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## Total synthesis and complete configurational assignment of amphirionin-2†

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Amphirionin-2 is a linear polyketide metabolite that exhibits potent and selective cytotoxic activity against certain human cancer cell lines. We disclose herein the first total synthesis of amphirionin-2 and determination of its absolute configuration. Our synthesis featured an extensive use of cobalt-catalyzed Mukaiyama-type cyclization of  $\gamma$ -hydroxy olefins for stereoselective formation of all the tetrahydrofuran rings found in the natural product, and a late-stage Stille-type coupling for convergent assembly of the entire carbon backbone. Four candidate diastereomers of amphirionin-2 were synthesized in a unified, convergent manner, and their spectroscopic/chromatographic properties were compared with those of the authentic material. The present study culminated in the reassignment of the C5/C7 relative configuration, assignment of the C12/C18 relative configuration, and determination of the absolute configuration of amphirionin-2.

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### Introduction

Marine polyketides are an important source of new chemotherapeutic agents for the treatment of cancer.<sup>1</sup> As such, the structure, synthesis, and biological function of this class of natural products have gained significant interest from the chemical community.<sup>2</sup> Marine polyketides are mostly non-crystalline, scarcely available substances from natural sources, and their complex structures are characterized mainly by NMR spectroscopic analysis. Integrated with quantum chemical calculations that enable the prediction of chemical shifts and <sup>3</sup>J<sub>H,H</sub> values,<sup>3,4</sup> NMR-based structural assignment of stereochemically complex natural products has become more feasible than ever. Unfortunately, however, configurational assignment of remote stereogenic centers between which only negligible, if any, stereoelectronic and/or steric interactions exist, is still beyond the reach of NMR spectroscopic analysis and computational simulations.<sup>5</sup> Orchestration of chemical synthesis, NMR and other spectroscopic techniques and, where appropriate, chromatographic analysis is indispensable for achieving complete configurational assignment of complex natural products.<sup>6–8</sup>

Amphirionin-2 (putative structures **1** and **2**, Fig. 1) is a linear polyketide metabolite, isolated from cultured cells of the marine

benthic dinoflagellate *Amphidinium* sp. KCA09051 strain.<sup>9</sup> Amphirionin-2 exhibited potent cytotoxic activity against the human colon carcinoma Caco-2 cell line and the human non-small cell lung adenocarcinoma A549 cell line with IC<sub>50</sub> values of 0.1 and 0.6  $\mu\text{g mL}^{-1}$ , respectively, whereas it showed only moderate cytotoxicity against the human cervix adenocarcinoma HeLa cell line (20% growth inhibition at 1  $\mu\text{g mL}^{-1}$ ). Furthermore, amphirionin-2 displayed *in vivo* antitumor activity against murine tumor P388 cells (T/C 120% at 0.5 mg kg<sup>-1</sup>).

The gross structure of amphirionin-2 was determined on the basis of extensive 2D-NMR analyses. The relative configurations of two unique hexahydrofuro[3,2-*b*]furan moieties were individually characterized based on NOESY correlations. The relative configurations of C4/C5 and C5/C7 were deduced from conformational analyses based on *J* values and NOESY correlations. The absolute configuration of C5 was assigned on the basis of a modified Mosher analysis.<sup>10</sup> However, the relative configuration between two remote stereogenic centers C12 and

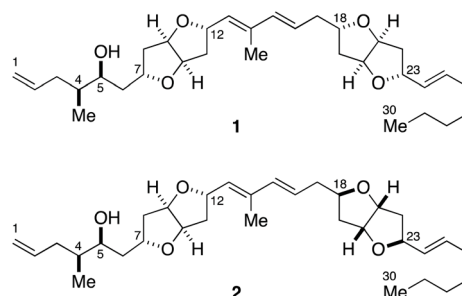


Fig. 1 Putative structures **1** and **2** of amphirionin-2.

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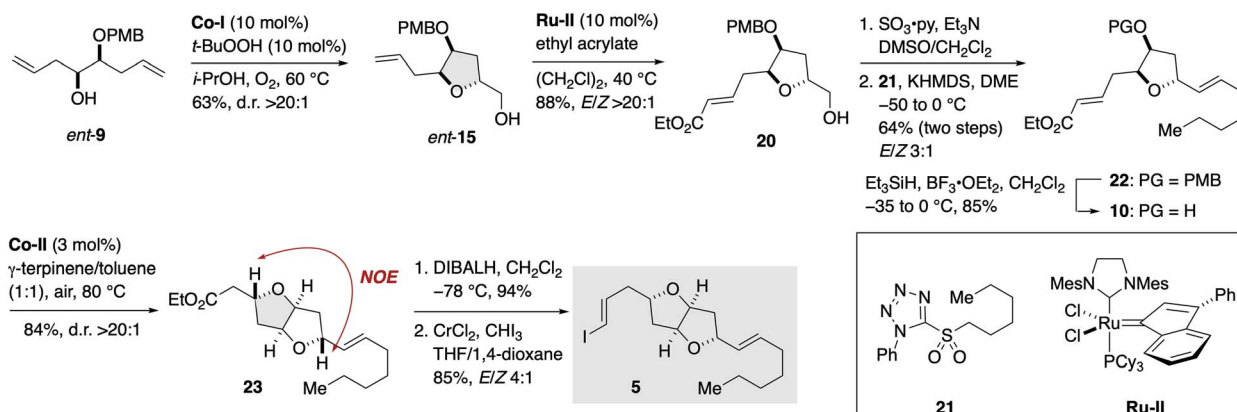
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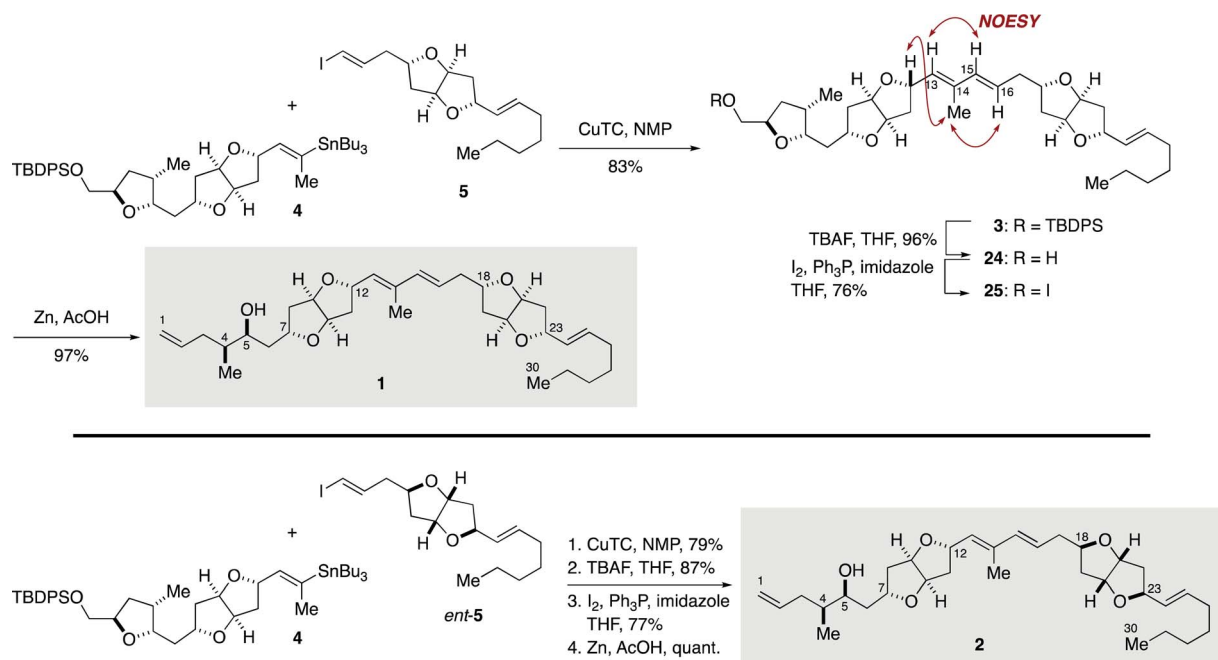
Scheme 3 Synthesis of iodoolefin 5.

be separated by flash column chromatography using silica gel. The enantiomer of 5, *i.e.*, *ent*-5, was prepared in the same manner from  $\gamma$ -hydroxy olefin 9 (see Scheme S5, ESI† for details).

Completion of the total synthesis of 1 and 2 is illustrated in Scheme 4. Stille-type reaction of vinylstannane 4 (1 equiv.) with iodoolefin 5 (1.1 equiv.) was non-trivial and required optimization of reaction conditions. Initial experiments showed that the reaction under palladium catalysis ([Pd<sub>2</sub>(dba)<sub>3</sub>·CHCl<sub>3</sub>]/Ph<sub>3</sub>As with or without CuI) provided (*E,E*)-diene 3 in only low yield and resulted in significant side reactions, including isomerization of the C15–C16 double bond and homodimerization of 5 (Table S3, ESI†). It was eventually found that the reaction was best performed by using CuTC in NMP<sup>33</sup> at room temperature, giving 3 in 83% yield with essentially no erosion of the configuration of the double bonds. The configuration of the diene moiety of 3 was confirmed to be *E,E* by NOESY

correlations and a coupling constant ( $^3J_{H_{15},H_{16}} = 15.6$  Hz). Cleavage of the TBDPS ether of 3 with TBAF delivered alcohol 24 in 96% yield. After iodination (I<sub>2</sub>, Ph<sub>3</sub>P, imidazole, 76%), the derived iodide 25 was exposed to excess zinc dust in acetic acid to furnish 1 (97%). The diastereomer 2 was synthesized from 4 and *ent*-5 in the same manner.

The <sup>1</sup>H and <sup>13</sup>C NMR spectra of 1 and 2 revealed that both compounds were not identical with natural amphirionin-2 (for assignment of <sup>1</sup>H and <sup>13</sup>C NMR signals, see Tables S4 and S5, ESI†). These results indicated the necessity of re-examination of the original structural assignment of the natural product. The <sup>1</sup>H NMR chemical shifts of the C1–C12 moiety of synthetic 1 and 2 were significantly deviated from those of the corresponding moiety of the natural product, whereas the <sup>13</sup>C NMR signals of synthetic 1 and 2 were similar to those of the authentic material and inconsistencies were limited to the



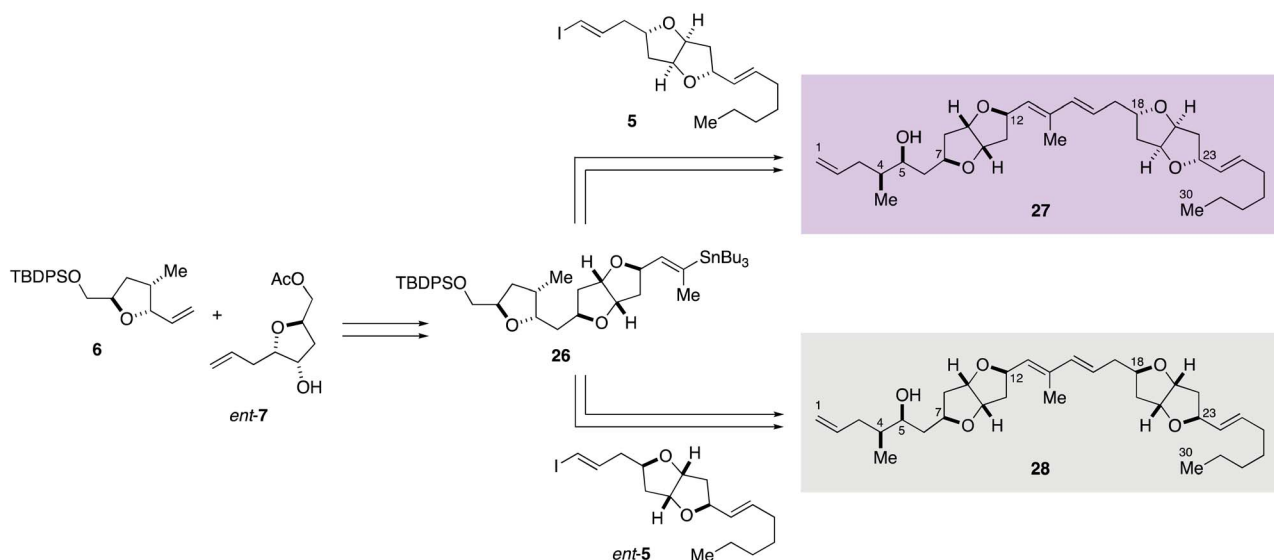
Scheme 4 Total syntheses of putative structures 1 and 2 of amphirionin-2.



C5–C10 moiety. Significantly, **1** and **2** were distinguishable from each other by  $^1\text{H}$  NMR analysis despite the C12 and C18 stereogenic centers being separated by six carbon–carbon bonds. Careful comparison of the  $^1\text{H}$  NMR spectra of **1** and **2** revealed subtle differences in signals assigned for H-11, H-16, H-17, and H-18 (Fig. S2, ESI $^\dagger$ ). With respect to these protons, the  $^1\text{H}$  NMR chemical shifts of **2** rather than **1** were in better agreement with those of natural amphirionin-2. It is known that stereoelectronic and/or steric interactions between two stereogenic centers separated by two or more methylene units are negligible by NMR spectroscopy.<sup>5</sup> In the present case, the C13–C16 conjugated diene would be responsible for unusual long-range stereochemical interactions between the C12 and C18 stereogenic centers.<sup>34</sup>

We considered that the relative configuration of C12/C18 of natural amphirionin-2 might be same as that of synthetic **2**, and that the relative configuration of C4/C5 and/or C5/C7 of the original stereochemical assignment should have been incorrectly assigned. Re-examination of NOESY correlations and  $^3J_{\text{H,H}}$  values of natural amphirionin-2 suggested that the relative configuration of C5/C7 of the natural product might be opposite to that of the proposed structures **1** and **2** (Fig. S1, ESI $^\dagger$ ).

Accordingly, diastereomers **27** and **28** were synthesized from olefins **6** and *ent*-**7** via Stille-type coupling of vinylstannane **26** and iodoolefins **5**/*ent*-**5** (Scheme 5, details are provided in Schemes S7 and S8, ESI $^\dagger$ ). The  $^1\text{H}$  NMR spectra of synthetic **27** and **28** were almost identical with each other, as anticipated, but small but significant differences were observed in H-11, H-16, H-17, H-18, and H-19 signals (Fig. 2). The apparent



Scheme 5 Syntheses of correct structure **27** of amphirionin-2 and its diastereomer **28**.

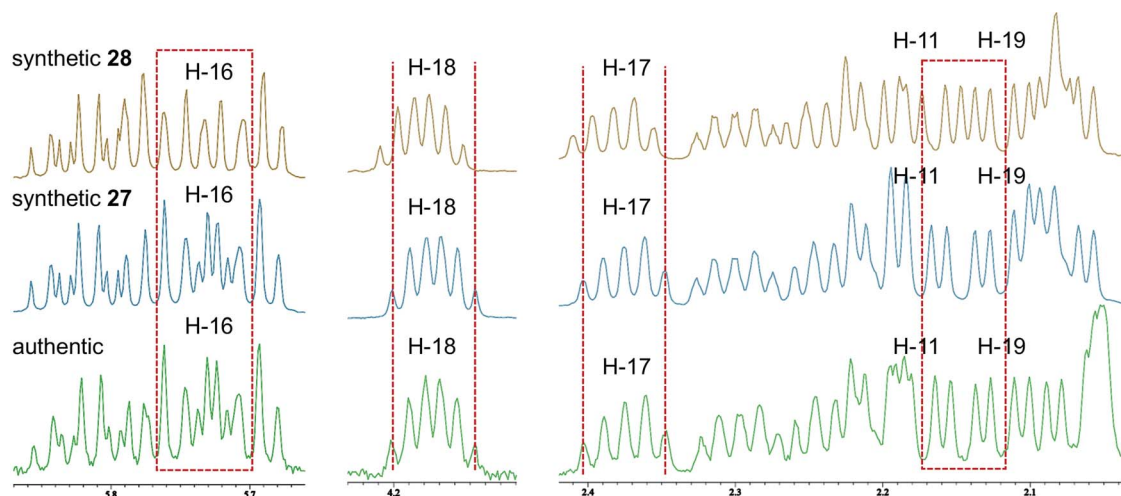


Fig. 2 Comparison of  $^1\text{H}$  NMR spectra of **27**, **28**, and natural amphirionin-2. Inconsistency observed around 2.04–2.10 ppm is ascribable to 5-OH signal.



inconsistency around 2.04–2.10 ppm in the  $^1\text{H}$  NMR spectra of synthetic **27** and **28** and natural amphirionin-2 was due to the 5-OH signal, which was firmly assigned on the basis of COSY experiments. The 5-OH signal disappeared upon addition of a drop of  $\text{D}_2\text{O}$  (Fig. S3 and S4, ESI $^\dagger$ ). Therefore, we determined that the  $^1\text{H}$  NMR spectrum of **27** matched that of natural amphirionin-2. The  $^{13}\text{C}$  NMR spectra of synthetic **27** and **28** were completely indistinguishable from each other.

Moreover, chiral HPLC analysis (Chiralpak IB N-5: 4.6 mm I.D.  $\times$  250 mm; eluent: 10% i-PrOH/*n*-hexane; flow rate: 1.0 mL  $\text{min}^{-1}$ ; UV detection: 254 nm) demonstrated that the retention time of **27**, **28**, and natural amphirionin-2 was 8.9, 7.7, and 8.8 min, respectively (Fig. S5, ESI $^\dagger$ ). Co-Injection of synthetic **27** and authentic amphirionin-2 resulted in a single peak under the above analytical conditions. Thus, we concluded that the relative configuration of amphirionin-2 is same as that of **27**. The specific rotation value of **27** ( $[\alpha]_{\text{D}}^{20} + 2.5$  (*c* 0.18,  $\text{CHCl}_3$ )) was in accordance with that of the authentic material ( $[\alpha]_{\text{D}}^{20} + 5$  (*c* 0.8,  $\text{CHCl}_3$ )). Because of the small magnitude of the specific rotation value, it was further confirmed that the circular dichroism (CD) spectrum of natural amphirionin-2 was consistent with that of **27** (Fig. S6, ESI $^\dagger$ ). Accordingly, we established that the absolute configuration of amphirionin-2 is represented by the structure **27**.

Finally, we evaluated the cytotoxic activity of our synthetic **1**, **2**, **27**, and **28** against a small panel of human cancer cell lines, including the non-small cell lung adenocarcinoma A549, the cervix adenocarcinoma HeLa, the acute T cell leukemia Jurkat, and the chronic myelogenous leukemia K562 cell lines by WST-8 assay (Fig. 3). A549 cells showed biphasic response to synthetic **1**, **2**, **27**, and **28**. The viability of A549 cells decreased to 39–65% at 10  $\mu\text{M}$ , increased to 63–73% at 30  $\mu\text{M}$ , and then underwent to 11–38% at 100  $\mu\text{M}$ . The sensitivity of A549 cells

toward synthetic **27** was more moderate than that expected from the results reported in the isolation paper.<sup>9</sup> This apparent discrepancy would be ascribable to the difference of the source of cells and/or experimental conditions, as a similar difference in potency was observed for the positive control doxorubicin:  $\text{IC}_{50}$  0.6  $\mu\text{M}$  (this work) *versus*  $\text{IC}_{50}$  0.07  $\mu\text{M}$  (ref. 9). A similar biphasic response was observed for HeLa cells upon exposure to our synthetic compounds; the cell viability declined to around 60% at 3  $\mu\text{M}$ , restored to 86–98% at 30  $\mu\text{M}$ , and dropped to 10–51% at 100  $\mu\text{M}$ . These results suggested the possibility that our synthetic compounds would have at least two different mechanisms of action in A549 and HeLa cells. In contrast, Jurkat and K562 cells responded to our synthetic compounds in a dose-dependent manner. Jurkat cells were found to be more sensitive than K562 cells toward these compounds. Overall, our synthetic **1**, **2**, **27**, and **28** showed cell line-dependent cytotoxic activity, whilst their stereochemistry did not have significant correlation with their cytotoxic potency.

## Conclusions

A unified total synthesis of four candidate stereoisomers **1**, **2**, **27**, and **28** of amphirionin-2 was completed in 17 linear steps from diol **11** or 19 linear steps from benzyloxyacetaldehyde. The salient feature of the present work is an extensive use of cobalt-catalyzed Mukaiyama-type cyclization of  $\gamma$ -hydroxy olefins in stereocontrolled construction of all the tetrahydrofuran rings of amphirionin-2. The present study illuminates the versatility of Hartung–Mukaiyama cyclization of  $\gamma$ -hydroxy olefins, and also expands the reaction scope by demonstrating the synthesis of complex 2,5-*trans*-substituted tetrahydrofurans from internal olefins (*e.g.*, **16E/16Z**  $\rightarrow$  **17**). A late-stage CuTC-mediated Stille-type reaction for convergent assembly of two hexahydrofuro[3,2-*b*]furan moieties is another important feature of our synthesis, which enabled rapid access to four candidate stereoisomers.

The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopic data of originally assigned structures **1** and **2** showed non-identity of these compounds with natural amphirionin-2 and suggested the necessity to reassign the C5/C7 relative configuration. Eventually, the relative configuration of this natural product was fully established on the basis of the  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopic data and chiral HPLC chromatograms of **27** and **28** with authentic reference. The absolute configuration was determined by comparing the specific rotation value and CD spectroscopic data of **27** and **28** with those of the authentic material. Thus, it was concluded that the absolute configuration of amphirionin-2 is shown by the structure **27**.

## Author contributions

H. F. conceived the project and directed the research. S. K. and D. M. executed synthetic experiments and collected compound characterization data. H. F., S. K., D. M., T. S. performed NMR analysis of key compounds. M. T. provided authentic amphirionin-2 and significant intellectual contribution for its configurational assignment. All authors composed the manuscript and the ESI $^\dagger$ .

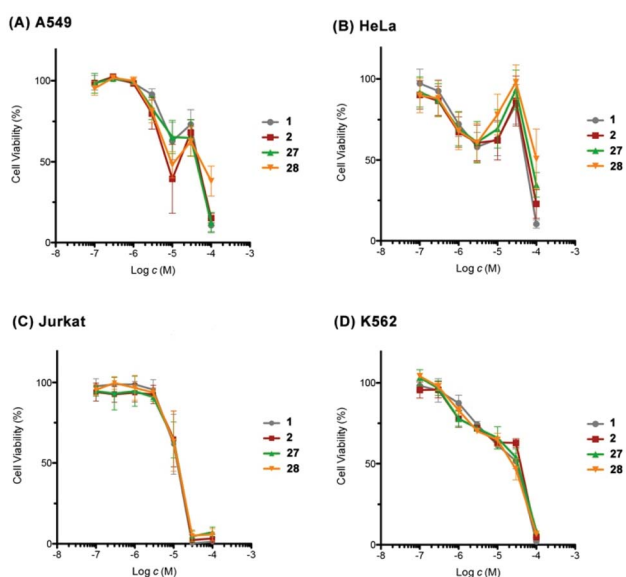


Fig. 3 Cytotoxic activity of synthetic amphirionin-2 (**27**) and its diastereomers **1**, **2**, and **28**. For an enlarged version of this figure, see Fig. S7, ESI $^\dagger$ .



## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

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## Notes and references

- For recent reviews, see: (a) C. Jiménez, *ACS Med. Chem. Lett.*, 2018, **9**, 959–961; (b) C. Alves, J. Silva, S. Pinteus, H. Gaspar, M. C. Alpoim, L. M. Botana and R. Pedrosa, *Front. Pharmacol.*, 2018, **9**, 777; (c) K.-H. Altmann, *Chimia*, 2017, **71**, 646–652; (d) D. J. Newman and G. M. Cragg, *J. Nat. Prod.*, 2016, **79**, 629–661.
- For recent reviews, see: (a) A. R. Carroll, B. R. Copp, R. A. Davis, R. A. Keyzers and M. R. Prinsep, *Nat. Prod. Rep.*, 2020, **37**, 175–223; (b) A. R. Carroll, B. R. Copp, R. A. Davis, R. A. Keyzers and M. R. Prinsep, *Nat. Prod. Rep.*, 2019, **36**, 122–173.
- For selected recent examples, see: (a) G. Lauro, P. Das, R. Riccio, D. S. Reddy and G. Bifulco, *J. Org. Chem.*, 2020, **85**, 3297–3306; (b) M. M. Zanardi, M. O. Marcarino and A. M. Sarotti, *Org. Lett.*, 2020, **22**, 52–56; (c) J. B. Neupane, R. P. Neupane, Y. Luo, W. Y. Yoshida, R. Sun and P. G. Williams, *Org. Lett.*, 2019, **21**, 8449–8453; (d) A. G. Kutateladze and T. Holt, *J. Org. Chem.*, 2019, **84**, 8297–8299; (e) E. D. Shepherd, B. S. Dyson, W. E. Hak, Q. N. N. Nguyen, M. Lee, M. J. Kim, T. Sohn, D. Kim, J. W. Burton and R. S. Paton, *J. Org. Chem.*, 2019, **84**, 4971–4991; (f) J.-W. Tang, H.-C. Xu, W.-G. Wang, K. Hu, Y.-F. Zhou, R. Chen, X.-N. Li, X. Du, H.-D. Sun and P.-T. Puno, *J. Nat. Prod.*, 2019, **82**, 735–740; (g) E. Roulland, H. Solanki, K. Calabro, M. Zubia, G. Genta-Jouve and O. P. Thomas, *Org. Lett.*, 2018, **20**, 2311–2314; (h) J. Wu, P. Lorenzo, S. Zhong, M. Ali, C. P. Butts, E. L. Myers and V. K. Aggarwal, *Nature*, 2017, **547**, 436–440. and references cited therein.
- For recent reviews, see: (a) G. Bifulco, P. Dambruoso, L. Gomez-Paloma and R. Riccio, *Chem. Rev.*, 2007, **107**, 3744–3779; (b) S. Di Micco, M. G. Chini, R. Riccio and G. Bifulco, *Eur. J. Org. Chem.*, 2010, 1411–1434; (c) M. Elyashberg, A. J. Williams and K. Blinov, *Nat. Prod. Rep.*, 2010, **27**, 1296–1328; (d) M. W. Lodewyk, M. R. Siebert and D. J. Tantillo, *Chem. Rev.*, 2012, **112**, 1839–1862; (e) N. Grimblat and A. M. Sarotti, *Chem.–Eur. J.*, 2016, **22**, 12246–12261.
- Y. Kobayashi, C.-H. Tan and Y. Kishi, *Angew. Chem., Int. Ed.*, 2000, **39**, 4279–4281.
- For reviews, see: (a) K. C. Nicolaou and S. A. Snyder, *Angew. Chem., Int. Ed.*, 2005, **44**, 1012–1044; (b) M. E. Maier, *Nat. Prod. Rep.*, 2009, **26**, 1105–1124; (c) Y. Usami, *Mar. Drugs*, 2009, **7**, 314–330; (d) T. L. Suyama, W. H. Gerwick and K. L. McPhail, *Bioorg. Med. Chem.*, 2011, **19**, 6675–6701; (e) T. F. Molinski and B. I. Morinaka, *Tetrahedron*, 2012, **68**, 9307–9343; (f) B. K. Chhetri, S. Lavoie, A. M. Sweeney-Jones and J. Kubanek, *Nat. Prod. Rep.*, 2018, **35**, 514–531; (g) I. Paterson and N. Lam, *Eur. J. Org. Chem.*, 2020, 2310–2320.
- For selected recent examples, see: (a) Y. Wakamiya, M. Ebine, N. Matsumori and T. Oishi, *J. Am. Chem. Soc.*, 2020, **142**, 3472–3478; (b) M. Hönig and E. M. Carreira, *Angew. Chem., Int. Ed.*, 2020, **59**, 1192–1196; (c) A. S. Burns and S. D. Rychnovsky, *J. Am. Chem. Soc.*, 2019, **141**, 13295–13300; (d) P. Sondermann and E. M. Carreira, *J. Am. Chem. Soc.*, 2019, **141**, 10510–10519; (e) N. A. Isley, Y. Endo, Z.-C. Wu, B. C. Covington, L. B. Bushin, M. R. Seyedsayamdost and D. L. Boger, *J. Am. Chem. Soc.*, 2019, **141**, 17361–17369; (f) C. L. Hugelshofer, V. Palani and R. Sarpong, *J. Am. Chem. Soc.*, 2019, **141**, 8431–8435; (g) Z. Feng, T. K. Allred, E. E. Hurlow and P. G. Harran, *J. Am. Chem. Soc.*, 2019, **141**, 2274–2278; (h) B. Görlicke, M. F. Bieber, K. E. Mohr and D. Menche, *Angew. Chem., Int. Ed.*, 2019, **58**, 13019–13023; (i) K. Nishikawa, K. Morita, S. Hashimoto, A. Hoshino, T. Ikeuchi, M. Kumagai and Y. Morimoto, *Angew. Chem., Int. Ed.*, 2019, **58**, 10168–10172.
- For our studies on the structure elucidation of complex natural products *via* chemical synthesis, see: (a) H. Fuwa, M. Ebine, A. J. Bourdelais, D. G. Baden and M. Sasaki, *J. Am. Chem. Soc.*, 2006, **128**, 16989–16999; (b) H. Fuwa, K. Ishigai, K. Hashizume and M. Sasaki, *J. Am. Chem. Soc.*, 2012, **134**, 11984–11987; (c) K. Ishigai, H. Fuwa, K. Hashizume, R. Fukazawa, Y. Cho, M. Yotsu-Yamashita and M. Sasaki, *Chem.–Eur. J.*, 2013, **19**, 5276–5288; (d) H. Fuwa, T. Muto, K. Sekine and M. Sasaki, *Chem.–Eur. J.*, 2014, **20**, 1848–1860; (e) H. Fuwa, Y. Okuaki, N. Yamagata and M. Sasaki, *Angew. Chem., Int. Ed.*, 2015, **54**, 868–873; (f) H. Fuwa, N. Yamagata, Y. Okuaki, Y. Ogata, A. Saito and M. Sasaki, *Chem.–Eur. J.*, 2016, **22**, 6815–6829; (g) R. Isaka, L. Yu, M. Sasaki, Y. Igarashi and H. Fuwa, *J. Org. Chem.*, 2016, **81**, 3638–3647; (h) K. Sakamoto, A. Hakamata, M. Tsuda and H. Fuwa, *Angew. Chem., Int. Ed.*, 2018, **57**, 3801–3805; (i) K. Sakamoto, A. Hakamata, A. Iwasaki, K. Suenaga, M. Tsuda and H. Fuwa, *Chem.–Eur. J.*, 2019, **25**, 8528–8542; (j) K. Sakamoto and H. Fuwa, *J. Synth. Org. Chem., Jpn.*, 2019, **77**, 831–840.
- K. Kumagai, M. Minamida, M. Akakabe, M. Tsuda, Y. Konishi, A. Tominaga, M. Tsuda, E. Fukushima and J. Kawabata, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 635–638.
- I. Ohtani, T. Kusumi, Y. Kashman and H. Kakisawa, *J. Am. Chem. Soc.*, 1991, **113**, 4092–4096.
- (a) S. Inoki and T. Mukaiyama, *Chem. Lett.*, 1990, 67–70; (b) T. Mukaiyama and T. Yamada, *Bull. Chem. Soc. Jpn.*, 1995, **68**, 17–35.
- (a) B. M. Pérez, D. Schuch and J. Hartung, *Org. Biomol. Chem.*, 2008, **6**, 3532–3541; (b) S. Ali, H. Milanezi, T. M. F. Alves, C. F. Tormena and M. A. B. Ferreira, *J. Org. Chem.*, 2018, **83**, 7694–7713.
- (a) D. Schuch, P. Fries, M. Dönges, B. M. Pérez and J. Hartung, *J. Am. Chem. Soc.*, 2009, **131**, 12918–12920; (b)





- P. Fries, D. Halter, A. Kleinschek and J. Hartung, *J. Am. Chem. Soc.*, 2011, **133**, 3906–3912.
- 14 For reviews, see: (a) J. K. Stille, *Angew. Chem., Int. Ed. Engl.*, 1986, **25**, 508–524; (b) V. Farina, V. Krishnamurthy and W. J. Scott, *Organomet. React.*, 1997, **50**, 1–652; (c) P. Espinet and A. M. Echavarren, *Angew. Chem., Int. Ed.*, 2004, **43**, 2–32; (d) M. M. Heravi, E. Hashemi and F. Azimian, *Tetrahedron*, 2014, **70**, 7–21.
- 15 For a review, see: S. J. Connon and S. Blechert, *Angew. Chem., Int. Ed.*, 2003, **42**, 1900–1923.
- 16 For representative examples of Mukaiyama cyclization in total synthesis, see: (a) S. Takahashi, A. Kubota and T. Nakata, *Angew. Chem., Int. Ed.*, 2002, **41**, 4751–4753; (b) P. A. Evans, J. Cui, S. J. Gharpure, A. Polosukhin and H.-R. Zhang, *J. Am. Chem. Soc.*, 2003, **125**, 14702–14703; (c) J. Wang and B. L. Pagenkopf, *Org. Lett.*, 2007, **9**, 3703–3706; (d) L. C. Dias and M. A. B. Ferreira, *J. Org. Chem.*, 2012, **77**, 4046–4062; (e) N. A. Morra and B. L. Pagenkopf, *Tetrahedron*, 2013, **69**, 8632–8644; (f) G. Valot, D. Mailhol, C. S. Regens, D. P. O'Malley, E. Godineau, H. Takikawa, P. Philipps and A. Fürstner, *Chem.–Eur. J.*, 2015, **21**, 2398–2408; (g) D. C. Akwaboah, D. Wu and C. J. Forsyth, *Org. Lett.*, 2017, **19**, 1180–1183; (h) A. Roushanbakhti, Y. Liu, P. C. M. Winship, M. J. Tucker, W. M. Akhtar, D. S. Walter, G. Wrigley and T. J. Donohoe, *Angew. Chem., Int. Ed.*, 2017, **56**, 14883–14887; (i) S. Takahashi, D. Satoh, M. Hayashi, K. Takahashi, K. Yamaguchi, T. Nakamura and H. Koshino, *J. Org. Chem.*, 2016, **81**, 11222–11234; (j) M. Heinrich, J. J. Murphy, M. K. Ilg, A. Letort, J. Flasz, P. Philipps and A. Fürstner, *Angew. Chem., Int. Ed.*, 2018, **57**, 13575–13581; (k) M. Heinrich, J. J. Murphy, M. K. Ilg, A. Letort, J. T. Flasz, P. Philipps and A. Fürstner, *J. Am. Chem. Soc.*, 2020, **142**, 6409–6422.
- 17 Compound **11** is commercially available. Also, it can be prepared from benzyloxyacetaldehyde in two steps, see the ESI† for details.
- 18 (a) C. Palmer, N. A. Morra, A. C. Stevens, B. Bajtos, B. P. Machin and B. L. Pagenkopf, *Org. Lett.*, 2009, **11**, 5614–5617; (b) N. A. Morra and B. L. Pagenkopf, *Eur. J. Org. Chem.*, 2013, 756–760; (c) B. Morra, N. A. Morra, D. G. MacDonald and B. L. Pagenkopf, *Synthesis*, 2020, **52**, 847–852.
- 19 An attempted hydrogenolysis of the TBDPS-protected **13** was too slow to complete within a reasonable time.
- 20 (a) D. B. Dess and J. C. Martin, *J. Am. Chem. Soc.*, 1991, **113**, 7277–7287; (b) K. Takai, Y. Hotta, K. Oshima and H. Nozaki, *Tetrahedron Lett.*, 1978, 2417–2420.
- 21 The substrate scope of this new sequential Dess–Martin oxidation/Takai methylenation protocol will be reported in due course.
- 22 R. N. MacCoss, E. P. Balskus and S. V. Ley, *Tetrahedron Lett.*, 2003, **44**, 7779–7781.
- 23 Compound **9** was available in four steps from *trans*-1,4-dichloro-2-butene or in eight steps from D-(–)-diethyl tartrate. See the ESI† for details.
- 24 H. Fuwa, T. Suzuki, H. Kubo, T. Yamori and M. Sasaki, *Chem.–Eur. J.*, 2011, **17**, 2678–2688.
- 25 M. Scholl, S. Ding, C. W. Lee and R. H. Grubbs, *Org. Lett.*, 1999, **1**, 953–956.
- 26 B. M. Pérez and J. Hartung, *Tetrahedron Lett.*, 2009, **50**, 960–962.
- 27 E. J. Corey and P. L. Fuchs, *Tetrahedron Lett.*, 1972, **13**, 3769–3772.
- 28 (a) J.-F. Betzer, F. Delalogue, B. Muller, A. Pancrazi and J. Prunet, *J. Org. Chem.*, 1997, **62**, 7768–7780; (b) D. Zurwerra, J. Gertsch and K.-H. Altmann, *Org. Lett.*, 2010, **12**, 2302–2305. See also ref. 8i.
- 29 L. Jafarpour, H.-J. Schanz, E. D. Stevens and S. P. Nolan, *Organometallics*, 1999, **18**, 5416–5419.
- 30 (a) P. R. Blakemore, W. J. Cole, P. J. Kocienski and A. Morley, *Synlett*, 1998, 26–28; (b) P. R. Blakemore, *J. Chem. Soc., Perkin Trans. 1*, 2002, 2563–2585; (c) C. Aïssa, *Eur. J. Org. Chem.*, 2009, 1831–1844.
- 31 T.-S. Li, J.-T. Li and H.-Z. Li, *J. Chromatogr. A*, 1995, **715**, 372–375.
- 32 (a) K. Takai, K. Nitta and K. Utimoto, *J. Am. Chem. Soc.*, 1986, **108**, 7408–7410; (b) D. A. Evans and W. C. Black, *J. Am. Chem. Soc.*, 1993, **115**, 4497–4513.
- 33 (a) G. D. Allred and L. S. Liebeskind, *J. Am. Chem. Soc.*, 1996, **118**, 2748–2749; (b) H. Prokopcová and C. O. Kappe, *Angew. Chem., Int. Ed.*, 2009, **48**, 2276–2286.
- 34 For related examples, see: (a) J. Willwacher, N. Kausch-Busies and A. Fürstner, *Angew. Chem., Int. Ed.*, 2012, **51**, 12041–12046; (b) J. Zhou, B. Gao, Z. Xu and T. Ye, *J. Am. Chem. Soc.*, 2016, **138**, 6948–6951; (c) B. Y. Han, N. Y. S. Lam, C. I. MacGregor, J. M. Goodman and I. Paterson, *Chem. Commun.*, 2018, **54**, 3247–3250.

