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## Can solvent induced surface modifications applied to screen-printed platforms enhance their electroanalytical performance?†

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In this paper the effect of solvent induced chemical surface enhancements upon graphitic screen-printed electrodes (SPEs) is explored. Previous literature has indicated that treating the working electrode of a SPE with the solvent *N,N*-dimethylformamide (DMF) offers improvements within the electroanalytical response, resulting in a 57-fold increment in the electrode surface area compared to their unmodified counterparts. The protocol involves two steps: (i) the SPE is placed into DMF for a selected time, and (ii) it is cured in an oven at a selected time and temperature. Beneficial electroanalytical outputs are reported to be due to the increased surface area attributed to the binder within the bulk surface of the SPEs dissolving out during the immersion step (step i). We revisit this exciting concept and explore these solvent induced chemical surface enhancements using edge- and basal-plane like SPEs and a new bespoke SPE, utilising the solvent DMF and explore, in detail, the parameters utilised in steps (i) and (ii). The electrochemical performance following steps (i) and (ii) is evaluated using the outer-sphere redox probe hexammineruthenium(III) chloride/0.1 M KCl, where it is found that the largest improvement is obtained using DMF with an immersion time of 10 minutes and a curing time of 30 minutes at 100 °C. Solvent induced chemical surface enhancement upon the electrochemical performance of SPEs is also benchmarked in terms of their electroanalytical sensing of NADH (dihydropyridine adenine dinucleotide reduced form) and capsaicin both of which are compared to their unmodified SPE counterparts. In both cases, it is apparent that a marginal improvement in the electroanalytical sensitivity (*i.e.* gradient of calibration plots) of 1.08-fold and 1.38-fold are found respectively. Returning to the original exciting concept, interestingly it was found that when a poor experimental technique was employed, only then significant increases within the working electrode area are evident. In this case, the insulating layer that defines the working electrode surface, which was not protected from the solvent (step ii) creates cracks within the insulating layer exposing the underlying carbon connections and thus increasing the electrode area by an unknown quantity. We infer that the origin of the response reported within the literature, where an extreme increase in the electrochemical surface area (57-fold) was reported, is unlikely to be solely due to the binder dissolving but rather poor experimental control over step (i).

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## 1. Introduction

The utilisation of carbon materials for applications within electroanalytical sensors have become a major factor over recent decades, as researchers strive to find cheap and effective electrochemical systems that possess the capabilities for the simplistic retrieval of analytical data. Modification of electrode surfaces has been a method commonly utilised<sup>1–4</sup> with materials such as graphene,<sup>5</sup> carbon nanotubes,<sup>6</sup> metal phthalocyanines,<sup>7</sup> chitosan,<sup>8,9</sup> enzymatic materials,<sup>10</sup> metallic nanoparticles<sup>11,12</sup> to name a just a few, with the aim of improving electroanalytical sensitivity's compared to their bare unmodified counterparts.



In addition to the modification of carbon materials, for improved electrochemical responses, the utilisation of porous electrodes have also been considered, with such electrodes offering a large surface area with no need for further modification.<sup>13–15</sup> There have been many beneficial electro-analytical applications of these porous electrode materials, where the increased surface area creates an easily improved electrochemical/electroanalytical performance.<sup>14,16</sup> Such advances of utilising porous electrodes has been identified by Wang *et al.*<sup>17</sup> who have presented a concise overview of electro-analytical capabilities of this electrode configuration. For example, Friedrich *et al.*<sup>18</sup> reported the use of reticulated vitreous carbon as an electrode material for the electroanalytical determination of NADH (dihydropyridine adenine dinucleotide reduced form), ascorbic acid and glucose. Additionally, it has been reported by Figueiredo-Filho *et al.*<sup>15</sup> that porous freestanding graphene foams provide beneficial electroanalytical performances towards uric acid, acetaminophen and dopamine. The application of porous electrodes have expanded towards the sensing of heavy metal ions, for example Niu *et al.*<sup>19</sup> have applied a highly porous carbon paste electrode for the detection of Cd(II), Pb(II) and Ni(II) within real water samples, which due to the large analytical current/signal provided by the porosity/increase in surface area allows for extreme improvements in the electroanalytical sensitivity.

It has become apparent within recent literature that a different approach for improving electroanalytical capabilities of screen-printed electrodes (SPEs) has been undertaken, exploiting the nature of the ink used in the manufacture process, utilising the solvent *N,N*-dimethylformamide (DMF).<sup>20</sup> It has been reported that exposure of SPEs to DMF results in an extreme increase of the electrochemically active surface area, thus increasing the electroanalytical sensitivity towards target analytes. It is hypothesised that the solvent dissolves the binder within the SPE creating large areas of “freed” graphite available at the working electrode surface.<sup>20</sup> This is exemplified by Washe *et al.*<sup>20</sup> who reported a 57-fold increment in the effective surface area evaluated using the redox probe potassium ferrocyanide utilising SPEs fabricated using commercially available inks. The reported solvent induced surface modification protocol involves two steps: (i) the SPE is placed into the solvent DMF for an optimised time after which, (ii) the SPEs are removed and cured for a selected time and temperature in an oven.

In this paper we explore solvent induced surface modifications, *via* steps (i) and (ii), in detail and benchmark these to their unmodified counterparts utilising the redox probe hexaammineruthenium(III) chloride. Solvent induced surface modified SPEs are also used and evaluated towards the electro-analytical sensing of NADH and capsaicin. In addition, we have performed control experiments to understand the exact origin of the response reported by Washe *et al.*<sup>20</sup> The results arising from this work show that the electrochemical improvement from utilising steps (i) and (ii) are not as beneficial as previously reported; reasons for this are given.<sup>20</sup>

## 2. Experimental

All chemicals used were obtained from Sigma-Aldrich at analytical grade and were used without any further purification. All solutions were prepared with deionised water of resistivity not less than 18.2 MΩ cm. Voltammetric measurements were carried out using an Emstat (Palmsens Instruments BV, The Netherlands) potentiostat. When necessary, Parafilm™ (Sigma-Aldrich) was used to define the working area of the screen-printed electrodes (explained later). SEM images were obtained with a JEOL JSM-5600LV model. The SPEs fabricated here have been extensively characterised *via* RAMAN and XPS analysis and have been published within recent literature.<sup>21</sup>

Screen-printed graphite electrodes were fabricated in-house with appropriate stencil designs using a microDEK 1760RS screen-printing machine (DEK, Weymouth, UK). Two commercially available graphite inks were utilised and a further bespoke ink developed; these are detailed in Table 1.<sup>21</sup> For each of the screen-printed electrodes, a carbon-graphite ink formulation was first screen-printed onto a polyester flexible film (Autostat, 250 μm thickness). This layer was cured in a box fan oven with extraction at 60 °C for 30 min. Next, a silver/silver chloride (40:60) reference electrode was applied by screen-printing Ag/AgCl paste (Product Code: C2040308P2; Gwent Electronic Materials Ltd, UK) onto the plastic substrate. This layer was once more cured in an oven at 60 °C for 30 min. Last, an insulating dielectric paste ink (Product Code: D2070423D5; Gwent Electronic Materials Ltd, UK) was printed to cover the connections and define the 3 mm diameter graphite working electrode. After curing at 60 °C for 30 min the screen-printed electrodes are ready to use. Scanning electron microscopic (SEM) images of above fabricated working electrodes at different magnifications are shown in ESI Fig. 1(A, C, E) and 2.†

The procedure to facilitate the enhancement of the active surface of the screen-printed electrodes (SPEs) involves two key steps, as reported within the literature.<sup>20</sup> Unless otherwise stated, the insulating layer of the devices was protected with Parafilm™, which is a DMF inert material, prior to any chemical treatment of the SPEs. The first step involves immersing the SPEs (working electrode only; counter and reference electrodes removed prior to this step) into DMF and leaving for an optimised period of time. The second step involves removing the SPEs from the DMF and curing at an optimised tempera-

**Table 1** The range of screen-printed inks utilised within this study to explore the effects of solvent treatment across an array of commercially available materials. Screen-printed electrodes made from these inks have been previously extensively characterised and reported upon<sup>7,21,30</sup>

Graphite ink	Notation	Product code
Edge plane	ESPE	Gwent Electronic Materials – C2000802P2
Basal plane	BSPE	Peters – SD 2843 HAL
Bespoke	MSPE	50/50 mix of above inks



ture and time in a fan oven. In all instances, the solvent (DMF) utilised were unadulterated and not changed in any way prior to utilisation following the reported protocol detailed by Washe *et al.*<sup>20</sup> Following the second step, the SPEs are ready to use and are connected *via* an edge connector to ensure a secure electrical connection.<sup>22</sup> A saturated calomel reference electrode (SCE) and a nickel counter electrode were used throughout for all experiments.

Electrochemical characterisation of the electrodes utilised throughout this work were *via* two routes, the standard heterogeneous rate constant ( $k^0$ ) and the effective electroactive area ( $A_{\text{eff}}$ ). The  $k^0$  was evaluated using the data obtained with the redox probe hexaammineruthenium(III) chloride in 0.1 M KCl. The Nicholson method is routinely used to estimate the  $k^0$ , for quasi-reversible systems using the following eqn (1):<sup>23</sup>

$$\varphi = k^0 [\pi D n \nu F / (RT)]^{-1/2} \quad (1)$$

where  $\varphi$  is the kinetic parameter,  $D$  is the diffusion coefficient ( $9.1 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$ ) of the electrochemical redox probe used,  $n$  is the number of electrons involved in the electrochemical process,  $F$  is the Faraday constant,  $\nu$ , the voltammetric scan rate,  $R$  the universal gas constant, and the experiments are performed at a temperature  $T$ . The kinetic parameter,  $\varphi$  is tabulated as a function of peak-to-peak separation ( $\Delta E_p$ ) at a set temperature (298 K) for a one-step, one electron process. The function of  $\varphi(\Delta E_p)$ , which fits Nicholson's data, for practical usage (rather than producing a working curve) is given by eqn (2):<sup>24</sup>

$$\varphi = (-0.628 + 0.0021X / (1 - 0.017X)) \quad (2)$$

where  $X = \Delta E_p$  is used to determine  $\varphi$  as a function of  $\Delta E_p$  from the experimentally obtained voltammetry. From this, a plot of  $\varphi$  against  $[\pi D n \nu F / (RT)]^{-1/2}$  can be produced graphically allowing the  $k^0$  to be readily determined, however  $\Delta E_p$  values that exceed 212 mV within the Nicholson table have to rely upon the following eqn (3):<sup>25</sup>

$$k^0 = [2.18(DanF\nu/RT)^{0.5}] \exp[-(\alpha^2 nF/RT) \times \Delta E_p] \quad (3)$$

where the constants are the same as described in eqn (1), apart from the transfer coefficient,  $\alpha$  which is assumed to correspond to 0.5. The corresponding  $k^0$  values were calculated for the unmodified/bare ESPEs, MSPEs and BSPEs (see Table 1) using the above equations, with values found to correspond to  $1.8 \times 10^{-3} \text{ cm s}^{-1}$ ,  $7.1 \times 10^{-4} \text{ cm s}^{-1}$  and  $1.23 \times 10^{-5} \text{ cm s}^{-1}$ , respectively. In these cases, the range in values is due to the different rates of electron transfer associated with these materials. We assign the term ESPE since it gives a relatively fast  $k^0$  akin to edge plane pyrolytic graphite electrodes, and also the term BSPE since the  $k^0$  is akin to basal plane pyrolytic graphite electrodes, *i.e.* relatively slow. In the case of MSPEs, a mixture of ESPEs and BSPEs inks, and hence the  $k^0$  is between that of the ESPEs and BSPEs.

To evaluate the effective electrode area,  $A_{\text{eff}}$ , given that for a co-planar macro electrode in the electrochemically quasi-reversible case, the Randles-Ševčík equation (at 298 K) is pre-

sented in eqn (4), where the notation is the same as above and  $C$  is the concentration of electroactive substance:<sup>5</sup>

$$I_p^{\text{Quasi}} = 2.65 \times 10^5 n^{3/2} D^{1/2} \nu^{1/2} C A_{\text{eff}} \quad (4)$$

### 3. Results and discussion

We initially build upon the exciting report by Washe *et al.*<sup>20</sup> who focused upon the use of DMF (as mentioned in the introduction) to facilitate impressive improvements within the voltammetric signal. It was reported that there was a 57-fold increase in the geometric area and a 100-fold increase within the electron transfer kinetics as a result of the solvent induced modification of carbon based screen-printed electrodes (SPEs).<sup>20</sup> SPEs which have been previously characterised and reported upon<sup>21,26</sup> were selected to explore the basis of this exciting phenomena. The graphitic ink formulations used within this work to fabricate the SPEs are presented in Table 1. The procedure to generate solvent modified SPE surfaces involves two key steps: (i) the SPE is placed into the DMF solvent for a selected time, and (ii) is cured in an oven at a selected time and temperature with the purpose of evaporating the residual solvent and re-curing the working electrode.<sup>20</sup>

#### Optimisation of the experimental procedure

Following the procedure detailed above, steps (i) and (ii), the SPEs were immersed in DMF for 10 minutes, followed by a curing step of 30 minutes at 100 °C. Fig. 1 shows typical cyclic voltammograms obtained within 1 mM hexaammineruthenium(III) chloride/0.1 M KCl, this probe was chosen since it is an outer-sphere redox probe, which is affected only by the electronic structure (*i.e.* edge plane like-sites/defects).<sup>27</sup> It is clear

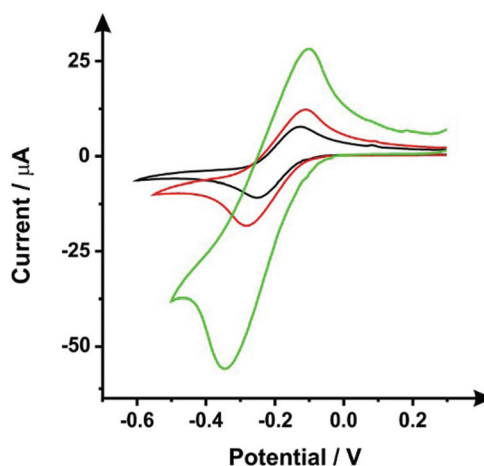


Fig. 1 Cyclic voltammograms recorded within 1 mM hexaammineruthenium(III) chloride/0.1 M KCl using unmodified (black line) and DMF-modified (10 minutes solvent immersion, 30 minutes of curing at 100 °C) ESPEs with (red line) and without (green line) an additional protecting DMF inert film over the insulating dielectric layer. Scan rate: 50  $\text{mV s}^{-1}$  (vs. SCE).



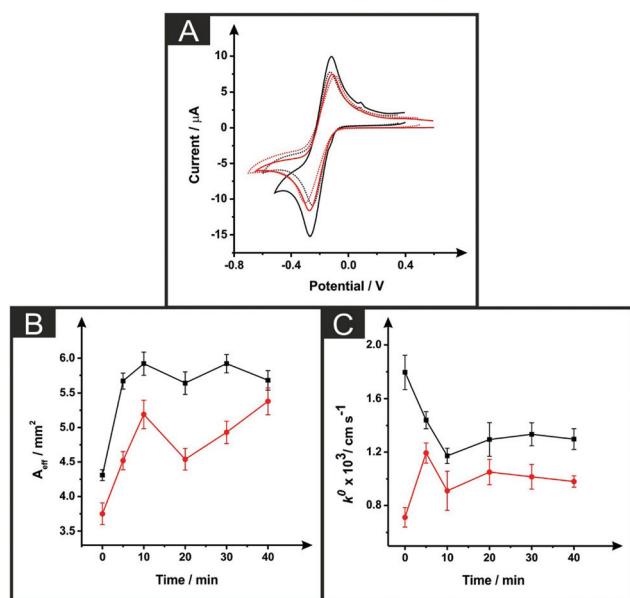
that the DMF induced surface modification shows a slight improvement of the voltammetric signal, as the peak current attained was relatively larger compared to an unmodified ESPE. It would therefore seem apparent that the DMF treatment can remove, to some degree, a proportion of the polymeric binder from within the ink of the working electrode, which is used to keep the carbon robustly adhered together. However, data presented in Fig. 1 shows that there is a substantial increase in the voltammetric peak current when the insulating layers (used to define the working electrode and protect the carbon tracks from the solution) are not protected from the solvent. Therefore, to avoid this scenario, which could increase the working electrode surface area (in an uncontrolled manner), we have taken precautions through the use of a protective DMF resistant film (see Experimental section) to cover the dielectric insulating layer.

### Optimisation of the DMF immersion time (step i)

The effect of exposing the SPEs towards DMF was evaluated through the monitoring of the standard heterogeneous rate constants,  $k^0$  and effective electroactive area,  $A_{\text{eff}}$  resulting from a range of different immersion times from 5 to 40 minutes. It is important to note, that in this situation a curing temperature of 100 °C for 1 hour was chosen in order to ensure full evaporation of the solvent and re-curing of the electrode surface. Fig. 2A depicts typical cyclic voltammetric

data for unmodified and solvent modified ESPE towards the redox probe hexammineruthenium(III) chloride, where an improvement in the peak-to-peak separation ( $\Delta E_p$ ) is apparent, with values corresponding to 115 mV and 150 mV respectively. In the case of the MSPEs (see Fig. 2B), the values of  $\Delta E_p$  corresponded to 200 mV and 155 mV for unmodified and DMF modified MSPEs respectively. In both cases, there is an apparent increase in the voltammetric peak height, indicating a possible increase in the electroactive area. Note that multiple attempts were made to explore the modification of the BSPEs *via* the above procedure (steps (i) and (ii)) but due to the lack of relative adherence of the carbon ink to the polyester film these were discarded; the effect of solvent exposure is shown in ESI Fig. 3.†

Returning to the ESPE and MSPEs longer DMF immersion times were next explored and the results are summarised within Fig. 2B and C, in terms of the  $A_{\text{eff}}$  and the  $k^0$ , respectively. In Fig. 2B, it is apparent that the effective area is improved in comparison to the unmodified SPEs for all the DMF exposure times assayed. Clearly, in the case of the ESPEs, the  $A_{\text{eff}}$  increases from 31 to 37%, whilst the improvement in the case of MSPEs varies between 21 and 44% relative to the bare/unmodified SPEs. Upon utilisation of an immersion time of 5 minutes, the DMF treated MSPE was found to have a slightly improved  $k^0$ , with values corresponding to  $7.2 \times 10^{-4} \text{ cm s}^{-1}$  and  $1.2 \times 10^{-3} \text{ cm s}^{-1}$  for unmodified and modified MSPE respectively. Increasing the immersion time within DMF implies an improved electrochemical reversibility compared to unmodified MSPEs in most cases apart from 5 minutes. However, in the case of the ESPE, the solvent treatment has no significant change, as evaluated using the  $k^0$ , (a modest change from  $1.8 \times 10^{-3} \text{ cm s}^{-1}$  to  $1.2 \times 10^{-3} \text{ cm s}^{-1}$  is observed). Overall, upon increasing the solvent immersion time it is dependent upon the starting electrode reactivity, it is apparent that the utilisation of an electrode platform with faster electrode kinetics (ESPE) results in a slight decrease in its voltammetric performance compared to that of a slower electrode platform (MSPE) where an increase is observed. During the solvent modification step applied to the MSPE, the DMF reveals more electroactive sites (*i.e.* removal of binder) upon the electrode's surface, which gives rise to an improvement in the electrochemical reversibility. However, in the case of the ESPE which has a large percentage of electroactive sites are already present in its unmodified form, the effect is that the binder is dissolved but then likely reforms on these active sites which results in an apparent reduced electrochemical response.



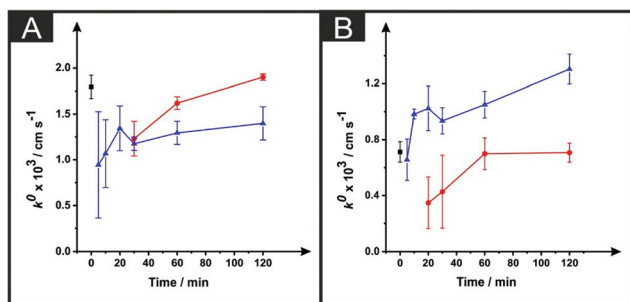
**Fig. 2** The effect of DMF exposure time (at a constant curing time and temperature of 1 hour at 100 °C) as evaluated *via* cyclic voltammetry using 1 mM hexammineruthenium(III) chloride/0.1 M KCl (A) utilising an unmodified ESPE (dotted line), modified ESPE (solid black line) and MSPE (solid red line) following DMF immersion at a time of 5 minutes. Additionally presented are corresponding plots of  $A_{\text{eff}}$  (B) and  $k^0$  (C) as a function of DMF immersion time utilising an ESPE (black) and MSPE (red). Data for the  $A_{\text{eff}}$  and  $k^0$  were deduced using eqn (3) and (4) respectively. Represented data indicate the averages and standard deviations ( $N = 3$ ). Scan rate:  $50 \text{ mV s}^{-1}$  (vs. SCE).

### Optimisation of SPE curing conditions (step ii)

Following the optimisation of the DMF immersion time, as described above, we next turn to exploring the effect of varying and optimising the curing temperature (40, 60 and 100 °C) and time (5–120 minutes). Fig. 3 shows the effect of the curing temperature upon the  $k^0$  which is depicted for both the ESPEs and MSPEs. It is clear that increasing the curing time equates to an improvement in the  $k^0$ . Fig. 3A shows that



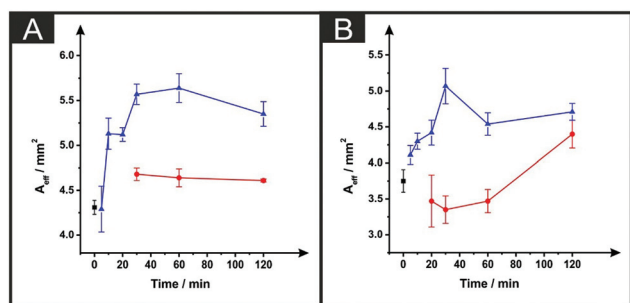




**Fig. 3** The effect of drying variables (temperature and time) upon the  $k^0$  utilising DMF-modified and unmodified (squares) ESPEs (A) and MSPEs (B). Note: prior to the DMF immersion step, the SPEs insulating layer was further protected with a DMF inert film. Experimental parameters: 20 minutes DMF modification followed by curing temperatures of: 60 °C (circles) and 100 °C (triangles). Data for the  $k^0$  were deduced using eqn (3). Represented data indicate the average and standard deviation ( $N = 3$ ).

the application of 100 °C reduces the magnitude of the  $k^0$  when compared with 60 °C or the use of the unmodified electrodes. This result is in accordance with Fig. 2C in which the use of the same parameters leads to a slower  $k^0$  value. In the case of the MSPEs (represented in Fig. 3B) and, unlike Fig. 3A for ESPEs, the  $k^0$  improves when the higher temperature is utilised instead of 60 °C.

In Fig. 4 the effect of the drying variables upon the electroactive surface area is presented. In the case of the ESPEs, in general for both the time and temperature of the curing process used, there is an increase in the effective area. When the effective area was deduced for the MSPEs, the signal increased in respect to the unmodified electrode at 100 °C and upon a relatively long curing time at 60 °C was applied; the largest increase in the electroactive area is found to correspond to 31% (ESPE) and 35% (MSPE). In the case of 40 °C as the chosen temperature, the electrochemistry of hexaammine-



**Fig. 4** The effect of drying variables (temperature and time) upon the  $A_{\text{eff}}$  utilising DMF-modified and unmodified (squares) ESPEs (A) and MSPEs (B). Note: prior to the DMF immersion step, the SPEs insulating layer was further protected with a DMF inert film. Experimental parameters: 20 minutes DMF modification followed by curing temperatures of: 60 °C (circles) and 100 °C (triangles). Data for the  $A_{\text{eff}}$  were deduced using eqn (4). Represented data indicate the average and standard deviation ( $N = 3$ ).

ruthenium(III) chloride was hindered when compared to the unmodified SPEs in terms of peak current,  $\Delta E_p$  and repeatability. In addition, when the drying time of the modified electrode was reduced to less than 30 minutes at 60 °C, the same electrochemical performance was observed therefore; such data was not taken into account in Fig. 3 and 4.

Last, although the solvent treatment can improve the electrochemical signal, possibly due to the solvent removing the polymeric binder within the working electrode, additionally incomplete removal of DMF also has a detrimental effect upon the electrochemical variables of interest, as seen in ESI Fig. 4.† As the boiling temperature of DMF is  $\sim 150$  °C, a drying temperature of 40 and 60 °C over relatively short times do not evaporate the solvent completely, giving rise to unreproducible voltammetric signatures, as shown in ESI Fig. 4,† where the residual DMF upon the surface is hindering the electrochemical performance of the electrode.

In an attempt to further understand the above processes, where solvent treatment can change the voltammetric response, SEM analysis was performed (depicted in ESI Fig. 1.†), where comparisons were made between modified and unmodified ESPEs that were subjected to DMF exposure for a period of 10 minutes. Inspection of the SEMs clearly show that the DMF does not seem to alter the surface morphology of the SPE. This can be seen in ESI Fig. 1,† which are representative of the bare and modified surfaces, respectively. However, SEM effectively takes snapshots of the electrode surface and not the overall surface; it could be quite likely that the binder is removed but SEM imaging cannot identify these changes. We also noted that the thickness of the printed working electrode, which is  $\sim 20$  microns (defined by the screen; see Experimental section) did not appear to alter following solvent treatment as evaluated using SEM (cross-sectional analysis) and White Light Profilometry. Overall, trying to optimise steps (i) and (ii) is not straightforward; that said, we utilise optimised steps (i) and (ii) and next explore solvent modified SPEs as potential beneficial electroanalytical sensors.

### Exploring solvent induced surface modified SPEs evaluated towards the sensing of NADH and capsaicin

Next, the electroanalytical performance of the modified ESPEs was considered towards the potential sensing of dihydronicotinamide adenine dinucleotide (NADH, reduced form) and capsaicin. These are chosen as model analytes with which to compare the electroanalytical response of the solvent modified electrode platforms. The importance of NADH is due to its central role in mammalian metabolism as NADH/NAD<sup>+</sup> pair redox couple and is the most common reducing power source between biochemical reactions.<sup>26</sup> Additionally, the sensing of capsaicin was chosen since it has been shown that there is a good correlation between capsaicin content in chilli peppers (and derivatives) measured by electroanalytical devices (modified with carbon nanotubes) and Scoville units<sup>28,29</sup> offering an interesting approach to determining the hotness of chilli pepper and related products through the sensing of capsaicin. We used these interesting analytes to benchmark the solvent



induced procedure of these SPE platforms, in optimised conditions of the chosen solvent (DMF), time in which solvent and sensor are in contact (10 minutes), curing temperature (100 °C) and fan oven term (30 minutes). The immersion time in DMF of 10 minutes was chosen because, as shown in Fig. 2B, the effective area was increased in respect to the unmodified devices and reached a plateau. As shown in Fig. 4A, the use of 100 °C improved the effective area more than when 60 °C was chosen as curing temperature. In addition, in this Fig. 4A it is shown that for 30 minutes of fan oven treatment, the effective area attained a plateau and being maximum.

The electroanalytical performance of an unmodified and solvent modified ESPE towards the detection of NADH was first evaluated. Additions of NADH were made into a pH 7 PBS solution over the range 2.5–100  $\mu\text{M}$ . As depicted in Fig. 5A, the increase within the NADH concentration relates to a higher peak current for the electrochemical oxidation of NADH, as expected. It is well reported that a single anodic wave is seen at  $\sim +0.50$  V (*vs.* SCE) due to the irreversibility of the process. The plots of NADH concentration *vs.* peak current are shown in Fig. 5B (black points for bare devices, red points for the modified). The response of both the unmodified and DMF modified ESPEs were linear over the studied concentration range with regression equations of  $I_p$  ( $\mu\text{A}$ ) =  $8.63 \times 10^{-3} \mu\text{M}^{-1} - 9.71 \times 10^{-3} (\mu\text{A})$  ( $R^2 = 0.999$ ,  $N = 3$ ) and  $I_p$  ( $\mu\text{A}$ ) =  $9.36 \times 10^{-3} \mu\text{M}^{-1} - 1.49 \times 10^{-2} (\mu\text{A})$  ( $R^2 = 0.99$ ,  $N = 3$ ) respectively. Therefore, the modified electrode showed a slightly improved sensitivity towards NADH, a 1.08-fold increment. The limits of detection ( $3\sigma$ ), based on the calculation using the standard deviation of the peak current with the lowest NADH concentration studied, corresponded to 1.3 and 1.8  $\mu\text{M}$  for bare and modified ESPEs, respectively; these limits of detection are competitive and analytically useful.

Next, the electroanalytical detection of capsaicin was explored with unmodified ESPEs and with the modified counterparts, subjected to the optimised modification procedure: immersion time of 10 minutes within DMF and a

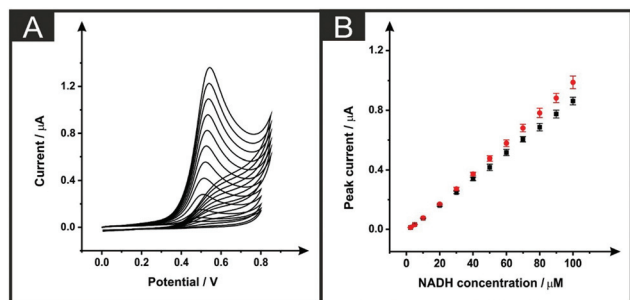


Fig. 5 Typical cyclic voltammograms (A) as a result of increasing concentrations of NADH (2.5–100  $\mu\text{M}$ )/pH 7.1 PBS using a DMF-modified ESPE (10 minutes within the solvent and a curing temperature of 100 °C for 30 minutes) and corresponding calibration plots (B) over the concentration range using an unmodified ESPE (squares) and DMF-modified ESPE (circles). Represented data indicate the average and standard deviations ( $N = 3$ ). Scan rate:  $50 \text{ mV s}^{-1}$  (*vs.* SCE).

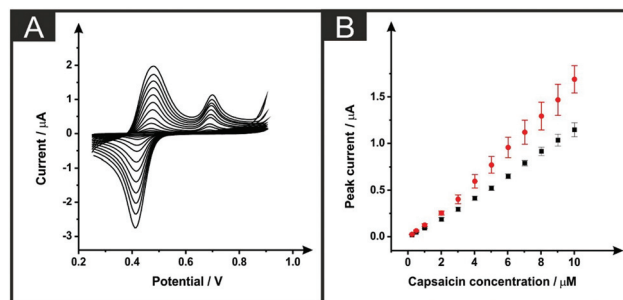


Fig. 6 Cyclic voltammograms (A) resulting from increasing concentrations of capsaicin (0.25–10  $\mu\text{M}$ )/0.1 M phosphoric acid solution using a DMF-modified ESPE (10 minutes within the solvent and a curing temperature of 100 °C for 30 minutes). The respective calibration plots (B) of the anodic signal at +0.48 V (*vs.* SCE) of the second consecutive cyclic voltammogram utilising unmodified ESPE (squares) and DMF-modified ESPEs (circles) are shown. Represented data indicate the average and standard deviations ( $N = 3$ ). Scan rate of  $50 \text{ mV s}^{-1}$  (*vs.* SCE).

curing temperature of 100 °C for 30 minutes. The redox mechanism of this interesting molecule includes first an oxidation that occurs at a potential of  $\sim +0.70$  V (*vs.* SCE), as is depicted within Fig. 6A (in a 0.1 M phosphoric acid aqueous solution).<sup>28,29</sup> In the reverse cathodic scan, the generated molecule with an *o*-benzoquinone unit is reduced at  $\sim +0.42$  V to produce an *o*-benzenediol derivative that is subjected to an anodic process occurring at  $\sim +0.48$  V of the second consecutive scan (at a scan rate of  $50 \text{ mV s}^{-1}$ ), which is in agreement with previous reports; no changes in the peak-to-peak separation of the voltammetric response of capsaicin on solvent modified and unmodified SPEs were evident.<sup>28,29</sup> Fig. 6A shows the response of capsaicin additions made over a concentration range of 0.25 to 10  $\mu\text{M}$  where both the bare and solvent modified electrodes show linear responses (Fig. 6B) with regression equations of  $I_p$  ( $\mu\text{A}$ ) =  $0.109 \mu\text{M}^{-1} - 8.54 \times 10^{-3} (\mu\text{A})$  ( $R^2 = 0.997$ ;  $N = 3$ ) and  $I_p$  ( $\mu\text{A}$ ) =  $0.151 \mu\text{M}^{-1} - 1.38 \times 10^{-2} (\mu\text{A})$  ( $R^2 = 0.98$ ;  $N = 3$ ) for bare ESPEs and modified ESPEs respectively. Therefore, a modest improvement within the sensitivity is observed for capsaicin with a 1.38-fold increment. Although the higher slope was obtained with the modified electrodes, the associated average standard deviation was almost twice of the bare, 12% and 7%, respectively for the modified platforms and for the bare devices (13 calibration points,  $N = 3$ ). The limits of detection are found to correspond to 0.14 and 0.17  $\mu\text{M}$  for bare and DMF-modified devices, respectively. These LODs are competitive and comparable to literature reports, for example, as that reported by Randviir *et al.*<sup>28</sup>

### Comparison with the prior literature reports

As mentioned previously, Washe *et al.*<sup>20</sup> reported impressive improvements in the voltammetric signals following solvent modification where an improvement of 57-fold was reported using the redox probe potassium ferrocyanide over that of an unmodified electrode. In this paper, the increase within the



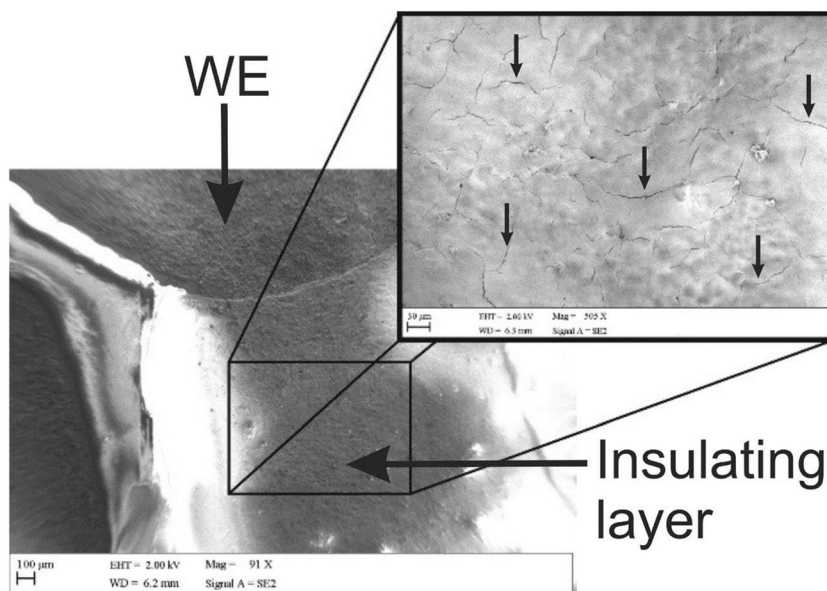


Fig. 7 Typical SEM imaging of the working electrode (WE) and insulating layer of an ESPE after 30 minutes exposure to DMF and curing step in a fan oven at 60 °C for 30 minutes, with focus upon the insulating layer. The insulating layer of the device was in direct contact with the solvent. Note: this SPE was not protected with the DMF inert film and thus the whole sensor including part of the insulating layer was purposely immersed into DMF.

effective electroactive area as evaluated in Fig. 1 (green curve) is less than 50% even when the insulative layer is attacked and partially dissolved by the DMF revealing the underlying carbon-graphite connective tracks; the electrochemical evidence of a SPE without protection of the insulating layer is depicted in Fig. 1.

Further to this, if we consider the effect of DMF exposure upon the areas encompassing the working electrode of the screen-printed electrode, that is the non-conductive insulating layer utilised to cover the carbon-graphite electrode tracks and critically define the working electrode area, we observed significant shortcomings with the aim of improving just the electrochemical response of the working electrode. Fig. 7 depicts a SEM image obtained after 20 minutes of exposure to DMF, but this time focussing upon the insulating layer, which due to its polymeric composition is attacked/degraded, by the DMF. Clearly, as can be seen in the form of severe pitting, the DMF significantly distorts the insulating layer exposing the underlying carbon-graphite connective tracks. In this scenario, when the electrochemistry is performed, not only is the signal due to the working electrode area, but also the underlying exposed carbon tracks contributes which provides a significantly increased electrode area. This is in agreement with data presented in Fig. 1 (green line) where a substantially large voltammetric current was observed when the insulating layer was not protected from the DMF solvent.

Last, we revisit the work of Washe *et al.*,<sup>20</sup> and re-explore their reported claims of a 57-fold improvement in the voltammetric response as demonstrated using 2 mM potassium ferricyanide. Since they report the use of an electrode area of 0.07 cm<sup>2</sup>, a 57-fold increase would correspond to an electrode

area of 3.99 cm<sup>2</sup>. If we input this value into the Randles-Ševčík (eqn (4)) for a quasi-reversible electrochemical process, the expected peak current equations to  $\sim 8.2 \times 10^{-4}$  A. Inspection of Fig. 5 published in the paper by Washe *et al.*<sup>20</sup> reveals that this is not the case and rather peak currents are of the  $\sim 10^{-6}$  A order of magnitude. This casts doubt on the hypothesis that the binder of the SPE working electrode is dissolved solely from the working electrode giving rise to a larger electrode area. Given our work above, we infer that in the work of Washe *et al.*<sup>20</sup> that poor experimental control, where the insulating layer is attacked by DMF to reveal the underlying carbon tracks, highly likely contributes to the overall electrode area; this is exemplified in our work (see Fig. 7).

## 4. Conclusions

In this paper we have explored the effect of solvent modified SPEs evaluated using a range of electroactive compounds namely, hexaammineruthenium(III) chloride, NADH and capsaicin. In all cases, following optimisation of the solvent immersion time (step i) and curing time (step ii), for the electroanalytical sensing of NADH and capsaicin increments of 1.08-fold and 1.38-fold respectively, only are observed relative to their unmodified counterparts. Consequently, returning to the title question of this paper, *Can Solvent Induced Surface Modifications Applied to Screen-Printed Platforms Enhance their Electroanalytical Performance?*, in the examples presented it is not an appropriate strategy to enhance the electrochemical/electroanalytical performance of SPEs for the range of SPEs and analytes studied here.





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## References

- 1 R. W. Murray, A. G. Ewing and R. A. Durst, *Anal. Chem.*, 1987, **59**, A379–A390.
- 2 S. A. Wring and J. P. Hart, *Analyst*, 1992, **117**, 1215–1229.
- 3 A. V. Kolliopoulos, J. P. Metters and C. E. Banks, *Anal. Methods*, 2013, **5**, 851–856.
- 4 A. P. Ruas de Souza, C. W. Foster, A. V. Kolliopoulos, M. Bertotti and C. E. Banks, *Analyst*, 2015, **140**, 4130–4136.
- 5 D. A. C. Brownson, C. W. Foster and C. E. Banks, *Analyst*, 2012, **137**, 1815–1823.
- 6 A. P. Ruas de Souza, M. Bertotti, C. W. Foster and C. E. Banks, *Electroanalysis*, 2015, **27**, 2295–2301.
- 7 C. W. Foster, J. P. Metters and C. E. Banks, *Electroanalysis*, 2013, **25**, 2275–2282.
- 8 Y. Xin, L. Guanghan, W. Xiaogang and Z. Tong, *Electroanalysis*, 2001, **13**, 923–926.
- 9 N. Hernández-Ibáñez, L. García-Cruz, V. Montiel, C. W. Foster, C. E. Banks and J. Iniesta, *Biosens. Bioelectron.*, 2016, **77**, 1168–1174.
- 10 A. Eremenko, A. Makower, W. Jin, P. Rüger and F. Scheller, *Biosens. Bioelectron.*, 1995, **10**, 717–722.
- 11 E. Bernalte, C. Marín Sánchez and E. Pinilla Gil, *Sens. Actuators. B*, 2012, **161**, 669–674.
- 12 L. Yuan, J. Zhang, P. Zhou, J. Chen, R. Wang, T. Wen, Y. Li, X. Zhou and H. Jiang, *Biosens. Bioelectron.*, 2011, **29**, 29–33.
- 13 S. Cho, K.-H. Shin and J. Jang, *ACS Appl. Mater. Interfaces*, 2013, **5**, 9186–9193.
- 14 D. A. C. Brownson, L. C. S. Figueiredo-Filho, X. Ji, M. Gomez-Mingot, J. Iniesta, O. Fatibello-Filho, D. K. Kampouris and C. E. Banks, *J. Mater. Chem. A*, 2013, **1**, 5962–5972.
- 15 L. C. S. Figueiredo-Filho, D. A. C. Brownson, O. Fatibello-Filho and C. E. Banks, *Electroanalysis*, 2014, **26**, 93–102.
- 16 X. Dong, X. Wang, L. Wang, H. Song, H. Zhang, W. Huang and P. Chen, *ACS Appl. Mater. Interfaces*, 2012, **4**, 3129–3133.
- 17 J. Wang, *Electrochim. Acta*, 1981, **26**, 1721–1726.
- 18 J. M. Friedrich, C. Ponce-de-León, G. W. Reade and F. C. Walsh, *J. Electroanal. Chem.*, 2004, **561**, 203–217.
- 19 P. Niu, C. Fernández-Sánchez, M. Gich, C. Ayora and A. Roig, *Electrochim. Acta*, 2015, **165**, 155–161.
- 20 A. P. Washe, P. Lozano-Sánchez, D. Bejarano-Nosas and I. Katakis, *Electrochim. Acta*, 2013, **91**, 166–172.
- 21 A. Gomis-Berenguer, M. Gomez-Mingot, V. Montiel, A. Canals, T. Thiemann, R. O. Kadara, C. E. Banks and J. Iniesta, *RSC Adv.*, 2012, **2**, 7735–7742.
- 22 F. E. Galdino, C. W. Foster, J. A. Bonacin and C. E. Banks, *Anal. Methods*, 2015, **7**, 1208–1214.
- 23 C. W. Foster, A. P. de Souza, J. P. Metters, M. Bertotti and C. E. Banks, *Analyst*, 2015, **140**, 7598–7612.
- 24 C. W. Foster, R. O. Kadara and C. E. Banks, *Screen-Printing Electrochemical Architectures*, Springer International Publishing, 1st edn, 2015.
- 25 I. Lavagnini, R. Antiochia and F. Magno, *Electroanalysis*, 2004, **16**, 505.
- 26 E. P. Randviir, D. A. C. Brownson, J. P. Metters, R. O. Kadara and C. E. Banks, *Phys. Chem. Chem. Phys.*, 2014, **16**, 4598–4611.
- 27 L. R. Cumba, C. W. Foster, D. A. C. Brownson, J. P. Smith, J. Iniesta, B. Thakur, D. R. do Carmo and C. E. Banks, *Analyst*, 2016, DOI: 10.1039/C6AN00167J.
- 28 E. P. Randviir, J. P. Metters, J. Stainton and C. E. Banks, *Analyst*, 2013, **138**, 2970–2981.
- 29 R. T. Kachosangi, G. G. Wildgoose and R. G. Compton, *Analyst*, 2008, **133**, 888–895.
- 30 A. V. Kolliopoulos, J. P. Metters and C. E. Banks, *Anal. Methods*, 2013, **5**, 3490–3496.

