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Synthetically tuneable biomimetic artificial photosynthetic reaction centres that closely resemble the natural system in purple bacteria†

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Porphyrin-based photosynthetic reaction centre (PRC) mimics, **ZnPQ-Q2HP-C$_{60}$** and **MP$_2$Q-Q2HP-C$_{60}$** (M = Zn or 2H), designed to have a similar special-pair electron donor and similar charge-separation distances, redox processes and photochemical reaction rates to those in the natural PRC from purple bacteria, have been synthesised and extensive photochemical studies performed. Mechanisms of electron-transfer reactions are fully investigated using femtosecond and nanosecond transient absorption spectroscopy. In benzonitrile, all models show picosecond-timescale charge-separations and the final singlet charge-separations with the microsecond-timescale. The established lifetimes are long compared to other processes in organic solar cells or other organic light harvesting systems. These rigid, synthetically flexible molecules provide the closest mimics to the natural PRC so far synthesised and present a future direction for the design of light harvesters with controllable absorption, redox, and kinetics properties.

1. Introduction

Long-lived photochemical charge separation provides means of harvesting solar energy for subsequent use in *e.g.*, photovoltaic or photosynthetic devices,[1–5] and many different types of systems have been investigated in the last two years.[6–32] Other possible applications include as photosensitizers for cancer treatment,[33–40] modulators of DNA,[41] neuron controllers,[42,43] biosensors,[44,45] logic switches,[46] and injecting charges into nanomaterials.[47] A long-standing objective has been the mimicking of biological photosynthetic reaction centres (PRCs)[48,49] in order to design efficient charge-separation units.[50–57] In the PRC from purple bacteria, the solar to electrical energy conversion process starts with a multistep photoinduced electron transfer sequence between a special-pair bacterio-chlorophyll dimer (P) and ubiquinone (Q$_B$), and the mechanism has been thoroughly studied over the past few decades[48,49] (for recent mini-reviews see *e.g.* ref. 58–62). Knowledge about the photoinduced-electron transfer mechanism has led to the development of numerous artificial photosynthetic systems that show the possibility for photovoltaic and photonic applications.[57,63–65] Indeed, the natural systems are often taken as model devices to be mimicked in artificial solar-energy generation and solar-photochemical processes.[1,57,66–71]

The purple bacteria *Rhodopseudomonas sphaeroides*[72,73] and *Rhodopseudomonas viridis*[74,75] are known to have a high degree of similarity in the structures of their PRCs. Similar arrangements are also found in the more complex photosystems I and II of the cyanobacterium *Synechococcus elongatus*,[76,77] implying that at least some features of the arrangement of the porphinoid pigments are generally useful in biological photosynthesis. Organic solar-energy capture systems usually mimic these natural systems in a variety of ways, including aspects of exciton transfer, primary charge separation, and secondary charge separation. Here we explore what happens when artificial systems are designed to mimic in considerable detail the chemical, structural and kinetics properties of natural photosystems[48,49] (for recent mini-reviews see *e.g.* ref. 58–62), choosing the known structure of PRCs from purple bacteria, as illustrated in Fig. 1. The mimics have improved properties for use in device applications, however, instead of bacteriochlorophylls and quinones, we use the related compounds, porphyrins and fullerenes, as these are chemically much more stable. Also we use rigid linkers to control structure rather than an external

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Fig. 1 (Left) Arrangement of the chromophores in the PRC from *Rhodopseudomonas viridis*, with the inter-chromophoric distances and process rates shown. (Right) The structure of **(MP)₂Q-Q2HP-C₆₀** (synthesised in this work) and the inter-chromophoric distances and analogous rates shown. P = 'special pair', B_L = auxiliary bacterio-chlorophyll, H_L = primary-acceptor bacteriopheophytin, and Q_A = secondary-acceptor menaquinone; M = Zn(II) or 2H, Ar = 3,5-di-*tert*-butylphenyl.

protein environment, maintaining structural integrity to affect similar degrees of charge separation in each step but done in a more easily controllable way. An interesting functional similarity is that porphyrin–fullerene molecules have been shown to support long-lived (1 ms at room temperature) spin-polarized long-distance charge-separated states,[78] somewhat akin to natural PRCs.[79–82] Such species are also particularly relevant in modern device contexts[55,83] but could lead to photochemical damage and so chemical means to control them in artificial systems are required.

In search of biomimetic charge-separating units, previously we designed and investigated the photophysical properties of the tris- and tetrakis-porphyrin arrays, **ZnPQ-Q2HPQ-QAuP⁺** and **ZnPQ-Q2HPQ-Q2HPQ-QAuP⁺**,[84–87] that to some degree also resemble the arrangement of the natural PRCs. Indeed, these compounds contain the quinoxaline Tröger's base linkage that establishes the C_2 symmetry in the 'special pair' (see Fig. 1), and the biquinoxalinyl linkage provides a very similar charge-separation distance to that between the 'special pair' and the primary electron acceptor. Both arrays show multistep positive charge shift from the gold(III) porphyrin to the zinc(II) porphyrin at the other end of the array after photoexcitation, and the final charge-shift states, namely **ZnPQ•⁺-Q2HPQ-QAuP** and **ZnPQ•⁺-Q2HPQ-Q2HPQ-QAuP**, are long-lived in benzonitrile. Besides, the addition of the chloride ligand has been shown to control

the gradient of energy and charge-shift processes,[86,87] and is able to afford the long-lived final charge-shift state in a non-polar solvent (*e.g.*, toluene), which is close to the dielectric constant of the natural PRC. This showed the importance of controlling the redox gradient in achieving long-lived charge-separation, a critical feature in any organic artificial photovoltaic device.

However, these model compounds were also found to have several deficiencies, including: (i) the quinoxaline Tröger's base linkage separates the 'special pair' porphyrins too far apart for an efficient cation radical delocalization, and (ii) the system lacks an electron acceptor to mimic the quinones and iron–sulfur complexes of the natural systems. Here we introduce the modified photosynthetic mimics **(MP)₂Q-Q2HP-C₆₀** (where M = Zn or 2H) shown in Fig. 1 to remove these differences. The inclusion of a 'special pair' is not essential in an artificial device and most compounds in current use are more like our previous designs. However, in biology the function of the special pair is readily controlled by protein mutation effects,[88] and the inclusion of a similar feature in artificial compounds can be subsequently explored using established synthetic strategies to allow similar control in an artificial environment.

In **(MP)₂Q-Q2HP-C₆₀**, the 'special pair' porphyrins "**(MP)₂**" are connected directly through a Tröger's base bridge on the porphyrin β-pyrrolic position without the use of the quinoxalinyl groups that linked the chromophores in **ZnPQ-Q2HPQ-QAuP⁺** and in **ZnPQ-Q2HPQ-Q2HPQ-QAuP⁺**.[84–87] The **(MP)₂** unit is closely modelled by the Tröger's base linked porphyrin dimer **(ZnP)₂**, and for this molecule an X-ray crystal structure confirms the pseudo C_2 symmetry of the porphyrin dimer that mimics the "special-pair" of natural PRCs.[89,90] Significantly, the intra-dimer porphyrin-centre to porphyrin-centre distance is reduced from 16.8 Å[84,85] for the quinoxalinyl-bridged mimics to 6.2 Å for this Tröger's base dimer, close to the inter-chromophore separation of 7.0 Å[75] found within the special pair in the natural PRC. Also, the new compound **(MP)₂Q-Q2HP-C₆₀** includes additional functionality through the bonding of the fullerene electron acceptor so as to permit secondary charge-separation processes to occur. This group mimics the electron acceptor function of the quinones in the natural system. Numerous studies showed that the use of fullerene as an electron acceptor significantly reduced the reorganization energy for electron transfer due to its symmetrical shape, large size, and delocalized π-electron system,[91–93] a change from the natural system likely to bring benefit in device applications.

As Fig. 1 shows, the critical aspects of these biomimetic compounds is that they separate charges over similar distances to the natural system, keep them separated for similar times, are readily controllable, and could show similar effects such as long-lived charge-separated triplet-state production.[78] However, the mechanisms driving exciton transport, charge separation, and charge combination are quite different. In the natural system, through-space interactions mediated dynamically by intervening matter are critical.[48,49,58–62,94] Such effects are difficult to control in artificial systems and so are replaced with

through-bond coupling mechanisms in rigid molecules,[78,84,95–103] and a critical design criteria is therefore to establish similar couplings using this device-robust mechanism. A great advantage of the chemical routes presented to biomimetic solar-energy harvesting is that it can readily be modified to make broadband or narrowband absorbers, for example, for use in solar cells or phototherapy, respectively,[40,96,98,104] for upconversion units with unprecedented efficiency,[105–107] and as tailor-made components for determining mechanism and improving solar cell functions.[108]

We report the details of the synthesis of the new photosynthetic mimic **(MP)₂Q-Q2HP-C₆₀**, along with model systems **ZnPQ-Q2HP-C₆₀**, **Q2HP-C₆₀**, **Q2HP-Ph**, **(ZnP)₂**, **(2HP)₂**, **ZnPQ**, **2HPQ**, **ZnP** and **2HP** (Chart 1). The electrochemical and photophysical properties of these systems are also presented in detail, and its performance is compared to that of natural PRCs.

2. Experimental section

2.1 General

Ultraviolet-visible spectra were recorded on a Cary 5E UV-Vis spectrophotometer in stated solvents. Steady-state fluorescence spectra were recorded on a Varian Cary Eclipse

Chart 1 Model compounds studied in this work.

fluorescence spectrophotometer in deacidified benzonitrile. ^{1}H NMR spectra were recorded on a Bruker AVANCE300 (300 MHz) spectrometer or Bruker DPX400 (400 MHz) spectrometer. Samples were dissolved in deacidified CDCl₃. MALDI-TOF mass spectrometry was recorded on a Waters (Micromass) TOF SPEC 2E mass spectrometer. Infrared spectra were recorded on a Shimadzu Model 8400 FT-IR as solutions in deacidified CHCl₃.

2.2 Electrochemical measurements

The cyclic voltammetry measurements were performed on a CHI 900B electrochemical analyser in deaerated PhCN containing 0.10 M n-Bu₄NPF₆ as supporting electrolyte at 298 K. A gold working electrode was polished with BAS polishing alumina suspension and rinsed with acetone before use. A platinum wire was used as a counter electrode and a silver wire was used as a reference electrode. All experiments were followed by the addition of ferrocene with the ferrocene/ferrocenium couple used as an internal reference.

2.3 Spectroelectrochemical measurements

UV-Vis spectroelectrochemical measurements were performed with a thin-layer quartz spectroelectrochemical cell and a platinum gauze working electrode. Potentials were applied and monitored using a CHI 900B electrochemical analyser. UV-Vis spectra were recorded on a Cary 4E UV-Vis spectrophotometer.

2.4 Spectral measurements and fitting

Femtosecond transient absorption spectroscopy experiments were conducted at Osaka University, Japan, using an ultrafast source: Integra-C (Quantronix Corp.), an optical parametric amplifier: TOPAS (Light Conversion Ltd.) and a commercially available optical detection system: Helios provided by Ultrafast Systems LLC. The source for the pump and probe pulses were derived from the fundamental output of Integra-C (780 nm, 2 mJ per pulse and fwhm = 130 fs) at a repetition rate of 1 kHz. As much as 75% of the fundamental output of the laser was introduced into TOPAS which has optical frequency mixers, resulting in a tuneable range from 285 to 1660 nm, while the rest of the output was used for white light generation. Prior to generating the probe continuum, a variable neutral density filter was inserted into the path in order to generate a stable continuum, the laser pulse was then fed to a delay line that provides an experimental time window of 3.2 ns with a maximum step resolution of 7 fs. A wavelength between 350 and 450 nm of the TOPAS output, which are the fourth harmonic of the signal or idler pulses, was chosen as the pump beam. As this TOPAS output consists of not only the desirable wavelength but also unnecessary wavelengths, the latter were deviated using a wedge prism with wedge angle of 18°. The desirable beam was irradiated at the sample cell with a spot size of 1 mm diameter where it was merged with the white probe pulse in a close angle (<10°). The probe beam after passing through the 2 mm sample cell was focused on a fibre optic cable that was connected to a CCD spectrograph for recording the time-resolved spectra (410–800 nm). Typically, 2500 excitation
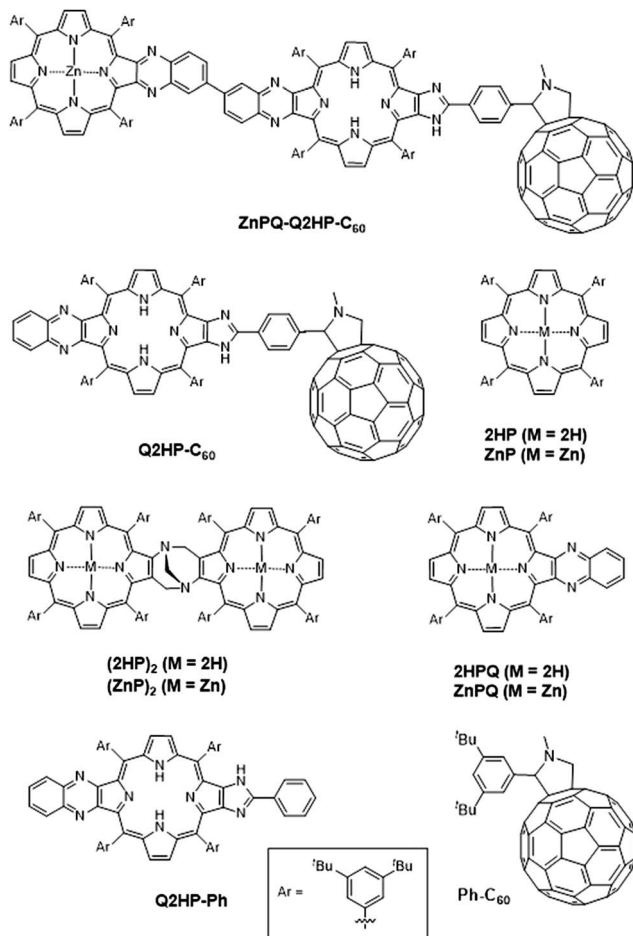
pulses were averaged for 5 s to obtain the transient spectrum at a set delay time. Kinetic traces at appropriate wavelengths were assembled from the time-resolved spectral data. All measurements were conducted at room temperature (RT), 298 K, using porphyrin concentrations of 1.0×10^{-6} M (fs transient spectra and fluorescence spectra).

Spectral components were obtained from the time-dependent spectra by fitting the observed data to kinetics schemes involving the unexcited ground-state reactant R and up to six transient or final photochemical products named in rough order of production P_1 up to P_6. Different rate constants are then ascribed to observed unimolecular reactions amongst these species. The time sequence of observed transient absorption spectra $\Delta A(\nu, t)$ was then fitted to reveal the transient spectrum of each component $\Delta A_i(\nu)$ and the set of rate constants k for each of the included reactions. These rate constants are reported as the isolated-process lifetimes $\tau = 1/k$. The transient absorption ΔA is the difference in sample absorption caused by the excitation pulse at $t = 0$. The fitting procedure involved first smoothing and interpolating the raw experimental data using Gaussian convolution. Prior to fitting, the data was assembled on a linear frequency scale and a logarithmic time scale so as to properly weight the different spectral and temporal regions. The fit was optimized to minimize the residual between the observed and fitted spectra using over-determined linear least-squares analysis to fit the spectral components, combined with Newton–Raphson optimization of the rate constants in an external loop.

A shortcoming of the fitting procedure is that typically there is insufficient data available to determine the properties of low-yield processes and pathways. For example, the absorption of light will excite a mixture of initial products arising from individual absorptions on each chromophore, each of which will react independently. The signal to noise ratio of the observed data, however, typically supports only the identification of the resultant state mix rather than each chemical component of the mix. As a result, the extracted spectra of P_1 and subsequent species each reflect differently weighted sums of individual molecular component spectra.

Nanosecond laser flash photolysis experiments were performed at Osaka University, Japan. Porphyrins in deaerated benzonitrile solutions (3.0×10^{-6} M) were excited by a Panther OPO pumped by a Nd:YAG laser (Continuum, SLII-10, 4–6 ns fwhm) at $\lambda = 355$ nm with a power of 16 mJ per pulse. The photochemical reactions were monitored by continuous exposure to a Xe-lamp (150 W) as a probe light and a photomultiplier tube (Hamamatsu 2949) as a detector. For transient absorption spectra in the near-IR region (800–1200 nm), monitoring light from a pulsed Xe-lamp was detected with a Ge-avalanche photodiode (Hamamatsu Photonics, B2834). All experiments were performed at 298 K.

2.5 Synthesis

The synthesis[109] and characterization of: zinc(II) 12/13-nitro{(formylphenyl)imidazo}porphyrin **2**, 12/13-nitro{(fullerene-N-methylpyrrolidinylphenyl)imidazo}porphyrin **3**, {6'-(3,4-diamino

benzene)quinoxalino}{(fullerene-N-methylpyrrolidinylphenyl)-imidazo}porphyrin **5**, (12/13-nitroquinoxalino)(fullerene-imidazo)quinoxalino}bisporphyrin **7**, (Tröger's base-quinoxalino){(fullereneimidazo)quinoxalino}trisporphyrin **(2HP)$_2$Q-Q2HP-C$_{60}$**, and (di-zinc(II) Tröger's base-quinoxalino){(fullereneimidazo)quinoxalino}trisporphyrin **(ZnP)$_2$Q-Q2HP-C$_{60}$** is described in detail in ESI Section S1.†

3. Results and discussion

3.1 Synthesis

The synthesis of **(MP)$_2$Q-Q2HP-C$_{60}$** is outlined in Scheme 1. The precursor zinc(II) nitroporphyrin-2,3-dione **1** was prepared according to the method published elsewhere.[109] Following the imidazole-forming condensation method developed earlier,[110] zinc(II) nitroporphyrin-dione **1** was reacted with an excess of terephthalaldehyde (5 eq.) in the presence of ammonium acetate in refluxing CHCl$_3$ and gave zinc(II) formyl-imidazo-porphyrin **2** in 90% yield. Remarkably, zinc(II) chelation is a crucial step in this particular reaction as using free-base formyl-imidazoporphyrin **6** gave only 40% yield under the same conditions. The aldehyde functionality provides a convenient means to attach a fullerene by cycloaddition of the azomethine ylide formed upon condensation of N-methylglycine with **2** to give **3**.[111,112] This reaction exhibits an opposite effect in term of metal chelation as the imidazole-forming condensation reaction (step i), where this reaction afforded a higher yield of fullerene appended nitroporphyrin using free-base porphyrin as a precursor (75% for free base porphyrin vs. 37% for zinc(II) porphyrin) under the same conditions. Zinc(II) formyl-imidazoporphyrin **2** was thereafter demetallated by stirring in a mixture of hydrochloric acid and CH$_2$Cl$_2$ before promoting the azomethine–fullerene reaction. Conversion of the nitro into the amino using tin(II) chloride reduction, followed by photooxidation afforded the fullerene appended porphyrin-dione **4** in 54% yield for the two steps. Condensation of **4** with 3,3'-diaminobenzidine (1 eq.) gave the diaminobenzidine-quinoxalinyl porphyrin **5**.[84,85,113] To minimize the formation of 2 : 1 adduct, a solution of **4** was added slowly over 1 h to a dilute solution of 3,3'-diaminobenzidine. Condensation of **5** with **6** afforded the fullerene-appended bisporphyrin **7** in 56% yield. Finally reduction of the nitro group followed by the acid-catalysed condensation of the resultant aminoporphyrin **9** (~2 eq.) and an excess of formaldehyde,[84,85,113,114] with heating at 70 °C under nitrogen for four days gave **(2HP)$_2$Q-Q2HP-C$_{60}$** which was purified by size exclusion chromatography. **(ZnP)$_2$Q-Q2HP-C$_{60}$** was simply prepared by stirring **(2HP)$_2$Q-Q2HP-C$_{60}$** with zinc acetate in CH$_3$OH and CH$_2$Cl$_2$ mixture. Interestingly, zinc metallation occurred preferentially at the 'special pair' porphyrin rings that have porphyrin and 'chlorin-like' quinoxalinoporphyrin aromatic delocalisation pathways and hence more acidic inner protons rather than at the less acidic 'bacteriochlorin-like' quinoxalino-imidazolo-porphyrin that has the appended fullerene. The difference in rate of metallation of chlorin-like and bacteriochlorin-like rings was recognised and exploited in earlier work on the synthesis of extended porphyrins.[115]
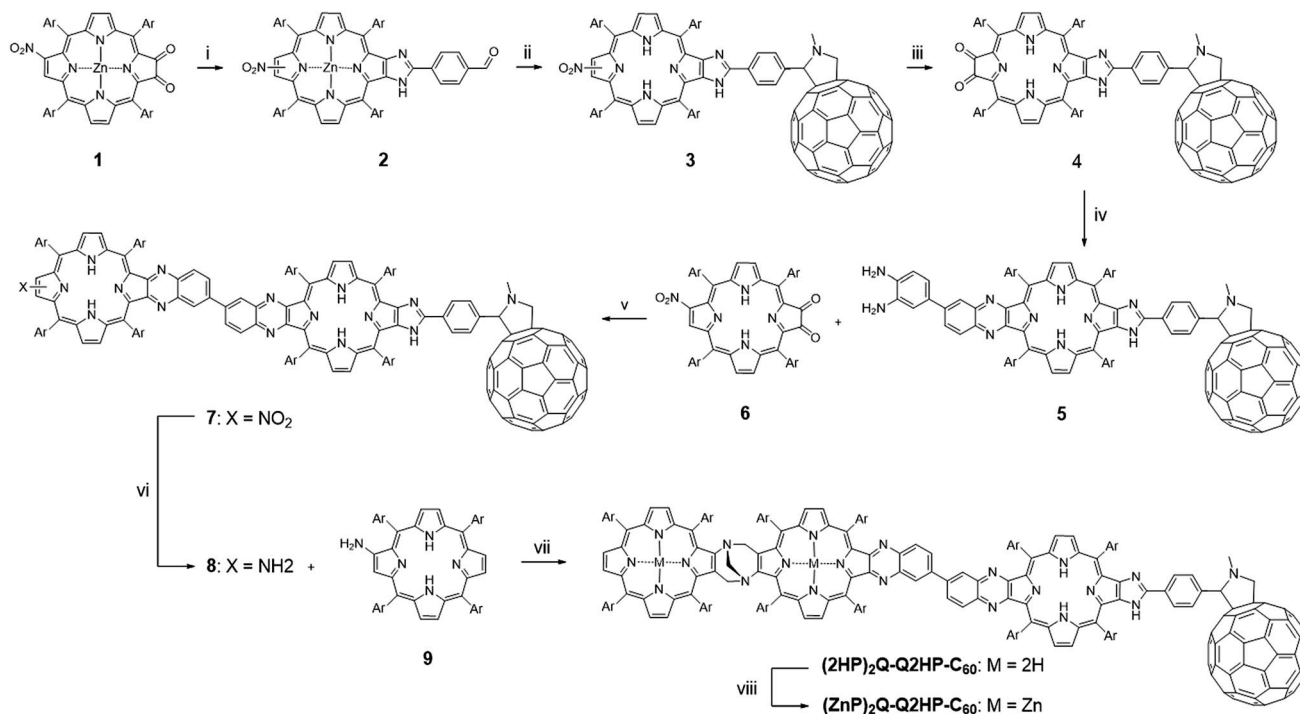
Scheme 1 *Reagents and conditions*: (i) terephthalaldehyde, NH_4OAc, $CHCl_3$, reflux (90%); (ii) **1**, CH_2Cl_2, HCl, stir (89%); **2**, fullerene, *N*-methylglycine, toluene, reflux (75%); (iii) **1**, $SnCl_2 \cdot 2H_2O$, HCl/Et_2O, stir; **2**, CH_2Cl_2, photooxidation (54%); (iv) 3,3′-diaminobenzidine, CH_2Cl_2, stir (80%); (v) CH_2Cl_2, stir (56%); (vi) $SnCl_2 \cdot 2H_2O$, HCl, CH_2Cl_2, stir; (vii) THF, HCl/EtOH, formaldehyde, stir, 70 °C (16%); (viii) $Zn(OAc)_2 \cdot 2H_2O$, $CH_2Cl_2/MeOH$, stir (58%). Ar = 3,5-di-*tert*-butylphenyl.

3.2 Steady-state absorption and emission spectroscopy

To identify the products formed at each stage of the photochemical reactions, detailed information concerning the assignment of the absorption spectra of the photoexcited molecule is required. Spectra for the dyad, triad and tetrad molecules studied, plus analogous spectra for each molecular fragment of these compound systems, are presented in ESI

Section S2 Fig. S1–S5.† This absorption data is therein interpreted by comparison with emission spectra.[53,85,97,116–129] Table 1 lists the key results required for the analyses of the photochemical experiments, including the origin band energies of the lowest transition observed in both absorption and emission, the Stoles shift between them, and the estimated origin energy E_{0-0}, while detailed assignment information is discussed in the text.

Table 1 Energy of the lowest excited-state (E_{0-0}) of **Q2HP-C$_{60}$**, **ZnPQ-Q2HP-C$_{60}$**, **(ZnP)$_2$Q-Q2HP-C$_{60}$** and **(2HP)$_2$Q-Q2HP-C$_{60}$**, and their model compounds **ZnP**, **2HP**, **ZnPQ**, **2HPQ**, **(ZnP)$_2$** and **(2HP)$_2$** in deaerated PhCN

Molecule	Chromophore	Abs.[a] (cm^{-1})	Emiss.[b] (cm^{-1})	Stokes shift[c] (cm^{-1})	E_{0-0}[d] (eV)
ZnP	**ZnP**	16 670	16 470	200	2.05
ZnPQ	**ZnPQ**	16 150	15 700	450	1.97
2HP	**2HP**	15 390	15 290	100	1.90
2HPQ	**2HPQ**	15 370	15 270	100	1.90
(ZnP)$_2$	**(ZnP)$_2$**	16 420	15 820	600	2.00
(2HP)$_2$	**(2HP)$_2$**	14 850	14 300	550	1.81
Q2HP-C$_{60}$	**2HPQ**	15 270	15 170	100	1.89
ZnPQ-Q2HP-C$_{60}$	**ZnPQ**	16 100	15 650[e]	450[e]	1.97
	2HPQ	15 270	15 170	100	1.89
(ZnP)$_2$Q-Q2HP-C$_{60}$	**(ZnP)$_2$Q**	15 650 ± 100	15 700[f] ± 175	[450–600][f]	1.95 ± 0.02
	2HPQ	15 270	15 170	100	1.89
(2HP)$_2$Q-Q2HP-C$_{60}$	**(2HP)$_2$Q**	14 640 ± 150	14 340	300 ± 150	1.80 ± 0.01
	2HPQ	15 220	15 120[g]	100[g]	1.88

[a] The wavenumber of the origin Qx band obtained from absorption spectrum. [b] The wavenumber of the origin Qx band obtained from emission spectrum. [c] Difference between the absorption and emission spectra of the origin Qx band. [d] The energy of the lowest excited-state calculated as the average energy of origin Qx bands in the absorption and the emission spectra. [e] Estimated based on the Stokes shift of **ZnPQ**. [f] Estimated based on the stokes shift of **ZnPQ** and **(ZnP)$_2$**. [g] Estimated based on the stokes shift of **2HPQ**.

Besides these critical quantitative results, the spectra also provide qualitative insight into the chemical interactions present within the compounds. All chromophores interact with each other only weakly so that the absorption spectra of dyad, triad, and tetrad species are simply the sum of those of the isolated chromophores. However, bands do shift considerably, indicating, *e.g.*, the presence of significant π conjugation likely to contribute significantly to the observed charge separation and recombination mechanisms.[78,99]

Alternatively, emission spectra can show significant differences when comparing that from dyads, triads, or tetrads to that from their molecular components, indicative of the photochemical processes that take place. By assigning emission to a particular chromophore, the nature of the lowest energy states are determined (using Kasha's rule),[119] Stokes shifts determined, and components of the much more complex absorption spectra assigned. Asymmetry between absorption and emission indicates that symmetric Troger's base dimers **(ZnP)₂** and **(2HP)₂** adsorb with excitation localized on their individual porphyrins but emit through an exciton-delocalized state. However, in extended Troger's base compounds containing **(ZnP)₂Q** units, the asymmetry prevents delocalization.

The arguments made make extensive use of the expected symmetry between absorption and emission coming from the same chromophore, and the difference between Franck–Condon allowed vibrational progressions and vibronic intensification arising from the Herzberg–Teller principle.[124] A critical feature discussed in ESI† is that the for a Franck–Condon progression the intensity ratio of the origin and side bands in traditional spectra gets distorted by a factor of $[(\Delta E - h\nu)/\Delta E]^6 \sim (14\,000/15\,500)^6 = 0.54$, where ΔE is the origin energy and $h\nu$ the vibrational sideband spacing. This and vibronic effects can dramatically reduce perceived asymmetry, and understanding them proved critical to the recent assignment of the Q-band spectrum of chlorophyll-a, a feature critical to photosynthesis understanding that defied mankind for over 50 years.[130,131]

3.3 Electrochemistry

Cyclic voltammetry was carried out in 0.1 M n-Bu₄NPF₆ solution in benzonitrile and redox potentials of investigated compounds are summarized in Table 2. Cyclic voltammograms of **Q2HP-C₆₀**, **ZnPQ-Q2HP-C₆₀**, **(ZnP)₂Q-Q2HP-C₆₀** and **(2HP)₂Q-Q2HP-C₆₀** are shown in ESI Section S3 Fig. S6,† while those for model systems **ZnP**, **(ZnP)₂**, **2HP**, **(2HP)₂**, **Q2HP-Ph** and **Ph-C₆₀** are given in Fig. S7.† From the observed redox potentials given in Table 2, the free-energy changes associated with all the possible charge-separation and charge-recombination processes for **Q2HP-C₆₀**, **ZnPQ-Q2HP-C₆₀**, **(ZnP)₂Q-Q2HP-C₆₀** and **(2HP)₂Q-Q2HP-C₆₀** can be estimated and are given in Table 3.

As expected, the attachment of a phenyl group to an imidazole on the β-pyrrolic position and the introduction of the quinoxalinyl group extents the conjugation and significantly affects the unoccupied orbitals (Table 2). The P/P·⁻ reduction hence becomes easier by *ca.* 23 mV, comparing **2HP** with **Q2HP-Ph**. The C₆₀/C₆₀·⁻ reduction is harder by *ca.* 28 mV in **Q2HP-C₆₀** compared to that of **Ph-C₆₀**. Such effects are also observed on **ZnPQ-Q2HP-C₆₀**, **(ZnP)₂Q-Q2HP-C₆₀** and **(2HP)₂Q-Q2HP-C₆₀**. More significantly, the Tröger's base porphyrin dimers **(ZnP)₂** and **(2HP)₂** are easier to oxidize by 90 and 140 mV, respectively. A split of 90 mV is observed for the first oxidation of the porphyrins for **(2HP)₂**. This suggests that one of the porphyrin macrocycles is oxidized in advance and yields H₂P·⁺–H₂P, while the oxidation of the second porphyrin macrocycle occurs subsequently to give H₂P·⁺–H₂P·⁺. This potential difference of 90 mV is due the Coulomb repulsion between the two positive charges. A similar result is also observed for **(ZnP)₂** in which the redox signal is broadened and the splitting cannot be resolved.

These estimates include screened coulombic interactions between the zwitterionic regions of the excited-states, expressed in terms of the distance R between the centres of positive and negative charge obtained from the calculated dipole moments at DFT-optimized molecular structures. The energies of the lowest excited-state (E_{0-0}) are determined from the average

Table 2 Standard potentials (*vs.* Fc⁺/Fc, in mV)[a] of Q2HP-C₆₀, ZnPQ-Q2HP-C₆₀, (ZnP)₂Q-Q2HP-C₆₀ and (2HP)₂Q-Q2HP-C₆₀, and their model compounds ZnP, 2HP, (ZnP)₂, (2HP)₂, Q2HP-Ph and Ph-C₆₀ in deaerated PhCN

| Molecule | $E°$/mV (*vs.* Fc⁺/Fc) in PhCN | | | | |
	P²⁺/P·⁺	P·⁺/P	C₆₀/C₆₀·⁻	C₆₀·⁻/C₆₀²⁻	P/P·⁻
ZnP	693	306	—	—	−1849
2HP	886	506	—	—	−1671
Ph-C₆₀	—	—	−1023	−1449	—
(ZnP)₂	585	213	—	—	—
(2HP)₂	—	363/454[b]	—	—	−1755
Q2HP-Ph	—	503	—	—	−1648
Q2HP-C₆₀	—	500	−1048	−1458	−1655
ZnPQ-Q2HP-C₆₀	—	328 (522)[c]	−1053	−1450	∼−1560
(ZnP)₂Q-Q2HP-C₆₀	—	228 (533)[c]	−1042	−1463	−1560
(2HP)₂Q-Q2HP-C₆₀	—	371[d] (502)[c]	−1038	−1456	∼−1660 (−1785)[e]

[a] The standard potentials were determined as $E° = (1/2)(E_{pa}° + E_{pc}°)$ by cyclic voltammetry in deaerated PhCN using n-Bu₄NPF₆ (0.1 M) as supporting electrolyte. [b] Two distinct first oxidation potentials of free-base porphyrin were observed (ESI Fig. S2). [c] The first oxidation potential of C₆₀-appended **2HPQ** is shown in parentheses. [d] Peak potential at a scan rate 100 mV s⁻¹ for irreversible reaction. [e] The first reduction potential of the free-base porphyrin dimer **(2HP)₂** is shown in parentheses.

Table 3 Free-energy changes ΔG (eV) for various possible sequences (observed sequence shown in bold where alternatives are presented) of electron-transfer reactions in PhCN, deduced from the observed electrochemical potentials after correction for the Coulomb interaction between the charges[a]

Reaction	Seq.	EN	CS_1	CR_1	CS_2	CR_2
Q2HP*-C$_{60}$ → Q2HP$^{•+}$-C$_{60}$$^{•-}$	—		−0.38	−1.51	—	—
ZnPQ*-Q2HP-C$_{60}$ → ZnPQ-Q2HP*-C$_{60}$ → ZnPQ-Q2HP$^{•+}$-C$_{60}$$^{•-}$ → ZnPQ$^{•+}$-Q2HP-C$_{60}$$^{•-}$	a	−0.08	−0.35	−1.54	−0.17	−1.36
ZnPQ*-Q2HP-C$_{60}$ → ZnPQ-Q2HP*-C$_{60}$ → ZnPQ$^{•+}$-Q2HP$^{•-}$-C$_{60}$ → ZnPQ$^{•+}$-Q2HP-C$_{60}$$^{•-}$	b	−0.08	−0.03	−1.86	−0.49	−1.36
ZnPQ*-Q2HP-C$_{60}$ → ZnPQ$^{•+}$-Q2HP$^{•-}$-C$_{60}$ → ZnPQ$^{•+}$-Q2HP-C$_{60}$$^{•-}$	c	—	−0.11	−1.86	−0.49	−1.36
(ZnP)$_2$Q*-Q2HP-C$_{60}$ → (ZnP)$_2$Q-Q2HP*-C$_{60}$ → (ZnP)$_2$Q-Q2HP$^{•+}$-C$_{60}$$^{•-}$ → (ZnP)$_2$Q$^{•+}$-Q2HP-C$_{60}$$^{•-}$	a	−0.06[b]	−0.35[b]	−1.54	−0.28	−1.25
(ZnP)$_2$Q*-Q2HP-C$_{60}$ → (ZnP)$_2$Q-Q2HP*-C$_{60}$ → (ZnP)$_2$Q$^{•+}$-Q2HP$^{•-}$-C$_{60}$ → (ZnP)$_2$Q$^{•+}$-Q2HP-C$_{60}$$^{•-}$	b	−0.06[b]	−0.14[b]	−1.75	−0.51	−1.24
(ZnP)$_2$Q*-Q2HP-C$_{60}$ → (ZnP)$_2$Q$^{•+}$-Q2HP$^{•-}$-C$_{60}$ → (ZnP)$_2$Q$^{•+}$-Q2HP-C$_{60}$$^{•-}$	c	—	−0.20[b]	−1.75	−0.51	−1.24
(2HP)$_2$Q-Q2HP*-C$_{60}$ → (2HP)$_2$Q*-Q2HP-C$_{60}$ → (2HP)$_2$Q$^{•+}$-Q2HP$^{•-}$-C$_{60}$ → (2HP)$_2$Q$^{•+}$-Q2HP-C$_{60}$$^{•-}$	a	−0.08[c]	0.20[c]	−2.00	−0.61	−1.39
(2HP)$_2$Q-Q2HP*-C$_{60}$ → (2HP)$_2$Q-Q2HP$^{•+}$-C$_{60}$$^{•-}$ → (2HP)$_2$Q$^{•+}$-Q2HP-C$_{60}$$^{•-}$	b	—	−0.38	−1.50	−0.11	−1.39
(2HP)$_2$Q-Q2HP*-C$_{60}$ → (2HP)$_2$Q$^{•+}$-Q2HP$^{•-}$-C$_{60}$ → (2HP)$_2$Q$^{•+}$-Q2HP-C$_{60}$$^{•-}$	c	—	0.12	−2.00	−0.61	−1.39

[a] $\Delta G_{CR_1} = E^0$(anion red.) − E^0(cation ox.) + $e/4\pi\varepsilon_0\varepsilon_s R$ is for charge-recombination from the primary charge-separated state where R is the charge-separation distance determined from DFT; $\Delta G_{CS_1} = -\Delta G_{CR_1} - \Delta E_{0-0}$ is the driving force for primary charge-separation where ΔE_{0-0} is the energy of the lowest excited-state calculated as the average of the energy of the (0–0) band in the absorption and the emission spectra and listed in Table 1; $\Delta G_{CS_2} = E^0$(new group ox.) − E^0(old group ox.) + $(e/4\pi\varepsilon_0\varepsilon_s)(1/R_{old} - 1/R_{new})$ is the driving force for secondary charge-separation where R_{old} is the distance from the counter ion to the originally charged group and R_{new} is the distance from the counter ion to the newly charged group; $\Delta G_{CR_2} = \Delta G_{CR_1} - \Delta G_{CS_2}$ is the driving force for the charge-recombination from the fully charge-separated state; and EN is the excitation energy transfer obtained as the difference of the ΔE_{0-0} values of the excited states. The parameters used are: $\varepsilon_0 = 25.2$ for PhCN, $R(P_{dimer}-C_{60}) = 32.5$ Å, $R(P_{C60-appended}-C_{60}) = 15$ Å, $R(P_{dimer}-P_{C60-appended}) = 19.5$ Å and $R(ZnPQ-C_{60}) = 31$ Å (for **ZnPQ-QH$_2$P-C$_{60}$**). [b] ±0.02 eV. [c] ±0.01 eV.

energy of origin Qx bands in the absorption and emission spectra (Table 1).

3.4 Transient absorption spectroscopy

Femtosecond and nanosecond laser flash photolysis experiments were used to characterize the electron-transfer processes in **Q2HP-C$_{60}$**, **ZnPQ-Q2HP-C$_{60}$**, **(ZnP)$_2$Q-Q2HP-C$_{60}$** and **(2HP)$_2$Q-Q2HP-C$_{60}$**. The resulting spectra $\Delta A(\nu, t)$ display the combined absorption of the active photochemical species with positive sign at time t after excitation minus the spectrum of the ground-state molecule. The femtosecond spectra are fitted to give spectral components $\Delta A_i(\nu)$ and rate constants $k = 1/\tau$. Full details of the observed spectra, the fitted spectra, and the deduced component concentrations as a function of time are provided in Section S4 Fig. S8–S11 of the ESI.†

Q2HP-C$_{60}$. The femtosecond transient absorption spectra $\Delta A(\nu, t)$ of the dyad **Q2HP-C$_{60}$** in deaerated benzonitrile taken after excitation of the porphyrin Soret band at the 430 nm are shown in Fig. 2a. This shows the change in absorption at energy $h\nu$ as a function of time t after excitation. These are then fitted to the kinetics scheme shown in Fig. 2e, with the resultant component spectra shown in Fig. 2b; process rates are shown in Fig. 2e, and fitted spectra and component concentration profiles as a function of time are provided in ESI Fig. S8.† The spectrum of the first identified component, P$_1$, depicts the initial formation of the ^{1}P(Soret)* excited state after $\tau_1 = 220$ fs. This is followed by relaxation from ^{1}P(Soret)* to the lowest-energy singlet porphyrin Q band ^{1}P(S$_1$)* (P$_2$) with a rise time of $\tau_2 = 800$ fs *via* internal conversion (IC). This component is characterized by a strong new absorption feature around 490 nm and well resolved negative peaks at 530, 566, 605 and 656 nm, indicating loss of the standard ground-state Q-band transitions.

Subsequent to the formation of the ^{1}P(S$_1$)*, an electron-transfer process occurs with $\tau_3 = 120$ ps forming the charge-separated species Q2HP$^{•+}$-C$_{60}$$^{•-}$ (P$_4$). The visible region (450–780 nm) of the observed component spectrum (ESI Fig. S5b†) is consistent with the spectrum of 2HPQ$^{•+}$ reported earlier.[86] The observed spectrum of this component in the 950–1050 nm region also matches that expected for C$_{60}$$^{•-}$,[53,126,132,133] unambiguously identifying the product state as Q2HP$^{•+}$-C$_{60}$$^{•-}$. The charge-separated state of **Q2HP-C$_{60}$** recombines on a timescale of $\tau_4 = 880$ ps leaving behind a final component with an estimated quantum yield of about 2%.

Fig. 2c shows results from nanosecond transient absorption spectroscopy. A long-lived component with a similar spectrum to the final component observed in the femtosecond experiments is observed. This species is identified as ^{3}2HPQ(T$_1$)*, most likely produced from ^{1}P(S$_1$)* through intersystem crossing (ISC) with the time constant determined from its yield to be 7 ns. This rate is slightly faster than the observed fluorescence decay rate of **2HP**, 14 ns, and slower than the lower bound found for free-base porphyrin-fullerene dyad without the fusion of quinoxaline, 4 ns.[78,134] While the energy of ^{3}2HPQ(T$_1$)* has not been measured, comparison of the energies of ^{3}ZnPQ(T$_1$)* (1.32 eV)[135] and ^{3}ZnP(T$_1$)* (1.53 eV)[53] suggests a value near 1.1 eV which is clearly much lower than that of the charge-separated state (1.51 eV). Hence the triplet state could also have been produced by charge-recombination and the observed data do not exclude this possibility. As ISC alone can account for the observed triplet yield, this process appears unlikely to have occurred, however. The nanosecond data indicates that ^{3}2HPQ(T$_1$)* decays by phosphorescence with a time constant of 9 μs.

ZnPQ-Q2HP-C$_{60}$. Fig. 3a shows the time-resolved transient absorption spectra $\Delta A(\nu, t)$ of **ZnPQ-Q2HP-C$_{60}$** measured by femtosecond laser flash photolysis in benzonitrile. These are
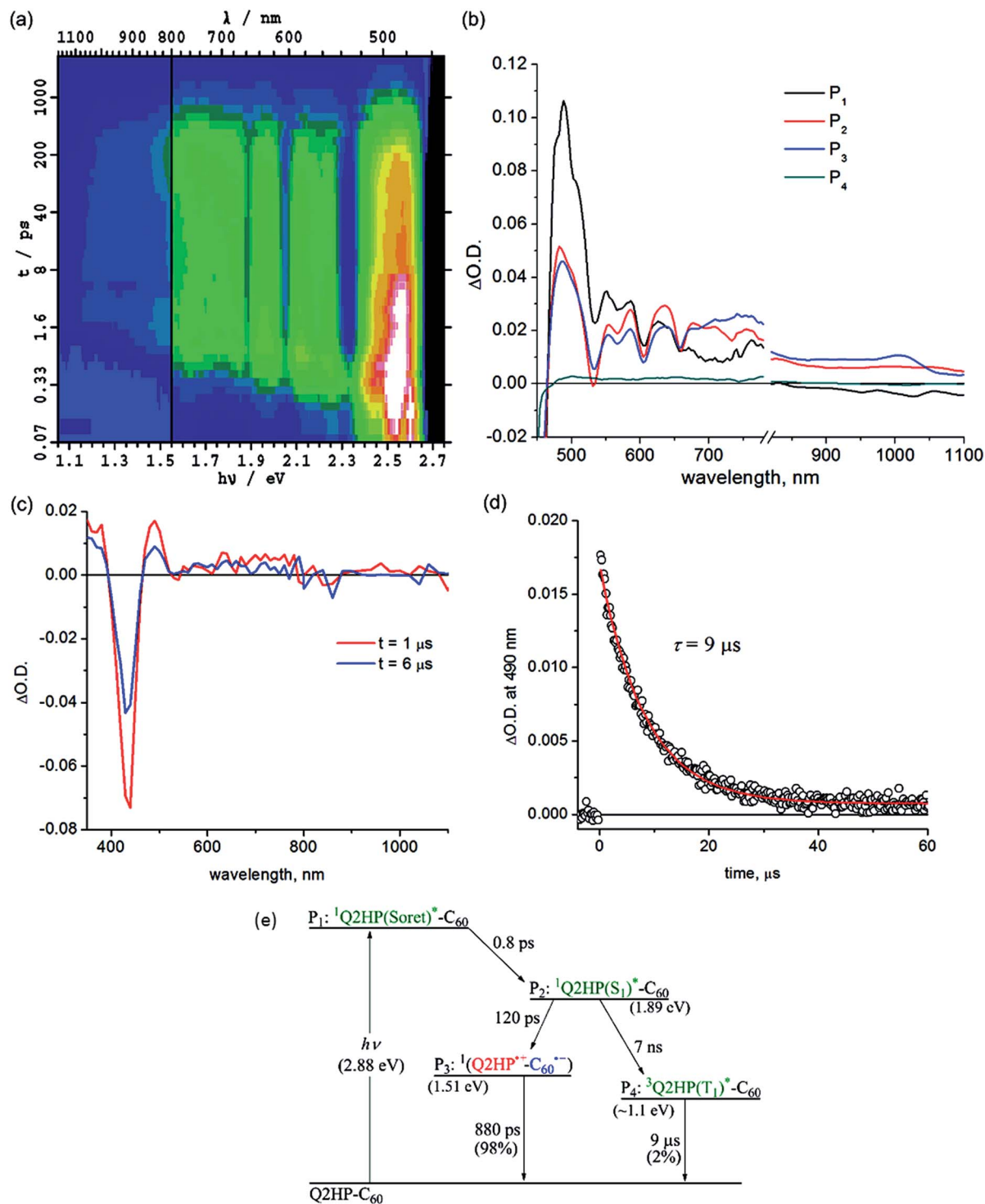
Fig. 2 Transient absorption spectra of **Q2HP-C$_{60}$** following excitation at 430 nm in PhCN at 298 K: (a) $\Delta A(\nu, t)$ at frequency ν following femtosecond laser pulse irradiation at prior time t; (b) component spectra $P_i(\nu)$ (shown as a function of wavelength); (c) 1 and 6 μs after nanosecond laser pulse; (d) the time–decay profile of the optical density at 490 nm; and (e) the reaction scheme used with fitted process lifetimes.

then fitted to the kinetics scheme shown in Fig. 3e; the fitted rate constants and excitation partitioning are given in Fig. 3e, whilst the deduced spectra of the resolved components are shown in Fig. 3b and the fitted time-resolved spectral data and the relative component concentrations as a function of time are

provided in ESI Fig. S9.† The first resolved product component P$_1$ depicts the combined ^{1}P(Soret)* excited-states of each chromophore as observed for all other molecules, forming $\tau_1 = 240$ fs after excitation. Component P$_2$ is an intermediate that forms $\tau_2 = 0.3$ ps later and leads to components P$_3$ and P$_4$ 1.0 ps later

Fig. 3 Transient absorption spectra of **ZnPQ-Q2HP-C$_{60}$** following excitation at 430 nm in PhCN at 298 K: (a) $\Delta A(\nu, t)$ at frequency ν following femtosecond laser pulse irradiation at prior time t; (b) component spectra $P_i(\nu)$ (shown as a function of wavelength); (c) 4 and 20 μs after nanosecond laser pulse; (d) the time–decay profile of the optical density at 1000 nm; and (e) the reaction scheme used with fitted process lifetimes τ. Dotted lines indicate expected but unobserved processes.

with yields determined to be 50 : 50. The transient absorption spectrum of component P_3 shows dips at 525 and 626 nm indicating the formation of ^{1}ZnPQ(S_1)*; similarly, the dips for component P_4 at 580, 608 and 655 nm arise from ^{1}2HPQ(S_1)*. The equal yield of these products could arise from their similar absorption coefficients at the excitation wavelength (430 nm), but in addition exciton coupling between the ^{1}P(Soret)* states of each chromophore would most likely occur on a timescale much faster than τ_1 and/or τ_2, redistributing the energy.

Intramolecular singlet–singlet energy transfer from ^{1}ZnPQ(S_1)* to the energetically lower lying ^{1}2HPQ(S_1)* state takes place with a time constant of 30 ps, eliminating the ZnPQ bleach whilst intensifying the **2HPQ** bleach from the transient absorption spectra. After this energy transfer, an electron is injected into the fullerene unit through process CS_{1a} (Table 3) to form the primary charge-separated state ZnPQ-Q2HP$^{•+}$-C$_{60}$$^{•−}$, component P_5, with a time constant of 100 ps. Two other possible charge-separation reactions, namely CS_{1b} and CS_{1c} (see Table 3), could also have occurred. However, the yields of these processes are too low to facilitate the identification of the resulting spectral components. Charge recombination from ZnPQ-Q2HP$^{•+}$-C$_{60}$$^{•−}$ was difficult to quantify as the spectra are only measured for up to 3000 ps and the process is of low yield. This charge-recombination lifetime is much slower than that observed for **Q2HP-C$_{60}$**, 880 ps from Fig. 2e, despite primary charge-separation occurring at a similar rate, 100 ps compared to 120 ps.

The final component P_6 observed in the femtosecond transient absorption spectra has a visible spectrum matching that for ZnPQ$^{•+}$ reported earlier,[86,135] while the C$_{60}$$^{•−}$ signal in the near-infrared region remains. Hence this component results from the secondary charge-separation process CS_{2a} forming the final charge-separated state ZnPQ$^{•+}$-Q2HP-C$_{60}$$^{•−}$. The lifetime of this species is out of the measurable range of this femtosecond-timescale pulsed experiment.

Nanosecond transient absorption spectra taken at 4 and 20 μs are shown in Fig. 3c. These spectra retain all the spectral features of the final component P_6 determined from the femtosecond measurements, ZnPQ$^{•+}$-Q2HP-C$_{60}$$^{•−}$. From the transient decay profiles also shown in this figure, the lifetime of ZnPQ$^{•+}$-Q2HP-C$_{60}$$^{•−}$ is determined to be 20 μs; this species is seen to decay to the ground state rather than to a locally excited triplet state. No long-lived triplet charge-separated species or ^{3}ZnP(T_1)* is observed as the energy of ^{3}2HPQ(T_1)* (~1.1 eV) lies below that of ^{3}(ZnPQ$^{•+}$-Q2HP-C$_{60}$$^{•−}$) (1.54 eV) and ^{3}ZnPQ$(T_1)$* (1.32 eV),[135] so that only long-lived ^{3}2HPQ(T_1)* is expected. Based on the known intersystem crossing rates for **Q2HP-C$_{60}$** and **ZnP-Ph** (2 ns), a net yield of less than 2% is expected for ZnPQ-^{3}2HPQ(T_1)*-C$_{60}$ with a lifetime near 9 μs. The observed data in Fig. 3 does not facilitate the identification of such a component as the major product decays over the same period.

(ZnP)$_2$Q-Q2HP-C$_{60}$. Fig. 4a shows the time-resolved transient absorption spectra $\Delta A(\nu, t)$ of **(ZnP)$_2$Q-Q2HP-C$_{60}$** measured by femtosecond laser flash photolysis in benzonitrile. These are then fitted to the kinetics scheme shown in Fig. 4e; the fitted rate constants and excitation partitioning are given in Fig. 4e, whilst the deduced spectra of the resolved components are

shown in Fig. 4b and the fitted time-resolved spectral data and the relative component concentrations as a function of time are provided in ESI Fig. S10.† The photochemical reactions identified for **(ZnP)$_2$Q-Q2HP-C$_{60}$** are directly analogous to those identified for **ZnPQ-Q2HP-C$_{60}$**. Direct excitation of the porphyrin Soret bands rapidly initiates internal conversions to component P_3, readily identified as ^{1}(ZnP)$_2$Q(S_1)* and component P_4, readily identified as ^{1}2HPQ(S_1)*. The yield of each species is fitted to be 50% whereas the relative absorption coefficients would suggest a ratio of 2 : 1. This result indicates that there was significant exciton transfer between the excited Soret states of each porphyrin before internal conversion was complete.

The contribution of ^{1}(ZnP)$_2$Q(S_1)* to the transient spectra diminishes 20 ps after internal conversion is complete, leaving behind only the contribution of ^{1}2HPQ(S_1)*. This indicates intramolecular singlet–singlet energy transfer from zinc to free-base porphyrin. Subsequent to this energy transfer, primary charge-transfer CS_{1a} occurs after 40 ps, generating component P_5, (ZnP)$_2$Q-Q2HP$^{•+}$-C$_{60}$$^{•−}$. An alternate process CS_{1b}, involving hole transfer from Q2HP to **(ZnP)$_2$Q** is also thermodynamically possible (Table 3), but the component spectrum provides no indication of (ZnP)$_2$Q$^{•+}$-Q2HP$^{•−}$-C$_{60}$ formation. Secondary charge-separation CS_{2a} then occurs to form the final charge-separated species (ZnP)$_2$Q$^{•+}$-Q2HP-C$_{60}$$^{•−}$ with at least 80% yield after 500 ps, in competition with weak primary charge recombination that occurs on a timescale of at least 2 ns.

The final charge-separated state is also observed in the nanosecond transient absorption spectra shown in Fig. 4c. From the decay curves at 700 and 1000 nm, the lifetime of (ZnP)$_2$Q$^{•+}$-Q2HP-C$_{60}$$^{•−}$ is determined to be 5 μs. While the energies of both ^{3}2HPQ(T_1)* and ^{3}(ZnP)$_2$Q(T_1)* are expected to be less than that for (ZnP)$_2$Q$^{•+}$-Q2HP-C$_{60}$$^{•−}$, no evidence for charge recombination to triplet states is obtained. The yield of these triplet states following ISC from the corresponding singlets is estimated to be 1%, and, as their estimated lifetime is ca. 9 μs, these components are not detectable in the observed transient absorption spectra.

(2HP)$_2$Q-Q2HP-C$_{60}$. Fig. 5a shows the time-resolved transient absorption spectra $\Delta A(\nu, t)$ of **(2HP)$_2$Q-Q2HP-C$_{60}$** measured by femtosecond laser flash photolysis in benzonitrile. These are then fitted to the kinetics scheme shown in Fig. 5e; the fitted rate constants and excitation partitioning are given in Fig. 5e, whilst the deduced spectra of the resolved components are shown in Fig. 5b and the fitted time-resolved spectral data and the relative component concentrations as a function of time are provided in ESI Fig. S11.† The kinetics scheme used is similar to that shown in Fig. 4e for **(ZnP)$_2$Q-Q2HP-C$_{60}$** except that the relative energies of the Tröger's-base porphyrins and the central porphyrin Q-bands are inverted, necessitating the inclusion of the up-hill energy transfer process from ^{1}(2HP)$_2$Q(S_1)* to ^{1}2HPQ(S_1)* in the reaction scheme.

Other processes with yields too small to be observed in **(ZnP)$_2$Q-Q2HP-C$_{60}$** also become manifest for **(2HP)$_2$Q-Q2HP-C$_{60}$** such as phosphorescence, requiring more component spectra and more rate constants to be extracted from the data to complete the chemical model. Attempts to extract all of this
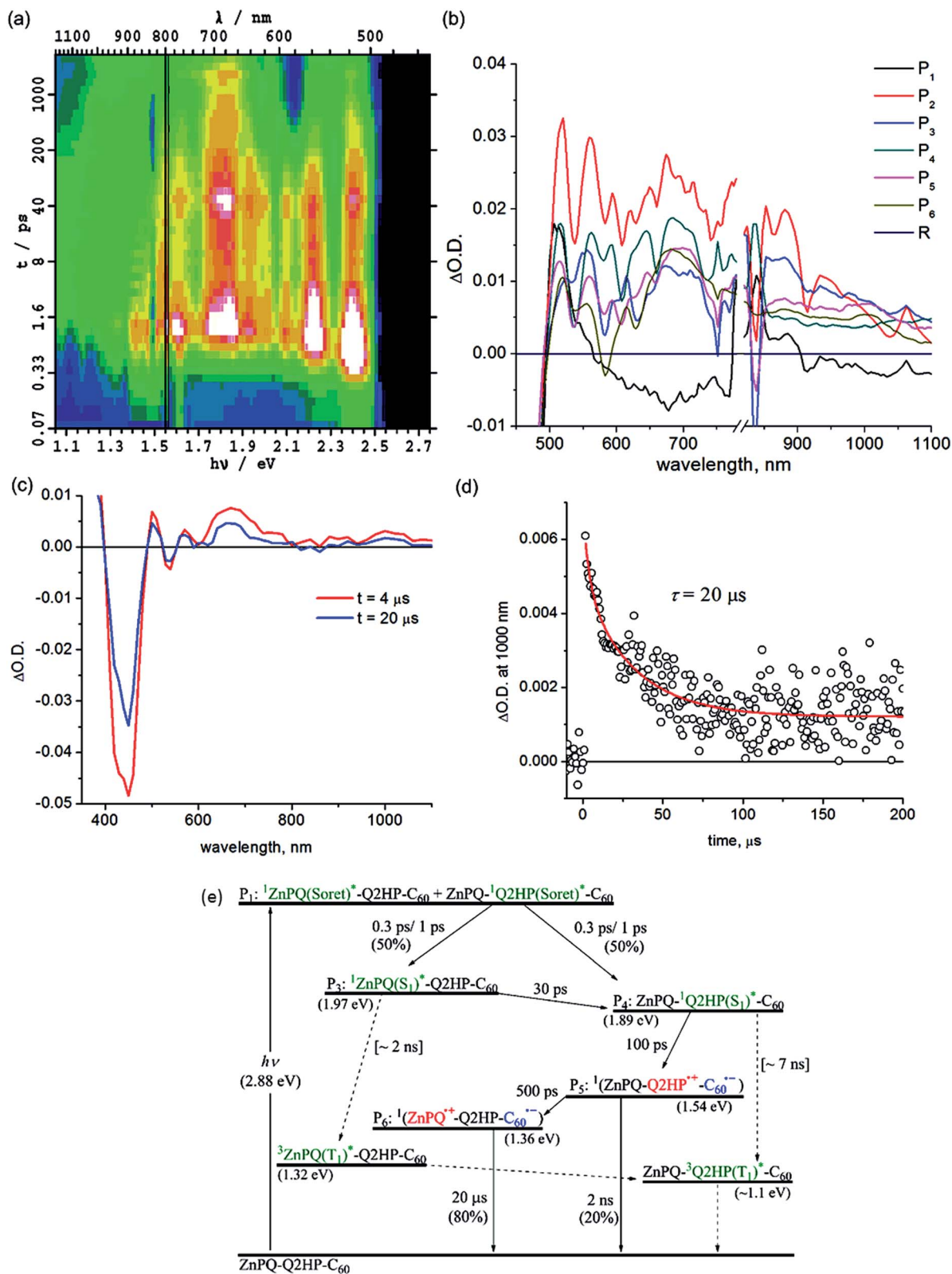
Fig. 4 Transient absorption spectra of **(ZnP)$_2$Q-Q2HP-C$_{60}$** following excitation at 430 nm in PhCN at 298 K: (a) $\Delta A(\nu, t)$ at frequency ν following femtosecond laser pulse irradiation at prior time t; (b) component spectra $P_i(\nu)$ (shown as a function of wavelength); (c) 0.2 and 6 μs after nanosecond laser pulse; (d) the time–decay profile of the optical density at 1000 nm; and (e) the reaction scheme used with fitted process lifetimes τ. Dotted lines indicate expected but unobserved processes.

Fig. 5 Transient absorption spectra of (2HP)$_2$Q-Q2HP-C$_{60}$ following excitation at 430 nm in PhCN at 298 K: (a) $\Delta A(\nu, t)$ at frequency ν following femtosecond laser pulse irradiation at prior time t; (b) component spectra $P_i(\nu)$ (shown as a function of wavelength); (c) 2 and 20 μs after nanosecond laser pulse; (d) the time–decay profile of the optical density at 770 nm; and (e) the reaction scheme used with fitted process lifetimes τ. Dotted lines indicate expected but unobserved processes.
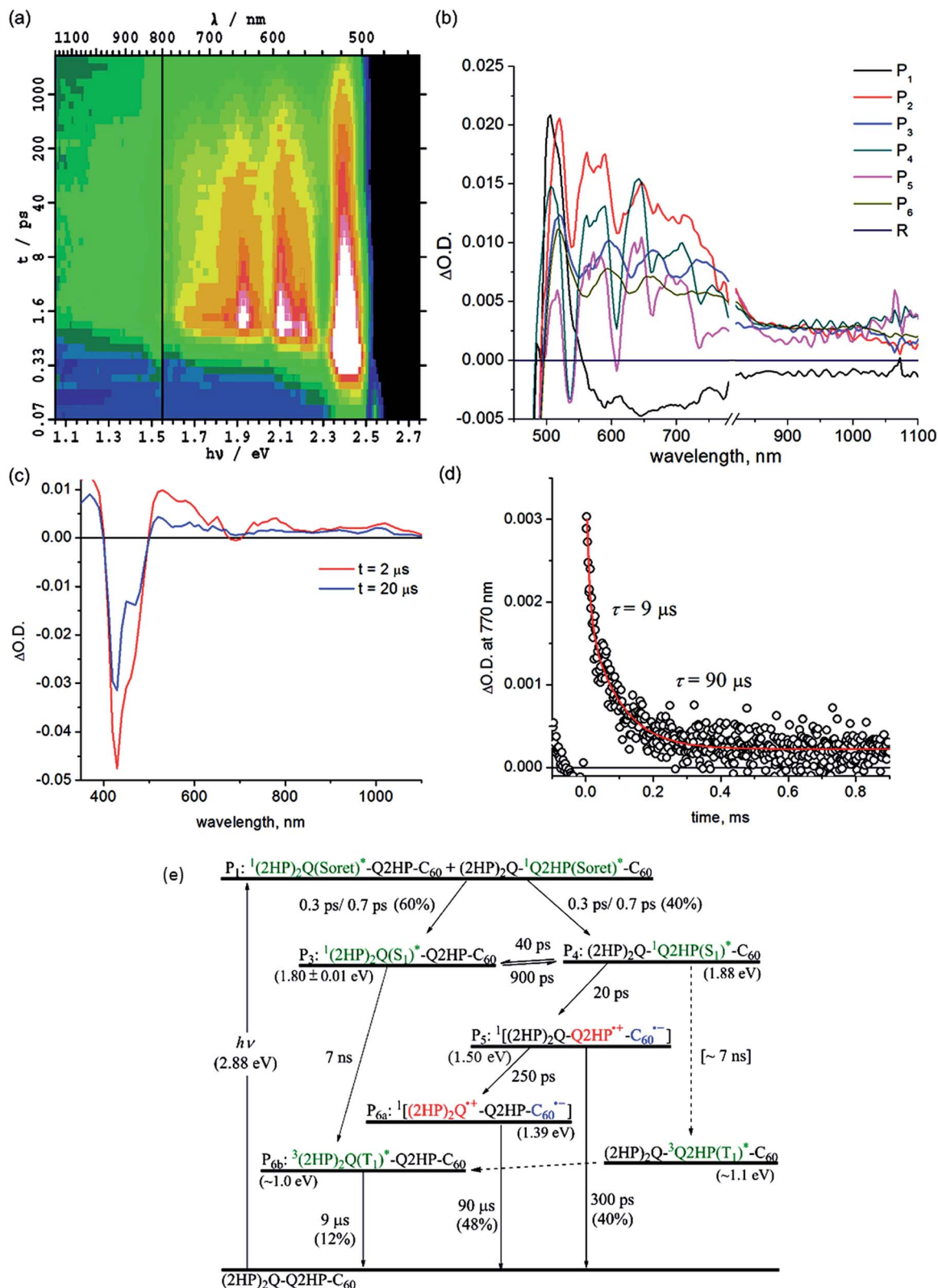
information were unsuccessful as the fitting procedure proved to be underdetermined. In particular, somewhat similar spectra of long-lived components, P_{6a} $(2HP)_2Q^{\bullet+}$-Q2HP-$C_{60}^{\bullet-}$ and P_{6b} $^3(2HP)_2Q(T_1)^*$, could not be differentiated and so a combined spectrum is reported as component P_6. With this restriction, determination of unique rate constants becomes possible in principle, but five rate constants are required to describe the same temporal region while only three were required for $(ZnP)_2Q$-Q2HP-C_{60}; as a result, a range of viable solutions could be obtained for each parameter.

To aid the spectral fitting process, a ratio of 22 for the forward and reverse rate constants for the exciton transfer from P_3 $^1(2HP)_2Q(S_1)^*$ to P_4 ^{1}2HPQ$(S_1)^*$ was determined from the observed free-energy difference of 0.08 eV; however, the free energy different is uncertain to at least ± 0.01 eV, allowing this ratio to vary from 15 to 33, and good fits to the spectral data could be obtained using any ratio in this range. The extracted rate constants are thus only accurate to at most a factor of two.

The yields of component P_3, $^1(2HP)_2Q(S_1)^*$, and component P_4, ^{1}2HPQ$(S_1)^*$, are fitted to be 60 and 40%, respectively, close to the ratio of the extinction coefficients of each chromophore. Primary and secondary charge-separation initiated by ^{1}2HPQ$(S_1)^*$ are deduced to occur in 20 and 250 ps, respectively, both processes being faster than those observed for **ZnPQ-Q2HP-C_{60}** (100 and 500 ps) and **$(ZnP)_2Q$-Q2HP-C_{60}** (40 and 500 ps).

However, the activated exciton transfer between $^1(2HP)_2Q(S_1)^*$ and ^{1}2HPQ$(S_1)^*$, with forward and reverse reaction times of 900 and 40 ps, respectively, ensures that $^1(2HP)_2Q(S_1)^*$ remains present for a long time. The fitted component concentrations reported in Fig. S11† reveal that the proportion of $^1(2HP)_2Q(S_1)^*$ is 12% after 2 ns, indicating the significance of this metastable trapped species.

Component P_5 is identified as $(2HP)_2Q$-Q2HP$^{\bullet+}$-$C_{60}^{\bullet-}$, the product of primary charge-separation process CS_{1b}, through assignment of the visible and NIR spectra to 2HPQ$^{\bullet+}$ and $C_{60}^{\bullet-}$, respectively. Component P_{6a} is identified as $(2HP)_2Q^{\bullet+}$-Q2HP-$C_{60}^{\bullet-}$, generated *via* the secondary charge-separation process CS_{2b}, based on the observed transient absorption spectrum of $(2HP)_2^{\bullet+}$ produced electrochemically, that is shown in ESI Fig. S13b,† combined with the continuity of the NIR absorption of $C_{60}^{\bullet-}$. In contrast to the observed spectra for the other molecules considered, the observed signal strength decreases significantly in the nanosecond regime, indicating that primary charge-recombination CR_{1b} competes with secondary charge-separation. The fitted lifetime for this process is 300 ps, shorter than the value of 880 ps deduced for **Q2HP-C_{60}** and much shorter that the value of >2 ns deduced for **ZnPQ-Q2HP-C_{60}** and **$(ZnP)_2Q$-Q2HP-C_{60}**.

The lifetime of the final charge-separated state is determined to be 90 μs by nanosecond laser flash photolysis (Fig. 5c). Notably, the time decay profiles at 770 nm (Fig. 5d) show biexponential character with a fast decay component of 9 μs (20%) and a slow component of 90 μs (80%). The time profiles at 1000 nm (ESI Fig. S12†), however, show only single exponential decay with a lifetime of 90 μs. As the 1000 nm transient absorption is attributed to $C_{60}^{\bullet-}$, it is clear that the slow component

corresponds to the charge-recombination of the final charge-separated state $(2HP)_2Q^{\bullet+}$-Q2HP-$C_{60}^{\bullet-}$. The fast decay component is only observed in the visible region and clearly comes from the porphyrin macrocycle. This process is hence associated with phosphorescence from the lowest-energy triplet state, $^3(2HP)_2Q(T_1)^*$. It is possible that this species is produced by charge-recombination of $(2HP)_2Q^{\bullet+}$-Q2HP-$C_{60}^{\bullet-}$, either directly or *via* ^{3}Q2HP$(T_1)^*$. In Fig. 5e, however, this process is not included, only production *via* ISC from in particular $^1(2HP)_2Q(S_1)^*$. This process is expected to be important as $^1(2HP)_2Q(S_1)^*$ is a long-lived trap in the photochemical process, and indeed the fitted lifetime of $^1(2HP)_2Q(S_1)^*$ is 7 ns, in good agreement with expectations based on the observed value of 7 ns for **Q2HP-C_{60}** (Fig. 2e).

3.5 Extraction of solvent reorganization energies and electronic couplings from the observed lifetimes

The observed rate constants of the deduced components of all the compounds studied by transient absorption spectroscopy are summarized in Table 4. With the free-energy changes listed in Table 3, the coupling parameters V and solvent reorganization energies λ_o are determined (Table 4) using the semiclassical rate equation:[136]

$$k_{ET} = \frac{2\pi}{\hbar} H_{rp}{}^2 (4\pi\lambda_o k_B T)^{-\frac{1}{2}} \sum_n P(E_{Rn}) \sum_m \langle \chi_{Pm} | \chi_{Rn} \rangle^2$$
$$\times \exp\left[-\frac{(\Delta G^\circ + E_{Pm} - E_{Rn} + \lambda_o)}{4\lambda_o k_B T} \right]$$

The Franck–Condon factors associated with changes in the intramolecular modes required for this procedure are evaluated from DFT frequency calculations of the vibrational modes of the molecular fragments in their various states of ionization; the vibrational parameters deduced by this procedure are given in full in ESI Table S1.†

The coupling V and solvent reorganization energy λ_o for **Q2HP-C_{60}** are extracted using the rates for charge-separation and charge-recombination, using the common assumption that the coupling and reorganization energy for both processes are the same. While calculated reorganization energies for these processes typically are very similar, coupling strengths can vary by an order of magnitude and so this analysis is actually quite approximate.[78] The coupling V and solvent reorganization energies λ_o are determined to be 19 cm^{-1} and 0.69 eV, respectively. A reorganization energy of 0.64 eV and a coupling of 20 cm^{-1} are obtained for the primary charge-separation process of **ZnPQ-Q2HP-C_{60}**. For secondary charge-recombination (CR_2), using 0.6 eV < λ_o < 0.8 eV, the values deduced for V range from 0.041 to 0.102 cm^{-1} for recombination on the singlet manifold. Similarly, the reorganization energies λ_o and couplings V of the primary charge-separation process for **$(ZnP)_2Q$-Q2HP-C_{60}** are determined to be 0.60 eV and 25 cm^{-1}, and for **$(2HP)_2Q$-Q2HP-C_{60}** are 0.64 eV and 40 cm^{-1}. For CR_2, the couplings V are deduced to be 0.07–0.163 cm^{-1} and 0.021–0.055 cm^{-1} for **$(ZnP)_2Q$-Q2HP-C_{60}** and **$(2HP)_2Q$-Q2HP-C_{60}**, respectively. The non-planar Qx–Qx linkage reduces the conjugation and

Table 4 Kinetic and energetic data extracted from the results of the femtosecond and nanosecond transient absorption spectra, Fig. S8–S11: λ_o is the solvent reorganization energy, while V is the coupling strength on the singlet manifolds

Molecule	Process	τ	λ_o (eV)	V (cm^{-1})
Q2HP-C$_{60}$	^{1}P(Soret)^a	0.22 ps	—	—
	^{1}IC^b	0.8 ps	—	—
	ISC	7 ns	—	—
	CS$_1$	120 ps	0.69	19
	CR$_1$	880 ps	0.69	19
	^{3}P*^{e}	9 μs	—	—
ZnPQ-Q2HP-C$_{60}$	^{1}P(Soret)^a	0.24	—	—
	^{1}IC^b	0.3, 1.0 ps	—	—
	EN^c	30 ps	—	—
	CS$_{1a}^d$	100 ps	0.64	20
	CR$_{1a}^d$	2 ns	0.64	20
	CS$_{2a}^d$	500 ps	[0.65–0.85]	31–84
	CR$_2$	20 μs	[0.6–0.8]	0.041–0.102
(ZnP)$_2$Q-Q2HP-C$_{60}$	^{1}P(Soret)^a	0.3 ps	—	—
	^{1}IC^b	0.5, 1.6 ps	—	—
	EN^c	20 ps	—	—
	CS$_{1a}^d$	40 ps	0.60	25
	CR$_{1a}^d$	2 ns	0.60	25
	CS$_{2a}^d$	500 ps	[0.6–0.8]	12–30
	CR$_2$	5 μs	[0.55–0.75]	0.070–0.163
(2HP)$_2$Q-Q2HP-C$_{60}$	^{1}P(Soret)^a	0.24 ps	—	—
	^{1}IC^b	0.3, 0.7 ps	—	—
	ISC	7 ns	—	—
	CS$_{1b}^d$	20 ps	0.64	40
	CR$_{1b}^d$	300 ps	0.64	40
	CS$_{2b}^d$	250 ps	[0.65–0.85]	70–190
	CR$_2$	90 μs	[0.6–0.8]	0.021–0.055

^{a} The instrument-limited rise time of the initial excited-state. ^{b} Internal conversion from the excited Soret-states of the porphyrin ^{1}P(Soret)* to the lowest-energy Q-state of the porphyrin ^{1}P(S$_1$)*. ^{c} The intramolecular singlet–singlet energy-transfer from the energetically higher lying ^{1}P(S$_1$)* to the lower lying ^{1}P(S$_1$)*. ^{d} The available charge-separation process options are defined in Table 3, CS$_{1a}$ and/or CS$_{1b}$ etc., may in principle be observed. ^{e} Phosphorescence lifetime.

therefore weakens the couplings. Chemical substitutions on the molecules to artificially control the non-planarity of these groups can therefore be used as an easy way of controlling photochemical properties.

4. Conclusions

The photoinduced electron-transfer reactions of photosynthetic reaction centre (PRC) models, **Q2HP-C$_{60}$**, **ZnPQ-Q2HP-C$_{60}$**, **(ZnP)$_2$Q-Q2HP-C$_{60}$** and **(2HP)$_2$Q-Q2HP-C$_{60}$**, were synthesised and studied using the femtosecond and nanosecond transient absorption spectroscopy and the lifetimes of the photochemical reaction, the solvent reorganization energies and couplings were extracted (Table 4). All molecules show picosecond-timescale charge-separations, **ZnPQ-Q2HP-C$_{60}$**, **(ZnP)$_2$Q-Q2HP-C$_{60}$** and **(2HP)$_2$Q-Q2HP-C$_{60}$** restrained singlet charge-separations to

the microsecond-timescale, whereas this process occurs on the picosecond-timescale for **Q2HP-C$_{60}$**. The long-lived component is maintained on the singlet manifold and the quantum yield for its production is high. **ZnPQ-Q2HP-C$_{60}$**, **(ZnP)$_2$Q-Q2HP-C$_{60}$** and **(2HP)$_2$Q-Q2HP-C$_{60}$** increase the distance between the final charge-separated ion pair while the internal biquinoxalinyl linkage reduces the coupling, affording the longer-lived species on the singlet manifold in these molecules. Also, all charge-recombination is observed to occur to the ground state rather than to the energetically accessible local triplet states.

The PRC model molecules **(ZnP)$_2$Q-Q2HP-C$_{60}$** and **(2HP)$_2$Q-Q2HP-C$_{60}$** incorporating porphyrin arrays and fullerene connected by Tröger's base, biquinoxalinyl and imidazole linkers were designed to have similar inter-chromophoric distances to those in the natural PRC. The centre-to-centre distance of 6.2 Å in the Tröger's base porphyrin dimer is close to that of 7.0 Å for the special pair in the natural PRC, see Fig. 1, suggesting that control over the system can be achieved by introducing small chemical modifications,[78,84,95–103] just the way in which mutagenesis controls the natural PRC.[88,137] Also, the distances for electron-transfer reactions are 18.7 and 15.0 Å for these models that is very close to that of 18.0 and 14.3 Å found in the natural system, so the electron-transport properties are quite similar.

While the processes driving charge separation and recombination are quite different in the natural and artificial systems, the resulting process lifetimes (Fig. 1) are comparable. In the natural system, charge separation to the first state stable for times longer than its production time is to species P$^+$H$_L^-$, that takes 4–7 ps to appear,[62] secondary charge separation then occurs[62] in 200–250 ps ahead of primary charge recombination (0.6 to 10 ns),[62] and then secondary charge recombination takes 100 ms.[94] The corresponding times for the biomimetic compounds are 20–40 ps, 250–500 ps, 0.3–2 ns, and 5–90 μs, respectively. All are within an order of magnitude of the natural system except for the secondary charge recombination times, which are 3–4 orders of magnitude shorter. This discrepancy is not necessarily of concern, however, as processes that take and utilize separated charges typically occur on the scale of a few μs and so the model compounds are stable enough, more so than most alternatives that have been considered. Related compounds show secondary charge separation lasting for over 1 ms at room temperature,[78] so synthetic methods are available to meet specific requirements.

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Notes and references

1 S. Fukuzumi, K. Ohkubo and T. Suenobu, *Acc. Chem. Res.*, 2014, **47**, 1455.

2 G. Li, R. Zhu and Y. Yang, *Nat. Photonics*, 2012, **6**, 153.

3 H. Imahori, T. Umeyama and S. Ito, *Acc. Chem. Res.*, 2009, **42**, 1809.

4 D. Gust, T. A. Moore and A. L. Moore, *Acc. Chem. Res.*, 2009, **42**, 1890.

5 M. J. Llansola-Portoles, R. E. Palacios, D. Gust, T. A. Moore and A. L. Moore, in *From Molecules to Materials: Pathways to Artificial Photosynthesis*, 2015, p. 71.

6 S. Zhou, M. Yamamoto, G. A. D. Briggs, H. Imahori and K. Porfyrakis, *J. Am. Chem. Soc.*, 2016, **138**, 1313.

7 T. Miura, R. Tao, S. Shibata, T. Umeyama, T. Tachikawa, H. Imahori and Y. Kobori, *J. Am. Chem. Soc.*, 2016, **138**, 5879.

8 G. J. Huang, M. A. Harris, M. D. Krzyaniak, E. A. Margulies, S. M. Dyar, R. J. Lindquist, Y. Wu, V. V. Roznyatovskiy, Y. L. Wu, R. M. Young and M. R. Wasielewski, *J. Phys. Chem. B*, 2016, **120**, 756.

9 M. Rudolf, S. V. Kirner and D. M. Guldi, *Chem. Soc. Rev.*, 2016, **45**, 612.

10 T. Higashino, T. Yamada, M. Yamamoto, A. Furube, N. V. Tkachenko, T. Miura, Y. Kobori, R. Jono, K. Yamashita and H. Imahori, *Angew. Chem., Int. Ed.*, 2016, **55**, 629.

11 L. Martín-Gomis, G. Rotas, K. Ohkubo, F. Fernández-Lázaro, S. Fukuzumi, N. Tagmatarchis and A. Sastre-Santos, *Nanoscale*, 2015, **7**, 7437.

12 S. Fukuzumi, in *Chemical Science of π-Electron Systems*, ed. T. Akasada, A. Osuka, S. Fukuzumi, H. Kandori and Y. Aso, Springer, Japan, 2015, p. 529.

13 R. C. Huber, A. S. Ferreira, R. Thompson, D. Kilbride, N. S. Knutson, L. S. Devi, D. B. Toso, J. R. Challa, Z. H. Zhou, Y. Rubin, B. J. Schwartz and S. H. Tolbert, *Science*, 2015, **348**, 1340.

14 T. Kamimura, K. Ohkubo, Y. Kawashima, S. Ozako, K. I. Sakaguchi, S. Fukuzumi and F. Tani, *J. Phys. Chem. C*, 2015, **119**, 25634.

15 A. Arrigo, A. Santoro, F. Puntoriero, P. P. Lainé and S. Campagna, *Coord. Chem. Rev.*, 2015, **304–305**, 109.

16 M. Vizuete, M. J. Gómez-Escalonilla, M. Barrejón, J. L. G. Fierro, M. Zhang, M. Yudasaka, S. Iijima, P. Atienzar, H. García and F. Langa, *Phys. Chem. Chem. Phys.*, 2016, **18**, 1828.

17 C. Stangel, C. Schubert, S. Kuhri, G. Rotas, J. T. Margraf, E. Regulska, T. Clark, T. Torres, N. Tagmatarchis, A. G. Coutsolelos and D. M. Guldi, *Nanoscale*, 2015, 7, 2597.

18 S. V. Kirner, C. Henkel, D. M. Guldi, J. D. Megiatto Jr and D. I. Schuster, *Chem. Sci.*, 2015, **6**, 7293.

19 S. V. Kirner, D. M. Guldi, J. D. Megiatto Jr and D. I. Schuster, *Nanoscale*, 2015, **7**, 1145.

20 M. Rudolf, O. Trukhina, J. Perles, L. Feng, T. Akasaka, T. Torres and D. M. Guldi, *Chem. Sci.*, 2015, **6**, 4141.

21 S. V. Kirner, D. Arteaga, C. Henkel, J. T. Margraf, N. Alegret, K. Ohkubo, B. Insuasty, A. Ortiz, N. Martín, L. Echegoyen, S. Fukuzumi, T. Clark and D. M. Guldi, *Chem. Sci.*, 2015, **6**, 5994.

22 L. Moreira, J. Calbo, R. M. Krick Calderon, J. Santos, B. M. Illescas, J. Aragó, J. F. Nierengarten, D. M. Guldi, E. Ortí and N. Martín, *Chem. Sci.*, 2015, **6**, 4426.

23 O. Trukhina, M. Rudolf, G. Bottari, T. Akasaka, L. Echegoyen, T. Torres and D. M. Guldi, *J. Am. Chem. Soc.*, 2015, **137**, 12914.

24 D. Gust, *Faraday Discuss.*, 2015, **185**, 9.

25 J. Arero, G. Kodis, R. A. Schmitz, D. D. Mendez-Hernandez, T. A. Moore, A. L. Moore and D. Gust, *J. Porphyrins Phthalocyanines*, 2015, **19**, 934.

26 N. L. Bill, M. Ishida, Y. Kawashima, K. Ohkubo, Y. M. Sung, V. M. Lynch, J. M. Lim, D. Kim, J. L. Sessler and S. Fukuzumi, *Chem. Sci.*, 2014, **5**, 3888.

27 V. M. Blas-Ferrando, J. Ortiz, K. Ohkubo, S. Fukuzumi, F. Fernández-Lázaro and Á. Sastre-Santos, *Chem. Sci.*, 2014, **5**, 4785.

28 M. Supur, Y. Kawashima, Y. X. Ma, K. Ohkubo, C. F. Chen and S. Fukuzumi, *Chem. Commun.*, 2014, **50**, 15796.

29 T. Zhang and W. Lin, *Chem. Soc. Rev.*, 2014, **43**, 5982.

30 S. Pillai, J. Ravensbergen, A. Antoniuk-Pablant, B. D. Sherman, R. Van Grondelle, R. N. Frese, T. A. Moore, D. Gust, A. L. Moore and J. T. M. Kennis, *Phys. Chem. Chem. Phys.*, 2013, **15**, 4775.

31 V. Garg, G. Kodis, P. A. Liddell, Y. Terazono, T. A. Moore, A. L. Moore and D. Gust, *J. Phys. Chem. B*, 2013, **117**, 11299.

32 M. Maiuri, J. J. Snellenburg, I. H. M. Van Stokkum, S. Pillai, K. Wongcarter, D. Gust, T. A. Moore, A. L. Moore, R. Van Grondelle, G. Cerullo and D. Polli, *J. Phys. Chem. B*, 2013, **117**, 14183.

33 C. Zhou, Q. Liu, W. Xu, C. Wang and X. Fang, *Chem. Commun.*, 2011, **47**, 2982.

34 M. G. Alvarez, C. Prucca, M. E. Milanesio, E. N. Durantini and V. Rivarola, *Int. J. Biochem. Cell Biol.*, 2006, **38**, 2092.

35 X. F. Zhang and W. Guo, *J. Photochem. Photobiol., A*, 2011, **225**, 117.

36 F. F. Sperandio, S. K. Sharma, M. Wang, S. Jeon, Y. Y. Huang, T. Dai, S. Nayka, S. C. O. M. de Sousa, L. Y. Chiang and M. R. Hamblin, *Nanomed.: Nanotechnol., Biol. Med.*, 2013, **9**, 570.

37 Y. Chen, D. Zhao and Y. Liu, *Chem. Commun.*, 2015, **51**, 12266.

38 J. Shi, L. Wang, J. Gao, Y. Liu, J. Zhang, R. Ma, R. Liu and Z. Zhang, *Biomaterials*, 2014, **35**, 5771.

39 Y. Y. Huang, S. K. Sharma, R. Yin, T. Agrawal, L. Y. Chiang and M. R. Hamblin, *J. Biomed. Nanotechnol.*, 2014, **10**, 1918.

40 S. A. Dingsdag, B. C. M. Yap, N. Hunter and M. J. Crossley, *Org. Biomol. Chem.*, 2015, **13**, 98.

41 A. P. N. Singh, M. A. Harris, R. M. Young, S. A. Miller, M. R. Wasielewski and F. D. Lewis, *Faraday Discuss.*, 2015, **185**, 105.

42 Y. Takano, T. Numata, K. Fujishima, K. Miyake, K. Nakao, W. D. Grove, R. Inoue, M. Kengaku, S. Sakaki, Y. Mori, T. Murakami and H. Imahori, *Chem. Sci.*, 2016, **7**, 3331.

43 J. G. Rohan, Y. R. Citron, A. C. Durrell, L. E. Cheruzel, H. B. Gray, R. H. Grubbs, M. Humayun, K. L. Engisch, V. Pikov and R. H. Chow, *ACS Chem. Neurosci.*, 2013, **4**, 585.

44 H. Takakura, R. Kojima, M. Kamiya, E. Kobayashi, T. Komatsu, T. Ueno, T. Terai, K. Hanaoka, T. Nagano and Y. Urano, *J. Am. Chem. Soc.*, 2015, **137**, 4010.

45 C. Schubert, M. Wielopolski, L.-H. Mewes, G. d. M. Rojas, C. van der Pol, K. C. Moss, M. R. Bryce, J. E. Moser, T. Clark and D. M. Guldi, *Chem.–Eur. J.*, 2013, **19**, 7575.

46 G. Copley, T. A. Moore, A. L. Moore and D. Gust, *Adv. Mater.*, 2013, **25**, 456.

47 L. Wibmer, L. M. O. Lourenço, A. Roth, G. Katsukis, M. G. P. M. S. Neves, J. A. S. Cavaleiro, J. P. C. Tomé, T. Torres and D. M. Guldi, *Nanoscale*, 2015, **7**, 5674.

48 R. E. Blankenship, *Molecular mechanisms of photosynthesis*, Blackwell Science, Oxford, 2002.

49 W. W. Parson and A. Warshel, in *The Purple Prototropic Bacteria*, Springer, Dordrecht, 2009, p. 355.

50 J. M. Warman, M. P. d. Haas, M. N. Paddon-Row, E. Cotsaris, N. S. Hush, H. Oevering and J. W. Verhoeven, *Nature*, 1986, **320**, 615.

51 M. R. Wasielewski, D. G. Johnson, W. A. Svec, K. M. Kersey and D. W. Minsek, *J. Am. Chem. Soc.*, 1988, **110**, 7219.

52 M. R. Wasielewski, *Chem. Rev.*, 1992, **92**, 435.

53 C. Luo, D. M. Guldi, H. Imahori, K. Tamaki and Y. Sakata, *J. Am. Chem. Soc.*, 2000, **122**, 6535.

54 D. Gust, T. A. Moore and A. L. Moore, *Acc. Chem. Res.*, 2001, **34**, 40.

55 Z. E. X. Dance, Q. Mi, D. W. McCamant, M. J. Ahrens, M. A. Ratner and M. R. Wasielewski, *J. Phys. Chem. B*, 2006, **110**, 25163.

56 S. Fukuzumi, *Phys. Chem. Chem. Phys.*, 2008, **10**, 2283.

57 B. D. Sherman, M. D. Vaughn, J. J. Bergkamp, D. Gust, A. L. Moore and T. A. Moore, *Photosynth. Res.*, 2014, **120**, 59.

58 M. Malferrari, A. Mezzetti, F. Francia and G. Venturoli, *Biochim. Biophys. Acta, Bioenerg.*, 2013, **1827**, 328.

59 A. G. Yakovlev and V. A. Shuvalov, *J. Theor. Biol.*, 2014, **343**, 92.

60 K. Gibasiewicz, M. Pajzderska, A. Dobek, J. Karolczak, G. Burdzinski, K. Brettel and M. R. Jones, *Phys. Chem. Chem. Phys.*, 2013, **15**, 16321.

61 K. Gibasiewicz, M. Pajzderska, A. Dobek, K. Brettel and M. R. Jones, *J. Phys. Chem. B*, 2013, **117**, 11112.

62 K. Gibasiewicz, R. Białek, M. Pajzderska, J. Karolczak, G. Burdziński, M. R. Jones and K. Brettel, *Photosynth. Res.*, 2016, 1.

63 A. C. Benniston, *Phys. Chem. Chem. Phys.*, 2007, **9**, 5739.

64 K. Ohkubo and S. Fukuzumi, *Bull. Chem. Soc. Jpn.*, 2009, **82**, 303.

65 Y. Pellegrin and F. Odobel, *Coord. Chem. Rev.*, 2011, **255**, 2578.

66 S. Kirner, M. Sekita and D. M. Guldi, *Adv. Mater.*, 2014, **26**, 1482.

67 M. E. El-Khouly, S. Fukuzumi and F. D'Souza, *ChemPhysChem*, 2014, **15**, 30.

68 Z.-Y. Gu, J. Park, A. Raiff, Z. Wei and H.-C. Zhou, *ChemCatChem*, 2014, **6**, 67.

69 M. K. Panda, K. Ladomenou and A. G. Coutsolelos, *Coord. Chem. Rev.*, 2012, **256**, 2601.

70 S. Yang, Z. Wu, X. Wan and J. Yan, *Progr. Chem.*, 2011, **23**, 1123.

71 R. Tange, K. Inai, T. Sagawa and S. Yoshikawa, *J. Mater. Res.*, 2011, **26**, 306.

72 C. H. Chang, D. Tiede, J. Tang, U. Smith, J. Norris and M. Schiffer, *FEBS Lett.*, 1986, **205**, 82.

73 J. P. Allen, G. Feher, T. O. Yeates, H. Komiya and D. C. Rees, *Proc. Natl. Acad. Sci. U. S. A.*, 1987, **84**, 5730.

74 J. Deisenhofer and H. Michel, *Science*, 1989, **245**, 1463.

75 S. G. Boxer, R. A. Goldstein, D. J. Lockhart, T. R. Middendorf and L. Takiff, *J. Phys. Chem.*, 1989, **93**, 8280.

76 P. Jordan, P. Fromme, H. T. Witt, O. Klukas, W. Saenger and N. Krauss, *Nature*, 2001, **411**, 909.

77 A. Zouni, H.-T. Witt, J. Kern, P. Fromme, N. Krauss, W. Saenger and P. Orth, *Nature*, 2001, **409**, 739.

78 S.-H. Lee, A. G. Larsen, K. Ohkubo, Z.-L. Cai, J. R. Reimers, S. Fukuzumi and M. J. Crossley, *Chem. Sci.*, 2012, **3**, 257.

79 L. P. Dutton, J. S. Leigh and M. Seibert, *Biochem. Biophys. Res. Commun.*, 1972, **46**, 406.

80 M. C. Thurnauer, J. J. Katz and J. R. Norris, *Proc. Natl. Acad. Sci. U. S. A.*, 1975, **72**, 3270.

81 A. J. Hoff, *Q. Rev. Biophys.*, 1984, **17**, 153.

82 M. Volk, A. Ogrodnik and M. E. Michel-Beyerle, in *Anoxygenic photosynthetic bacteria*, ed. R. E. Blankenship, M. T. Madigan and C. E. Bauer, Kluwer, Dordrecht, 1995, p. 595.

83 A. Rao, P. C. Y. Chow, S. Gélinas, C. W. Schlenker, C. Z. Li, H. L. Yip, A. K. Y. Jen, D. S. Ginger and R. H. Friend, *Nature*, 2013, **500**, 435.

84 M. J. Crossley, P. J. Sintic, R. Walton and J. R. Reimers, *Org. Biomol. Chem.*, 2003, **1**, 2777.

85 M. J. Crossley, P. J. Sintic, J. A. Hutchison and K. P. Ghiggino, *Org. Biomol. Chem.*, 2005, **3**, 852.

86 J. A. Hutchison, P. J. Sintic, M. J. Crossley, T. Nagamura and K. P. Ghiggino, *Phys. Chem. Chem. Phys.*, 2009, **11**, 3478.

87 J. A. Hutchison, P. J. Sintic, P. R. Brotherhood, C. Scholes, I. M. Blake, K. P. Ghiggino and M. J. Crossley, *J. Phys. Chem. C*, 2009, **113**, 11796.

88 J. R. Reimers and N. S. Hush, *J. Am. Chem. Soc.*, 2004, **126**, 4132.

89 M. J. Crossley, L. G. Mackay and A. C. Try, *J. Chem. Soc., Chem. Commun.*, 1995, 1925.

90 P. R. Brotherhood, R. A. S. Wu, P. Turner and M. J. Crossley, *Chem. Commun.*, 2007, 225.

91 D. M. Guldi, *Chem. Commun.*, 2000, 321.

92 H. Imahori, *Bull. Chem. Soc. Jpn.*, 2007, **80**, 621.

93 D. M. Guldi, B. M. Illescas, C. M. Atienza, M. Wielopolski and N. Martin, *Chem. Soc. Rev.*, 2009, **38**, 1587.

94 G. Feher, J. P. Allen, M. Y. Okamura and D. C. Rees, *Nature*, 1989, **339**, 111.

95 M. J. Crossley, A. C. Try and R. Walton, *Tetrahedron Lett.*, 1996, **37**, 6807, submitted.

96 J. R. Reimers, L. E. Hall, M. J. Crossley and N. S. Hush, *J. Phys. Chem. A*, 1999, **103**, 4385.

97 P. J. Sintic, W. E, Z. Ou, J. Shao, J. A. McDonald, Z.-L. Cai, K. M. Kadish, M. J. Crossley and J. R. Reimers, *Phys. Chem. Chem. Phys.*, 2008, **10**, 268. This article was unfortunately duplicated later in 10(4), p. 515–527, their note 10 P7328.

98 W. E, K. M. Kadish, P. J. Sintic, T. Khoury, L. J. Govenlock, Z. Ou, J. Shao, K. Ohkubo, J. R. Reimers, S. Fukuzumi and M. J. Crossley, *J. Phys. Chem. A*, 2008, **112**, 556.

99 D. Curiel, K. Ohkubo, J. R. Reimers, S. Fukuzumi and M. J. Crossley, *Phys. Chem. Chem. Phys.*, 2007, **9**, 5260.

100 K. Sendt, L. A. Johnston, W. A. Hough, M. J. Crossley, N. S. Hush and J. R. Reimers, *J. Am. Chem. Soc.*, 2002, **124**, 9299.

101 M. J. Crossley and P. L. Burn, *J. Chem. Soc., Chem. Commun*, 1991, 1569.

102 J. R. Reimers, T. X. Lü, M. J. Crossley and N. S. Hush, *Chem. Phys. Lett.*, 1996, **256**, 353.

103 T. X. Lü, J. R. Reimers, M. J. Crossley and N. S. Hush, *J. Phys. Chem.*, 1994, **98**, 11878.

104 T. Khoury and M. J. Crossley, *New J. Chem.*, 2009, **33**, 1076.

105 A. Nattestad, Y. Y. Cheng, R. W. MacQueen, T. F. Schulze, F. W. Thompson, A. J. Mozer, B. Fueckel, T. Khoury, M. J. Crossley, K. Lips, G. G. Wallace and T. W. Schmidt, *J. Phys. Chem. Lett.*, 2013, **4**, 2073.

106 T. F. Schulze, J. Czolk, Y. Y. Cheng, B. Fuckel, R. W. MacQueen, T. Khoury, M. J. Crossley, B. Stannowski, K. Lips, U. Lemmer, A. Colsmann and T. W. Schmidt, *J. Phys. Chem. C*, 2012, **116**, 22794.

107 Y. Y. Cheng, B. Fueckel, T. Khoury, R. G. C. R. Clady, M. J. Y. Tayebjee, N. J. Ekins-Daukes, M. J. Crossley and T. W. Schmidt, *J. Phys. Chem. Lett.*, 2010, **1**, 1795.

108 P. C. Dastoor, C. R. McNeill, H. Frohne, C. J. Foster, B. Dean, C. J. Fell, W. J. Belcher, W. M. Campbell, D. L. Officer, I. M. Blake, P. Thordarson, M. J. Crossley, N. S. Hush and J. R. Reimers, *J. Phys. Chem. C*, 2007, **111**, 15415.

109 M. J. Crossley, C. S. Sheehan, T. Khoury, J. R. Reimers and P. J. Sintic, *New J. Chem.*, 2008, **32**, 340.

110 M. J. Crossley and J. A. McDonald, *J. Chem. Soc., Perkin Trans. 1*, 1999, 2429.

111 M. Maggini, G. Scorrano and M. Prato, *J. Am. Chem. Soc.*, 1993, **115**, 9798.

112 T. Drovetskaya, C. A. Reed and P. Boyd, *Tetrahedron Lett.*, 1995, **36**, 7971.

113 M. J. Crossley, A. C. Try and R. Walton, *Tetrahedron Lett.*, 1996, **37**, 6807.

114 M. J. Crossley, T. W. Hambley, L. G. Mackay, A. C. Try and R. Walton, *J. Chem. Soc., Chem. Commun.*, 1995, 1077.

115 M. J. Crossley, L. J. Govenlock and J. K. Prashar, *J. Chem. Soc., Chem. Commun.*, 1995, 2379.

116 Z. Ou, W. E, J. Shao, P. L. Burn, C. S. Sheehan, R. Walton, K. M. Kadish and M. J. Crossley, *J. Porphyrins Phthalocyanines*, 2005, **9**, 142.

117 K. M. Kadish, W. E, P. J. Sintic, Z. Ou, J. Shao, K. Ohkubo, S. Fukuzumi, L. J. Govenlock, J. A. McDonald, A. C. Try, Z.-L. Cai, J. R. Reimers and M. J. Crossley, *J. Phys. Chem. B*, 2007, **111**, 8762.

118 M. J. Crossley and L. A. Johnston, *Chem. Commun*, 2002, 1122.

119 P. Atkins and J. de Paula, *Atkins' Physical Chemistry*, Oxford University Press, Oxford, 9th edn, 2010.

120 M. Gouterman, *J. Mol. Spectrosc.*, 1961, **6**, 138.

121 R. L. Fulton and M. Gouterman, *J. Chem. Phys.*, 1961, **35**, 1059.

122 R. L. Fulton and M. Gouterman, *J. Chem. Phys.*, 1964, **41**, 2280.

123 M. Gouterman, *J. Chem. Phys.*, 1965, **42**, 351.

124 G. Fischer, *Vibronic Coupling*, Academic Press, London, 1984.

125 E. K. L. Yeow, P. J. Sintic, N. M. Cabral, J. N. H. Reek, M. J. Crossley and K. P. Ghiggino, *Phys. Chem. Chem. Phys.*, 2000, **2**, 4281.

126 H. Imahori, K. Tamaki, D. M. Guldi, C. Luo, M. Fujitsuka, O. Ito, Y. Sakata and S. Fukuzumi, *J. Am. Chem. Soc.*, 2001, **123**, 2607.

127 K. Tamaki, H. Imahori, Y. Sakata, Y. Nishimura and I. Yamazaki, *Chem. Commun.*, 1999, 625.

128 S. I. Yang, R. K. Lammi, J. Seth, J. A. Riggs, T. Arai, D. Kim, D. F. Bocian, D. Holten and J. S. Lindsey, *J. Phys. Chem. B*, 1998, **102**, 9426.

129 K. Kilsa, J. Kajanus, J. Martensson and B. Albinsson, *J. Phys. Chem. B*, 1999, **103**, 7329.

130 J. R. Reimers, Z.-L. Cai, R. Kobayashi, M. Rätsep, A. Freiberg and E. Krausz, *Sci. Rep.*, 2013, **3**, 2761.

131 M. Rätsep, Z.-L. Cai, J. R. Reimers and A. Freiberg, *J. Chem. Phys.*, 2011, **134**, 024506.

132 D. M. Guldi, *Chem. Soc. Rev.*, 2002, **31**, 22.

133 D. M. Guldi, *Phys. Chem. Chem. Phys.*, 2007, **9**, 1400.

134 Y. Kashiwagi, K. Ohkubo, J. A. McDonald, I. M. Blake, M. J. Crossley, Y. Araki, O. Ito, H. Imahori and S. Fukuzumi, *Org. Lett.*, 2003, **5**, 2719.

135 K. Ohkubo, P. J. Sintic, N. V. Tkachenko, H. Lemmetyinen, W. E, Z. Ou, J. Shao, K. M. Kadish, M. J. Crossley and S. Fukuzumi, *Chem. Phys.*, 2006, **326**, 3.

136 A. Warshel, *J. Phys. Chem.*, 1982, **86**, 2218.

137 J. R. Reimers, J. M. Hughes and N. S. Hush, *Biochemistry*, 2000, **39**, 16185.