



Cite this: *Soft Matter*, 2023,  
19, 2339

# Influence of solute association on the phase behavior of 12-hydroxystearic acid/*n*-alkane solutions†

Tzu-Yu Lai,<sup>‡a</sup> Fardin Khabaz<sup>ID ab</sup> and Kevin A. Cavicchi<sup>ID \*a</sup>

The phase behavior of 12-hydroxystearic acid (12-HSA) in even-numbered alkanes ranging from octane (C<sub>8</sub>) to hexatriacontane (C<sub>36</sub>) was measured by visual observation of liquid + solid to liquid and liquid–liquid to liquid cloud points and liquid + solid to liquid + liquid transitions. In general solid phases were stabilized to low concentration and higher temperature with increasing alkane length. Liquid–liquid immiscibility was observed in larger alkanes starting with octadecane. The liquidus lines of shorter alkanes (octane to hexadecane) showing only liquid to liquid + solid transitions were fit with an attenuated associated solution model based on the Flory–Huggins lattice model assuming that 12-HSA forms a carboxylic acid dimer over all concentrations investigated. The fit results show that 12-HSA forms associated structures with degrees of association ranging from 3.7–4.5 dimers in the neat 12-HSA. At low concentrations, the 12-HSA is dissociated into dimers, however the free energy cost of dissociation stabilizes the solid phase giving a sharp knee at low concentrations. The role of 12-HSA association in its phase behavior and gelation behavior are discussed. More broadly, the importance of solute association in small molecule organogelators and its potential as a molecular design parameter similar to other component thermodynamic parameters, such as melting temperature and heat of fusion, is discussed.

Received 4th January 2023,  
Accepted 27th February 2023

DOI: 10.1039/d3sm00013c

rsc.li/soft-matter-journal

## Introduction

Molecular organogelators are small molecules that gel or immobilize an organic fluid by their self-assembly into structure spanning, solid networks, at low concentrations. This field has grown rapidly in the past three decades producing a wide-range of chemically distinct organogelators.<sup>1–5</sup> A large class of organogelators are molecular crystals that that gel a solution through nucleation and growth with preferential growth along a particular crystal axis to form three dimensional self-assembled fibrillar networks (SAFiNs).<sup>6,7</sup> The reversible nature of the non-covalent interactions governing the SAFiN formation allows for stimuli-responsive properties, such as the sol–gel transition. While thermally responsive gels dominate, there are

also examples of pH, light, anion/chemical, and mechano-responsive gels.<sup>8</sup> This broad combination of chemical motifs and stimuli has led to their application as rheological modifiers, structure directing agents, and sensors in a range of fields including plastics processing, separations, drug delivery, cosmetics, and remediation of crude oil spills.<sup>9–13</sup>

A long-standing challenge in the field of molecular organogels is the difficulty in predicting whether a molecule will gel a particular solvent.<sup>14</sup> This is most vexing in systems where seemingly minor structural variations made in either the solvent or the organogelator lead to large variations in the gel properties.<sup>15–17</sup> These cases highlight the complex interdependence of the solvent–solvent, solvent–organogelator, and organogelator–organogelator interactions on the structure of the organogelator, where it is difficult to independently tune these interactions and it therefore is an active area of study.<sup>3,18</sup> For example, there has been good success in correlating solvent parameters to gelation ability,<sup>19–22</sup> most notably with Hansen solubility parameters to map gelation spheres.<sup>23–28</sup>

Another avenue of study of the structure–property relationships of organogelators is the measurement of the phase behavior of different organogelator solutions.<sup>29</sup> For thermally-responsive organogels that form through the nucleation and growth of organogel fibers, strong correlations have been

<sup>a</sup> School of Polymer Science and Polymer Engineering, University of Akron, Akron, OH, 44325, USA. E-mail: kac58@uakron.edu

<sup>b</sup> Department of Chemical, Biomolecular, and Corrosion Engineering, University of Akron, Akron, OH, 44325, USA

† Electronic supplementary information (ESI) available: Document containing molar volume and solubility parameters of alkanes, example cloud-point measurements, stearic acid/dodecane phase behavior, and derivation of the attenuated associated solution model. Matlab files for curve fitting. Excel file of raw data for curve fitting. See DOI: <https://doi.org/10.1039/d3sm00013c>

‡ Current address: 3M Company, St. Paul, MN 55144, USA.

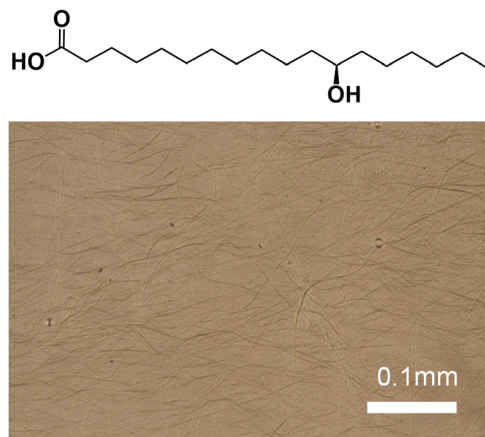


Fig. 1 (R)-12-Hydroxystearic Acid (12-HSA) and an optical micrograph of 5 vol% 12-HSA in docosane.

observed among the gel properties (*e.g.* gel transition temperature, minimum gelation concentration), the phase behavior (*e.g.* solubility or liquidus curve, miscibility gap), and the processing conditions (*e.g.* cooling rate and extent of undercooling/supersaturation).<sup>30–40</sup> In particular, conditions that produce sufficient undercooling<sup>30</sup> (or supersaturation<sup>36</sup>) at low organogelator concentration and direct liquid to liquid + solid phase transitions tend to favor gelation for “good” organogelators. Understanding the thermodynamics governing the phase behavior of an organogel provides a means to link the physical properties of the solution and the gel performance.

The focus of this paper is the solution phase behavior of *R*-12-hydroxystearic acid (12-HSA, Fig. 1) in a series of *n*-alkanes and the relationship to its gelation behavior. 12-HSA is known as an excellent organogelator that has a propensity to form fibrillar networks.<sup>41–50</sup> It is also an inexpensive, bio-derived material produced from the hydrogenation of ricinoleic acid from castor oil.<sup>51</sup> It has been used as an organogelator in a number of fields, such as food science,<sup>52</sup> cosmetics,<sup>53</sup> drug delivery,<sup>54,55</sup> and oil spill remediation.<sup>56</sup> An interesting property of 12-HSA solutions is the formation of aggregated or associated structures in the sol state, which has been observed in both experiment and simulation. The fitting of carbon NMR data of 12-HSA solutions in benzene by Sakurai *et al.* showed an equilibrium between monomer, dimer, and tetramer in the solution state.<sup>48</sup> Simulation of 12-HSA has shown that stable cyclic structures are formed from the dimerization of the acid head group and cyclization of six hydroxyl groups.<sup>57,58</sup> 12-HSA solutions should be associating solutions where the degree of association of 12-HSA with temperature and concentration influences the phase behavior and therefore should be a key factor in its gelation ability.<sup>59,60</sup> There have been numerous reports focusing on the influence of solvent–organogelator interactions in 12-HSA systems and the ability of polar solvent to disrupt organogelator–organogelator interaction is recognized.<sup>61–63</sup> However, to our knowledge, besides the work of Sakurai *et al.*, there are no direct investigations of the role solution state gelator–gelator interactions play in the solution phase behavior and gelation ability of 12-HSA.

As a model system, the phase behavior of 12-HSA in *n*-alkanes ranging from octane to hexatriacontane was investigated. Two reasons this system was chosen were, first, aliphatic alkanes do not have any functional groups that strongly compete with the hydrogen bonding in 12-HSA to limit concentration and solvent dependent solvent–gelator interactions. Second, the solubility parameter of *n*-alkanes varies over a small range, increasing at low carbon lengths and leveling off at higher carbon lengths, while the molar volume systematically increases (Fig. S1, ESI†). Consequently, this system is an excellent choice for isolating the effect of gelator–gelator interactions on the phase behavior of 12-HSA solutions.

The structure of this paper is as follows. First, the measurements of the phase behavior and the construction of phase diagrams are presented. Second, the liquidus lines are fit using an attenuated associating solution model that allows the variation of the associated structure of 12-HSA *vs.* concentration in solution to be assessed. A significant finding is that gelator association is a relevant factor at low concentrations where 12-HSA is used as an organogelator. Third, the role of gelator–gelator association in 12-HSA solutions is more broadly discussed based on the associated solution model.

## Experimental section

### Materials

Octane (>98% Alfa Aesar), decane (>99% TCI), dodecane (>99% TCI), hexadecane (>98% TCI), tetradecane (>99% TCI), hexadecane (≥98.5% Fisher Scientific), octadecane (99% Alfa Aesar), eicosane (>98% TCI), docosane (99% Acros), tetracosane (99% Aldrich), octacosane (99% Aldrich), dotriacontane (97% Aldrich), and hextriacontane (98% Aldrich) were used without further purification. 12-Hydroxystearic acid (12-HSA) with purity above 80.0% was purchased from TCI and further purified through repeated process of recrystallization by slowly cooling in hexane/ethyl acetate (19 : 1 *v* : *v*) cosolvent.<sup>41,64</sup> Stearic acid (>98% TCI) was also used as received. Solution samples were prepared by weighing out appropriate amounts of 12-HSA or stearic acid and solvent. The volume fractions of samples were calculated by converting the mass of each component to its volume using component volume = (mass)/(molar volume)/(molar mass) where the molar volume was calculated using group contribution method.<sup>65</sup>

### Differential scanning calorimetry (DSC)

DSC measurements were made using a DSC 8500 (PerkinElmer) under nitrogen atmosphere. Samples (~2–6 mg) were prepared and sealed in hermetic pans (DSC Consumables, Inc). Samples were initially cooled to 20 °C for 2 min then heated at a rate of 2 °C min<sup>−1</sup> to 100 °C. The molar heat of fusion ( $\Delta H_f^\circ$ ) of pure 12-HSA was determined calorimetrically by peak integration on cooling at 5 °C min<sup>−1</sup>. Melting points were taken from the endpoint of the melting endotherm.

### Tube inversion

Tube inversion measurements were used to measure the sol–gel transition temperature where the sample no longer was able to

support its own weight when inverted.<sup>66</sup> Additionally, the solubility was measured by recording the temperature at which the sol sample became optically clear. In the case of liquid–liquid phase separation, the cloud point ( $T_{cp}$ ) was determined as the temperature at which the sample became optically clear. To prepare samples for measurement, *ca.* 1.5 g of 12-HSA and solvent were loaded in 8 mL vials. Solutions were heated to 130 °C to form transparent and homogeneous solution, which were then air-cooled down to ambient temperature at the workbench.

The as-prepared samples were heated using a hot plate equipped with an aluminum heating block which can hold up to 40 samples at a time. Temperature ramps were started from 25 °C, at steps of 1 °C with annealing for 20 min after each temperature change. After annealing, sample vials were tilted to a 90° angle. The temperature at which flow behavior was first observed was recorded at  $T_{gel}$ . Two phenomena occurred above  $T_{gel}$ . Either the sample gradually turned optically transparent from the dissolution of the 12-HSA crystals or the sample underwent a transition to a liquid–liquid phase separated structure with droplets of one phase in the matrix of the second phase, followed by a transition to an optically transparent solution. The temperature where the solution turned homogeneously transparent was recorded as the cloud point,  $T_{cp}$ . The temperature where liquid–liquid phase separated was first observed was recorded as  $T_{L-L}$ . Each sample was tested through three heating ramps and the average values for each sample are presented. Identical sample preparation methods and tube inversed measurements were applied to solutions of stearic acid (SA) in dodecane.

### Cloud point apparatus

An OptiMelt MPA100 (Stanford Research System) was used to measure the melting point of the samples where a sharp increase in the optical transparency was recorded. To prepare samples a hot, transparent solution was transferred into a capillary tube and air-cooled down to the room temperature.

In the instrument, samples were heated at a rate of 2 °C min<sup>−1</sup> from ambient temperature to 120 °C.

## Results and discussion

### Phase diagrams of 12-HSA/alkane solutions

Three methods were used to determine the phase diagrams of 12-HSA in different *n*-alkanes, visual observations during tube inversion measurements, automated cloud point measurements, and differential scanning calorimetry (DSC). Fig. 2 displays the transitions temperatures recorded as function of volume fraction of 12-HSA in two solutions, 12-HSA/decane and 12-HSA/octadecane.  $T_{gel}$  was measured by tube inversion as the temperature at which the sample exhibited flow when tilted 90° from vertical. Technically, gels are only formed at low concentration of 12-HSA where the sample is primarily composed of the solvent. Rather than try to differentiate the samples based on this criterion  $T_{gel}$  is used to delineate the rheological observation of the transition where the self-supporting solid structure of the 12-HSA disappears.  $T_{cp}$  is the cloud point temperature above which the sample was homogeneous and optically transparent as determined either by visual inspection or by the cloud point apparatus.  $T_m$  is the endpoint of the endothermic melting peak measured by DSC, corresponding to the dissolution of the 12-HSA crystals.  $T_{L-L}$  is the temperature above which only a two-phase liquid was observed by visual inspection. In decane, these measurements gave qualitatively similar curves, corresponding to the liquidus line separating the one phase liquid from the two phase solid + liquid regions. While there is some spread in the temperatures among the different curves it is only *ca.* 3–6 °C, with the largest spread at small 12-HSA volume fractions. The  $T_{gel}$  curve is generally lower than the others, which is to be expected as the mechanical failure of the sample would occur during partial dissolution of the 12-HSA. The  $T_{cp}$  agree well with that gathered from the cloud point apparatus (raw data in Fig. S2, ESI†) and visual inspection.

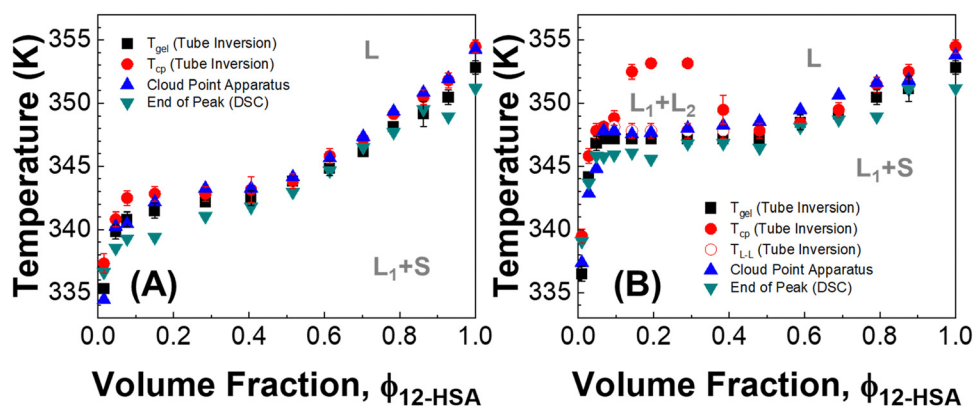


Fig. 2 Phase diagram of (A) 12-HSA/decane and (B) 12-HSA/octadecane. The transition temperatures correspond to  $T_{gel}$ : the temperature when samples started to flow or fractured during tube inversion; End of Peak ( $T_m$ ): the end of melting peak measured by DSC;  $T_{cp}$  the temperature when sample became homogeneous and optically transparent; and  $T_{L-L}$ : the temperature above which liquid–liquid phase separation was observed. The instrument or technique used to measure each point is labeled in the key on each plot.

In the 12-HSA/octadecane system a liquid–liquid miscibility gap is observed as an additional feature. The cloud-point measurement of the miscibility gap is only observed with the visual cloud point measurement. In the cloud point apparatus, the detected cloud point corresponds to the melting of the solid 12-HSA. Under the miscibility gap, the transition temperature for 12-HSA is roughly invariant as would be expected for a monotectic phase diagram. As the visual results obtained during tube inversion measurements are consistent with these other measurements and offer measurements of both dissolution and liquid–liquid phase separation in a single experiment, this method was used as the primary route to determine the phase diagrams in the other 12-HSA/*n*-alkane systems.

The DSC heating traces of samples prepared in decane and octadecane are shown in Fig. 3. In decane, multiple endothermic peaks are observed above a volume fraction of  $\phi_{12\text{-HSA}} = 0.14$ . The most likely origin for this phenomenon is a solid–solid transition. Previously, transition between different 12-HSA polymorphs were observed by Takeno and co-workers in a study of the phase behavior of 12-HSA in phenyl methyl silicone.<sup>67</sup> Using small angle X-ray scattering and polarized optical microscopy, three crystal forms were identified (form I, form II, and form III). An enantiotropic Gibbs free energy profile was proposed where form I is thermodynamically stable at low temperature ( $< ca. 70^\circ\text{C}$ ) and form II is stable at higher temperature ( $> ca. 70^\circ\text{C}$ ). Form III was proposed to be unstable at any temperature. The energy barrier between form III and form I was smaller than the energy barrier between form I and form II, which may explain its metastable formation. Similar behavior was observed in 12-HSA/ionic liquid solutions where the same three crystal forms was observed and the solid–solid transition between form I and form II was observed at  $70\text{--}75^\circ\text{C}$ .<sup>68</sup>

Similar DSC behavior to the 12-HSA/decane solution is observed in the 12-HSA/octadecane DSCs. It appears that the solid–solid transition temperature is below the monotectic temperature ( $T_{\text{mt}} \approx 75^\circ\text{C}$ ). Therefore, there should be an additional peak in the DSC trace at compositions to the right of the monotectic point due to melting at the monotectic

temperature and the liquidus temperature.<sup>69–71</sup> Based on the composition where single melting peaks first appear on moving from high to low composition, the low temperature stable phase of 12-HSA appears to be stable below  $66^\circ\text{C}$  in decane and  $72^\circ\text{C}$  in octadecane. One source for this temperature difference is kinetic hindrance in the solid–solid transition as measured by DSC.<sup>72</sup> Therefore, the lower solubility of 12-HSA in octadecane compared to decane may result in higher kinetic hindrance and a higher observed solid–solid transition temperature. For example, in stearic acid, which is structurally similar to 12-HSA, similar solid–solid transition temperature was measured in decane, methanol, and butanone by solubility measurements, but a higher transition temperature was measured by DSC in decane.<sup>73,74</sup>

The exact origin of these solid–solid transitions is not clear. As detailed in a recent review by Guenet, solid–solid transitions can be categorized into two main types.<sup>7</sup> In the first case, the solid–solid transition occurs due to the formation of an alloy or a molecular compound below the peritectic temperature. In the absence of a solid solution phase containing both the solvent and solute at higher composition, this would result in an invariant melting temperature with solute composition. If this occurred in the phase diagrams shown in Fig. 3, they would be consistent with the formation of an incongruently melting compound where at solute concentrations below the stoichiometric composition of the compound this crystal melts to form a solid phase plus a liquid phase rather than melting directly to the liquid in the case a congruently melting compound. In the second case, there is a metatectic transition where the solid undergoes a solid–solid phase transition at the metatectic temperature. This would also give an invariant solid–solid transition temperature, however the likely presence of a solid–solution at higher solute concentration would give a solidus line below the liquidus resulting in the melting temperature increasing with solute composition and three melting endotherms could be detected on heating. At low temperatures below the peritectic and metatectic temperatures, and therefore low solute concentration, the liquid would directly solidify into the molecular compound or second solid phase, different from

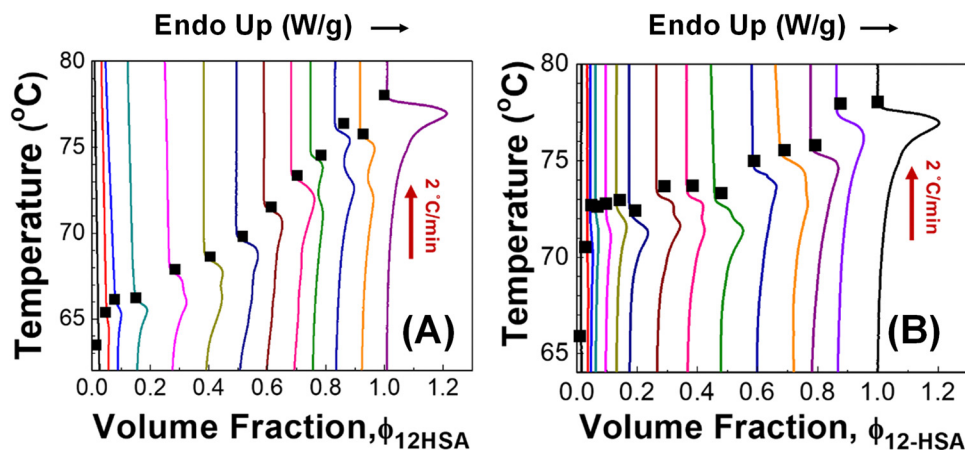


Fig. 3 DSC heating traces of (A) 12-HSA/decane and (B) 12-HSA/octadecane systems.



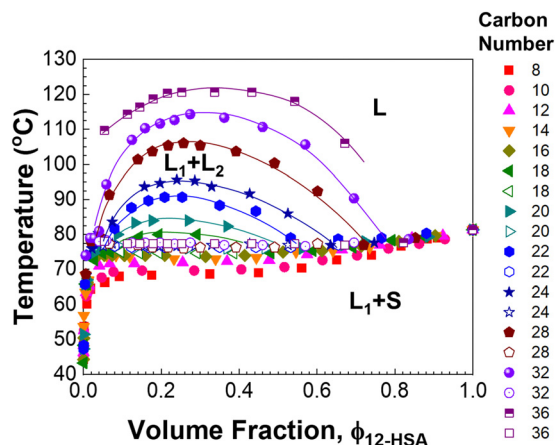


Fig. 4 Phase diagram of 12-HSA/*n*-alkanes mixtures (*n* = 8–36) measured by tube inversion. Solid symbols indicate the phase transition from two components (solid–liquid or liquid–liquid) to one component, and open symbols indicate the solid–liquid to liquid–liquid phase transition. The solid lines on the binodal are guides to the eye. The 12-HSA/dodecane data was previously reported in ref. 29. Adapted with permission from ref. 29. Copyright 2018 American Chemical Society.

the bulk crystal stable just below the neat compound's melting temperature.

More detailed DSC, optical microscopy and potentially scattering measurements would be needed to investigate the solid–solid phase transitions more fully to delineate between the type of solid–solid transition and determine the solidus line.<sup>75,76</sup> The thermodynamic modeling in the next section assumes a simpler system where the solid–solid transitions are not considered. The implications of this approximation on the fitting results are discussed.

Fig. 4 displays the liquidus lines and miscibility gaps measured by visual inspection of 12-HSA in alkanes ranging from octane to hexatriacontane. A variation is observed in the positions of the miscibility gap and liquidus line with the length of the alkanes (carbon number, *n*). The plateau temperature at intermediate concentration moves up with an increasing length of the alkane. At octadecane, a miscibility gap appears, with a composition invariant melting temperature beneath it. This miscibility gap grows as the length of the alkane increases. As the solubility parameter difference between the alkanes and 12-HSA ( $\delta_{12\text{-HSA}} = 18.1 \text{ MPa}^{1/2}$ )<sup>61</sup> decreases with increasing alkane length (see Fig. S1, ESI<sup>†</sup>), the increasing liquid–liquid immiscibility is likely driven by the lower entropy of mixing with increasing alkane length, such as is observed in polymer solutions. Interestingly the miscibility gap is asymmetric with its maximum at  $\phi_{12\text{-HSA}} < 0.5$ . This is similar to what is observed in polymer solutions and associated solutions where the molar volume of the solute is greater than the solvent.<sup>77</sup> However, it is difficult to fit the binodal line defining the miscibility gap analytically. Therefore more quantitative investigation of the phase behavior focuses on the solutions in hexadecane and shorter alkanes that do not show liquid–liquid phase separation above the liquidus line.

### Solution model fitting of the liquidus line

An approach to further investigate the solution state structure of 12-HSA is to fit the liquidus lines to a thermodynamic solution model. The liquidus line separates the pure liquid phases from the mixed liquid and solid phases. At the temperatures and compositions on the liquidus line the chemical potential of 12-HSA is equal in the liquid and solid phase. Assuming that 12-HSA forms pure crystals in the solid-state (*i.e.* no solid solution phase), the enthalpy and entropy of fusion are not temperature dependent, the interfacial tension between the liquid and solid phases is small, the crystals are large, and the crystals form the same polymorph as the neat 12-HSA, the chemical potential of the liquid at composition  $\phi_{12\text{-HSA}}$ ,  $\mu_{12\text{-HSA}}(\text{l})$  is equal to the chemical potential of the pure solid,  $\mu_{12\text{-HSA}}^{\circ}(\text{s})$ , taken as the standard state of the solid. The difference in the chemical potential between the 12-HSA in the liquid at composition,  $\phi_{12\text{-HSA}}$ ,  $\mu_{12\text{-HSA}}(\text{l})$ , and the neat 12-HSA liquid,  $\mu_{12\text{-HSA}}^{\circ}(\text{l})$ , taken as the standard state, is,

$$\begin{aligned}\Delta\mu_{12\text{-HSA}}(\text{l}) &= \mu_{12\text{-HSA}}(\text{l}) - \mu_{12\text{-HSA}}^{\circ}(\text{l}) \\ &= \mu_{12\text{-HSA}}^{\circ}(\text{s}) - \mu_{12\text{-HSA}}^{\circ}(\text{l}) \\ &= -\Delta H_{\text{f},12\text{-HSA}}^{\circ} T_{\text{m}} \left( \frac{1}{T_{\text{m}}} - \frac{1}{T_{\text{m}}^{\circ}} \right)\end{aligned}\quad (1)$$

where *R* is the gas constant,  $\Delta H_{\text{f},12\text{-HSA}}^{\circ}$  is the heat of fusion of the neat 12-HSA,  $T_{\text{m}}$  is the melting point of 12-HSA at composition  $\phi_{12\text{-HSA}}$ , and  $T_{\text{m}}^{\circ}$  is the melting point of the neat 12-HSA.<sup>78</sup>

This chemical potential difference,  $\Delta\mu_{12\text{-HSA}}(\text{l})$ , may also be found from the free energy of mixing of the one-phase liquid. As it is assumed that the 12-HSA molecules can associate through the hydrogen bonding of carboxylic acid groups to form dimers and the alcohol units to form cyclic structures an associating solution model should be used for the free energy of mixing. The lattice model used for an associating solution was first derived by Flory for treating a polymer solution with a molecular weight dispersity<sup>79</sup> and was applied to alcohol–hydrocarbon mixtures.<sup>80</sup> More recently, this model was applied to the solubility of asphaltenes in oil, which has some similarity to 12-HSA/alkane systems, motivating the use of this model.<sup>77,81–83</sup> While the main equations are discussed below, the full derivation of this model following previous references is given in the ESI.<sup>†</sup><sup>77,79–81,84</sup> For solutions where 12-HSA forms associated aggregates in a non-associating solvent,  $\Delta\mu_{12\text{-HSA}}(\text{l})$  is given by,

$$\begin{aligned}\Delta\mu_{12\text{-HSA}}(\text{l}) &= RT \left( \ln \left( \frac{\phi_{12\text{-HSA}_1}}{\phi_{12\text{-HSA}_1}^{\circ}} \right) - \frac{r_{12\text{-HSA}_1} V_{\text{r}}}{V} + \frac{r_{12\text{-HSA}_1} V_{\text{r}}}{V^{\circ}} + r_{12\text{-HSA}_1} \phi_{\text{SZ}}^2 \right)\end{aligned}\quad (2)$$

where  $\phi_{12\text{-HSA}_1}$  is the volume fraction of the 12-HSA monomer,  $\phi_{12\text{-HSA}_1}^{\circ}$  is the volume fraction of 12-HSA monomer in the neat 12-HSA liquid,  $r_{12\text{-HSA}_1}$  is the ratio of the molar volume of the

12-HSA monomer,  $V_{12\text{-HSA}_1}$  to a reference lattice cell volume,  $V_r$ ,  $V$  is the volume of 1 true mole of solution,  $V^\circ$  is the molar volume of the neat 12-HSA liquid,  $\phi_s$  is the volume fraction of the solvent, and  $\chi$  is the Flory-Huggins interaction parameter. To compare eqn (1) and (2), the standard state,  $\mu_{12\text{-HSA}}^\circ(1)$ , is the pure 12-HSA liquid. 12-HSA is a supramolecular polymer with a molecular weight distribution with an average degree of association. On mixing with the solvent, the degree of association of 12-HSA will shift with temperature and composition to minimize the free energy. The first three terms in the parentheses on the right-hand side of eqn (2) describe the partial molar entropy of mixing and the partial molar free energy change from breaking the hydrogen bonds. It should be noted that the composition is expressed in terms of the monomer volume fraction rather than the 12-HSA volume fraction. This simplification is allowed from the results of Prigogine where the chemical potential of component A ( $\mu_A$ ) is equal to the chemical potential of its monomer ( $\mu_{A1}$ ).<sup>85</sup> The last term on the right hand side describes the non-specific enthalpic interactions between the liquid 12-HSA and solvent using the standard  $\chi$  parameter from the Flory-Huggins model given by,

$$\chi = \frac{V_r}{RT}(\delta_{12\text{-HSA}} - \delta_s)^2 + B = \frac{A}{T} + B \quad (3)$$

where  $\delta_{12\text{-HSA}}$  and  $\delta_s$  are the solubility parameters of 12-HSA and the solvent, respectively.  $B$  is an empirical parameter that is typically *ca.* 0.3.<sup>86</sup>

Combining eqn (1) and (2) and rearranging gives an equation for the liquidus line as,

$$\frac{1}{T_{m,12\text{-HSA}}} - \frac{1}{T_{m,12\text{-HSA}}^\circ} = -\frac{R}{\Delta H_{f,12\text{-HSA}}^\circ} \left( \ln \left( \frac{\phi_{12\text{-HSA}_1}}{\phi_{12\text{-HSA}_1}^\circ} \right) - \frac{r_{12\text{-HSA}_1} V_r}{V} + \frac{r_{12\text{-HSA}_1} V_r}{V^\circ} + r_{12\text{-HSA}_1} \phi_s^2 \chi \right) \quad (4)$$

To fit the experimental liquidus lines a specific association model is needed for the variation of  $\phi_{12\text{-HSA}_1}$  vs.  $\phi_s$  and the value of  $\phi_{12\text{-HSA}_1}^\circ$ . A standard model is a continuous association model where the association of a monomer A with an aggregate of degree of polymerization  $i$  is described as an equilibrium reaction,<sup>77</sup>



Assuming that reactivity is not dependent on the size of the aggregate, the equilibrium constant,  $K$ , is given in terms of the

volume fraction of the components as,

$$K = \frac{\phi_{A_{i+1}}}{\phi_{A_i} \phi_{A_1}} \frac{i}{i+1} \quad (6)$$

In the case of 12-HSA there is a limit to the size of the associated aggregate as the carboxylic acid groups tend to form dimers while the alcohol groups form cyclic structures. A simple model to use in this case is an attenuated association model where  $K$  is reduced with increasing aggregate size by dividing the left-hand side of eqn (6) by  $i+1$  to give,

$$\frac{K}{i+1} = \frac{\phi_{12\text{-HSA}_{i+1}}}{\phi_{12\text{-HSA}_i} \phi_{12\text{-HSA}_1}} \frac{i}{i+1} \quad (7)$$

This equation gives a general equation for  $\phi_{12\text{-HSA}_i}$  in terms of the equilibrium constant and 12-HSA monomer concentration as,

$$\phi_{12\text{-HSA}_i} = \frac{K^{i-1} \phi_{12\text{-HSA}_1}^i}{(i-1)!} \quad (8)$$

The overall volume fraction of 12-HSA is,

$$\begin{aligned} \phi_{12\text{-HSA}} &= \sum_{i=1} \phi_{12\text{-HSA}_i} = \sum_{i=1} \frac{K^{i-1} \phi_{12\text{-HSA}_1}^i}{(i-1)!} \\ &= \phi_{12\text{-HSA}_1} \exp(K \phi_{12\text{-HSA}_1}) \end{aligned} \quad (9)$$

Eqn (9) allows the second and third terms in the parentheses on the right-hand side of eqn (4) to be written in terms of  $K$ ,  $\phi_{12\text{-HSA}_1}$ , and  $\phi_{12\text{-HSA}_1}^\circ$  as,

$$\begin{aligned} \frac{r_{12\text{-HSA}_1} V_r}{V} &= \sum_{i=1} \frac{\phi_{12\text{-HSA}_i}}{i} + \frac{r_{12\text{-HSA}_1} \phi_s}{r_s} \\ &= \frac{1}{K} (\exp(K \phi_{12\text{-HSA}_1}) - 1) + \frac{r_{12\text{-HSA}_1} \phi_s}{r_s} \end{aligned} \quad (10)$$

where  $r_s$  is the ratio of the molar volume of the solvent to the reference volume ( $V_s/V_r$ ) and,

$$\frac{r_{12\text{-HSA}_1} V_r}{V^\circ} = \frac{1}{K} (\exp(K \phi_{12\text{-HSA}_1}^\circ) - 1) \quad (11)$$

Substituting eqn (10) and (11) into eqn (4) gives,

$$\begin{aligned} \frac{1}{T_{m,12\text{-HSA}}} - \frac{1}{T_{m,12\text{-HSA}}^\circ} &= -\frac{R}{\Delta H_{f,12\text{-HSA}}^\circ} \left( \ln \left( \frac{\phi_{12\text{-HSA}_1}}{\phi_{12\text{-HSA}_1}^\circ} \right) - \frac{1}{K} \exp(K \phi_{12\text{-HSA}_1}) \right. \\ &\quad \left. - \frac{r_{12\text{-HSA}_1} \phi_s}{r_s} + \frac{1}{K} \exp(K \phi_{12\text{-HSA}_1}^\circ) + r_{12\text{-HSA}_1} \phi_s^2 \chi \right) \end{aligned} \quad (12)$$

Using  $\chi = A/T + B$  and rearranging eqn (12) to solve for  $T_{m,12\text{-HSA}}$  gives,

$$T_{m,12\text{-HSA}} = \frac{\left( \frac{\Delta H_{f,12\text{-HSA}}^\circ}{R} + r_{12\text{-HSA}_1} \phi_s^2 A \right)}{\frac{1}{K} \exp(K \phi_{12\text{-HSA}_1}) - \frac{1}{K} \exp(K \phi_{12\text{-HSA}_1}^\circ) - \ln \left( \frac{\phi_{12\text{-HSA}_1}}{\phi_{12\text{-HSA}_1}^\circ} \right) + \frac{r_{12\text{-HSA}_1} \phi_s}{r_s} - r_{12\text{-HSA}_1} \phi_s^2 B + \frac{\Delta H_{f,12\text{-HSA}}^\circ}{RT_{m,12\text{-HSA}}^\circ}} \quad (13)$$

Table 1 Liquidus line fitting parameters

Compound	$\delta^a$ (MPa <sup>1/2</sup> )	$V^b$ (cm <sup>3</sup> mol <sup>-1</sup> )	$\Delta H_f^{\circ c}$ (kJ mol <sup>-1</sup> )	$T_m^{\circ d}$ (K)	M (g mol <sup>-1</sup> )
Octane	15.5	163.6	—	—	114.23
Decane	15.7	195.8	—	—	142.28
Dodecane	16.0	228.0	—	—	170.33
Tetradecane	16.2	260.2	—	—	198.39
Hexadecane	16.3	292.4	—	—	226.44
12-HSA	18.1	312.5	50.45	354.5	300.48
Stearic acid	17.5	319.6	64.28	344.2	284.48

<sup>a</sup> Previously reported values for all compounds except tetradecane, which was calculated by the group contribution method using the values of Fedors.<sup>61,65</sup> <sup>b</sup> Calculated by group contribution methods using the values of Fedors.<sup>65</sup> <sup>c</sup> Previously reported values measured by DSC on heating for moles of molecules (*i.e.* unimers).<sup>35</sup> <sup>d</sup> Measured by visual cloud point measurement.

Matlab was used to non-linear least squares fit eqn (13) with  $K$  and  $B$  as the two fitting parameters. Example Matlab code and input data for 12-HSA in octane is included in the ESI.† To better match the attenuation model, the 12-HSA 'monomer' was assumed to be the carboxylic acid dimer with twice the molar volume of 12-HSA ( $V_{12\text{-HSA}_1} = 2V_{12\text{-HSA}}$ ).  $\Delta H_{f,12\text{-HSA}_1}^\circ$  was also calculated using the moles of 12-HSA dimer ( $\Delta H_{f,12\text{-HSA}_1}^\circ(\text{dimer}) = 2\Delta H_{f,12\text{-HSA}_1}^\circ(\text{unimer})$ ). This is driven by literature reports that the equilibrium constant for carboxylic acid dimers (*e.g.* 3200 L mol<sup>-1</sup> acetic acid in hexane at 25 °C)<sup>87</sup> is much greater than that of alcohol dimers (12.5 L mol<sup>-1</sup> for 1-hexanol in alkane at 25 °C).<sup>88</sup> Therefore,

by assuming that 12-HSA forms dimers over the entire concentration range, the attenuation model captures the variation in alcohol association with concentration. This assumption was tested by measuring the liquidus line of stearic acid in dodecane and fitting  $T_m$  vs. the volume fraction of stearic acid assuming the stearic acid was a dimer over the entire concentration range with  $B$  as the only fitting parameter. The stearic acid fitting is shown in the ESI† (Fig. S3). A reasonable fit is obtained with  $B = 0$ . The fit overpredicts the melting point at the lowest concentration by *ca.* 5 K. We believe this is mostly due to the uncertainty in the solubility limit at this low temperature, rather than a low equilibrium constant for dimerization. The fitting constants are listed in Table 1. The fits for the 12-HSA data are shown in Fig. 5A ( $\phi_{12\text{-HSA}}$  vs.  $T_m$ ) and Fig. 5B ( $\ln \phi_{12\text{-HSA}}$  vs.  $1/T_m$ ),  $K$  and  $B$  are plotted in Fig. 5C. The number average degree of association of the 12-HSA dimers is given by,<sup>77</sup>

$$\bar{N}_{12\text{-HSA}_1} = \frac{\sum_{i=1}^{\infty} N_{12\text{-HSA}_1} i}{N_{12\text{-HSA}_1}} = \frac{\sum \phi_{12\text{-HSA}_i}}{\sum (\phi_{12\text{-HSA}_i}/i)} = \frac{\phi_{12\text{-HSA}} K}{(e^{K\phi_{12\text{-HSA}}} - 1)} \quad (14)$$

$\bar{N}_{12\text{-HSA}_1}$  vs.  $\phi_{12\text{-HSA}}$  is plotted in Fig. 5D.

The fits of the liquidus line are best for the shorter alkanes (octane, decane and dodecane). In tetradecane and hexadecane the liquidus line has a sinusoidal shape at intermediate concentration that is not captured as well with this fitting model. As the molar volume of the solvent increases, its

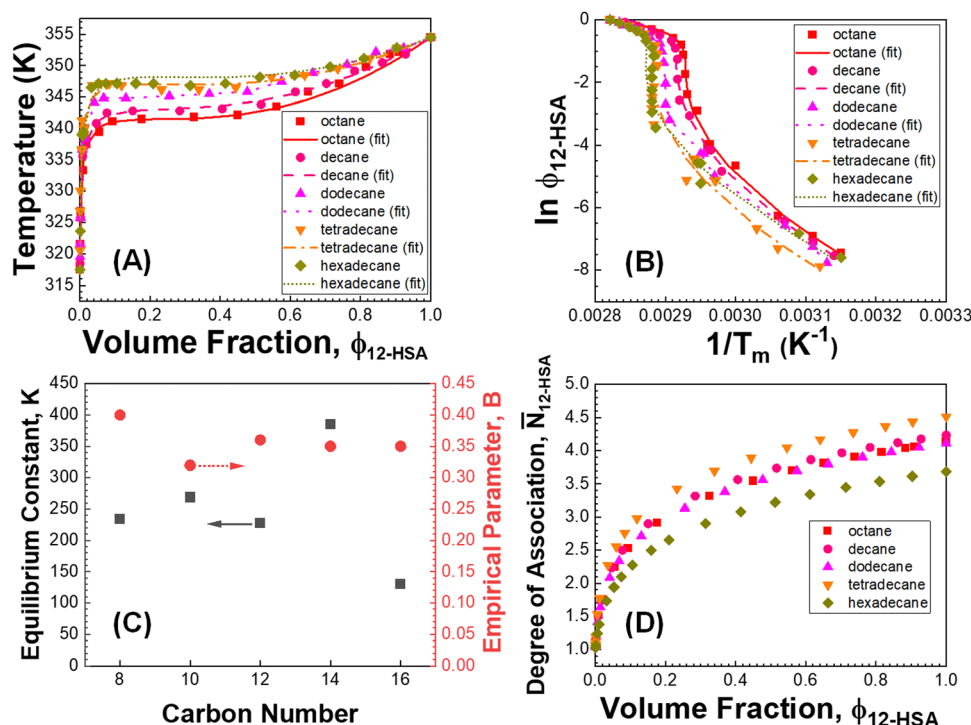


Fig. 5 (A and B) Fitting results of 12-HSA alkane solutions. (C) Fitting parameters vs. the number of backbone carbons in each alkane. (D) The degree of association of 12-HSA dimers vs. 12-HSA volume fraction in different alkanes. The 12-HSA/dodecane data in (A) was previously reported in ref. 29. Adapted with permission from ref. 29. Copyright 2018 American Chemical Society.

contribution to the entropy of mixing decreases, which magnifies the contributions of the 12-HSA to liquidus line. Therefore, the model is most challenged in these larger alkanes. More exact models could be used, for example using separate equilibrium constants for the carboxylic acid dimer, alcohol dimer and cyclic alcohol polymer.<sup>88</sup> However, the increased number of fitting constants makes it more difficult to find a unique local minima and are best applied when associated structures and equilibrium constants can be independently measured, such as by FTIR spectroscopy.<sup>88</sup> Three other simplifying assumptions of these fit are, first, the temperature dependence of  $K$  is not accounted for and therefore the fits give an average  $K$  over the temperature range. Second, the polymorph formation at low concentration is ignored. A lower value of  $T_{m,12-HSA}^{\circ}$ , a higher value of  $\Delta H_{f,12-HSA}^{\circ}$ , and potentially an additional interaction parameter is needed in this region where the lower melting point, higher heat of fusion polymorph is stable. The model assuming one solid polymorph is formed therefore likely underpredicts the liquidus line at lower 12-HSA concentration. It is possible that the molecular compound formation is more favorable in longer alkanes giving rise to more fitting error. Third, solid solution formation at high 12-HSA concentration would result in a solidus line below the liquidus line. This would introduce additional terms into eqn (13) due to the free energy of mixing in the solid solution that depend on the composition of the solidus line at the same temperature as the liquidus line and potentially a second interaction parameter between the liquid and solid phases.<sup>89,90</sup> As the solidus line is likely much steeper than the liquidus line, it would have a smaller composition variation, so that it could potentially be approximated as an additional constant term in eqn (13), which would be similar to allowing  $\Delta H_{f,12-HSA}^{\circ}$  or  $T_{m,12-HSA}^{\circ}$  to be used as a fitting term. The value of the lattice model is in finding reasonable agreement to the data to allow interpretation of the different component contributions to the phase behavior, where gross disagreement with the experimental and fit curves indicate more complexity and the need for less simplifying assumptions and a more complex model. The good agreement

of the fits with the data, especially in shorter alkanes, show that the model is adequate description of the solution behavior.

Examining the plots in Fig. 5D, the average degree of association at  $\phi_{12-HSA} = 1$  ranges from 3.7–4.5, with excellent agreement between octane (4.1) decane (4.2) and dodecane (4.1) with more variation in tetradecane (4.5) and hexadecane (3.7) in-line with fitting results. This an excellent order of magnitude agreement with the previous work of Gordon *et al.* showing 12-HSA dimers forms six-member rings,<sup>57</sup> and the work of Sakurai that fit the association of 12-HSA in benzene to a monomer, dimer, tetramer model.<sup>91</sup> In addition, the previous fitting of alcohol alkane solutions showed a degree of association ranging from 3–5 with lower degree of association for secondary alcohols compared to primary alcohols.<sup>88</sup> At lower concentrations of 12-HSA in the concentration range where gelation is typically studied the alcohol groups are not strongly associated, but the 12-HSA still primarily exist as carboxylic acid dimers based on the fitting model. An important feature of the association model is that the dissociation of the alcohol groups at low concentrations contributes to a higher entropy of mixing and stabilizes the liquid, there is also the free-energy cost of dissociation, which favors crystallization. This is the topic of the next section.

#### Role of self-association in 12-HSA solution phase behavior

The influence of the self-association of the 12-HSA is visualized in Fig. 6A by plotting the predicted  $T_m$  vs.  $\phi_{12-HSA}$  using eqn (13) for the 12-HSA/octane solution for  $B = 0.4$  and  $K = 234$  (original fit), 10 and 0.01 and the case where  $K = 0.01$  and the 12-HSA monomer is assumed to a single 12-HSA molecule (*i.e.* 12-HSA unimer) with half the molar volume and half the heat of fusion of the 12-HSA dimer. In Fig. 6B,  $\phi_{12-HSA_1}$  vs.  $\phi_{12-HSA}$  is plotted. In the case of  $K = 0.01$ ,  $\phi_{12-HSA_1} = \phi_{12-HSA}$  over the entire concentration range. Going from the 12-HSA unimer to the dimer at  $K = 0.01$  to higher values of  $K$ , the liquidus line shows a more persistent plateau at lower concentrations with a sharper knee where the melting point drops off at low concentrations. Therefore, it is clear that even though the degree of association

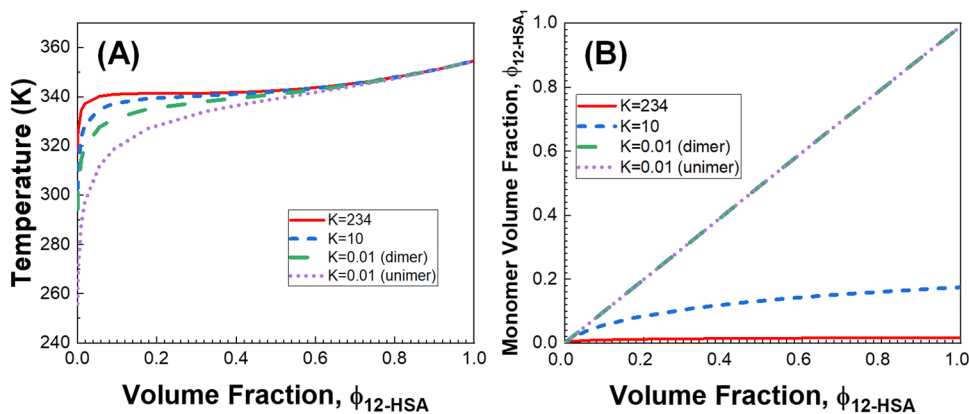


Fig. 6 (A) Predicted liquidus lines for 12-HSA in octane at  $K = 234$ , 10, and 0.01 for 12-HSA dimers and  $K = 0.01$  for a 12-HSA unimer using eqn (13) with  $B = 0.4$ ,  $\Delta H_{f,12-HSA}^{\circ} = 100\,900\text{ J mol}^{-1}$  (dimer) and  $50\,450\text{ J mol}^{-1}$  (unimer),  $V_{12-HSA} = 625\text{ cm}^3\text{ mol}^{-1}$  (dimer) and  $312.5\text{ cm}^3\text{ mol}^{-1}$  (unimer),  $V_s = 163.6\text{ cm}^3\text{ mol}^{-1}$ ,  $V_r = 163.6\text{ cm}^3\text{ mol}^{-1}$ . (B) Monomer volume fraction ( $\phi_{12-HSA_1}$ ) vs. 12-HSA volume fraction for the predicted liquidus lines shown in (A).



is low at low concentrations solute association still significantly influences the phase behavior. This also can be seen by examining eqn (13) where  $K$  and  $\phi_{12\text{-HSA}_1}^\circ$  are found in two concentration independent terms in the denominator (second and third terms). These terms are both negative and therefore act to increase  $T_m$ . They have a similar effect of a high bulk melting temperature,  $T_{m,12\text{-HSA}}^\circ$ , which also is in a concentration independent term in the denominator.

A likely key to the ability of 12-HSA to efficiently gel a wide-range of solvents is the balance achieved in its physical properties. While stronger directional interactions would raise  $T_m$  and stabilize the solid phase, this would also likely increase  $K$ , which increases the possibility of liquid–liquid phase separation as larger associated structures would persist to lower concentration. Liquid–liquid phase separation is typically not advantageous for gelation, as it forms dispersed organogelator rich phases, that lowers the concentration of the fine structure spanning organogelator fibers form from the liquid to liquid + solid phase transition.<sup>29</sup> Similarly, the size limiting effect of the cyclization of the hydroxyl groups also provides a significant free energy penalty from the disassociation of a large number of alcohol bonds, which stabilizes the liquidus line to lower concentrations, while not producing supramolecular polymers with a significantly large degree of association,<sup>92</sup> which could also drive liquid–liquid phase separation from the large effective molar volume of the supramolecular species.

This associated solution phase behavior also provides a rationale for why solvent–gelator interactions are able to influence the gelation behavior of 12-HSA. For example, Gao *et al.* examined the gelation behavior of 12-HSA in a number of different classes of solvents (*e.g.* alkanes, ketones, alcohols).<sup>61</sup> It was observed that in solvents with higher Hansen hydrogen bonding solubility parameters ( $\delta_h$ ) higher critical gelation concentrations were observed (*i.e.* the concentrations below which gels would not form on cooling a heated solution to room temperature). In addition, a shift from clear gels, to opaque gels to no gelation was observed with increasing  $\delta_h$ . In these systems at low concentrations, as the temperature is lowered association becomes more favorable, however cross-association between the 12-HSA and solvent in hydrogen-bonding systems may compatibilize the solution and make crystallization less favorable. At the same time this may result in larger  $\chi$  parameters and larger associated solvent molecules, that could drive liquid–liquid phase separation resulting in cloudy gels. A second example is the comparison of the gelation behavior of 12-HSA in alcohols and the corresponding diols.<sup>16</sup> While 12-HSA is not able to gel the alcohols, the diols could be gelled. This may be due to the diols forming larger associated aggregates, which stabilize the liquidus line providing more favorable conditions for gelation. While this discussion is speculative, it motivates revisiting these systems and investigating their phase behavior to understand the role of solvent and solute association on the phase diagrams and gelation behavior. In addition, while the solution solute structure strongly influences the phase behavior, it is only facet of the gelation behavior, which also includes the solid-state structure of the

organogelator, and its ability to crystallize quickly in one-dimension to form fibrillar networks. Therefore, it would also be useful to study the interplay between the liquid-state and solid-state structure of the organogelator. For example, are the preference between different polymorphs related to the degree of association in the solution?

## Conclusions

It has been demonstrated that the phase behavior of 12-hydroxystearic acid (12-HSA)/alkane solutions may be interpreted by using an associated solution model. The liquidus lines in shorter alkanes (octane to hexadecane with even number of carbons) were fit to an attenuated solute association model with only two fitting parameters that give results consistent with the association of 12-HSA dimers into cyclic aggregates through alcohol hydrogen bonding at high 12-HSA concentration. At low 12-HSA concentration, 12-HSA dimers are primarily formed, but the influence of the high concentration associated aggregates is present at low concentration through the free energy cost of dissociating the alcohol groups. Therefore, the monomer concentration of 12-HSA dimers in the neat 12-HSA plays a similar role to the melting temperature in setting the liquidus line. As many different small molecule organogelators use hydrogen bonding groups to drive gelation, solute association offers additional design parameters, such as the neat monomer concentration or the association mechanism, to tune the phase behavior similar to other solute parameters, such as the melting point, heat of fusion and solubility parameter. As the design of organogelator has always been limited by some measure of empirical, trial-and-error variations in structure, it would therefore be extremely useful to revisit some widely used organogelator systems to see the extent that solute association models can be used to interpret differences in the structure–property relationships with systematic variation of the molecular structure.

## Author contributions

Conceptualization T-YL, KAC; data curation T-YL; formal analysis T-YL, KAC, FK; funding acquisition KAC; investigation T-YL; methodology T-YZL, KAC; project administration KAC; resources KAC, software T-YL, KAC, FK; supervision KAC; validation T-YL, KAC; visualization T-YL, KAC; writing – original draft T-YL, KAC; writing – review & editing T-YL, KAC, FK.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

Acknowledgement is made to the donors of the American Chemical Society Petroleum Research Fund for partial support of this research.

## References

- 1 R. G. Weiss, *J. Am. Chem. Soc.*, 2014, **136**, 7519–7530.
- 2 P. Terech and R. G. Weiss, *Chem. Rev.*, 1997, **97**, 3133–3159.
- 3 E. R. Draper and D. J. Adams, *Chem.*, 2017, **3**, 390–410.
- 4 P. R. A. Chivers and D. K. Smith, *Nat. Rev. Mater.*, 2019, **4**, 463–478.
- 5 R. G. Weiss, *Molecular Gels: Structure and Dynamics*, The Royal Society of Chemistry, 2018.
- 6 R. G. Weiss and P. Terech, *Molecular Gels: Materials with Self-Assembled Fibrillar Networks*, Springer, Dordrecht, Netherlands, 2006.
- 7 J.-M. Guenet, *Gels*, 2021, **7**, 65.
- 8 M. D. Segarra-Maset, V. J. Nebot, J. F. Miravet and B. Escuder, *Chem. Soc. Rev.*, 2013, **42**, 7086–7098.
- 9 P. Kirilov, S. Rum, E. Gilbert, L. Roussel, D. Salmon, R. Abdayem, C. Serre, C. Villa, M. Haftek, F. Falson and F. Pirot, *Int. J. Cosmet. Sci.*, 2014, **36**, 336–346.
- 10 K. J. Skilling, F. Citossi, T. D. Bradshaw, M. Ashford, B. Kellam and M. Marlow, *Soft Matter*, 2014, **10**, 237–256.
- 11 D. B. Amabilino and J. Puigmarti-Luis, *Soft Matter*, 2010, **6**, 1605–1612.
- 12 A. Singh, F. I. Auzanneau and M. A. Rogers, *Food Res. Int.*, 2017, **97**, 307–317.
- 13 N. M. Sangeetha and U. Maitra, *Chem. Soc. Rev.*, 2005, **34**, 821–836.
- 14 M. de Loos, B. L. Feringa and J. H. van Esch, *Eur. J. Org. Chem.*, 2005, 3615–3631.
- 15 S. Abraham, Y. Lan, R. S. H. Lam, D. A. S. Grahame, J. J. H. Kim, R. G. Weiss and M. A. Rogers, *Langmuir*, 2012, **28**, 4955–4964.
- 16 Y. Lan and M. A. Rogers, *CrystEngComm*, 2015, **17**, 8031–8038.
- 17 M. Suzuki, M. Yumoto, M. Kimura, H. Shirai and K. Hanabusa, *Chem. – Eur. J.*, 2003, **9**, 348–354.
- 18 J.-M. Guenet, *Organogels: Thermodynamics, structure, solvent role, and properties*, Springer, 2016.
- 19 G. Zhu and J. S. Dordick, *Chem. Mater.*, 2006, **18**, 5988–5995.
- 20 W. Edwards, C. A. Lagadec and D. K. Smith, *Soft Matter*, 2011, **7**, 110–117.
- 21 M. Bielejewski, A. Łapiński, R. Luboradzki and J. Tritt-Goc, *Langmuir*, 2009, **25**, 8274–8279.
- 22 J. Makarević, M. Jokić, B. Perić, V. Tomišić, B. Kojić-Prodić and M. Žinić, *Chem. – Eur. J.*, 2001, **7**, 3328–3341.
- 23 C. Liu, M. Corradini and M. A. Rogers, *Colloid Polym. Sci.*, 2015, **293**, 975–983.
- 24 J. Gao, S. Wu and M. A. Rogers, *J. Mater. Chem.*, 2012, **22**, 12651–12658.
- 25 J. Bonnet, G. Suissa, M. Raynal and L. Bouteiller, *Soft Matter*, 2014, **10**, 3154–3160.
- 26 M. A. Rogers and A. G. Marangoni, *Langmuir*, 2016, **32**, 12833–12841.
- 27 M. Raynal and L. Bouteiller, *Chem. Commun.*, 2011, **47**, 8271–8273.
- 28 Y. Lan, M. G. Corradini, X. Liu, T. E. May, F. Borondics, R. G. Weiss and M. A. Rogers, *Langmuir*, 2014, **30**, 14128–14142.
- 29 K. A. Cavicchi, M. Pantoja and T.-Y. Lai, *ACS Symp. Ser.*, 2018, **1296**, 246–265.
- 30 A. R. Hirst, I. A. Coates, T. R. Boucheteau, J. F. Miravet, B. Escuder, V. Castelletto, I. W. Hamley and D. K. Smith, *J. Am. Chem. Soc.*, 2008, **130**, 9113–9121.
- 31 C. Lagadec and D. K. Smith, *Chem. Commun.*, 2011, **47**, 340–342.
- 32 E. A. Wilder, C. K. Hall, S. A. Khan and R. J. Spontak, *Langmuir*, 2003, **19**, 6004–6013.
- 33 X. Huang, P. Terech, S. R. Raghavan and R. G. Weiss, *J. Am. Chem. Soc.*, 2005, **127**, 4336–4344.
- 34 M. Lescanne, A. Colin, O. Mondain-Monval, F. Fages and J. L. Pozzo, *Langmuir*, 2003, **19**, 2013–2020.
- 35 J. F. Toro-Vazquez, J. Morales-Rueda, A. Torres-Martínez, M. A. Charó-Alonso, V. A. Mallia and R. G. Weiss, *Langmuir*, 2013, **29**, 7642–7654.
- 36 R. Wang, X.-Y. Liu, J. Xiong and J. Li, *J. Phys. Chem. B*, 2006, **110**, 7275–7280.
- 37 R. J. Twieg, T. P. Russell, R. Siemens and J. F. Rabolt, *Macromolecules*, 1985, **18**, 1361–1362.
- 38 E. Christ, C. Blanc, A. Al Ouahabi, D. Maurin, R. Le Parc, J.-L. Bantignies, J.-M. Guenet, D. Collin and P. J. Mésini, *Langmuir*, 2016, **32**, 4975–4982.
- 39 E. Christ, D. Collin, J.-P. Lamps and P. J. Mésini, *Phys. Chem. Chem. Phys.*, 2018, **20**, 9644–9650.
- 40 L. Feng and K. A. Cavicchi, *Soft Matter*, 2012, **8**, 6483–6492.
- 41 V. A. Mallia, M. George, D. L. Blair and R. G. Weiss, *Langmuir*, 2009, **25**, 8615–8625.
- 42 N. M. Sangeetha and U. Maitra, *Chem. Soc. Rev.*, 2005, **34**, 821–836.
- 43 Y. Lan, M. G. Corradini, R. G. Weiss, S. R. Raghavan and M. A. Rogers, *Chem. Soc. Rev.*, 2015, **44**, 6035–6058.
- 44 V. A. Mallia and R. G. Weiss, *Soft Matter*, 2015, **11**, 5010–5022.
- 45 D. A. S. Grahame, C. Olauson, R. S. H. Lam, T. Pedersen, F. Borondics, S. Abraham, R. G. Weiss and M. A. Rogers, *Soft Matter*, 2011, **7**, 7359–7365.
- 46 T. Taro and K. Hideko, *Bull. Chem. Soc. Jpn.*, 1969, **42**, 3422–3424.
- 47 T. Tachibana and K. Hori, *J. Colloid Interface Sci.*, 1977, **61**, 398–400.
- 48 T. Sakurai, Y. Masuda, H. Sato, A. Yamagishi, H. Kawaji, T. Atake and K. Hori, *Bull. Chem. Soc. Jpn.*, 2010, **83**, 145–150.
- 49 V. A. Mallia and R. G. Weiss, in *Gels and Other Soft Amorphous Solids*, American Chemical Society, 2018, vol. 1296, ch. 12, pp. 227–243.
- 50 M. A. Rogers, S. Abraham, F. Bodondics and R. G. Weiss, *Cryst. Growth Des.*, 2012, **12**, 5497–5504.
- 51 H. Mutlu and M. A. R. Meier, *Eur. J. Lipid Sci. Technol.*, 2010, **112**, 10–30.
- 52 A. K. Zetzel, A. G. Marangoni and S. Barbut, *Food Funct.*, 2012, **3**, 327–337.
- 53 P. Kirilov, S. Rum, E. Gilbert, L. Roussel, D. Salmon, R. Abdayem, C. Serre, C. Villa, M. Haftek and F. Falson, *Int. J. Cosmet. Sci.*, 2014, **36**, 336–346.
- 54 C. L. Esposito, V. Tardif, M. Sarrazin, P. Kirilov and V. G. Roullin, *Mater. Sci. Eng., C*, 2020, **114**, 110999.

- 55 B. Martin, G. Garrait, E. Beyssac, D. Goudouneche, E. Perez and S. Franceschi, *Pharm. Res.*, 2020, **37**, 92.
- 56 P. Lee and M. A. Rogers, *Langmuir*, 2013, **29**, 5617–5621.
- 57 R. Gordon, S. T. Stober and C. F. Abrams, *J. Phys. Chem. B*, 2017, **121**, 9223–9233.
- 58 R. Gordon, S. T. Stober and C. F. Abrams, *J. Phys. Chem. B*, 2019, **123**, 534–541.
- 59 K. Marsh and F. Kohler, *J. Mol. Liq.*, 1985, **30**, 13–55.
- 60 A. Apelblat, *J. Mol. Liq.*, 2006, **128**, 1–31.
- 61 J. Gao, S. Wu and M. A. Rogers, *J. Mater. Chem.*, 2012, **22**, 12651–12658.
- 62 V. A. Mallia and R. G. Weiss, *J. Phys. Org. Chem.*, 2014, **27**, 310–315.
- 63 A. Bui and N. Virgilio, *Ind. Eng. Chem. Res.*, 2013, **52**, 14185–14191.
- 64 P. Fei, S. J. Wood, Y. Chen and K. A. Cavicchi, *Langmuir*, 2015, **31**, 492–498.
- 65 C. M. Hansen, *Hansen's solubility Parameters: A Users Handbook*, CRC Press, Boca Raton, FL, 2007.
- 66 S. R. Raghavan and B. H. Cipriano, in *Molecular Gels: Materials with Self-Assembled Fibrillar Networks*, ed. R. G. Weiss and P. Terech, Springer Netherlands, Dordrecht, 2006, pp. 241–252.
- 67 H. Takeno, M. Yanagita, Y. Motegi and S. Kondo, *Colloid Polym. Sci.*, 2015, **293**, 199–207.
- 68 H. Takeno and M. Kozuka, *Adv. Mater. Sci. Eng.*, 2017, **2017**, 4762379.
- 69 J.-M. Guenet, *Thermochim. Acta*, 1996, **284**, 67–83.
- 70 Y. K. Lin, G. Chen, J. Yang and X. L. Wang, *Desalination*, 2009, **236**, 8–15.
- 71 L. Rycerz, *J. Therm. Anal. Calorim.*, 2013, **113**, 231–238.
- 72 D. Giron, *J. Therm. Anal. Calorim.*, 2001, **64**, 37–60.
- 73 K. Sato, K. Suzuki, M. Okada and N. Garti, *J. Cryst. Growth*, 1985, **72**, 699–704.
- 74 E. Moreno, R. Cordobilla, T. Calvet, M. A. Cuevas-Diarte, G. Gbabode, P. Negrier, D. Mondieig and H. A. J. Oonk, *New J. Chem.*, 2007, **31**, 947–957.
- 75 M. C. Costa, M. P. Rolemberg, A. J. A. Meirelles, J. A. P. Coutinho and M. A. Krähenbühl, *Thermochim. Acta*, 2009, **496**, 30–37.
- 76 J. M. Guenet, B. Demé, O. Gavet, E. Moulin and N. Giuseppone, *Soft Matter*, 2022, **18**, 2851–2857.
- 77 P. Painter, B. Veytsman and J. Youtcheff, *Energy Fuels*, 2014, **28**, 2472–2480.
- 78 R. Koningsveld, W. H. Stockmayer and E. Nies, *Polymer Phase Diagrams*, Oxford University Press, New York, 2001.
- 79 P. J. Flory, *J. Chem. Phys.*, 1944, **12**, 425–438.
- 80 C. B. Kretschmer and R. Wiebe, *J. Chem. Phys.*, 1954, **22**, 1697–1701.
- 81 P. Painter, B. Veytsman and J. Youtcheff, *Energy Fuels*, 2015, **29**, 2120–2133.
- 82 P. Painter, B. Veytsman and J. Youtcheff, *Energy Fuels*, 2015, **29**, 7048–7057.
- 83 P. Painter, B. Veytsman and J. Youtcheff, *Energy Fuels*, 2015, **29**, 2951–2961.
- 84 J. William and E. Acree, *Thermodynamic Properties of Non-electrolyte Solutions*, Academic Press, Orlando, FL, 1984.
- 85 I. Prigogine, A. Bellemans and M. Mathot, *The Molecular Theory of Solutions*, Interscience Publishers, Inc., New York NY, 1957.
- 86 A. F. M. Barton, *Chem. Rev.*, 1975, **75**, 731–753.
- 87 Y. Fujii, H. Yamada and M. Mizuta, *J. Phys. Chem.*, 1988, **92**, 6768–6772.
- 88 N. Asprion, H. Hasse and G. Maurer, *Fluid Phase Equilib.*, 2001, **186**, 1–25.
- 89 K. Nakasone, K. Shiokawa, Y. Urabe and N. Nemoto, *J. Phys. Chem. B*, 2000, **104**, 7483–7489.
- 90 R. A. Matkar and T. Kyu, *J. Phys. Chem. B*, 2006, **110**, 12728–12732.
- 91 T. Sakurai, Y. Masuda, H. Sato, A. Yamagishi, H. Kawaji, T. Atake and K. Hori, *Bull. Chem. Soc. Jpn.*, 2010, **83**, 145–150.
- 92 D. Zhao and J. S. Moore, *Org. Biomol. Chem.*, 2003, **1**, 3471–3491.