica prisms *via* atomic transfer radical polymerization (ATRP) according to a previous report.¹⁰ The thicknesses of the identical polymer brushes prepared on silicon wafers were measured to be 25–30 nm using an alpha-SE ellipsometer (J. A. Woollam).

Animal handling

One blue mussel (*Mytilus edulis*) was placed on each plate and held in place with two rubber bands. Mussels used were adults of ~6–8 cm length. Five mussels, one on each plate, were contained in a 10 gallon aquarium in 4 °C, 3.5 g/100 mL salinity water with constant aeration. Additional details of the laboratory aquarium system can be found in a previous report.³¹

Mussel plaque collection for SFG spectroscopy

Animals deposited adhesive plaques onto uncoated surfaces of fused silica and polymer coatings of PS, PMMA, SBMA, and OEGMA. For each type of surface, the substrates were adhered to a larger piece of glass with double sided tape or epoxy to provide a larger area for animals to deposit glue. One mussel was held in place on each surface with a rubber band. Animals were contained separately in a 10 gallon aquarium and exposed to surfaces for a minimum of 5 days. The adhesive was collected until several plaques were deposited onto each surface. For the surfaces of OEGMA and SBMA, animals and surfaces were placed on PVC stands to encourage plaque deposition on coatings rather than on the glass aquarium tank bottom.

Surface spectroscopy

Sum frequency generation (SFG) spectroscopy is a nonlinear optical spectroscopic method well suited to examine chemistry at interfaces.^{19–22} Selection rules allow observation of only species immediately at a surface or within an interface for which there is no inversion symmetry. For example, the available inversion symmetry of free water does not bring about an SFG signal whereas water bound at the interface with order does appear in spectra.^{19–22}

SFG spectroscopy was implemented according to the protocol reported previously.¹³ Visible and infrared (IR) input beams were overlapped spatially and temporally at the interfaces between mussel adhesive plaques and the substrates immersed in water, or the interfaces between the substrates and water without the



plaques. All SFG spectra were collected using the ssp (s-polarized sum frequency output, s-polarized visible input, and p-polarized IR input) polarization combination.³² More introduction to SFG spectroscopy can be found in the ESI.†

Adhesion data collection

Adhesion testing was carried out three days after placement of mussels and plates into aquaria. Animals on plates were removed from aquaria and their threads were cut leaving behind attached plaques and threads attached to surfaces. Photographs were taken of plaques to determine the area by digital image analysis. Removal force was collected on an Instron 5544 materials testing system. Each thread was pulled perpendicular to the surface until failure. Adhesion of plaques was calculated by dividing the removal force by plaque area. Additionally, adhesive plaque production (Table S1†) and material failure modes (Table S3†) were recorded. At least 10 animals were examined for each surface (fused silica, PMMA, PS, OEGMA, and SBMA) and 75 animals for aluminum controls. Adhesion was gathered in at least 2 rounds where each round contained five mussels on a separate plate per coating type.

Adhesion for the first round of coatings was collected over 6 consecutive days, testing each coating type independently. Aluminum controls were tested on the same day of each surface for consistency. A second round of adhesion testing was gathered over 6 consecutive days with controls. Animals deposited less plaques on the surfaces of PMMA, OEGMA and SBMA and

therefore a third round of adhesion testing was performed. Here, data were gathered over 4 consecutive days. During the second and third rounds of testing, animals on plates were placed on top of PVC stands to encourage plaque deposition onto test surfaces rather than the glass tanks.

Combining all rounds of adhesion testing, 10 animals were examined on each surface of fused silica and PS. Due to the additional round of testing, 15 animals were examined on each of PMMA, OEGMA, and SBMA surfaces. Aluminum controls were tested alongside all coatings thus resulting in the collection of more animals and plaques than on other surfaces.

Results and discussion

Mussel behavior

Mussels were placed atop a series of substrates ranging from high to low surface energies. The animals attached by depositing a flowing adhesive precursor that cures into a final plaque formation.³³ Control materials investigated in this study on which biofouling occurs included fused silica (water contact angle of $\sim 20^\circ$), PMMA ($\sim 71^\circ$), aluminum ($\sim 75^\circ$), and PS ($\sim 87^\circ$). Also included in this research were the antifouling zwitterionic polymer (grafted SBMA, $\sim 20^\circ$) and PEG material (grafted OEGMA, 42°).¹⁰ As discussed above, zwitterionic³⁴ and OEGMA³⁵ surfaces have gained a measure of fame for antifouling properties (*i.e.*, preventing adhesion) within biomedical as well as marine shipping contexts.

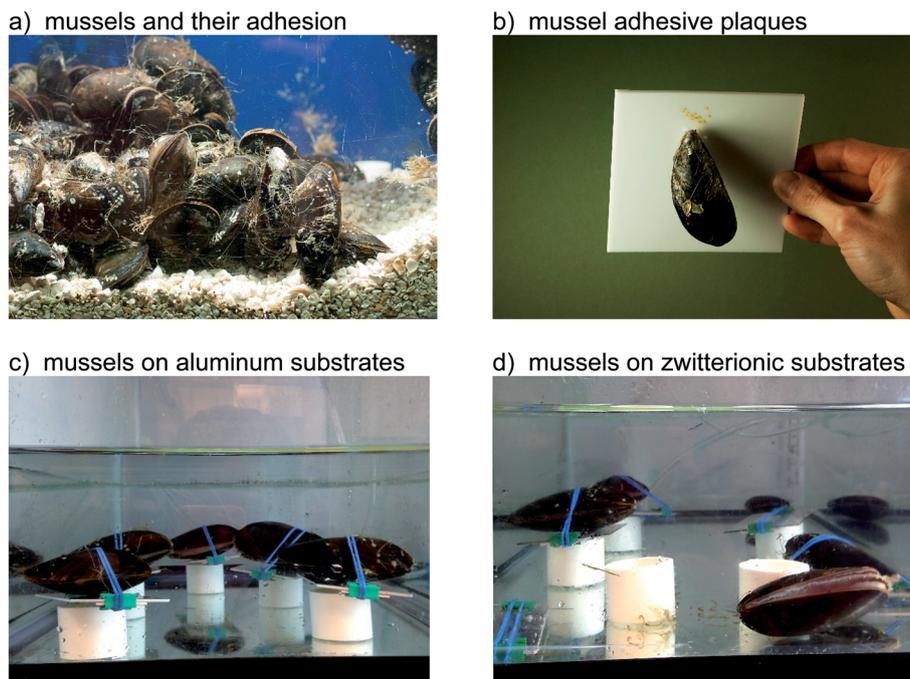


Fig. 1 Mussel adhesive and animal behavior on different surfaces. (a) Mussels using an adhesive for attaching to each other and the side of an aquarium tank. (b) A mussel bonding to a sheet of polytetrafluoroethylene (Teflon), showing the adhesive plaques and threads. (c) Photograph showing mussels two days after binding to aluminum substrates. Note how all animals remain attached. Top down views of mussels on the substrates are provided in Fig. S1 (ESI†). Each mussel and substrate is resting on a piece of plastic pipe to prevent the animal from placing its adhesive on the more stable aquarium bottom. (d) Mussels after two days on a zwitterionic SBMA surface. Most animals have moved off the substrates.



When placed onto surfaces, marine mussels attach by depositing their adhesive plaques (Fig. 1a and b).³⁶ Prior to settlement, the animals may move around in search of the most appealing location. Consequently, we used rubber bands to keep the animals in one place and force adhesion to a given substrate. For the aluminum (Fig. 1c), fused silica, PS, and PMMA substrates, mussels remained in place and attached with their plaques. The zwitterionic SBMA (Fig. 1d) and OEGMA surfaces, by contrast, resulted in different animal behavior. Most of the mussels were able to work their way off from these antifouling substrates. In order to understand the origin of this unusual action, we monitored the animals over several days. ESI Video 1† shows that the mollusks opened and closed their shells repeatedly. In doing so, they were able to wiggle free from their constraints, but only when on the zwitterionic SBMA and OEGMA surfaces.

The above observation of the mussel behavior demonstrated clearly that animals could easily settle on the four “control” or “fouling” surfaces including silica, PMMA, PS, and aluminum, but do not want to settle down on the two antifouling polymer SBMA and OEGMA surfaces. We believe that the drastically different animal behaviors on fouling and antifouling surfaces are caused by different surface hydration on these materials. Likely the control materials have weak surface hydration, while the antifouling SBMA and OEGMA have strong surface hydration, which will be investigated in more detail below.

Surface hydration in water

SFG spectra could be collected from the material/water interfaces *in situ* in real time to probe the molecular structure of a solid/liquid interface. Extensive research has been performed to investigate the interfacial water structure using SFG.^{37–45} Typically two broad peaks centered around 3200 and 3400 cm^{-1} could be detected, sometimes along with a relatively narrower peak at around 3700 cm^{-1} .^{37–45} It is agreed that the ~ 3700 peak is due to the free OH stretching, while debates still exist for the assignment for the other two peaks. It is generally believed that the 3200 cm^{-1} and 3400 cm^{-1} peaks come from the strongly and weakly (or loosely) hydrogen bonded water molecules at the interface.

We have collected SFG spectra from a variety of surfaces in water.^{11,46–51} It was found in our previous research that stronger surface hydration leads to better antifouling behavior.^{10–13,52,53} The strong hydration is characterized by a strong 3200 cm^{-1} peak, a weak (or absence of) 3400 cm^{-1} signal, and a strong overall water signal intensity.^{10–13} For example, SFG spectra collected from the antifouling SBMA and OEGMA surfaces in water are both dominated by the ~ 3200 cm^{-1} peak, showing that the surfaces are covered by strongly bonded water molecules, leading to excellent antifouling properties.¹⁰ For a control PMMA surface, both 3200 cm^{-1} and 3400 cm^{-1} peaks were detected, indicating that the PMMA surface has a substantial amount of loosely bonded water molecules and thus PMMA is not a good antifouling material.¹²

Fig. 2a shows a picture of a mussel adhesive plaque on a surface, while Fig. 2b shows the schematic of the SFG sample geometry.

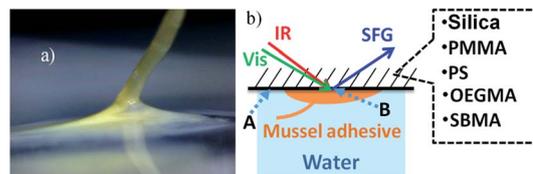


Fig. 2 Mussel adhesive plaque and SFG sample geometry. (a) Picture of a mussel adhesive plaque on a surface. (b) Schematic showing the SFG sample geometry used to collect SFG spectra from the sample/water interface (position A) and the sample/mussel adhesive interface (position B). The input laser beams penetrate the substrates (silica or silica with polymer thin films) to reach the sample/water and the sample/mussel adhesive interfaces. Aluminum was not studied because it is not transparent for the input laser beams.

geometry. SFG spectra can be collected from the sample/water interface (position A in Fig. 2b) and the sample/mussel adhesive plaque interface (position B in Fig. 2b).

Fig. 3 shows SFG spectra collected from the surfaces and interfaces of five samples, while aluminum was not included owing to a lack of optical transparency. Note that the y-axes are on different scales to aid visualization of spectral features. We first discuss the SFG spectra collected from the material/water interface in Fig. 3, shown in blue. It is worth mentioning that these SFG spectra were not collected from the “clean” material/water interface, instead, they were detected from the samples after the mussel deposition experiments, from the surface regions where there were no adhesives deposited (e.g., position A in Fig. 2b).

The O–H stretching SFG spectra collected from three control samples, fused silica, PMMA, and PS in water show some common features (blue spectra in Fig. 3a–c). In addition to the signals at around ~ 3200 cm^{-1} , all three blue spectra have strong ~ 3400 cm^{-1} signals, showing substantial coverage of loosely bonded water molecules on the surfaces, or weak hydration. This observation matches our previous observation on the “clean” PMMA/water interface well.¹² In contrast, SFG spectra collected from the SBMA and OEGMA surfaces are dominated by the ~ 3200 cm^{-1} signal (blue spectra in Fig. 3d and e), showing strong surface hydration, similar to those observed from the “clean” SBMA/water and OEGMA/water interfaces.¹⁰ The surface hydration detected by SFG can be well correlated with the above animal behavior on various sample surfaces: On the weakly hydrated silica, PMMA, and PS surfaces, mussels easily settled and deposited adhesives. On the strongly hydrated SBMA and OEGMA surfaces, mussel tried to move away from the surfaces, even after tying with rubber bands.

Our previous research shows that for SBMA, protein molecules cannot disrupt the surface hydration of SBMA at the SBMA/protein solution interface,¹⁰ due to the strong SBMA surface hydration. On the OEGMA/protein solution interfaces, protein molecules could disrupt the interfacial water structure and loosely deposit onto the surface, which can be easily washed off by water. Both SBMA and OEGMA are antifouling materials. For fouling surfaces, protein molecules can disrupt the surface hydration and deposit to the surfaces permanently.



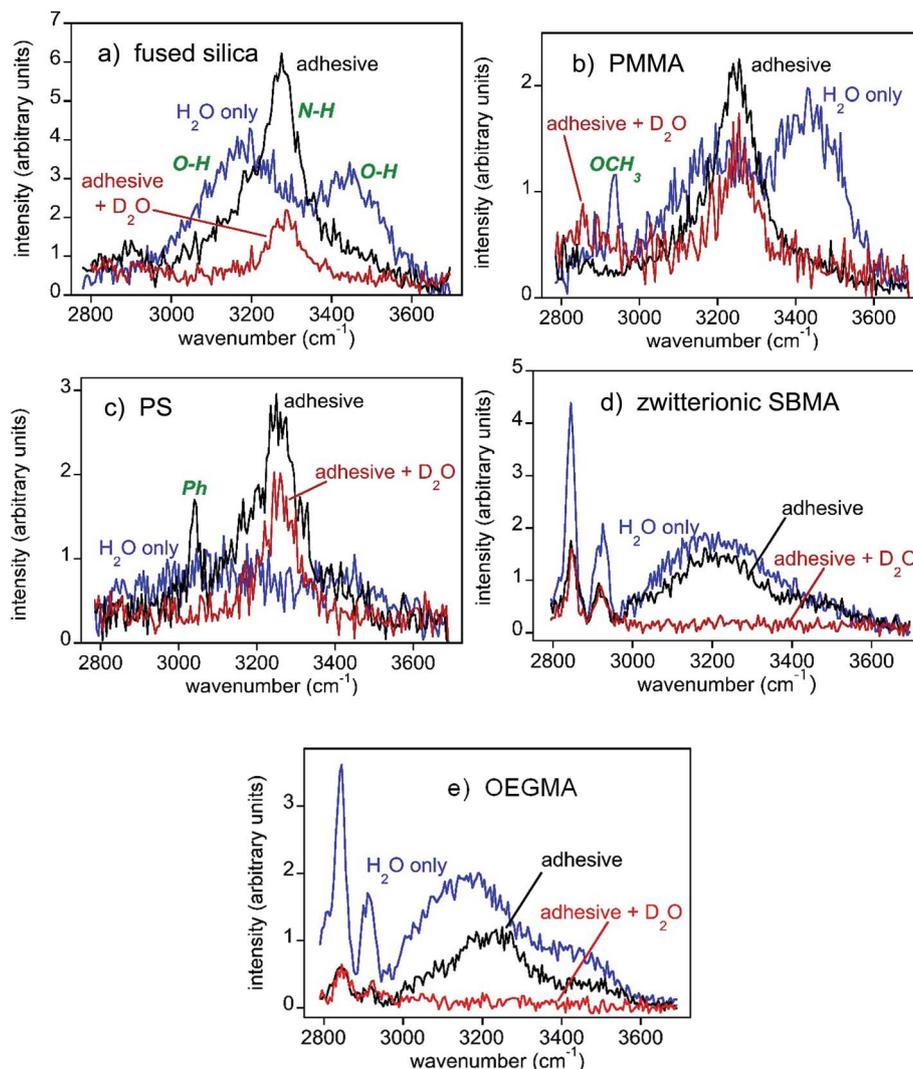


Fig. 3 SFG spectra of the substrate/water interface (region A in Fig. 2(b), blue curves), substrate/mussel adhesive interface (region B in Fig. 2(b), black curves), and the substrate/mussel adhesive interface after placing the sample in D_2O for a while (red curves). Note that the y-axes differ between panels in order to better visualize differences in spectra. The SFG intensities in different panels are different because of the different water orientation and ordering at various interfaces due to different interfacial interactions. SFG spectra collected from mussel adhesive plaque/substrate interfaces for the different plaques on the same type of substrate are similar and the results are reproducible. (a) Fused silica. General water and amine peak assignments are shown in green. These same assignments apply to all other spectra shown. (b) PMMA. In the water spectrum, prior to adhesive deposition, a substrate methyl peak was observed. (c) PS. The adhesive spectrum also shows a phenyl peak from the substrate. (d) Zwitterionic SBMA. (e) OEGMA.

Here instead of protein molecules, we used SFG to study interfacial hydration between various surfaces and mussel adhesives deposited by real live animals, the results of which will be presented below.

Dehydration at interfaces between mussel adhesives and control surfaces

To understand bio-adhesion and biofouling in detail, SFG spectra were collected from the buried interfaces between the three control samples (silica, PMMA, and PS) and mussel adhesives respectively (black spectra in Fig. 3a–c). After mussel attachment and partial plaque coverage, the interfacial SFG spectra of all three buried solid/solid interfaces between the

control samples and adhesives exhibit some common features: They all shifted from showing strong water signals (O–H stretching at $\sim 3200\text{ cm}^{-1}$ and $\sim 3450\text{ cm}^{-1}$) to those of amines from the protein-based adhesive (N–H stretching at $\sim 3280\text{ cm}^{-1}$). Spectral fitting results in Fig. S2 (ESI†) showed that a single amine peak can describe the spectra well, indicating that no ordered water exists at the three control material/mussel adhesive interfaces, and likely dehydration occurred at these interfaces.

SFG detects signals from a medium with no inversion symmetry. The absence of the SFG water signal at the control sample/mussel adhesive interfaces indicates that there is no ordered water at the interfaces. To confirm the interfacial



dehydration and to exclude the existence of disordered water (e.g., water molecules adopting a random orientation) at the interfaces, the above three samples were placed in D₂O and SFG spectra were then collected. The SFG spectral regions shown in Fig. 3 will not detect O-D and N-D stretching signals. If an interface is hydrated by disordered water molecules, then changing a sample from residing in H₂O (black spectra) to D₂O (red spectra) will diminish the interfacial N-H signals because of the H-D exchange between the N-H groups and D₂O molecules accessing the interfaces. Placement of adhesive-fused silica into D₂O (Fig. 3a) shrank the amine N-H signal with some intensity persisting. This change indicated that water (D₂O here) could access some of the interface to exchange protons with proteins (e.g., N-H to N-D). The remaining N-H signal indicated that water could not access the entire interface with the rest of the system being dehydrated, without ordered or disordered water. For PMMA, the amine N-H peak for the adhesive/sample interface only decreased a little (Fig. 3b), showing that the buried PMMA/mussel adhesive interface is mostly dehydrated and cannot be accessed by D₂O, preventing the H-D exchange for interfacial N-H groups. Further support for such a conclusion was obtained by having the animals deposit glue atop deuterated PMMA to yield similar results (Fig. S2, ESI†). PS (Fig. 3c) was generally similar to PMMA – a small N-H decrease when in D₂O, and analogous results when using a deuterated plastic surface (Fig. S2, ESI†). The H-D exchange experiments using D₂O clearly demonstrated the dehydration of the buried interfaces between the three control samples and mussel adhesives. Therefore we concluded that for biofouling to occur, mussels displaced the water on the sample surface and dehydrated the surface while depositing adhesives on the surface. These happened when a material surface does not have strong hydration.

Hydration at interfaces between mussel adhesives and antifouling SBMA as well as OEGMA

As presented above, most mussels left the SBMA or OEGMA surfaces without depositing adhesive plaques even when tied with rubber bands. Only a small amount of mussels stayed on the SBMA or OEGMA surfaces and deposited adhesives. SFG spectra were collected from the buried interfaces between these mussel adhesives and antifouling polymers, SBMA or OEGMA. For zwitterionic SBMA (Fig. 3d), plaque deposition decreased the SFG water signal, but only to a small extent. Furthermore, no prominent amine N-H stretching peak was observed. Quantitative spectral fitting results in Fig. S2 (ESI†) attested to the absence of the amide N-H signal. A water signal at ~3200 cm⁻¹ indicated water that was hydrogen bonded strongly. These data showed hydrogen bonded interfacial water residing between the mussel adhesive and the surface. From the OEGMA spectra (Fig. 3e), it can be seen that plaque binding decreased the water signal to yield a continued presence of water with a weak N-H contribution (Fig. S2, ESI†). Here, too, interfacial water appeared to persist between the adhesive and surface.

Soaking the adhesive-SBMA sample in D₂O eliminated almost all of the SFG signal (red spectrum in Fig. 3d), indicating that the entire interface was accessible by water (or D₂O). Furthermore, there appeared to be no dehydrated, ordered N-H functionalities at this adhesive-SBMA interface. A similar result was found for OEGMA (Fig. 3e). A weak N-H may have been present for adhesive-OEGMA (Fig. S2, Table S6, ESI†). Nonetheless, the entire SBMA and OEGMA systems appeared to be water accessible, with no regions of dehydration. The interfacial hydration between mussel adhesives and control samples *vs.* antifouling materials is drastically different. At the mussel adhesive/control sample interfaces, dehydration occurred. At the antifouling polymer/mussel adhesive interfaces, dehydration was not observed.

Adhesion testing

To further understand antifouling and bio-adhesion, we measured the adhesion strengths of mussel adhesives on control surfaces and antifouling surfaces. The adhesive performance of mussel glue on each substrate was quantified using an established method (Fig. 4).^{36,54} Briefly, adhesion values (in kPa) were obtained by pulling each plaque perpendicular to the surface until failure and the maximum force was divided by the plaque contact area on the surface. Data in Fig. 4 and Table S1† show that this adhesive functioned in a generally classic manner, with stronger binding to substrates with higher surface energies. Such behavior is in agreement with how adhesives stick better to, for example, metals than hydrophobic plastics.⁵⁵ However, mussel adhesion does not track perfectly with surface energy. Aluminum brings about particularly high strengths, possibly due to observations of changes in the surface (e.g., shiny to dull) after hours of residence in sea water.³⁶ Condition index tests ensured that animal health was not influenced by any substrate (Table S2†).^{56,57}

A particularly interesting finding here is that, in the rare instances when mussels attached to the OEGMA and

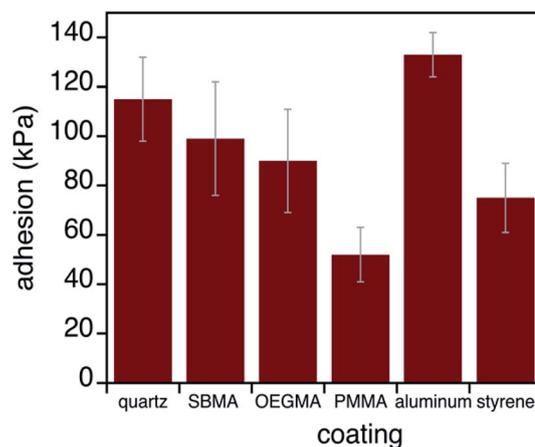


Fig. 4 Performance of mussel adhesives on different substrates. Average adhesion of mussel plaques on each surface. The substrates are ordered from the highest to lowest surface energies. Error bars shown are 99% confidence intervals.



zwitterionic SBMA substrates, adhesion strengths were quite substantial. This was different from what we expected. Since most adhesives and glues fail in wet environments, we thought that the interfacial hydration would lead to weak adhesion between mussel adhesives and SBMA or OEGMA. This was not observed and will be discussed in more detail below.

Discussion on antifouling

Our previous studies demonstrated that the surface hydration is well correlated with the antifouling activity of a material.^{10–13} For control samples or materials which do not possess antifouling properties, surface hydration is weak, as evidenced by the observation of both 3200 cm^{-1} and 3400 cm^{-1} peaks. Mussels can easily settle on such surfaces, dehydrate the surfaces, and deposit adhesives. Interfacial dehydration was observed *in situ* using SFG between mussel adhesives and the three control samples. The dehydration process and the adhesive deposition process could occur at the same time: The adhesive proteins are deposited onto the surface to replace interfacial water molecules. Adhesive proteins could also bind water molecules initially, but when they came into contact with the control sample directly, squeezing the initially bound water from the protein and the adhesive/control material interface, leading to interfacial dehydration. For the two antifouling materials SBMA and OEGMA, strong surface hydration was observed, as evidenced by the dominating SFG signals at 3200 cm^{-1} . With strong surface hydration, mussels do not like to settle on these surfaces and try to escape even when bound by rubber bands. The established antifouling properties of these surfaces may derive primarily from mussels avoiding the hydrating surfaces. Once attached, however, bonding tracked well with the high surface energies, which is not related to the antifouling behavior of the material.

Discussion on bio-adhesion

How might we explain the attachment strength of mussels to substrates without consistently strong protein peaks and without completely dehydrated surfaces? The first column of Table 1 lists these five substrates in the order of decreasing surface energy or increasing water contact angles (column 2). After mussel attachment, the SFG spectra collected from the buried mussel adhesive/sample interfaces were fit to approximate relative intensities of water O–H *versus* protein amine N–H signals (columns 3 and 4). Measured adhesion values are in

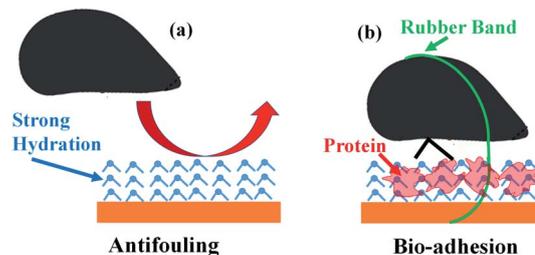


Fig. 5 Schematics (not drawn to scale) showing proposed mechanisms of antifouling (a) and strong bio-adhesion (b) of mussels on SBMA or OEGMA surfaces. Because of the strong surface hydration, mussels do not want to stay on the SBMA or OEGMA surface, leading to the excellent antifouling performance of SBMA and OEGMA (a). If mussels are forced to stay on SBMA or OEGMA (e.g., tied with a rubber band), the interfacial water can strongly interact with both surface and mussel adhesive proteins, resulting in strong bio-adhesion (b).

column 5. Quite striking is that, contrary to what one might expect, adhesion did not correlate with the detected presence of protein (N–H signal) on all surfaces. We can explain this underwater bonding with either water O–H or protein N–H signals. Such results lead to the conclusion that adhesion may be brought about either by the protein alone, which can dehydrate some substrates,^{16–18} or a combination of disordered protein and ordered interfacial water. Such ordered interfacial water can be the initial hydration layer on the SBMA or OEGMA surface and/or the protein bound water at the interface (protein bound water in the adhesive bulk should not contribute the SFG signal due to inversion symmetry). Water appears to help adhesion on some surfaces.

Speaking generally, water inhibits most adhesion phenomena. Water bound to surfaces prevents proteins or polymers from generating strong contacts. Cohesive forces can also be eliminated by hydration, thus “hiding” proteins or polymers from the counterparts and minimizing interactions. Reducing the amount of water present at surfaces appears to be a logical means of generating adhesive contacts. However, the practical aspects of such drying are not easy to achieve. Placement of hydrophobic species such as oils atop submerged high energy substrates may be able to remove bulk hydration, but water remains attached at the surface.^{58–60} Our current understanding of the importance of removing water to gain adhesive function may be in flux. Having a hydrophobic polymer appears to be helpful, if not essential, for displacing water and achieving strong adhesion between submerged substrates.⁶¹ One of the

Table 1 Examination of substrates as functions of water contact angles, SFG signal intensities, and mussel adhesion. Arbitrary units are denoted by “a.u.”

Surface	Water contact angle (°)	Water intensity with plaque (a.u.)	N–H intensity with plaque (a.u.)	Adhesion (kPa)
Fused silica	20	0	2.2 ± 0.2	115 ± 17
SBMA	20	1.2 ± 0.2	0	99 ± 23
OEGMA	42	1.1 ± 0.1	0	90 ± 21
PMMA	71	0	1.4 ± 0.1	52 ± 11
PS	87	0	1.5 ± 0.2	75 ± 14



highest performing underwater adhesives is made from an all hydrophobic polymer host with pendant catechol groups.⁶² Yet some data indicate that increasing the polymer dielectric constant aids underwater bonding.⁶³ These results may imply that hydrophilicity is actually beneficial for underwater bonding. Results found here indicate that viewing surfaces to be only wet and bad at adhesion *versus* dry and good for bonding is likely too simplistic of a view.

The hard, dry adhesives of insects and geckos provide a potential analogy to explain how interfacial water might contribute to the bonding of a wet, flowing adhesive. These terrestrial organisms attach temporarily to surfaces, *via* van der Waals forces, using hard and structured materials often shaped into nanoscale hair⁶⁴ or pad⁶⁵ structures. In general, they stick well when dry and less well or not at all when substrates are underwater.^{14,15} However, when in between dry and submerged, high humidity environments can actually increase the bonding of hard bioadhesives.^{66,67} Just the right amount of water fills gaps between the hard adhesive and substrate, contributing capillary forces and strengthening bonding.¹⁵ At an even smaller scale, nanoconfined water between the surface and adsorbate can be structured, no longer behaving in the way that we usually think of bulk solvent and possibly illustrating how surface hydration can be beneficial.⁶⁸ With data shown here, we may now be seeing the first known instance of interfacial hydration contributing to the underwater bonding of a curing bio-adhesive.

Conclusion

In this research, SFG has been applied to investigate the molecular structures of buried solid/liquid and solid/solid interfaces *in situ* in real time to understand antifouling and bio-adhesion, supplemented by the marine animal (mussel) activity study and adhesion measurements. It was found that both antifouling zwitterionic polymer SBMA and PEG containing polymer OEGMA have strong surface hydration in water. Because of the strong surface hydration, mussels do not want to settle on the SBMA or OEGMA surface to deposit adhesive plaque (Fig. 5a). Oppositely, on control sample surfaces, only weak surface hydration was detected. It is easy for mussels to settle on such surfaces, dehydrate the surfaces, and deposit adhesive plaques on these surfaces. Therefore surface hydration is important for antifouling activity of a material.

It is widely observed that bio-adhesion is weak in water. Our SFG studies show that dehydration occurs when mussels deposit adhesives on control sample surfaces, leading to strong adhesion, as expected. It has been shown in the literature that mussel adhesive proteins can replace interfacial water.^{16,69} When interfacial dehydration occurred, the interfacial SFG water O–H stretching signal was replaced by the interfacial protein N–H stretching signal. Interestingly, it was found that for a small amount of mussels which deposited adhesives on SBMA or OEGMA, interfacial dehydration did not occur. Under such circumstances, strong adhesion was also measured between the mussel adhesive and SBMA or OEGMA surface, with the presence of interfacial water. We therefore concluded

that under certain conditions, interfacial water can enhance the interfacial adhesion, with the help of interfacial proteins (Fig. 5b).

The detailed mechanism on how water and mussel adhesive protein interact at the interface to enhance adhesion needs further investigation in the future. It is well known that cross-linking of 3,4-dihydroxyphenylalanine (DOPA) containing mussel adhesive proteins plays an important role in bio-adhesion of mussels. We believe that on SBMA and OEGMA, interfacial water molecules can be strongly hydrogen bonded with both cross-linked mussel adhesive proteins and polymer surfaces, leading to strong bio-adhesion. Certainly other related or different effects such as chelating or capillary forces may also play roles.^{15,66,67,70}

This study demonstrates the power of using SFG to probe interfacial hydration/dehydration, and the importance of surface hydration/dehydration for antifouling and bio-adhesion. The knowledge obtained from this study will help the design and development of antifouling materials with improved performance and polymer materials with desired bio-adhesion properties.

Conflicts of interest

The authors declare no competing interests.

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