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A sodium alginate-based sustained-release IPN hydrogel and its applications

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Interpenetrating polymer network (IPN) hydrogels are crosslinked by two or more polymer networks, providing free volume space in the three-dimensional network structure, and providing conditions for the sustained and controlled release of drugs. The IPN hydrogels based on the natural polymer sodium alginate can form a stable porous network structure. Due to its excellent biocompatibility, the loaded drug can be sustained to the maximum extent without affecting its pharmacological effect. Sodium alginate-based IPN hydrogels have broad application prospects in the field of sustained and controlled drug release. This paper begins with an overview of the formation of alginate-based IPN hydrogels; summarizes the types of alginate-based IPN hydrogels; and discusses the pharmaceutical applications of alginate-based IPN hydrogels. We aim to give an overview of the research on IPN hydrogels based on sodium alginate in sustained and controlled drug release systems.

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Introduction

Hydrogels are colloids composed of a hydrophilic polymer with a high water content and a three-dimensional (3D) crosslinked networks structure,¹ widely used in tissue engineering, biomedicine and other fields.^{2–4} Traditional hydrogels have defects such as poor mechanical properties⁵ and single performance,⁶ and their application in biomedicine is significantly limited. In recent years, with the development of life sciences, people have submitted higher requirements for hydrogels. It is well-known that hydrogels can be widely used in the sustained and controlled release of drugs, and good biocompatibility and biodegradability are critical factors for their success.⁷

Interpenetrating polymer network (IPN) hydrogels are cross-linked by two or more polymer networks (such as chitosan, alginate, starch, polyacrylamide, polyacrylic acid, polyvinyl alcohol, *etc.*). In semi-IPN hydrogels, a network with linear polymers is embedded within the first network. IPN hydrogels contain two or more polymers, which can be cross-linked in the presence of each other to give a 3D network structure, producing a free volume for the easy encapsulation of drugs,⁸ and provide conditions for sustained and controlled release of drugs. IPN hydrogels also have mechanical strengthening,⁹ biodegradation kinetics,¹⁰ *in vivo* dual polymer activity,¹¹ new nanostructures¹² and other characteristics, which provide biocompatibility and convenience for molecules ranging from non-steroidal anti-inflammatory drugs (NSAIDs) to proteins and peptides and stable drug delivery systems.

Natural polymers have excellent biocompatibility and biodegradability, and are widely distributed and easily obtained.^{13,14} Among them, sodium alginate can be used as a matrix to prepare macroporous hydrogels.^{15,16} Sodium alginate is a linear block copolymer comprising 1,4-linked β -D-mannuronic and α -L-guluronic acid residues.¹⁷ It is considered to be non-toxic and biocompatible,¹⁸ and its compatibility with drugs does not affect the pharmacological effects, showing that it has more natural advantages in the sustained and controlled release of drugs. At present, scientists have conducted a lot of research on IPN hydrogels made from the natural polymer sodium alginate. This review discusses the formation and types of alginate-based IPN hydrogels, and their pharmaceutical application.

IPN hydrogels

IPNs are alloys of cross-linked polymers¹⁹ that are formed by interpenetrating entanglement of two or more cross-linked polymers. When only one polymer in the system is cross-linked, it is called a semi-IPN. IPNs are often created for the purpose of conferring key attributes of one of the components while maintaining the critical attributes of another. In some cases, entirely new, and sometimes surprising properties are exhibited by the IPN that are not observed in either of the two single networks alone.²⁰

IPNs and semi-IPNs used in biomedicine include films,²¹ microspheres²² and hydrogels.²⁰ They are used as scaffold materials for tissue engineering,²³ drug delivery systems²⁴ and in many other biomedical fields. IPN-based hydrogels have attracted the attention of researchers in the biomedical field

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Review

due to their excellent properties. Hydrogels prepared from IPNs have various properties, including better mechanical strength,²⁴ stability,²⁵ and stimulus responsiveness.²⁶

Sodium alginate is a polymer based on natural polysaccharides. In the presence of divalent cations, it exhibits unique gelation characteristics in aqueous solutions, but the gel is accompanied by shortcomings such as instability and fragility. It can be combined with polymers with temperature responsiveness, electric field responsiveness and other responsiveness, which can not only make up for the original defects, but also prepare various intelligent IPN hydrogels with the help of IPN technology. The fruitful precedent of combining sodium alginate with different polymers has promoted the development of IPN hydrogels, which have sustained and controlled release of drugs,²⁷ high porosity²⁸ and multiple responsiveness.^{28–30}

Formation of alginate-based IPN hydrogels

The main chain of sodium alginate contains a large number of $-\text{COO}^-$ reactive groups, and the anions in aqueous solution exhibit aggregation behavior, so it has some adhesiveness and can be used as a drug carrier for the treatment of mucosal tissues.³¹ The pH sensitivity of sodium alginate also originates from the $-\text{COO}^-$ group. Under acidic conditions, $-\text{COO}^-$ is converted to $-\text{COOH}$, and the hydrogen bond interaction between the carboxyl groups of sodium alginate molecules is enhanced, which causes the molecular chain to shrink.³² When the pH value increases, the $-\text{COOH}$ group tends to ionize and produce $-\text{COO}^-$.³³ At that time, the hydrophilicity of sodium alginate also increases,³⁴ so that the molecular chain can be extended.

Sodium alginate hydrogels have a wide range of applications in sustained and controlled release of drugs,^{35–37} and its pH sensitivity can be a key element of its selective release in the stomach.³⁸ Sodium alginate has unique gelation properties in aqueous solution in the presence of divalent cations.³⁹ The sodium ion of guluronic acid exchanges with the divalent cation, and the divalent cationic association proximal fraction belongs to carboxyl groups on different sodium alginate molecular chains to form a hydrogel with a cross-linked network structure.⁴⁰ Such a 3D network structure provides a large number of drug-loading sites⁴¹ and is used to manufacture various drugs, proteins and even cellular controlled release systems.^{42,43} Because sodium alginate can have extremely mild gelation conditions, it has the stability required for preparation of pharmaceutical excipients,⁴⁴ which also prevents the inactivation of pharmaceutically active substances.⁴⁵

Under normal circumstances, the burst release of drug molecules from sodium alginate hydrogels which formed by Ca^{2+} cross-linking (Fig. 1) is a common issue in the field of controlled drug delivery systems, leading to suddenly burst release.¹⁶ In recent years, a lot of work has been conducted to improve the performance of sodium alginate hydrogels as drug

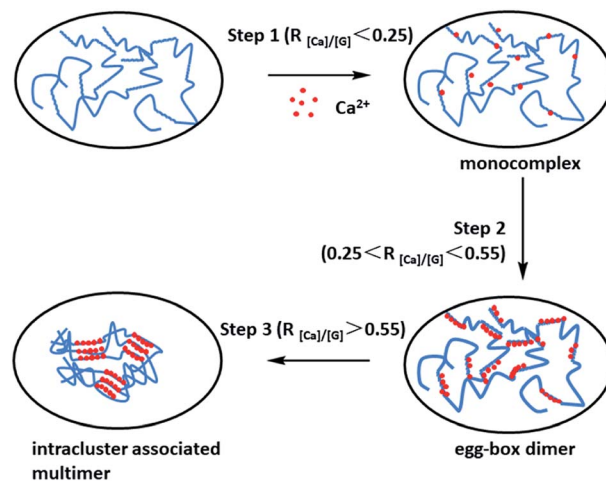


Fig. 1 Schematic drawing of the multiple-step binding of Ca^{2+} to alginate with a considerable chain length. The zigzag lines, smooth lines, and dots stand for G-block, M-block, and Ca^{2+} , respectively. $R_{[\text{Ca}]/[\text{G}]}$ is the feeding ratio of added Ca^{2+} to unit G residue. This figure has been adapted from ref. 50 with permission from American Chemical Society, Copyright © 2007.

carriers. Compounds of sodium alginate with other polymers such as polyvinyl alcohol (PVA),⁴⁶ gelatin,⁴⁷ or hydroxypropyl methyl cellulose,⁴⁸ and the coating of sodium alginate beads with cationic polymers such as chitosan has been found to improve the problems of drug leaking and encapsulation rate.⁴⁹ Cross-linking of sodium alginate and ovalbumin alone has also been successful.⁵⁰ The formation of hydrogels may be the product of one or more cross-linking methods.

IPN technology can use polymers with certain special properties to improve the deficiencies of sodium alginate. Taking sodium alginate/polyvinyl alcohol (PVA) hydrogels as an example, the PVA molecule is soft, and the sodium alginate gel is rigid, and the introduction of PVA can increase the elasticity of the gel.⁵¹

Jao⁵² *et al.* chose not to use chemical cross-linking agents but physical cross-linking-prepared semi-IPN hydrogel, which relied on Ca^{2+} cross-linking between alginate chains and hydrogen bonding interaction between PVA chains (Fig. 2). In

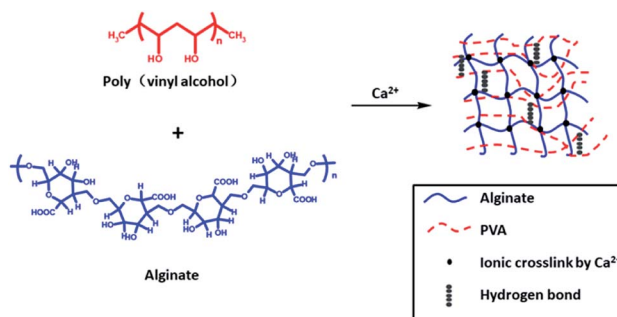


Fig. 2 Physically cross-linked sodium alginate-based semi-IPN hydrogel film formation mechanism. This figure has been adapted from ref. 52 with permission from John Wiley & Sons, Ltd., Copyright © 2008.





Fig. 3 Formation mechanism of chemically cross-linked sodium alginate-based IPN hydrogel. This figure has been adapted from ref. 46 with permission from Elsevier B.V., copyright © 2010.

such systems, the PVA network can provide mechanical enhancement and temperature sensitivity, while the alginate network can provide pH sensitivity. They showed that when the pH value is 4, the swelling is minimal, and the swelling ratio increases with pH value. The enthalpy of alginate/PVA hydrogel suddenly increases between 37 and 45 °C, which may be caused by hydrogen bond dissociation of alginate/PVA hydrogel. The elastic modulus and effective cross-linking density of the composite increases with increase of alginate content, which may be due to the increase of hydrophobic interaction and polymer interaction. Alginate/PVA hydrogels are sensitive to pH and temperature and can be invoked as controlled release systems in biomedical fields.

Kulkarni⁴⁶ and others used glutaraldehyde as a cross-linking agent to prepare sodium alginate/PVA IPN hydrogel membranes by chemical cross-linking (Fig. 3), which was used for the sustained release of the antihypertensive drug prazosin hydrochloride. Prazosin hydrochloride is a widely used drug for the treatment of hypertension and congestive heart failure that requires multiple doses to maintain blood levels of the drug. The IPN hydrogel membranes extend the drug release up to 24 h, while sodium alginate and PVA hydrogel membranes discharge the drug quickly. Cineole has shown greater enhancement in drug permeation. A study of skin irritation and skin histopathology has indicated that the prepared IPN hydrogel membranes are perfectly safe for transdermal drug delivery applications.

Boppana⁵³ *et al.* used a Ca^{2+} + glutaraldehyde double cross-linking method to prepare IPN microspheres with ketoprofen-preloaded, polyacrylamide-grafted gum ghatti and sodium



Fig. 4 Formation mechanism of sodium alginate-based IPN gel microspheres with double cross-linking of Ca^{2+} + GA. This figure has been adapted from ref. 53 with permission from Elsevier B.V., copyright © 2015.

alginate (Fig. 4). Ketoprofen is an NSAID currently being utilized to treat musculoskeletal and joint diseases. It is easily absorbed from the gastrointestinal tract and has a short half-life of only 2 h. After oral administration, gastric irritation, ulcer, bleeding and other adverse reactions occurred. Therefore, in order to effectively treat musculoskeletal diseases and patient compliance, IPN hydrogel microspheres prepared in this study were used to control the release of ketoprofen and avoid its release in the stomach. As IPN microspheres contain $-\text{COO}^-$ reactive groups, they are un-ionized in the stomach pH, leading to small amounts of drug release. In contrast, the microspheres undergo ionization in the intestinal pH, leading to greater drug release. *In vivo* experiments in Wistar rats also confirmed this conclusion. The pH-sensitive IPN microspheres blocked the release of drugs in the stomach and reduced ulcers, bleeding and mucosal erosion. The controlled release effect of ketoprofen was also confirmed. Therefore, IPN microspheres are useful for gastro-protective controlled delivery of NSAIDs.

Wen⁵⁴ *et al.* prepared IPN hydrogels composed of gelatin and sodium alginate by the double cross-linking method of enzymes and ions (Fig. 5). The gel can support cell adhesion and diffusion *in vitro*. Surprisingly, IPN hydrogels can even withstand traditional steam sterilization, which may seriously damage the morphology and mechanical properties of traditional hydrogels. The enzymatic and calcium cross-linking methods are straightforward, efficient and free of harmful and toxic chemicals. The results show that the sodium alginate IPN hydrogel



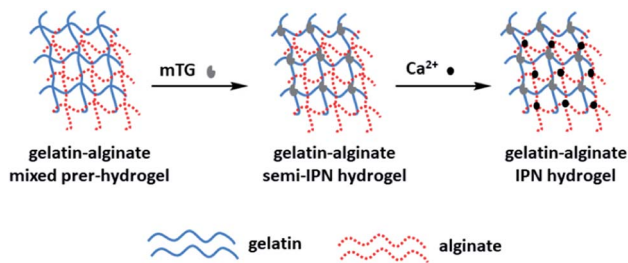


Fig. 5 Formation mechanism of sodium alginate-based IPN hydrogel with double cross-linked of Enzyme and ion. This figure has been adapted from ref. 54 with permission from WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim, copyright © 2014.

has excellent biocompatibility and robust mechanical properties, and provides an opportunity to be used as a tissue engineering scaffold.

Types of IPN hydrogels based on sodium alginate

In recent years, researchers have controlled the release of drugs by changing the structure of hydrogels, and can sense changes in environmental information such as temperature,⁵⁵ pH value,⁵⁶ or chemical substances⁵⁷ at the lesion site to achieve the effect of sustained and controlled release, increasing the safety of drug-loaded hydrogels, and other advantages. Although a single component sodium alginate hydrogel is pH sensitive, no matter what kind of cross-linking method is used, the sustained and controlled release of drugs is not ideal. Therefore, sodium alginate is generally used in combination with other polymer compounds, and researchers have prepared a series of smart IPN hydrogels with the help of IPN technology.

The pH-sensitive hydrogels

Khalid⁵⁸ *et al.* prepared PVA-grafted poly(acrylic acid)/sodium alginate semi-IPN hydrogel by chemical cross-linking, and the hydrogels supported loxoprofen sodium. The prepared hydrogels had a high pH sensitivity, and showed maximal swelling and drug loading and release at pH 7.4, and lower at pH 1.2. The biodegradability and biocompatibility of sodium alginate and poly(methacrylic acid) semi-IPN hydrogel has been used to control the release of theophylline.⁵⁹ Upadhyay and others⁶⁰ used the natural polymers locust bean gum and sodium alginate to develop an IPN sodium alginate hydrogel for capecitabine. *In vitro* drug release studies have shown sustained release for 12 h. Basu⁶¹ *et al.* developed a nano-silver composite semi-IPN hydrogel of sodium alginate and acrylamide, and explored its drug-release characteristics. The encapsulated nano-silver particles and ciprofloxacin molecules showed sustained release.

Due to the addition of the second component, the pH-responsive sodium alginate-based IPN hydrogels show better sustained and controlled release of drugs, as well as excellent mechanical strength.

Temperature/pH double-sensitive hydrogels

It can be seen from the above that sodium alginate hydrogels have pH sensitivity. However, single responsive hydrogels can no longer respond to the needs of contemporary medicine. IPN hydrogels prepared by various cross-linking methods from two or more non-interfering, single sensitive polymer monomers have double/multiple responsiveness. In addition to temperature changes,⁶² the focal site of patients with cancer or inflammation often has localized acidification. These two characteristics usually could be used for specific drug delivery.⁶³ Recent in-depth study of multiple responsive hydrogels has broadened the potential applications of smart hydrogels. Research hotspots are concentrated on the preparation of temperature/pH dual-response hydrogels and their applications in the field of biomedicine. This is because temperature and pH are important physiological parameters in human systems, and they are also easy to sense and control stimulus signals.^{64–67}

The IPN hydrogel formed by doxorubicin (DOX)-loaded sodium alginate and poly(*N*-isopropylacrylamide) copolymer showed sustained-release of the drug at physiological pH and temperature (Fig. 6). DOX released from the hydrogels is encapsulated in the micelles formed by the co-polymers, *i.e.*, the hydrogels release DOX-encapsulated micelles in a sustained manner that has been found to be effective in cancer treatment.²⁹ Eswaramma and others¹⁹ prepared IPN hydrogel microbeads based on sodium alginate and functionally modified guar gum. They showed that the microbeads may be sensitive to both pH and temperature. At 37 °C and pH 7.4, the release time of zidovudine (an anti-HIV drug) was extended to 34 h. Jalababu and others⁶⁸ used modified sodium alginate, *N*-isopropylacrylamide and acrylamide as raw materials, and adopted a single-step free radical polymerization reaction to cross-link with *N,N'*-methylenebisacrylamide to synthesize a new polymer. The pH/temperature dual-response type was used to control the IPN hydrogel released by the anticancer drug was extended to 48 h. Vasanta Madhavi and others⁶⁹ used locust bean gum and sodium alginate to prepare IPN hydrogels beads through glutaraldehyde cross-linking to study the controlled release of the anti-inflammatory drug nimesulide. *In vitro* dissolution experiments showed that nimesulide had prolonged release of 48 h.

The sustained release capability of these newly developed drug-loaded temperature/pH dual-response IPN hydrogels may

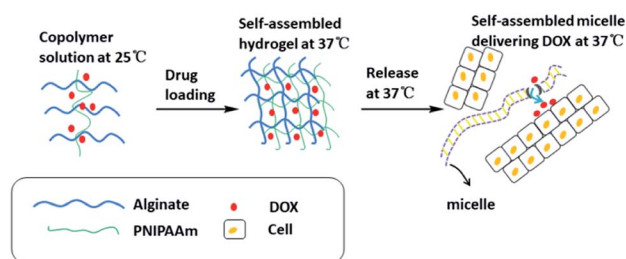


Fig. 6 Mechanism of temperature/pH double-sensitive IPN hydrogels based on sodium alginate release at physiological temperature. This figure has been adapted from ref. 29 with permission from American Chemical Society, copyright © 2017.



be advantageous because it can achieve the purpose of reducing the dosing interval and is beneficial to patient compliance. The temperature/pH dual-response IPN hydrogels can be a promising candidate preparation for targeted and sustained release of anti-cancer and anti-inflammatory drugs.

Electric field/pH double-sensitive hydrogels

Electric-field-sensitive hydrogels control the release of drugs by monitoring the on/off of electrical signals. Sodium alginate is a polymer polyelectrolyte, and its hydrogels are responsive to electrical stimulation.

Shi³⁰ *et al.* prepared a semi-IPN hydrogel based on bacterial cellulose nanofibers and sodium alginate. This semi-IPN is a double-stimulated drug release system. The system has the characteristics of swelling and drug release in response to pH and electric field stimuli. The swelling rate of sodium alginate bacterial cellulose increased from <8 times (pH 1.5) to >13 times when the pH value was 11.8. In addition, the swelling rate is impacted by the electric field. When the electric field changes from 0 to 0.5 V, the expansion rate of the semi-IPN system increases from 8 to 14 times in the dry state. This kind of IPN hydrogel uses the change in its swelling rate in the electric field to realize the electric field control of the drugs release. Researchers have used this semi-IP system to study the loading and release of the NSAID ibuprofen. The drug release can be controlled by deprotonation or protonation of sodium alginate in hydrogels under different pH conditions. The system demonstrates that the drug release is faster under alkaline conditions and slower under acidic conditions.

Salt/pH double-sensitive hydrogels

The characteristic of salt-sensitive hydrogels is that the swelling properties of the gel can undergo abrupt changes under the

action of added salt. Salt is one of the most important substances for human survival, and can regulate the osmotic balance between cells and blood as well as normal water and salt metabolism.

Yin²⁸ *et al.* synthesized an IPN super-porous hydrogel containing sodium alginate. The results show that these hydrogels have a large number of interconnected pores and IPNs at the same time. Their swelling rate is lower than that of super-porous hydrogels, with an increased sodium alginate/monomer ratio and lower. The *in situ* lactate dehydrogenase test was conducted on AD293 cells by the thiazole blue method, and morphological observation of the rat intestinal tissue was performed. The results showed that the polymer caused no obvious cell and mucosal damage to the rat intestinal tissue. The good news is that the rapid swelling of this hydrogel, superior mechanical properties, pH responsiveness, salt sensitivity and biocompatibility indicate that it is a potential candidate in the field of drug delivery systems.

Application of sodium alginate-based IPN sustained and controlled release hydrogels in pharmacy

Sodium alginate has the characteristics of degradability, good biocompatibility, non-toxicity, easy availability, low cost, and no immunogenicity, so that it has an excellent performance in the field of biomedicine. IPN sustained and controlled release hydrogels based on sodium alginate have been extensively studied for targeted drug and protein administration.

Delivery of small chemical drugs

Inherent pH sensitivity of sodium alginate makes it an ideal agent for colon-specific drug delivery. It has been used

Table 1 Sodium alginate-based IPN sustained and controlled release hydrogels and its latest progress in small chemical drug delivery

Route of administration	Components of IPN	Drugs	Entrapment efficiency	
Oral	Poly(vinyl alcohol), xanthan gum	Norfloxacin	83% (ref. 79)	
	Locust bean gum	Nimesulide	74% (ref. 69)	
	Locust bean gum, methacrylic acid	Simvastatin	70% (ref. 80)	
	Polyacrylamide, Gum ghatti	Ketoprofen	90% (ref. 53)	
	Locust bean gum	Capecitabine	81% (ref. 60)	
	Carboxymethyl cellulose	Melbine	73% (ref. 81)	
	<i>N</i> -Acryloyl-L-phenylalanine, ethylene glycol vinyl ether, hydroxyethyl acrylate	Curcumin	73% (ref. 82)	
	Locust bean gum	Capecitabine	84% (ref. 83)	
	Carboxymethyl locust bean gum	Glipizide	93% (ref. 84)	
	<i>O</i> -Carboxymethyl tamarind gum	Acyclovir	70% (ref. 85)	
	Functional modified guar gum	Zidovudine	68% (ref. 19)	
	Modified karaya gum	D-Penicillamine	71% (ref. 86)	
	Starch, poly(acrylic acid)	Diclofenac sodium	72% (ref. 87)	
	Acrylic acid-divinyl benzene	Ciprofloxacin	92% (ref. 88)	
	Karaya gum	Fluorine formamide	74% (ref. 89)	
	Subcutaneously	Poly(<i>N</i> -isopropyl acrylamide)	Adriamycin	— ²⁹
		Poly(ethylene glycol) Methacrylate, <i>N</i> -isopropyl acrylamide	Diclofenac sodium	— ⁹⁰
Transdermal	Poly(vinyl alcohol)	Prazosin hydrochloride	— ⁴⁶	



extensively as a drug carrier because of its ability to encapsulate and release multiple drugs in a gentle and biocompatible manner. After oral, subcutaneous, vascular, mucosal, pulmonary and transdermal administration of sodium alginate,^{70–76} biocompatibility *in vivo* has been verified. Sodium alginate-based IPN hydrogels can be used for continuous and local administration of traditional small chemical drugs. Traditional methods of administration (mostly oral administration) often result in the burst release of the drug, and poor targeting, leading to adverse effects and ineffectiveness. Oral preparations are now the most common and most preferred method in pharmaceutical applications. The design of oral dosage forms generally follows one of the following two principles: (1) the entire drug dose is in the same physical unit; or (2) the dose is encapsulated in an assembly of small subunits.⁷⁷ In the latter case, the subunits are further compressed into tablets, thereby creating a barrier that can provide controlled release properties. The sodium alginate hydrocolloid immediately hydrates, forming a high viscosity layer around it, acting as a diffusion barrier, thereby reducing the migration of small chemical drugs and maintaining drug delivery. By adjusting the drug-sodium alginate interaction, the release kinetics of low molecular weight drugs in the sodium alginate gel can be monitored. When there is no chemical reaction between the drug and the polymer, the release mainly depends on the charge polarization of the molecule; that is, the hydrophilic molecule diffuses quickly, while the hydrophobic drug diffuses slowly through the pores of the gel.⁷⁸ Table 1 summarizes the latest progress of sodium alginate-based IPN slow-release controlled hydrogels and their delivery in small chemical drugs.

It can be seen from the above table that oral dosing of the drug dosing way great proportion, because oral drug administration has the characteristics of convenience, economy and safety; however, short residence time of drugs in the gut limits the use of the oral route. The inherent pH responsiveness of sodium alginate makes them into an excellent agent for colon-specific drug delivery. The IPN between various functional polymers and sodium alginate gives hydrogels multiple responses, among which the temperature response is the most commonly used. They can be used as targeted control drug delivery systems. A large number of sodium alginate-based IPN hydrogels have been developed for controlled release of targeted drugs. The potential of sodium alginate IPN hydrogels as sustained drug delivery formulations has been demonstrated. Reports in the literature of alginate-based IPN hydrogels

indicate that the drug release time can be extended from several hours to several days.

Protein delivery

Route of administration. The inside of sodium alginate is almost an inert aqueous environment. Its organic solvent-free encapsulation process and high gel porosity at room temperature can achieve a high diffusion rate of macromolecules. Not only that, the porosity can be easily controlled through a simple procedure, and the system can be biodegraded under normal physiological conditions. These unique characteristics make it a matrix for the encapsulation and delivery of multiple protein drugs.⁹¹ In order to realize the sustained release of protein drugs, the protein diffuses out *via* the porosity. There have been some reports on the controlled release of proteins in sodium alginate-based IPN hydrogels. Table 2 summarizes them.

As with IPN hydrogels for delivery of small chemotherapeutic agents, oral delivery is still the main route for protein drug delivery, but in the oral delivery route, the drug must overcome several obstacles before getting into the systemic circulation. In the gastrointestinal tract, protein drugs can be affected by digestive enzymes. Sodium alginate has strong biological adhesion, which can help the contact time between drugs and the conjunctival mucosa. Many unique characteristics make it a substrate for the encapsulation and delivery of various protein drugs. Bovine serum albumin has served as a model drug to evaluate the drug delivery effect of hydrogels. Surprisingly, the drug's release lasted for about 15 days. These hydrogel controlled-release proteins have been recorded to validate their effectiveness as a continuous protein delivery system for improving patient compliance.

Pharmaceutical excipients. Momoh and others⁹⁶ conducted a study in which they delivered an IPN hydrogel dressing made of sodium alginate and glycerin containing therapeutic proteins to the wound, and found that the protein could be released for more than 72 h. This dressing also shows ideal moisture content, which is required for protein conformation, and shows a good balance between flexibility and toughness.

Tissue engineering. Pescosolido⁹⁷ *et al.* have developed an IPN hydrogels based on a physical network of alginate, interpenetrated with a chemical one based on hydroxyethyl-methacrylate-derivatized dextran. In about 15 days, bovine serum albumin was gradually released from the hydrogel. They showed that these IPN hydrogels had the potential to serve as a scaffold for the proliferation and differentiation of encapsulated cells.

Table 2 Sodium alginate-based IPN sustained and controlled release hydrogels and its latest progress in protein delivery

Route of administration	Components of IPN	Protein	Entrapment efficiency
Oral	Sodium acrylate, <i>N</i> -vinylpyrrolidone	BSA	— ⁹²
	L-Arginine	BSA	— ⁹³
	Carboxymethyl chitosan	BSA	— ¹⁶
Subcutaneously	Poly(ethylene glycol) methacrylate, <i>N</i> -isopropyl acrylamide	BSA	— ⁹⁰
	Dextran methacrylate derivative	Myoglobin	— ⁹⁴
Pulmonary	Modified <i>N</i> -phthalyl chitosan	BSA	90% (ref. 95)



Conclusions

Alginate-based IPN hydrogels have been extensively used for the delivery of drugs and proteins or as basic materials for tissue repair and regeneration. They have potential research applications in the field of sustained and controlled drug release.

However, at present, most of the sodium alginate hydrogels are still limited to the theoretical research stage in the laboratory. Their actual application still needs to be improved considerably. Not only that, sodium alginate is highly hydrophilic, which results in poor sustained and controlled release of water-soluble drugs. Single-response and dual-response sodium alginate-based IPN hydrogels have been reported in a large number of studies, and most of them are only for one drug. Today, the treatment of diseases often involves the synergy of multiple drugs or some diseases are subject to a considerable variety of complications. The development of multiple-response intelligent sodium alginate hydrogels that can release multiple drugs accurately and in an orderly manner, and how to improve the targeting of drug-loaded hydrogels will be important directions in future research.

Conflicts of interest

The authors declare no conflict of interest.

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Notes and references

- 1 S. Li, S. Dong, W. Xu, S. Tu, L. Yan, C. Zhao, J. Ding and X. Chen, *Adv. Sci.*, 2018, **5**, 1700527.
- 2 Y. Zhang, J. Yu, K. Ren, J. Zuo, J. Ding and X. Chen, *Biomacromolecules*, 2019, **20**, 1478–1492.
- 3 C. Wang, N. Feng, F. Chang, J. Wang, B. Yuan, Y. Cheng, H. Liu, J. Yu, J. Zou and J. Ding, *Adv. Healthcare Mater.*, 2019, **8**, 1900312.
- 4 D. S. Lee, J. I. Kang, B. H. Hwang and K. M. Park, *Macromol. Res.*, 2019, **27**, 205–211.
- 5 R. Na, Y. Liu, N. Lu, S. Zhang, F. Liu and G. Wang, *Chem. Eng. J.*, 2019, **374**, 738–747.
- 6 G. Wu, K. Jin, L. Liu and H. Zhang, *Soft Matter*, 2020, **16**, 3319–3324.
- 7 Y. Wang, Z. Jiang, W. Xu, Y. Yang, X. Zhuang, J. Ding and X. Chen, *ACS Appl. Mater. Interfaces*, 2019, **11**, 8725–8730.
- 8 H. Qiu, H. Guo, D. Li, Y. Hou, T. Kuang and J. Ding, *Trends Biotechnol.*, 2020, **38**, 579–583.
- 9 S. Park, S. Edwards, S. Hou, R. Boudreau, R. Yee and K. J. Jeong, *Biomater. Sci.*, 2019, **7**, 1276–1280.
- 10 J. Liao and H. Huang, *Cellulose*, 2020, **27**, 825–838.
- 11 G. G. de Lima, J. K. Elter, B. S. Chee, W. L. E. Magalhães, D. M. Devine, M. J. Nugent and M. J. de Sá, *Biomed. Mater.*, 2019, **14**, 054101.
- 12 T. Tominaga, V. R. Tirumala, E. K. Lin, J. P. Gong, H. Furukawa, Y. Osada and W.-l. Wu, *Polymer*, 2007, **48**, 7449–7454.
- 13 Z. Shi, X. Gao, M. W. Ullah, S. Li, Q. Wang and G. Yang, *Biomaterials*, 2016, **111**, 40–54.
- 14 Y. Jiang, Y. Wang, Q. Li, C. Yu and W. Chu, *Curr. Med. Chem.*, 2019, **27**, 2631–2657.
- 15 C. Chang, B. Duan and L. Zhang, *Polymer*, 2009, **50**, 5467–5473.
- 16 Y. Hu, J. Peng, L. Ke, D. Zhao, H. Zhao and X. Xiao, *J. Polym. Res.*, 2016, **23**, 129.
- 17 A. Stanisci, A. Tøndervik, M. Gaardløs, A. Lervik, G. Skjåk-Bræk, H. Sletta and F. L. Aachmann, *ACS Omega*, 2020, **5**, 4352–4361.
- 18 A. H. King, *Food Hydrocolloids*, 1983, **2**, 115–188.
- 19 S. Eswaramma and K. K. Rao, *Carbohydr. Polym.*, 2017, **156**, 125–134.
- 20 D. Myung, D. Waters, M. Wiseman, P. E. Duhamel, J. Noolandi, C. N. Ta and C. W. Frank, *Polym. Adv. Technol.*, 2008, **19**, 647–657.
- 21 C. Silan, A. Akcali, M. T. Otkun, N. Ozbey, S. Butun, O. Ozay and N. Sahiner, *Colloids Surf., B*, 2012, **89**, 248–253.
- 22 K. Mallikarjuna Reddy, V. Ramesh Babu, K. Krishna Rao, M. Subha, K. Chowdoji Rao, M. Sairam and T. Aminabhavi, *J. Appl. Polym. Sci.*, 2008, **107**, 2820–2829.
- 23 J. George, J. Onodera and T. Miyata, *J. Biomed. Mater. Res., Part A*, 2008, **87**, 1103–1111.
- 24 L. Alka, S. Garima, B. S. Sankar and V. Anurag, *J. Drug Delivery*, 2014, **2014**, 1–11.
- 25 S. Ekici and D. Saraydin, *Drug Delivery*, 2004, **11**, 381–388.
- 26 W. Wu and D.-s. Wang, *React. Funct. Polym.*, 2010, **70**, 684–691.
- 27 S. Mandal, S. K. Basu and B. Sa, *Carbohydr. Polym.*, 2010, **82**, 867–873.
- 28 L. Yin, L. Fei, C. Tang and C. Yin, *Polymer International*, 2007, **56**, 1563–1571.
- 29 M. Liu, X. Song, Y. Wen, J.-L. Zhu and J. Li, *ACS Appl. Mater. Interfaces*, 2017, **9**, 35673–35682.
- 30 X. Shi, Y. Zheng, G. Wang, Q. Lin and J. Fan, *RSC Adv.*, 2014, **4**, 47056–47065.
- 31 K. Moebus, J. Siepmann and R. Bodmeier, *Eur. J. Pharm. Sci.*, 2012, **45**, 358–366.
- 32 K.-L. Deng, Y.-B. Gou, L.-R. Dong, Q. Li, L.-B. Bai, T. Gao, C.-Y. Huang and S.-L. Wang, *Frontiers of Materials Science in China*, 2010, **4**, 353–358.
- 33 S. Hua, H. Ma, X. Li, H. Yang and A. Wang, *Int. J. Biol. Macromol.*, 2010, **46**, 517–523.
- 34 A. C. Hodsdon, J. R. Mitchell, M. C. Davies and C. D. Melia, *J. Controlled Release*, 1995, **33**, 143–152.
- 35 C. K. Kuo and P. X. Ma, *Biomaterials*, 2001, **22**, 511–521.
- 36 R. Censi, P. Di Martino, T. Vermonden and W. E. Hennink, *J. Controlled Release*, 2012, **161**, 680–692.



- 37 S. J. Bidarra, C. C. Barrias and P. L. Granja, *Acta Biomater.*, 2014, **10**, 1646–1662.
- 38 R. Schellekens, F. Stellaard, D. Mitrovic, F. Stuurman, J. Kosterink and H. Frijlink, *J. Controlled Release*, 2008, **132**, 91–98.
- 39 A. Blandino, M. Macías and D. Cantero, *Enzyme Microb. Technol.*, 2000, **27**, 319–324.
- 40 T. W. Chung, J. Yang, T. Akaike, K. Y. Cho, J. W. Nah, S. I. Kim and C. S. Cho, *Biomaterials*, 2002, **23**, 2827–2834.
- 41 M. Rajaonarivony, C. Vauthier, G. Couarraze, F. Puisieux and P. Couvreur, *J. Pharm. Sci.*, 1993, **82**, 912–917.
- 42 Y. S. Kim, H. W. Kim, S. H. Lee, K. S. Shin, H. W. Hur and Y. H. Rhee, *Int. J. Biol. Macromol.*, 2007, **41**, 36–41.
- 43 P. Sriamornsak, J. Nunthanid, M. Luangtana-anan and S. Puttipipatkachorn, *Eur. J. Pharm. Biopharm.*, 2007, **67**, 227–235.
- 44 H. H. Tønnesen and J. Karlsen, *Drug Dev. Ind. Pharm.*, 2002, **28**, 621–630.
- 45 M. Grassi, I. Colombo and R. Lapasin, *J. Controlled Release*, 2001, **76**, 93–105.
- 46 R. V. Kulkarni, V. Sreedhar, S. Mutalik, C. M. Setty and B. Sa, *Int. J. Biol. Macromol.*, 2010, **47**, 520–527.
- 47 U. Bhutani, A. Laha, K. Mitra and S. Majumdar, *Mater. Lett.*, 2016, **164**, 76–79.
- 48 Y. Hu, S. Zhang, D. Han, Z. Ding, S. Zeng and X. Xiao, *J. Polym. Res.*, 2018, **25**, 148.
- 49 K. Ramesh, M. Krishnapriya, A. Paul and C. N. Sreeja, *Asian J. Pharm. Clin. Res.*, 2017, **10**, 271–276.
- 50 Y. Fang, S. Al-Assaf, G. O. Phillips, K. Nishinari, T. Funami, P. A. Williams and L. Li, *J. Phys. Chem. B*, 2007, **111**, 2456–2462.
- 51 M. Yonese, K. Baba and H. Kishimoto, *Polym. J.*, 1992, **24**, 395–404.
- 52 W. C. Jao, H. C. Chen, C. H. Lin and M. C. Yang, *Polym. Adv. Technol.*, 2009, **20**, 680–688.
- 53 R. Boppana, G. K. Mohan, U. Nayak, S. Mutalik, B. Sa and R. V. Kulkarni, *Int. J. Biol. Macromol.*, 2015, **75**, 133–143.
- 54 C. Wen, L. Lu and X. Li, *Macromol. Mater. Eng.*, 2014, **299**, 504–513.
- 55 H. Huang, X. Qi and Y. Chen, *Saudi Pharm. J.*, 2019, **27**, 990–999.
- 56 C. Spedalieri, C. Sicard, M. Perullini, R. Brayner, T. Coradin, J. Livage, S. A. Bilmes and M. Jobbágy, *J. Mater. Chem. B*, 2015, **3**, 3189–3194.
- 57 T. Ogoshi, Y. Takashima, H. Yamaguchi and A. Harada, *J. Am. Chem. Soc.*, 2007, **129**, 4878–4879.
- 58 I. Khalid, M. Ahmad, M. U. Minhas and K. Barkat, *Polym. Bull.*, 2018, **75**, 1075–1099.
- 59 S. Ganguly, P. P. Maity, S. Mondal, P. Das, P. Bhawal, S. Dhara and N. C. Das, *Materials Science and Engineering: C*, 2018, **92**, 34–51.
- 60 M. Upadhyay, S. K. R. Adena, H. Vardhan, S. Pandey and B. Mishra, *Drug Dev. Ind. Pharm.*, 2018, **44**, 511–521.
- 61 S. Basu, H. S. Samanta and J. Ganguly, *Soft Mater.*, 2018, **16**, 7–19.
- 62 Y. Lu, A. A. Aimetti, R. Langer and Z. Gu, *Nat. Rev. Mater.*, 2016, **2**, 1–17.
- 63 C.-Y. Chen, T. H. Kim, W.-C. Wu, C.-M. Huang, H. Wei, C. W. Mount, Y. Tian, S.-H. Jang, S. H. Pun and A. K.-Y. Jen, *Biomaterials*, 2013, **34**, 4501–4509.
- 64 K. N. Plunkett, K. L. Berkowski and J. S. Moore, *Biomacromolecules*, 2005, **6**, 632–637.
- 65 N. Bhattarai, H. R. Ramay, J. Gunn, F. A. Matsen and M. Zhang, *J. Controlled Release*, 2005, **103**, 609–624.
- 66 C. De las Heras Alarcón, S. Pennadam and C. Alexander, *Chem. Soc. Rev.*, 2005, **34**, 276–285.
- 67 Y. Zhao, J. Kang and T. Tan, *Polymer*, 2006, **47**, 7702–7710.
- 68 R. Jalababu, S. S. Veni and K. S. Reddy, *J. Drug Delivery Sci. Technol.*, 2018, **44**, 190–204.
- 69 C. Madhavi, P. Kumara Babu, Y. Maruthi, A. Parandhama, O. Sreekanth Reddy and K. Chowdoji Rao, *Int. J. Pharmacol. Pharm. Sci.*, 2017, **9**, 245–252.
- 70 M. Afshar, G. Dini, S. Vaezifar, M. Mehdikhani and B. Movahedi, *J. Drug Delivery Sci. Technol.*, 2020, **56**, 101530.
- 71 K. Campbell, D. Hadley, D. Kukis and E. Silva, *PLoS One*, 2017, **12**, e0181484.
- 72 J. Amirian, T. T. T. Van, S.-H. Bae, H.-I. Jung, H.-J. Choi, H.-D. Cho and B.-T. Lee, *Int. J. Biol. Macromol.*, 2017, **105**, 143–153.
- 73 H.-J. Kao, Y.-L. Lo, W.-J. Vong, Y.-J. Lin and H.-R. Lin, *J. Biomater. Sci., Polym. Ed.*, 2006, **17**, 1191–1205.
- 74 M. J. Martín, A. C. Calpena, F. Fernández, M. Mallandrich, P. Gálvez and B. Clares, *Carbohydr. Polym.*, 2015, **117**, 140–149.
- 75 J. Du, PhD thesis, The University of Texas at Austin, 2015.
- 76 P. Yuan, X. Qiu, R. Jin, Y. Bai, S. Liu and X. Chen, *Biomater. Sci.*, 2018, **6**, 820–826.
- 77 K. Nahar, M. K. Hossain and T. A. Khan, *J. Pharm. Sci. Res.*, 2017, **9**, 606.
- 78 A. D. Augst, H. J. Kong and D. J. Mooney, *Macromol. Biosci.*, 2006, **6**, 623–633.
- 79 K. Ghosal, S. Adak, C. Agatemor, G. Praveen, N. Kalarikkal and S. Thomas, *J. Polym. Res.*, 2020, **27**, 1–11.
- 80 M. Chinthia, S. R. Obireddy, P. Areti, S. Marata Chinna Subbarao, C. R. Kashayi and J. K. Rapoli, *J. Dispersion Sci. Technol.*, 2019, 1–11.
- 81 B. Y. Swamy and Y.-S. Yun, *Int. J. Biol. Macromol.*, 2015, **77**, 114–119.
- 82 V. P. Rudhrabatla, R. Jalababu, K. K. Rao and K. S. Reddy, *J. Drug Delivery Sci. Technol.*, 2019, **51**, 438–453.
- 83 M. Upadhyay, S. K. R. Adena, H. Vardhan, S. K. Yadav and B. Mishra, *Int. J. Biol. Macromol.*, 2018, **115**, 907–919.
- 84 P. Dey, B. Sa and S. Maiti, *Int. J. Biol. Macromol.*, 2015, **76**, 176–180.
- 85 S. Jana, R. Sharma, S. Maiti and K. K. Sen, *Int. J. Biol. Macromol.*, 2016, **92**, 1034–1039.
- 86 O. S. Reddy, M. Subha, T. Jithendra, C. Madhavi and K. C. Rao, *Polym. Polym. Compos.*, 2020, 0967391120904477.
- 87 A. Chang, *Iran. Polym. J.*, 2015, **24**, 161–169.
- 88 A. M. Rajesh, S. A. Bhatt, H. Brahmabhatt, P. S. Anand and K. M. Popat, *Asian J. Pharm. Sci.*, 2015, **10**, 331–340.
- 89 G. Nagarjuna, P. K. Babu, Y. Maruthi, A. Parandhama, C. Madhavi, M. Subha and K. Chowdojirao, *J. Appl. Pharm. Sci.*, 2016, **6**, 011–019.



- 90 J. Zhao, X. Zhao, B. Guo and P. X. Ma, *Biomacromolecules*, 2014, **15**, 3246–3252.
- 91 W. R. Gombotz and S. Wee, *Adv. Drug Delivery Rev.*, 1998, **31**, 267–285.
- 92 D. S. Lima, E. T. Tenório-Neto, M. K. Lima-Tenório, M. R. Guilherme, D. B. Scariot, C. V. Nakamura, E. C. Muniz and A. F. Rubira, *J. Mol. Liq.*, 2018, **262**, 29–36.
- 93 M. S. M. Eldin, E. A. Kamoun, M. A. Sofan and S. M. Elbayomi, *Arabian J. Chem.*, 2015, **8**, 355–365.
- 94 P. Matricardi, M. Pontoriero, T. Coviello, M. A. Casadei and F. Alhaique, *Biomacromolecules*, 2008, **9**, 2014–2020.
- 95 I. M. El-Sherbiny and H. D. Smyth, *Int. J. Pharm.*, 2010, **395**, 132–141.
- 96 F. U. Momoh, J. S. Boateng, S. C. Richardson, B. Z. Chowdhry and J. C. Mitchell, *Int. J. Biol. Macromol.*, 2015, **81**, 137–150.
- 97 L. Pescosolido, T. Vermonden, J. Malda, R. Censi, W. J. Dhert, F. Alhaique, W. E. Hennink and P. Matricardi, *Acta Biomater.*, 2011, **7**, 1627–1633.

