

Cite this: *Chem. Sci.*, 2020, 11, 10594

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 6th June 2020

Accepted 7th July 2020

DOI: 10.1039/d0sc03164j

rsc.li/chemical-science

The evolution of molecular machines through interfacial nanoarchitectonics: from toys to tools

Katsuhiko Ariga *ab

Molecular machines are often regarded as molecular artworks and sometimes as fancy molecular toys. However, many researchers strive to operate molecular machines as useful tools for realistic practical applications. In this perspective article, shifting the working environment of molecular machines from solution to interfacial media is discussed from the viewpoint of their evolution from scientific toys to useful tools. Following a short description of traditional research into molecular machines in solution and their nanotechnological manipulation on clean solid surfaces, pioneering research into molecular machine operation at dynamic interfaces, such as liquid surfaces, is discussed, along with cutting-edge research into molecular machine functions in living cells and their models. Biomolecular machines within organisms are the products of evolution over billions of years. We may nanoarchitect such sophisticated functional systems with artificial molecular machines within much shorter periods.

Introduction

For various issues facing society, including energy,^{1–4} environmental,^{5–8} and biomedical^{9–12} problems, the development of sophisticated material systems with superior efficiency and specificity is required. In addition to improving the intrinsic properties of materials themselves, precise structural control of material components and their organization is an undoubtedly important factor for optimizing their function.^{13–16} Ideal functional systems can be seen in biological systems, in which highly complex functions are commonly found, as exemplified

by photosynthetic processes and signal transduction pathways.^{17,18} Functional molecules such as proteins and nucleic acids co-operatively work like machines and devices. Learning from these biological examples, molecular machines and molecular devices could also be key players in achieving highly sophisticated materials to meet our societal demands.

The importance of molecular machines was recognized in previous centuries. With pioneering research efforts for the design and synthesis of molecular machines, the 2016 Nobel Prize in Chemistry was awarded to Jean-Pierre Sauvage,¹⁹ Sir J. Fraser Stoddart²⁰ and Bernard L. Feringa.²¹ In addition to the work of these awardees, excellent examples of molecules with machine-like functions have been continuously reported.^{22–31} Some of these molecules can be regarded as molecular artworks and sometimes as fancy molecular toys. However, many researchers have dreams to operate molecular machines as useful tools in realistic practical applications, although it is not an easy process. High-level control of molecular machines and their sophisticated organization would be required to accomplish this dream.

Molecular machines originated as products of transitional chemistry fields, such as organic chemistry and supramolecular chemistry, where the observation of a single molecular machine is not always easy. This insufficiency is fortunately compensated for by developments in nanotechnology. Various probe microscopies and advanced analytical methods now enable us to directly observe, analyze, and manipulate a single molecular machine.^{32,33}

For the next stage, the organization of molecular machines using nanotechnology techniques is necessary. This new phase of molecular machine research can be supported by an emerging concept called nanoarchitectonics,^{34–36} which couples

^aWPI Research Center for Materials Nanoarchitectonics (MANA), National Institute for Materials Science (NIMS), 1-1 Namiki, Tsukuba, Ibaraki 305-0044, Japan. E-mail: ARIGA.Katsuhiko@nims.go.jp

^bGraduate School of Frontier Sciences, The University of Tokyo, 5-1-5 Kashiwanoha, Kashiwa, Chiba 277-8561, Japan



Katsuhiko Ariga received his PhD from the Tokyo Institute of Technology in 1990. He joined the National Institute for Materials Science (NIMS) in 2004 and is currently the leader of the Supermolecules Group and principal investigator of the World Premier International (WPI) Research Centre for Materials Nanoarchitectonics (MANA), NIMS. He has also been appointed as a professor in The University of Tokyo.



nanotechnology and other research fields including organic chemistry, supramolecular chemistry, and biochemistry, as pioneered by Masakazu Aono.^{37,38} The aim of nanoarchitectonics is to use molecular units to construct functional material systems with advanced features, which can be fabricated into interfacial structures exhibiting hierarchical organization.^{39–41} In order to make molecular machines really useful, they have to be fixed and organized onto defined sites such as interfaces. Interfaces are beneficial environments for connecting molecular functions with device outputs and biological mechanisms.^{42–44} Thus, molecular machines can evolve from toys to tools through the use of interfacial nanoarchitectonics.

In this perspective article, a shift of the working environments of molecular machines from solution to interfacial media is discussed from the viewpoint of the evolution of molecular machines from scientific toys to useful tools (Fig. 1). Following a short description of traditional research into molecular machines in solution and their nanotechnological manipulation at clean solid surfaces, pioneering research into molecular machine operation at dynamic interfaces such as liquid surfaces is introduced, in addition to cutting-edge research into molecular machine functions in living cells and their models.



Fig. 1 The shift in the working environments of molecular machines, for the evolution of molecular machines from scientific toys to useful tools.

Molecular machines in solution

Pioneering operations of molecular machines were demonstrated in solution, where intentional motions of well-designed molecules and supermolecules were induced upon application of external stimuli. As exemplified by the Nobel prize machines (Fig. 2), rotational re-arrangement of integrated molecular rings (catenane),^{45,46} switching of ring positions within a rotaxane chain (molecular shuttle),⁴⁷ and stepwise and directional rotation of half of a molecule relative to the remaining half (molecular rotor and/or molecular motor),⁴⁸ were realized with inputs of external stimuli. These examples can be regarded as masterpiece molecules, achieved through expert design employing organic chemistry and stimulus-responsive strategies in supramolecular chemistry.

Molecular machines in solution environments have been actively researched on the strong basis of organic chemistry and supramolecular chemistry. Undoubtedly, these research activities support the fundamental development of molecular machine science. Various designs and motion patterns of molecular machines have been proposed. For example, Qu and co-workers successfully synthesized a molecular machine, which exhibits a coupling of the translational motion of a molecular shuttle and rotational motion of a molecular motor (Fig. 3).⁴⁹ A ring part of



Fig. 2 Pioneering examples of molecular machines (a catenane, a molecular shuttle, and a molecular motor). These schemes were provided courtesy of Dr Masayuki Takeuchi, National Institute for Materials Science.





Fig. 3 A molecular machine with the coupling of the translational motion of a molecular shuttle and the rotational motion of a molecular motor. Reproduced with permission ref. 49. Copyright 2019, American Chemical Society.

a bistable [1]rotaxane unit is also linked to a molecular motor. Photo-driven *cis-trans* isomerization of the motor induces a sliding motion of the molecular shuttle. The use of nanoarchitectonics to couple different molecular motions is an important step in combining individual machines into a unified functional system. Well-considered molecular design sometimes induces larger amplitude molecular motions by the same operational principal. The molecular shuttle shown in Fig. 4 possesses branched chain structures, which enable shuttle motions between two stations to cause larger molecular shape shifts named molecular zippers.⁵⁰ This type of wisely designed nanoarchitectonics would be useful to convert molecular motion into larger actions such as material actuation.

Molecular machines on clean solid surfaces

As another playground for molecular machines, clean solid surfaces are recognized as ideal environments for the



Fig. 4 A molecular shuttle with branched chain structures for shape shifts, named a molecular zipper. Reproduced with permission ref. 50. Copyright 2019, American Chemical Society.

evaluation of molecular shapes and motions with excellent structural precision. Although in most cases operational conditions are impractical, such as ultralow temperature and high vacuum, the direct and precise observation of a single molecule provides indispensable insights for molecular machine science.^{51,52} Experimental approaches under ultra-high vacuum can avoid the contribution of Brownian motion. High-resolution observations of molecular machines are usually obtained with scanning tunneling microscopy (STM). Driving fuels can be electronically supplied from the STM tip.

One representative category of molecular machine research on clean solid surfaces is centered around the nanocar (or molecular car).^{53,54} Nanocars can be regarded as molecular machines with automobile-like motions. Mechanisms of a single molecule were semi-classically explained on the basis of quantum incoherence of their mechanical degrees of freedom by Joachim and Gimzewski.^{55,56} Tour and co-workers demonstrated that the motional mechanisms of their nanocar on a solid surface are almost classical under specific conditions.⁵⁷ Built upon these basic understandings, molecular machine operations on clean solid surfaces have progressed in the last decade. As a demonstration of progress in this research field, the worldwide nanocar race was held in 2017, with 6 teams from France, Germany, Switzerland, USA, USA/Austria, and Japan.^{58,59} Operations of nanocars with regulated translational motions driven by electrical stimuli from the STM tip without touching the nanocar molecule were demonstrated in public.

In addition to experimental work, simulation approaches have been applied to figure out the principles of molecular machine motions, because a single molecule fixed on a clean solid surface is an ideal situation for theoretical research. For example, Abbasi-Pérez, Kantorovich, and co-workers simulated the motions of chiral molecular walkers on a clean solid surface.⁶⁰ The diffusive motions of the chiral molecular walker, 1,3-bis(imidazol-1-ylmethyl)-5(1-phenylethyl)benzene (Fig. 5), on a Cu(110) surface along the Cu rows were estimated by kinetic Monte Carlo simulations. The molecular walker moves from one position to the nearest lattice in an inchworm motion, whereby the walker steps first with the rear foot and next with the front foot. This two-step mechanism with its asymmetric

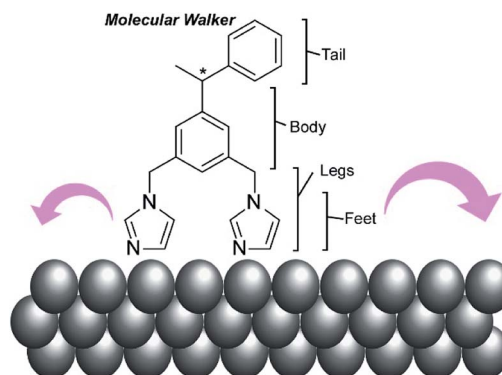


Fig. 5 A chiral molecular walker, 1,3-bis(imidazol-1-ylmethyl)-5(1-phenylethyl)benzene, on a Cu(110) surface.



design makes the walker diffuse preferentially in a particular direction with Brownian ratchets. The obtained knowledge is also useful for the separation of enantiomers through controlled diffusion on the surface upon applying appropriate external signals.

The accumulation and synchronization of the motions of numerous molecular machines nanoarchitected on surfaces are rational ways to connect molecular-level motions to macroscopic mechanical material deformations. For example, the synchronized photo-isomerizations of huge numbers of azobenzene arrays on a surface can induce motion and orientation control of bulk liquids and liquid crystalline phases (Fig. 6A).⁶¹ The accumulation of molecular shuttle motions on a surface of cantilevers can induce mechanical deformation of much larger objects of the cantilevers (Fig. 6B).⁶² In more basic research, Soe *et al.* visually demonstrated train-like actions of single molecular gears on a Pb(111) surface using low-temperature STM (Fig. 7).⁶³ The molecular gears with six teeth and a diameter of 1.2 nm are fixed on a single Cu adatom with an exact interval of 1.9 nm. Exact positioning between neighboring gears enables



Fig. 7 The train-like actions of single molecular gears on a Pb(111) surface.

the transmission of gear rotations reversibly through tooth-to-tooth molecular mechanics. The concept of coupling gear motions could be extended to a molecular-level mechanical calculating machine.

Molecular machines at dynamic liquid interfaces

Although clean solid surfaces are ideal environments to analyze detailed motions and functions of molecular machines, these media are not always adaptable for practical applications under ambient conditions. As more dynamically adaptable interfaces, liquid interfaces, such as the air–water interface, have been used for molecular machine functions. The air–water interface is actually an attractive medium for supramolecular chemistry, as it possesses several unique properties such as highly enhanced capabilities of molecular interaction and significantly shifted equilibrium.^{64,65} In addition, the extremely anisotropic nature of motional freedom at the air–water interface with macroscopically deformable lateral dimensions and a nanoscopically confined vertical dimension can couple macroscopic mechanical motions and molecular functions.^{66–68} The latter characteristics of the air–water interface enable us to operate molecular machine functions through hand-like macroscopic motions. For example, cavity-flat conversions of a molecular catcher (steroid cyclophane) embedded in an air–water interface can be repeatedly and reversibly accomplished through compression and expansion of their monolayer (Fig. 8).^{69,70} The capture and release of a target molecule by the molecular catcher can be controlled with hand-like macroscopic motions.

The conformation of molecular receptors can be subtly tuned through mechanical processes to intentionally alter binding efficiency and selectivity to target guest molecules. For example, lateral compression of a monolayer of cholesterol-substituted triazacyclononane, as a molecular receptor for nucleic acid bases, optimizes discrimination between uracil and thymine derivatives,⁷¹ which cannot be distinguished by naturally occurring DNA and RNA. Reversion of *enantio*-selectivity of aqueous amino acids depending on lateral pressures



Fig. 6 The accumulation and synchronization of motions of numerous molecular machines nanoarchitected on surface environments: (A) synchronized actions of the photo-isomerization of huge numbers of azobenzene arrays on a surface for controlling bulk liquids; and (B) the accumulation of molecular shuttle motions on a surface of cantilevers for the mechanical deformation of cantilevers.





Fig. 8 Cavity-flat conversions of a molecular catcher (steroid cyclophane) embedded at an air–water interface, accomplished through the compression and expansion of the monolayer.

was also realized through tuning the conformations of a molecular receptor, cholesterol-armed cyclen complex, by mechanical compression of the receptor monolayers (Fig. 9).⁷² In Fig. 10, the mechanical tuning of molecular receptors is compared with the traditional doctrines of molecular recognition. The basic principle of molecular recognition relies on the most stable complex structure between the host (receptor) and guest (Fig. 10A).^{73–75} If this can be regarded as the first generation of molecular recognition, the second generation of molecular recognition should be the switching of receptor structures by external stimuli, as seen in the pioneering example of photo-switchable azobenzene-type receptors by Shinkai (Fig. 10B).^{76,77} This switching mechanism utilizes two or more stable states to control recognition capability by external stimuli. Unlike these two traditional mechanisms, receptor tuning at a dynamic interface is based on selection and optimization from numerous conformational candidates of receptors (Fig. 10C).^{78,79} In this third-generation methodology, desirable efficiency and selectivity of molecular recognition can be tuned among numerous possibilities during continuous deformation of receptor structures.

Conformation tuning of molecules can be done in both analogue mode and digital mode with interfacial nanoarchitectonics. The analogue tuning of the dihedral angles of the binaphthyl unit as a model unit was demonstrated by the continuous mechanical compression of a monolayer of

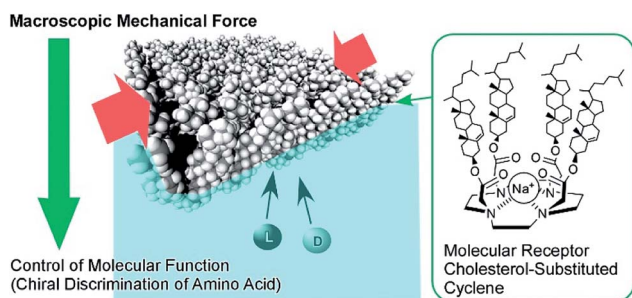


Fig. 9 The *enantio*-selectivity control of aqueous amino acids through tuning the conformations of a molecular receptor, a cholesterol-armed cyclen complex, *via* mechanical compression.



Fig. 10 Molecular recognition modes: (A) fixed state mode; (B) switching mode; and (C) tuning mode.

molecular pliers, in which two oligo(ethylene oxide) hydrophilic heads and two alkyl hydrophobic tails are attached to the binaphthyl core (Fig. 11A).⁸⁰ Mechanical compression of the molecular pliers monolayer induces continuous shifts of the dihedral angles of the binaphthyl unit. This analogue molecular tuning demonstrates that molecular conformations can be precisely manipulated using macroscopic lateral forces at the air–water interface. The applied forces are calculated to pN levels corresponding to those for delicate biological events such as DNA hybridization, myosin walking, and protein rotation and are much smaller than artificial processes such as photo-



Fig. 11 The conformation tuning of a binaphthyl unit: (A) analogue mode with molecular pliers; and (B) digital mode with a simple binaphthyl compound in a matrix lipid.



isomerization. When non-amphiphilic binaphthyl molecules are embedded within a monolayer of conventional amphiphilic molecules, the dihedral angles of the binaphthyl molecules are changed discontinuously upon compression of the mixed monolayer (Fig. 11B).⁸¹ The latter digital control of molecular conformation is accompanied by phase separation of binaphthyl crystals and dissolution of the crystals in a two-dimensional amphiphilic solvent. While analogue tuning of amphiphilic molecular pliers allows small continuous changes of the binaphthyl conformation in a cisoid to cisoid (closed to closed) transformation, the latter digital conversion is accompanied with the helical transformation of cisoid to transoid (closed to open, right to left) conversion.

The regulation of molecular motions and conformations can be converted to macroscopic properties at a dynamic interface. Gengler, Feringa, and co-workers demonstrated bidirectional control of the surface tension of a macroscopic liquid surface (Fig. 12).⁸² They utilized photo-switching of a Langmuir monolayer of a photoactive amphiphile at the air–water interface. They used an amphiphile containing a bis(thiaxanthylidene) unit as a central core, with two hydrophilic tails and two hydrophobic tails at each side of the central core. Photoisomerization of the *anti*-folded to *syn*-folded geometry of the bis(thiaxanthylidene) unit as a central core induced up and down shifts of the surface tension of the water. Surface pressure increased upon photoisomerization at the two-dimensional low-density state, whereas a high-density state of the amphiphile resulted in a drop in surface pressure upon photoirradiation, governed by the stacking state of the *syn*-folded conformers. Thus, the control of stacking states by addition of a matrix amphiphile, dipalmitoylphosphatidylcholine, modifies the regulation of surface tension. This means that not only is the photo-isomerization of responsive molecules an important factor for regulating surface tension, but the interfacial nanoarchitectonics of the components also are.



Fig. 12 The bidirectional control of the surface tension of a macroscopic liquid surface through the photo-switching of a photoactive amphiphile with a bis(thiaxanthylidene) unit.

Photonic properties, such as light emission, are sometimes regulated by rotational motions of molecular rotors. Therefore, controlling the two-dimensional environments of molecular rotor motions using interfacial nanoarchitectonics can lead to regulated conversion of molecular motion into photonic outputs. At the air–water interface, the free volume for rotation of molecular rotors can be tuned by lateral pressures and co-existing matrix amphiphiles. In the case of the BODIPY-type molecular rotor, 4-farnesyloxyphenyl-4,4-difluoro-4-bora-3a,4a-diaza-*s*-indacene, the restriction of rotational motions results in fluorescence emission (Fig. 13A).⁸³ With this molecular design, the cross-sectional area is not large enough to give sufficient free volume for BODIPY rotor rotation in the condensed state. Therefore, fluorescence emission from the BODIPY rotor can be controlled by various influential factors such as two-dimensional surface pressures and the compressibility of the matrix monolayer. In contrast, the rotation of another molecular rotor, a julolidine derivative with a cholesteryl hydrophobic moiety, is not hindered even in a highly condensed two-dimensional state at the air–water interface, because the bulky cholesteryl moiety reserves free volume for rotor rotation (Fig. 13B).⁸⁴ However, the collapse of the two-dimensional ordered phase into a three-dimensional disordered phase results in the loss of rotational free volume, accompanied with fluorescence emission. These examples indicate that the nanoarchitectonics of ordered two-dimensional structures would provide many opportunities in the motional regulation of molecular machines.

Another specific feature of the air–water interface is a drastic change in dielectric nature across the interface. The control of molecular motion across the air–water interface can induce drastic changes in physical properties, with dependence on dielectric environments. One example was recently demonstrated as a novel concept called submarine emission (Fig. 14).⁸⁵ In the submarine emission process, the floating-up motion of submarine molecules from the water phase into the air phase across a sub-nanometer level interfacial region induces a drastic increase of fluorescence emission. The submarine molecules employed are double paddle-type platinum dinuclear complexes, consisting of two mononuclear pyrazolato Pt(II)



Fig. 13 Molecular rotor behaviors at an air–water interface: (A) BODIPY rotor; and (B) julolidine rotor.





Fig. 14 Submarine emission with a drastic increase of fluorescence emission through the floating-up motion of submarine molecules from the water phase into the air phase across a sub-nanometer level interfacial region.

complexes doubly linked by alkyl spacers. The H-shaped complexes of the submarine molecules are capable of taking various orientations at the air–water interface. At lower pressures, the submarine molecules are submerged in water, where they exhibit a weak emission. The submarine molecules are forced to float upward toward the air phase by mechanical compression of their monolayer. The weak interaction between the complex planes is broken so that one plane is exposed to the air phase. The complex plane in the air phase is free from dispersion of excitation energy due to molecular contacts under water and thus shows strong emission. Prior to mechanical compression of the monolayer of the submarine molecules, their phosphorescence is quenched but subsequently increases upon mechanical compression of the monolayer. Manipulation of the molecules at interfacial media with different dielectric constants would be an effective way to control their medium-induced photonic properties.

Molecular machine functions in living cells and model membranes

Among various research efforts into molecular machines and related functional molecules, their usage in biological systems would be the most attractive. In particular, cell membranes provide many opportunities for molecular machines to express their potential capabilities.

Membrane transport

One of the major roles of cell membranes is the regulation of chemical potential gradients between the inside and outside of the cell. The control of material transport by artificial molecules

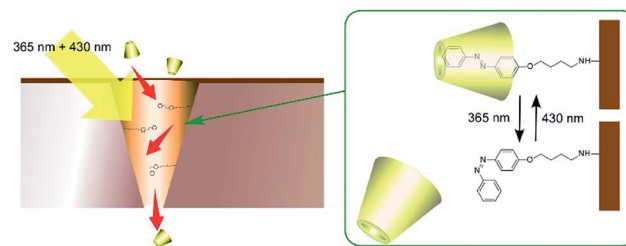


Fig. 15 Photo-driven molecular motors for mass transport across model polymer membranes.

such as molecular carriers and channels has been researched with much interest.

Molecular machines can be used for the regulation of material transport across the model membranes. Wen and co-workers demonstrated photo-driven molecular motors for mass transport across model polymer membranes (Fig. 15).⁸⁶ On a polyimide membrane, conical-shape nanochannels were first fabricated with ion-track etching. The inside surfaces of the nanochannels were then modified with photo-sensitive azobenzene-type molecular motors. The modified nanochannels were found to be capable of autonomous transport of selective guest molecules through simultaneous irradiation with UV and visible light. In the case of β -cyclodextrin transport, β -cyclodextrin is trapped by the *trans*-form of azobenzene upon visible-light irradiation and the trapped β -cyclodextrin is released by UV light irradiation. Interestingly, balanced



Fig. 16 Molecular swings, for ion transport across model lipid bilayer membranes.



intensity between UV and visible light can facilitate mass transport better than strong intensity of a single wavelength.

Zeng and co-workers developed a new type of molecular machine, a molecular swing, for ion transport across model lipid bilayer membranes (Fig. 16).⁸⁷ The molecular swing has a 15-crown-5 part as a carrier unit with ion binding and transport capabilities, together with a lag bolt component to anchor the swing into the hydrophilic regions of the membrane. Even in the cholesterol-rich conditions of lipid membranes, high ion transport behavior was observed. The ion transport mechanism is based on numerous pathways, making the flux for transmembrane K^+ transport highly efficient, corresponding to 27% better than that for gramicidin A. Anticancer activity can even be achieved by further structure optimization.

As reported by Zhang, Long, and co-workers, the motion of a DNA molecular train with azobenzene gears within aerolysin pores was evaluated with high temporal and spatial resolution through electrical signals.⁸⁸ A cytolytic pore-forming toxin known as aerolysin is embedded in lipid membranes. Photoisomerization of azobenzene attached to the molecular train can modulate the speed of the train in the aerolysin pore. Based on such motional control of molecular machines in biological pores, single-molecule computing may become possible upon logic operation with selected inputs of multiple external stimuli.

Drilling cell membrane

Rotational motion of well-designed molecular motors can induce hole opening in cell membranes through a drilling effect, as demonstrated by García-López *et al.* (Fig. 17A).⁸⁹ Molecular motors are adsorbed on the membranes of living cells, and conformational changes of the molecular motors upon application of external stimuli such as UV light irradiation induce disturbances that open holes in the cell membranes. Diffusion of only a trace amount of molecular motors can lead to necrosis and diffusion of outer chemicals into live cells. By introducing short peptide segments to the arms of the molecular motor, the mechanical actions of the molecular motor selectively occurred at target cells with corresponding recognition sites. Because the size of the molecular motors is at the 1 nm level, ruptures of thicker cell membranes by

nanomechanical effects are not always immediate. Concerted motions of 1 nm size motors can dislocate membrane components significantly.

Orozco *et al.* developed more useful systems. Their molecular motor can be activated with much safer visible light instead of UV light (Fig. 17B).⁹⁰ Pancreatic cancer cells are actually killed by the newly synthesized molecular motors with visible light irradiation (405 nm light). The proposed system could be applied to skin cancer cells because the affected areas can be easily exposed to visible light. Unlike healthy cells, cancer cells lack a protective stratum corneum layer. Therefore, artificial molecular motors are expected to adsorb selectively on cancer cells.

Coupling with bio-machines

In addition to introducing artificial molecular machines into biological systems, the introduction of biomolecular machines into artificial supramolecular systems has become a hot topic. In natural biological systems, there are many kinds of highly complex biomolecular machines operating at biological interfacial media, such as cell membranes. Therefore, nano-architecting biomolecular machines into supramolecular films is a rational approach to generating bio-mimetic functional systems. For example, Li and co-workers successfully immobilized a motor protein together with a photoacid generator and proton conductor in a layer-by-layer structural motif (Fig. 18).⁹¹ On a quartz slide, a multilayer layer-by-layer film of poly(allylamine hydrochloride) and graphene oxide is first assembled, into which 1-hydroxy pyrene molecules as photoacid generators are loaded through π - π interactions with graphene oxide. Nafion is next coated as a proton conductor on the surface of the layer-by-layer film, and then proteoliposomes containing the motor protein, ATP synthase, are spread on top. The nano-architected artificial membrane with biomolecular motors can work as photo-activated ATP producer. Photo-irradiation deprotonates 1-hydroxy pyrene to release protons that can transmit through the membrane *via* the proton conductor. The

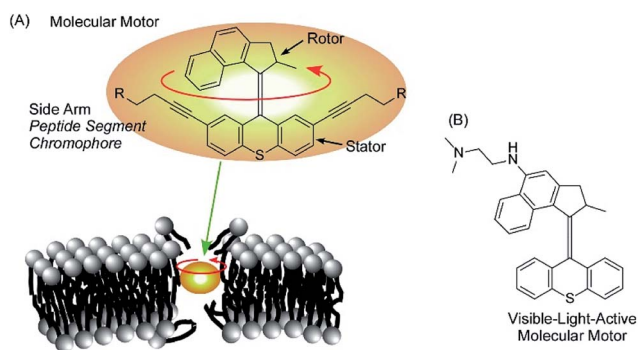


Fig. 17 The hole opening of cell membranes through the drilling effect of molecular motors (A) and a visible-light-active version of a motor (B).



Fig. 18 Layer-by-layer structures with a motor protein together with a photoacid generator and proton conductor for photo-driven ATP synthesis. Reproduced with permission ref. 91. Copyright 2017, American Chemical Society.



Notes and references

- D. Guo, R. Shibuya, C. Akiba, S. Saji, T. Kondo and J. Nakamura, *Science*, 2016, **351**, 361–365.
- K. Maeda and T. E. Mallouk, *Bull. Chem. Soc. Jpn.*, 2019, **92**, 38–54.
- J. Zhang, Q. Zhou, Y. Tang, L. Zhang and Y. Li, *Chem. Sci.*, 2019, **10**, 8924–8929.
- N. Roy, N. Suzuki, C. Terashima and A. Fujishima, *Bull. Chem. Soc. Jpn.*, 2019, **92**, 178–192.
- J. Li, X. Wang, G. Zhao, C. Chen, Z. Chai, A. Alsaedi, T. Hayat and X. Wang, *Chem. Soc. Rev.*, 2018, **47**, 2322–2356.
- L. Liu, W. Li, W. Song and M. Guo, *Sci. Total Environ.*, 2018, **633**, 206.
- H. Kobayashi and A. Fukuoka, *Bull. Chem. Soc. Jpn.*, 2018, **91**, 29–43.
- R. Kas, K. Yang, D. Bohra, R. Kortlever, T. Burdyny and W. A. Smith, *Chem. Sci.*, 2020, **11**, 1738–1749.
- F. Ding, Y. Zhan, X. Lu and Y. Sun, *Chem. Sci.*, 2018, **9**, 4370–4380.
- E. Kim and H. Koo, *Chem. Sci.*, 2019, **34**, 7835–7851.
- J. Kobayashi and T. Okano, *Bull. Chem. Soc. Jpn.*, 2019, **92**, 817–824.
- H. Chen, J. Guo, C. Wang, F. Luo, X. Yu, W. Zhang, J. Li, D. Zhao, D. Xu, Q. Gong, J. Liao, H. Yang, W. Hou and Y. Zhang, *Lancet*, 2020, **395**, 809–815.
- Y. Dong, T. Qiao, D. Kim, D. Parobek, D. Rossi and D. H. Son, *Nano Lett.*, 2018, **18**, 3716–3722.
- Z. Hu, B. Shao, G. T. Geberth and D. A. V. Bout, *Chem. Sci.*, 2018, **9**, 1101–1111.
- A. Azhar, Y. Li, Z. Cai, M. B. Zakaria, M. K. Masud, M. S. A. Hossain, J. Kim, W. Zhang, J. Na, Y. Yamauchi and M. Hu, *Bull. Chem. Soc. Jpn.*, 2019, **92**, 875–904.
- B. Roy and T. Govindaraju, *Bull. Chem. Soc. Jpn.*, 2019, **92**, 1883–1901.
- X. Wei, X. Su, P. Cao, X. Liu, W. Chang, M. Li, X. Zhang and Z. Liu, *Nature*, 2016, **534**, 69–74.
- V. Cherezov, D. M. Rosenbaum, M. A. Hanson, S. G. F. Rasmussen, F. S. Thian and T. S. Kobilk, *Science*, 2007, **318**, 1258–1265.
- J.-P. Sauvage, *Angew. Chem., Int. Ed.*, 2017, **56**, 11080–11093.
- J. F. Stoddart, *Angew. Chem., Int. Ed.*, 2017, **56**, 11094–11125.
- B. L. Feringa, *Angew. Chem., Int. Ed.*, 2017, **56**, 11060–11078.
- A. Harada, *Acc. Chem. Res.*, 2001, **34**, 456–464.
- K. Kinbara and T. Aida, *Chem. Rev.*, 2005, **105**, 1377–1400.
- W. R. Browne and B. L. Feringa, *Nat. Nanotechnol.*, 2006, **1**, 25–35.
- S. Saha and J. F. Stoddart, *Chem. Soc. Rev.*, 2007, **36**, 77–92.
- S. Erbas-Cakmak, D. A. Leigh, C. T. McTernan and A. L. Nussbaumer, *Chem. Rev.*, 2015, **115**, 10081–10206.
- S. Kassem, T. van Leeuwen, A. S. Lubbe, M. R. Wilson, B. L. Feringa and D. A. Leigh, *Chem. Soc. Rev.*, 2017, **46**, 2592–2621.
- T. Takata, *Bull. Chem. Soc. Jpn.*, 2019, **92**, 409–426.
- A. Colin-Molina, M. J. Jellen, E. García-Quezada, M. E. Cifuentes-Quintal, F. Murillo, J. Barroso, S. Perez-Estrada, R. A. Toscano, G. Merino and B. Rodríguez-Molina, *Chem. Sci.*, 2019, **10**, 4422–4429.
- H. V. Schroeder and C. A. Schalley, *Chem. Sci.*, 2019, **10**, 9626–9639.
- X. Liang, L. Li, J. Tang, M. Komiyama and K. Ariga, *Bull. Chem. Soc. Jpn.*, 2020, **93**, 581–603.
- L. Gross, B. Schuler, N. Pavliček, S. Fatayer, Z. Majzik, N. Moll, D. Peña and G. Meyer, *Angew. Chem., Int. Ed.*, 2018, **57**, 3888–3908.
- A. Mukherjee, A. Sanz-Matias, G. Velpula, D. Waghray, O. Ivasenko, N. Bilbao, J. N. Harvey, K. S. Mali and S. De Feyter, *Chem. Sci.*, 2019, **10**, 3881–3891.
- K. Ariga, Q. Ji, W. Nakanishi, J. P. Hill and M. Aono, *Mater. Horiz.*, 2015, **2**, 406–413.
- K. Ariga, K. Minami, M. Ebara and J. Nakanishi, *Polym. J.*, 2016, **48**, 371–389.
- M. Aono and K. Ariga, *Adv. Mater.*, 2016, **28**, 989–992.
- K. Ariga, Q. Ji, J. P. Hill, Y. Bando and M. Aono, *NPG Asia Mater.*, 2012, **4**, e17.
- K. Ariga, M. Nishikawa, T. Mori, J. Takeya, L. K. Shrestha and J. P. Hill, *Sci. Technol. Adv. Mater.*, 2019, **20**, 51–95.
- K. Ariga, J. Li, J. Fei, Q. Ji and J. P. Hill, *Adv. Mater.*, 2016, **28**, 1251–1286.
- K. Ariga and Y. Yamauchi, *Chem.-Asian J.*, 2020, **15**, 718–728.
- K. Ariga, X. Jia, J. Song, J. P. Hill, D. T. Leong, Y. Jia and J. Li, *Angew. Chem., Int. Ed.*, DOI: 10.1002/anie.202000802.
- K. Ariga, S. Watanabe, T. Mori and J. Takeya, *NPG Asia Mater.*, 2018, **10**, 90–106.
- K. Ariga, M. Ito, T. Mori, S. Watanabe and J. Takeya, *Nano Today*, 2019, **28**, 100762.
- K. Ariga, X. Jia, J. Song, C.-T. Hsieh and S.-h. Hsu, *ChemNanoMat*, 2019, **5**, 692–702.
- C. O. Dietrich-Buchecker and J. P. Sauvage, *Chem. Rev.*, 1987, **87**, 795–810.
- A. Livoreil, J.-P. Sauvage, N. Armaroli, V. Balzani, L. Flamigni and B. Ventura, *J. Am. Chem. Soc.*, 1997, **119**, 12114–12124.
- R. A. Bissell, E. Córdova, A. E. Kaifer and J. F. Stoddart, *Nature*, 1994, **369**, 133–137.
- N. Koumura, R. W. J. Zijlstra, R. A. van Delden, N. Harada and B. L. Feringa, *Nature*, 1999, **401**, 152–155.
- J.-J. Yu, L.-Y. Zhao, Z.-T. Shi, Q. Zhang, G. London, W.-J. Liang, C. Gao, M.-M. Li, X.-M. Cao, H. Tian, B. L. Feringa and D.-H. Qu, *J. Org. Chem.*, 2019, **84**, 5790–5802.
- M. Dumartin, M. C. Lipke and J. F. Stoddart, *J. Am. Chem. Soc.*, 2019, **141**, 18308–18317.
- Y. Shirai, A. J. Osgood, Y. Zhao, Y. Yao, L. Saudan, H. Yang, C. Yu-Hung, L. B. Alemany, T. Sasaki, J.-F. Morin, J. M. Guerrero, K. F. Kelly and J. M. Tour, *J. Am. Chem. Soc.*, 2006, **128**, 4854–4864.
- P. Mishra, J. P. Hill, S. Vijayaraghavan, W. Van Rossam, S. Yoshizawa, M. Grisolia, J. Echeverria, T. Ohno, K. Ariga, T. Nakayama, C. Joachim and T. Uchihashi, *Nano Lett.*, 2015, **15**, 4793–4798.
- G. Vives and J. M. Tour, *Acc. Chem. Res.*, 2009, **42**, 473–487.
- K. Ariga, T. Mori and W. Nakanishi, *Chem.-Asian J.*, 2018, **13**, 1266–1278.



- 55 T. A. Jung, R. R. Schlittler, J. K. Gimzewski, H. Tang and C. Joachim, *Science*, 1996, **271**, 181–184.
- 56 H. Tang, M. T. Cuberes, C. Joachim and J. K. Gimzewski, *Surf. Sci.*, 1997, **386**, 115–123.
- 57 Y. Shirai, J. F. Morin, T. Sasaki, J. M. Guerrero and J. M. Tour, *Chem. Soc. Rev.*, 2006, **35**, 1043–1055.
- 58 D. Castelvetti, *Nature*, 2017, **544**, 278–279.
- 59 G. Rapenne and C. Joachim, *Nat. Rev. Mater.*, 2017, **2**, 17040.
- 60 D. Abbasi-Pérez, H. Sang, L. Pérez-García, A. Floris, D. B. Amabilino, R. Raval, J. Manuel Recio and L. Kantorovich, *Chem. Sci.*, 2019, **10**, 5864–5874.
- 61 T. Seki, *Bull. Chem. Soc. Jpn.*, 2018, **91**, 1026–1057.
- 62 Y. Liu, A. H. Flood, P. A. Bonvallet, S. A. Vignon, B. H. Northrop, H.-R. Tseng, J. O. Jeppesen, T. J. Huang, B. Brough, M. Baller, S. Magonov, S. D. Solares, W. A. Goddard, C.-M. Ho and J. F. Stoddart, *J. Am. Chem. Soc.*, 2005, **127**, 9745–9759.
- 63 W.-H. Soe, S. Srivastava and C. Joachim, *J. Phys. Chem. Lett.*, 2019, **10**, 6462–6467.
- 64 K. Ariga and T. Kunitake, *Acc. Chem. Res.*, 1998, **31**, 371–378.
- 65 K. Ariga, H. Ito, J. P. Hill and H. Tsukube, *Chem. Soc. Rev.*, 2012, **41**, 5800–5835.
- 66 K. Ariga, T. Mori and J. P. Hill, *Adv. Mater.*, 2012, **24**, 158–176.
- 67 K. Ariga, Y. Yamauchi, T. Mori and J. P. Hill, *Adv. Mater.*, 2013, **25**, 6477–6512.
- 68 K. Ariga, M. Ishii and T. Mori, *Chem.–Eur. J.*, 2020, **26**, 6461–6472.
- 69 K. Ariga, Y. Terasaka, D. Sakai, H. Tsuji and J. Kikuchi, *J. Am. Chem. Soc.*, 2000, **122**, 7835–7836.
- 70 K. Ariga, T. Nakanishi, Y. Terasaka, H. Tsuji, D. Sakai and J. Kikuchi, *Langmuir*, 2005, **21**, 976–981.
- 71 T. Mori, K. Okamoto, H. Endo, J. P. Hill, S. Shinoda, M. Matsukura, H. Tsukube, Y. Suzuki, Y. Kanekiyo and K. Ariga, *J. Am. Chem. Soc.*, 2010, **132**, 12868–12870.
- 72 T. Michinobu, S. Shinoda, T. Nakanishi, J. P. Hill, K. Fujii, T. N. Player, H. Tsukube and K. Ariga, *J. Am. Chem. Soc.*, 2006, **128**, 14478–14479.
- 73 J. M. Lehn, *Angew. Chem., Int. Ed. Engl.*, 1988, **27**, 89–112.
- 74 D. J. Cram, *Angew. Chem., Int. Ed. Engl.*, 1988, **27**, 1009–1020.
- 75 C. J. Pedersen, *Angew. Chem., Int. Ed. Engl.*, 1988, **27**, 1021–1027.
- 76 S. Shinkai, T. Ogawa, T. Nakaji, Y. Kusano and O. Nanabe, *Tetrahedron Lett.*, 1979, **20**, 4569–4572.
- 77 S. Shinkai, *Proc. Jpn. Acad., Ser. B*, 2019, **95**, 590–601.
- 78 K. Ariga, T. Mori and J. Li, *Langmuir*, 2019, **35**, 3585–3599.
- 79 K. Ariga, *ChemNanoMat*, 2020, **6**, 870–880.
- 80 D. Ishikawa, T. Mori, Y. Yonamine, W. Nakanishi, D. Cheung, J. P. Hill and K. Ariga, *Angew. Chem., Int. Ed.*, 2015, **54**, 8988–8991.
- 81 T. Mori, D. Ishikawa, Y. Yonamine, Y. Fujii, J. P. Hill, I. Ichinose, K. Ariga and W. Nakanishi, *ChemPhysChem*, 2017, **18**, 1470–1474.
- 82 J. Cheng, P. Štacko, P. Rudolf, R. Y. N. Gengler and B. L. Feringa, *Angew. Chem., Int. Ed.*, 2017, **56**, 291–296.
- 83 T. Mori, H. Chin, K. Kawashima, H. Ngo, M.-J. Cho, W. Nakanishi, J. P. Hill and K. Ariga, *ACS Nano*, 2019, **13**, 2410–2419.
- 84 T. Mori, H. Komatsu, N. Sakamoto, K. Suzuki, J. P. Hill, M. Matsumoto, H. Sakai, K. Ariga and W. Nakanishi, *Phys. Chem. Chem. Phys.*, 2018, **20**, 3073–3078.
- 85 J. Adachi, T. Mori, R. Inoue, M. Naito, N. H.-T. Le, S. Kawamorita, J. P. Hill, T. Naota and K. Ariga, *Chem.–Asian J.*, 2020, **15**, 406–414.
- 86 G. Xie, P. Li, Z. Zhao, X.-Y. Kong, Z. Zhang, K. Xiao, H. Wang, L. Wen and L. Jiang, *Angew. Chem., Int. Ed.*, 2018, **57**, 16708–16712.
- 87 C. Ren, F. Chen, R. Ye, Y. S. Ong, H. Lu, S. S. Lee, J. Y. Ying and H. Zeng, *Angew. Chem., Int. Ed.*, 2019, **58**, 8034–8038.
- 88 Y.-L. Ying, Z.-Y. Li, Z.-L. Hu, J. Zhang, F.-N. Meng, C. Cao, Y.-T. Long and H. Tian, *Chem*, 2018, **4**, 1893–1901.
- 89 V. García-López, F. Chen, L. G. Nilewski, G. Duret, A. Aliyan, A. B. Kolomeisky, J. T. Robinson, G. Wang, R. Pal and J. M. Tour, *Nature*, 2017, **548**, 567–572.
- 90 C. A. Orozco, D. Liu, Y. Li, L. B. Alemany, R. Pal, S. Krishnan and J. M. Tour, *ACS Appl. Mater. Interfaces*, 2020, **12**, 410–417.
- 91 Y. Xu, J. Fei, G. Li, T. Yuan and J. Li, *ACS Nano*, 2017, **11**, 10175–10183.
- 92 G. Li, J. Fei, Y. Xu, B. Sun and J. Li, *Angew. Chem., Int. Ed.*, 2019, **58**, 1110–1114.
- 93 Y. Zhang, H. Kersell, R. Stefak, J. Echeverria, V. Iancu, U. G. E. Perera, Y. Li, A. Deshpande, K.-F. Braun, C. Joachim, G. Rapenne and S.-W. Hla, *Nat. Nanotechnol.*, 2016, **11**, 706–713.
- 94 Y. Zhang, J. P. Calupitan, T. Rojas, R. Tumbleson, G. Erbland, C. Kammerer, T. M. Ajayi, S. Wang, L. A. Curtiss, A. T. Ngo, S. E. Ulloa, G. Rapenne and S. W. Hla, *Nat. Commun.*, 2019, **10**, 3742.
- 95 M. Clemente-León, A. Credi, M.-V. Martínez-Díaz, C. Mingotaud and J. F. Stoddart, *Adv. Mater.*, 2006, **18**, 1291–1296.

