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Synthesis, Structures and Properties of Self-Assembling Quaternary Ammonium Dansyl Fluorescent Tags for Porous and Non-Porous Surfaces

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A series of H_2O and/or EtOH soluble, self-assembling guaternary ammonium salts containing a dansyl (DNS) fluorescent moiety suitable for attachment to both porous ([DNS-NH-(CH₂)₃-NMe₂- $R^{+}[X^{-}]$ (**2**; R = -Si(OMe)₃, X⁻ = Cl⁻) and non-porous (**3a**; R = -PO(OEt)₂, **3b**; -PO(O/-Pr)₂, **3c**; -(PO(OH)₂, X⁻ = Br), **4**; ([DNS-NH-(CH₂)₃-NMe₂-(CH₂)₃-NH(CH₂PO(OEt)₂)₂⁺][Br⁻], **5**; R = - $((CH_2)_3SCOCH_3, X^- = CI^-), 6\alpha; (R = -(CH_2)_nO(C_6H_4)CO(C_6H_5), n = 3, X^- = Br^-, 6b; n = 3, X^- = CI^-, Ab; n = 3, X^-, Ab; n =$ **óc**; n = 3, X⁻ = I⁻, **ód**; n = 4, X⁻ = Br⁻, **óe**; n = 4, X⁻ = I⁻, **óf**; n = 6, X⁻ = Br⁻, **óg**; n = 6, X⁻ = Cl⁻, **óh**; n = 6, X^- = 1), 7; (R = -CH₂-CH=CH₂, X^- = Br) surfaces were prepared from the precursor dansyl amine (1; DNS-NH-(CH₂)₃-NMe₂). Compounds 1-7 were characterized by NMR (¹H, ¹³C, ²⁹Si (2), ³¹P (**3a-c**, **4**)) spectroscopy, HRMS, UV-vis and Fluorescence spectroscopy. Additional characterization of compounds 1 and 7 were carried out by X-ray structure determinations. Physical attachment of compound 2 to cotton surfaces after immersion in solutions containing fluorescent dyes was verified by exposure to UV light and by complexation with bromo-phenol blue that rendered the surfaces visibly blue in colour. Phosphorus containing dansyl fluorescent dye, **3c**, was attached to a stainless steel surface by exposure to an aqueous solution containing this dye, resulting in the formation of a self-assembled fluorescent monolayer. UV cure of plastic surfaces (polypropylene, silicon medical tubing) coated with compound **6a** resulted in the covalent attachment of the dyes.

Introduction

Infection control and reduction of the spread of microorganisms such as fungi and bacteria is a key challenge facing health care providers. Many microorganisms have the ability to attach to surfaces and proliferate, forming colonies called biofilms.¹ The use of antibiotics to treat infectious diseases, including those caused by biofilms, is one of the most significant medical developments over the last century. However, after widespread use of these antibiotics, as well as other chemicals used for the purpose of disinfection, several strains of microorganisms (e.g. bacteria), have developed resistance.²⁻⁴ For the growing number of microorganisms with clinical importance (pathogens), there is either no efficacious therapy or only one or two effective antibiotics that are difficult to administer, expensive and/or have increasingly toxic side effects. Furthermore, when growing on surfaces as biofilms, microorganisms are generally

more persistent, and it is now estimated that the majority of infections (65-80%) involve these biofilms in some form.⁴ Biofilms also pose a notable threat of contamination in food processing facilities and spoilage of other products susceptible to microbial attack.⁵ One approach in preventing pathogenic biofilm formation, and thus the potential to cause spoilage or infection, is the use of antimicrobial coatings on surfaces that are not susceptible to the development of resistance by the target microorganisms.^{1,6,7} These coatings have bacteriostatic (inhibiting) or bactericidal (killing)⁸⁻¹² properties and thus afford a preventative strategy compared to disinfection, which is reactive, often after some damage or infection has occurred. In contrast to conventional antibiotics, bacteria do not readily develop resistance to antimicrobial coatings that inhibit microorganisms in a mechanical, as opposed to a chemical, fashion. This important distinction, and the related alarming

rate at which the number of effective antibiotics has declined, is a primary reason for the rapidly growing interest in antimicrobial coatings in recent years.¹³⁻²⁰



Quaternary ammonium compounds ("QAC's") including benzylalkonium salts (Figure 1), are well established as surfactants, possess antimicrobial activity and are used as disinfectants.^{21,22} QAC's consist of an irreversibly positively charged quaternary nitrogen atom where at least one substituent is often a long aliphatic chain. The synthesis of these compounds involves the quaternization of a tertiary amine following a Menshutkin reaction (i.e. a reaction of a tertiary amine with an alkyl halide).²³⁻²⁵ Surface bound antimicrobial QAC surface coatings, such as dimethyloctadecyl[3-(trimethoxysilyl)propyl]ammonium chloride, were first described by Isquith et al. who bound dimethyloctadecyl ammonium chloride via a silane anchor onto a variety of polyhydroxy surfaces.^{8,26-28} Research into developing alternative ways of grafting QAC's to a multitude of surfaces is hampered by the lack of simple, rapid and non-destructive detection methods for these coatings after they are applied. Traditionally, detection of surface silane bound QAC's on textiles has been achieved by visible staining of treated surfaces with a dilute aqueous solution of the bromophenol blue dye, (4,4'-(1,1-dioxido-3H-2,1-benzoxathiole-3,3-diyl)bis(2,6-

dibromophenol).²⁹ Bromophenol blue is a water soluble anionic dye that readily binds to QAC's and is characterized by a strong blue coloured ion pair that is visible to the naked eye. However, this dye often irreversibly binds to the textile, visibly staining the surface and destroying the material for future use. One solution that we envisioned was the use of surface anchored, water soluble QAC's functionalized with a fluorescent dansyl tag as an inexpensive, rapid and simple alternative to bromophenol blue for the facile detection of product surface coverage and coating uniformity. Thus, treated surfaces would glow fluorescent under exposure to a low power UV lamp, identifying areas of poor adhesion as well as missed areas during application. Additionally, it can also act as a unique product identifier and security feature for treated surfaces when added in trace amounts to functional antimicrobial solutions.

The dansyl fluorophore, capable of intermolecular chargetransfer, possesses high emission quantum yields and displays solvatochromism in the excited state giving rise to interesting and useful fluorescence properties.^{30,31} Along with the chemical flexibility of the sulfonic acid group, the dansyl amide has been utilized in various applications ranging from chemical sensing materials for the detection of explosives, biological weapons, metal ions, pH and as a probe/label for amino acids in the study of various diseases and metabolic pathways. For example, dansyl-functionalized organosilane self-assembled monolayers (SAM's) on glass surfaces were used to sense explosives (e.g. nitroarenes),³² whereas SAM's on silicon were used to sense pH, Hg^{2+} , Cl⁻, and Br⁻ ions.³³ Other functionalized dansyl probes have been used to study various diseases. In a study of the effect of copper on Alzheimer's and Wilson's diseases, Pitchumani designed dansyl-thiol silver nanoparticles for sensing Cu²⁺ ions.³¹ Meanwhile, Viirre and his group utilized Pd catalysis chemistry to incorporate a dansyl probe for the study of protein-protein interactions in cystic fibrosis research.³⁴ Dansyl phosphonic acid was also recently used to help elucidate organophosphonate metabolism in bacteria.³⁵

Experiments and Methods

Materials

The 3-(dimethylamino)propylamine, reagents, 3-(chloropropyl)trimethoxysilane, diethvl(3bromopropyl)phosphonate, N,N'-dimethylethylenediamine, 3chlorpropylthioacetate, allyl bromide, ACN, EtOH, were used as received (Sigma Aldrich). Anhydrous DCM and Et₂O were obtained by passage through a bed of activated molecular sieves under an atmosphere of dry $N_{2(g)}$. Dansyl chloride (> 97%) was used as received (Alfa Aesar). All fluorescent compounds were stored in glass vials in the absence of light. Nuclear magnetic resonance (NMR) experiments were recorded on a 400 MHz Bruker Avance II Spectrometer using deuterated chloroform (CDCl₃) as the solvent unless otherwise indicated. ¹H and ¹³C $\{^{1}H\}$ NMR spectra were referenced to the residual CHCl₃ ($\delta_{\rm H}$ = 7.26 ppm) and the central peak of CDCl₃ ($\delta_{\rm C}$ = 77.0 ppm) solvent signal respectively. The ${}^{31}P{}^{1}H{}$ spectra were referenced externally to 85% phosphoric acid ($\delta_P = 0.00$ ppm). Proton chemical shift assignments given in δ (ppm) were interpreted with the aid of 2-D COSY spectra, while carbon chemical shift assignments given in δ (ppm) were interpreted with the aid of 2-D HSQC spectra. Microwave reactions were performed in sealed glass reaction tubes utilizing the Biotage® Initiator Microwave Synthesizer (2.45 GHz). UV-VIS and fluorescence measurements were recorded on a Perkin Elmer Spectrophotometer (Lambda 20) and a Perkin Elmer Luminescence Spectrophotometer (LS 50B) respectively at the Ryerson University Analytical Centre (RUAC). High resolution mass spectra (HRMS) were recorded by direct analysis in real time by DART-TOF or by electrospray time-of-flight (ESI-TOF) at the Advanced Instrumentation for Molecular Structure (AIMS) laboratory at the University of Toronto. X-ray crystal structure analysis was performed with a Bruker-Nonius Kappa-CCD diffractometer at the University of Toronto X-ray diffraction facilities. Thin Layer Chromatography (TLC) was carried out on Silica gel 60 aluminium backed plates and visualized with a UV lamp or staining with (KMnO₄ or Ninhydrin). Melting points were measured using a Fischer Scientific melting point apparatus. UV curing of samples was achieved with a Novocure UV curing system from EFOS. UV curing of samples was initiated by UV curing at UV(A) (320-390 nm) and UV(V) (395-445 nm) wavelengths by exposing 1

× 1 cm² samples for 2.5 mins at distance of 5 cm from the light source. Samples were subjected to an average UV dosage of $3.80 \pm 0.369 \text{ J/cm}^2 \text{ UV}(\text{A})$ and $2.32 \pm 0.242 \text{ J/cm}^2 \text{ UV}(\text{V})$ and irradiance of $3.05 \times 10^{-2} \pm 1.90 \times 10^{-3} \text{ W/cm}^2 \text{ UV}(\text{A})$ and $1.90 \times 10^{-2} \pm 1.90 \times 10^{-3} \text{ W/cm}^2 \text{ UV}(\text{V})$. Dose and irradiance measurements were measured with an EIT Power Puck.

Synthesis of Precursors for the Dansyl Quaternization Reaction

Tetraethyl(((3-iodopropyl)azanediyl)bis(methylene))

bis(phosphonate)

Journal Name

A mixture of 3-(bis((diethoxyphosphoryl)methyl)amino)propyl methanesulfonate (1.0 g: 2.21 mmol) and NaI (0.69 g: 4.6 mmol) in acetone (3 mL) were placed, with a magnetic stirring bar, into a 5 mL μ W glass reaction tube and sealed. The reaction mixture was placed in microwave and, with constant stirring, run at 100°C for 5 min. The yellow solid was transferred to a 100 mL round bottom flask and washed with acetone (50 mL). Volatiles were removed with the aid of rotary evaporation and the crude material diluted with brine (10 mL) and then extracted with CHCl₃ (10 mL). The organic layer was separated, dried over anhydrous MgSO₄, filtered, and dried on a vacuum line and recovered as a yellow oil. Yield: 0.80 g (75%). TLC (10% w/w NH₄OH/acetone): $R_f = 0.85$. ¹H NMR (400 MHz, CDCl₃, δ): 4.20-4.09 (m, 8H: H6), 3.24 (t, 2H, J = 7.0Hz: H5), 3.15 (d, 4H, J = 8.7 Hz: H4), 2.89 (t, 2H, J = 6.6 Hz: H3), 2.03-1.94 (m, 2H: H2), 1.33 (t, 12H, J = 7.0 Hz: H1); ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 61.92 (t, ² J_{C-P} = 3.4 Hz, C6), 55.08 (C3), 49.46 (C4), 31.09 (C5), 30.96 (C2), 16.49 (t, ${}^{3}J_{C-P} = 2.9$ Hz: C1); ${}^{31}P{}^{1}H{}$ NMR (121.45 MHz, CDCl₃, δ): 24.79 ppm.

4-(3-bromopropyoxy)benzophenone^{36,37}

1,3-dibromopropane (6.14 mL: 60.5 mmol), K₂CO₃ (4.18 g: 30.2 mmol) and 4-hydroxybenzophenone (3.0 g: 15.1 mmol) were stirred in ACN (20 mL) under reflux for 24 h. The crude product was recrystallized in toluene/hexanes to give an off white coloured powder. Yield: 3.22 g (66.7%). Mp = $52-53^{\circ}\text{C}$. ¹H NMR (400 MHz, CDCl₃, δ): 7.83 (d, 2H, J = 8.7 Hz: H1), 7.77 (d, 2H, J = 8.3 Hz: H7), 7.58 (t, 1H, J = 7.4 Hz: H3), 7.48 (t, 2H, J = 7.7 Hz; H2), 6.98 (d, 2H, J = 8.7 Hz; H8), 4.20 (t, 2H, J = 5.8 Hz: H10), 3.63 (t, 2H, J = 6.3 Hz: H12), 2.37 (q, 2H, J = 6.0 Hz: H11) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 195.51 (C5), 162.30 (C9), 138.23 (C4), 132.58 (C3), 131.94 (C7), 130.36 (C6), 129.74 (C1), 128.21 (C2), 114.04 (C8), 65.53 (C10), 32.13 (C12), 29.74 (C11) ppm. This compound was previously prepared by Krichelodorf et al., but not characterized by NMR.³⁸ HRMS-DART (m/z): [MH⁺] calculated for C₁₆H₁₅BrO₂, 319.0334; found, 319.0329.

4-(3-chloropropoxy)benzophenone^{37,38}

1-bromo-3-chloropropane (5.00 mL: 50.4 mmol), K_2CO_3 (3.49 g: 25.3 mmol) and 4-hydroxybenzophenone (2.50 g: 12.6 mmol) were stirred in ACN (20 mL) under reflux for 24 h. The crude product was filtered off and recrystallized in toluene/hexanes to yield an off white coloured powder. Yield: 1.21 g (34.9%). Mp = 60-61°C. ¹H NMR (400 MHz, CDCl₃, δ):

7.83 (d, 2H, J = 8.4 Hz: H1), 7.76 (d, 2H, J = 7.2 Hz: H7), 7.56 (t, 1H, J = 7.3 Hz: H3), 7.48 (t, 2H, J = 7.5 Hz: H2), 6.96 (d, 2H, J = 8.4 Hz: H8), 4.23-4.16 (m, 2H: H10), 3.76 (t, 2H, J = 5.9 Hz: H12), 2.28 (t, 2H, J = 11.4 Hz: H11) ppm; $^{13}C{^{1}H}$ NMR (100 MHz, CDCl₃, δ): 195.53 (C5), 162.34 (C9), 138.25 (C4), 132.58 (C3), 131.93 (C7), 130.37 (C6), 129.74 (C1), 128.21 (C2), 114.04 (C8), 64.50 (C10), 41.30 (C12), 32.07 (C11) ppm. HRMS-DART (m/z): [MH⁺] calculated for C₁₆H₁₅ClO₂, 275.0838; found, 275.0841. (This compound was previously reported by Saettone *et al.*, but not fully characterized).⁴⁰

4-(3-iodopropoxy)benzophenone³⁷

4-(3-bromopropyoxy)benzophenone (1.00 g: 3.13 mmol) and NaI (1.41 g: 9.40 mmol) were mixed in acetone (10 mL) under reflux for 24 h. The crude product was filtered off and recrystallized in toluene/hexanes (1:2) to afford a yellow coloured powder. Yield: 0.59 g (51%). Mp = 72-74°C. ¹H NMR (400 MHz, CDCl₃, δ): 7.85-7.80 (m, 2H: H1), 7.77-7.73 (m, 2H: H7), 7.56 (ddt, 1H, J = 8.6 Hz, J = 6.6 Hz, J = 1.3 Hz: H3), 7.50-7.43 (m, 2H: H2), 6.99-6.89 (m, 2H: H8), 4.13 (t, 2H, J = 5.8 Hz: H10), 3.38 (t, 2H, J = 6.7 Hz: H12), 2.35-2.27 (m, 2H: H11) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 195.54 (C5), 162.30 (C9), 138.24 (C4), 132.59 (C3), 131.94 (C7), 130.39 (C6), 129.75 (C1), 128.21 (C2), 114.06 (C8), 67.52 (C10), 32.72 (C11), 2.13 (C12) ppm. HRMS-DART (m/z): [MH⁺] calculated for C₁₆H₁₅IO₂, 367.0195 found, 367.0202.

4-(4-bromobutoxy)benzophenone³⁷

1,4-dibromobutane (6.02 mL: 50.4 mmol), K_2CO_3 (3.49 g: 25.3 mmol) and 4-hydroxybenzophenone (2.50 g: 12.6 mmol) were stirred in ACN (20 mL) under reflux for 24 h to give a crude product of 4-(4-bromobutoxy)benzophenone which was recrystallized in toluene/hexanes to yield a pale yellow powder. Yield: 3.84 g (91.6%). Mp = 50-51°C. ¹H NMR (400 MHz, CDCl₃, δ): 7.81 (d, 2H, *J* = 8.8: H1), 7.74 (d, 2H, *J* = 7.2: H7), 7.55 (t, 1H, *J* = 7.4: H3), 7.46 (t, 2H, *J* = 7.7: H2), 6.94 (d, 2H, *J* = 8.8 Hz: H8), 4.07 (t, 2H, *J* = 5.9 Hz: H10), 3.50 (t, 2H, *J* = 6.5 Hz: H13), 2.14-2.05 (m, 2H: H11), 2.04-1.95 (m, 2H: H12) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 195.53 (C5), 162.52 (C9), 138.29 (C4), 132.59 (C3), 131.90 (C7), 130.20 (C6), 129.73 (C1), 128.20 (2), 113.99 (C8), 67.13 (C10), 33.29 (C13), 29.36 (C12), 27.77 (C11) ppm. HRMS-DART (m/z): [MH⁺] calculated for C₁₇H₁₇BrO₂, 333.0490 found, 333.0486.

4-(4-iodobutoxy)benzophenone³⁹

4-(4-bromobutoxy)benzophenone (1.00 g : 3.00 mmol) and NaI (0.900 g: 6.00 mmol) were mixed in acetone (10 mL) under reflux for 24 h to give crude product of 4-(4-iodobutoxy)benzophenone which was recrystallized in toluene/hexanes (1:2) to afford a pale yellow powder. Yield: 1.03 g (90.2%). Mp = 84-86°C. ¹H NMR (400 MHz, CDCl₃, δ): 7.80 (d, 2H, J = 8.7: H1), 7.74 (d, 2H, J = 7.3: H7), 7.54 (t, 1H, J = 7.3: H3), 7.47 (t, 2H, J = 7.6: H2), 6.93 (d, 2H, J = 8.8 Hz: H8), 4.06 (t, 2H, J = 5.9 Hz: H10), 3.26 (t, 2H, J = 6.7 Hz: H13), 2.10-2.02 (m, 2H: H11), 2.00-1.90 (m, 2H: H12) ppm;

¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 195.54 (C5), 162.52 (C9), 138.29 (C4), 132.59 (C3), 131.90 (C7), 130.20 (C6), 129.73 (C1), 128.20 (C2), 113.99 (C8), 66.93 (C10), 30.06 (C12), 30.02 (C11), 6.15 (C13) ppm. HRMS-DART (m/z): [MH⁺] calculated for $C_{17}H_{17}IO_2$, 381.0351 found, 381.0339. (This data is in good agreement with previously reported values).³⁹

4-(6-bromohexoxy)benzophenone^{38,39}

1,6-dibromohexane (6.21 mL: 40.4 mmol), K₂CO₃ (2.79 g: 20.2 mmol) and 4-hydroxybenzophenone (2.00 g: 10.1 mmol) were stirred in ACN (20 mL) under reflux for 24 h to give a crude product of 4-(6-bromohexoxy)benzophenone which was recrystallized in toluene/hexanes to yield a white coloured powder. Yield: 1.50 g (42.7%). Mp = $62-64^{\circ}$ C. ¹H NMR (400 MHz, CDCl₃, δ): 7.84-7.80 (m, 2H: H1), 7.77-7.41 (m, 2H: H7), 7.59-7.54 (m, 1H: H3), 7.50-7.44 (m, 2H: H2), 6.97-6.93 (m, 2H: H8), 4.05 (t, 2H, J = 6.4 Hz: H10), 3.43 (t, 2H, J = 6.8Hz: H15), 1.95-1.81 (m, 4H: (H11, H12)), 1.55-1.52 (m, 4H: (H13, H14)) ppm; ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃, δ): 195.64 (C5), 162.96 (C9), 138.22 (C4), 132.57 (C3), 131.07 (C7), 129.98 (C6), 129.78 (C1), 128.21 (C2), 113.98 (C8), 67.99 (C10), 34.02 (C15), 32.74 (C14), 29.15 (C13), 27.87 (C11), 25.59 (C12) ppm. HRMS-DART (m/z): [MH⁺] calculated for C₁₉H₂₁BrO₂, 361.0803; found, 361.0796. (This data is in good agreement with previously reported values).³⁹

4-(6-chlorohexoxy)benzophenone³⁷

1-bromo-6-chlorohexane (2.77 mL: 13.9 mmol), K₂CO₃ (3.49 g: 25.2 mmol) and 4-hydroxybenzophenone (2.50 g: 12.6 mmol) were stirred in ACN (20.0 mL) under reflux for 24 h to give a crude product of 4-(6-chlorohexoxy)benzophenone which was recrystallized in toluene/hexanes to yield an off white coloured powder. Yield: 3.07 g (76.8%). Mp = 65-66°C. ¹H NMR (400 MHz, CDCl₃, δ): 7.81 (d, 2H, J = 8.7 Hz: H1), 7.75 (d, 1H, J = 7.4 Hz: H7), 7.55 (t, 2H, J = 7.4 Hz: H3), 7.47 (t, 2H, J = 7.6 Hz: H2), 6.94 (d, 2H, J = 8.7 Hz: H8), 4.04 (t, 2H, J = 6.4 Hz: H10), 3.55 (t, 2H, J = 6.6 Hz: H15), 1.85-1.80 (m, 4H: H11, H12), 1.55-1.51 (m, 4H: H13, H14) ppm; $^{13}C{^{1}H}$ NMR (100 MHz, CDCl₃, δ): 195.55 (C5), 162.77 (C9), 138.35 (C4), 132.58 (C3), 131.86 (C7), 130.01 (C6), 129.72 (C1), 130.01 (C2), 114.01 (C8), 68.01 (C10), 44.98 (C15), 32.48 (C14), 28.98 (C13), 26.61 (C11), 25.38 (C12) ppm. HRMS-DART (m/z): $[MH^+]$ calculated for $C_{19}H_{21}ClO_2$, 317.1308; found, 317.1311.

4-(6-iodohexoxy)benzophenone³⁹

4-((3-bromohexoxy)benzophenone (0.500 g: 1.38 mmol) and NaI (0.622 g: 4.15 mmol) were mixed in acetone (10.0 mL) under reflux for 24 h to give crude product of 4-(6-iodohexoxy)benzophenone which was recrystallized in toluene/hexanes (1:2) to afford an off white coloured powder. Yield: 0.480 g (85.0%). Mp = 55-57°C. ¹H NMR (400 MHz, CDCl₃, δ): 7.77 (m, 4H: H1, H7), 7.51 (m, 3H: H2, H3), 6.95 (m, 2H: H8), 4.05 (m, 2H: H10), 3.21 (m, 2H: H15), 1.85 (m, 4H: H11, H12), 1.55 (m, 4H: H13, H14) ppm; ¹³C{¹H} NMR (CDCl₃, 100 MHz): δ 195.61 (C5), 162.62 (C9), 138.35 (C4),

132.57 (C3), 131.85 (C7), 130.00 (C6), 129.72 (C1), 128.17 (C2), 113.89 (C8), 68.13 (C10), 33.12 (C14), 30.17 (C11), 29.04 (C13), 25.02 (C12), 6.85 (C15) ppm. (This data is in good agreement with previously reported values).³⁹

5-(dimethylamino)-N-(3-(dimethylamino)propyl)naphthalene-1-sulfonamide $1^{40,41}$

To a flame dried 500 mL round bottom flask with a reflux condenser connected to an inert atmosphere manifold, anhydrous DCM (300 mL) was added followed by dansyl chloride (10.0 g: 37.07 mmol), Et₃N (8 mL: 55.6 mmol). While the solution was stirring at room temperature (23 \pm 2°C), 3-(dimethylamino)propylamine (7.0 mL: 55.6 mmol) was added drop wise via an inert syringe resulting in a colour change from orange to lime-green. After stirring for 1 h, HCl (g) was bubbled through the solution until a pH of 2 was achieved. The resulting mixture was evaporated to dryness, re-dissolved in saturated brine H₂O (100 mL) and brought to pH 11 with 6N NaOH (15 mL) at 0°C until a white-yellow coloured precipitate was observed. The mixture was refrigerated overnight enhancing further precipitation of product. The precipitate was filtered and washed (2 \times 30 mL) with H₂O and the filtrate extracted with DCM (500 mL) and evaporated to dryness to afford a white coloured solid, 1. The crude sample was recrystallized using a minimum volume (25 mL) of 80% EtOH/H₂O. Yield: 12.1 g (97%). Mp = 122-124°C. TLC (5% w/w NH₄OH:Acetone), $R_f = 0.72$. UV-Vis (MeOH, 1×10^{-3} M), $\lambda_{Abs max} = 516 \text{ nm}, \epsilon = 447 \text{ M}^{-1} \text{ cm}^{-1}$. ¹H NMR (400 MHz, $CDCl_3$, δ): 8.52 (d, 1H, J = 8.5 Hz: H8), 8.31 (d, 1H, J = 8.6Hz: H5), 8.23 (dd, 1H, ${}^{1}J = 1.2$ Hz, ${}^{2}J = 7.3$ Hz: H10), 7.58 – 7.50 (m, 2H: H4, H9), 7.17 (d, 1H, J = 7.5: H3), 2.94 (t, 2H, J =5.5 Hz: H13), 2.88 (s, 6H: H1), 2.21 (t, 2H, J = 5.5 Hz: H15), 2.12 (s, 6H: H16), 1.58-1.52 (m, 2H: H14) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 151.90 (C2), 134.77 (C11), 129.98 (C6), 128.89 (C8), 129.71 (C4), 129.65 (C9), 128.07 (C10), 123.17 (C5), 119.03 (C7), 115.00 (C3), 59.58 (C15), 45.42 (C1, C16), 44.54 (C13), 24.61 (C14) ppm. HRMS-DART (m/z): [M⁺] calculated for C₁₇H₂₆N₃O₂S₁, 336.1736; found, 336.1745. (This compound has also been prepared on a 25 g, 92.6 mmol scale).

General Procedure for the Synthesis of Dansyl Quaternary Ammonium salts

In a glass screw cap vial 5-(dimethylamino)-*N*-(3-(dimethylamino)propyl)naphthalene-1-sulfonamide **1** (1.0 equiv.) and the appropriate alkyl halide (1.0-2.0 equiv.) were dissolved in ACN or EtOH (0.5-1.0 M). The resultant solution was stirred at reflux (4-24 h) until TLC analysis showed the disappearance of the starting materials (EtOAc/hexanes 1:4). The residue was then precipitated from the resultant solution by the addition of cold Et_2O (5 × 5 mL) followed by decanting the solvent and placing the vial under high vacuum (~ 1 h) to afford the desired product.

3-(5-(dimethylamino)naphthalene-1-sulfonamido)-*N*,*N*dimethyl-*N*-(3-(trimethoxysilyl)propyl)propan-1-aminium chloride **2**

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To a flame dried 25 mL round bottom flask with a reflux condenser connected to an inert atmosphere manifold, ACN (20 mL) was added followed by 5-(dimethylamino)-N-(3-(dimethylamino)propyl)naphthalene-1-sulfonamide (3.35 g: 10 mmol). While stirring, (3-chloropropyl)trimethoxysilane (5 mL: 25 mmol) was added via syringe. The solution was stirred for 48 h at 110°C, with the solution turning to yellow-brownish coloured oil. The oil was then precipitated in cold (1:1) DCM:Et₂O mixture, forming two layers; a gummy layer and a white liquid layer. The white liquid layer was separated inside a 10 mL syringe and collected in a second 25 mL round bottom flask; the gummy layer was dissolved using DCM and collected in a 25 mL round bottom flask. Excess DCM was evaporated using rotary evaporator resulting in the recovery of a yellow coloured solid, 2. Yield: 3.85 g (72.0%). Mp = 85-87°C. UV-Vis (MeOH, 1×10^{-3} M), $\lambda_{Abs max} = 340$ nm, $\epsilon = 413$ M⁻¹ cm⁻¹. ¹H NMR (400 MHz, CDCl₃, δ): 8.48 (d, 2H, J = 8.6 Hz: H8, H5), 8.37 (s, 1H, H12), 8.19 (d, 1H, J = 7.3 Hz: H10), 7.59 (t, 1H, J = 7.9 Hz: H4), 7.49 (t, 1H, J = 8.2 Hz: H9), 7.13 (d, 1H, J = 7.6 Hz: H3), 3.57-3.52 (m, 2H: H17), 3.49 (s, 9H: H20), 3.27-3.20 (m, 2H: H15), 3.10 (s, 6H: H16), 3.08-3.03 (m, 2H: H13), 2.84 (s, 6H: H1), 2.01-1.90 (m, 2H: H14), 1.71-1.60 (m, 2H: H18), 0.54 (t, 2H, J = 7.8 Hz: H19) ppm; ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃, δ): 151.70 (C2), 135.06 (C11), 130.18 (C4), 129.75 (C6), 129.58 (C8), 128.93 (C9), 128.53 (C10), 123.35 (C5), 119.45 (C7), 115.14 (C3), 65.62 (C15), 62.27 (C17), 50.79 (C16), 50.68 (C20), 45.40 (C1), 39.81 (C13), 26.00 (C14), 16.16 (C18), 5.35 (C19) ppm; ²⁹Si{¹H} NMR (79.4 MHz, CD₃OD, δ): -68.86 ppm. HRMS-DART (m/z): [M⁺ - Cl⁻, - CH₃], calculated for C₂₃H₄₀ClN₃O₅SSi, 484.2314; found 484.2315.

3-(diethoxyphosphoryl)-*N*-(3-(5-(dimethylamino)naphthalene-1-sulfonamido)propyl)-*N*,*N*-dimethylpropan-1-aminium bromide **3a**

To a stirred solution of 1 (2.06 g: 6.13 mmol) in refluxing ACN (15 mL) was added diethyl(3-bromopropyl)phosphonate (2.40 g: 9.2 mmol) via syringe, and the vial was capped and refluxed for 20 h. To purify, the mixture was first triturated, washed with Et₂O (2 \times 40 mL) to remove any unreacted starting material followed by drying under high vacuum overnight to afford 3a as a yellow coloured solid. Yield: (3.64 g: 90%). Mp = 37-39°C. UV-Vis (MeOH, 1×10^{-3} M), $\lambda_{Abs max} = 334$ nm, $\epsilon = 505$ M^{-1} cm⁻¹. ¹H NMR (400 MHz, CDCl₃, δ): 8.51 (d, 1H, J = 8.3Hz: H8), 8.43 (d, 1H, J = 8.6 Hz: H5), 8.20 (dd, 1H, ${}^{1}J = 1.0$ Hz, ${}^{2}J = 7.3$ Hz: H10), 7.71 (s, 1H: H12), 7.59 (t, 1H, J = 8.4Hz: H9), 7.50 (t, 1H, J = 7.3 Hz: H4), 7.16 (d, 1H, J = 7.4 Hz: H3), 4.11-4.02 (m, 4H: H20), 3.67-3.56 (m, 4H: H15, H17), 3.17 (s, 6H: H16), 3.10-2.99 (m, 2H: H13), 2.86 (s, 6H: H1), 2.10-1.95 (m, 4H: H14, H18), 1.89-1.77 (m, 2H: H19), 1.28 (t, 6H, J = 7.1 Hz: H21) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 151.79 (C2), 134.62 (C11), 130.29 (C4), 129.79 (C6), 129.47 (C8), 129.34 (C9), 128.60 (C10), 123.28 (C5), 119.28 (C7), 115.30 (C3), 62.23-62.16 (overlap, C15, C17, C20), 51.31 (C16), 45.43 (C1), 39.73 (C13), 24.75-22.83 (C14, C18, C19), 16.45 (d, ${}^{2}J = 6.0$ Hz: C21) ppm; ${}^{31}P{}^{1}H{}$ NMR (121.45 MHz, $CDCl_{3,} \delta$): 29.29 ppm. HRMS-DART (m/z): [M⁺ - Br⁻] calculated for $C_{24}H_{41}BrN_3O_5P_1S_1$, 514.2504; found, 514.2519.

3-(diisopropoxyphosphoryl)-*N*-(3-(5-(dimethylamino) naphthalene-1-sulfonamido)propyl)-*N*,*N*-dimethylpropan-1aminium bromide **3b**

To a stirred solution of 1 (0.5 g: 1.5 mmol) in refluxing ACN (3 mL) was added diisopropyl(3-bromopropyl)phosphonate (0.46 g: 1.6 mmol) via syringe, and the vial was capped and refluxed for 7 h. To purify, the mixture was first triturated, washed with Et_2O (2 × 15 mL) to remove any unreacted starting material, followed by drying under high vacuum overnight to afford 3b as a yellow coloured solid. Yield: 0.653 g: (70%). Mp = 36-38°C; UV-Vis (MeOH, 1×10^{-3} M), $\lambda_{Abs max} = 340$ nm, $\varepsilon = 519$ M^{-1} cm⁻¹. ¹H NMR (400 MHz, CDCl₃, δ): 8.50 (d, 1H, J = 8.3Hz: H8), 8.43 (d, 1H, J = 8.6 Hz: H5), 8.22 (dd, 1H, ${}^{1}J = 1.0$ Hz, ${}^{2}J = 7.3$ Hz: H10), 7.75 (s, 1H: H12), 7.59 (t, 1H, J = 8.4Hz: H9), 7.49 (t, 1H, J = 7.3 Hz: H4), 7.15 (d, 1H, J = 7.4 Hz: H3), 4.68-4.56 (m, 2H: H20), 3.66-3.49 (m, 4H: H15, H17), 3.16 (s, 6H: H16), 3.09-2.97 (m, 2H: H13), 2.85 (s, 6H: H1), 2.05-1.90 (m, 4H: H14, H18), 1.80-1.69 (m, 2H: H19), 1.26 (t, 12H, J = 7.1 Hz: H21) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 151.79 (C2), 134.62 (C11), 130.31 (C4), 129.87 (C6), 129.49 (C8), 129.31 (C9), 128.64 (C10), 123.28 (C5), 119.28 (C7), 115.30 (C3), 70.69 (C20), 62.23-62.16 (overlap, C15, C17), 51.19 (C16), 45.42 (C1), 41.80 (C13), 39.60 (C21), 33.76 (C19), 24.08 (C14, C18) ppm; ³¹P{¹H} NMR (121.45 MHz, CDCl₃ δ): 27.32 ppm. HRMS-ESI-TOF (m/z): [M⁺ - Br⁻] calculated for C₂₆H₄₅BrN₃O₅P₁S₁, 542.2813; found, 542.2815.

3-(5-(dimethylamino)naphthalene-1-sulfonamido)-N,N-

dimethyl-*N*-(3-phosphonopropyl)propan-1-aminium bromide **3c** Inside a flame dried and evacuated 20 mL screw cap vial *N*-(3-(diethoxyphosphoryl)propyl)-*N*,*N*-dimethyloctadecan-1-

ammonium bromide (0.35 g: 0.58 mmol) was dissolved in anhydrous DCM (5 mL). To the clear solution, with stirring, was added trimethylsilyl bromide (0.23 mL: 1.76 mmol) through a rubber septum via syringe and the reaction was then stirred at room temperature overnight. Completion of the reaction was monitored by ³¹P NMR, after which the reaction was quenched with EtOH (10 mL) and stirred for 1 h followed by the addition of H₂O (1 mL). Volatiles were removed under high vacuum and the crude product was first triturated then washed with Et_2O (2 × 10 mL) to remove brown colored impurities. Further purification entailed extraction with $NH_4OH:H_2O$ (1:10, 10 mL) and washed with Et_2O (1 × 5 mL). The aqueous fluorescent layer was evaporated from ACN (50 mL) to give 3c as a light vellow coloured solid. Yield: 0.25 g: (79%). Mp = 166-168°C; UV-Vis (MeOH, 1×10^{-3} M), $\lambda_{Abs max}$ = 342 nm, ε = 449 M⁻¹ cm⁻¹. ¹H NMR (400 MHz, D₂O, δ): 8.58 (d, 1H, J = 8.5 Hz: H8), 8.35 (d, 1H, J = 8.5 Hz: H5), 8.23 (d, 1H, J = 7.0 Hz: H10), 7.70–7.58 (m, 2H: H4, H9), 7.31 (d, 1H, J = 7.5 Hz: H3), 3.32 (s, 6H: H16), 3.30-3.22 (m, 2H: H17) overlap), 3.19-3.11 (m, 2H: H15), 2.97 (t, 2H, J = 6.0 Hz: H13), 2.89 (s, 6H: H1), 1.93-1.86 (m, 4H: H14, H18), 1.57-1.49 (m, 2H, H19) ppm; ${}^{13}C{}^{1}H{}$ NMR (100 MHz, D₂O, δ): 133.80

(C2), 131.15 (C11), 130.15 (C6), 129.76 (C8), 128.99 (C4), 128.61 (C9), 128.40 (C10), 124.54 (C5), 119.81 (C7), 116.59 (C3), 64.32 (d, ${}^{3}J_{C-P} = 18.8$ Hz, C17), 61.33 (C15), 53.99 (C16), 45.08 (C1), 39.04 (C13), 24.77 (d, ${}^{1}J_{C-P} = 136.1$ Hz, C19), 21.96 (C14), 16.85 (d, ${}^{2}J_{C-P} = 3.2$, C18) ppm; ${}^{31}P{}^{1}H{}$ NMR (121.45 MHz, D₂O, δ): 20.99 ppm. HRMS-ESI-TOF (m/z): [M⁺ - Br⁻] calculated for C₂₀H₃₃BrN₃O₅P₁S₁, 458.1873; found, 458.1868.

3-(bis((diethoxyphosphoryl)methyl)amino)-N-(3-(5-

(dimethylamino)naphthalene-1-sulfonamido)propyl)-*N*,*N*-dimethylpropan-1-aminium iodide **4**

To a stirred solution of 1 (0.658 g: 1.35 mmol) in refluxing ACN (3 mL) was added tetraethyl (((3iodopropyl)azanediyl)bis(methylene))bis(phosphonate) (0.658 g: 1.35 mmol), and the vial was capped and refluxed for 7 h. To purify, the mixture was triturated, washed with Et₂O (2 \times 15 mL) to remove any unreacted starting material followed by drying under high vacuum overnight to afford 4 as a yellow solid. Yield: 0.626 g: (60%). Mp = 40-42°C; UV-Vis (MeOH, 1 × 10⁻³ M), $\lambda_{Abs max} = 340$ nm, $\epsilon = 526$ M⁻¹ cm⁻¹. ¹H NMR (400 MHz, CDCl₃, δ): 8.48-8.38 (m, 2H: H8), 8.37-8.29 (m, 1H: H5), 8.19-8.12 (m, 1H: H10), 7.72 (s, 1H: H12), 7.56-7.38 (m, 2H: H4, H9), 7.15 (m, 1H: H2), 4.16-3.97 (m, 8H: H21), 3.62-3.25 (m, 4H: H15, H17), 3.05-2.85 (s, 6H: H16), 3.10-2.91 (m, 8H: H19, H20, H13), 2.81 (s, 6H, H1), 1.99-1.75 (m, 4H: H14, H18), 1.25 (t, 12H, J = 7.0 Hz: H22) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 151.85 (C2), 135.16 (C11), 130.18 (C6), 129.76 (C8), 129.47 (C4), 129.12 (C9), 128.38 (C10), 123.34 (C5), 119.34 (C7), 115.19 (C3), 62.74 (C15), 62.24 (t, ${}^{2}J_{C-P} =$ 3.5 Hz: C21), 61.84 (C17), 53.78 (C19), 51.65 (C16), 50.49 (C20), 45.38 (C1), 39.61 (C13), 22.79 (C14), 21.22 (C18), 16.51 (d, ${}^{2}J_{C-P} = 2.5$ Hz: C22) ppm; ${}^{31}P{}^{1}H{}$ NMR (121.45 MHz, CDCl_{3.} δ): 24.42 ppm. HRMS-ESI-TOF (m/z): [MH⁺ - Γ] calculated for C₃₀H₅₅IN₄O₈P₂S₁, 693.3210; found, 693.3213.

3-(acetylthio)-*N*-(3-(5-(dimethylamino)naphthalene-1sulfonamido)propyl)-*N*,*N*-dimethylpropan-1-aminium chloride 5

To a stirred solution of 1 (1.0 g: 2.98 mmol) in refluxing EtOH (4 mL) was added 3-chloropropylthioacetate (90%) (0.65 mL: 4.9 mmol) via syringe, and the vial capped and refluxed for 24 h. To purify, the mixture was first triturated, washed with Et₂O $(2 \times 15 \text{ mL})$ to remove any unreacted starting material followed by drying under high vacuum overnight to afford 5 as a yellow coloured solid. Yield: 1.1 g (63%). Mp = 35-36°C. UV-VIS (MeOH, 1×10^{-3} M), $\lambda_{Abs max} = 334$ nm, $\epsilon = 439$ M⁻¹ cm⁻¹. ¹H NMR (400 MHz, CDCl₃, δ): 8.49 (d, 1H, *J* = 8.4 Hz: H8), 8.43 (d, 1H, J = 8.5 Hz: H5), 8.18 (d, 1H, J = 7.3 Hz: H10), 7.57 (t, 1H, J = 8.0 Hz: H4), 7.49 (t, 1H, J = 8.2 Hz: H9), 7.15 (d, 1H, J= 7.5 Hz: H3), 3.57-3.48 (m, 2H: H15), 3.40-3.32 (m, 2H: H17), 3.10 (s, 6H: H16), 3.05-2.98 (m, 2H: H13), 2.86 (s, 6H, H1), 2.83-2.77 (m, 2H: H19), 2.28 (s, 3H: H21), 2.03-1.85 (m, 4H: H14, H18) ppm; ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃, δ): 195.80 (C21), 149.79 (C2), 134.73 (C11), 130.34 (C4), 129.67 (C6), 129.43 (C8), 128.32 (C9), 128.71 (C10),124.64 (C5),

123.52 (C7), 115.45 (C3), 62.92 (C15), 62.73 (C17), 55.14 (C16), 45.32 (C1), 42.90 (C13), 30.77 (C21), 25.57 (C19), 23.16 (C18), 23.00 (C14) ppm. HRMS-ESI-TOF (m/z): $[M^+ - C\Gamma]$ calculated for $C_{22}H_{34}ClN_3O_3S_2$, 452.2036; found, 452.2037.

 $3-(4-benzoylphenoxy)-N-(3-((5-(dimethylamino)naphthalene-1-sulfonamido)propyl)-N,N-dimethylpropan-1-aminium bromide <math>6a^{37}$

To a stirred solution of compound 1 (0.5 g: 1.5 mmol) in refluxing ACN (4 mL) was added (4 - (3 bromopropoxy)benzophenone (0.72 g: 2.25 mmol) via syringe, and the vial was capped and refluxed for 24 h. To purify, the mixture was first triturated and washed with Et₂O (2 \times 15 mL) to remove any unreacted starting material followed by drying under high vacuum overnight and recovered 6a as a yellow coloured powder. Yield: 0.91 g (93%). Mp = 75-76°C; UV-VIS (MeOH, 1×10^{-3} M), $\lambda_{Abs max} = 335$ nm, $\epsilon_1 = 524$ M⁻¹ cm⁻¹, λ_{Abs} $max = 288 \text{ nm}, \epsilon_2 = 327 \text{ M}^{-1} \text{ cm}^{-1}$. ¹H NMR (400 MHz, CDCl₃, δ): 8.49-8.43 (m, 2H: H5, H10), 8.18 (d, 1H, J = 7.3 Hz: H8), 7.76-7.68 (m, 5H: H28, H22, H12, H9), 7.59-7.52 (m, 2H: H27), 7.47-7.42 (m, 3H: H26, H4), 7.07 (d, 1H, J = 7.6 Hz: H3), 6.82 (d, 2H, J = 8.8 Hz: H21), 4.00 (t, 2H, J = 5.4 Hz: H19), 3.73-3.69 (m, 2H: H15), 3.62-3.59 (m, 2H: H17), 3.22 (s, 6H: H16), 3.11-3.03 (m, 2H: H13), 2.81 (s, 6H: H1), 2.29-2.09 (m, 2H: H14), 2.05-1.95 (m, 2H: H18) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 195.47 (C24), 161.69 (C20), 151.86 (C2), 137.99 (C11), 134.86 (C25), 132.42 (C6), 132.06 (C28), 130.51 (C23), 130.37 (C8), 129.71 (C22), 129.44 (C26), 129.23 (C4), 129.20 (C9), 128.71 (C10), 128.25 (C27), 123.36 (C5), 119.39 (C7), 115.30 (C3), 114.15 (C21), 65.82 (C17), 64.50 (C15), 62.50 (C19), 51.37 (C16), 45.34 (C1), 39.87 (C13), 22.90 (C18), 15.26 (C14) ppm. HRMS-ESI-TOF (m/z): [M⁺ -Br⁻] calculated for C₃₃H₄₀BrN₃O₄S, 574.2749; found, 574.2734.

3-(4-benzoylphenoxy)-N-(3-(5-(dimethylamino)naphthalene-1-sulfonamido)propyl)-N,N-dimethylpropan-1-ammonium chloride **6b**³⁷

To a stirred solution of compound 1 (0.870 mmol: 0.291 g) and (4-(3-chloropropoxy)benzophenone (0.790 mmol: 0.250 g) were dissolved in ACN (2 mL) and left to stir in a 100°C sand bath for 24 h. The resultant residue was precipitated using cold Et₂O (4 mL) to afford the desired product 6b as a yellow coloured powder. Yield: 0.25 g (51.9%). Mp = $73-75^{\circ}$ C. ¹H NMR (400 MHz, CDCl₃, δ): 8.54-8.48 (m, 2H: H5, H10), 8.29 (d, 1H, J = 7.3 Hz; H8), 7.77-7.67 (m, 5H; H28, H9, H22),7.60-7.42 (m, 5H: H27, H26, H4), 7.19-7.12 (m, 1H: H3), 7.00-6.92 (m, 2H: H21), 4.23-4.16 (m, 2H: H19), 3.79-3.72 (m, 2H: H15), 3.63-3.59 (m, 2H: H17), 3.23 (s, 6H: H16), 2.81-2.79 (m, 2H: H13), 2.20 (s, 6H: H1), 2.32-2.28 (m, 2H: H14), 2.15-1.92 (m, 2H: H18) ppm; ${}^{13}C{}^{1}H{}$ NMR (100 MHz, CDCl₃, δ): 195.53 (C24), 162.33 (C20), 151.90 (C2), 138.16 (C11), 134.75 (C25), 132.57 (C6), 131.93 (C28), 130.00 (C23), 129.87 (C8), 129.72 (C22), 129.68 (C26), 129.62 (C4), 128.23 (C9), 128.20 (C10), 128.10 (C27), 123.17 (C5), 118.99 (C7), 115.01 (C3), 114.02 (C21), 64.48 (C17), 64.45 (C15), 59.49 (C19), 45.41

 $3-(4-benzoylphenoxy)-N-(3-(5-(dimethylamino)naphthalene-1-sulfonamido)propyl)-N,N-dimethylpropan-1-ammonium iodide <math>6c^{37}$

To a stirred solution of compound 1 (0.252 g: 0.750 mmol) and (4-(3-iodopropoxy)benzophenone (0.250 g: 0.680 mmol) were dissolved in ACN (2 mL) and left to stir in a 100°C sand bath for 24 h. The resultant residue was precipitated using cold Et₂O (4 mL) to afford the desired product 6c as a vellow coloured powder. Yield: 0.267 g: (56%) Mp = $73-75^{\circ}$ C. ¹H NMR (400 MHz, CDCl₃, δ): 8.49-8.39 (m, 2H: H5, H10), 8.18 (d, 1H, J = 7.7 Hz: H8), 7.75-7.65 (m, 5H: H28, H22, H12, H9), 7.59-7.49 (m, 2H: H27), 7.48-7.38 (m, 3H: H26, H4), 7.09 (d, 1H, J = 7.6 Hz: 1H, H3), 6.85 (d, 2H, J = 7.4 Hz: H21), 4.05-3.95 (m, 2H: H19), 3.65-3.56 (m, 4H: H15, H17), 3.15 (s, 6H: H16), 3.11-3.04 (m, 2H: H13), 2.80 (s, 6H: H1), 2.23-2.10 (m, 2H: H14), 2.05-1.93 (m, 2H: H18) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 195.45 (C24), 161.68 (C20), 151.88 (C2), 137.75 (C11), 134.33 (C25), 132.53 (C6), 132.45 (C28), 132.04 (C23), 130.55 (C8), 129.74 (C22), 129.29 (C26), 129.27 (C4), 128.91 (C9), 128.26 (C10), 128.10 (C27), 123.03 (C5), 118.94 (C7), 115.30 (C3), 113.80 (C21), 68.72 (C17), 64.00 (C15), 58.43 (C19), 51.42 (C16), 45.37 (C1), 39.03 (C13), 22.99 (C18), 18.42 (C14) ppm. HRMS-ESI-TOF (m/z): $[M^+ - I^-]$ calculated for C₃₃H₄₀IN₃O₄S, 574.2734; found, 574.2753.

4-(4-benzoylphenoxy)-*N*-(3-(5-(dimethylamino)naphthalene-1-sulfonamido)propyl)-*N*,*N*-dimethylbutan-1-ammonium bromide **6d**³⁷

To a stirred solution of compound 1 (0.240 g: 0.717 mmol) and (4-(4-bromobutoxy)benzophenone (0.721 mmol, 0.240 g) were dissolved in ACN (2 mL) and left to stir in a 100°C sand bath for 24 h. The resultant residue was precipitated using cold Et₂O (4 mL) to afford the desired product 6d. Yield: 0.168 g (35.0%). Mp = 73-75°C. ¹H NMR (400 MHz, CDCl₃, δ): 8.51-8.44 (m, 2H: H5, H10), 8.19 (d, 1H, J = 7.3 Hz: H8), 7.77-7.67 (m, 5H: H29, H23, H9), 7.60-7.51 (m, 2H: H28), 7.48-7.42 (m, 3H: H27, H4), 7.12 (d, 1H, J = 7.6 Hz: H3), 6.90 (d, 2H, J = 8.9 Hz: H22), 4.06-3.97 (m, 2H: H20), 3.71-3.60 (m, 2H: H15), 3.56-3.43 (m, 2H: H17), 3.16 (s, 6H: H16), 3.14-3.00 (m, 2H: H13), 2.82 (s, 6H: H1), 2.08-1.96 (m, 2H: H19), 1.94-1.65 (m, 4H: H18, H14) ppm; ${}^{13}C{}^{1}H{}$ NMR (100 MHz, CDCl₃, δ): 195.56 (C25), 162.22 (C21), 151.63 (C2), 138.09 (C11), 134.73 (C26), 132.51 (C6), 132.00 (C29), 130.35 (C24), 130.18 (C8), 129.73 (C23), 129.46 (C27), 129.26 (C4), 128.90 (C9), 128.74 (C28), 128.23 (C10), 123.44 (C5), 119.12 (C7), 114.87 (C3), 114.11 (C22), 67.23 (C17), 64.00 (C15), 61.66 (C20), 51.18 (C16), 45.39 (C1), 39.87 (C13), 25.74 (C19), 23.00 (C14), 19.75 (C18) ppm. HRMS-ESI-TOF (m/z): $[M^+ - Br^-]$ calculated for C₃₄H₄₂BrN₃O₄S, 588.2908; found, 588.2890.

To a stirred solution of compound 1 (0.201 g: 0.598 mmol) and (4-(4-iodobutoxy)benzophenone (0.250 g: 0.658 mmol) were dissolved in ACN (2 mL) and left to stir in a 100°C sand bath for 24 h. The resultant residue was precipitated using cold diethyl ether (4 mL) to afford the desired product 6e as a yellow coloured powder. Yield: 0.244 g (56.9%). Mp = $73-75^{\circ}$ C. ¹H NMR (400 MHz, CDCl₃, δ): 8.49 (d, 1H, J = 8.20 Hz: H5), 8.40 (d, 1H, J = 8.6 Hz: H10), 8.18 (d, 1H, J = 7.3 Hz: H8), 7.77-7.70 (m, 5H: H29, H23, H9), 7.61-7.53 (m, 2H: H28), 7.50-7.42 (m, 3H: H27, H4), 7.12 (d, 1H, J = 7.4 Hz: H3), 6.91 (d, 2H, J = 8.6 Hz: H22), 4.10-3.99 (m, 2H: H20), 3.86-3.54 (m, 2H: H15), 3.53-3.43 (m, 2H: H17), 3.14 (s, 6H: H16), 3.10-3.01 (m, 2H: H13), 2.84 (s, 6H: H1), 2.09-1.78 (m, 6H: H19, H18, H14) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 195.45 (C25), 162.22 (C21), 151.92 (C2), 138.05 (C11), 134.37 (C26), 132.58 (C6), 132.51 (C29), 132.04 (C24), 130.55 (C8), 130.19 (C24), 129.74 (C27), 129.32 (C24), 128.91 (C9), 128.25 (C28), 128.20 (C10), 123.62 (C5), 119.37 (C7), 115.53 (C3), 114.20 (C22), 66.99 (C17), 64.41 (C15), 62.73 (C20), 51.39 (C16), 45.40 (C1), 39.62 (C13), 29.81 (C19), 25.74 (C14), 19.77 (C18) ppm. HRMS-ESI-TOF (m/z): $[M^+ - I^-]$ calculated for C₃₄H₄₂IN₃O₄S, 588.2890; found, 588.2904.

6-(4-benzoylphenoxy)-N-(3-(5-(dimethylamino)naphthalene-1-sulfonamido)propyl)-N,N-dimethylhexan-1-ammonium bromide $6f^{37}$

To a stirred solution of compound 1 (0.211 g: 0.629 mmol) and (4-((6-bromohexoxy)benzophenone (0.250 g: 0.692 mmol) were dissolved in ACN (2 mL) and left to stir in a 100°C sand bath for 24 h. The resultant residue was precipitated using cold Et₂O (4 mL) to afford the desired product 6f as a yellow coloured powder. Yield: 0.385 g (87.8%). Mp = $70-71^{\circ}$ C. ¹H NMR (400 MHz, CDCl₃, δ): 8.49 (d, 1H, *J* = 8.8 Hz: H5), 8.46 (d, 1H, J = 8.7 Hz: H10), 8.21-8.19 (m, 1H: H8), 7.78-7.71 (m, 5H: H31, H25, H12, H9), 7.60-7.53 (m, 2H: H30), 7.50-7.42 (m, 3H: H29, H4), 7.13 (d, 1H, J = 7.4 Hz: H3), 6.89 (d, 2H, J = 8.4 Hz: H24), 3.90 (t, 2H, J = 6.4 Hz: H21), 3.70-3.58 (m, 2H: H15), 3.39-3.28 (m, 2H: H17), 3.15 (s, 6H: H16), 3.13-3.00 (m, 2H: H13), 2.87 (s, 6H: H1), 1.79-1.62 (m, 6H: H21, H18, H14), 1.57-1.41 (m, 2H: H20), 1.40-1.28 (m, 2H: H19) ppm; $^{13}C{^{1}H}$ NMR (100 MHz, CDCl₃, δ): 195.58 (C27), 162.68 (C23), 151.21 (C2), 138.20 (C11), 134.94 (C28), 132.56 (C6), 132.50 (C31), 131.92 (C26), 129.86 (C8), 129.68 (C25), 129.45 (C30), 129.17 (C4), 128.69 (C9), 128.20 (C29), 128.17 (C10), 123.17 (C5), 119.13 (C7), 115.30 (C3), 114.06 (C24), 67.81 (C17), 64.65 (C15), 55.01 (C22), 50.77 (C16), 45.38 (C1), 39.20 (C13), 33.76 (C21), 32.59 (C19), 28.70 (C20), 27.78 (C18), 25.42 (C14) ppm. HRMS-ESI-TOF (m/z): [M⁺ - Br⁻] calculated for C₃₆H₄₆BrN₃O₄S, 616.3224; found, 616.3203.

 $6-(4-benzoylphenoxy)-N-(3-(5-(dimethylamino)naphthalene-1-sulfonamido)propyl)-N,N-dimethylhexan-1-ammonium chloride <math>6g^{37}$

To a stirred solution of compound 1 (0.291 g: 0.870 mmol) and (4-(6-chlorohexoxy)benzophenone (0.250 g: 0.790 mmol) were dissolved in ACN (2 mL) and left to stir in a 100° C sand bath

for 24 h. The resultant residue was precipitated using cold Et₂O (4 mL) to afford the desired product 6g as a vellow coloured powder. Yield: 0.435 g (84.5%). Mp = $65-67^{\circ}$ C. ¹H NMR (400 MHz, CDCl₃, δ): 8.50 (d, 1H, J = 8.5 Hz: H5), 8.28 (d, 1H, J = 8.7 Hz: H10), 8.22-8.20 (m, 1H: H8), 7.80-7.71 (m, 5H: H31, H25, H12, H9), 7.60-7.42 (m, 5H: H30, H29, H4), 7.15 (d, 1H, J = 7.5 Hz: H3), 6.93 (d, 2H, J = 8.8 Hz: H24), 4.03 (t, 2H, J =6.4 Hz: H22), 3.54 (t, 2H, J = 6.6 Hz: H15), 3.30-3.26 (m, 2H: H17), 3.10 (s, 6H: H16), 3.08-3.06 (m, 2H: H13), 2.87 (s, 6H: H1), 1.84-1.78 (m, 6H: H21, H18, H14), 1.55-1.25 (m, 4H: H20, H19) ppm; ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃, δ): 195.58 (C27), 162.76 (C23), 151.89 (C2), 138.31 (C11), 134.74 (C28), 132.57 (C6), 132.53 (C31), 131.96 (C26), 130.01 (C8), 129.91 (C25), 129.70 (C30), 129.68 (C4), 128.62 (C9), 128.20 (C29), 128.17 (C10), 123.17 (C5), 118.98 (C7), 115.02 (C3), 113.99 (C24), 67.99 (C17), 67.77 (C15), 59.44 (C22), 50.93 (C16), 45.41 (C1), 44.41 (C13), 32.46 (C21), 28.95 (C19), 26.59 (C20), 25.46 (C18), 24.64 (C14) ppm. HRMS-ESI-TOF (m/z): $[M^+ - Cl^-]$ calculated for $C_{36}H_{46}ClN_3O_4S$, 616.3221; found, 616.3203.

6-(4-benzoylphenoxy)-N-(3-(5-(dimethylamino)naphthalene-1-sulfonamido)propyl)-N,N-dimethylhexan-1-ammonium iodide**6h**³⁷

To a stirred solution of compound 1 (0.272 g: 0.366 mmol) and (4-((6-iodohexoxy)benzophenone (0.136 g: 0.333 mmol) were dissolved in ACN (2 mL) and left to stir in a 100°C sand bath for 24 h. The resultant residue was precipitated using cold Et₂O (4 mL) to afford the desired product as a yellow coloured powder **6h**. Yield: (0232 g (93.5%). Mp = 67-68°C. ¹H NMR (400 MHz, CDCl₃, δ): 8.48 (d, 1H, J = 8.5 Hz: H5), 8.41 (d, 1H, J = 8.7 Hz: H10), 8.20-8.17 (m, 1H: H8), 7.78-7.70 (m, 5H: H31, H25, H12, H9), 7.61-7.42 (m, 5H: H30, H29, H4), 7.13 (d, 1H, J = 7.2 Hz: H3), 6.90 (d, 2H, J = 8.9 Hz: H24), 3.95 (t, 2H, J = 6.3 Hz: H22), 3.56-3.49 (m, 2H: H15), 3.33-3.29 (m, 2H: H17), 3.10 (s, 6H: H16), 3.08-3.06 (m, 2H: H13), 2.83 (s, 6H: H1), 1.76-1.59 (m, 6H: H21, H18, H14), 1.49-1.40 (m, 2H: H20), 1.36-1.24 (m, 2H: H19) ppm; ¹³C{¹H} NMR (100 MHz, CDCl₃, δ): 195.63 (C27), 162.69 (C23), 151.87 (C2), 138.21 (C11), 132.53 (C28), 131.94 (C6), 131.76 (C31), 130.55 (C26), 129.92 (C8), 129.72 (C25), 129.45 (C30), 129.33 (C4), 128.89 (C9), 128.22 (C29), 128.18 (C10), 123.62 (C5), 119.36 (C7), 115.54 (C3), 114.11 (C24), 67.23 (C17), 64.65 (C15), 59.09 (C22), 51.42 (C16), 45.42 (C1), 39.22 (C13), 28.71 (C21), 25.69 (C19), 25.47 (C20), 22.96 (C18), 22.59 (C14) ppm. HRMS-ESI-TOF (m/z): $[M^+ - I^-]$ calculated for $C_{36}H_{46}IN_3O_4S_4$, 616.3217; found, 616.3203.

N-(3-(5-(dimethylamino)naphthalene-1-sulfonamido)propyl)-N,N-dimethylprop-2-en-1-aminium bromide 7^{42}

In a 5 mL microwave vial equipped with a magnetic stir bar, compound 1 (0.355 g: 1.06 mmol) was dissolved in ACN (1 mL). Allylbromide (0.09 mL, 1 mmol) was carefully added to the solution with a syringe and the reaction vessel was slightly heated, using a heat gun, to allow the reactants to fully dissolve. The capped vial was then placed in the microwave and, with

constant stirring, run at 150°C for 2 min. To the resulting yellow mixture was added Et₂O (~10 mL) and the vial placed in the refrigerator overnight. The excess solvent was decanted out of the vial and a yellow precipitate layer attached to the bottom of the vial remained. The vial was then attached to a highvacuum pump for 1 h to remove any remaining solvent. A vellow coloured oil was recovered and the crude product recrystallized in minimal amount of MeOH (2 mL). After 48 h, yellow solid crystals suitable for X-ray crystallography were obtained. Yield: 0.36 g (75%). Mp = 164-166°C; $R_f = 0.38$ (5%, NH₄OH:Acetone). UV-Vis (MeOH, 1×10^{-3} M), $\lambda_{Abs max} =$ 339 nm, $\varepsilon = 413 \text{ M}^{-1} \text{ cm}^{-1}$. $\varepsilon = 560 \text{ M}^{-1} \text{ cm}^{-1}$.¹H NMR (CD₃OD, 400 MHz) δ 8.58 (d, 1H, J = 8.5 Hz: H9), 8.35 (d, 1H, J = 8.4 Hz: H6), 8.23 (d, 1H, J = 8.5 Hz: H11), 7.66-7.59 (m, 4H: H5, H10), 7.30 (d, 1H, J = 7.4 Hz: H4), 5.96-5.86 (m, 1H: H18), 5.67-5.63 (m, 2H: H19), 3.83 (d, 2H, J = 7.35 Hz: H17), 3.21-3.17 (m, 2H: H15), 2.97 (t, 2H, J = 6.09 Hz: H13), 2.90 (s, 6H:H16), 2.88 (s, 6H: H1), 1.89-1.84 (m, 2H: H14) ppm; ${}^{13}C{}^{1}H{}$ NMR (CD₃OD, 100.6 MHz) δ 151.94 (C2), 135.17 (C11), 130.08 (C8), 129.76 (C7), 129.37 (C9), 129.17 (C10), 128.31 (C6), 128.15 (C19), 124.54 (C18), 123.20 (C4), 118.94 (C5), 115.25 (C3), 66.23 (C17), 61.50 (C15), 47.42 (C16), 44.44 13 (C1), 39.26 (C13), 22.66 (C14) ppm. HRMS (ESI-TOF) (m/z): $[M^+ - Br^-]$ calculated for $C_{20}H_{30}BrN_3O_2S$: 376.2057; found 376.2053.

Results and Discussion

Synthesis and Characterization

Quaternary ammonium dansyl fluorescent tags were synthesized by a Menshutkin reaction between the dansyl amine precursor 1 (Scheme 1) and various linkers (Scheme 2). Compound 1 was obtained in high yield (97%) by a modified procedure from literature and purified by avoiding both a problematic acid/base extraction and column chromatography.41 Previous preparations of 1 had called for quenching of the phosphate buffer and or reaction with sequential acidifying/basifying steps followed by extraction into DCM, which in our hands lead to inseparable emulsions and low yields. A more direct work up included the initial acidification of the reaction, removal of the DCM, followed by the addition of base (6N KOH in brine at -20°C, O/N) and finally precipitation of the desired compound. Precursor 1 was isolated by vacuum filtration, washed (4 \times 50 mL) with cold H₂O and recrystallized from 20% (v/v) EtOH/H₂O to afford Xray quality crystals. Compound 1 can be readily scaled up (25 g) and used in the preparation of new dansyl QAC's (Scheme 2) displaying a variety of functional linkers for grafting onto porous and non-porous surfaces.

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All quaternization reactions were performed in sealed glass vials with refluxing acetonitrile (ACN) for 7-48 h depending on the alkyl halide. Reaction completion, monitored by NMR (¹H: CDCl₃) for products 2-7, revealed the characteristic disappearance of the dimethylamino protons at ~ 2.2 ppm of 1 and the appearance of two new upfield resonances at ~ 3.5 ppm $[(-CH_2-N^+(Me_2)-CH_2-)]$ and at ~ 3.3 ppm $[(-N^+-(CH_3)_2)]$ (Figure 2). The phosphorus containing compounds, **3a-b**, prepared from precursors possessing a bromo leaving group formed within 7 h, while the chloro-containing silane (2) and thioacetate (5) compounds prepared from precursors with chloro leaving groups required substantially longer reaction times (24-48 h). A simple and convenient purification method was developed based on the insolubility in Et₂O of the newly formed QACs. Compounds 2-7 precipitated directly from solutions using cold Et₂O. They are initially isolated as viscous oils attached to the walls of the vial after several cycles of rinsing and decantation. Further drying under reduced pressure resulted in the isolation of light yellow fluorescent powders in modest to good yield (35-94%) and high purity. The quaternization reaction for the silane derivative 2 showed, along with the two new upfield proton resonances, a ²⁹Si{¹H} NMR (CD₃OD) resonance at δ = -68.4 ppm.



R	X	Product	Yield %
-0 -0-si -0	Cl	2	72
	Br	3 a	90
O PrOi P OiPr	Br	3b	70
EtO_p EtO_N O=P-OEt OEt	ľ	4	60
o L _s ~~x	Cl	5	69
	Br ⁻ Cl ⁻ I ⁻	6a 6b 6c	93 52 56
	Br ⁻ I ⁻	6d 6e	35 57
	Br ⁻ Cl ⁻ I ⁻	6f 6g 6h	88 85 94
N N N N N N N N N N N N N N N N N N N	Br ⁻	7	75

Scheme 2: Preparation of fluorescent anchors for binding to porous and non-porous surfaces.

The dansyl phosphonate QAC's, 3a and 3b, were twice dealkylated to the corresponding phosphonic acid 3c by refluxing in mineral acids (Scheme 3). As expected, faster de-alkylation was accomplished with HBr when compared to HCl, and with the *i*-PrO leaving group versus that of the EtO group. It was also necessary to use aqueous HBr as anhydrous HBr (20% in EtOH) was significantly slower at cleaving the phosphonate ester and could not be forced to reach completion even after prolonged heating. In aqueous HBr, the reaction with 3a or 3b was complete after 1-2 h, whereas after 48 h of heating 3a in ethanolic HBr, the reaction was only 71% complete by NMR. The NMR $({}^{1}H, {}^{31}P{}^{1}H{}$: CDCl₃) spectrum for the diphosphonate esters **3a** and **3b** (31 P NMR, $\delta_P = 29.29$ and $\delta_P =$ 27.32 ppm respectively) showed a slight upfield shift in ${}^{31}P$ resonance to 20.99 ppm (D₂O) and the disappearance of the hydrogen resonances of the Et ($\delta_{\rm H}$ = 4.06, 1.28 ppm) and i-Pr groups ($\delta_{\rm H}$ = 4.69, 1.26 ppm) after ester hydrolysis to the free phosphonic acid **3c** (Figure 2). Dansyl α -bisphosphonate, **4**, was also synthesized from 1 and 3-iodopropyl- α bisphosphonate with NMR (CDCl₃) analysis showing the expected hydrogen ($\delta_{\rm H}$ = 3.06 ppm), carbon ($\delta_{\rm C}$ = 50.49 ppm) and phosphorus ($\delta_P = 24.42$ ppm) resonances characteristic of the aminophosphonate group. However, no dealkylation of 4 to the free phosphonic acid was investigated.



The thioacetate protected dansyl derivative, 5, selected for potential binding onto noble metals, was also successfully from 1 and commercially available 3prepared chloropropylthioacetate. NMR analysis (¹H, ¹³C: CDCl₃) of 5 revealed characteristic acetate hydrogen ($\delta_{\rm H} = 2.28$ ppm) and carbon resonances ($\delta_{\rm C} = 195.80$ and $\delta_{\rm C} = 30.77$ ppm). However, an attempted deprotection of 5 to the free thiol (MeOH/KOH) was unsuccessful with the reaction mixture changing colour from a light yellow to green and finally to a deep purple colour. This is likely due to decomposition of the free thiol. No further attempts to make the free thiol derivative were carried out.



Scheme 3: Cleavage of dansyl phosphonate and attempted cleavage of the dansyl thioacetate.

A series of benzophenone terminated quaternary ammonium materials, with different counter ions and alkyl chains, were prepared from the reaction of **1** with the appropriate (n-haloalkoxy)benzophenone derivative (100°C for 24 h). NMR analysis (¹H, ¹³C{¹H}: CDCl₃) of compounds **6a-h** confirmed their expected structures; with the ¹³C NMR spectra revealing the expected ketone carbon resonance at $\delta_C \approx 195$ ppm with the aromatic carbon bound to oxygen shifted slightly upfield at $\delta_C \approx 162$ and 152 ppm. Compound **1**, allyl bromide, and ACN were combined in a reaction vessel and stirred with microwave heating at 150°C for 2 min. The reaction was cooled, filtered

and compound 7 recovered as an oily, yellow coloured semisolid. Recrystallization in a minimal amount of MeOH provided yellow crystals in good yield (75%). Analysis of 7 by NMR (CD₃OD) revealed characteristic vinyl hydrogen ($\delta_H =$ 5.89 and $\delta_H =$ 5.64 ppm) and carbon ($\delta_C =$ 128.15 and $\delta_C =$ 124.54 ppm) resonances. With the exception of 6 and 7, all dansyl QAC derivatives with an electronegative atom present in the linker (i.e. 2, 3a, 3b, 3c, 4, 5) are H₂O soluble. Compounds 6a-h and 7, bearing the hydrophobic benzophenone and/or allyl group are only slightly soluble in H₂O, but readily dissolved in EtOH.

X-ray Crystallography of 1 and 7

Schematic representations of a unit cell molecule of precursor dansyl compound 1 and the ammonium dansyl allyl salt 7 are depicted in Figure 3. Crude 1 was re-crystallized using a 20 % (v/v) EtOH/H₂O mixture, resulting in the isolation of fine, clear prismatic crystals. Crystal structure analysis of 7 reveals C-N bond lengths typical for an ammonium salt (~ 1.50Å).⁴³



Figure 3: ORTEP representations of the unit cell components of 1 and 7.

UV-Vis and Fluorescence Spectroscopy

As expected, all dansyl QAC derivatives analysed in MeOH exhibit nearly identical absorption spectra ($\lambda_{Abs max} = 340$ nm, Figure 4) and fluorescence emission spectra ($\lambda_{Em max} = 516$ nm, Figure 5) characteristic of the green-yellow dansyl tag.³⁵ Only compound **6a** exhibited a major absorption wavelength outside typical values with $\lambda_{Abs max}$ centered closer to ~ 300 nm. This is likely the result of strong π - π * absorption of the benzophenone group. In aqueous environments at low pH (0.1 N HCl), fluorescence quenching of the dansyl fluorophore **2** was observed likely due to protonation of the aromatic *N*,*N*-dimethylamine moiety (see ESI[±], Figure S84). At pH 7 (phosphate buffer), no quenching was observed and a characteristic dansyl fluorescence spectra, similar to the fluorescence in MeOH solutions, was observed. At high pH (0.1 N NaOH), the absorption was shifted to ~ 310 nm resulting in a lower intensity emission at 516 nm when excited at 340 nm (see ESI[±], Figures S83 and S84).





Fig. 4 Absorption spectra of compounds **1-7** in MeOH (λ_{Abs} max = 340 nm).



Fig. 5 Fluorescence spectra of compounds 1-7 in MeOH (λ_{Em} max = 516 nm).

Coating Experiments and the Grafting Mechanism

Functional quaternary ammonium dansyl fluorescent tags attach to selective porous and non-porous surfaces via a self-assembly process. Upon application, either by dip or spray coating, anchors from the reactive dansyl molecule presumably form bonds with complementary functional groups on the substrate surface, thereby immobilizing an assumed monolayer of the compound on the surface. Both silane, 2, and phosphonic acid, **3c**, dansyl compounds can form monolayers on –OH and metal oxides surfaces respectively after a thermal curing step. In the process, H₂O is eliminated as the by-product resulting in M-O-Si or M-O-P (where M = Titanium, Stainless steel) to the surface. If improperly cured, the monolayer may easily become displaced with a rinsing step because it is non-covalently bound to the surface. The benzophenone 6a-h and vinyl 7 dansyl compounds require UV light to initiate binding (Figure 6 c-d). Excitation by UV can create radicals within the compounds and/or at the substrate surface that facilitate grafting (Figure 6cd).



Fig. 6 Mechanism of grafting (a) silanes on polyhydroxy surfaces; (b) phosphonic acids on metal oxide surfaces; (c) benzophenone and (d) allyl groups onto plastic (C-H) surfaces

Fluorescent coatings (Figure 7) of **2** on to cotton and silica nanoparticles (NP's) and **3c** on to stainless steel were prepared by dip coating or immersion of pre-cleaned samples in 0.01% (w/v) H₂O or MeOH solution of **2** or **3c** followed by heating (80° C, 5 min) and drying. Grafting of **6a** or **7** onto polypropylene (PP) and silicone was performed by first precleaning of all plastic samples (3.5×2.5 cm rectangles) by rinsing in distilled H₂O, followed by immersion in a MeOH bath and finally air drying prior to use. The clean substrates were then dipped into a 0.01% (w/v) EtOH solution of **6** or **7** and irradiated with UV for 2.5 min. Any unbound material was then rinsed off from substrates using distilled H₂O prior to visualization with UV light.



Fig.7 Coating procedure of dansyl QAC tags onto porous and non-porous surfaces.

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Fig.8 Images of A) medical grade silicone tubing coated with **6a** under UV light (control to the left), B) silica NP's with control left and treated with **2** right, C) polypropylene sample UV coated with **6a** and stained with bromophenol blue, D) polypropylene UV coated with **6a** under UV light, E) Stainless steel control top and a **3c** coated sample bottom under visible light and F) under UV light.

The dansyl fluorescent coatings **2**, **3c**, and **6a** were evaluated on a number of porous and non-porous surfaces (Figure 8). All fluorescent coatings adhered to the surfaces without leaching, as evident by the evaluation of the washings for fluorescence active residue.

Conclusions

Novel dansyl-functionalized QAC's bearing various chemical anchors specific for grafting onto porous and non-porous surfaces were successfully prepared via a Menschutkin reaction with the dansyl amine, 1. Compounds 2-7 were readily purified by trituration with Et2O without resorting to column chromatography and fully characterized by NMR (1H, 13C, 29Si (2), ³¹P (3a-3c, 4)), HRMS, UV-VIS and fluorescence spectroscopy. X-ray crystal structures were obtained for fluorophores (1, 7). The dansyl-silane 2 was prepared on a large scale and grafted onto textiles and silica NP's. Dansyl phosphonic acid 3c was successfully grafted to stainless steel coupons, while the dansyl QAC 6a containing the benzophenone photoactive crosslinker was used to coat plastic surfaces (PP, silicone). The inclusion of a trace amount of covalently bound dansyl molecules to surfaces treated with an antimicrobial QAC coating is expected to provide a simple and useful way of determining product identity and coverage quality.

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Notes and references

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Electronic Supplementary Information (ESI) available: This ESI contains the (a) NMR (¹H, ¹³C, and where identified, ²⁹Si, ³¹P NMR data) for all synthesized compounds, (b) DART or ESI-TOF MS spectra for new compounds (c) X-ray crystallographic data for compounds 1 and 7. Structure data was also deposited with the Cambridge Crystallographic Data Centre for compounds 1 (CCDC 971464) and 7 (CCDC 971465). See DOI: 10.1039/b000000x/

References

- 1 L. Ferreira, and A. Zumbuehl, J. Mater. Chem., 2009, 19, 7796-7806.
- 2 D. Campoccia, L. Montanaro, P. Speziale, and C. R. Arciola, *Biomaterials*, 2010, **31**, 6363-6377.
- 3 S. L. Warnes, C. J. Highmore, and C. W. Keevil, *mBio*, 2012, **3**, e00489-12.
- 4 D. Lebeaux, A. Chauhan, O. Rendueles, and C. Beloin, *Pathogens*, 2013, 2, 288-356.
- 5 K. Page, M. Wilson and I. P. Parkin, J. Mater, Chem., 2009, 19, 3819-3831.
- 6 J. C. Tiller, C.-J. Liao, K. Lewis, and A. M. Klibanov, *PNAS*, 2001, 98, 5981-5985.
- 7 A. M. Klibanov, J. Mater. Chem., 2007, 17, 2479-2482.
- 8 A. J. Isquith, E.A. Abbott, and P.A. Walters, *Appl. Microbiol.*, 1972, 24, 859-863.
- 9 J. Amalric, P. H. Mutin, G. Guerrero, A. Ponche, A. Sotto and J.-P. Lavigne, J. Mater. Chem., 2009, 19, 141-149.
- 10 P. Thebault, E. Taffin de Givenchy, R. Levy, Y. Vandenberghe, F. Guittard and S. Géribaldi, *Eur. J. Med. Chem.*, 2009, **44**, 4227-4234.
- 11 P. E. Thebault, R. Taffin de Givenchy, Y. Levy, F. Vandenberghe, F. Guittard and S. Géribaldi, *Eur. J. Med. Chem.*, 2009, 44, 717-724.
- 12 P. H. Mutin, J. Amalric, G. Guerrero, A. Ponche, J.-P. Lavigne, Eur Cell Mater., 2011, 21, 63.
- 13 J. C. Tiller, Antimicrobial Surfaces, *Bioactive Surfaces*, Advances in Polymer Science, Springer, Berlin / Heidelberg, 2011, 1-25.
- 14 V. P. Dhende, S. Samanta, D. M. Jones, I. R. Hardin and J. Locklin, ACS Appl. Mater. Interfaces, 2011, 3, 2830-2837.
- 15 H. Han, J. Wu, C. W. Avery, M. Mizutani, X. Jiang, M. Kamigaito, Z. Chen, C. Xi and K. Kuroda, *Langmuir*, 2011, **27**, 4010-4019.

ARTICLE

- 16 F. Costa, I. F. Carvalho, R. C. Montelaro, P. Gomes and M. C. L. Martins, *Acta Biomater.*, 2011, 7(4), 1431-1440.
- 17 A. D. Fuchs and J. C. Tiller, Angew. Chem. Int. Ed., 2006, 45, 6759-6762.
- 18 H. Murata, R. R. Koepsel, K. Matyjaszewski and A. J. Russell, *Biomaterials*, 2007, 28, 4870-4879.
- 19 K. Lewis and A. M. Klibanov, Trends Biotechnol., 2005, 23, 343-348.
- 20 Z. Li, D. Lee, X. Sheng, R. E. Cohen and M. F. Rubner, *Langmuir*, 2006, **22**, 9820-9823.
- 21 C. J. Ioannou, G. W. Hanlon, S. P. Denyer, Antimicrob. Agents Chemother., 2007, 51, 296-306.
- 22 M. Diz, A. Manresa, A. Pinazo, P. Erra and M.R. Infante, J. Chem. Soc. Perkin Trans. 2.,1994, 8, 1871-1876.
- 23 K. J. Stanger, J.-J. Lee, B. D. Smith, J. Org. Chem., 2007, 72, 9663-9668.
- 24 J. D. Reinheimer, J. D. Harley and W. W. Meyers, J. Org. Chem., 1963, 28, 1575-1579.
- 25 N. Menshutkin, Ann. Chem. Pharm., 1865, 133, 317-320.
- 26 M. Andresen, P. Stenstad, T. Møretrø, S. Langsrud, K. Syverud, L.-S. Johansson and P. Stenius, *Biomacromolecules*, 2007, 8, 2149-2155.
- 27 H. Nikawa, K. Ishida, T. Hamada, T. Satoda, T. Murayama, T. Takemoto, M. Tamamoto, H. Tajima, S. Shimoe, H. Fujimoto, *Dent. Mater. J.*, 2005, 24, 570-582.
- 28 S. M. Abou El Ola, R. Kotek, W. C. White, J. A. Reeve, P. Hauser and J. H. Kim, *Polymer*, 2004, **45**, 3215-3225.
- 29 R. R. Egan, J. Amer. Oil Soc. Chem., 1978, 55, 118-121.
- 30 Y. H. Li, L. M. Chan, L. Tyler, R. T. Moody, C. M. Himel and D. M. Hercules, J. Amer. Chem. Soc., 1975, 97, 3118-3126.
- 31 V. Tharmaraj and K. Pitchumani, J. Mater. Chem. B, 2013, 1, 1962-1967.
- 32 L. Ding and Y. Fang, Chem. Soc. Rev., 2010, 39, 4258-4273.
- 33 L. Mu, W. Shi, G. She, J. C. Chang and S.-T. Lee, Angew. Chem. Int. Ed., 2009, 48, 3469-3472.
- 34 B. Alkhouri, R. A. Denning, P. K. Chiaw, P. D. W. Eckford, W. Yu, C. Li, J. J. Bogojeski, C. E. Bear and R. D. Viirre, *J. Med. Chem.*, 2011, **54**, 8693-8701.
- 35 S.-M. He, Y. Luo, B. Hove-Jensen, D. L. Zechel, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 5954-5957.
- 36 H. R. Kricheldorf and R. Burger, *Makromol.Chem.*, 1993, **194**, 1197-1207.
- 37 A. Mocella, UV Cured Benzophenone Terminated Quaternary Ammonium Antimicrobials for Plastics, BSc. Thesis, Ryerson University, April 2011.
- 38 M.F. Saettone, C. Alderigi, B. Giannaccini, C. Anselmi, M.G. Rossetti, M. Scotton, R. Cerini, *Int. J. Cos. Sci.*, 1988, 10, 99-109.
- 39 R. Acosta, M. Garcia, A. Rosales, C. Gonzalez, L. Berlanga, G. Arias, and N. S. Allen, *Polym. Degrad. Stab.*, 1996, **52**, 11-17.
- 40 K. Mistry, New Self-assembling Quaternary Ammonium Fluorescent Compounds, BSc. Thesis, Ryerson University, April 2011.
- 41 X. Wang and H. Schneider, J. Chem. Soc. Perkin Trans. 2., 1998, 6, 1323-1328.
- 42 H. Deng, Vinyl Terminated Quaternary Ammonium Antimicrobials for Plastics and Metals, BSc. Thesis, Ryerson University, April 2013.
- 43 F. H. Allen, O. Kennard, D. G. Watson, L. Brammer, A. G. Orpen, R. Taylor, J. Chem. Soc. Perkin Trans. 2, 1897, 2, S1–S19.