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Tetrazine-based inverse-electron-demand Diels–Alder reaction: a powerful tool for fabrication and functionalization of polymeric materials

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The advent of the click chemistry paradigm has profoundly transformed the design of functional polymeric materials. Among the various reactions in the click chemistry toolkit, the inverse electron demand Diels–Alder (IEDDA) reaction is particularly notable for its rapid kinetics, high specificity, and bioorthogonality. Of the different electron-deficient dienes, the tetrazine (Tz) building block has become the most extensively employed component in the IEDDA reaction-based synthesis and functionalization of polymeric materials. This review begins with an overview of the IEDDA reaction involving Tz, followed by a discussion of examples where the Tz moiety has been utilized as a reactive handle for polymer synthesis and functionalization. We then focus on the application of Tz-based IEDDA reactions in the development of functional hydrogels and surface modifications. The selected examples highlight the diversity and versatility of this highly efficient chemistry. This review aims not only to survey recent progress but also to demonstrate the significant potential of IEDDA chemistry in advancing the design of functional polymeric materials.

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1. Introduction

In recent years, the inverse electron demand Diels–Alder (IEDDA) reaction has witnessed an increasing utilization in the synthesis and functionalization of polymeric materials.¹ While electron-rich dienes and electron-poor dienophiles are employed in the Diels–Alder reaction, reactive counterparts with the opposite electronic demand are used in the IEDDA reaction. Although its usage is relatively new in polymer science, since its inception, the IEDDA reaction has been extensively used in organic synthesis, especially in heterocyclic chemistry.^{2,3} Until recently, normal Diels–Alder and hetero Diels–Alder reactions have been widely used in polymer science to create a wide range of compositionally and architecturally complex macromolecular structures.^{4–13} Compared to the normal Diels–Alder cycloaddition reactions that have been employed over the past several decades in the synthesis and functionalization of polymeric materials, the widespread use of the IEDDA reactions has been limited due to the lack of ready availability of suitable diene and dienophile partners that could be readily incorporated into polymeric materials. Unlike the electron-rich dienes such as butadiene, isoprene,

and furan, among many others, the most widely used electron-deficient diene, namely, the 1,2,4,5-tetrazine (Tz) derivatives, usually required multistep synthesis. For researchers with some synthetic experience, a relatively simpler synthetic approach for obtaining various Tz derivatives was reported in due course.¹⁴ Nonetheless, the attractive attributes of the IEDDA reaction, such as its high efficiency and selectivity, have ensured continued efforts to increase its usage in polymer science. The Tz-based diene reacts with a variety of alkenes and alkynes and generally possesses higher reactivity toward the strained derivatives (Fig. 1). The reactivity of the IEDDA can be tuned by the selection of appropriately substituted dienes and dienophiles. As expected, the presence of electron-withdrawing substituents increases the reactivity of the diene, while the presence of electron-donating groups reduces the reactivity. Thus, using an appropriately substituted Tz derivative, it is possible to dial in a desirable reactivity profile, as necessitated for the intended application. While the electronic nature of the substituents affects the reactivity of the nitrogen-rich diene, the geometric strain of the dienophiles dictates their reactivity.

Although the first report of the IEDDA reaction of Tz was disclosed by Carboni and Lindsey,¹⁵ its usage had been primarily limited to synthetic organic chemistry.^{16,17} Also, most of the early reports of the utilization of the IEDDA reactions, after the introduction of the concept of “click” chemistry by Sharpless and coworkers in 2001,¹⁸ primarily focused on its usage in areas such as bioconjugation and *in vitro* and *in vivo* applications, due to its biorthogonal nature.^{19,20}

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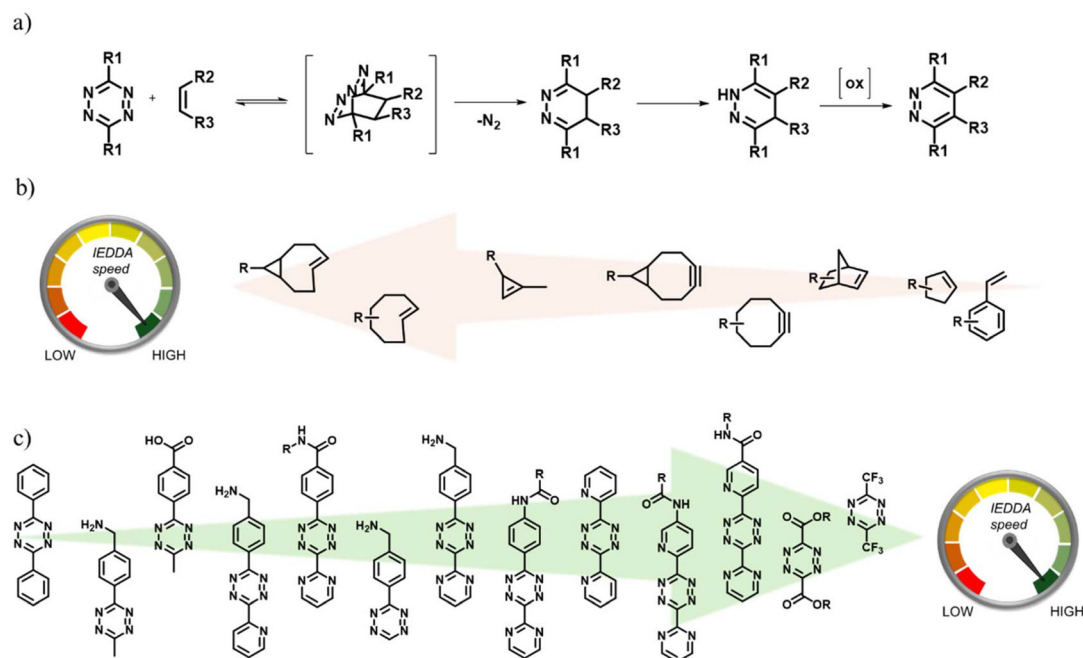


Fig. 1 (a) The IEDDA cycloaddition reaction, (b) commonly employed dienophiles,¹ and (c) typical examples of Tz-based dienes.¹ Adapted with permission from Knall *et al.*¹ Copyright © The Royal Society of Chemistry 2013.

In this review, while we summarize the pertinent milestones in utilizing the IEDDA reaction in polymer science, the versatility this chemistry offers for creating functional polymeric materials is highlighted through various examples (Scheme 1). First, examples of the Tz-based IEDDA in synthesizing and functionalizing polymers are discussed. The section surveys polymers incorporating the Tz group as side-chain or chain end residues and highlights examples of polymers that can undergo efficient post-polymerization functionalization with Tz-containing molecules. This discussion is followed by an illustration of how the IEDDA reaction has been used to fabricate and functionalize crosslinked networks. Finally, examples of functionalization of polymer-coated surfaces using the IEDDA reactions are discussed. While the article would provide a thorough overview of this area to the beginner, it is expected that the survey of examples will promote researchers already working in the design and application of polymers in various areas of materials and biomedical sciences to consider utilizing this highly efficient chemistry.

2. Tetrazine-IEDDA reaction in the synthesis and functionalization of polymers

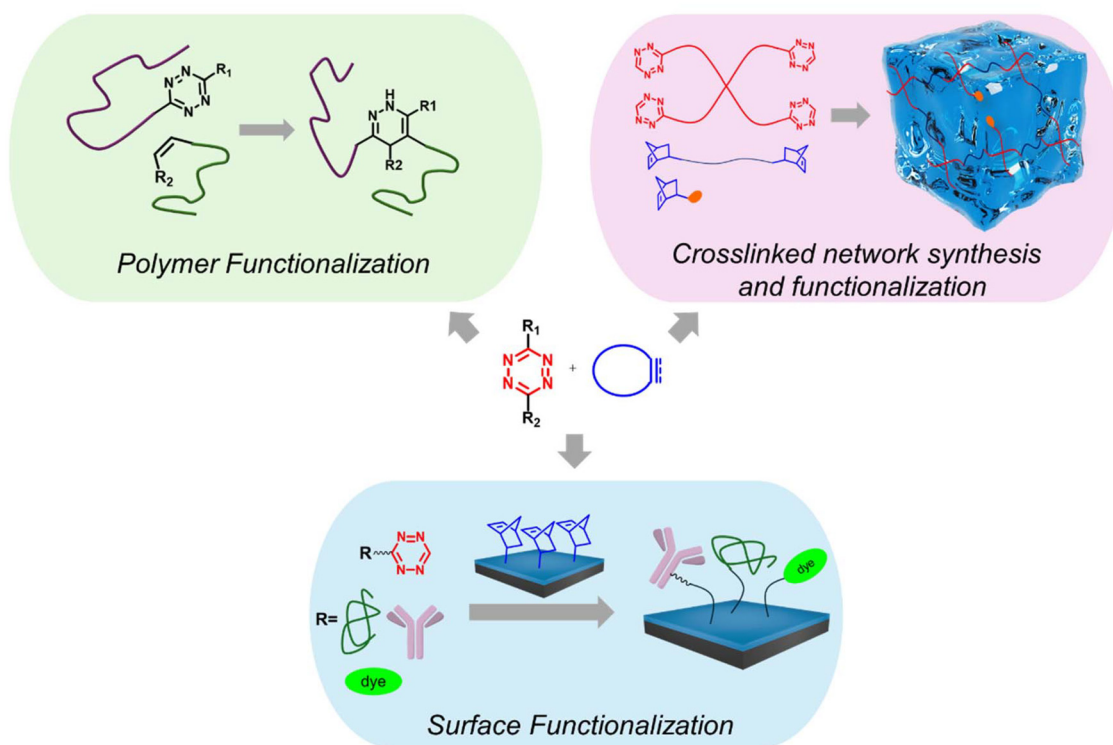
2.1. Polymers bearing Tz groups as reactive handles

Among the several methodologies of obtaining IEDDA-reactive Tz-containing polymers, a much-sought strategy relies on incorporating Tz-bearing small molecules onto existing

polymer scaffolds *via* various post-polymerization reactions.²¹ While synthesis of polymers involving the formation of the Tz group in the backbone are rare, suitably modified Tz group containing monomers has been reported to obtain step-growth polymers.²² In an interesting approach, Sayed and Wiggins utilized a dipolar addition mediated step-growth polymerization to obtain poly(1,4-diphenyl-1,4-dihydro-1,2,4,5-tetrazine-3,6-diyl-2,6-pyridinediyl) from bis-nitrilimine.²³ Other applications include the utilization of Tz functional ROP (ring-opening polymerization) initiators for polymerization of cyclic monomers and chain transfer agents for radical polymerization of methacrylates. Albeit the conjugated polymers with Tz groups on polymer backbones have widespread utilization in the fabrication of photovoltaic, energetic, and electrochromic materials,^{24–26} in limited examples, IEDDA functionalizations were reported on polymers obtained from Tz-based monomers. The following paragraphs summarize several studies that exploited the abovementioned strategies to synthesize polymers with IEDDA-clickable tetrazine groups as reactive handles.

In an early example of post-polymerization modification-based incorporation of Tz unit onto a polymer scaffold, Panek and coworkers reported IEDDA reactions of resin-bound tetrazines with various electron-rich dienophiles.²⁷ The Tz-immobilized polystyrene complexes were prepared by esterifying a hydroxyl-containing Tz analog onto acyl chloride groups of carboxylated polystyrene resin. Subsequently, elaborated azadiene groups were reacted with various electron-rich enamine, dihydrofuran, and dihydropyran dienophiles to yield resin-bound 1,2-diazines with increased regioselectivity. The





Scheme 1 Schematic illustration of functionalization of polymers, synthesis, and modification of crosslinked networks, as well as functionalization of solid surfaces via IEDDA "click" reaction.

obtained cycloadducts were efficiently removed from the resins by basic hydrolysis, granting the practical nature of the IEDDA reaction on solid supports.

Bioorthogonal methodologies for preparing polymeric conjugates *in vivo* have recently gained intense interest since this class of reactions proceeds with rapid kinetics and high chemical selectivity in biological environments without interfering with biological processes.^{28,29} The bioorthogonal nature of these chemical reactions minimizes the side reactions towards endogenous functional groups present in the cellular systems. Especially in the last decade, bioorthogonal reactions have played pivotal roles in preparing polymeric conjugates that serve in biomedical imaging, genetic code expansion, drug delivery, metabolic engineering, and drug target identification.³⁰ Of these only a handful of reactions, such as the IEDDA-based transformations have been the method of choice due to their unmatched reaction kinetics, superior orthogonality, and biocompatibility.^{1,20}

In an impressive example, to demonstrate *in vivo* labeling in the tumor microenvironment, Weissleder, and coworkers have used Tz-modified and ¹⁸F-labeled dextran polymers ($M_n = 10\,000\text{ g mol}^{-1}$) in targeting A33 glycoproteins, which are over-expressed in colorectal cancers (Fig. 2a).³¹ In this study, amino-dextran polymers were partially reacted with fluorescent succinimidyl ester fluorophore, and subsequently, the remaining amine groups were modified by adding Tz-NHS. The conjugate labeling was done by reacting the Tz groups with a [¹⁸F]

labeled *trans*-cyclooctene (TCO) derivative. The *in vivo* conjugation studies performed on LS174T tumor xenografts, which were modified on cell surfaces with excess anti-A33 TCO monoclonal antibodies, revealed that the dextran polymers were able to accumulate in the TCO-tagged tumors. In a later study, Meyer *et al.* conceptualized a TCO scavenger approach based on the utilization of Tz-modified small dextran polymers (DP-Tz) to mask the remaining TCO-moieties on the circulating monoclonal antibodies which did not interact with antigen-expressing tumor tissue (Fig. 2b).³² The pre-targeting experiments employing DP-Tz demonstrated significantly improved tumor delineation, reduced uptake in vital organs, and enhanced PET image quality.

In an elegant study to demonstrate the potential of bioorthogonal tetrazine ligation chemistry in preparation for smart cancer theranostics, Wang and coworkers designed a pH-responsive polymer containing tertiary amine and Tz groups at side chains.³³ In the synthesis step, the Tz groups were installed onto the side chains of an atom transfer radical polymerization (ATRP)-generated copolymer ($M_n = 15\text{ kDa}$) via post-polymerization amidation reaction, with a yield of 98%. These polymers were shown to self-assemble into micelles under neutral conditions (pH 7.4), concurrently enclosing Tz groups and shielding their activities. In the acidic tumor microenvironment (pH 6.5), these micelles collapsed rapidly, therefore promoting the accessibility of Tz groups. A polymeric prodrug (CyPVE), a vinyl ether functional hemicyanine dye



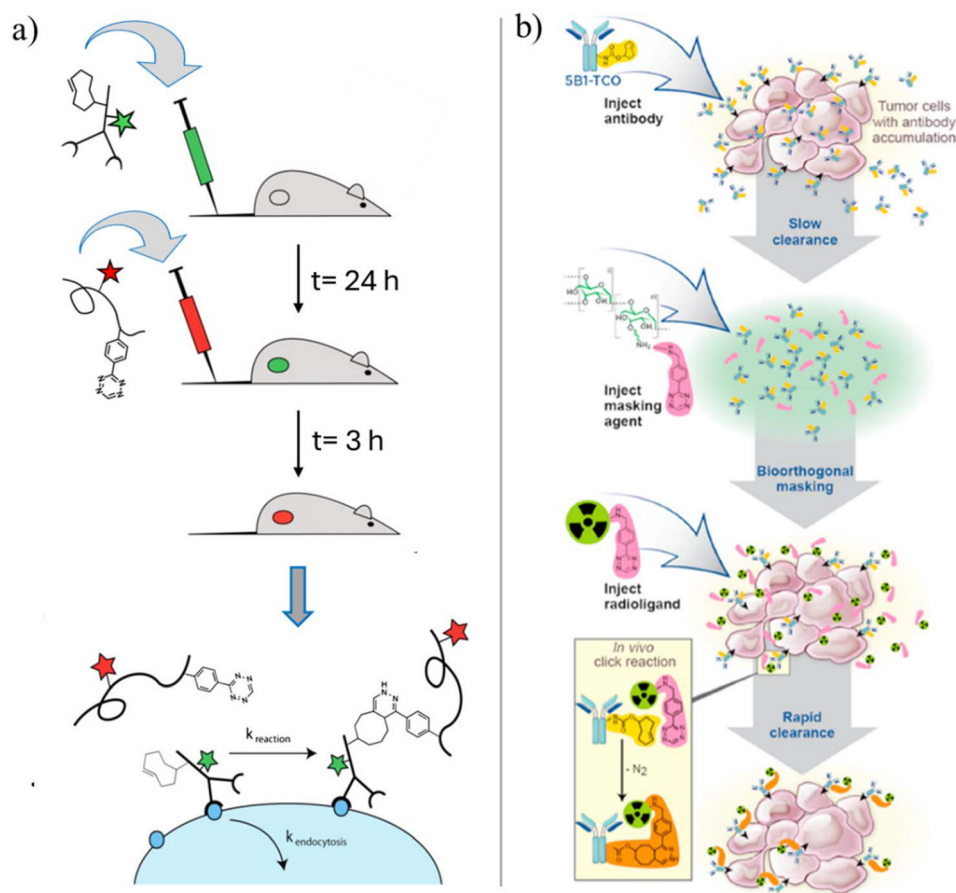


Fig. 2 (a) Displaying of *in vivo* bioorthogonal reaction and *in vivo* multistep delivery of imaging agent. First, a slow-clearing targeting agent (green) is administered and is given 24 hours for localization. This is followed by injecting a low molecular weight secondary imaging agent (red), which gets localized through an IEDDA reaction with the targeting agent. Adapted with permission from Devaraj *et al.*³¹ Copyright (2012) National Academy of Sciences, and (b) illustration of the IEDDA-based small molecule radioligand labeling of TCO-functional antibodies following injection of the masking agent before the radioligand conjugation. Adapted with permission from Meyer *et al.*³² Copyright © 2018, American Chemical Society.

(CyPOH) with a poly(ethylene glycol) (PEG) chain was utilized to impart a bioorthogonal prodrug activation. Under acidic microenvironment conditions, Tz functional collapsed micelles were reacted with vinyl ether groups of CypVE, leading to ligation-elimination of vinyl ether groups. The phototoxicity and NIR fluorescence properties were activated after removing the vinyl ether group from latent phenolic hemicyanine dye. *In vivo*, experiments on multicellular spheroids (MCSs) derived from 4T1 cells revealed that, while the fluorescence intensity of dye conjugate was only lit up around the periphery at pH 7.4, it could reach a 100 mm depth at pH 6.5 conditions. These findings were reported as a potential application of bioorthogonal Tz IEDDA chemistry to prepare smart cancer theranostics.

Fox, Jia, and coworkers employed the post-polymerization installation of tetrazine groups at chain ends of PEG to prepare hybrid multiblock copolymers based on complementary bifunctional monomers.³⁴ The interfacial copolymerization *via* Tz ligation was carried out using hydrophilic bis-Tz monomers with PEG spacers and a bis-TCO monomer (Fig. 3).

The results showed that the judicious selection of IEDDA-*clickable* monomers allows the synthesis of high molecular weight polymers at the immiscible solvent interface and enables incorporating functional peptides into the multiblock copolymer structure to tune the polymer properties. In a subsequent study, the Fox group employed bis Tz end-functional PEG polymer ($M_n = 7500 \text{ g mol}^{-1}$) and a bis-TCO containing monomer tethering dihydrotetrazine unit at the center for interfacial polymerization.³⁵ After obtaining fibrous copolymers by continuous pulling, the latent Tz functionalities on polymer side chains were converted to IEDDA-*clickable* Tz groups by photooxidation. Several peptidic ligands and imaging agents were shown to be efficiently conjugated to the synthesized polymers *via* post-polymerization modification with TCO derivatives. Selectively, RGD-TCO and RGD-TCO-attached polymers displayed promoted cell adhesion and contact guidance.

Maynard and coworkers reported the synthesis of bis-Tz end-functional poly(*N*-isopropylacrylamide) (pNIPAAm) and PEG polymers that underwent Tz ligation with a TCO-func-



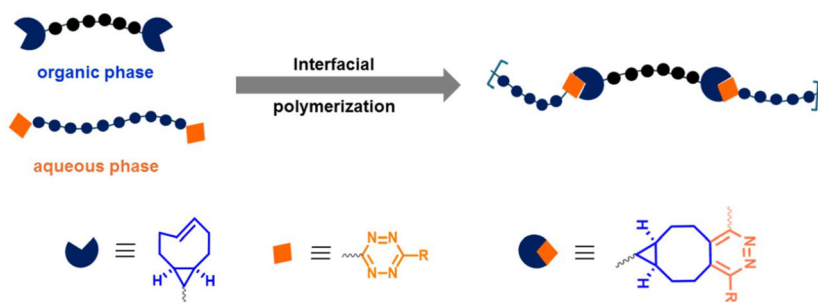


Fig. 3 Synthesis of multiblock copolymer fibers through interfacial bioorthogonal polymerization.³⁴

tionalized lysozyme to generate protein dimers.³⁶ Telechelic Tz functional polymer was obtained by reversible addition–fragmentation chain-transfer (RAFT) polymerization of NIPAAm monomer with a bis carboxylic acid containing chain transfer agent and subsequent post-polymerization esterification of a hydroxyl bearing Tz molecule to carboxylic end groups. In the study, the protein homodimerization of Tz telechelic polymers was compared with a bis-maleimide end-functional pNIPAAm polymer ($M_n = 2000 \text{ g mol}^{-1}$). Accordingly, the IEDDA-based “click” reaction demonstrated higher protein conjugation efficiency and dimer formation for both telechelic Tz functional pNIPAAm (38%) and PEG (37%) polymers in a 1 h conjugation duration, compared to thiol-maleimide Michael addition (<1%). In a screening study, O’Reilly and coworkers extensively studied several important bioconjugation reactions in the organic solvent medium for the efficient covalent conjugation of DNA to one chain end of pNIPAAm.³⁷ Among the amide coupling, thiol–ene, and Tz ligation reactions, the IEDDA reaction between Tz and norbornene (NB) groups complementarily found on reacting molecules was found to be the most effective ligation tool to give DNA-polymer conjugates.

One of the critical drawbacks of the post-polymerization installation of Tz groups onto polymers is the requirement for a separate synthesis of reactive polymer and anchoring Tz derivative. In most cases, cumbersome reaction procedures, several side reactions, and non-orthogonal reacting functional groups can lead to tedious purification steps and overall diminished reaction yields. In an attempt to directly install the Tz functionalities onto the side chains of poly(*N*-isopropylacrylamide-*co*-acrylonitrile) (p(NIPAAm-*co*-AN) and p(PEGMA-*co*-acrylonitrile) copolymers, Kahveci and coworkers have employed an elegant one-pot two-step post-polymerization modification approach.³⁸ In the first step, side chain nitrile groups of copolymers were cross-coupled with an aromatic nitrile-containing molecule in Lewis acid-catalyzed conditions. In the following step, NaNO_2 oxidized the 1,4-dihydropyridazine groups to Tz functionalities in acidic conditions. Modification efficiency determination *via* ^1H NMR for p(NIPAAm-*co*-AN) and p(PEGMA-*co*-acrylonitrile) copolymers revealed 7.9% and 17.9% transformation yields (based on present nitrile groups on copolymers), respectively. It was also shown that Tz-modified copolymers could be efficiently

functionalized with a TCO-containing small molecule in less than a minute. In another study, the same group reported the preparation of three-arm star block copolymers *via* Tz ligation.³⁹ First, three-arm polylactide (PLA) polymers with Tz end groups were prepared by ring opening polymerization of *D*, *L*-lactide monomer, and subsequent Tz end group installation. Then, a mono methoxy PEG polymer with a TCO end group was reacted with a three-arm Tz functional PLA polymer under exact stoichiometric conditions. It was demonstrated that a high polymer–polymer coupling efficiency (up to 95.8%) was achieved in about an hour, leading to 3-arm star PLA-*b*-PEG copolymers ($M_n = 23\,400 \text{ g mol}^{-1}$). The post-polymerization introduction of Tz groups onto poly(styrene-*co*-acrylonitrile) polymer and subsequent TCO functionalization was also reported in this context.⁴⁰

The importance of synthesizing functional polymers that allow site-specific attachment of desired molecules is prevalent in preparing polymeric conjugates for several biomedical applications. For example, in the preparation of polymer–protein conjugates, site-specific conjugation of proteins to polymers is imperative because the interaction of bioconjugates with target proteins, receptors, or cells is strongly related to the size, shape, and binding ligand density.^{41–43} Precise and well-defined polymeric systems are also crucial for quantitatively assessing such conjugates. In the preparation of polymer bioconjugates, end functional polymers provide both the opportunities of controlling position and quantifying attached biomolecules.^{44,45} Synthesis of such polymers are generally performed by choosing suitable functional initiators or chain transfer agents that allow the covalent attachment of the desired molecules.

To synthesize polymers containing the Tz groups at polymer chain ends, a preferred strategy is to use functional initiators to start the polymerization. The judicious choice of an appropriate initiator allows the installation of IEDDA-*clickable* Tz units at the specific end groups of polymers. This concept was initially reported by Hansell *et al.* to synthesize Tz-terminated PEG, poly(δ -valerolactone) (PVL) for IEDDA-based conjugation to NB-terminated polymers.⁴⁶ A Tz-containing alcohol was utilized with an organo-phosphorus catalyst, which yielded a polymer with 1.7 kDa molecular weight and 1.27 dispersity, using ring opening polymerization (ROP). In a



approaches include the employment of dihalide functional Tz monomers in nucleophilic aromatic substitution and metal-mediated cross-coupling polymerizations. Post-polymerization modification of such Tz group containing polymers *via* IEDDA-based ligation allows desired functionalization. In a recent study, Adronov and coworkers employed a bisbromofuran group containing Tz compound and a fluorene diboronate-based monomer to prepare soluble poly(Tz) polymers.⁵³ The polymerization reactions employed a palladium-catalyzed Suzuki polycondensation reaction, which yielded high molecular weight conjugated polymers containing reactive Tz units along the backbones. Quantitative functionalization of poly(Tz)s with a series of TCO derivatives and subsequent oxidation resulted in corresponding conjugated poly(pyridazine) polymers. The analytic measurements *via* absorption and emission studies revealed that the functionalized polymers display substantial hypsochromic shifts in absorption and increased emission intensity relative to the parent poly(Tz) polymer. In another study that employs a dichloro functional Tz monomer, Carrillo and coworkers fabricated polymer microspheres that display tailored dynamic covalent networks.⁵⁴ Polymers were obtained by nucleophilic aromatic substituting chloro groups of Tz monomer with either multiphenol or multi-thiol-containing compounds (70% to quantitative yield). Taking advantage of dynamic covalent chemistry reactions based on the nucleophilic aromatic substitution of Tz,⁵⁵ the authors were able to prepare degradable and recyclable networks. IEDDA-based conjugation of Tz networks with a dicarboxylic acid functional NB derivative resulted in the tuning of polymer properties, such as water solubility. Notably, these networks were concomitantly locked to the exchange reaction after Tz ligation, thus stabilizing the polymer against depolymerization. Recently, the post-polymerization modification of the main-chain Tz group containing copolymers with aryl alkynes was reported.⁵⁶

2.2 Polymer functionalization using Tz-containing functional molecules

The applications of IEDDA reaction in the fabrication and functionalization of polymeric materials principally stem from the course of the reaction that proceeds on “click” chemistry principles. On the other hand, for efficient implementation of this cycloaddition reaction on a macromolecular construct, the electronic natures of diene and dienophile functionalities present on the counterpart reacting moieties have the utmost importance in attaining high coupling efficiencies and fast reaction kinetics. In the case of IEDDA-based Tz ligation with electron-rich dienophiles, the latter reaction components are usually chosen from strained alkenes or vinylic compounds neighboring electron-donating groups. Since Tz groups have limited compatibility with nucleophiles, radical sources, redox species, and metal donors that might impede the retention of Tz structure during polymerization,⁵⁷ a commonly practiced approach for IEDDA-based polymer functionalization relies on the post-polymerization attachment of Tz containing small molecules onto polymeric scaffolds that bear counterpart

reacting dienophiles. Among the various IEDDA-clickable groups that can be orthogonally conjugated with Tz units, strained NB alkenes have widespread utilization in modifying polymeric materials. The suitability of NBs in thermal and UV-initiated thiol-ene radical additions,^{58,59} and their archetypal nature in ring-opening metathesis polymerizations⁶⁰ is prominent. Due to their highly strained ring structure, NB derivatives are ideal reacting partners in the IEDDA reactions.

Building on the ROP initiator strategy of placing NB groups at polymer chain ends, Dove and coworkers demonstrated post-polymerization functionalization of NB-modified polylactic acid *via* IEDDA “click” reaction.⁶¹ The ligation reaction of NB units with 3,6-di-2-pyridyl-1,2,4,5-Tz resulted in complete end group modification without degradation of the polymer backbone. In the same study, side chain NB functional lactide polymer poly(spiro[6-methyl-1,4-dioxane-2,5-dione-3,20-bicyclo-[2.2.1]hept[5]ene]) was also synthesized and modified similarly with Tz-based compound. In another study, Dove, O'Reilly, and coworkers adopted a similar strategy to obtain NB-functional poly(carbonate) scaffolds.⁶² After obtaining degradable polymers *via* organo-catalyzed ROP of NB-containing cyclic carbonate monomer, post-polymerization modification of pendent NB units was successfully conducted by various complementary reactive groups, including azides *via* a 1,3-dipolar cycloaddition, Tzs in IEDDA reaction and thiols *via* radical thiol-ene reaction. The orthogonal nature of these chemistries was also demonstrated by one-pot sequential modification of NB functional polymer. Synthesis and Tz ligation modification of degradable polyesters with NB functional side chains was reported within this scope.⁶³ Other than the NB partner, Tz-based IEDDA reaction was also successfully implemented in the functionalization of allyl ether side groups containing anionic ROP copolymers.⁶⁴

Due to its modular, fast, efficient, and biorthogonal nature, Tz-IEDDA chemistry is attractive for fabricating polymeric bioconjugates. In an early study reported by Bawendi and coworkers, polymeric imidazole ligands were prepared by RAFT polymerization and used to coat quantum dots (QDs) through ligand exchange.⁶⁵ NB groups were subsequently installed on the surface of QDs *via* amidation reaction. To illustrate the application of NB-decorated QDs for live-cell imaging, they successfully targeted A431 human carcinoma cells, previously decorated with a Tz-modified epidermal growth factor. In another work by Zhang *et al.*, side-chain NB functional polymers with desired molecular weights (7–30 kDa) with dispersities between 1.2–1.8 were synthesized by RAFT polymerization of a NB containing acrylamide monomer (Fig. 5).⁶⁶ In polymerization, metal affinity chromatography deliberately employed a histidine-tagged chain transfer agent to allow straightforward purification of later-prepared polymer-antibody conjugates. The polymer bearing reactive NB groups and a histidine tag was conjugated to antibody Herceptin *via* amidation to prepare a polymeric bioconjugate amenable to Tz ligation. After the IEDDA-based attachment of a Tz-containing fluorescent dye molecule to the bioconjugate, the polymer-antibody scaffold provided an 83-fold increase in fluorescence



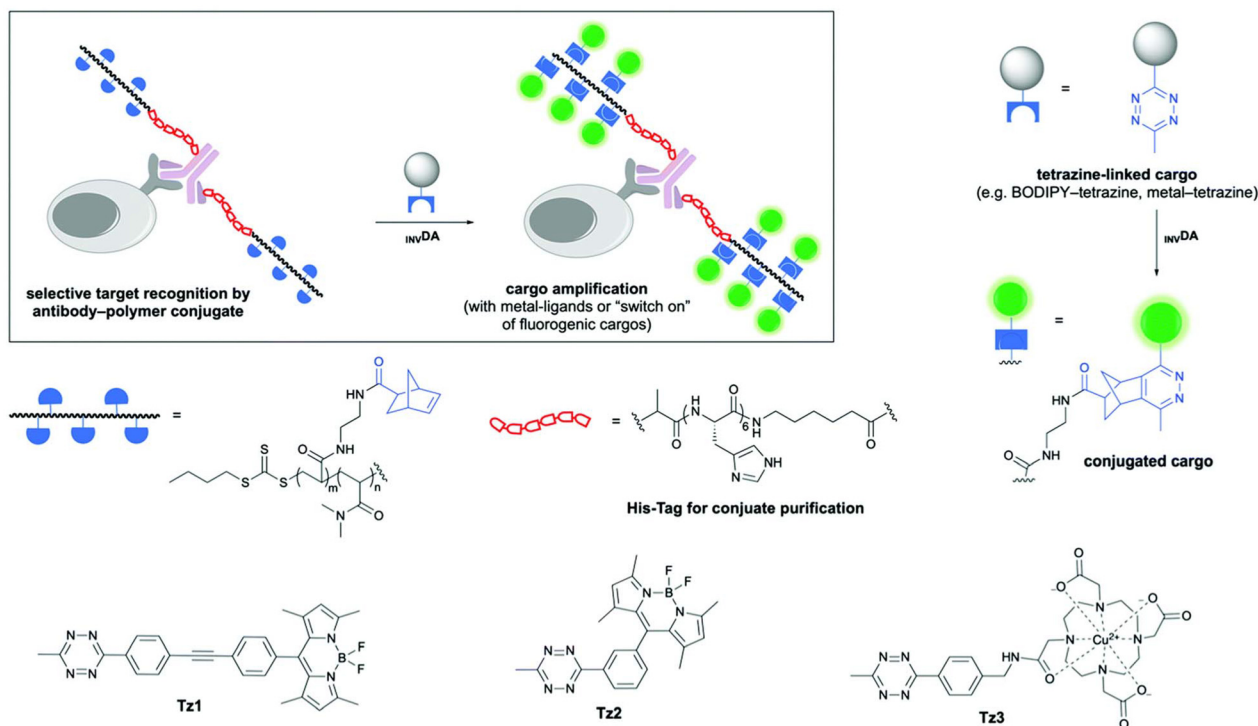


Fig. 5 Preparation of Tz-IEDDA reaction-mediated polymer-protein conjugates for fluorescent signal amplification. Adapted with permission from Zhang *et al.*⁶⁶ Copyright © 2020, The Royal Society of Chemistry.

intensity compared to the control antibody labeled with a standard small molecule fluorophore. In a recent study, biorthogonal Tz-NB and aminoxy-aldehyde ligation reactions were concomitantly employed in the *in situ* generation of synthetic dendrimers on live cells.⁶⁷ The authors achieved cancer cell targeting on live tumor tissue by growing the dendritic structure on antibody Herceptin.

The application of IEDDA-*clickable* side chain NB functional polymers in the preparation of polymeric micelles and single-chain polymer nanoparticles (SCNPs) was investigated by the O'Reilly group.^{68,69} In the self-assembly generated formation of micellar scaffolds, a RAFT-mediated amphiphilic diblock copolymer carrying two orthogonal “click”-compatible functionalities in the polystyrene block core and PEG-based shell was utilized. Upon the formation of micellar structures in water, functionalizations of NB groups in the core structure and terminal alkynes in the shell of aggregates have been carried out by Tz-NB chemistry and copper-catalyzed azide-alkyne reaction. It was shown that this single micellar scaffold can be used either in separate core-shell functionalizations or in a one-pot sequential manner that allows both core- and shell functionalization using two orthogonal “click” reactions. Later, the same group employed the Tz-NB ligation reaction to prepare single-chain polymer nanoparticles. Using polystyrene polymers decorated with pendent NB groups and a bifunctional Tz crosslinker, SCNPs of diameters below 10 nm were prepared. In an elegant work by Li and coworkers, IEDDA-based Tz-NB chemistry was also employed to prepare

polymeric micelles from bacteria templating.⁷⁰ In this study, either doxorubicin (DOX) or α -tocopheryl succinate (TOS) anti-cancer agents were attached to the one-chain end of a PEG polymer through redox-responsive disulfide linkers. The DOX and TOS loading levels in MDOX and MTOS micelles were approximately 18.9% and 14.9%. A NB derivative was installed through an acid-labile linker to the other end of polymers. These polymeric platforms were later IEDDA-conjugated to *Escherichia coli* Nissle 1917 (EcN) bacterial carrier decorated with Tz groups on the cellular surface. Upon introducing the bacterial microbot drug carrier system to the slightly acidic tumor microenvironment, drug-loaded copolymers were released from the bacterial surface and concomitantly arranged into hybrid micellar structures. It was demonstrated that after intravenous administration of bacterial carrier system into tumor-bearing mice, selective tumor accumulation and prolonged retention of synergistic anticancer drugs at target tissue were achieved.

The Tz-based IEDDA chemistry has been recently employed to end chain norbornadiene group-containing polymers, in which, in this case, the ligation chemistry allowed the unveiling of latent reactive cyclopentadiene functionalities. In a report by Hawker, De Alaniz, and coworkers, norbornadiene groups, previously installed at the chains ends of polymers during polymerization *via* ROP, ATRP, and RAFT processes, were employed in polymer functionalization and block copolymer synthesis through one-pot, sequential Tz-NB IEDDA and cyclopentadiene-maleimide Diels-Alder reactions (Fig. 6).⁷¹ It



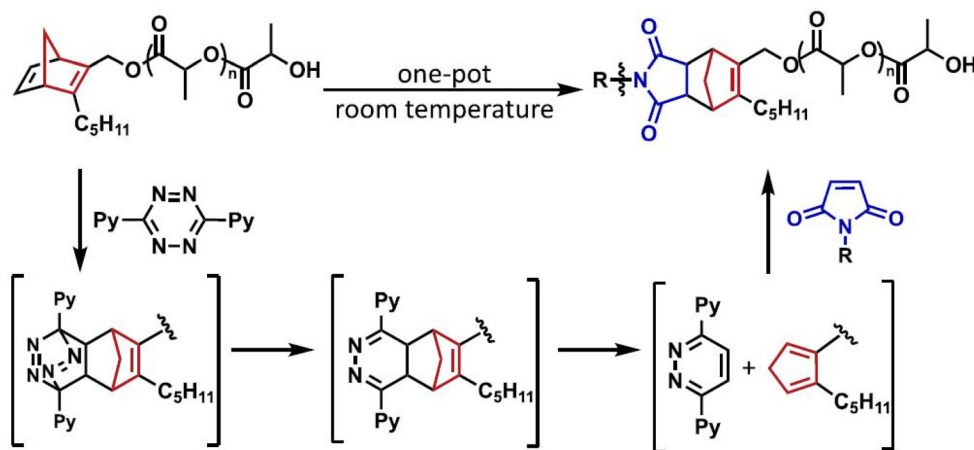


Fig. 6 One-pot functionalization of norbornadiene end-functional polymer with maleimides. Adapted with permission from Amant *et al.*⁷¹ Copyright © 2019, American Chemical Society.

was shown that initial Tz cycloaddition with norbornadienes reveals a cyclopentadiene derivative selectively at polymer chain end, which can subsequently participate in Diels–Alder cycloaddition with maleimide. In a later work, the same group employed this cascade “click” modification methodology to the norbornadiene end group containing polymers in which the norbornadiene groups were installed at polymer chains *via* post-polymerization modification.⁷²

Besides NB, one of the most potent reactive dienophiles is TCO and its derivatives for the Tz-based IEDDA reactions. The ligation reactions between TCO and Tz groups could reach the fastest known reaction kinetics (rate constants up to $10^6 \text{ M}^{-1} \text{ s}^{-1}$), making them exceptionally useful tools for bioconjugation studies.^{73,74} Important drawbacks of TCO compounds, however, are their low commercial availability and complex synthesis.⁷⁵ The applications of Tz-TCO ligation chemistry in polymeric material synthesis and functionalization predominantly include preparing polymer bioconjugates suited to cell imaging, protein modification, and cell targeting.

Herth, Radchenko and coworkers reported radiolabeling of polyglutamic acid (pGlu) ($M_n = 23\,300 \text{ g mol}^{-1}$) with ^{177}Lu and ^{225}Ac radioligands using pGlu modified with pendant TCO groups and radiolabeled Tz chelates.⁷⁶ The method yielded conjugates with excellent stability in human plasma. In another study, Zeglis and coworkers combined two bioorthogonal chemistries, strain-promoted azide–alkyne cycloaddition (SPAAC) and Tz-TCO ligation, to produce antibody-attached dendrimer scaffolds, that can be used in signal amplification in pre-targeted positron-emission tomography (PET) imaging.⁷⁷ After site-specific SPAAC conjugation of peripheral TCO-functional dendrimer to huA33 antibody, *in vivo* tumor targeting was carried out using a murine model of colorectal carcinoma. The labeling of resulting TCO-bearing immunoconjugates by a Tz-modified radioligand demonstrated highly enhanced PET imaging efficiency compared to an analogous dendrimer lacking immunoconjugate. By Herth and coworkers, the pre-targeted imaging strategy was extended to the

TCO polypeptide-*g*-polypeptoid polymers that were utilized in nuclear imaging.⁷⁸ After tumor accumulation of the ROP-generated TCO-functional ‘peptobrush’ (polyglutamic acid-*g*-poly-sarcosine) polymers, biorthogonal conjugation of a Tz modified [^{111}In] contrast agent provided efficient tumor radiolabeling. In a follow-up study, the Herth group employed TCO functional peptobrush polymers for radiolabeling tumor tissues with [^{111}C]–Tz markers.⁷⁹ A pretargeting strategy developed for the passive delivery of side chain TCO-functional RAFT star polymers to tumor tissue was reported by Goos and coworkers.⁸⁰ In the study, while star-shaped polymeric carriers loaded with Gd-based MRI components provided passive tumor targeting, a Tz functional [^{18}F] radiolabel conjugated to the TCO groups allowed nuclear imaging.

Xu *et al.* have broadened the applications of Tz-TCO ligation chemistry to the site-selective protein modification established on disulfide rebridging strategy.⁸¹ A Tz-based disulfide reagent was employed for site-selective installation of the Tz group into both disulfide-containing hormone somatostatin and the antigen-binding fragment of human immunoglobulin G. A set of TCO-containing substrates such as dyes, polymers, and enzymes were conjugated to the Tz modified proteins. The fast and efficient conjugation results demonstrated that IEDDA reaction based on Tz and TCO substrates could be concurrently applied with other bioconjugation tools to prepare the bioconjugates in a site-selective manner.

Although, due to its bioorthogonal nature and fast reaction kinetics, Tz-IEDDA chemistry has found extensive utilization in preparing polymer bioconjugates, the applications have also been directed to the covalent modification of industrial polymers. These polymers are commercial and produced on massive scales; thus, their covalent modifications *via* highly efficient “click” reactions have attracted attention to preparing polymers with unique properties.^{82–84} In an initial report, Bagge *et al.* demonstrated the modification of polybutadienes with 3,6-dichloro-1,2,4,5-Tz.⁸⁵ Polymeric foams with enhanced chain rigidities were obtained by the liberation of nitrogen gas



during the cycloaddition process. The incorporation of 1,4-dihydropyridazine group to the polymers also enhanced the antioxidant properties of polymers as deduced by the free radical inhibition experiments. Similarly, Li, Wu, and co-workers applied 3,6-di(2-pyridyl)-1,2,4,5-Tz to modify and introduce nitrogen-containing groups into styrene-butadiene rubber chains covalently.⁸⁶ These metal chelating groups were further demonstrated to coordinate Cu²⁺ ions, allowing fabrication of mechanically robust and reversibly processable materials. By the same groups, the same chemistry was applied to nitrile-butadiene rubber to obtain tough, recyclable and healable polymers.⁸⁷

3. Tetrazine-IEDDA mediated synthesis and functionalization of crosslinked networks

The distinguished characteristics of IEDDA reactions, such as fast reaction kinetics, high monomer conversions, and catalyst-free conditions, make them ideal reaction tools for fabricating crosslinked homogenous polymer networks.^{88–91} The Tz-IEDDA reaction has been utilized in fabricating hydrogels, nanogels, organogels, and metal-crosslinked polymer scaffolds for diverse applications.⁹² Particularly, the biorthogonal and biocompatible nature of the Tz-IEDDA reaction serves utility for fabricating hydrogels for various biomedical applications, including cell encapsulation, bioconjugation, tissue engineering, and sustained drug/protein delivery.

One of the earlier examples of Tz-IEDDA reaction in hydrogel synthesis was reported by Johnson and coworkers in which A2 + B3 type cycloaddition of bisnorbornene end functional PEG ($M_n = 6$ kDa and 10 kDa) and a three-Tz containing crosslinker resulted in gel networks.⁹³ The study demonstrated that the Tz-IEDDA reaction could provide an efficient and additive-free crosslinking process by complementing the reaction procedure with other “click” chemistry methodologies. Besides several hydrogelation systems that employed end group functional polymer precursors,⁹⁴ Tz-IEDDA reaction has also been exemplified in smooth crosslinking hydrogelation using side chain Tz functional polymers.⁹⁵ The rapid reaction kinetics of Tz-IEDDA reaction has been extended to highly efficient interfacial crosslinking at oil-water^{96–98} and liquid-gel interface,^{99,100} as well as annealing hydrogel microparticles.^{101–105} Although Tz-IEDDA has primarily served in preparation of cast and injectable hydrogel-based materials, this chemistry also has the edge on synthesizing patterned hydrogels,^{106,107} nanogels^{108–110} and cryogels.¹¹¹

The orthogonal nature of Tz-IEDDA reaction to various other “click” chemistry-based methodologies benefits dual-click crosslinking for generating functional networks. Along with the IEDDA reaction with Tzs, NBs, and strained alkynes are reaction partners in thiol-ene and thiol-yne-based synthetic approaches. In the case of targeting mechanically robust 3D structures, which is especially desirable in biomedical

applications, the dual-click crosslinking strategy might allow the generation of rigid and stiff crosslinked networks. Dove and coworkers exploited this approach to fabricate high-compressive strength hydrogels with two interpenetrating networks in the structure.¹¹² In a one-pot tandem “click” procedure that combines all the reaction components, the initial formation of the loose network was achieved by Tz-IEDDA reaction of NB-functionalized chitosan with a linear PEG-Tz crosslinker (Fig. 7). Subsequently, the dense network was constructed by thiol-yne reaction of a 4-arm PEG-tetra alkyne with a linear PEG-dithiol. It was demonstrated that these mechanically strong and biocompatible hydrogels are ideal platforms for encapsulation of human cells. In another report that shares a similar dual crosslinking methodology, photo-initiated thiol-NB addition was subsequently followed by Tz-IEDDA reaction to fabricate dual crosslinked networks.¹¹³ Incorporation of a mono Tz end functional polymer into thiol-NB partially crosslinked networks also enhanced the network stiffening through supramolecular interactions.¹¹⁴

While the Tz-IEDDA reaction offers an attractive way to prepare biorthogonal hydrogels, the susceptibility of the Tz group towards different chemical environments should be considered during precursor preparation and cross-linking processes. For example, Tzs with electron-withdrawing carboxylate substitutions are not stable in an aqueous environment despite their high reactivity with dienophiles.¹¹⁵ Tzs might also display cross-reactivity towards free thiol groups, which are abundant in biological media.³⁰ To address such challenges, a preferred strategy is the on-demand generation of Tz groups by oxidation of dihydrotetrazine (dHTz) precursors. Forsythe, Truong, and coworkers utilized *in situ* generation of Tz to obtain hydrogels using light-triggered chemistry.¹¹⁶ Activation of dHTz, which is more stable against reductants, in the presence of methylene blue and red-light (625 nm) furnishes hydrogel *via* the *in situ* IEDDA reaction (Fig. 8). Impressively, the gelation occurred behind a dermal tissue model with a thickness of 1 cm. The non-toxic nature allows the encapsulation of hMSC within the gel matrix. The same group also reported the fabrication of PEG-gelatin hydrogels using an enzyme-activated Tz-IEDDA reaction. The addition of horse radish peroxidase enzyme to an aqueous solution of 4-arm PEG-dHTz ($M_n = 10$ kDa) and gelatin-NB induced gelation. Obtained hydrogels were highly stable under physiological conditions, allowing 3D culture of human mesenchymal stem cells (hMSCs) for 32 days.¹¹⁷ In another study, Fox and co-workers reported an efficient method for mild oxidation of dihydrogen Tz to Tz using silicon-rhodamine dyes as cyto-compatible far-red (660 nm) photocatalysts.¹¹⁸

The structural tailoring of the network matrix can be conveniently carried out to induce hydrolytic stability or degradability in Tz-IEDDA hydrogels. Goepferich and coworkers prepared hydrogels that show long-term hydrolytic stability and controlled protein release over time to obtain structurally stable crosslinked networks or promote hydrolytic matrix degradation over time.¹¹⁹ PEG-based hydrogels prepared using 8-arm poly(ethylene glycol)s ($M_n = 10, 20, \text{ or } 40$ kDa) of Tz and



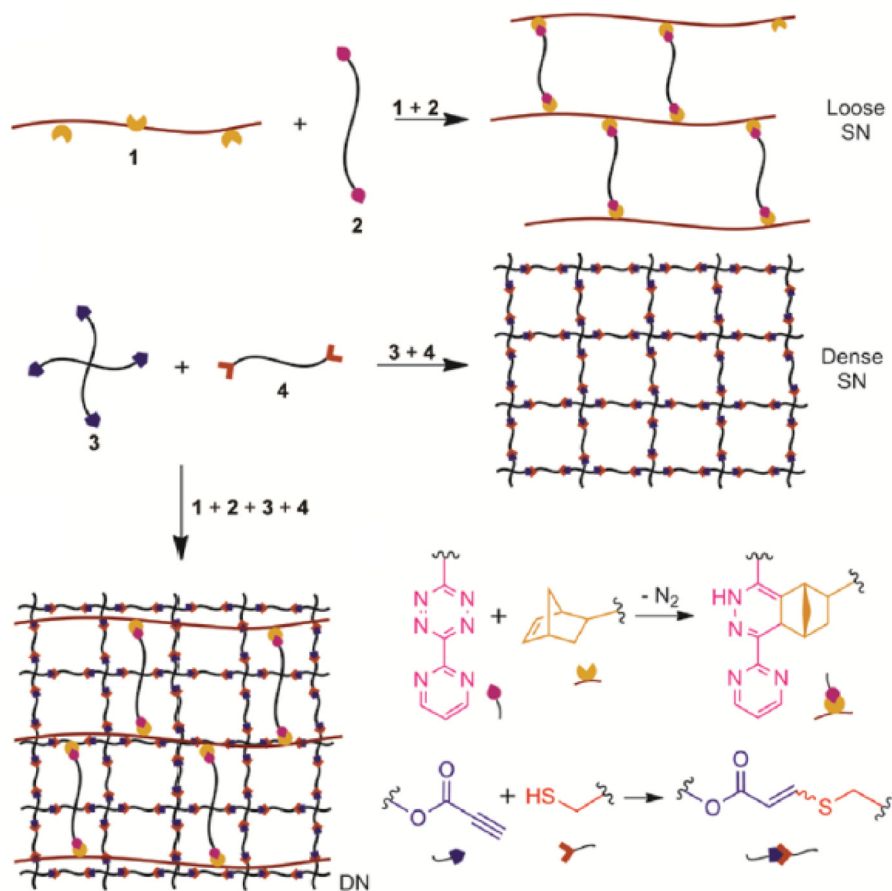


Fig. 7 Schematic of double network hydrogel formation via dual "click" approach. Adapted with permission from Truong *et al.*¹¹² Copyright © 2015, American Chemical Society.

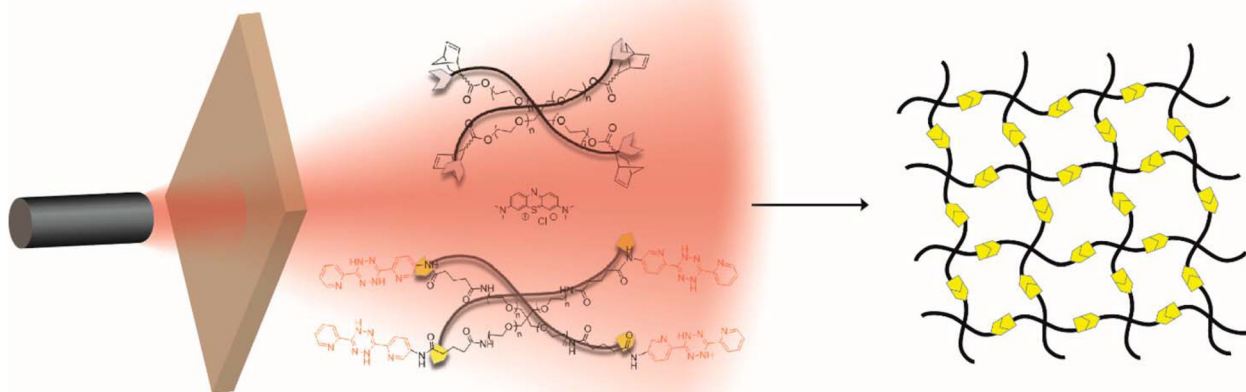


Fig. 8 The schematic illustration of hydrogel fabrication using red-light catalyzed Tz-IEDDA reaction. Adapted with permission from Truong *et al.*¹¹⁶ Copyright © 2017, American Chemical Society.

NB end functionalities showed complete degradation *in vitro* in a span of ~150 to 500 days, depending on the molecular weight of PEG used. These depots allowed a long-term control

release (over 250 days) for glucose oxidase protein encapsulated within hydrogels. In a subsequent study, the group placed hydrolyzable ester structures in the PEG-NB polymer



precursor to increase the hydrolytic degradation of the hydrogels.¹²⁰ Among the various carbonate, carbamate, and tetrahydrophthalimide hydrolyzable linkers, considerable degradation acceleration was accounted for in the case of phenyl carbonate ester inclusion. Instead of changing the nature of the linkages to the precursor polymers, it is also possible to tune the degradability of the hydrogels by varying the structure of the reactive groups. This was recently demonstrated by Lin and co-workers, who investigated hydrolytically degradable PEG-based hydrogels prepared *via* the IEDDA reaction using substituted Tz and NB derivatives (Fig. 9).¹²¹ For example, carbic anhydride was introduced to 5-norbornene-2-carboxylic acid functionalized multiarm poly(ethylene glycol) to achieve fast degradation. In addition to carbic anhydride, utilization of (methyl) tetrazine-modified macromers to form hydrogel provided highly tunable degradation. The work demonstrates that introducing small changes in the functional groups profoundly affects the stability of obtained materials. The authors postulated that such PEG-based degradable hydrogel systems may be helpful in tissue regeneration and delivery applications.

Apart from the installation of crosslinks through the reaction of Tz with appropriate dienophiles, the nitrogen-rich Tz building block and the resultant pyridazine moieties are known to coordinate with metal ions in which this attribute has been utilized in the fabrication of various metal-coordinated polymer networks.¹²² In one of their studies, Johnson and coworkers utilized the Tz-IEDDA reaction to prepare a drug-containing polymer precursor for the hydrogel, as well as harnessed the metal coordination of the Tz group to obtain supramolecular hydrogels.¹²³ Briefly, 3,6-bis(2-pyridyl)-1,2,4,5-Tzs (bptz) groups were conjugated to the end of the PEG chains ($M_n = 2000 \text{ g mol}^{-1}$) with a yield of 71%, which were utilized to obtain hydrogels in the presence of Fe^{2+} and

Ni^{2+} ions (Fig. 10). The bptz-terminated PEG chains could be functionalized using NB-containing functional molecules. In particular, an *exo*-norbornene-imide linked to doxorubicin with an *ortho*-nitrobenzyl alcohol unit and to tryptamine glycylamide unit through a chymotrypsin-cleavable peptide linker, to impart photo- and enzymatic-responsive release, respectively, were introduced into the gels through mono-functionalization of the precursor polymers using the IEDDA reaction. Obtained gels demonstrated photo- and enzymatically stimulated release of the conjugated drugs. Moreover, the gels were found to undergo complete degradation after one week, which is an attractive feature warranting the clearance of these materials for *in vivo* applications. In an alternative approach, Knall and coworkers reported the functionalization of macroporous poly(dicyclopentadiene) foams to obtain metallo-gels.¹²⁴ The macroporous poly(dicyclopentadiene) monoliths were functionalized with di(pyridyl)-pyridazines using the IEDDA reaction to introduce dipyridyl(pyridazine) units, which were used to coordinate europium(III) nitrate to impart these materials with bright red long-lived emission.

Hydrogels are indispensable materials for fabricating 3D macroscopic tissue scaffolds for encapsulation of cells. For cell encapsulation, it is essential to maintain a suitable chemical environment to promote their proliferation and function. Thus, the chemo-selective and bioorthogonal nature of the Tz-IEDDA reaction has been extensively employed in fabricating hydrogel scaffolds that efficiently incorporate cells with high post-encapsulation viability. In a seminal contribution, Anseth and coworkers reported one of the earlier examples of Tz-IEDDA “click” crosslinked hydrogels in which simple mixing of a Tz functionalized multi-armed PEG ($M_n \approx 20\,000 \text{ g mol}^{-1}$, 75% Tz functionalization) and dinorbornene-conjugated degradable peptide yielded hydrogels¹²⁵ (Fig. 11).

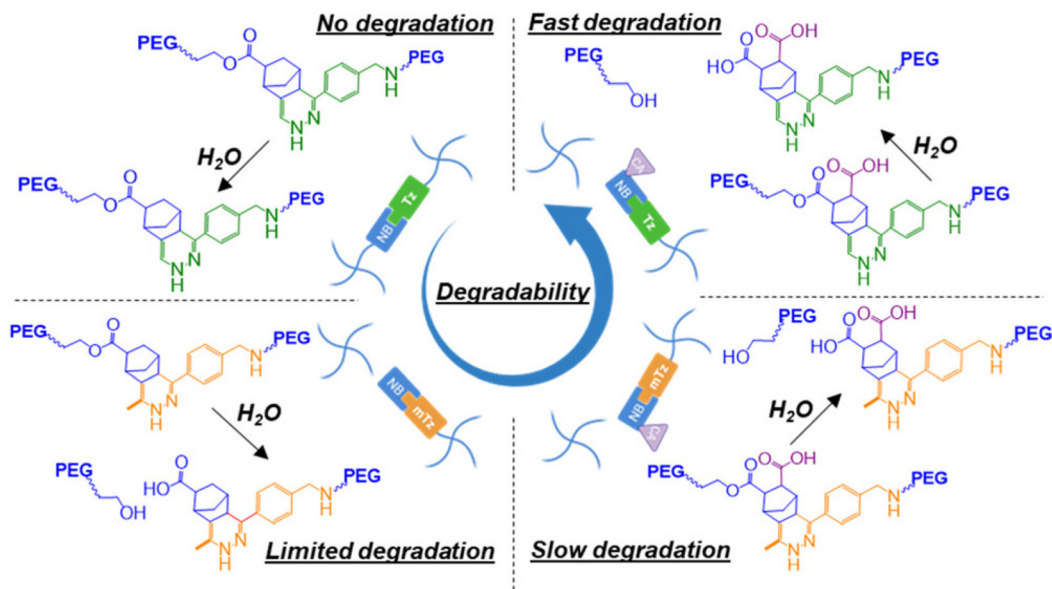


Fig. 9 Hydrolytic degradation of PEG-based Tz-IEDDA “click” hydrogels. Adapted with permission from Dimmitt *et al.*¹²¹ Copyright © 2022, American Chemical Society.



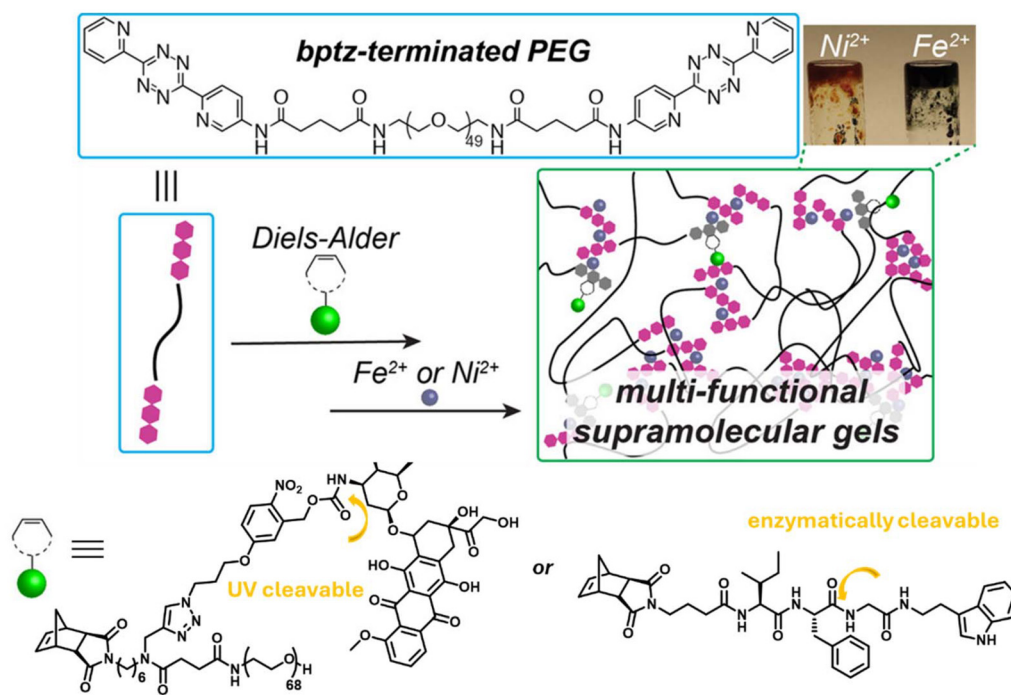


Fig. 10 The schematic illustration of metallo-hydrogel using IEDDA reaction and metal coordination. Adapted with permission from Kawamoto *et al.*¹²³ Copyright © 2015, American Chemical Society.

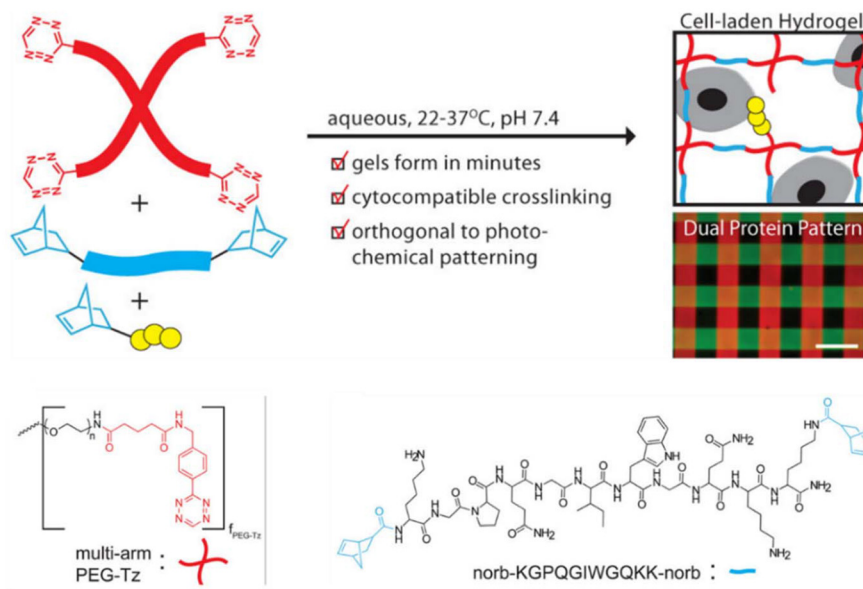


Fig. 11 Overview of hydrogel formation by Tz-NB IEDDA reaction using multi-functional PEG-Tz and cell degradable dinorbornene crosslinker peptide. Adapted with permission from Alge *et al.*¹²⁵ Copyright © 2013, American Chemical Society.

Mononorbornene-functionalized peptides introduced during gelation provided hydrogels suitable for cell encapsulation and spatially-controlled protein patterning. Notably, the cyto-compatible nature of the system led to high post-encapsulation viability of human mesenchymal stem cells.

Mooney and coworkers extensively utilized the Tz-IEDDA reaction to fabricate structurally diverse hydrogel scaffolds in

cell proliferation studies. In a study, the group reported the fabrication of alginate-based hydrogels by crosslinking individually prepared Tz and NB functionalized alginate polymers.¹²⁶ To promote cell adhesion, a thiol-modified cell adhesion peptide (cysteine bearing RGD) was also conjugated to the network structure *via* thiol-ene reaction by employing the NB groups. The alginate-based platform was reported to allow



good cell adhesion and proliferation, bringing minimal inflammatory response and high structural stability. In a subsequent study, the group extended the Tz-IEDDA-based crosslinking to hydrolytically-degradable alginate hydrogels.¹²⁷ A similar thiol-ene-based cell adhesion peptide conjugation to the network structure promoted high cellular growth and viability.

Tailorable functionalization of natural polymers or biopolymers with Tz groups is a convenient strategy to tune the network properties for efficient cell encapsulation and spreading. Since gel porosity and mechanical properties are essential to achieve a suitable scaffold for cell adhesion and spreading, tailorable swelling profiles, rheological, and morphological properties could be attained by changing the total polymer concentration and amount of complementary “click” functional groups. The biocompatible natures of these polymers also have a pivotal role in assisting enhanced cell viability and proliferation. Accordingly, Tz-modified hyaluronic acid,^{128–132} methylcellulose,¹³³ gelatin^{134,135} and chondroitin sulfate^{136,137} have been reported in IEDDA-based hydrogel synthesis for cell culture, neural stem cells delivery, and oxygen-releasing micro-particle generation.

The catalyst-free reaction conditions and bioorthogonal nature of the Tz-IEDDA reaction provide ideal cross-linking chemistry that generates sustained release depots for therapeutic proteins and drugs. The enhanced chemoselectivity of Tz-based ligation chemistry is especially advantageous in encapsulating therapeutic proteins in hydrogel scaffolds by minimizing the possible protein-matrix chemical interactions. Famili and Rajagopal exploited Tz-IEDDA reaction with NBs to fabricate hydrogels for encapsulation and release of a model protein, Fab1.¹³⁸ Hydrogels were obtained using Tz-appended hyaluronic acid (200 kg mol⁻¹) and NB-conjugated PEG (2 kg mol⁻¹) for *in situ* encapsulation of Fab1 (Fig. 12). Complete release of Fab1 from the hydrogel scaffold occurred over several weeks, and the protein was released in pristine form, without any effect on antigen binding capacity. Raghavan and coworkers reported that IEDDA-generated hyaluronic acid hydrogels can induce antibacterial activity by the slow release

of BSA-conjugated silver nanoparticles, which were encapsulated *in situ* in the gel matrix.¹³⁹

Injectable hydrogels are essential in designing deliverable drug formulations, local tissue engineering scaffolds, and dermal fillers on site. Due to fast reaction kinetics under mild conditions with high chemo-selectivity between the reactive partners, the Tz-IEDDA “click” reaction provides an attractive avenue for the fabrication of injectable hydrogels.¹⁴⁰ Since the network physicochemical properties are paramount to attain the desired application features, easy installation of Tz and complementary alkene functionalities to a diverse set of polymer structures allows fine-tuning of matrix properties after gel formation.^{141,142} The bioorthogonal and biocompatible nature of this reaction has proven especially useful in 3D cell encapsulation studies.^{143,144} In injectable systems, cells can be pre-suspended in the hydrogel precursor solutions and locally administered to the target body. This approach was demonstrated in Tz-IEDDA based *in situ* encapsulation and delivery of stem cells for osteogenic differentiation^{145,146} and cartilage tissue engineering.^{147,148} Tz-IEDDA injectable hydrogels have been efficiently utilized for *in vivo* delivery of chemical or protein therapeutic agents to articular joints,¹⁴⁹ inner ear,¹⁵⁰ intervertebral discs,¹⁵¹ bone tissue¹⁵² and dorsal skin.^{153,154}

Tz-IEDDA-based *in situ* gelation could provide a suitable platform for the delivery of anticancer agents to the target tumor tissue.¹⁵⁵ In a study to prepare injectable polypeptide hydrogels for localized cisplatin release, Chen and coworkers employed NB-modified poly(L-glutamic acid) (5.4% degree of substitution) and Tz-functionalized four-arm PEG ($M_n = 10\,000\text{ g mol}^{-1}$) (Fig. 13).¹⁵⁶ The antitumor agent cisplatin was attached to the network structure through polymer-metal complexation with the carboxylic acid groups of poly(L-glutamic acid). A rapid hydrogel formation was attained after subcutaneous injection of the gel formulation in rats, and good stability was observed *in vivo*. This cisplatin-loaded delivery platform exhibited reduced systemic toxicity and improved antitumor efficacy in an MCF-7-bearing mice model.

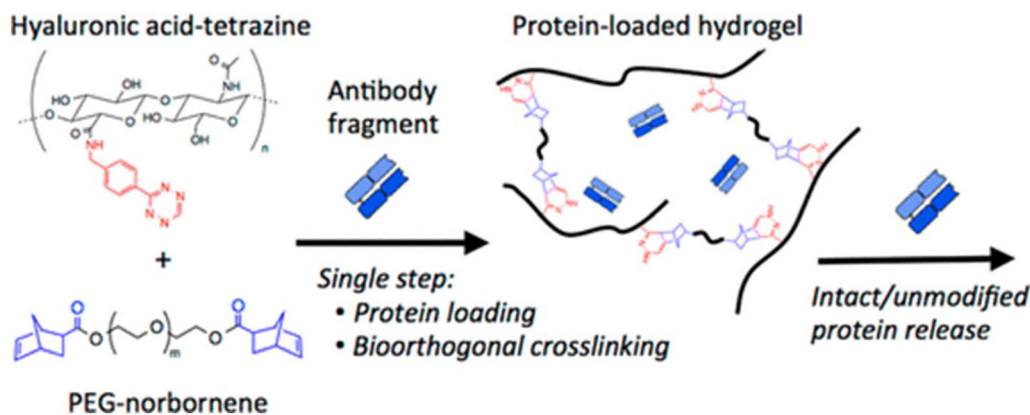


Fig. 12 Schematic illustration of fabricating protein-releasing hydrogels using Tz-IEDDA-based crosslinking of hyaluronic acid and PEG polymers. Adapted with permission from Famili *et al.*¹³⁸ Copyright © 2017, American Chemical Society.



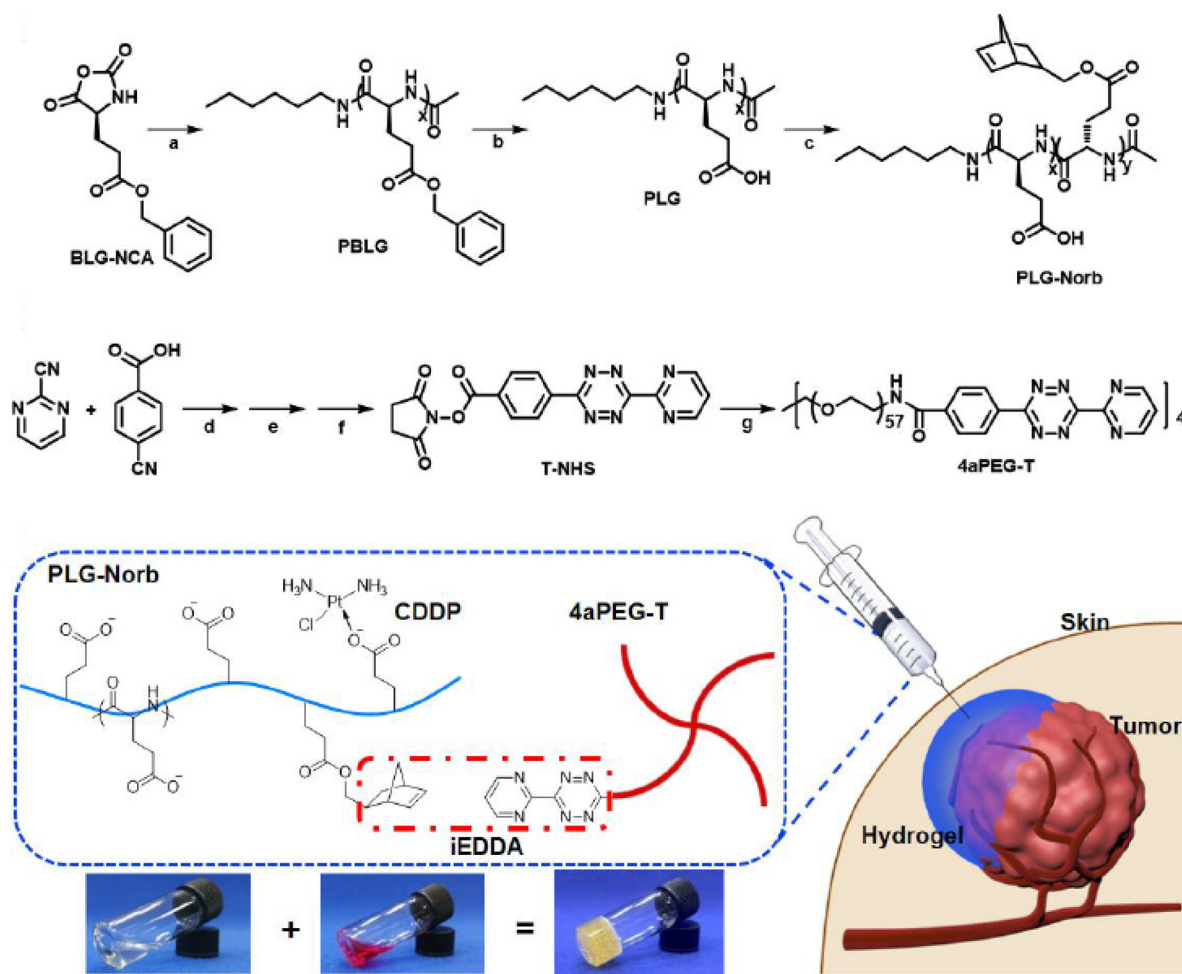


Fig. 13 Schematic representation of preparation and delivery of injectable hydrogel system loaded with antitumor agent. Adapted with permission from Zhang *et al.*¹⁵⁶ Copyright © 2020, Molecular Diversity Preservation International (MDPI).

Stimuli-responsive hydrogels have emerged as an important subclass of hydrogels, where the triggered release of bioactive agents in the presence of internal or external stimuli provides increased control.^{157,158} In several studies, Tz-IEDDA reaction has been prompted for incorporation of stimuli-responsive linkages into hydrogels for controlled drug release applications. For example, Lim and coworkers reported an injectable redox-responsive hydrogel by utilizing alginate functionalized with pendant NB units and a Tz-based PEG cross-linker where the Tz group is linked through disulfide linkages (Fig. 14).¹⁵⁹ The authors demonstrated that the encapsulated doxorubicin was efficiently (~92%) released in the presence of glutathione (GSH) (10 mM) with minimal release (<25%) in the physiological buffer.

In another related work, Lim and coworkers reported Tz-IEDDA generated diselenide-containing hydrogels for stimuli-responsive drug release responsive to near-infrared (NIR) irradiation.¹⁶⁰ Hydrogels were obtained by using NB-functionalized alginate polymer (25.53% NB functionalization) and a diselenide-containing Tz crosslinker with high conversions of

92–96% (Fig. 15). NIR-sensitive indocyanine green (ICG) dye and DOX were loaded into the hydrogel matrix during gelation. ICG generated reactive oxygen species (ROSS) upon NIR-light irradiation, which triggered hydrogel degradation through cleavage of the diselenide bonds and allowed the release of the encapsulated drug. In another work, the group reported reduction-responsive and biorthogonal carboxymethyl cellulose hydrogels obtained using a similar approach.¹⁶¹ In this work, DOX release from hydrogels was achieved through the cleavage of diselenide bonds in the presence of reducing agent GSH. In a subsequent study, alginate-based biocompatible hydrogels containing redox-responsive disulfide linkers for GSH-responsive DOX release were reported.¹⁶² Recently, the group extended their scope to hyaluronic acid-based Tz-IEDDA hydrogels that show multi-stimuli responsive drug release.¹⁶³ It was demonstrated that diselenide crosslinked networks can undergo increased DOX release triggered by exposure to DTT, H₂O₂, and NIR irradiation.

In addition to approaches that utilize the Tz-IEDDA reaction for chemical crosslinking of polymers, this chemistry has also



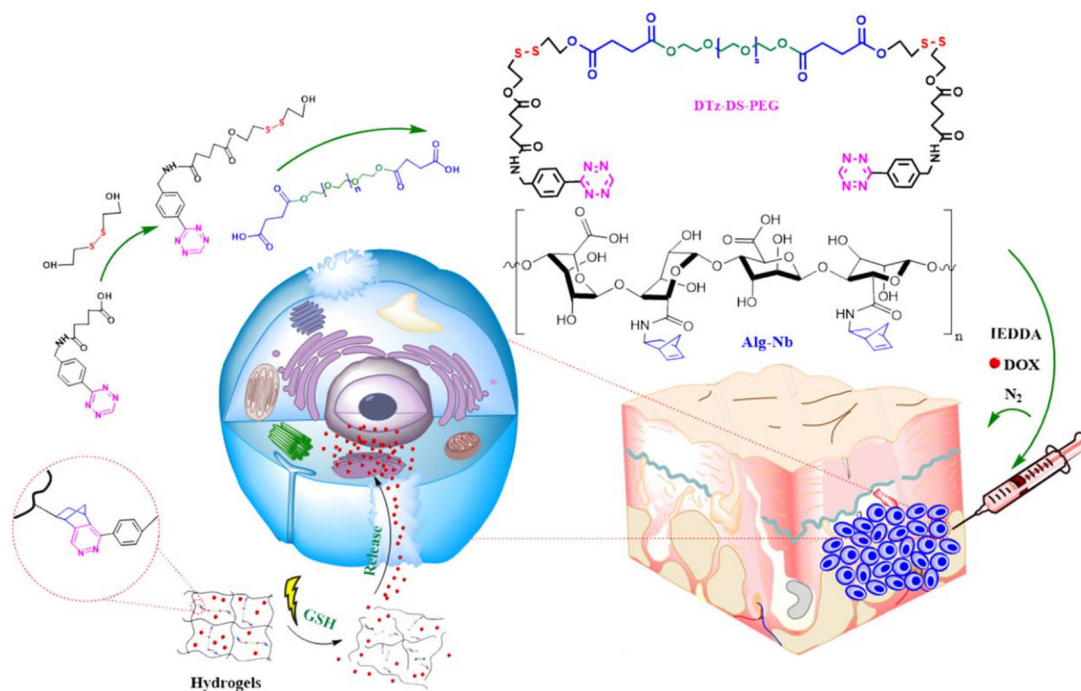


Fig. 14 Fabrication of an alginate-based injectable and reduction-responsive hydrogels. Adapted with permission from Vu et al.¹⁵⁹ Copyright © 2021 Elsevier.

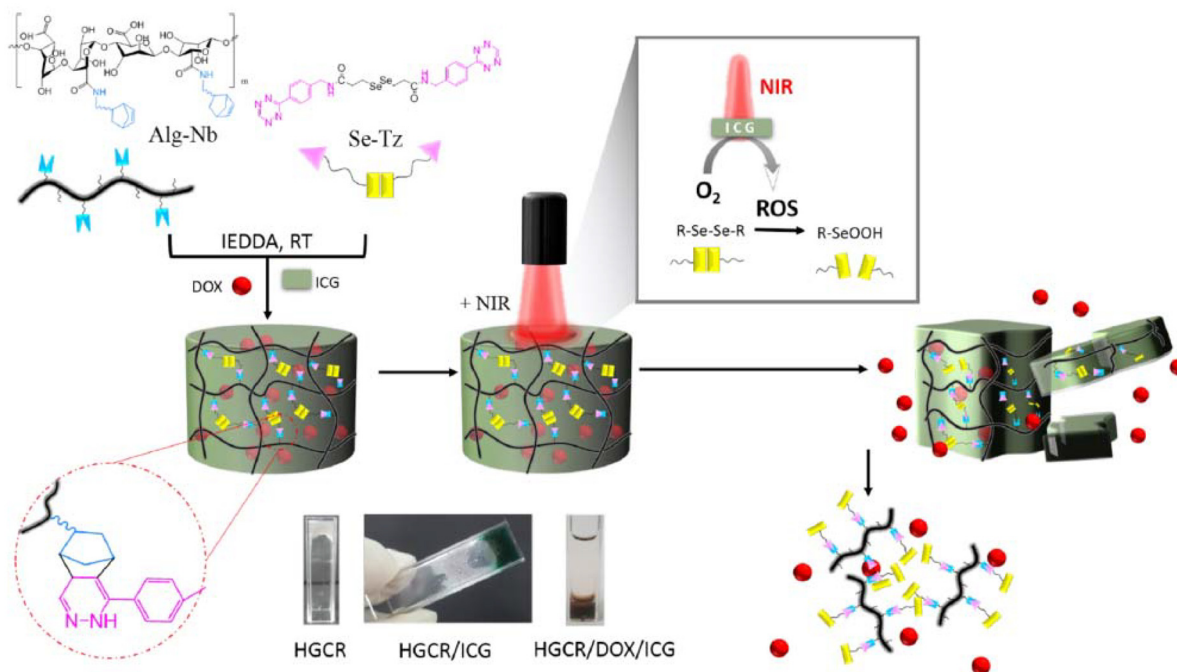


Fig. 15 Fabrication of Tz-IEDDA generated NIR-responsive alginate hydrogels. Adapted with permission from Anugrah et al.¹⁶⁰ Copyright © 2019, Elsevier.

been exploited in the post-polymerization functionalization of pre-fabricated hydrogels. In a report, Alge and coworkers demonstrated the preparation and functionalization of hydrogel microparticles *via* sequential thiol-ene addition and

Tz-IEDDA “click” reactions.¹⁶⁴ Hydrogels were synthesized by photo-polymerizing tetra-functional PEG-NB macromer/dithiothreitol mixtures emulsified in the dextran phase. Hydrogel microparticles with controlled sizes were obtained by adjusting



the stoichiometry between NB and thiol groups. It was shown that the unreacted strained alkene functionalities can efficiently be conjugated with Tz-containing model proteins such as fluorescein-labeled ovalbumin, as well as bioactive proteins alkaline phosphatase (ALP) and glucose oxidase (GOx). After functionalization, both ALP and GOx maintained their bioactivity, as demonstrated by enzyme activity assays. In a later study, the group employed a similar procedure to functionalize nucleophilic/radical thiol-ene-generated peptide hydrogels *via* a Tz-based IEDDA reaction.¹⁶⁵ Installation of Tz groups onto pre-fabricated hydrogels is another strategy for further functionalization of the templates by using appropriately modified bioactive molecules. This was demonstrated by Yi and coworkers to install Tz groups onto chitosan-polyacrylamide¹⁶⁶ and chitosan-poly(ethylene glycol)¹⁶⁷-based hydrogel microspheres.

4. Tetrazine-mediated polymer functionalization on solid surfaces

Reactive polymeric coatings that are applied on organic/inorganic solid platforms including metal surfaces, polymer substrates, glass slips or wafers are indispensable platforms in various biomedical applications.¹⁶⁸ While polymeric coatings can bestow range of important properties to applied surfaces, such as anti-biofouling, wear resistance, corrosion protection and biocompatibility; deliberately installed reactive units on these coatings might endow specific covalent attachment of desired molecules.^{169–171} Such functional platforms have especially held promise in cell culture, aligned cell growth, peptide purification, enzyme immobilization applications, and microarray technologies.^{172,173}

In the past decade, Tz-based IEDDA reaction has found a growing interest in modifying reactive polymeric coatings that are applied on solid surfaces. In an early report, Ravoo and coworkers used a microcontact printing technique and surface-initiated ATRP polymerization to obtain patterned glass and silicon surfaces with reactive Tz groups.¹⁷⁴ By using a Tz functional ATRP initiator, poly(methyl acrylate) brushes were efficiently grown in grafting from the approach on 10-undecyl trichlorosilane-modified surfaces. These reactive patterned surfaces were subsequently modified with both NB and cyclooctyne-containing substrates *via* IEDDA reaction in short reaction times and room temperature conditions. In a recent study, Kahveci and coworkers developed a grafting technique for light-induced IEDDA modification of glass surfaces (Fig. 16).¹⁷⁵ TCO-decorated surfaces were grafted with latent Tz functional dihydrotetrazine end group containing p(NIPAAm) and PLA polymers. It was demonstrated that the conversion of dihydrotetrazine groups to IEDDA *clickable* Tz functionalities could be achieved by a light-promoted photooxidation process allowing on-demand modification of glass surfaces with relevant polymers or biomolecules.

Titanium has become the material of choice in many implant applications originating from unique advantages that

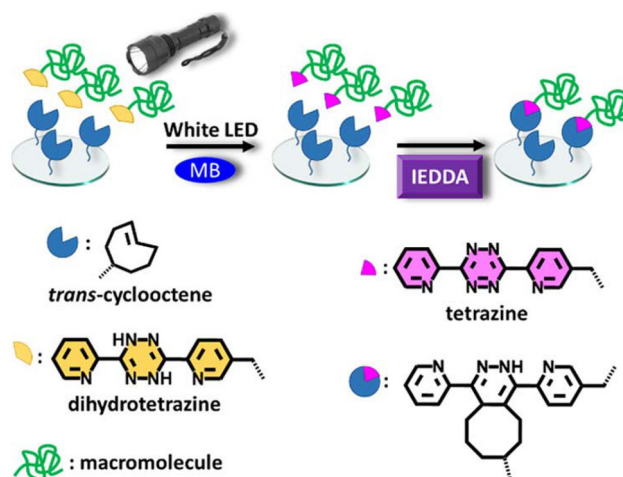


Fig. 16 Schematic illustration of a surface grafting approach based on light-induced inverse electron demand Diels–Alder (photo-IEDDA) reaction. Adapted with permission from Ozbek *et al.*¹⁷⁵ Copyright © 2021 American Chemical Society.

this metal can offer *i.e.* outstanding mechanical properties, resistance to erosion, and durability. On the other hand, biocompatibility to promote cell adhesion or osseointegration on bio-material is essential for the long-term success of titanium-based implants. To enhance the surface adhesive properties and proliferation of cells, a practiced strategy is to attach specific cell-binding moieties on the metal surface covalently. Pagel *et al.* employed a mussel-derived peptide containing strong metal affinity rendering DOPA (1-3,4-dihydroxyphenylalanine) groups and two cell binding motifs to increase the cell adhesion on the titanium surface.¹⁷⁶ Two “click” chemistry-based transformations, IEDDA and copper-catalyzed azide-alkyne cycloaddition (CuAAC) reaction, were utilized to conjugate a cyclic RGD peptide and a heparin-binding peptide to the reactive platform. It was shown that the conjugation of two cell-binding moieties on the peptide allowed the improved spreading, proliferation, and viability of cells on the titanium surface, rendering well-developed actin cytoskeleton formation. In a similar approach, Czuban *et al.* used a DOPA-containing Tz functional dendritic polyglycerol polymer to coat titanium surfaces.¹⁷⁷ An antimicrobial agent with a carbamate-TCO linker was then coupled onto the polymeric coating to facilitate the slow drug release. Bacterial experiments demonstrated that coated titanium surfaces can activate the release of anti-infective medication after IEDDA modification. This bioorthogonally reactive polymeric coating was conceptualized to prevent orthopedic implant-associated infections. In another work, Sanyal and coworkers demonstrated the fabrication of titanium surface-bound hydrogels amenable to functionalization with several “click” chemistry reactions (Fig. 17).¹⁷⁸ Hydrogel layers incorporating furan-protected maleimide groups as reactive handles were prepared by UV-based free radical crosslinking approach. In the study, oxanorbornene groups were efficiently functionalized with IEDDA



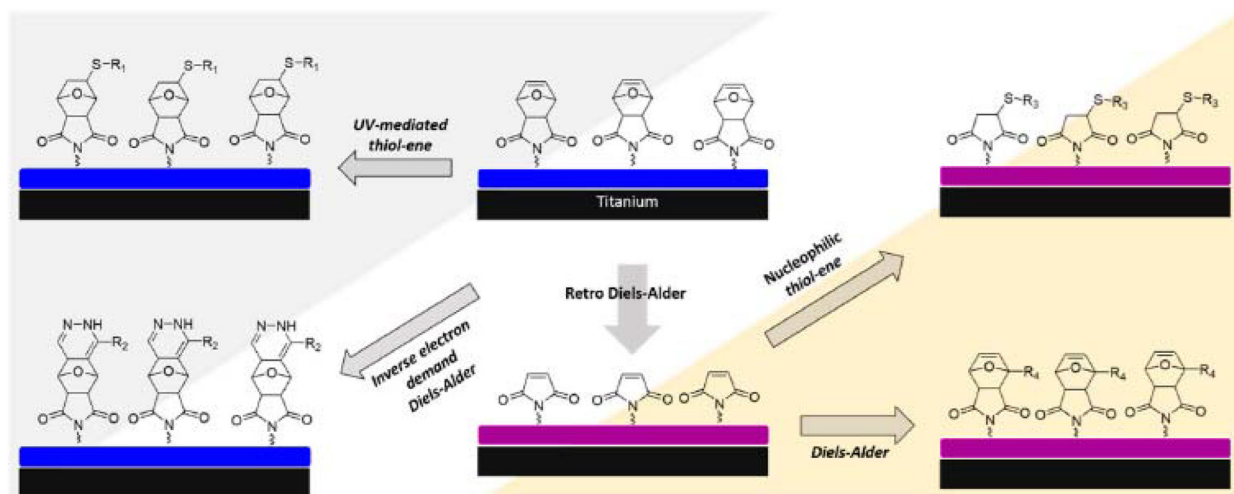


Fig. 17 Multi-functionalization of titanium surface bound hydrogels using different “click” chemistry reactions, including the IEDDA reaction. Adapted with permission from Gevrek et al.¹⁷⁸ Copyright © 2020, Molecular Diversity Preservation International (MDPI).

and radical thiol-ene reactions by covalent attachment of molecular cues with complementary Tz and thiol functionalities. It was also demonstrated that, after furan deprotection and liberation of maleimide groups, nucleophilic thiol-maleimide addition and Diels-Alder chemistries could also be applied to modify these surface-bound hydrogel platforms covalently. In another work, the same group extended the fabrication of a multi-functionalizable bioconjugation platform concept to surface-immobilized nanofibers.¹⁷⁹ Electrospun PLA-based polymers carrying orthogonal oxanorbornene and azide groups were efficiently functionalized with Tz and cyclooctyne-bearing bioactive cues *via* sequential SPAAC and IEDDA bioorthogonal “click” reactions. Recently, sequential SPAAC and IEDDA functionalization strategy was used by Huskens and coworkers on gold and silicon dioxide surfaces coated with either Tz or cyclooctyne functional poly-L-lysine (PLL) polymers.¹⁸⁰ It was shown that DNA probe molecules with complementary azide and TCO groups can be selectively conjugated to the surfaces within less than 20 min incubation period. Notably, while modified surfaces showed hybridization with entirely complementary DNA sequences, they did not respond to non-complementary DNA sequences.

Dankers and coworkers investigated the supramolecular elastomeric materials based on ureido pyrimidinone matrices as anti-fouling coating materials.¹⁸¹ The supramolecular polymer scaffolds facilitating Tz groups as IEDDA reactive handles were surface modified by cyclooctyne end group containing PEG polymers with different topologies. The protein adsorption experiments revealed that the efficiency of anti-fouling behavior is directly related to the PEG polymer applied, *i.e.*, the best performance was accounted for by using a star PEG polymer ($M_n = 10\,000\text{ g mol}^{-1}$) rendering four cyclooctyne groups, then monofunctional and bifunctional PEG-cyclooctyne derivatives ($M_n = 2000\text{ g mol}^{-1}$ and 5000 g mol^{-1}). In a follow-up study, using a TCO-functional peptide derivative,

these Tz functional supramolecular substrates fabricated as cast films or electrospun fiber mats were employed in specific protein immobilization.¹⁸²

5. Outlook and conclusions

The preceding sections highlight the significant potential of IEDDA cycloaddition reactions in designing and synthesizing functional polymeric materials. While IEDDA reactions follow the core principles of “click” chemistry, their biorthogonal nature extends their utility to new applications. Notably, *in vivo* studies have demonstrated the reaction’s effectiveness in living systems. Furthermore, the high efficiency of IEDDA reactions makes them invaluable for post-polymerization modifications, particularly when rapid transformations are needed, such as the conjugation of short-lived species like radioisotopes.

Current research predominantly focuses on incorporating Tz moieties through post-polymerization modifications. However, developing polymerization methods that directly yield polymers with these reactive handles would be advantageous. This approach poses challenges with vinylic monomers due to the reactivity of electron-deficient dienes with double bonds, but it may be more feasible with non-vinylic monomers, such as those used in ROP. As reviewed, the excellent selectivity and high yield of IEDDA reactions also make them suitable for fabricating network structures. Although numerous examples of such applications exist, integrating inorganic components into hybrid materials remains limited. Advancing the development of biomolecule-polymer hybrid materials could lead to innovative functional materials for biomedical applications. Nonetheless, challenges related to the multi-step synthesis of reactive counterparts persist. Continued progress in synthesizing Tz-bearing linkers and their complementary reactive partners is expected. Ultimately,



the remarkable efficiency of the IEDDA reactions in addressing specific challenges in fabricating functional polymeric materials for targeted applications will continue to drive their broader adoption and utilization.

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

Data availability

No primary research results have been included and no new data were generated or analyzed as part of this review.

Conflicts of interest

There are no conflicts to declare.

References

- 1 A.-C. Knall and C. Slugovc, *Chem. Soc. Rev.*, 2013, **42**, 5131–5142.
- 2 A. M. Prokhorov and D. N. Kozhevnikov, *Chem. Heterocycl. Compd.*, 2012, **48**, 1153–1176.
- 3 N. Saracoglu, *Tetrahedron*, 2007, **63**, 4199–4236.
- 4 Z. Geng, J. J. Shin, Y. Xi and C. J. Hawker, *J. Polym. Sci.*, 2021, **59**, 963–1042.
- 5 M. A. Tasdelen, *Polym. Chem.*, 2011, **2**, 2133–2145.
- 6 A. Sanyal, *Macromol. Chem. Phys.*, 2010, **211**, 1417–1425.
- 7 G. Hizal, U. Tunca and A. Sanyal, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 4103–4120.
- 8 M. Cetin, C. Esen, O. Daglar, S. Luleburgaz, G. Hizal, H. Durmaz and U. Tunca, *Polym. Chem.*, 2016, **7**, 7094–7100.
- 9 A. Dag, H. Durmaz, U. Tunca and G. Hizal, *J. Polym. Sci., Part A: Polym. Chem.*, 2008, **47**, 178–187.
- 10 H. Sun, C. P. Kabb, Y. Dai, M. R. Hill, I. Ghiviriga, A. P. Bapat and B. S. Sumerlin, *Nat. Chem.*, 2017, **9**, 817–823.
- 11 P. Espeel and F. E. Du Prez, *Macromolecules*, 2015, **48**, 2–14.
- 12 K. K. Oehlenschlaeger, N. K. Guimard, J. Brandt, J. O. Mueller, C. Y. Lin, S. Hilf, A. Lederer, M. L. Coote, F. G. Schmidt and C. Barner-Kowollik, *Polym. Chem.*, 2013, **4**, 4348–4355.
- 13 A. Bousquet, C. Barner-Kowollik and M. H. Stenzel, *J. Polym. Sci., Part A: Polym. Chem.*, 2010, **48**, 1773–1781.
- 14 J. Yang, M. R. Karver, W. Li, S. Sahu and N. K. Devaraj, *Angew. Chem., Int. Ed.*, 2012, **51**, 5222–5225.
- 15 R. A. Carboni and R. V. Lindsey, *J. Am. Chem. Soc.*, 1959, **81**, 4342–4346.
- 16 D. L. Boger, *Chem. Rev.*, 1986, **86**, 781–793.
- 17 J. Zhang, V. Shukla and D. L. Boger, *J. Org. Chem.*, 2019, **84**, 9397–9445.
- 18 H. C. Kolb, M. G. Finn and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2001, **40**, 2004–2021.
- 19 N. K. Devaraj and R. Weissleder, *Acc. Chem. Res.*, 2011, **44**, 816–827.
- 20 B. L. Oliveira, Z. Guo and G. J. L. Bernardes, *Chem. Soc. Rev.*, 2017, **46**, 4895–4950.
- 21 S. A. Busche, S. Peplau, L. Zuhse, D. Steimer, D. D. August and H. G. Börner, *Polym. Chem.*, 2023, **14**, 2888–2892.
- 22 A. R. Sayed and J. S. Wiggins, *J. Heterocycl. Chem.*, 2024, **61**, 118–126.
- 23 A. R. Sayed and J. S. Wiggins, *J. Appl. Polym. Sci.*, 2011, **120**, 623–630.
- 24 Z. Li and J. Ding, *Macromol. Chem. Phys.*, 2011, **212**, 2260–2267.
- 25 L. Peng, Y. Yu, J. Lu, P. He, G. Wang, M. Huang, B. Zhao, Y. Pei and S. Tan, *Dyes Pigm.*, 2019, **171**, 107717.
- 26 J. Ding, Z. Li, Z. Cui, G. P. Robertson, N. Song, X. Du and L. Scoles, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 3374–3386.
- 27 J. S. Panek and B. Zhu, *Tetrahedron Lett.*, 1996, **37**, 8151–8154.
- 28 S. L. Scinto, D. A. Bilodeau, R. Hincapie, W. Lee, S. S. Nguyen, M. Xu, C. W. Am Ende, M. G. Finn, K. Lang, Q. Lin, J. P. Pezacki, J. A. Prescher, M. S. Robillard and J. M. Fox, *Nat. Rev. Methods Primers*, 2021, **1**, 30.
- 29 W. R. Algar, D. E. Prasuhn, M. H. Stewart, T. L. Jennings, J. B. Blanco-Canosa, P. E. Dawson and I. L. Medintz, *Bioconjugate Chem.*, 2011, **22**, 825–858.
- 30 R. E. Bird, S. A. Lemmel, X. Yu and Q. A. Zhou, *Bioconjugate Chem.*, 2021, **32**, 2457–2479.
- 31 N. K. Devaraj, G. M. Thurber, E. J. Keliher, B. Marinelli and R. Weissleder, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 4762–4767.
- 32 J.-P. Meyer, K. M. Tully, J. Jackson, T. R. Dilling, T. Reiner and J. S. Lewis, *Bioconjugate Chem.*, 2018, **29**, 538–545.
- 33 Y. Dong, Y. Tu, K. Wang, C. Xu, Y. Yuan and J. Wang, *Angew. Chem., Int. Ed.*, 2020, **59**, 7168–7172.
- 34 S. Liu, H. Zhang, R. A. Remy, F. Deng, M. E. Mackay, J. M. Fox and X. Jia, *Adv. Mater.*, 2015, **27**, 2783–2790.
- 35 H. Zhang, W. S. Trout, S. Liu, G. A. Andrade, D. A. Hudson, S. L. Scinto, K. T. Dicker, Y. Li, N. Lazouski, J. Rosenthal, C. Thorpe, X. Jia and J. M. Fox, *J. Am. Chem. Soc.*, 2016, **138**, 5978–5983.
- 36 M. M. Lorenzo, C. G. Decker, M. U. Kahveci, S. J. Paluck and H. D. Maynard, *Macromolecules*, 2016, **49**(1), 30–37.
- 37 T. R. Wilks and R. K. O'Reilly, *Sci. Rep.*, 2016, **6**, 39192.
- 38 S. S. Kara, M. Y. Ateş, G. Devenci, A. Cetinkaya and M. U. Kahveci, *J. Polym. Sci., Part A: Polym. Chem.*, 2018, **57**, 673–680.
- 39 G. Bakirdogen, E. L. Sahkulubey Kahveci and M. U. Kahveci, *Eur. Polym. J.*, 2020, **140**, 110027.
- 40 E. Meyvacı and T. Öztürk, *ChemistrySelect*, 2022, **7**, e202200668.



- 41 M. Ambrosi, N. R. Cameron and B. G. Davis, *Org. Biomol. Chem.*, 2005, **3**, 1593–1608.
- 42 M. Mammen, S.-K. Choi and G. M. Whitesides, *Angew. Chem., Int. Ed.*, 1998, **37**, 2754–2794.
- 43 L. L. Kiessling, J. E. Gestwicki and L. E. Strong, *Angew. Chem., Int. Ed.*, 2006, **45**, 2348–2368.
- 44 C. Timperanza, H. Jensen, T. Bäck, S. Lindegren and E. Aneheim, *Pharmaceuticals*, 2023, **16**, 595.
- 45 I. Lepori, Y. Oz, J. Lm, N. Ghosh, M. Paul, U. S. Schubert and S. Fedeli, *Reactions*, 2024, **5**, 231–245.
- 46 C. F. Hansell, P. Espeel, M. M. Stamenovic, I. A. Barker, A. P. Dove, F. E. Du Prez and R. K. O'Reilly, *J. Am. Chem. Soc.*, 2011, **133**, 13828–13831.
- 47 K. Johann, D. Svatunek, C. Seidl, S. Rizzelli, T. A. Bauer, L. Braun, K. Koynov, H. Mikula and M. Barz, *Polym. Chem.*, 2020, **11**, 4396–4407.
- 48 S. A. van den Berg, H. Zuilhof and T. Wennekes, *Macromolecules*, 2016, **49**, 2054–2062.
- 49 O. Stetsiuk, A. Abherve and N. Avarvari, *Dalton Trans.*, 2020, **49**, 5759–5777.
- 50 S. Kramer, D. Svatunek, I. Alberg, B. Gräfen, S. Schmitt, L. Braun, A. H. A. M. van Onzen, R. Rossin, K. Koynov, H. Mikula and R. Zentel, *Biomacromolecules*, 2019, **20**, 3786–3797.
- 51 K. R. Aswani, S. Joshi, R. Ghosh and R. R. Malakalappalli, *Polym. Adv. Technol.*, 2023, **34**, 1331–1340.
- 52 L. Xu, W. Tao, M. Guan, X. Yang, M. Huang, H. Chen, J. Zhang, B. Zhao and S. Tan, *ACS Appl. Energy Mater.*, 2021, **4**, 11624–11633.
- 53 V. Kardelis, M. M. Denk and A. Adronov, *Angew. Chem., Int. Ed.*, 2021, **60**, 2980–2986.
- 54 D. S. Rivero, R. E. Paiva-Feener, T. Santos, E. Martín-Encinas and R. Carrillo, *Macromolecules*, 2021, **54**, 10428–10434.
- 55 T. Santos, D. S. Rivero, Y. Pérez-Pérez, E. Martín-Encinas, J. Pasán, A. H. Daranas and R. Carrillo, *Angew. Chem., Int. Ed.*, 2021, **60**, 18783–18791.
- 56 Z. M. Png, X. Y. D. Soo, J. X. D. Liew, M. H. Chua, S. Xiong, Q. Zhu and J. Xu, *Polym. Chem.*, 2023, **14**, 3347–3351.
- 57 R. N. Butler and A. G. Coyne, *Chem. Rev.*, 2010, **110**, 6302–6337.
- 58 N. B. Cramer, S. K. Reddy, A. K. O'Brien and C. N. Bowman, *Macromolecules*, 2003, **36**, 7964–7969.
- 59 C. E. Hoyle and C. N. Bowman, *Angew. Chem., Int. Ed.*, 2010, **49**, 1540–1573.
- 60 C. W. Bielawski and R. H. Grubbs, *Prog. Polym. Sci.*, 2007, **32**, 1–29.
- 61 I. A. Barker, D. J. Hall, C. F. Hansell, F. E. Du Prez, R. K. O'Reilly and A. P. Dove, *Macromol. Rapid Commun.*, 2011, **32**, 1362–1366.
- 62 R. J. Williams, I. A. Barker, R. K. O'Reilly and A. P. Dove, *ACS Macro Lett.*, 2012, **1**, 1285–1290.
- 63 R. Baumgartner, Z. Song, Y. Zhang and J. Cheng, *Polym. Chem.*, 2015, **6**, 3586–3590.
- 64 S. Jain, K. Neumann, Y. Zhang, J. Geng and M. Bradley, *Macromolecules*, 2016, **49**, 5438–5443.
- 65 H.-S. Han, N. K. Devaraj, J. Lee, S. A. Hilderbrand, R. Weissleder and M. G. Bawendi, *J. Am. Chem. Soc.*, 2010, **132**, 7838–7839.
- 66 Y. Zhang, A. Gambardella, M. Üçüncü, J. Geng, J. Clavadetscher, M. Bradley and A. Lilienkamp, *Chem. Commun.*, 2020, **56**, 13856–13859.
- 67 Y. Zhang, M. Üçüncü, A. Gambardella, A. Baibek, J. Geng, S. Zhang, J. Clavadetscher, I. Litzen, M. Bradley and A. Lilienkamp, *J. Am. Chem. Soc.*, 2020, **142**, 21615–21621.
- 68 C. F. Hansell and K. O'Reilly, *ACS Macro Lett.*, 2012, **1**, 896–901.
- 69 C. F. Hansell, A. Lu, J. P. Patterson and R. K. O'Reilly, *Nanoscale*, 2014, **6**, 4102–4107.
- 70 S. Xie, M. Chen, X. Song, Z. Zhang, Z. Zhang, Z. Chen and X. Li, *Acta Biomater.*, 2018, **78**, 198–210.
- 71 A. H. St Amant, E. H. Discekici, S. J. Bailey, M. S. Zayas, J.-A. Song, S. L. Shankel, S. N. Nguyen, M. W. Bates, A. Anastasaki, C. J. Hawker and J. R. de Alaniz, *J. Am. Chem. Soc.*, 2019, **141**, 13619–13624.
- 72 S. J. Bailey, E. H. Discekici, S. M. Barbon, S. N. Nguyen, C. J. Hawker and J. Read de Alaniz, *Macromolecules*, 2020, **53**, 4917–4924.
- 73 R. Rossin, S. M. van den Bosch, W. ten Hoeve, M. Carvelli, R. M. Versteegen, J. Lub and M. S. Robillard, *Bioconjugate Chem.*, 2013, **24**, 1210–1217.
- 74 F. Thalhammer, U. Wallfahner and J. Sauer, *Tetrahedron Lett.*, 1990, **31**, 6851–6854.
- 75 D. Svatunek, C. Denk, V. Rosecker, B. Sohr, C. Hametner, G. Allmaier, J. Fröhlich and H. Mikula, *Monatsh. Chem.*, 2016, **147**, 579–585.
- 76 V. Shalgunov, G. Engudar, L. Bohrmann, L. Wharton, K. Maskell, K. Johann, M. Barz, P. Schaffer, M. M. Herth and V. Radchenko, *Nucl. Med. Biol.*, 2022, **104–105**, 11–21.
- 77 B. E. Cook, R. Membreno and B. M. Zeglis, *Bioconjugate Chem.*, 2018, **29**, 2734–2740.
- 78 E. J. L. Stéen, J. T. Jørgensen, K. Johann, K. Nørregaard, B. Sohr, D. Svatunek, A. Birke, V. Shalgunov, P. E. Edem, R. Rossin, C. Seidl, F. Schmid, M. S. Robillard, J. L. Kristensen, H. Mikula, M. Barz, A. Kjær and M. M. Herth, *ACS Nano*, 2020, **14**, 568–584.
- 79 R. García-Vázquez, U. M. Battisti, V. Shalgunov, G. Schäfer, M. Barz and M. M. Herth, *Macromol. Rapid Commun.*, 2021, **43**, 2100655.
- 80 J. A. C. M. Goos, M. Davydova, T. R. Dilling, A. Cho, M. A. Cornejo, A. Gupta, W. S. Price, S. Puttick, M. R. Whittaker, J. F. Quinn, T. P. Davis and J. S. Lewis, *Nucl. Med. Biol.*, 2020, **84–85**, 63–72.
- 81 L. Xu, M. Raabe, M. M. Zegota, J. C. F. Nogueira, V. Chudasama, S. L. Kuan and T. Weil, *Org. Biomol. Chem.*, 2020, **18**, 1140–1147.
- 82 M. Arslan, G. Acik and M. A. Tasdelen, *Polym. Chem.*, 2019, **10**, 3806–3821.
- 83 M. Arslan and M. A. Tasdelen, *Polymers*, 2017, **9**, 499.
- 84 M. Arslan and M. A. Tasdelen, *Chem. Afr.*, 2019, **2**, 195–214.



- 85 R. E. Bagge, T. C. Mauldin, D. J. Boday, B. M. Kobilka and D. A. Loy, *Chem. Mater.*, 2017, **29**, 7953–7960.
- 86 Q. Wang, W. Wang, Q. Li and C. Wu, *Ind. Eng. Chem. Res.*, 2021, **60**, 2163–2177.
- 87 Q. Wang, Y. Shi, Q. Li and C. Wu, *Eur. Polym. J.*, 2021, **150**, 110415.
- 88 H. Zhou and J. A. Johnson, *Angew. Chem., Int. Ed.*, 2013, **52**, 2235–2238.
- 89 H. Zhou, J. Woo, A. M. Cok, M. Wang, B. D. Olsen and J. A. Johnson, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 19119–19124.
- 90 H. Zhou, E.-M. Schön, M. Wang, M. J. Glassman, J. Liu, M. Zhong, D. D. Díaz, B. D. Olsen and J. A. Johnson, *J. Am. Chem. Soc.*, 2014, **136**, 9464–9470.
- 91 A. Degirmenci, R. Sanyal and A. Sanyal, *Bioconjugate Chem.*, 2024, **35**, 433–452.
- 92 S. A. Busche, M. Traxler, A. Thomas and H. G. Börner, *ChemSusChem*, 2024, **17**, e202301045.
- 93 A. M. Cok, H. Zhou and J. A. Johnson, *Macromol. Symp.*, 2013, **329**, 108–112.
- 94 S. E. Holt, A. Rakoski, F. Jivan, L. M. Pérez and D. L. Alge, *Macromol. Rapid Commun.*, 2020, **41**, 2000287.
- 95 D. S. B. Anugrah, M. P. Patil, X. Li, C. M. Q. Le, K. Ramesh, G.-D. Kim, K. Hyun and K. T. Lim, *EXPRESS Polym. Lett.*, 2020, **14**, 248–260.
- 96 S. Liu, A. C. Moore, A. B. Zerdoum, H. Zhang, S. L. Scinto, H. Zhang, L. Gong, D. L. Burris, A. K. Rajasekaran, J. M. Fox and X. Jia, *Biomaterials*, 2018, **180**, 24–35.
- 97 O. J. George, J. Song, J. M. Benson, Y. Fang, H. Zhang, D. L. Burris, J. M. Fox and X. Jia, *Biomacromolecules*, 2022, **23**, 3017–3030.
- 98 R. P. Tello, S. Wang, F. Fontana, A. Correia, G. Molinaro, S. L. Cerdà, S. Hietala, J. Hirvonen, G. Barreto and H. A. Santos, *Biomater. Sci.*, 2023, **11**, 4972–4984.
- 99 H. Zhang, K. T. Dicker, X. Xu, X. Jia and J. M. Fox, *ACS Macro Lett.*, 2014, **3**, 727–731.
- 100 K. T. Dicker, A. C. Moore, N. T. Garabedian, H. Zhang, S. L. Scinto, R. E. Akins, D. L. Burris, J. M. Fox and X. Jia, *ACS Appl. Mater. Interfaces*, 2019, **11**, 16402–16411.
- 101 N. J. Darling, W. Xi, E. Sideris, A. R. Anderson, C. Pong, S. T. Carmichael and T. Segura, *Adv. Healthcare Mater.*, 2020, **9**, 1901391.
- 102 K. L. Wilson, S. C. L. Pérez, M. M. Naffaa, S. H. Kelly and T. Segura, *Adv. Mater.*, 2022, **34**, 2201921.
- 103 A. Isaac, F. Jivan, S. Xin, J. Hardin, X. Luan, M. Pandya, T. G. H. Diekwisch and D. L. Alge, *ACS Biomater. Sci. Eng.*, 2019, **5**, 6395–6404.
- 104 F. Jivan and D. L. Alge, *Adv. Ther.*, 2020, **3**, 1900148.
- 105 T. J. Tigner, G. Dampf, A. Tucker, Y.-C. Huang, V. Jagrit, A. J. Clevenger, A. Mohapatra, S. A. Raghavan, J. N. Dulin and D. L. Alge, *Adv. Healthcare Mater.*, 2024, e2303912.
- 106 A. Lueckgen, D. S. Garske, A. Ellinghaus, D. J. Mooney, G. N. Duda and A. Cipitria, *Acta Biomater.*, 2020, **115**, 185–196.
- 107 A. L. Rutz, E. S. Gargus, K. E. Hyland, P. L. Lewis, A. Setty, W. R. Burghardt and R. N. Shah, *Acta Biomater.*, 2019, **99**, 121–132.
- 108 A. Oehrl, S. Schötz and R. Haag, *Macromol. Rapid Commun.*, 2020, **41**, 1900510.
- 109 A. Oehrl, S. Schötz and R. Haag, *Colloid Polym. Sci.*, 2020, **298**, 719–733.
- 110 S. Schötz, F. Reisbeck, A.-C. Schmitt, M. Dimde, E. Quaas, K. Achazi and R. Haag, *Pharmaceutics*, 2021, **13**, 1276.
- 111 S. T. Koshy, D. K. Y. Zhang, J. M. Grolman, A. G. Stafford and D. J. Mooney, *Acta Biomater.*, 2018, **65**, 36–43.
- 112 V. X. Truong, M. P. Ablett, S. M. Richardson, J. A. Hoyland and A. P. Dove, *J. Am. Chem. Soc.*, 2015, **137**, 1618–1622.
- 113 M. R. Arkenberg, N. H. Dimmitt, H. C. Johnson, K. R. Koehler and C.-C. Lin, *Adv. Biosyst.*, 2020, **4**, 2000129.
- 114 S. E. Holt, J. Arroyo, E. Poux, A. Fricks, I. Agurcia, M. Heintschel, A. Rakoski and D. L. Alge, *Biomacromolecules*, 2021, **22**, 3040–3048.
- 115 M. R. Karver, R. Weissleder and S. A. Hilderbrand, *Bioconjugate Chem.*, 2011, **22**, 2263–2270.
- 116 V. X. Truong, K. M. Tsang, F. Ercole and J. S. Forsythe, *Chem. Mater.*, 2017, **29**, 3678–3685.
- 117 J. Carthew, J. E. Frith, J. S. Forsythe and V. X. Truong, *J. Mater. Chem. B*, 2018, **6**, 1394–1401.
- 118 C. Wang, H. Zhang, T. Zhang, X. Zou, H. Wang, J. E. Rosenberger, R. Vannam, W. S. Trout, J. B. Grimm, L. D. Lavis, C. Thorpe, X. Jia, Z. Li and J. M. Fox, *J. Am. Chem. Soc.*, 2021, **143**, 10793–10803.
- 119 C. E. Ziegler, M. Graf, M. Nagaoka, H. Lehr and A. M. Goepferich, *Biomacromolecules*, 2021, **22**, 3223–3236.
- 120 C. E. Ziegler, M. Graf, M. Nagaoka and A. M. Goepferich, *Macromol. Biosci.*, 2022, **22**, 2200226.
- 121 N. H. Dimmitt, M. R. Arkenberg, M. M. de Lima Perini, J. Li and C.-C. Lin, *ACS Biomater. Sci. Eng.*, 2022, **8**, 4262–4273.
- 122 Z. Yao, Z. Wang, Y. Yu, C. Zeng and K. Cao, *Polymer*, 2017, **119**, 98–106.
- 123 K. Kawamoto, S. C. Grindy, J. Liu, N. Holten-Andersen and J. A. Johnson, *ACS Macro Lett.*, 2015, **4**, 458–461.
- 124 A.-C. Knall, S. Kovacic, M. Hollauf, D. Reishofer, R. Saf and C. Slugovc, *Chem. Commun.*, 2013, **49**, 7325–7327.
- 125 D. L. Alge, M. A. Azagarsamy, D. F. Donohue and K. S. Anseth, *Biomacromolecules*, 2013, **14**, 949–953.
- 126 R. M. Desai, S. T. Koshy, S. A. Hilderbrand, D. J. Mooney and N. S. Joshi, *Biomaterials*, 2015, **50**, 30–37.
- 127 A. Lueckgen, D. S. Garske, A. Ellinghaus, R. M. Desai, A. G. Stafford, D. J. Mooney, G. N. Duda and A. Cipitria, *Biomaterials*, 2018, **181**, 189–198.
- 128 V. Delplace, P. E. B. Nickerson, A. Ortin-Martinez, A. E. G. Baker, V. A. Wallace and M. S. Shoichet, *Adv. Funct. Mater.*, 2020, **30**, 1903978.
- 129 K. A. Gultian, R. Gandhi, T. W. B. Kim and S. L. Vega, *Macromol. Biosci.*, 2023, **23**, 2200425.
- 130 J. Song, H. Gao, H. Zhang, O. J. George, A. S. Hillman, J. M. Fox and X. Jia, *ACS Appl. Mater. Interfaces*, 2022, **14**, 51669–51682.



- 131 Y. Hao, J. Song, A. Ravikrishnan, K. T. Dicker, E. W. Fowler, A. B. Zerdoum, Y. Li, H. Zhang, A. K. Rajasekaran, J. M. Fox and X. Jia, *ACS Appl. Mater. Interfaces*, 2018, **10**, 26016–26027.
- 132 M. Pol, H. Gao, H. Zhang, O. J. George, J. M. Fox and X. Jia, *Biomaterials*, 2023, **299**, 122180.
- 133 V. Delplace, A. J. Pickering, M. H. Hettiaratchi, S. Zhao, T. Kivijärvi and M. S. Shoichet, *Biomacromolecules*, 2020, **21**, 2421–2431.
- 134 N. C. Negrini, A. A. Volponi, P. T. Sharpe and A. D. Celiz, *ACS Biomater. Sci. Eng.*, 2021, **7**, 4330–4346.
- 135 M. R. Arkenberg, K. Koehler and C.-C. Lin, *Biomacromolecules*, 2022, **23**, 4141–4152.
- 136 E. de Sousa Araújo, T. D. Stocco, G. F. de Sousa, S. Afewerki, F. R. Marciano, M. A. F. Corat, M. M. M. de Paula, T. F. C. L. Verde, M. C. M. Silva and A. O. Lobo, *Colloids Surf., B*, 2021, **205**, 111850.
- 137 G. F. Sousa, S. Afewerki, D. Dittz, F. E. P. Santos, D. O. Gontijo, S. R. A. Scalzo, A. L. C. Santos, L. C. Guimaraes, E. M. Pereira, L. S. Barcelos, S. J. H. Do Monte, P. P. G. Guimaraes, F. R. Marciano and A. O. Lobo, *J. Funct. Biomater.*, 2022, **13**, 45.
- 138 A. Famili and K. Rajagopal, *Mol. Pharmaceutics*, 2017, **14**, 1961–1968.
- 139 B. Zewde, O. Atoyebi, A. Gugssa, K. J. Gaskell and D. Raghavan, *ACS Omega*, 2021, **6**, 11614–11627.
- 140 C. E. Ziegler, M. Graf, M. Nagaoka, J. Groner, R. Mietzner, M. Breunig and A. M. Goepferich, *Mol. Pharm.*, 2023, **20**, 2465–2476.
- 141 S. Li, J. Zhou, Y. Huang, J. Roy, N. Zhou, K. Yum, X. Sun and L. Tang, *Acta Biomater.*, 2020, **110**, 95–104.
- 142 S. Hong, J. Carlson, H. Lee and R. Weissleder, *Adv. Healthcare Mater.*, 2016, **5**, 421–426.
- 143 S. T. Koshy, R. M. Desai, P. Joly, J. Li, R. K. Bagrodia, S. A. Lewin, N. S. Joshi and D. J. Mooney, *Adv. Healthcare Mater.*, 2016, **5**, 541–547.
- 144 H. Zhan, H. de Jong and D. W. P. M. Löwik, *ACS Appl. Bio Mater.*, 2019, **2**, 2862–2871.
- 145 Y. Zhang, H. Chen, T. Zhang, Y. Zan, T. Ni, M. Liu and R. Pei, *Biomater. Sci.*, 2018, **6**, 2578–2581.
- 146 S. H. Park, J. Y. Park, Y. B. Ji, H. J. Ju, B. H. Min and M. S. Kim, *Acta Biomater.*, 2020, **117**, 108–120.
- 147 S. H. Park, J. Y. Seo, J. Y. Park, Y. B. Ji, K. Kim, H. S. Choi, S. Choi, J. H. Kim, B. H. Min and M. S. Kim, *NPG Asia Mater.*, 2019, **11**, 1–16.
- 148 Y. Zhang, M. Liu and R. Pei, *Mater. Adv.*, 2021, **2**, 4733–4742.
- 149 J. Seo, S. H. Park, M. J. Kim, H. J. Ju, X. Y. Yin, B. H. Min and M. S. Kim, *ACS Appl. Mater. Interfaces*, 2019, **11**, 24984–24998.
- 150 H. J. Ju, M. Park, J. H. Park, G. R. Shin, H. S. Choi, M.-W. Suh and M. S. Kim, *Materials*, 2020, **13**, 3070.
- 151 J. Luo, A. Darai, T. Pongkulapa, B. Conley, L. Yang, I. Han and K.-B. Lee, *Bioact. Mater.*, 2023, **23**, 551–562.
- 152 X. Liu, Y. Zhang, Z. Hussain, P. Zheng, M. Xu, H. Zhao, Y. Liu, Y. Cao, I. Ullah, A. Osaka and R. Pei, *Appl. Mater. Today*, 2023, **30**, 101693.
- 153 J. Y. Heo, J. H. Noh, S. H. Park, Y. B. Ji, H. J. Ju, D. Y. Kim, B. Lee and M. S. Kim, *Pharmaceutics*, 2019, **11**, 438.
- 154 M. Czuban, S. Srinivasan, N. A. Yee, E. Agustin, A. Koliszak, E. Miller, I. Khan, I. Quinones, H. Noory, C. Motola, R. Volkmer, M. Di Luca, A. Trampuz, M. Royzen and J. M. M. Oneto, *ACS Cent. Sci.*, 2018, **4**, 1624–1632.
- 155 T. T. Vu, S. Yadav, O. S. Reddy, S.-H. Jo, S.-B. Joo, B. K. Kim, E. J. Park, S.-H. Park and K. T. Lim, *Pharmaceutics*, 2023, **16**, 841.
- 156 Z. Zhang, C. He and X. Chen, *Polymers*, 2020, **12**, 884.
- 157 I. Ali, A. Rizwan, T. T. Vu, S.-H. Jo, C.-W. Oh, Y. H. Kim, S.-H. Park and K. T. Lim, *Int. J. Biol. Macromol.*, 2024, **260**, 129549.
- 158 A. Rizwan, M. Gulfam, S.-H. Jo, J.-W. Seo, I. Ali, T. T. Vu, S.-B. Joo, S.-H. Park and K. T. Lim, *Eur. Polym. J.*, 2023, **191**, 112019.
- 159 T. T. Vu, M. Gulfam, S.-H. Jo, S.-H. Park and K. T. Lim, *Carbohydr. Polym.*, 2022, **278**, 118964.
- 160 D. S. B. Anugrah, K. Ramesh, M. Kim, K. Hyun and K. T. Lim, *Carbohydr. Polym.*, 2019, **223**, 115070.
- 161 I. Ali, M. Gulfam, S.-H. Jo, J.-W. Seo, A. Rizwan, S.-H. Park and K. T. Lim, *Int. J. Biol. Macromol.*, 2022, **219**, 109–120.
- 162 S. A. P. Siboro, D. S. B. Anugrah, K. Ramesh, S.-H. Park, H.-R. Kim and K. T. Lim, *Carbohydr. Polym.*, 2021, **260**, 117779.
- 163 Y.-J. Jo, M. Gulfam, S.-H. Jo, Y.-S. Gal, C.-W. Oh, S.-H. Park and K. T. Lim, *Carbohydr. Polym.*, 2022, **286**, 119303.
- 164 F. Jivan, R. Yegappan, H. Pearce, J. K. Carrow, M. McShane, A. K. Gaharwar and D. L. Alge, *Biomacromolecules*, 2016, **17**, 3516–3523.
- 165 F. Jivan, N. Fabela, Z. Davis and D. L. Alge, *J. Mater. Chem. B*, 2018, **6**, 4929–4936.
- 166 S. Jung, J. H. Abel, J. L. Starger and H. Yi, *Biomacromolecules*, 2016, **17**, 2427–2436.
- 167 E. Y. Liu, S. Jung, D. A. Weitz, H. Yi and C.-H. Choi, *Lab Chip*, 2018, **18**, 323–334.
- 168 D. Mirta, E.-T. Kang and K. G. Neoh, *ACS Appl. Polym. Mater.*, 2021, **3**, 2233–2263.
- 169 T. N. Gevrek, I. Kosif and A. Sanyal, *ACS Appl. Mater. Interfaces*, 2017, **9**, 27946–27954.
- 170 T. N. Gevrek, K. Yu, J. N. Kizhakkedathu and A. Sanyal, *ACS Appl. Polym. Mater.*, 2019, **1**, 1308–1316.
- 171 A. Degirmenci, G. Y. Bas, R. Sanyal and A. Sanyal, *Bioconjugate Chem.*, 2022, **33**, 1672–1684.
- 172 J. Escorihuela, A. T. M. Marcelis and H. Zuilhof, *Adv. Mater. Interfaces*, 2015, **2**, 1500135.
- 173 A. Shahrokhtash and D. S. Sutherland, *ACS Appl. Mater. Interfaces*, 2024, **16**, 21534–21545.
- 174 O. Røling, A. Mardjukov, S. Lamping, B. Vönhören, S. Rinnen, H. F. Arlinghaus, A. Studera and B. J. Ravoo, *Org. Biomol. Chem.*, 2014, **12**, 7828–7835.
- 175 N. Ozbek, E. L. Sahkulubey Kahveci and M. U. Kahveci, *ACS Appl. Polym. Mater.*, 2021, **3**, 3721–3732.



- 176 M. Pagel, R. Hassert, T. John, K. Braun, M. Wießler, B. Abel and A. G. Beck-Sickinger, *Angew. Chem., Int. Ed.*, 2016, **55**, 4826–4830.
- 177 M. Czuban, M. W. Kulka, L. Wang, A. Koliszak, K. Achazi, C. Schlaich, I. S. Donskyi, M. Di Luca, J. M. M. Oneto, M. Royzen, R. Haag and A. Trampuz, *Mater. Sci. Eng., C*, 2020, **116**, 111109.
- 178 T. N. Gevrek, A. Degirmenci, R. Sanyal and A. Sanyal, *Polymers*, 2020, **12**, 1211.
- 179 O. I. Kalaoglu-Altan, R. Sanyal and A. Sanyal, *ACS Omega*, 2019, **4**, 121–129.
- 180 D. D. Iorio, A. Marti, S. Koeman and J. Huskens, *RSC Adv.*, 2019, **9**, 35608–35613.
- 181 O. J. G. M. Goor, J. E. P. Brouns and P. Y. W. Dankers, *Polym. Chem.*, 2017, **8**, 5228–5238.
- 182 M. Putti, S. M. J. de Jong, O. M. J. A. Stassen, C. M. Sahlgren and P. Y. W. Dankers, *ACS Appl. Polym. Mater.*, 2019, **1**, 2044–2054.

