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Exploiting complexity to implement function in chemical systems

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Chemistry deals with complex molecular systems that can be further connected by supramolecular interactions and reaction networks. However, chemists have taken little advantage of the intrinsic complexity of chemical systems, probably due to the lack of appropriate tools to analyse and understand complexity. In the last few decades, the concept of complexity has grown appealing: it allows the design of networks and dynamic systems expressing emerging properties and functions, which would be difficult to achieve from the mere addition of the components of the ensemble. Here we describe a personal overview of the recent state-of-the-art in the field, mainly focused on complex systems providing molecular recognition and catalysis. Far from being a thorough revision of the recent literature, we intend to illustrate the topic to attract the chemical community for considering complexity as an additional parameter in their research.

Introduction

Complexity is a very appealing concept present in Nature, since it is ubiquitous to living matter,¹ climate phenomena,² astrophysics,³ and biological ecosystems,⁴ but also social networks,⁵ economical markets⁶ and, in general, every network where individual entities interact with each other. The realisation of complexity has crystallized in a growing interest to understand the laws and properties governing the evolution and adaptation of complex systems to different conditions, such as the action of stimuli or external perturbations. The deep understanding of complex systems will allow taking advantage of emerging properties that are absent in the components of the systems but appear upon the construction of the corresponding network.⁷ By definition, molecular systems are in general complex dynamic systems.⁸ Even a mere dissolution of a molecule in a solvent can be considered a complex network where the physicochemical properties of both components (solute and solvent) change from the isolated pure states to the corresponding solution. In this simple example, the solute–solute, solute–solvent and even solvent–solvent interactions dramatically change in the mixture and can be also different depending on the conditions at which this mixture is formed (molar fraction, temperature, pressure, presence of heterogeneous domains and phases, *etc.*). Moreover, if we include in the system a chemical reaction⁹ or a recognition process,¹⁰ an additional level of complexity starts where temporal and spatial constitutional changes can occur. Thus, in

general, chemistry is a scientific discipline dealing with complex systems. In contrast, chemists have avoided to work with high levels of complexity in order to be able to understand the processes under study. However, chemists are recently designing and studying more complex systems thanks to the acquired knowledge and to the emergence of new analytical and computational technologies that now make possible to measure, analyse and examine several parameters and variables.

In the last decades, the proposal of systems chemistry¹¹ as a new paradigm to face dynamic chemical systems has emerged as a different viewpoint in chemical research. In parallel to the well-established field of systems biology,¹² the chemical partner aims to set up complex chemical systems to exploit their ability to produce emerging properties that arise from the network and are absent in the individual components of the ensemble. To that, dynamic combinatorial chemistry, self-assembling, self-replication, adaptation, molecular evolution or far-from-equilibrium concepts have entered in the chemists' jargon to explain processes and properties of complex systems.¹³ The beauty of complex chemical systems is especially remarkable when developing a given function that comes from the coordinated action of the different components of the network. Thus, we propose the implementation of complexity as a design vector in our chemical research, like an additional parameter to manipulate the outcome of a chemical reaction or device. Besides, complexity can be expressed at different levels (Fig. 1). The molecular complexity can be attributed to the actual properties of the molecules themselves, such as different functionalities, conformational flexibility or tautomerism. An additional level sources when molecules interact by non-covalent forces leading to supramolecular structures. In parallel, molecules can interact

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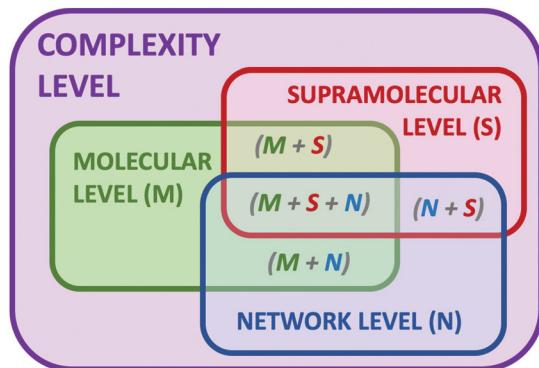


Fig. 1 Schematic representation of the different levels within chemical systems, connecting molecular (M), supramolecular (S) and network (N) levels. Complexity can be expressed at all levels, as well as through their corresponding combinations.

between them in a network of chemical reactions and processes, producing an additional level of complexity. Also consequently, we can imagine situations where any combination of two or the three levels can cooperate.

In this feature article, we will summarize our point of view in this area, specially by the implementation of complex chemical systems for a given function. We will illustrate the concept with selected examples that are not intended to deeply review the state of the art in the field, but to encourage the readership to think over the concept of complexity as an important additional variable to consider in future research planning.

Dynamic covalent chemistry in complex systems

The formation of chemical networks provides an excellent methodology for the generation and study of complex chemical systems.¹⁴ To this aim, Dynamic Covalent Chemistry (DCC) affords a valuable tool for the preparation of such networks.¹⁵ In the Dynamic Covalent Libraries (DCL) the members of the system are connected through the formation and exchange of reversible covalent bonds giving rise to combinatorial collections of compounds.¹⁶ Several dynamic bonds are suitable for that purpose being hydrazone and imine formation, and disulphide exchange maybe the most commonly employed.^{15a,e} DCC was first envisioned for the preparation and study of ligand–receptor systems.¹⁷ As DCLs are intrinsically dynamic and subjected to thermodynamic control the receptor with a better affinity towards the guest should be stabilised and therefore its concentration should increase. This ideal concept, however, is not always perfect as the system will evolve towards the distribution that minimizes the energy of the whole system itself, which does not necessarily mean that the best binder is amplified the most.¹⁸ On the other hand, amplification of suitable receptors allows the emergence of compounds that were almost non-detectable in the absence of the guest.¹⁹ The size of the library is also an important parameter: Otto's research group addressed this issue by simulating a set of

libraries leading to the conclusion that larger libraries offer the possibility to obtain stronger ligands and therefore complexity seems to deliver positive outcomes.²⁰ The concentration of the template also plays an important role: if too much template is present it can hamper the emergence of the best receptors by the formation of a large number of lower affinity, worse, ligands.²¹

In practical terms, DCC has been useful in the generation of efficient ligands, particularly in aqueous systems, which are especially relevant when pursuing biological applications because the binding and recognition of specific molecules by non-covalent, supramolecular interactions is competing with solvation interactions with the medium.²² Thus, several ligands with different structural features, from cyclic to linear scaffolds, have been described to target a large variety of compounds, from simple small molecules to bigger biomacromolecules.²³ Maybe the most attractive characteristic of the DCC methodology is that a detailed structural knowledge of the host–guest interaction is not needed, even being possible to work with mixtures. This feature is especially useful when trying to recognise macromolecules with large conformational freedom or with heterogeneous composition. We took advantage of the intrinsic nature of DCC for the recognition of glycosaminoglycans (GAGs) such as heparin (1 in Fig. 2A) using a library of polyamines and aromatic aldehydes.²⁴ We targeted two main binding motifs of this polysaccharide: the anionic sulphate and carboxylate functions on one side and the axial positions on the carbohydrates that are able to establish attractive CH–π interactions with aromatic rings. To do so, we used reductive amination reaction between spermine (2) as cationic platform at physiological pH and a mixture of five different aldehydes leading to a combinatorial output (Fig. 2A). When repeating the reaction in the presence of heparin we succeeded in identifying some amplified compounds. After preparative-scale synthesis of the potential best ligand 3AL we proceeded to study its interaction with heparin by means of NMR and fluorescence spectroscopies, as well as molecular modelling identifying a series of electrostatic, hydrogen bonding and C–H contacts as expected. Using Isothermal Titration Calorimetry (ITC, Fig. 2B) we were able to estimate a strong binding constant in aqueous medium between 3AL and heparin and, more importantly, this ligand was able to reverse the inhibitory effect of heparin in an enzymatic test related to human blood coagulation (Fig. 2C).

Encouraged by this success, we reasoned that an attractive and useful application to extend our research would be addressing the GAGs present in the cells. We anticipated that a suitable DCL against whole living cells should reflect the extracellular matrix composition. As we expected, using live A549 cancer cells as template and a dynamic library similar to that employed for the interaction with heparin, we could observe the amplification of a particular polyamine.²⁵ Importantly, another cell line gave a different amplification pattern showing the dependence in the library behaviour towards the composition of the specific extracellular matrix. In this case, using a combination of on-cell NMR and SPR techniques we could prove an interaction between the amplified component and a GAG within the extracellular matrix. Our success highlights



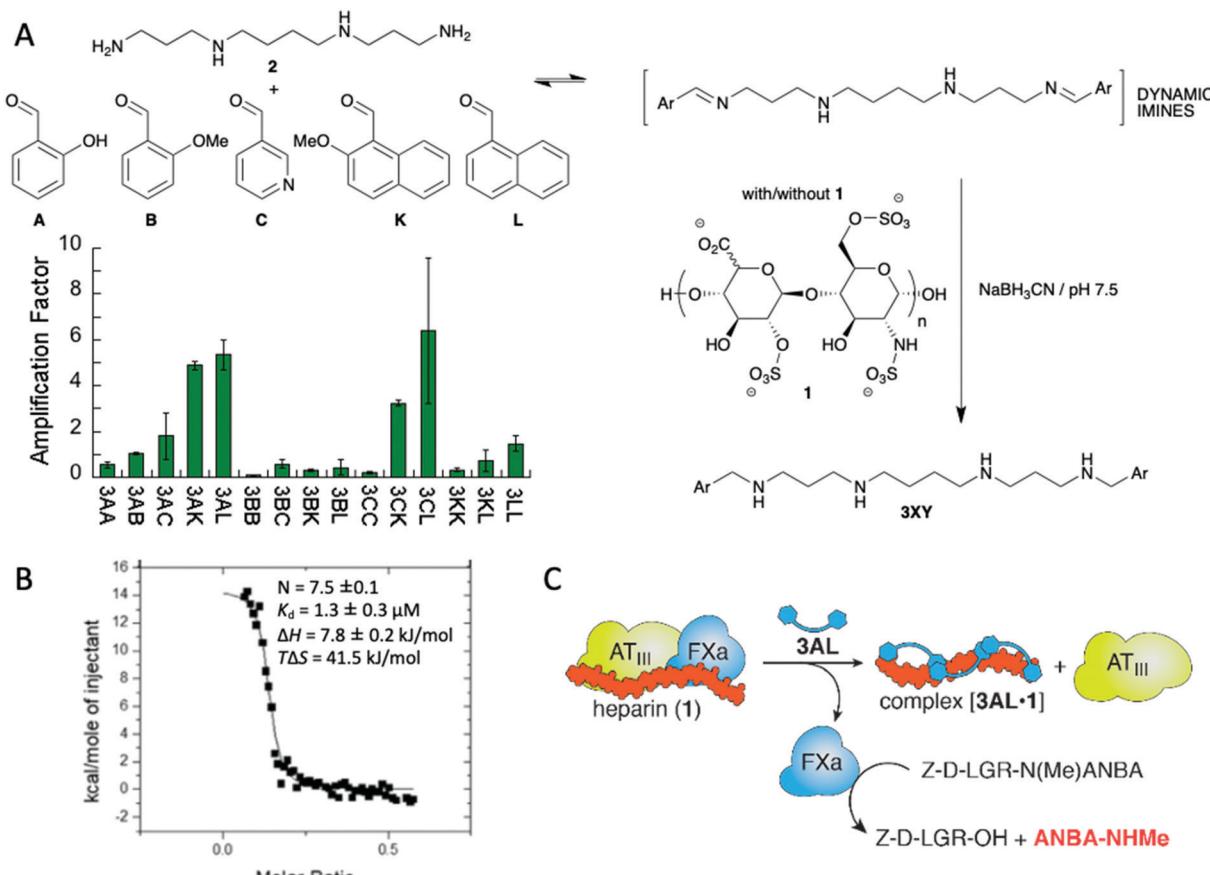


Fig. 2 (A) Discovery of a ligand for heparin (1) using a library of dynamic imines. (B) ITC trace for the binding of **3AL** to **1**. (C) Reversal of the heparin inhibitory effect in an enzymatic assay related to blood coagulation.

the power of dynamic combinatorial methodologies to address particularly challenging supramolecular processes specially when targeting not well-defined structures.

The adaptability of the chemical networks originated by DCC allows the sorting of their constituents in response to external stimuli or the presence of chemical effectors. Light,²⁶ pH changes,²⁷ temperature,^{27a} pressure²⁸ or salt concentration²⁹ proved to be efficient in shifting the species' distribution. Studying these changes, it is possible to obtain individual fingerprints for each stimuli. Thus, the groups of Anslyn,³⁰ Matile,³¹ Severin,³² Zonta³³ and Lehn³⁴ have used pattern-based techniques for the sensing of different analytes using the response of the system without the need to isolate, synthesise or study individual compounds. The analyte is monitored by the changes in the UV, fluorescence or even mass spectra and the corresponding chemometric analysis of the changes in the whole library leads to the identification of the analyte added. We started our research in this area analysing dynamic libraries of macrocyclic pseudopeptides and their response to several stimuli.^{29,35} We found out that these libraries responded to variations in the ionic strength leading to changes in the library composition that resemble the evolutionary patterns found in halophilic proteins.^{29a} Moreover, the knowledge acquired allowed us to design *de novo* chemical systems with intended behaviour

containing competing or cooperating species.^{29b} In addition, chemometrics such as principal component analysis (PCA) or Multivariate Curve Resolution-Alternating Least Squares (MCR-ALS) allowed us to quantify the effect of multiple stimuli in a dynamic library (Fig. 3). These tools proved to be especially useful when two inter-connected stimuli were combined in the same dynamic system.³⁶

As scientists have been able to deal with more complex systems thanks to the advanced analysis techniques, the complexity of the chemical networks has gained importance. Thus, despite that dynamic libraries usually contain just a dynamic bond, reaction networks containing more than one dynamic reaction have been described,³⁷ with up to four orthogonal reactions operating at the same time and studied thanks to computational deconvolutions.³⁸ Very interestingly, Furlan and co-workers showed that combining sequential reversible reactions gives the chemical pathway a direction. Thus, the thermodynamic systems that result from combining these reactions keep memory of the previous equilibration processes leading to different layers of information.³⁹

The group of Otto undertook a further step in the generation of complex systems by combining two different covalent chemistries operating simultaneously to generate diverse DCLs (Fig. 4).⁴⁰ Thiol-disulphide and thio-Michael reactions were

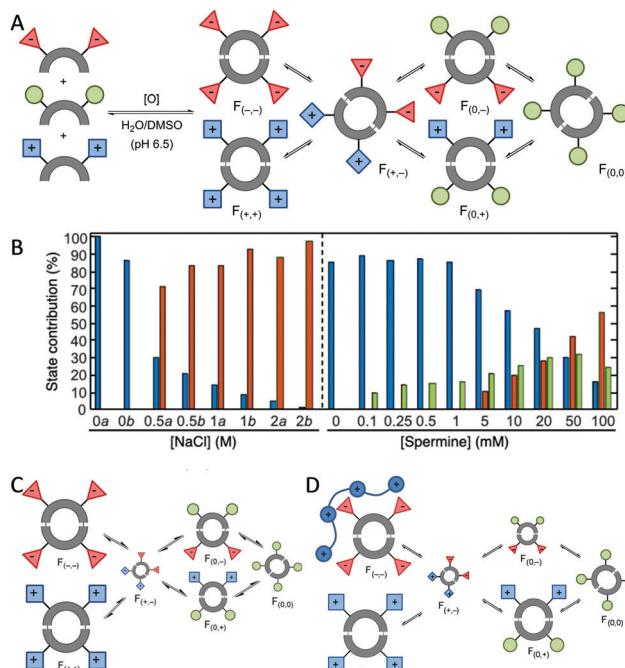


Fig. 3 Use of chemometrics to unravel multi-stimuli response in DCL of macrocyclic pseudopeptides: (A) schematic representation of the DCL generated by disulphide formation in aqueous media giving yield to six different families classified by their charge at neutral pH. (B) Bar plots of the scores of the three states resolved by MCR-ALS analysis of the HPLC in the presence of salt or spermine data sets. (C) Schematic representation of the changes in concentration of the families of macrocycles due to the salt effect (larger/smaller size symbols represent an increase/decrease of the concentration) (D) cartoon representation of the effect of the spermine polycation binding on the DCL.

combined using the same building blocks (BBs) and, consequently, as the products of one reaction grow in concentration the products of the other have to decrease making the two processes antiparallel. External control over their ratios can be exerted by the use of reducing or oxidizing agents.

This use of different chemistries was elegantly used by Withersides to build networks of reactions with autocatalytic, bistable and oscillatory behaviour. First, they developed a network based on amplification using thiols and thioesters. Addition of acrylamide delays the autocatalytic growth of the amplified species by the covalent capture of the thiols. Using a continuously stirred tank reaction that allowed a flux of species into and out of the system overtime they could design conditions in which the addition of acrylamide (in excess in the media) produced oscillations in the concentration of the amplified thiols.⁴¹ In a more recent study, the same authors showed that the oscillatory behaviour of thioester (AlaSET) can be 'trained' to other non-oscillatory related compounds (GlySET and AbuSET) even increasing the period of oscillating behaviour and thus increasing its robustness. Finally, they showed that individual components that do not show oscillatory behaviour when isolated are able to oscillate when mixed together.⁴² Remarkably, the emergence of complex behaviour from simple molecules, such as those described in these papers, has been related to the pre-biotic origin of life.

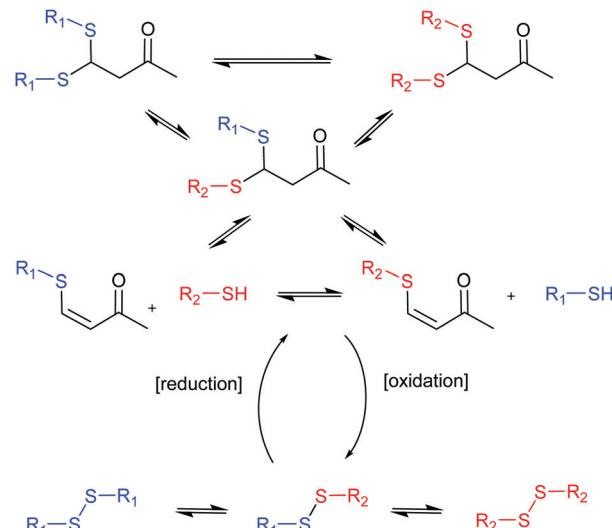


Fig. 4 Antiparallel chemistries described by Otto.⁴⁰ The final state of the system between thio-Michael chemistry (top) or disulphide chemistry (bottom) can be switched by addition of reducing or oxidising agents.

The time-dependence of systems has also been studied by Lehn using DCL where its constituents can be switched from kinetic products (imines) to thermodynamic products (oximes or acylhydrazones) based on the reactivities and relative stabilities of the members.⁴³

Another potential source of complexity arises from the number of functional groups involved in a given network and the combination of different geometries. In these scenarios one can envision the formation of large multimacrocycles or cage-like compounds among others. Building in the research performed by Sanders and Otto,⁴⁴ we became particularly interested in this area as a manner to expand the geometrical possibilities of our dynamic libraries beyond the formation of cyclic compounds. By mixing BBs having a different number of thiol groups we were able to generate a variety of molecules with different topologies, some of them with degenerate mass. Importantly, we also observed self-selection mechanisms leading to a simplification of the libraries by the complementarity of charge and structure between BBs.⁴⁵ Studying the molecular interactions that gave rise to these processes, we were able first to understand the mechanism leading to self-assembly and how the system evolved through time from complex kinetic mixtures to the thermodynamic final state. Strikingly, in some cases adding more components to the mixture resulted in the simplification in the number of the species formed (Fig. 5A). Mixing BB 4 having three thiol groups and dithiol 5 a complex library was formed. When cysteine 6 was added to the mixture, a single compound 7 containing all the BBs was formed which implies the covalent capture of cysteine in zwitterionic form.^{45a,b} Including a chromophore (8) in the library that was released after the addition of cysteine (as the oxidized dimer 8-8) the system was able to act as a sensor for this amino acid.⁴⁶

The particularity of our strategy is that it is the library itself and not any of the individual compounds that form the sensor



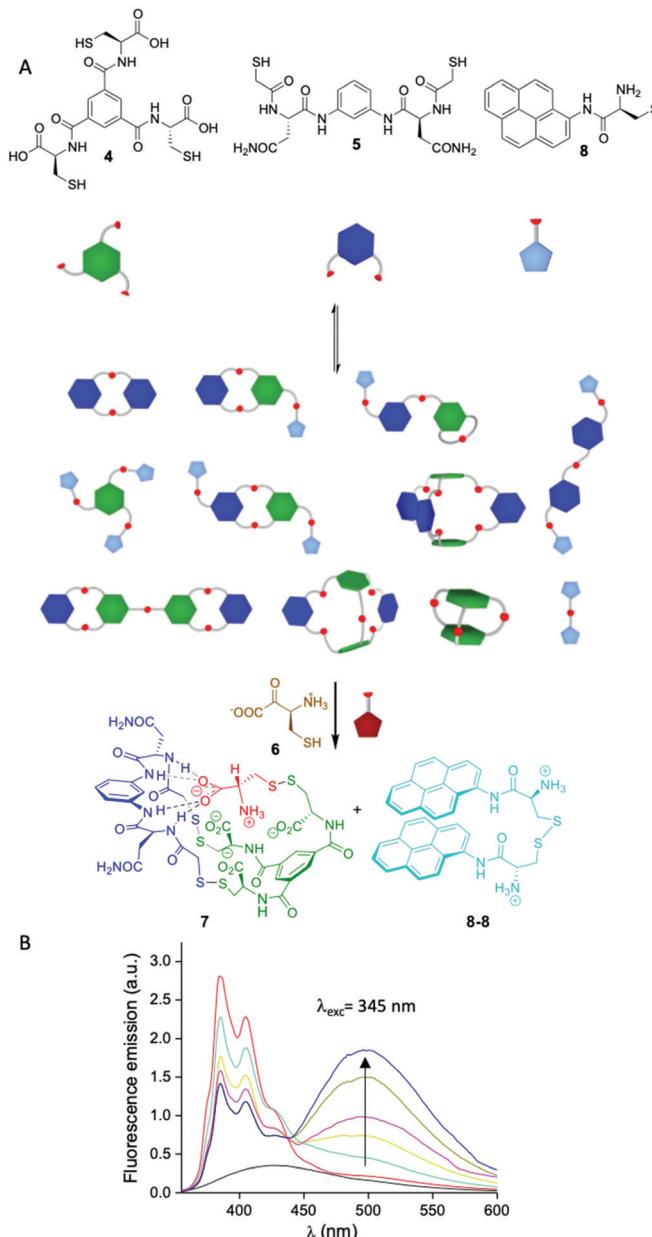


Fig. 5 (A) Schematic representation of the sensing network for cysteine and cystine described by our group.⁴⁶ (B) Fluorescence response of the dynamic network in response to increasing amounts of cysteine in human urine matrix (background urine fluorescence emission in black).

and the response is read by changes in fluorescence without the need to analyse the library composition once the system was optimised. This sensor was shown to be useful to detect cysteine in human urine being a potential tool for the detection and diagnosis of cystinuria (Fig. 5B).

Dynamic networks not only can react to external stimuli but also self-templation is an interesting phenomenon itself. Thus, unexpected complex structures such as catenanes,^{23c,47} knots⁴⁸ or self-folding molecules have been described.⁴⁹ In most cases their formation is driven by the hydrophobic effect as these compounds present a more compact structure with less areas

exposed to the solvent. A particular case of study arises when one of the members of the library can aggregate with good specificity favouring its formation and making effectively a self-replicator.⁵⁰ Minimal replicators are thought to be an essential requirement in the appearance of systems where life could emerge.⁵¹ Among other chemistries, dynamic covalent networks have been developed for the generation of replicating systems. Giuseppone using a dynamic equivalent of a replicator reported by Rebek⁵² described one of the first examples of the generation of replicators with DCC.⁵³ Later, Philp designed a network of reactions leading to the amplification of a synthetic replicator by coupling a pre-existent nitronate-based replicator to an imine-nitronate exchange.⁵⁴ In order to achieve new smart materials, Giuseppone and co-workers described the selection of a nano-structure resulting from phase separation (Fig. 6). They combined a kind of copolymers bearing a hydrophobic moiety (from **9**) and hydrophilic ones (**10–13**) connected by a reversible imine linkage (dynablocks). Interestingly, the imine bond resulted stable in aqueous media thanks to the spontaneous formation of supramolecular structures. Moreover, the self-assembled structures made from those dynablocks were able to replicate by catalysing the formation from their BBS and selecting the most efficient replicator.⁵⁵

The use of peptide supramolecular structures such as coiled-coil motifs had been employed by Ghadiri⁵⁶ for the generation of autocatalytic networks. More recently, Ashkenasy used the templating effect and folding properties of these structures to design a bistable network through the reversible thioester formation of a thiodesipeptide. Depending on the initial concentration, the system leads to two different steady states which are switchable applying the appropriate stimuli.⁵⁷ The same author reported the use of complex β -sheet peptidic networks from where the formation of a dominant functional polymer backbone can emerge.⁵⁸

In the last years, the group of Otto has put a lot of effort in the preparation of several replicator systems based on libraries

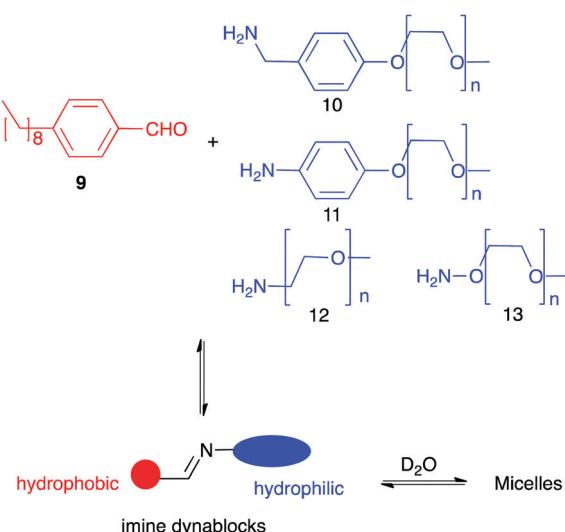


Fig. 6 Formation of supramolecular aggregates using imine dynablocks reported by Giuseppone.⁵⁵

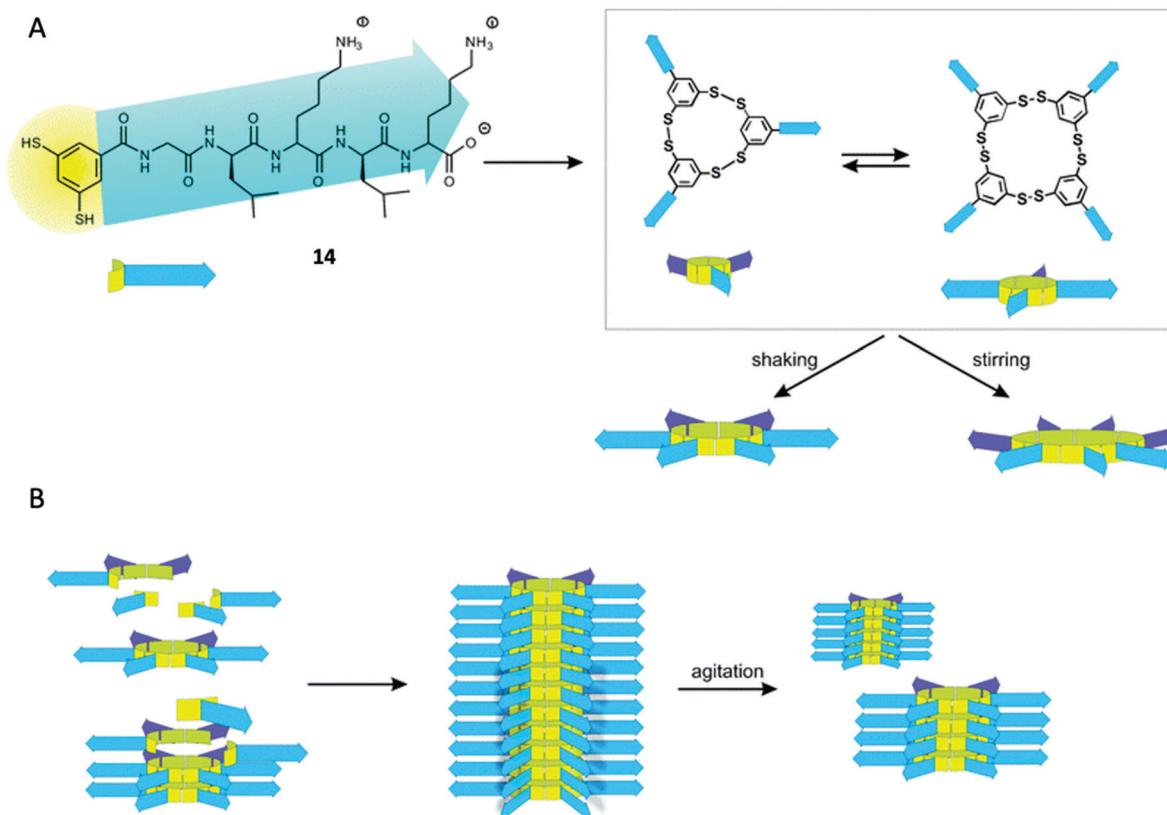


Fig. 7 General replication process for the pseudopeptidic replicators described by Otto. (A) Oxidation of dithiol **14** gives rise to different macrocyclic compounds (mainly trimer and tetramer). After agitation the emergence of larger macrocycles takes place (hexamer and heptamer). (B) The hexamer can form stacks of compound held together by β -sheet formation of the peptidic moieties promoting its own formation. Mechanical rupture of the stacks due to agitation increases the number of ends leading to exponential growth. Reproduced from ref. 14b with permission from the Royal Society of Chemistry.

of dithiol pseudopeptidic BBs (**14**, Fig. 7). As the components are oxidised, several macrocycles arise with different size and the particularity is that some of those members are able to stack on top of themselves therefore promoting their own formation. As these stacks are prone to mechanical rupture the number of ends from where the replicator can grow increases enabling exponential growth.⁵⁹

However, a certain macrocycle size is needed to afford sufficiently strong interactions to drive the process.⁶⁰ Thus, stacks of certain compound form fibres that can be obtained in controllable length and stabilized by covalent capture giving rise to self-synthesising materials. The dissimilar behaviour of **s** when they are alone or mixed with other species allowed the group to design a library that shows no replication until an effector is present.⁶¹ In this particular case, a BB capable of self-replication and a known to form a receptor in the presence of a template were combined together. In the absence of template, no self-catalysis is detected and the different components form mixed structures. Once the effector is introduced, the formation of a suitable receptor captures one of the BBs freeing the other one that is capable of self-templation, and autocatalytic behaviour is observed.

Remarkably, by studying different replicating systems, Otto's research group has been able to establish diverse relationships

inside the chemical networks. On one hand, they described a system in which a diversification of the replicators that are competing for two common BBs into two different sets of replicators was observed.⁶² On the other hand, the appearance of some replicator molecules may require the presence of a pre-existing replicator, that is, in some cases the replicator itself does not form unless it is first templated by a previously formed oligomer.⁶³ Moreover, the emergence of parasitic behaviours of replicators that consume those members to which they owe their own formation has also been witnessed.⁶⁴ These dissimilar behaviours demonstrate the variety of possibilities that arise from studying complex systems. These replicators, however, are invariably either in thermodynamic or kinetic trapped states and no source of energy is required to maintain this state. A step forward in complexity was reported by the group of Fletcher. Building on the work of Giuseppone they described the formation of a metastable surfactant that is capable of forming micelles which increase the surfactant formation and therefore convert the process in self-catalytic.

However, the micelles are subjected to disassembly to reach the thermodynamic products. The addition of hydrogen peroxide as chemical fuel regenerates the BBs for further replicator formation.⁶⁵ Following their work in phase separation autocatalysis, the same group reported the selection of components



in a system of reproducing lipids forming micelles. In these systems, the replicators are metastable and selection can occur from a pool of competitors and out-of-equilibrium populations can be maintained by feeding the system with starting materials. Importantly, no selection is observed when the components are dissolved in a single phase.⁶⁶

Controlled self-assembly and disassembly, coupled with functional molecular architectures is still a major challenge in the laboratory. One of the first examples of the use of dynamic libraries for the controlled formation of materials was described by Lehn using guanosine-5'-hydrazide and aldehydes to generate a library of gelators in the presence of metal cations. If a mixture of aldehydes was used, then the library evolved towards the formation of the most stable gel.⁶⁷ Self-assembly of peptidic materials has been studied extensively by Ulijn group. In an early example, the use of reversible peptidic bond formation catalysed by a protease was reported to generate a DCL of dipeptides. As described above, gelation of one of the components drives the selection of a single product and allows for the discovery of stable self-assembled nanostructures. In these systems, the nature of the self-assembly can be modulated using changes in pH according the acid/base properties of the amino acid residues.⁶⁸ In a more recent example, the group took a step further exploiting the molecular interactions resulting of incorporating amino acids into an organic semiconductor and thus electronic wires could be formed and degraded achieving temporary control over electroconductivity.⁶⁹ As we described above, transient assembly of materials can be fuelled by a chemical reaction provided the self-assembled material can be switched between a non-associating and an associating states by a chemical reaction consuming a fuel, and reversed back to the first state by a different mechanistic pathway. The group of van Esch elegantly made use of ester formation and hydrolysis to control the gel properties of molecular gelators tuned by the kinetics of fuel consumption (Fig. 8).⁷⁰

The kinetic lability of dynamic bonds introduced in polymeric compounds gives them unique properties allowing for the formation of smart materials with self-healing properties, shape memory or degradability. The use of pH sensitive covalent bonds is especially useful when pursuing biological applications. The group of Matile reported the ring opening polymerization initiated by a substrate bearing a thiol group that allows the delivery of dyes, proteins or drugs inside the cells. These polymers were shown to depolymerise rapidly inside the cell due to the presence of endogenous glutathione, minimizing thus their toxicity.⁷¹ A different approach was reported by the group of Ulrich who prepared dynamic covalent polymers by condensation of hydrazone linkers. Thanks to the presence of cationic monomers these materials were capable of effectively complexing DNA. Moreover, the design was demonstrated to be pH sensitive, disassembling rapidly at acidic pHs.⁷² In an extension of this work, using amino acid BBs (18 in Fig. 9) they were able to complex DNA and siRNA and deliver the last one into live cells.⁷³

Overall, the use of dynamic covalent bonds represents a suitable approach to implement diversity in a chemical system

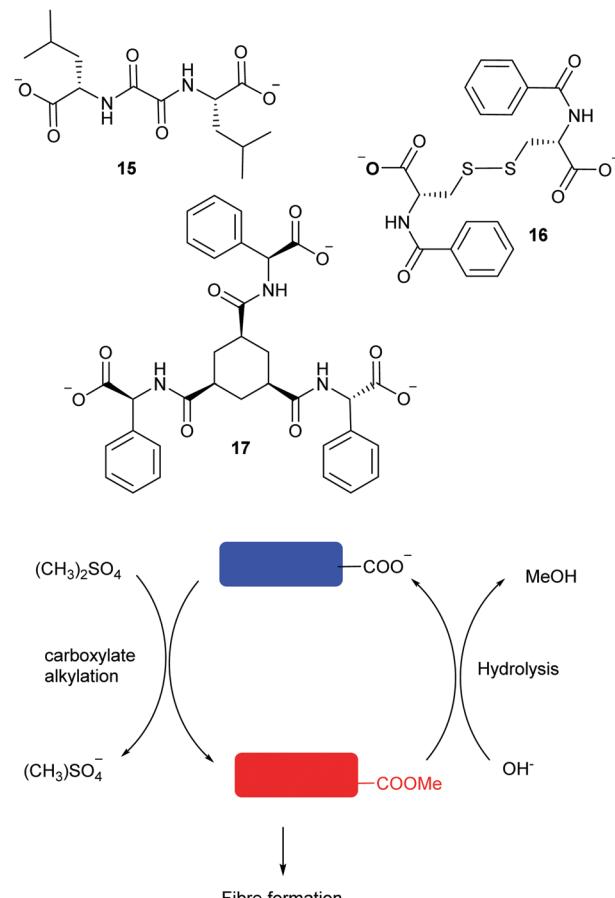


Fig. 8 Fuel-dependant modular pseudopeptidic gelators (15–17) reported by van Esch.⁷⁰

leading to the potential emergence of interesting properties and functions.

Complex systems leading to efficient catalysis

Catalytic network and metabolism are synonymous in living organisms, likewise catalyst and enzyme. Indeed, a single enzymatic reaction in a cell is pointless without the context of a living organism. It is the overall combined network of catabolic and anabolic reactions that defines a living being. The whole metabolic network is a clear example of complex catalytic system that goes beyond simple reaction acceleration: substrate recognition, specificity and selectivity, enzyme inhibition or activation through allosteric regulation, the activation of cascade pathways, and self-regulation are emerging properties of complexity in catalytic networks. Artificial systems might depict some of these emerging properties as well, and there is a continuous flow of new and exciting discoveries in this field.⁷⁴ Some illustrative examples from the last years that allow understanding the behaviour of complex catalytic networks are discussed below.

A very interesting case of substrate activation of a catalytic assembly was reported by Lei and Ju.⁷⁵ Mercury(II) was used as a



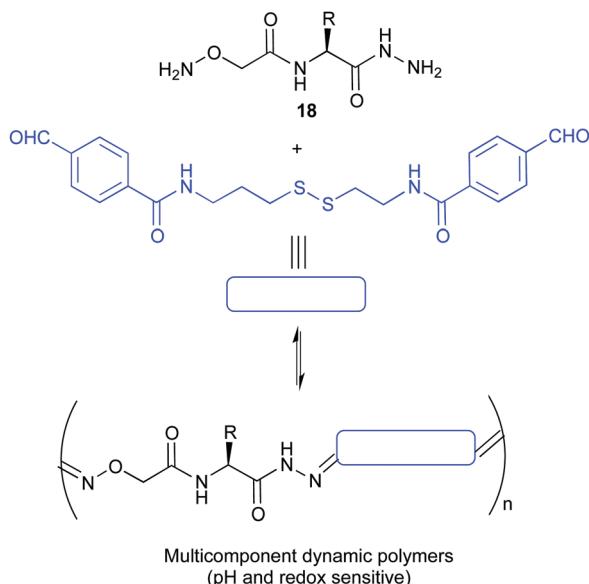


Fig. 9 Multicomponent pseudopeptidic dynamic copolymers described by Ulrich for the delivery of siRNA.⁷³

co-factor to trigger the self-assembly of gold nanoparticles functionalized with hemin groups in the presence of a bridging pyrimidine-type ligand. The Hg(II) assembly showed three-times as much peroxidase activity as the mercury-free system. Ultimately, this activity was used to detect Hg(II) at a sub-attomolar level (Fig. 10).

This is related to allosteric regulation as well, wherein a small molecule or metal regulates catalytic activity of an enzyme. Of course, supramolecular chemistry seems an ideal field for this approach. Remarkable examples have been developed in Mirkin's group.⁷⁶ Two of them are discussed below. In a first experiment a P,S ligand containing a diphenylurea group was developed to coordinate Pt and form a tweezer complex. This is the catalytically

active conformation of the complex, since the hydrogen bond donor properties of the adjacent urea groups enhance the Diels–Alder reaction. However, upon addition of chloride ions, the Pt–S bond breaks, destroying the tweezer conformation, promoting intermolecular aggregation of the urea groups, and ending catalytic activity in the DA reaction (Fig. 11A).⁷⁷ In a second example, two different P,S ligands designed to form a triple decker complex with Pt contained a catalytic squaramide group (hydrogen bond donor) or a self-quenching ester groups. This complex was catalytically active in the Michael addition because the ester groups were positioned far away from the squaramide. Upon addition of chloride ions, the Pt–S bond is broken again and the structure of the triple decker complex changes allowing the quenching of the squaramide by ester groups from other complexes. These oligomers are catalytically inactive (Fig. 11B).⁷⁸

We will turn our attention now to some intrinsically dynamic catalytic systems, either in the substrates or the catalyst, and how chemists have learnt to deal with such complexity. Ramström and co-workers have developed several complex catalytic systems with unusual activities. Dynamic systemic resolutions⁷⁹ of complex but reversible reaction networks were achieved through metal catalysis⁸⁰ and biocatalysis.⁸¹ These are excellent examples of substrate specificity even in the presence of mixtures of compounds in equilibrium, which somehow relates to dynamic kinetic resolutions in asymmetric catalysis. For example, a mixture of α -immonitriles in dynamic equilibrium obtained by mixing up to thirteen different aldehydes was subjected to azomethine ylide formation in the presence of a chiral Ag(I)–Taniaphos complex and base. After the addition of dimethyl fumarate as dipolarophile, two diastereomers derived from furfural (19 and 20) were isolated as the exclusive products (Fig. 12).⁸⁰

A similar process was developed using lipases as catalysts. A dynamic mixture was generated with two aromatic aldehydes (21a,b) and four related imines (22a–d). Then, 2-sulfanyl acetate (23) and 2-nitropropane (24) were added, which could lead to the reversible formation of multiple products arising from thioacetal or thioaminal formation, or Henry reactions. Indeed, Henry adducts were detected in the equilibrating mixture, but in the presence of CAL-B as lipase and phenyl acetate as acyl donor, the complex system was resolved into virtually two cyclized products (25a,b) out of eighteen possible products (Fig. 13).⁸¹

Nevertheless, the use of dynamic systems to generate new catalytic entities might be more common. In this sense, again in Ramström's group, a dynamic combinatorial approach to the discovery of a bifunctional organocatalyst for the Morita–Baylis–Hillman (MBH) reaction was undertaken (Fig. 14). A series of amines (26a–d, each with a different catalytic function) and aldehydes (27a–d) were subjected to reversible imine formation, and the resulting mixture tested in the MBH reaction. Significant catalytic activity was disclosed, which after deconvolution and independent preparation, led to the discovery of an efficient catalyst (28) for such reaction, although stereoselectivities were not reported.⁸²

A more elaborated approach by the same group led to a kinetic self-sorting of a dynamic covalent catalyst for the MBH

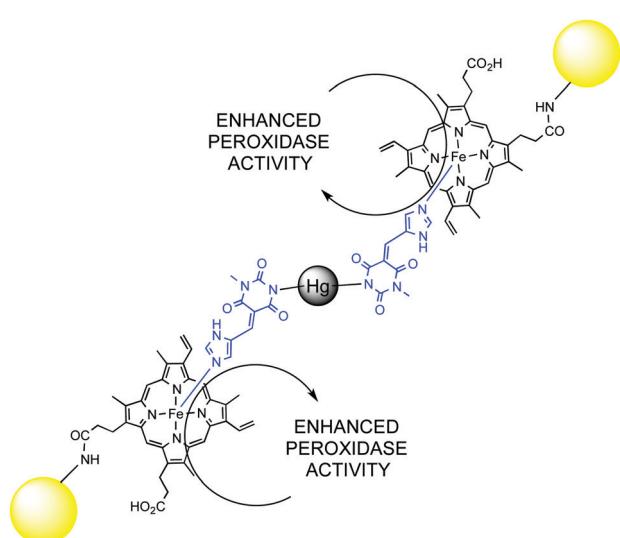


Fig. 10 Enhanced peroxidase activity of Au-NPs functionalized with hemin groups in the presence of Hg(II).



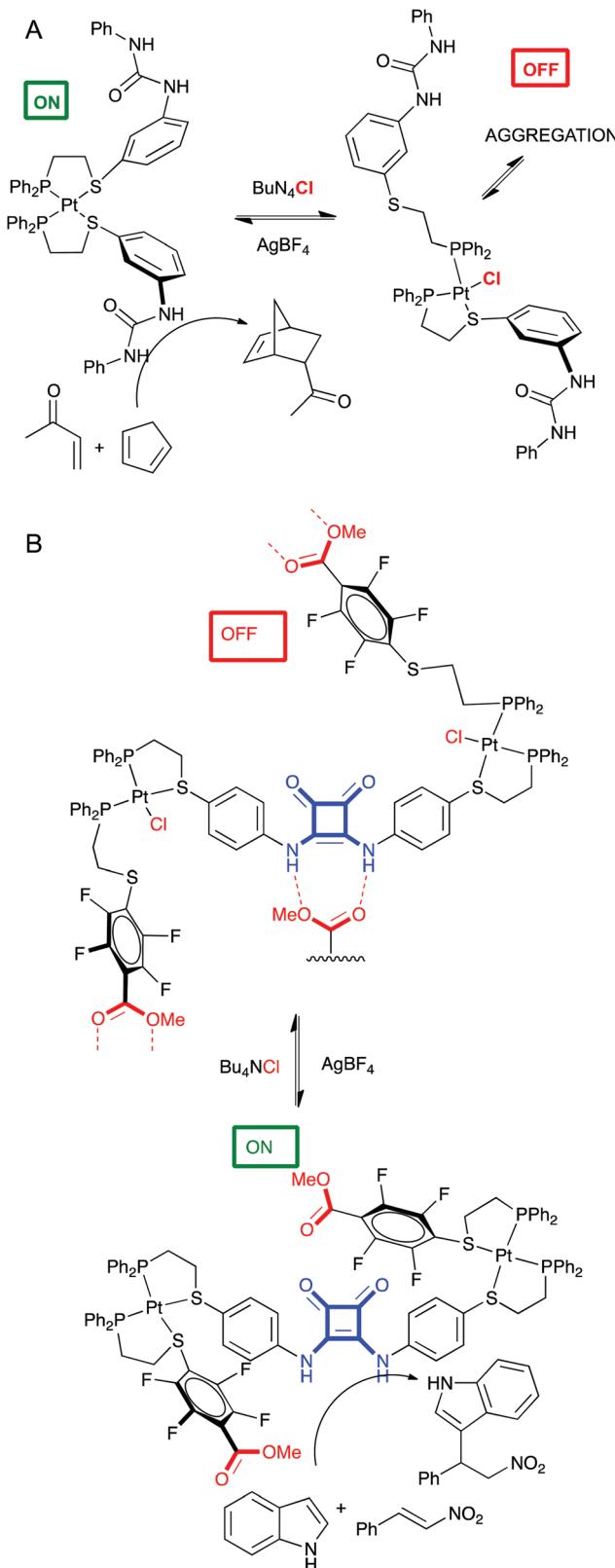


Fig. 11 Allosteric control of catalytic activity of a tweezer complex (A) or a triple decker complex (B) by chloride.

reaction, ultimately leading to feedback regulation. A family of quinuclidine catalysts with a reversible imine bond was tested

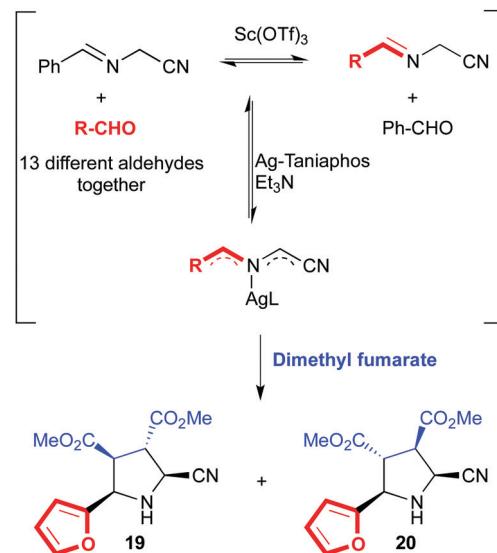


Fig. 12 Dynamic systematic resolution of α -immonitriles with silver(i)-Taniaphos.

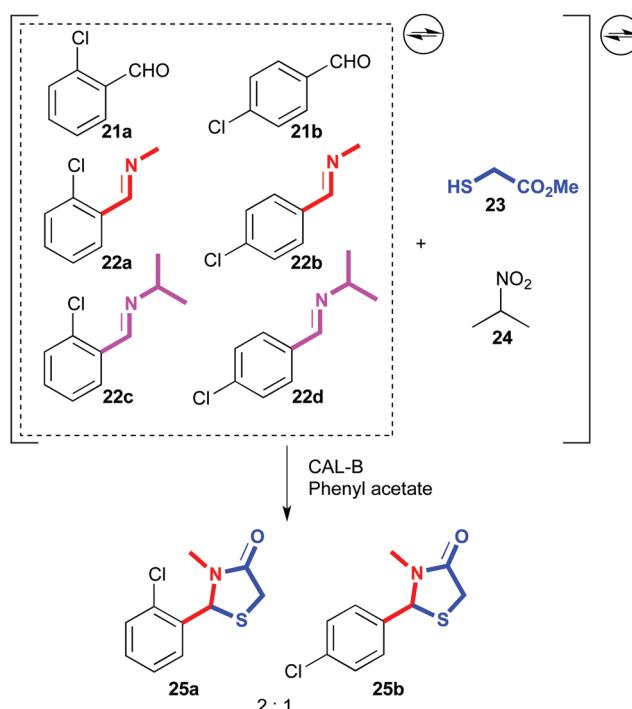


Fig. 13 Lipase-catalysed dynamic systematic resolution of aldehydes and imines.

in the above reaction showing a clear dependence of the reaction rate in the imine counterpart (arising from an aromatic aldehyde). Since an aromatic aldehyde was also used in the MBH reaction, this led to the development of a more or less active catalyst as a function of the aldehyde substrate chosen, thus being the substrate a key component to determine the catalyst structure and activity (Fig. 15).⁸³

Early examples of Dynamic Combinatorial Chemistry (DCC) applied to the discovery of new catalysts were actually

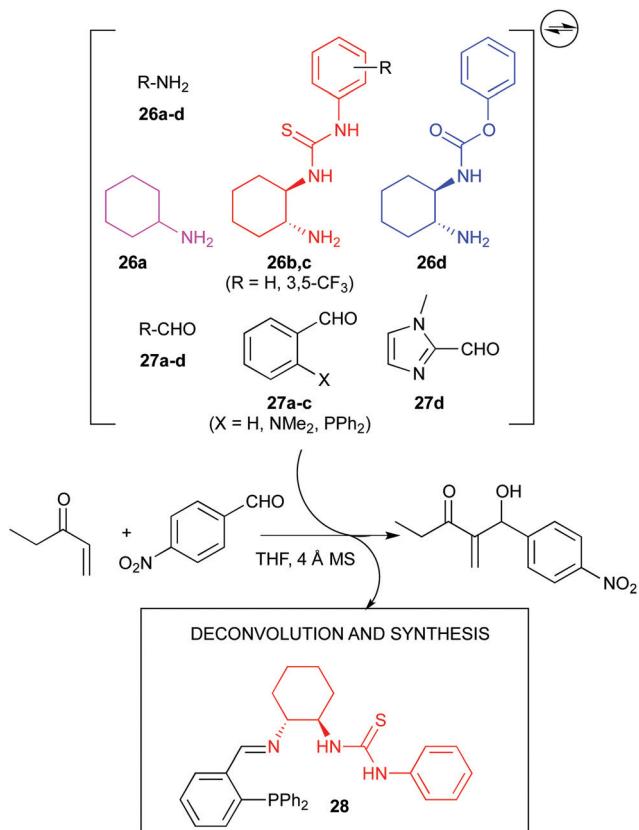


Fig. 14 Dynamic combinatorial approach to the discovery of an organo-catalyst for the MBH reaction.

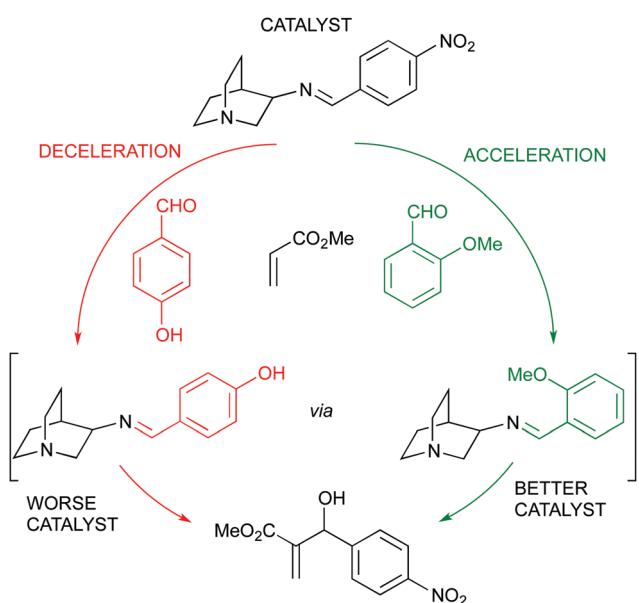


Fig. 15 Self-sorting of a dynamic covalent quinuclidine catalyst with substrate feedback regulation.

pioneered by Otto and co-workers, in particular through the use of disulphide exchange chemistry. A significant example is provided next: their initial findings focused on the acceleration

of the Diels–Alder (D–A) reaction through the adaptation of a library of disulphide ligands to a transition state analogue (TSA) as template. However, this catalytic reaction was hampered by product inhibition due to the similarities between the D–A adduct and the TSA.⁸⁴ Later on, they focused on the aza-Cope rearrangement (Fig. 16) using an ammonium-based TSA (**29**). With this strategy, the properly designed ammonium-containing substrate would be recognized by the DCC library and trigger the formation of its own catalyst, a mixture of tetrameric isomers (**30**). After the rearrangement, the product would hydrolyse, losing the positive charge and avoiding product inhibition. Moreover, the catalyst also disassembled after the reaction was complete, which can be used to control catalysis in complex chemical systems. Still, a stoichiometric quantity of catalyst was used.⁸⁵

Our own research group has been interested in complex catalytic systems, including the effect of water⁸⁶ and other additives⁸⁷ in reaction rate and stereoselectivity, as well as studying concentration effects.⁸⁸ We eventually developed a dynamic catalytic network based on the binding of functional pyridine ligands to zinc(II), which thus behaves as a structural co-factor, in order to generate bifunctional catalysts for the asymmetric aldol reaction. This catalytic network showed fast exchange rate, and no single species could be identified. Nevertheless, high conversion and stereoselectivity were observed.⁸⁹ Finally, in order to slow down the exchange process and favour the formation of more stable catalytic species, we developed the corresponding monofunctional bipyridine ligands **31** and **32** (Fig. 17).⁹⁰ We demonstrated that the complex network thus generated, with several potential catalysts, actually simplified under the reaction conditions making the bifunctional species to predominate in a kind of thermodynamic self-sorting. As expected, this species also provided the highest reaction acceleration and stereoselectivity, improving the performance of the previous pyridine-based system at lower catalyst loading. Moreover, its reversible nature made impossible the synthesis

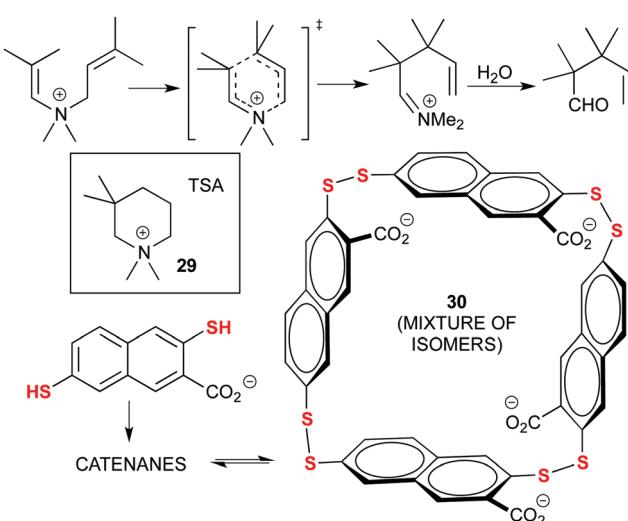


Fig. 16 Amplification of a tetrameric catalyst (**30**) from a DCC library of sulphide ligands for the aza-Cope rearrangement, by using a TSA (**29**) as the template.



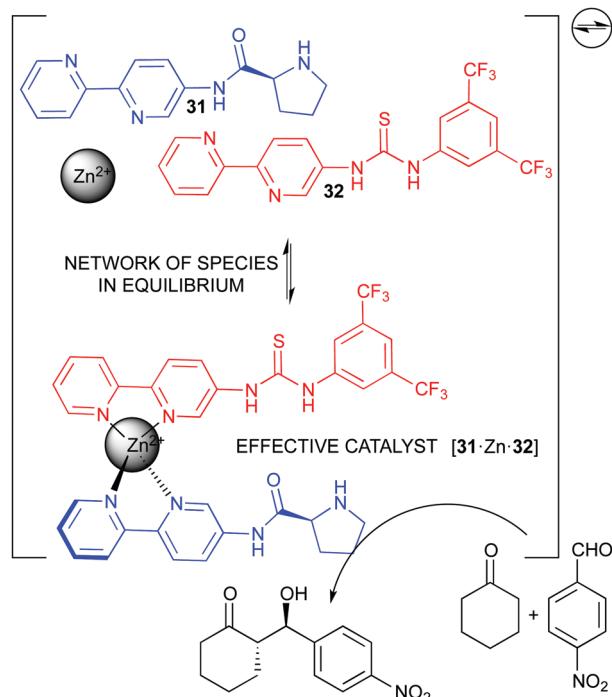


Fig. 17 Dynamic network of organocatalytic bipyridine ligands templated by Zn^{2+} .

and isolation of the catalytic complex, and the network generated with the three-components mixture was a convenient (and actually the only) manner of utilizing it.⁹⁰ Thus, our approach illustrated how a dynamic system can cooperatively work for the formation of an active catalyst, even in the presence of other species in equilibrium, which are intrinsically needed for the function of the system.

Other dynamic systems involve the use of vesicles or micelles due to the mobility of the self-assembled amphiphiles and therefore of the attached catalysts. Like in cell membranes, wherein enzymes and proteinic nanomachines are embedded in the lipidic bilayer, artificial vesicles or micelles provide a variety of new properties to catalysts or reactions different from the homogeneous systems. For instance, in our group we developed an amphiphilic catalyst for the asymmetric aldol reaction that featured an acylguanidinium group able to establish a strong hydrogen bonding network. Indeed, intramolecular hydrogen bonding in the acylguanidine group was identified as the key structural motif. This catalyst worked nicely in the organic/aqueous interphase, outperforming other amphiphilic catalysts.⁹¹ A recent and more sophisticated example from Chen's group shows how an amphiphile containing an *E*-photoresponsive group and a catalytic Zn^{2+} -triazacyclononane unit (33) self-assembles under visible light into a vesicle because of its *E*-configuration, which triggers catalytic activity towards transphosphorylation due to the cooperative action of the Zn^{2+} -containing groups on the vesicle's surface. On the contrary, upon UV irradiation, the *Z*-configuration predominates, therefore the vesicle disassembles and the catalytic activity is lost (Fig. 18). This is an excellent example of catalyst activity control

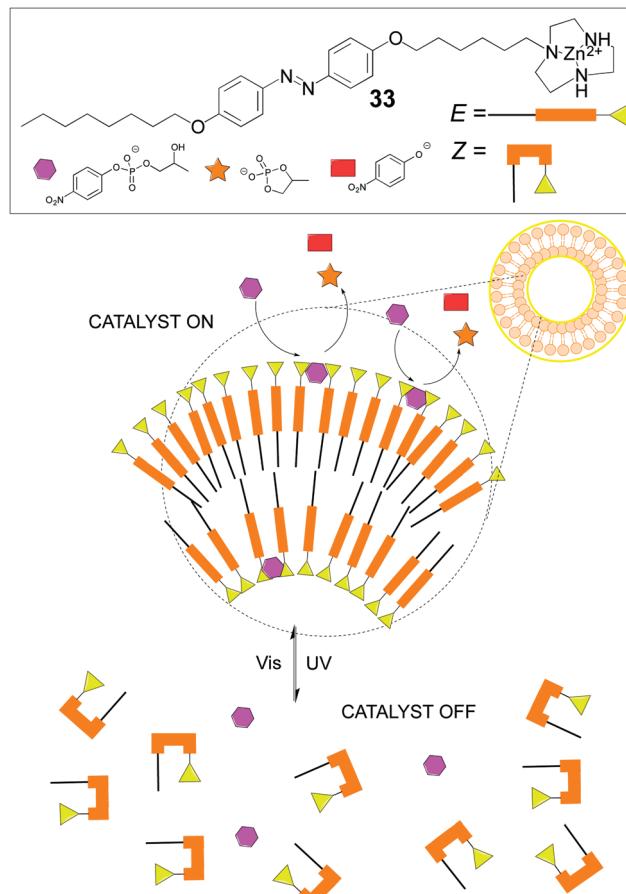


Fig. 18 Photoactivation–deactivation in an amphiphilic catalytic system.

through an external stimulus in a fully reversible fashion, with concomitant control of both the molecular catalyst and the structure of the corresponding assembly.⁹²

This section beautifully reflects the power of using complexity in catalysis with selected examples. The main advantages of such an approach are the regulation of catalytic activity by simple means and the self-assembly of complex catalysts from simpler BBs, as well as product selectivity, even when dealing with complex mixtures in equilibrium (either for substrates or catalysts). We expect that further developments in the field will lead eventually to cost-effective and practical catalysts suitable for industrial applications, but meanwhile it is clear that Nature is an endless source of inspiration for the creativity in supramolecular catalysis.

Conclusions

Complexity has been traditionally avoided in chemical research as an unwanted complication and undesired side effect for achieving efficiency. The improvement of analytical techniques for the characterization of complex mixtures in addition to the development of elaborated mathematical tools have paved the way for the design and controlled implementation of complexity as an additional tool or parameter in molecular science.



Following the growth of systems biology, the chemical counterpart systems chemistry has emerged as a subdiscipline with a bright future in different areas of research. Here, the use of dynamic combinatorial mixtures and stimuli-responsive networks for molecular recognition, sensing, chemical biology or catalysis has been illustrated with selected examples from the recent literature. Taking advantage of bioinspiration, several research groups have made this emergent field an appealing framework for the development of new functional materials and molecules. By constructing controlled mixtures of dynamic species, we and others have developed new chemical systems with potential applications in different fields. Although this can be considered a rather unconventional view to chemical design, it does not imply a very different methodological approach since complexity is inherent to molecular species. We think that complexity must be used in our benefit, instead of ignoring or avoiding its presence. We envision that the wise design of complex chemical systems will allow the emergence of new and even unexpected molecular properties and behaviours. With the progress of analytical instruments, we will be able to reduce the technical limitations of using complex systems in the near future, which will probably come from the capacity to rationalize and extract information from the data obtained. We have already shown that chemometric analysis may provide a solution to this bottleneck.³⁶

However, several challenges remain open for the future. The work in far-from-equilibrium systems is still in its infancy. The efficient implementation of fully synthetic systems able to replicate, merged with catalysis and compartmentalization would represent a big leap in the search for 'artificial life'.⁹³ Related to this, during the revision of the manuscript two important papers from the group of Otto have appeared. In a first study, the authors were able to combine a replicator and a cofactor that, when bound to the replicator, accelerates the formation of the components from which the replicator can emerge.⁹⁴ In a second study, the replicator is able to catalyse a retro-aldol reaction due to the local increased basicity. Moreover, the same replicator promotes the deprotection of a N-Fmoc-protected amino acid. The particularity here is that the product of the reaction is able to catalyse the thiol oxidation necessary to form the replicator, thus exerting a positive feedback on replication.⁹⁵

Last but not least, our final intention is not to create new terminology or to catalyse provocative discussion, but to encourage other chemists to reflect on the intrinsic complexity of molecules and their mixtures. This will hopefully create an opportunity to advance in basic knowledge, as well as on the impact of chemistry to solve the current societal challenges.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 N. Pearcy, N. Chuzhanova and J. J. Crofts, *J. Theor. Biol.*, 2016, **406**, 99–104.
- 2 J. Bascompte, M. B. García, R. Ortega, E. L. Rezende and S. Pironon, *Sci. Adv.*, 2019, **5**, eaav2539.
- 3 M. J. Aschwanden, N. B. Crosby, M. Dimitropoulou, M. K. Georgoulis, S. Hergarten, J. McAtee, A. V. Milovanov, S. Mineshige, L. Morales, N. Nishizuka, G. Pruessner, R. Sánchez, A. S. Sharma, A. Strugarek and V. Uritsky, *Space Sci. Rev.*, 2016, **198**, 47–166.
- 4 (a) J. M. Montoya, S. L. Pimm and R. V. Solé, *Nature*, 2006, **442**, 259–264; (b) S. Azalea, S. Pigolotti, J. R. Banavar and A. Maritan, *Nature*, 2006, **444**, 926–928.
- 5 S. P. Borgatti, A. Mehra, D. J. Brass and G. Labianca, *Science*, 2009, **323**, 892–895.
- 6 F. Schweitzer, G. Fagiolo, D. Sornette, F. Vega-Redondo, A. Vespignani and D. R. White, *Science*, 2009, **325**, 422–425.
- 7 M. Gosak, R. Markovič, J. Dolenšek, M. Slak Rupnik, M. Marhl, A. Stožer and M. Perc, *Phys. Life Rev.*, 2018, **24**, 118–135.
- 8 J.-M. Lehn, *Angew. Chem., Int. Ed.*, 2013, **52**, 2836–2850.
- 9 A. S. Y. Wong and W. T. S. Huck, *Beilstein J. Org. Chem.*, 2017, **13**, 1486–1497.
- 10 (a) E. Mattia and S. Otto, *Nat. Nanotechnol.*, 2015, **10**, 111–119; (b) I. Alfonso, *Chem. Commun.*, 2016, **52**, 239–250.
- 11 R. F. Ludlow and S. Otto, *Chem. Soc. Rev.*, 2008, **37**, 101–108.
- 12 (a) A. Aderem, *Cell*, 2005, **121**, 511–513; (b) Special issue devoted to systems biology: L. Chong and L. B. Ray, *Science*, 2002, **295**, 1661–1682.
- 13 G. Ashkenasy, T. Hermans, S. Otto and A. Taylor, *Chem. Soc. Rev.*, 2017, **46**, 2543–2554.
- 14 (a) J. R. Nitschke, *Nature*, 2009, **462**, 736–738; (b) S. Otto and A. R. Hunt, *Chem. Commun.*, 2011, **47**, 847–858; (c) J. Li, P. Nowak and S. Otto, *J. Am. Chem. Soc.*, 2013, **135**, 520–5126.
- 15 (a) S. J. Rowan, S. J. Cantrill, G. R. L. Cousins, J. K. M. Sanders and J. F. Stoddart, *Angew. Chem., Int. Ed.*, 2002, **41**, 898–952; (b) P. T. Corbett, J. Leclaire, L. Vial, K. R. West, J.-L. Wietor, J. K. M. Sanders and S. Otto, *Chem. Rev.*, 2006, **106**, 3652–3711; (c) J. N. H. Reek and S. Otto, *Dynamic Combinatorial Chemistry*, Wiley-VCH, Weinheim, 2010; (d) F. B. L. Cougnon and J. K. M. Sanders, *Acc. Chem. Res.*, 2012, **45**(12), 2211–2221; (e) Y. Jin, C. Yu, R. J. Denman and W. Zhang, *Chem. Soc. Rev.*, 2013, **42**, 6634–6654; (f) P. Frei, R. Hevey and B. Ernst, *Chem. – Eur. J.*, 2019, **25**, 60–73.
- 16 (a) J.-M. Lehn, *Chem. – Eur. J.*, 1999, **5**, 2455–2463; (b) J.-M. Lehn, *Chem. Soc. Rev.*, 2007, **36**, 151–160.
- 17 (a) R. L. E. Furlan, S. Otto and J. K. M. Sanders, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 4801–4804; (b) S. Otto, *Curr. Opin. Drug Discovery Dev.*, 2003, **6**, 509–520.
- 18 (a) P. T. Corbett, S. Otto and J. K. M. Sanders, *Chem. – Eur. J.*, 2004, **10**, 3139–3143; (b) I. Saur and K. Severin, *Chem. Commun.*, 2005, 1471–1473.
- 19 P. T. Corbett, S. Otto and J. K. M. Sanders, *Org. Lett.*, 2004, **6**, 1825–1827.
- 20 R. F. Ludlow and S. Otto, *J. Am. Chem. Soc.*, 2010, **132**, 5984–5986.
- 21 (a) I. Saur and K. Severin, *Chem. Commun.*, 2005, 1471–1473; (b) P. T. Corbett, J. K. M. Sanders and S. Otto, *J. Am. Chem. Soc.*, 2005, **127**, 9390–9392.
- 22 (a) S. Otto and S. Kubik, *J. Am. Chem. Soc.*, 2003, **125**, 7804–7805; (b) L. Vial, R. F. Ludlow, J. Leclaire, R. Pérez-Fernández and S. Otto, *J. Am. Chem. Soc.*, 2006, **128**, 10253–10257.
- 23 Selected examples: (a) S. Otto, R. L. E. Furlán and J. K. M. Sanders, *Science*, 2002, **297**, 590–593; (b) A. González-Álvarez, I. Alfonso, F. López-Ortiz, S. Aguirre, S. García-Granda and V. Gotor, *Eur. J. Org. Chem.*, 2004, 1117–1127; (c) T. S. R. Lam, A. Belenguer, S. L. Roberts, C. Naumann, T. Jarrosson, S. Otto and J. K. M. Sanders, *Science*, 2005, **308**, 667–669; (d) S. Ladame, A. Whitney



and S. Balasubramanian, *Angew. Chem., Int. Ed.*, 2005, **44**, 5736–5739; (e) A. Bugaut, K. Jantos, J. L. Wietor, R. Rodriguez, J. K. M. Sanders and S. Balasubramanian, *Angew. Chem., Int. Ed.*, 2008, **47**, 2677–2680; (f) S. Hamieh, V. Saggiomo, P. Nowak, E. Mattia, R. F. Ludlow and S. Otto, *Angew. Chem., Int. Ed.*, 2013, **52**, 12368–12372; (g) R. Brachvogel, F. Hampel and M. von Delius, *Nat. Commun.*, 2015, **6**, 7129.

24 M. Corredor, D. Carbajo, C. Domingo, Y. Pérez, J. Bujons, A. Messeguer and I. Alfonso, *Angew. Chem., Int. Ed.*, 2018, **57**, 11973–11977.

25 D. Carbajo, Y. Pérez, J. Bujons and I. Alfonso, *Angew. Chem., Int. Ed.*, 2020, **59**, 17202–17206.

26 (a) M. Samiappan, Z. Dadon and G. Ashkenasy, *Chem. Commun.*, 2011, **47**, 710–712; (b) X. Dadon, M. Samiappan, N. Wagner and G. Ashkenasy, *Chem. Commun.*, 2012, **48**, 1419–1421.

27 (a) N. Giuseppone and J.-M. Lehn, *Chem. – Eur. J.*, 2006, **12**, 1715–1722; (b) L. Tauk, A. P. Schröder, G. Decher and N. Giuseppone, *Nat. Chem.*, 2009, **1**, 649–656.

28 S. Sobczak, W. Drozd, G. I. Lampronti, A. M. Belenguer, A. Katrusiak and A. R. Stefankiewicz, *Chem. – Eur. J.*, 2018, **24**, 8769–8773.

29 (a) J. Atcher, A. Moura and I. Alfonso, *Chem. Commun.*, 2013, **49**, 487–489; (b) J. Atcher, A. Moura, J. Bujons and I. Alfonso, *Chem. – Eur. J.*, 2015, **21**, 6869–6878.

30 (a) J. Zhang, L. You, E. V. Anslyn and X. Qian, *Chem. – Eur. J.*, 2012, **18**, 1102–1110; (b) D. Zamora-Olivares, T. S. Kaoud, K. N. Dalby and E. V. Anslyn, *J. Am. Chem. Soc.*, 2013, **135**, 14814–14820; (c) L. You, D. Zhaa and E. V. Anslyn, *Chem. Rev.*, 2015, **115**, 7840–7892.

31 (a) J. Montenegro, P. Bonvin, T. Takeuchi and S. Matile, *Chem. – Eur. J.*, 2010, **16**, 14159–14166; (b) T. Takeuchi, J. Montenegro, A. Hennig and S. Matile, *Chem. Sci.*, 2011, **2**, 303–307; (c) T. Takeuchi and S. Matile, *Chem. Commun.*, 2013, **49**, 19–29.

32 (a) A. Buryak, A. Pozdnoukhov and K. Severin, *Chem. Commun.*, 2007, 2366–2368; (b) S. Rochat, J. Gao, X. Qian, F. Zaubitzer and K. Severin, *Chem. – Eur. J.*, 2010, **16**, 104–113; (c) Z. Köstereli, R. Scopelliti and K. Severin, *Chem. Sci.*, 2014, **5**, 2456–2460.

33 (a) F. A. Scaramuzzo, G. Licini and C. Zonta, *Chem. – Eur. J.*, 2013, **19**, 16809–16813; (b) C. Bravin, A. Guidetti, G. Licini and C. Zonta, *Chem. Sci.*, 2019, **10**, 3523–3528.

34 (a) A. Osypenko, S. Dhers and J.-M. Lehn, *J. Am. Chem. Soc.*, 2019, **141**, 12724–12737; (b) G. Men and J.-M. Lehn, *Chem. Sci.*, 2019, **10**, 90–98.

35 J. Atcher, J. Solà and I. Alfonso, *Org. Biomol. Chem.*, 2017, **15**, 213–217.

36 A. M. Valdivielso, F. Puig-Castellví, J. Atcher, J. Solà, R. Taulera and I. Alfonso, *Chem. – Eur. J.*, 2017, **23**, 10789–10799.

37 K.-D. Zhang and S. Matile, *Angew. Chem., Int. Ed.*, 2015, **54**, 8980–8983.

38 H. M. Seifert, K. R. Trejo and E. V. Anslyn, *J. Am. Chem. Soc.*, 2016, **138**, 10916–10924.

39 M. Martínez-Amezaga, A. G. Orrillo and R. L. E. Furlan, *Chem. Sci.*, 2019, **10**, 8338–8347.

40 B. M. Matysiak, P. Nowak, I. Cyrtila, C. G. Pappas, B. Liu, D. Komáromy and S. Otto, *J. Am. Chem. Soc.*, 2017, **139**, 6744–6751.

41 S. N. Semenov, L. J. Kraft, A. Ainla, M. Zhao, M. Baghbanzadeh, V. E. Campbell, K. Kang, J. M. Fox and G. M. Whitesides, *Nature*, 2016, **537**, 656–660.

42 B. J. Cafferty, A. S. Y. Wong, S. N. Semenov, L. Belding, S. Gmür, W. T. S. Huck and G. M. Whitesides, *J. Am. Chem. Soc.*, 2019, **141**, 8289–8295.

43 M. He and J.-M. Lehn, *J. Am. Chem. Soc.*, 2019, **141**, 18560–18569.

44 (a) K. West, K. Bake and S. Otto, *Org. Lett.*, 2005, **7**, 2615–2618; (b) A. R. Stefankiewicz and J. K. M. Sanders, *Chem. Commun.*, 2013, **49**, 5820–5822.

45 (a) J. Solà, M. Lafuente, J. Atcher and I. Alfonso, *Chem. Commun.*, 2014, **50**, 4564–4566; (b) M. Lafuente, J. Atcher, J. Solà and I. Alfonso, *Chem. – Eur. J.*, 2015, **21**, 17002–17009; (c) M. Lafuente, I. Alfonso and J. Solà, *ChemSystemsChem*, 2019, **1**, 25–31.

46 M. Lafuente, J. Solà and I. Alfonso, *Angew. Chem., Int. Ed.*, 2018, **57**, 1–5.

47 (a) K. R. West, R. F. Ludlow, P. T. Corbett, P. Besenius, F. M. Mansfeld, P. A. G. Cormack, D. C. Sherrington, J. M. Goodman, M. C. A. Stuart and S. Otto, *J. Am. Chem. Soc.*, 2008, **130**, 10834–10835; (b) M.-K. Chung, S. J. Lee, M. L. Waters and M. R. Gagne, *J. Am. Chem. Soc.*, 2012, **134**, 11430–11443; (c) F. B. L. Cougnon, N. Ponnuswamy, N. A. Jenkins, G. D. Pantos and J. K. M. Sanders, *J. Am. Chem. Soc.*, 2012, **134**, 19129–19135.

48 (a) N. Ponnuswamy, F. B. L. Cougnon, J. M. Clough, G. D. Pantos and J. K. M. Sanders, *Science*, 2012, **338**, 783–785; (b) N. Ponnuswamy, F. B. L. Cougnon, G. D. Pantos and J. K. M. Sanders, *J. Am. Chem. Soc.*, 2014, **136**, 8243–8251.

49 B. Liu, C. G. Pappas, E. Zangrandi, N. Demitri, P. J. Chmielewski and S. Otto, *J. Am. Chem. Soc.*, 2019, **141**, 1685–1689.

50 (a) A. J. Bissette and S. P. Fletcher, *Angew. Chem., Int. Ed.*, 2013, **52**, 12800–12826; (b) T. Kosikova and D. Philp, *Chem. Soc. Rev.*, 2017, **46**, 7274–7305.

51 (a) K. Ruiz-Mirazo, C. Briones and A. de la Escosura, *Chem. Rev.*, 2014, **114**, 285–366; (b) J. D. Sutherland, *Angew. Chem., Int. Ed.*, 2016, **55**, 104–121.

52 T. Tjivikua, P. Ballester and J. Rebek, *J. Am. Chem. Soc.*, 1990, **112**, 1249–1250.

53 S. Xu and N. Giuseppone, *J. Am. Chem. Soc.*, 2008, **130**, 1826–1827.

54 W. Sadownik and D. Philp, *Angew. Chem., Int. Ed.*, 2008, **47**, 9965–9970.

55 R. Nguyen, L. Allouche, E. Buhler and N. Giuseppone, *Angew. Chem., Int. Ed.*, 2009, **48**, 1093–1096.

56 K. Severin, D. H. Lee, J. A. Martinez, M. Vieth and M. R. Ghadiri, *Angew. Chem., Int. Ed.*, 1998, **37**, 126–128.

57 R. Mukherjee, R. Cohen-Luria, N. Wagner and G. Ashkenasy, *Angew. Chem., Int. Ed.*, 2015, **54**, 12452–12456.

58 J. Nanda, B. Rubinov, D. Ivnitski, R. Mukherjee, E. Shtelman, Y. Motro, Y. Miller, N. Wagner, R. Cohen-Luria and G. Ashkenasy, *Nat. Commun.*, 2017, **8**, 434.

59 J. M. A. Carnall, C. A. Waudby, A. M. Belenguer, M. C. A. Stuart, J. J.-P. Peyralans and S. Otto, *Science*, 2010, **327**, 1502–1506.

60 (a) M. Malakoutikhah, J. J.-P. Peyralans, M. Colomb-Delsuc, H. Fanlo-Virgos, M. C. A. Stuart and S. Otto, *J. Am. Chem. Soc.*, 2013, **135**(49), 18406–18417; (b) M. Colomb-Delsuc, E. Mattia, J. W. Sadownik and S. Otto, *Nat. Commun.*, 2015, **6**, 7427.

61 D. Komáromy, M. Tezcan, G. Schaeffer, I. Marić and S. Otto, *Angew. Chem., Int. Ed.*, 2017, **56**, 14658–14662.

62 J. W. Sadownik, E. Mattia, P. Nowak and S. Otto, *Nat. Chem.*, 2016, **8**, 264–269.

63 Y. Altay, M. Tezcan and S. Otto, *J. Am. Chem. Soc.*, 2017, **139**, 13612–13615.

64 M. Altay, Y. Altay and S. Otto, *Angew. Chem., Int. Ed.*, 2018, **139**, 10564–10568.

65 S. M. Morrow, I. Colomer and S. P. Fletcher, *Nat. Commun.*, 2019, **10**, 1011.

66 I. Colomer, A. Borissov and S. P. Fletcher, *Nat. Commun.*, 2020, **11**, 176.

67 J. Sreenivasachary and J.-M. Lehn, *Proc. Natl. Acad. Sci. U. S. A.*, 2005, **102**, 5938–5943.

68 S. Toledano, R. J. Williams, V. Jayawarna and R. V. Ulijn, *J. Am. Chem. Soc.*, 2006, **128**, 1070–1071.

69 M. Kumar, N. L. Ing, V. Narang, N. Wijerathne, A. I. Hochbaum and R. V. Ulijn, *Nat. Chem.*, 2018, **10**, 696–703.

70 J. Bokhoven, W. E. Hendriksen, G. J. M. Koper, R. Eelkema and J. H. van Esch, *Science*, 2015, **349**, 1075–1079.

71 (a) E.-K. Bang, G. Gasparini, G. Molinard, A. Roux, N. Sakai and S. Matile, *J. Am. Chem. Soc.*, 2013, **135**, 2088–2091; (b) G. Gasparini, E.-K. Bang, G. Molinard, D. V. Tulumello, S. Ward, S. O. Kelley, A. Roux, N. Sakai and S. Matile, *J. Am. Chem. Soc.*, 2014, **136**, 6069–6074.

72 C. Bouillon, D. Paolantoni, J. C. Rote, Y. Bessin, L. W. Peterson, P. Dumy and S. Ulrich, *Chem. – Eur. J.*, 2014, **20**, 14705–14714.

73 C. Bouillon, Y. Bessin, F. Poncet, M. Gary-Bobo, P. Dumy, M. Barboiu, N. Bettache and S. Ulrich, *J. Mater. Chem. B*, 2018, **6**, 7239–7246.

74 (a) M. Vlatkovic, B. S. L. Collins and B. L. Feringa, *Chem. – Eur. J.*, 2016, **22**, 17080–17111; (b) B. A. Grzybowski, K. Fitzner, J. Paczesny and S. Granick, *Chem. Soc. Rev.*, 2017, **46**, 5647–5678; (c) A. M. Lifschitz, M. S. Rosen, C. M. McGuirk and C. A. Mirkin, *J. Am. Chem. Soc.*, 2015, **137**, 7252–7261; (d) N. Kumagai and M. Shibasaki, *Catal. Sci. Technol.*, 2013, **3**, 41–57.

75 L. Zhang, F. Ma, J. Lei, J. Liu and H. Ju, *Chem. Sci.*, 2017, **8**, 4833–4839.

76 (a) A. I. d'Aquino, H. F. Cheng, J. Barroso-Flores, Z. S. Kean, J. Mendez-Arroyo, C. M. McGuirk and C. A. Mirkin, *Inorg. Chem.*, 2018, **57**, 3568–3578; (b) N. C. Gianneschi, S.-H. Cho, S. T. Nguyen and C. A. Mirkin, *Angew. Chem., Int. Ed.*, 2004, **43**, 5503–5507.



77 C. M. McGuirk, C. L. Stern and C. A. Mirkin, *J. Am. Chem. Soc.*, 2014, **136**, 4689–4696.

78 C. M. McGuirk, J. Méndez-Arroyo, A. M. Lifschitz and C. A. Mirkin, *J. Am. Chem. Soc.*, 2014, **136**, 16594–16601.

79 M. Sakulsombat, Y. Zhang and O. Ramström, *Top. Curr. Chem.*, 2012, **322**, 55–86.

80 L. Hu and O. Ramström, *Chem. Commun.*, 2014, **50**, 3792–3794.

81 (a) Y. Zhang and O. Ramström, *Chem. – Eur. J.*, 2014, **20**, 3288–3291; (b) Y. Zhang, P. Vongvilai, M. Sakulsombat, A. Fischer and O. Ramström, *Adv. Synth. Catal.*, 2014, **356**, 987–992.

82 F. Schaufelberger and O. Ramström, *Chem. – Eur. J.*, 2015, **21**, 12735–12740.

83 F. Schaufelberger and O. Ramström, *J. Am. Chem. Soc.*, 2016, **138**, 7836–7839.

84 B. Brisig, J. K. M. Sanders and S. Otto, *Angew. Chem., Int. Ed.*, 2003, **42**, 1270–1273.

85 H. Fanlo-Virg, A.-N. R. Alba, S. Hamieh, M. Colomb-Delsuc and S. Otto, *Angew. Chem., Int. Ed.*, 2014, **53**, 11346–11350.

86 C. Jimeno, *Org. Biomol. Chem.*, 2016, **14**, 6147–6164.

87 Z. I. Günler, X. Companyó, I. Alfonso, J. Burés, C. Jimeno and M. A. Pericàs, *Chem. Commun.*, 2016, **52**, 6821–6824.

88 Z. Inci Günler, I. Alfonso, C. Jimeno and M. A. Pericàs, *Synthesis*, 2017, 319–325.

89 (a) A. Serra-Pont, I. Alfonso, C. Jimeno and J. Solà, *Chem. Commun.*, 2015, **51**, 17386–17389; (b) A. Serra-Pont, I. Alfonso, J. Solà and C. Jimeno, *Org. Biomol. Chem.*, 2017, **15**, 6584–6591.

90 A. Serra-Pont, I. Alfonso, J. Solà and C. Jimeno, *Chem. Commun.*, 2019, **55**, 7970–7973.

91 A. M. Valdivielso, A. Catot, I. Alfonso and C. Jimeno, *RSC Adv.*, 2015, **5**, 62331–62335.

92 C. Z.-J. Ren, P. Solís Muñana, J. Dupont, S. S. Zhou and J. L.-Y. Chen, *Angew. Chem., Int. Ed.*, 2019, **58**, 15254–15258.

93 (a) P. Adamski, M. Eleveld, A. Sood, Á. Kun, A. Szilágyi, T. Czárán, E. Szathmáry and S. Otto, *Nat. Rev. Chem.*, 2020, **4**, 386–403; (b) E. A. J. Post and S. P. Fletcher, *Chem. Sci.*, 2020, **11**, 9434–9442.

94 G. M. Santiago, K. Liu, W. R. Browne and S. Otto, *Nat. Chem.*, 2020, **12**, 603–607.

95 J. Ottelé, A. S. Hussein, C. Mayer and S. Otto, *Nat. Catal.*, 2020, **3**, 547–553.

