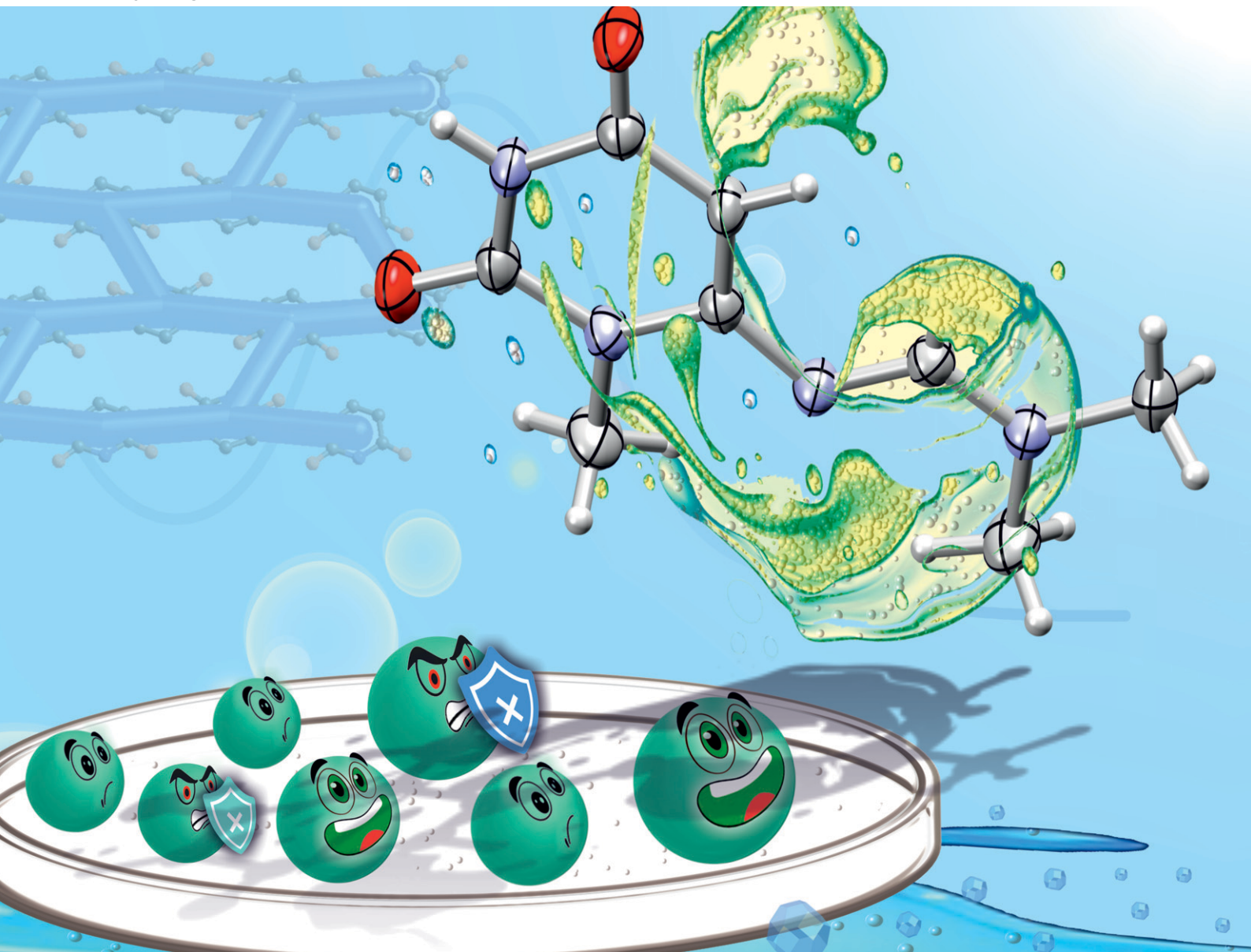


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**PAPER**

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Exploring uracil derivatives: synthesis, crystal structure  
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## Exploring uracil derivatives: synthesis, crystal structure insights, and antibacterial activity†

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As one of four nucleobases of RNA, uracil and its analogues represent an important class of bioactive pyrimidine derivatives. Their molecular arrangements in the solid state can be explored from the crystal engineering approach to obtain an understanding of structure–bioactivity correlations. In the present study, a series of uracil derivatives (compounds 1–4) was synthesized and fully characterized. The effect of the functionalization of the uracil core with different  $-NCHN(CH_3)_2$ ,  $-CH_3$ ,  $-Cl$ ,  $=S$ ,  $-NH_2$ , and  $-CH_2-COOH$  groups on stability, solubility, and antibacterial activity was investigated. The single-crystal structures of these compounds show that the hydrogen bonds formed by distinct synthons ( $R_2^2(8)$ ,  $R_4^4(12)$ ,  $C_1^1(6)$ ) contribute to framework stability. The presence of water molecules in the lattice is an important feature, as they provide additional H-bonding interactions that influence lattice energy and solubility. Lattice energy minimization, Hirshfeld surface analysis, and 2D fingerprint plots were employed to investigate intermolecular interactions and the stability of the obtained uracil derivatives, particularly the effect of functional groups. Although all compounds exhibit antibacterial behavior, the derivatives with small polar functional groups revealed an enhanced activity against Gram-negative bacteria, while the compounds with moderately polar substituents are more active against Gram-positive bacteria. The established discussions expand the comprehension of uracil chemistry and highlight the relationship between crystal structure and the resulting properties of the compounds, thus contributing to the rational development of new antibacterial agents.

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## Introduction

As one of the four nucleobases in RNA, uracil and its analogues form a crucial group of bioactive pyrimidine derivatives.<sup>1,2</sup> These are often functionalized at different positions of the pyrimidine ring, resulting in a variety of molecular arrangements (Scheme 1). Investigating these arrangements in the solid state can give insights into the relationship between structure and bioactivity.<sup>3–8</sup> In particular, uracil derivatives feature a broad range of bioactivity<sup>1,9–15</sup> that makes them appealing for exploring chemical and biological properties, aiming at the development of potential antiviral,<sup>16</sup> antitumor,<sup>17</sup> antiplasmodial,<sup>18</sup> antidiabetic,<sup>19</sup>

and antibacterial<sup>20,21</sup> molecules. However, the therapeutic application of uracil derivatives is still limited because of toxicity levels in several human body systems,<sup>10</sup> thus inspiring the synthesis of new derivatives with improved pharmacological and pharmacokinetic properties.<sup>1,10</sup>

Furthermore, the physicochemical and biological significance of these compounds is profoundly influenced by their capacity to establish hydrogen bonds and other non-covalent interactions, such as halogen bonding and  $\pi$ -stacking interactions.<sup>22–31</sup> Among the various pyrimidine bases, uracil derivatives exhibit both hydrogen bond donor and acceptor sites, thus acting as robust H-bonded synthons.<sup>22</sup> Despite being weaker than most covalent bonds,<sup>32,33</sup> hydrogen bonds are essential to consider in crystal engineering, given their prevalence in biological systems. Many aspects of biochemistry are a result of these weak interactions in an aqueous environment,<sup>32</sup> where H-bonding often determines the self-organization of organic molecules such as proteins, ribonucleic acid (RNA), and deoxyribonucleic acid (DNA).<sup>31</sup>

The elucidation of molecular arrangements in the solid state *via* crystal engineering has become a valuable tool, particularly for pyrimidine and uracil derivatives.<sup>34–36</sup> Distinct supramolecular synthons can stabilize crystal packing through diverse intermolecular interactions, enabling the design of

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**Scheme 1** Functionalization of uracil to derivatives 1–4. 1: (*E*)-*N,N*-dimethyl-*N'*-(3-methyl-2,6-dioxo-1,2,3,6-tetrahydropyrimidin-4-yl)formimidamide, C<sub>8</sub>H<sub>12</sub>N<sub>4</sub>O<sub>2</sub>. 2: 6-chloro-3-methylpyrimidine-2,4(1*H*,3*H*)-dione, C<sub>5</sub>H<sub>5</sub>N<sub>2</sub>O<sub>2</sub>Cl. 3: 2-(5-methyl-2,4-dioxo-3,4-dihydropyrimidin-1(2*H*)-yl)acetic acid, C<sub>7</sub>H<sub>8</sub>N<sub>2</sub>O<sub>4</sub>. 4: 6-amino-2-thioxo-2,3-dihydropyrimidin-4(1*H*)-one, C<sub>4</sub>H<sub>5</sub>N<sub>3</sub>OS.

materials with desirable properties.<sup>37–40</sup> Given the presence of uracil derivatives in some drugs, further diversification of uracil skeletons and synthesis of new molecules with prospective bioactivity is a current focus of synthetic chemistry, particularly with regard to fine-tuning of the physicochemical properties including solubility and stability.<sup>41–43</sup>

Inspired by our interest in uracil chemistry<sup>44–46</sup> and development of new antimicrobial compounds,<sup>47</sup> in the present work we focused on the synthesis, detailed structural characterization and antibacterial activity of several uracil derivatives (Scheme 1). The single-crystal X-ray structures of two new products, C<sub>8</sub>H<sub>12</sub>N<sub>4</sub>O<sub>2</sub>·H<sub>2</sub>O (**1**) and C<sub>7</sub>H<sub>8</sub>N<sub>2</sub>O<sub>4</sub>·2H<sub>2</sub>O (**3**), were determined and compared to related compounds such as uracil,<sup>48</sup> C<sub>5</sub>H<sub>5</sub>N<sub>2</sub>O<sub>2</sub>Cl (**2**),<sup>36</sup> and C<sub>4</sub>H<sub>5</sub>N<sub>3</sub>OS·H<sub>2</sub>O (**4**).<sup>49</sup> The molecular Hirshfeld surface analysis and energy frameworks were used to verify the differences in the crystal structures and establish possible correlations between structure and bioactivity. This work widens the family of bioactive uracil derivatives.

## Experimental

### Synthesis and crystallization

**Compound 1.** Compound **1** was synthesized by adapting a previously reported method.<sup>50</sup> 1-Methyl-6-aminouracil (1 mmol) and *N,N*-dimethylformamide dimethyl acetal (DMF-DMA, 1.1 mmol) were dissolved in *N,N*-dimethylformamide (DMF, 5 mL), and the resulting mixture was stirred at 40–50 °C for 24 h (Scheme 2). Then, the obtained mixture was filtered off and washed with diethyl ether. The crude product of **1** was dried in an oven and used for further characterization (yield: 94%). To obtain single crystals, **1** (0.25 mmol) was dissolved in warm methanol (10 mL). The resulting solution was filtered off to remove an insoluble part and kept for crystallization. After several days, the white colorless crystals appeared in the

solution and were collected manually for X-ray diffraction analysis. Melting point: 259–262 °C; <sup>1</sup>H-NMR (400 MHz, deuterated dimethyl sulfoxide, DMSO-*d*<sub>6</sub>) δ: 10.62 (s, 1H), 8.03 (s, 1H), 4.97 (s, 1H), 3.22 (s, 3H), 3.11 (s, 3H), 2.99 (s, 3H); <sup>13</sup>C-NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ: 163.61, 161.24, 156.26, 152.33, 82.71, 40.76, 34.86, 29.10.

**Compound 2.** 6-Chloro-3-methyluracil (0.25 mmol) was added into a methanol–water solvent mixture (8–2 mL) and heated until almost complete dissolution occurred. Subsequently, the resulting solution was subjected to filtration, and the filtrate was left in a vial for crystallization in air. Light yellow crystals of **2** emerged in a few days, which were collected manually for further characterization (yield: 72%).<sup>36</sup>

**Compound 3.** Compound **3** was synthesized by modifying a previously reported method.<sup>51</sup> In brief, KOH (4 mmol) was combined with thymine (1 mmol) in distilled water (5 mL). An aqueous solution of chloroacetic acid (1.5 mmol, 2 mL) was added and the resulting mixture was kept at 40 °C for 4 h (Scheme 3). It was then cooled, acidified with concentrated hydrochloric acid (HCl), and kept overnight in a refrigerator at 4 °C, producing white crystals of **3**. These were filtered off, washed with water, and dried in a vacuum oven to give **3** (yield: 89%). Melting point: 272–275 °C; <sup>1</sup>H-NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ: 13.09 (s, 1H), 11.32 (s, 1H), 7.48 (s, 1H), 4.35 (s,



**Scheme 2** Synthesis of compound **1**.





Scheme 3 Synthesis of compound 3.

2H), 1.73 (s, 3H);  $^{13}\text{C-NMR}$  (101 MHz,  $\text{DMSO-d}_6$ )  $\delta$ : 170.13, 164.83, 151.45, 142.26, 108.80, 48.85, 12.36.

**Compound 4.** The recrystallization of 4 was carried out in a similar way to that of 2, but using 6-amino-2-thiouracil (0.25 mmol) instead of 6-chloro-3-methyluracil. After several days, white colorless crystals emerged, which were collected manually and dried (yield: 63%).<sup>49</sup>

### Single-crystal X-ray diffraction

X-ray data for compounds 1 and 3 were acquired at room temperature (298 K) using a Rigaku Oxford Diffraction SuperNova diffractometer equipped with a high sensitivity EosS2 CCD detector and a Mo-K $\alpha$  radiation ( $\lambda = 0.71073 \text{ \AA}$ ). CrysAlisPro (ver. 1.171.40.69a, 2020)<sup>52</sup> program package was used for data collection, data reduction, and absorption correction. For compound 2, the X-ray data were collected on a Bruker AXS APEXIII diffractometer with PHOTON II CCD detector using Cu-K $\alpha$  radiation ( $\lambda = 1.54178 \text{ \AA}$ ). Data collection and reduction were performed using APEXIII<sup>53</sup> and SAINT.<sup>54</sup> SADABS was used for absorption corrections.<sup>55</sup> X-ray data for compound 4 were collected on a Bruker D8 Venture Diffractometer with PHOTON II detector using a Mo-K $\alpha$  radiation ( $\lambda = 0.71073 \text{ \AA}$ ) at room temperature (298 K). CrysAlisPro (ver. 1.171.40.69a, 2020) program package was used for data treatment and correction.<sup>52</sup> With Olex2<sup>56</sup> as the graphical interface, the crystal structures of all compounds were solved with the SHELXS structure solution program using direct methods (SHELXS). The structures were refined with the SHELXL<sup>57</sup> refinement package using the least squares minimization on  $F^2$ . All non-hydrogen atoms were refined anisotropically, while hydrogen atom positions were calculated geometrically and placed in idealized positions. Methyl hydrogen atoms were constrained by  $\text{C-H} = 0.96 \text{ \AA}$  and  $U_{\text{iso}}(\text{H}) = 1.5U_{\text{eq}}(\text{C})$ ; however, H atoms connected to nitrogen centers were constrained by  $\text{N-H} = 0.86 \text{ \AA}$  and  $U_{\text{iso}}(\text{H}) = 1.2U_{\text{eq}}(\text{N})$ . In the crystal structure of 1, a water molecule was found to be disordered over two positions. The disorder was modeled by splitting the oxygen atom into two sites, O3 and O3A, with fixed occupancies of 0.50 each, corresponding to a 50:50 ratio. The hydrogen atoms associated with each water position (H3A, H3B for O3; H3AA, H3AB for O3A) were placed in idealized positions and refined with distance restraints ( $\text{O-H} \approx 0.84 \text{ \AA}$ ,  $\text{H}\cdots\text{H} \approx 1.36 \text{ \AA}$ ) and angle constraints ( $\text{H-O-H} \approx 104.5^\circ$ ). The Mercury program<sup>58</sup> was used to draw the molecules, crystal packing, and motifs.

Crystal data and refinement parameters for compounds 1–4 are summarized in Table S1, ESI,<sup>†</sup> CCDC codes: 2306291 (1) and 2306292 (3).

### Hirshfeld surface analysis and energy frameworks

For initial geometry optimization, the experimental crystal structures of uracil (CSD code URACIL)<sup>48</sup> and its derivatives were obtained from crystallographic information files (cif files). The positional disorder in compound 1 was preserved as modeled throughout the single-crystal X-ray refinement before the structure was loaded into CrystalExplorer.<sup>59</sup> All model energies were calculated using molecular electron densities derived from wavefunctions computed by the Gaussian09 suite.<sup>60</sup> The intermolecular interactions were investigated using Hirshfeld surface<sup>61,62</sup> and 2D fingerprint plots,<sup>63,64</sup> in addition to all molecular clusters' calculations, with a radius ranging from 3.8 to 22  $\text{\AA}$ . These calculations were performed at the B3LYP/6-31G(d,p) level of theory using the Crystal Explorer package.<sup>59</sup> The energy frameworks and lattice energy were also calculated with the B3LYP/6-31G(d,p) level of theory, and lattice energies<sup>65</sup> were considered convergent when they did not present consistent changes of less than  $1 \text{ kJ mol}^{-1}$ , with the variation of the cut-off radius of interactions between pairs of molecules (convergence data can be found in the ESI,<sup>†</sup> Table S2, Fig. S1 and S2).

### Solubility studies

The solubility of compounds 1–4 was evaluated following a previously described procedure.<sup>35</sup> Distinct concentrations of each compound were prepared to establish a calibration curve (concentration–absorbance graph based on a  $\lambda_{\text{max}}$  value) employing UV-visible spectroscopy (Shimadzu UV-1780 spectrophotometer). Simultaneously, by adding an excess of each component to 2 mL of distilled water (pH = 6.8) and stirring for 24 h at room temperature, saturated solutions of each compound were prepared. Following this step, the resulting solution underwent centrifugation for 15 min. The clear solution obtained was subsequently diluted by a factor of 1000. The absorbance of the supernatant solution was evaluated, and then the concentration of each compound was calculated from the calibration curve. The solubility was determined by multiplying the concentration of the resulting solution by a factor of 1000. (detailed data can be found in Fig. S15 and S16, ESI<sup>†</sup>).

**Antibacterial Activity Investigation:** compounds 1–4, each at a consistent concentration, were dissolved in MilliQ-grade water. Prior to experimentation, all bacterial cells were subjected to purification through the streaking method to obtain single, isolated colonies. These isolated colonies were cultured in Luria Bertani (LB) broth for 7–8 h, depending on the microorganism under investigation. A known quantity of each compound was introduced into a fresh LB broth, along with a fixed number of bacterial cells, followed by incubation at 37  $^\circ\text{C}$ . Concurrently, a fixed concentration of cells was introduced into LB agar media to facilitate visual inspection



of bacterial growth. Optical density (OD) measurements were taken at 600 nm after 8–9 h of incubation.

## Results and discussion

### Structural features and solid-state studies

In this work, all uracil analogues are pyrimidine derivatives having a heterocyclic aromatic ring and a ketone group along with different substituents ( $-\text{NCHN}(\text{CH}_3)_2$ ,  $-\text{CH}_3$ ,  $-\text{Cl}$ ,  $=\text{S}$ ,  $-\text{NH}_2$ , and  $-\text{CH}_2-\text{COOH}$  groups), which represent the core pharmacophore of several drugs.<sup>19</sup> Four uracil derivatives were synthesized (**1–4**, Scheme 1) and their molecular structures (Fig. 1a–d) were investigated using single-crystal X-ray diffraction and theoretical approaches (Hirshfeld surface, 2D-finger plots, electrostatic potential isosurface, energy frameworks, and lattice energy) to identify and compare the effects of different functional groups.

Compound **1** ( $\text{C}_8\text{H}_{20}\text{N}_4\text{O}_2 \cdot \text{H}_2\text{O}$ ), a mono-methyl-amino substituted uracil, crystallizes in a triclinic system and space group  $P\bar{1}$ . There is some disorder that affects only one water molecule across the inversion center with a ratio of 50:50 at 0.7 Å distance. Thus, it is expected that the local energy minima are not affected by significant differences in total lattice energy. The mono-methyl-acid substituted uracil derivative, compound **3** ( $\text{C}_7\text{H}_{16}\text{N}_2\text{O}_4 \cdot 2\text{H}_2\text{O}$ ), crystallizes in the same crystallographic system (Fig. 1). Two water molecules present in the crystal structure of **3** are responsible for extending the H-bonding network. The presence of  $-\text{COOH}$  group is evident from the difference between the C–O distance of carboxylate group, being also in agreement with

the FTIR data ( $1703\text{ cm}^{-1}$ ). The protected amino group in **1** is practically planar to the uracil ring orientation, with the torsion C3–C4–N3–C5 angle of  $-8.64^\circ$ . However, this angle in **3** is  $110.78^\circ$  (C4–N2–C6–C7) for the substituted acid group, which is potentially a result of intermolecular H-bonds. The bonding parameters for compounds **1–4** are listed in Tables S5 and S7 (ESI†). The mono-methyl-choro derivative **2**, ( $\text{C}_5\text{H}_5\text{N}_2\text{O}_2\text{Cl}$ ), and the amino-thiooxo derivative **4**, ( $\text{C}_4\text{H}_5\text{N}_3\text{OS} \cdot \text{H}_2\text{O}$ ) crystallize in a system of high symmetry (monoclinic) with the space groups  $P2_1/c$  and  $C2/c$ , respectively (Fig. 1, Table S1†). The substitution of functional groups produces minor changes in the main uracil ring. The C=C double bond lengths in **1–4** range between 1.340 and 1.378 Å, and agree with the distance in uracil (1.340 Å).<sup>48</sup>

The molecular electrostatic potential isosurface map for the compounds **1–4** can be utilized for predicting non-covalent interactions.<sup>22</sup> The isosurface was calculated at the B3LYP/6-31G(d,p) level of theory using the Crystal Explorer package.<sup>59,66</sup> From the surface data in Fig. 1e–h, it is possible to note for **1** that the N1 atom exhibits an increased tendency to donate a proton (as indicated by the blue color). In contrast, the ketone oxygen atoms, shown in red, tend to accept a proton, according to the electrostatic potential representation. In compound **3**, the same tendency is observed. However, the  $-\text{COOH}$  group tends to donate and accept protons from water molecules in the crystal lattice. For compounds **2** and **4**, the acceptor atoms show a negative electrostatic potential with Cl atoms involved in donor sites with methyl-substituted groups. Intriguingly, the sulfur atom demonstrates a small partial region to accept protons (red



Fig. 1 Molecular structures of compounds (a) **1**, (b) **2**, (c) **3**, and (d) **4**, including atom labelling scheme. Thermal ellipsoids are drawn at the 50% probability level. H atoms were omitted for clarity. Molecular electrostatic potential isosurfaces for crystal structures of (e) **1**, (f) **2**, (g) **3**, and (h) **4**. The electron density surface drawn at  $\pm 0.05$  a.u. contour. The quantum mechanical properties obtained using the B3LYP/6-31G(d,p) wave function.





Fig. 2 Crystal packing diagrams of (a) **1** and (b) **2** showing the supramolecular  $R_2^2(8)$  and  $C_1^1(6)$  synthons (highlighted in light blue) and the arrangement of molecules.

color). Based on these data, the pyrimidinone ring protons show positive electrostatic potential and probably contribute to the non-covalent interactions with the available sites. This trend is similar in all analyzed crystal structures.

Because of differences in space group configurations, water molecules content, and unit cell features, each compound has a distinct crystal packing that is influenced by the electrostatic potential of the molecules and H-bonds.<sup>27,67</sup> Considering only short-range contact D–H···A type hydrogen bonds [ $H\cdots A \leq 2.75$  Å,  $D\cdots A \leq 3.50$  Å, and  $\angle(D-H\cdots A) > 120^\circ$ ], the main structural motif in **1** is a  $R_2^2(8)$  synthon (Fig. 2a), formed by molecular dimers with the N–H···O type hydrogen bonds [ $d_{N1\cdots O2} = 2.8658(19)$  Å]. There is a 1D zig-zag chain along the [201] direction through long-range contact H-bonds derived from the disordered water molecule [ $d_{O\cdots O} = 2.956(5)$  and  $2.637(11)$  Å]. As a result, the independent 1D chains are arranged into column motifs stabilized by H-bonding interactions (Fig. S10, ESI†). Compound **3** shows two main structural motifs – the synthons  $R_2^2(8)$  and  $R_4^4(12)$  produced by molecular dimers [ $d_{N1\cdots O2} = 2.860(3)$  Å] and water molecules [ $d_{O6\cdots O1} = 2.858(3)$  Å and  $d_{O6\cdots O2} = 2.917(3)$  Å] (Fig. 3a). These discrete units are connected by three types of H-bonds, resulting in the development of 2D sheets that are stacked by non-classic long-range H-bonds (C–H···O); all observations are consistent with electrostatic isosurfaces. Consequently, this compound tends to be more soluble in polar solvents, since these larger distances result in weaker and more dispersed interactions in the lattice. Differently, the structure of **2** has no crystallization solvent molecules and propagates in a 1D chain parallel to the *b*-axis by  $C_1^1(6)$  synthons [ $d_{N1\cdots O2} = 2.785(5)$  Å] as shown in Fig. 2b. In contrast, in the compound **4**, the molecules are arranged in parallel 1D column motifs along the *c*-axis and connected by the N–H···O hydrogen bonds [ $d_{N2\cdots O1} = 2.7364(17)$  Å and  $d_{N3\cdots O1} = 3.136(2)$  Å] in bifurcated mode with  $C_1^1(6)$  synthons (Fig. 3b). The interactions involving the  $NH_2$  group are not significant. The water molecules (trifurcated mode) connect the perpendicular 1D column motifs and establish a 3D network *via*



Fig. 3 Crystal packing diagrams of (a) **3** and (b) **4** showing the supramolecular  $R_2^2(8)$  and  $R_4^4(12)$  synthons in **3** and the  $C_1^1(6)$  synthon in **4** (highlighted in light blue), as well as the arrangement of molecules. Inset represents water molecules in trifurcate mode.

the O–H···O and N–H···O hydrogen bonding [ $d_{N1\cdots O2} = 2.8528(19)$  Å,  $d_{N3\cdots O2} = 3.109(2)$  Å and  $d_{O2\cdots O1} = 2.7632(19)$  Å] (Fig. S11, ESI†). Although the crystallographic data suggest a weak O–H···S interaction due to the long contact distance [ $d_{O2\cdots S1} = 3.4889(15)$  Å], the electrostatic potential surface reveals that such interaction is important for stabilization of the crystal structure.

The Hirshfeld surface (HS) analysis and 2D-fingerprint plots (2D-FP) were used to assess how the presence of different functional groups can affect the physicochemical properties and biological activity of compounds in the present study. The results were compared to the uracil precursor. In addition, energy framework and lattice energy parameters were used to get further insight on the molecular interactions and to predict some properties. Fig. 4 displays the HS mapped over  $d_{norm}$ , shape index, and 2D-fingerprint plot for the uracil molecule. The HS reveals the electron density accumulation and depletion regions around each atom, providing information on the nature of intermolecular interactions in the crystal structure.<sup>68,69</sup> The intense red spots over the  $d_{norm}$  surface indicate the short intermolecular contacts, which are less than the sum of the vdW (van der Waals) radii of interacting atoms.<sup>37,69</sup> The blue regions relate to longer contacts, and the white regions refer to the vdW contacts. Uracil shows three H-bonds of the N–H···O type with intense red areas ( $d_{norm}$ ), which represent a main  $R_6^6(32)$  synthon generated by molecular dimers (Fig. S12, ESI†), leading to the growth of 2D sheets stacked by weak interactions from pyrimidinone ring (Fig. 4b, red and blue triangular shapes above the rings) as expected to electrostatic potentials isosurface (Fig. S13†).

By examining the 2D-FP, it is possible to obtain the contribution of each type of interaction in stabilizing the crystal packing.<sup>63,69,70</sup> Most interactions in uracil involve symmetric N–H···O contacts (55%). In contrast, van der Waals interactions



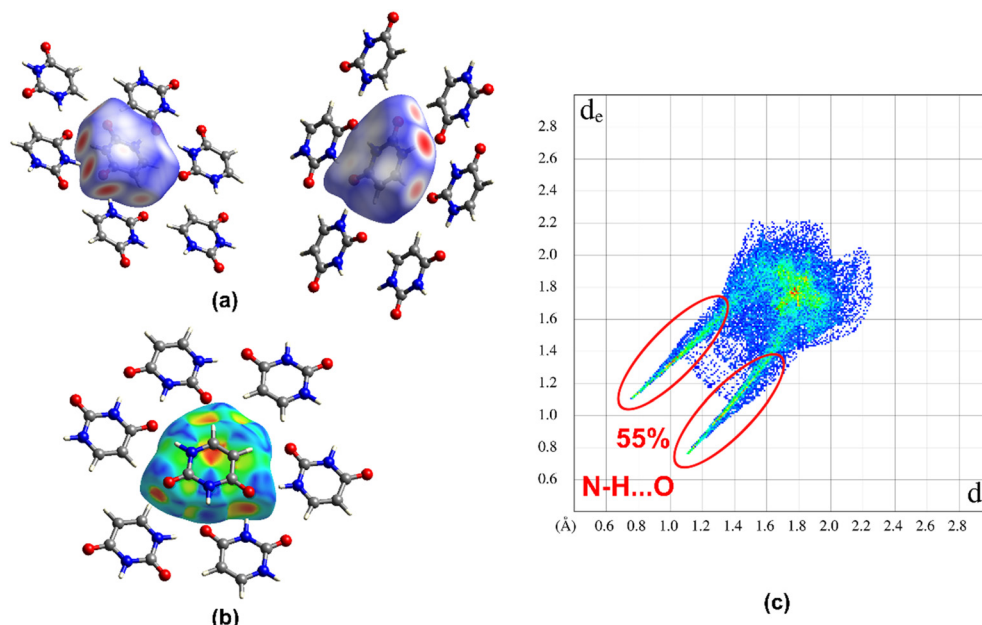


Fig. 4 Hirshfeld surface analysis of uracil. (a) Perpendicular view showing three short intermolecular contacts (red spots). (b) Shape index plots surface, proving the stacking of uracil molecules by weak pyrimidinone ring interactions. (c) 2D-fingerprint plot of N-H...O contacts.

represented by H...H contacts account for 15.9% of total interactions. These two types of interactions represent 70.9% of total interactions. The functionalization of the pyrimidinone ring *via* the incorporation of -CH<sub>3</sub> and -NCHN(CH<sub>3</sub>)<sub>2</sub> groups in **1** lead to a reduction of crystallographic symmetry and a change of the distribution of electron density in the main ring and, consequently, in all interactions. The 2D-FP of **1** reveals that the predominant interactions are H...H (46.2%), followed by the N-H...O and C-H...O hydrogen bonds (28.2%); these three interactions correspond to 70.4% of total interactions similar those of uracil (Fig. 5a). Disordered water molecules are present in the lattice and slightly influence on the stabilization of crystal packing (long-range D-H...A distance, Table S7†). The C...H and C...C contacts originating from methyl group show a secondary role in stabilizing the supramolecular structure of **1** (10.1% and 2.2%, respectively). In the methyl-chloro analogue (compound **2**), two N-H...O and C-H...O types of H-bonds display intense red spots with 29.4% of interactions (Fig. 5b). Nonetheless, a much longer intermolecular C-H...O intermolecular contact shows pale red spots. Contacts involving the Cl atom contribute to 20.9% of all contacts. In contrast to **1**, the water molecules in **3** are associated into a 2D sheet by the O-H...O and N-H...O hydrogen bonding along with C-H...O interactions (44.6% of contribution), thus directly affecting the stability of network (bright red spots, Fig. 5c). The contribution of the H...H contacts (vdW interactions) from methyl group is 34.9%. Particularly, the contribution of O...H contacts is increased (~20%) due to the presence of different substituents in **3** if compared to **1**. In **4**, the S...H contacts account for 19.5% of interactions. They derive from the -NH<sub>2</sub> functional group (Fig. 5d). The N-H...O interactions are more significant (26.4%) and responsible for the extension of crystal structure along the

*c*-axis. The H...H contacts present due to the incorporation of methyl group in **4** account for 24.3% (more details for 2D-FP see Fig. S14†).

Intermolecular interactions<sup>37</sup> can affect various properties of molecular compounds. The stability of **1-4** can be evaluated *via* lattice energy minimization ( $E_{\text{lattice}}$ ).<sup>65,71,72</sup> The lattice energies can be estimated by the direct sum of the total interaction energies between pairs *i* and *j* ( $E_{\text{lattice}} = 0.5 \sum_{R_{ij} < R} E_{\text{total}}^{ij}$ ), until the lattice energy converted to a limit of 1 kJ mol<sup>-1</sup>. All these calculations were performed in Crystal Explorer,<sup>59</sup> using a radius range of 3.8 to 22 Å (details about minimization process can be found in ESI†), and the calculated total energy values for all crystal structures are displayed in Scheme 4. The obtained values are consistent with those available in the literature for comparable compounds.<sup>43,72</sup> The presence of water in the lattice is seen as an essential feature, since H-bonds involving H<sub>2</sub>O molecules provide additional intermolecular interactions, which can increase the lattice energy (Table S2, ESI†). According to Scheme 4, the stability order (kJ mol<sup>-1</sup>) of compounds is as follows: **4** > **3** > **1** > uracil > **2**.

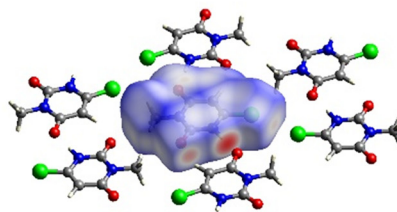
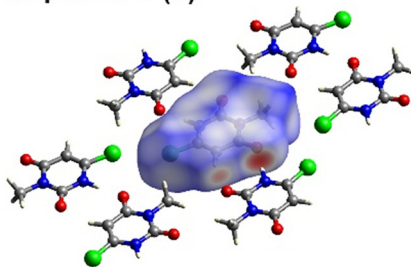
The analysis of the results reveals that the type of functional group affects the stability of uracil derivatives **1-4**, influencing on how the electrostatic potential on the isosurface is distributed. While in uracil the major contribution to the lattice energy is derived from the energies of H-bond interactions (N-H...O motifs), in compound **1** the dispersion energy assumes a greater contribution, directly affecting the solubility values. The presence of solvated water facilitates the organization of organic molecules and their stabilization by long-range H-bonds, which decreases the lattice energy and reduces solubility. As for **2**, the modification of uracil structure reduces the dimensionality and concentrates on the greatest energy contribution in a single



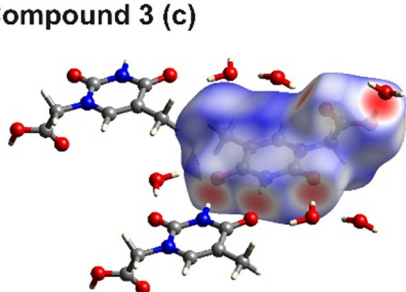
## Compound 1 (a)



## Compound 2 (b)



## Compound 3 (c)



## Compound 4 (d)



Fig. 5 Hirshfeld surfaces with short intermolecular contacts (red spots) in (a) 1, (b) 2, (c) 3, and (d) 4 (two perpendicular views). In the right column, 2D fingerprint plots of main contacts are represented.

direction along the crystallographic  $c$ -axis. The interactions caused by the presence of  $-Cl$  substituent (29%) present only a small contribution to total stability. Compound 3 features carboxylic acid group that affects the charge distribution, with Coulomb energy donating the most. It is worth noting that in 4, despite the formation of infinite 1D chains *via* intermolecular interactions, crystallization water molecules form H-bonds in a trifurcated mode, which provides 30% to the overall value of the lattice energy ( $-57.7 \text{ kJ mol}^{-1}$ ) and favors the formation of stable lattice. Despite the  $N-H\cdots O$  interactions are responsible for the formation of dimers between molecules of all the analyzed compounds (preferential binding mode is the double

$C=O\cdots H-N$  hydrogen bond). Consequently, in energy framework distribution, it is possible to notice that H-bonds from water molecules significantly influence the crystal lattice energy by contributing to the overall stability of the lattice and affecting the packing arrangement of molecules.

#### Antibacterial properties

Antibacterial activity of 1–4 was investigated using the pre-grown colonies of *E. coli*<sup>73</sup> (Gram-negative) and *S. aureus*<sup>74</sup> (Gram-positive), following the previously described protocol.<sup>75</sup> The percentage of bacterial growth inhibition by 1–4 as well as





**Scheme 4** Lattice energy profile and simplified energy frameworks for uracil and derivatives 1–4. Total framework interaction energies (blue) are shown as cylindrical tube size with 150 u.a. and cut-off 15 kJ mol<sup>-1</sup>.

solubility data are summarized in Table 1. Additional details on the solubility tests (Fig. S15 and S16) and antibacterial screening (Tables S8 and S9, Fig. S17–S24) are given in ESI.† The inhibitory activity (in %) of compounds 1–4 was determined at maximum concentrations of 5.6, 4.7, 12.5, and 2.2  $\mu\text{g mL}^{-1}$  relative to negative control (water), and results are shown in Fig. 6. A comparison of the inhibitory activity of 1–4 suggests that the inhibition of bacterial activity is not proportional to the concentration of uracil derivatives in solution (Table 1). The compounds show a varying degree of bacterial growth inhibition. The presence of different functional groups influences the nature of non-covalent interactions, which may affect the antibacterial activity. Gram-positive and Gram-negative bacteria respond differently to uracil derivatives, depending on the size and polarity of the incorporated functional groups. It reveals a correlation between the incorporation of functional groups and the electrostatic potential of the pyrimidine ring. Small polar functional groups enhance the activity against Gram-negative bacteria (compound 4), suggesting a favorable interaction with the bacterial cell surfaces. In contrast, the introduction of both larger and smaller polar groups with greater density dispersion favors

antibacterial activity against Gram-positive bacteria (compound 1) in the C<sub>4</sub> position.

Although compounds 1 and 4 exhibit only a small difference in lattice energy (10.57 kJ mol<sup>-1</sup>), they display opposite trends in the wavenumber of their main absorption bands. This suggests that hydrogen bonding may influence the electronic structures of the compounds and contribute to the observed bathochromic effect (Table S4, Fig. S9†). The bathochromic shift observed in compound 1 can be attributed to enhanced charge delocalization (Fig. 7), resulting in a partial positive charge on nitrogen atom (N4). The resulting resonance-stabilized system promotes coplanarity between the formimidamide moiety and the adjacent heterocycle, stabilizing the electronic structure and shifting the absorption band to lower energy. A similar delocalization pattern is observed in compound 4, which is consistent with the X-ray crystallographic data. The resembling structural features were observed in both compounds 1 and 4—specifically, the elongation of the C=C bond and the shortening of the C–C(=O) bond if compared to 2 and 3. Moreover, the variation in spectroscopic and structural properties observed among the compounds can

**Table 1** Solubility of 1–4 in water and their inhibitory activity against *E. coli* and *S. aureus*

Compound	$\lambda_{\text{max}}$	Saturation solubility ( $\mu\text{g mL}^{-1}$ )	Inhibition (%)	
			( <i>E. coli</i> )	( <i>S. aureus</i> )
1	305	5.6	17.8	12.9
2	263	4.7	15.9	3.3
3	272	12.5	31.1	7.1
4	275	2.2	12.9	2.1





Fig. 6 Images of Petri dishes revealing the antibacterial activity of compounds **1** (a and b), **2** (c and d), **3** (e and f), and **4** (g and h) against *E. coli* (a–g) and *S. aureus* (b–h). Maximum concentrations (saturation solubility) of **1–4** were used. Conditions are those of Table 1.



Fig. 7 Resonance and charge delocalization in compounds **1** and **4**.

also be further clarified by differences in hydrogen bonding and van der Waals interactions.

Notably, while the N–H...O and C–H...O interactions contribute similarly in both compounds, the H...H interactions exhibit a ratio of approximately 2 : 1, proportionally influencing the solubility and antibacterial activities of the compounds. In the case of **3**, the introduction of carboxylic acid functionality, which promotes the formation of H-bonds within the crystal lattice, results in a reduction of the lattice energy compared to compound **2** and uracil. However, the antibacterial activity of compound **3** is negatively influenced by these modifications. When comparing the solubility, **3** is the most soluble, as expected since it presents a 2D dispersion of H-bond interactions as previously described. Nonetheless, it displays a lower inhibition in contrast to the concentration used. While lattice energy of **2** exhibits values of  $-112.7 \text{ kJ mol}^{-1}$ , it is essential to note that this compound exists in an anhydrous form. The organization of H-bond interactions [ $C_1^1(6)$  synthons] plays an important role in stabilizing this structure, resulting in reduced solubility. The primary contributor to bioactivity remains the functional group rather than the solvation effects. These observations provide an elucidation of the key role of functional groups in customizing uracil derivatives for antibacterial applications.

## Conclusions

In the current work, we reported a simple synthetic procedure for two new uracil derivatives, **1** and **3**, as well as succeeded in the crystallization of two other uracil analogs, **2** and **4**. All compounds were analyzed using standard techniques (single crystal X-ray diffraction, FTIR, UV-vis and NMR spectroscopy). The structural details were further emphasized using Hirshfeld surface analysis, energy frameworks and 2D-fingerprint plots.

Despite the structural differences and space groups, the structures of compounds **1**, **2** and **4** reveal the formation of 1D H-bonded motifs, while compound **3** presents a 2D supramolecular arrangement with a greater dispersion of interactions. The presence of H-bonds increases the lattice energy, especially for compounds with crystallization water molecules (**1**, **2**, and **4**), where the arrangement of H-bonds can also affect the solubility. Furthermore, the antibacterial activity of these compounds against Gram-negative (*E. coli*) and Gram-positive (*S. aureus*) bacteria were evaluated, and the presence of different functional groups within the uracil core appeared to be closely associated with antibacterial activity. Small polar functional groups tend to increase activity against Gram-negative bacteria (compound **4**), while the introduction of larger and smaller polar groups with greater density dispersion (compound **1**) favors antibacterial activity against Gram-positive bacteria. Despite having greater solubility in aqueous medium, compound **3** presents a lower antibacterial activity.

The interaction of these compounds with bacterial cell surfaces is influenced by variations in the electrostatic potential of the pyrimidine ring driven by changes in functional groups and the distribution of non-covalent interactions in the structure. Hence, this work not only expands our knowledge about pyrimidine and uracil derivatives but also contributes to the rational design of new bioactive uracil derivatives.

## Data availability

The authors confirm that the data supporting the findings of this study are available within the article and included in the ESI.† Crystallographic data for the compounds have been deposited at the CCDC under CCDC numbers 2306291 and 2306292 and can be freely obtained from <https://www.ccdc.cam.ac.uk/structures/>.

## Conflicts of interest

There are no conflicts to declare.

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## References

- H. U. Rashid, M. A. U. Martines, A. P. Duarte, J. Jorge, S. Rasool, R. Muhammad, N. Ahmad and M. N. Umar, *RSC Adv.*, 2021, **11**, 6060–6098.
- O. Michalak, P. Cmoch, P. Krzeczynski, M. Cybulski and A. Leś, *Org. Biomol. Chem.*, 2019, **17**, 354–362.
- K. Kajal, R. Shakya, M. Rashid, V. Nigam, B. D. Kurmi, G. D. Gupta and P. Patel, *Sustainable Chem. Pharm.*, 2024, **37**, 101374.
- J. Ye, H. Pei, D. Liu, F. Wang, Y. Yu, J. Zhang and L. Zhang, *Tetrahedron Lett.*, 2024, **142**, 155101.
- K. N. Jarzemska, M. Kubsik, R. Kamiński, K. Wozniak and P. M. Dominiak, *Cryst. Growth Des.*, 2012, **12**, 2508–2524.
- B. Cui, B. Yan, K. Wang, L. Li, S. Chen and Z. Zhang, *J. Med. Chem.*, 2022, **65**, 12747–12780.
- P. Chakraborty and P. Dastidar, *Chem. Commun.*, 2019, **55**, 7683–7686.
- M. Katkevics, J. A. MacKay and E. Rozners, *Chem. Commun.*, 2024, **60**, 1999–2008.
- L. Baziar, L. Emami, Z. Rezaei, A. Solhjoo, A. Sakhteman and S. Khabnadideh, *Sci. Rep.*, 2024, **14**, 26538.
- A. Pałasz and D. Cież, *Eur. J. Med. Chem.*, 2015, **97**, 582–611.
- K. Jain, S. Sharma and D. Utreja, *ChemistrySelect*, 2024, **9**, e202303179.
- T. Das, P. Kumar, S. Kumar, S. Mal, S. Kumar, V. K. Rajana, S. Singh, A. Dasgupta, D. Mondal and S. Das, *ACS Infect. Dis.*, 2024, **10**, 4314–4326.
- C. Zhi, Z. Y. Long, A. Manikowski, N. C. Brown, P. M. Tarantino, K. Holm, E. J. Dix, G. E. Wright, K. A. Foster, M. M. Butler, W. A. LaMarr, D. J. Skow, I. Motorina, S. Lamothe and R. Storer, *J. Med. Chem.*, 2005, **48**, 7063–7074.
- V. A. Kezin, E. S. Matyugina, M. S. Novikov, A. O. Chizhov, R. Snoeck, G. Andrei, S. N. Kochetkov and A. L. Khandazhinskaya, *Molecules*, 2022, **27**, 2866.
- S. Djaković, L. Glavaš-Obrovac, J. Lapić, S. Maračić, J. Kirchofer, M. Knežević, M. Jukić and S. Raić-Malić, *Appl. Organomet. Chem.*, 2021, **35**, e6052.
- M. S. Novikov, M. P. Paramonova, I. M. Kirillov, O. A. Smirnova, A. L. Khandazhinskaya, A. O. Chizhov, S. N. Kochetkov, A. V. Ivanov and I. F. Fedyakina, *Mendeleev Commun.*, 2023, **33**, 537–538.
- G. Q. Lu, X. Y. Li, O. K. Mohamed, D. Wang and F. H. Meng, *Eur. J. Med. Chem.*, 2019, **171**, 282–296.
- H. Cui, G. F. Ruda, J. Carrero-Lérida, L. M. Ruiz-Pérez, I. H. Gilbert and D. González-Pacanowska, *Eur. J. Med. Chem.*, 2010, **45**, 5140–5149.
- D. Ramesh, B. G. Vijayakumar and T. Kannan, *Eur. J. Med. Chem.*, 2020, **207**, 112801.
- Shivangi, S. Das, S. Mal and S. Naskar, *ChemistrySelect*, 2024, **9**, e202400715.
- S. Meshcheryakova, A. Shumadalova, O. Beylerli, I. Gareev, S. Zhao and J. Wu, *Drug Dev. Res.*, 2022, **83**, 578–585.
- L. H. Al-Wahaibi, S. R. S. Bysani, S. S. Tawfik, M. S. M. Abdelbaky, S. Garcia-Granda, A. A. El-Emam, M. J. Percino and S. Thamocharan, *Cryst. Growth Des.*, 2021, **21**, 3234–3250.
- G. Portalone, J. O. Moilanen, H. M. Tuononen and K. Rissanen, *Cryst. Growth Des.*, 2016, **16**, 2631–2639.
- A. O. Surov, A. P. Voronin, K. V. Drozd, T. V. Volkova, N. Vasilev, D. Batov, A. V. Churakov and G. L. Perlovich, *Cryst. Growth Des.*, 2022, **22**, 2569–2586.
- Y. Roselló, M. Benito, M. Barceló-Oliver, A. Frontera and E. Molins, *Cryst. Growth Des.*, 2021, **21**, 4857–4870.
- S. J. Zamisa, N. P. Ngubane, A. A. Adeleke, S. B. Jonnalagadda and B. Omondi, *Cryst. Growth Des.*, 2022, **22**, 5814–5834.
- M. Barceló-Oliver, C. Estarellas, A. García-Raso, A. Terrón, A. Frontera, D. Quiñonero, I. Mata, E. Molins and P. M. Deyà, *CrystEngComm*, 2010, **12**, 3758–3767.
- B. Makiabadi, M. Zakarianezhad and E. Zeydabadi, *Struct. Chem.*, 2023, **34**, 755–767.
- S. Jena, J. Dutta, K. D. Tulsian, A. K. Sahu, S. S. Choudhury and H. S. Biswal, *Chem. Soc. Rev.*, 2022, **51**, 4261–4286.
- P. W. Kenny, *J. Med. Chem.*, 2022, **65**, 14261–14275.
- M. Zhang, Z. Li, Y. Zhu, L. Yan, X. Zhong, Y. Zhang, Z. Li, Y. Bai and H. Li, *CrystEngComm*, 2024, **26**, 261–267.
- E. D. Głowacki, M. Irimia-Vladu, S. Bauer and N. S. Sariciftci, *J. Mater. Chem. B*, 2013, **1**, 3742–3753.
- T. Steiner, *Angew. Chem., Int. Ed.*, 2002, **41**, 48–76.
- V. Kumar, P. K. Goswami, R. Thaimattam and A. Ramanan, *CrystEngComm*, 2018, **20**, 3490–3504.
- M. Kumar, G. Jaiswar, M. Afzal, M. Muddassir, A. Alarifi, A. Fatima, N. Siddiqui, R. Ayub, N. A. Y. Abduh, W. S. Saeed and S. Javed, *Molecules*, 2023, **28**, 2116.
- V. Gerhardt and E. Egert, *Acta Crystallogr., Sect. B: Struct. Sci., Cryst. Eng. Mater.*, 2015, **71**, 209–220.
- L. F. Diniz, C. H. J. Franco, D. F. Silva, L. S. Martins, P. S. Carvalho Jr, M. A. C. Souza, N. F. A. Reis, C. Fernandes and R. Diniz, *Int. J. Pharm.*, 2021, **605**, 120790.
- A. E. Krajewski and J. K. Lee, Gas-phase experimental and computational studies of 5-halouracils: intrinsic properties and biological implications, *J. Org. Chem.*, 2021, **86**, 6361–6370.
- F. B. Ilhami, Y. S. Birhan and C. C. Cheng, *ACS Biomater. Sci. Eng.*, 2023, **10**, 234–254.
- F. Jubeen, A. Liaqat, F. Amjad, M. Sultan, S. Z. Iqbal, I. Sajid, M. B. K. Niazi and F. Sher, *Cryst. Growth Des.*, 2020, **20**, 2406–2414.
- R. K. Shayakhmetova, E. M. Khamitov, A. G. Mustafin, S. P. Ivanov and S. L. Khursan, *J. Chin. Chem. Soc.*, 2017, **64**, 143–151.
- J. L. Sessler and J. Jayawickramarajah, *Chem. Commun.*, 2005, 1939–1949.
- S. A. Barnett, A. T. Hulme, N. Issa, T. C. Lewis, L. S. Price, D. A. Tocher and S. L. Price, *New J. Chem.*, 2008, **32**, 1761–1775.



- 44 D. Ramesh, A. K. Mohanty, A. De, B. G. Vijayakumar, A. Sethumadhavan, S. K. Muthuvel, M. Mani and T. Kannan, *RSC Adv.*, 2022, **12**, 17466–17480.
- 45 E. A. Basiony, A. A. Hassan, M. Elsayalhy, A. H. Adel, H. Mansour, R. K. Arafa and N. A. Hassan, *Bioorg. Chem.*, 2025, **154**, 108066.
- 46 T. H. Abdalla, A. G. Ibrahim, S. Elabbady, E. Nassar, A. A. Hamed and A. Aboelnaga, *Int. J. Biol. Macromol.*, 2025, **300**, 140327.
- 47 T. A. Fernandes, F. Macedo, R. G. Cabral, T. Guiu, C. H. Franco, P. Jorge, A. C. Sousa, V. Andre, N. Cerca and A. M. Kirillov, *RSC Appl. Interfaces*, 2024, **1**, 98–109.
- 48 R. F. Stewart and L. H. Jensen, *Acta Crystallogr.*, 1967, **23**, 1102–1105.
- 49 S. Swaminathan and K. K. Chacko, *Acta Crystallogr., Sect. B*, 1978, **34**, 3108–3110.
- 50 K. Dzieszowski, I. Barańska and Z. Rafiński, *J. Org. Chem.*, 2020, **85**, 6645–6662.
- 51 G. Marafon, D. Mosconi, D. Mazzier, B. Biondi, M. De Zotti and A. Moretto, *RSC Adv.*, 2016, **6**, 73650–73659.
- 52 Rigaku, *CrysAlisPro Software System, 1.171.40.69a*, 2020.
- 53 APEXIII, *APEXIII crystallography software suite*, 2016.
- 54 Bruker, *Bruker AXS: SAINT+*, 6.22, 2016.
- 55 Bruker, *Bruker AXS: SADABS*, 2005.
- 56 O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard and H. Puschmann, *J. Appl. Crystallogr.*, 2009, **42**, 339–341.
- 57 G. Sheldrick, *Acta Crystallogr., Sect. C: Struct. Chem.*, 2015, **71**, 3–8.
- 58 C. F. Macrae, I. Sovago, S. J. Cottrell, P. T. A. Galek, P. McCabe, E. Pidcock, M. Platings, G. P. Shields, J. S. Stevens, M. Towler and P. A. Wood, *J. Appl. Crystallogr.*, 2020, **53**, 226–235.
- 59 C. F. Mackenzie, P. R. Spackman, D. Jayatilaka and M. A. Spackman, *IUCrJ*, 2017, **4**, 575–587.
- 60 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, *Gaussian 09 Revision A.2.*, 2009.
- 61 M. A. Spackman and D. Jayatilaka, *CrystEngComm*, 2009, **11**, 19–32.
- 62 P. A. Wood, J. J. McKinnon, S. Parsons, E. Pidcock and M. A. Spackman, *CrystEngComm*, 2008, **10**, 368–376.
- 63 A. Parkin, G. Barr, W. Dong, C. J. Gilmore, D. Jayatilaka, J. J. McKinnon, M. A. Spackman and C. C. Wilson, *CrystEngComm*, 2007, **9**, 648–652.
- 64 M. A. Spackman and J. J. McKinnon, *CrystEngComm*, 2002, **4**, 378–392.
- 65 S. P. Thomas, P. R. Spackman, D. Jayatilaka and M. A. Spackman, *J. Chem. Theory Comput.*, 2018, **14**, 1614–1623.
- 66 M. A. Spackman, *Z. Kristallogr. - Cryst. Mater.*, 2018, **233**, 641–648.
- 67 D. Ž. Veljković, V. B. Medaković, J. M. Andrić and S. D. Zarić, *CrystEngComm*, 2014, **16**, 10089–10096.
- 68 M. Ziemniak, S. Pawłędzio, A. Zawadzka-Kaźmierczuk, P. M. Dominiak, D. Trzybiński, W. Koźmiński, R. Zieliński, I. Fokt, W. Priebe, K. Woźniak and B. Pająk, *RSC Adv.*, 2022, **12**, 8345–8360.
- 69 M. S. Cunha, C. E. P. Ribeiro, C. C. Corrêa and R. Diniz, *J. Mol. Struct.*, 2017, **1150**, 586–594.
- 70 P. K. Deb, A. M. Menon, F. N. PSR, I. Shruti, S. Nidal, K. N. Venugopala and D. Chopra, *CrystEngComm*, 2025, **27**, 2070–2085.
- 71 M. K. Singh, *J. Cryst. Growth*, 2014, **396**, 14–23.
- 72 S. L. Price and K. S. Wibley, *J. Phys. Chem. A*, 1997, **101**, 2198–2206.
- 73 J. B. Kaper, J. P. Nataro and H. L. Mobley, *Nat. Rev. Microbiol.*, 2004, **2**, 123–140.
- 74 N. A. Turner, B. K. Sharma-Kuinkel, S. A. Maskarinec, E. M. Eichenberger, P. P. Shah, M. Carugati, T. L. Holland and V. G. Fowler Jr, *Nat. Rev. Microbiol.*, 2019, **17**, 203–218.
- 75 J. Campbell, *Curr. Protoc. Chem. Biol.*, 2010, **2**, 195–208.

