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# Multicolor monitoring of cellular organelles by single wavelength excitation to visualize the mitophagy process†

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Multiplexed cellular organelle imaging using single wavelength excitation is highly desirable for unravelling cellular functions but remains challenging. This requires the design of organelle specific fluorophores with distinct emission but similar absorption. Herein, we present two unique aggregation-induced emission (AIE) probes to track mitochondria and lysosomes simultaneously with emission colors that can be distinguished from that of the nucleus stain Hoechst 33342 upon single wavelength excitation. Compared to conventional organelle stains, the two AIE probes have larger Stokes shifts and higher photostability, which endow them with the capability to monitor bioprocesses, such as mitophagy with strong and sustained fluorescent signals. Moreover, both probes can also stain intracellular organelles in zebrafish larvae with good cell-penetrating capabilities, showing their great potential to monitor bioprocesses *in vivo*.

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## Introduction

Mitochondria are the powerhouses of cells, and they also play important roles in calcium signaling and cell apoptosis.<sup>1</sup> However, damaged mitochondria tend to generate excessive reactive oxygen species, which is highly undesirable.<sup>2</sup> In fact, the accumulation of dysfunctional mitochondria is believed to be associated with a range of human pathologies.<sup>3</sup> Fortunately, the physiological system is able to maintain its health by regulating cellular metabolism through a specific mechanism known as mitophagy. In general, mitophagy serves as a key process to control mitochondrial quality as well as to facilitate autophagic processes to degrade damaged mitochondria.<sup>4</sup> Specifically, during the mitophagy process, damaged mitochondria are driven into autophagosomes and are then delivered to lysosomes to be degraded and recycled.

To investigate the mitophagy process, fluorescent probes that can stain mitochondria and lysosomes have been tested.<sup>5–7</sup> Among all the fluorescent moieties, organic dyes are the most

promising<sup>8,9</sup> because of their non-invasiveness and small size, although they share the drawbacks of having small Stokes shifts, being easily photobleached<sup>10</sup> and undergoing facile aggregation-caused quenching (ACQ).<sup>11</sup> In addition, two wavelengths are often needed to excite both dyes to emit different colors. Therefore, the wavelength of the excitation laser has to be constantly switched, or two lasers with different excitation wavelengths have to be used simultaneously during mitophagy monitoring.

To minimize the complexity of fluorescence detection and simplify instrumentation requirements, probes that can be excited by a single-excitation wavelength to emit different colors are highly attractive.<sup>12</sup> Specifically, two probes which are highly specific to mitochondria and lysosomes with different light emission upon single wavelength excitation would provide a convenient way of visualizing the mitophagy process. However, to date, dyes that can be excited with a single wavelength to generate different emission colors are quite limited. Although quantum dots (QDs) with broad excitation spectra<sup>13</sup> and dual-luminophore-doped nanoparticles with fluorescence resonance energy transfer (FRET) characteristics<sup>14</sup> are able to emit distinguishable fluorescence under single wavelength excitation, they are not suitable for organelle tracking or mitophagy monitoring. This is because it is highly complex to endow them with organelle-targeting abilities and they generally have low performance in organelle imaging.<sup>15</sup>

Recently, fluorogens with aggregation-induced emission (AIEgens) characteristics have been developed for numerous biological assays, including organelle imaging,<sup>16</sup> specific protein/enzyme detection,<sup>17</sup> bacteria identification,<sup>18</sup> and many others.<sup>19,20</sup> In contrast to traditional ACQ dyes, AIEgens show

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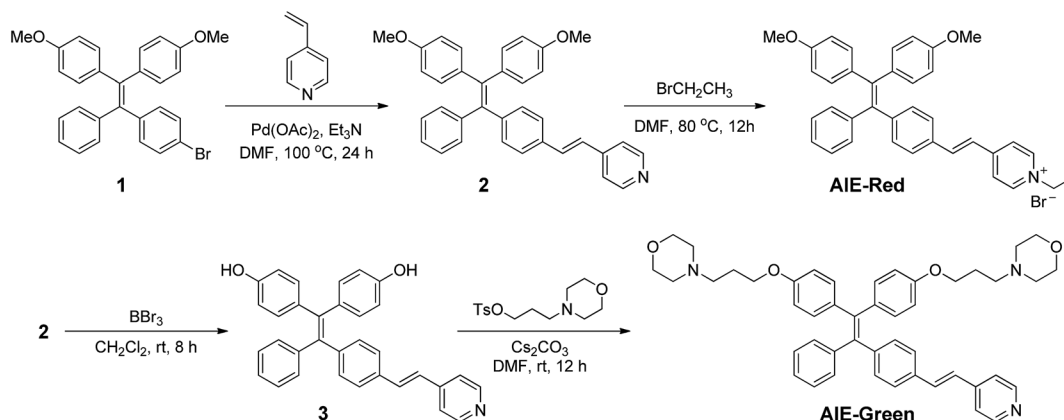
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Scheme 2 The synthetic routes to obtain AIE-Red and AIE-Green (DMF represents dimethyl formamide).

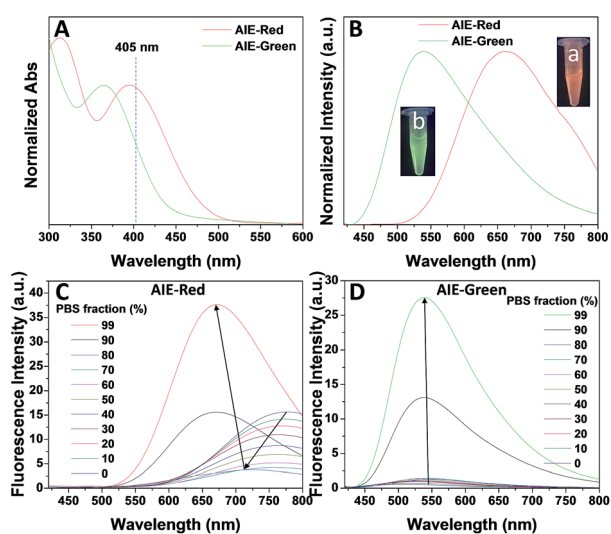


Fig. 1 The normalized absorption (A) and fluorescence (B) spectra of AIE-Red (10  $\mu$ M) and AIE-Green (10  $\mu$ M) in PBS buffer (pH = 7.4, containing 0.5% DMSO). The insets show the optical photographs of these solutions obtained under 365 nm light irradiation. (C and D) Fluorescence spectra of AIE-Red (10  $\mu$ M), (C) and AIE-Green (10  $\mu$ M), (D) in DMSO/PBS mixtures with different volume fractions of PBS from 0 to 99%. The excitation wavelength was 405 nm for all the fluorescence spectra.

introduction of different ICT does not lead to big differences in the absorption spectra of AIE-Red and AIE-Green. Upon incorporation of different electron-accepting groups to AIE-Red (vinyl pyridinium) and AIE-Green (vinyl pyridine), the difference of their emission maxima (127 nm) is much larger than that of their absorption maxima (27 nm). This enables the excitation of the two probes with a single wavelength for different emission colors. These unique photoproperties originate primarily from the large Stokes shifts of AIE-Red (271 nm) and AIE-Green (171 nm).

The AIE behaviors of AIE-Red and AIE-Green were investigated in a dimethyl sulfoxide (DMSO, good solvent) and PBS (poor solvent) mixture. As shown in Fig. 1C, AIE-Red exhibits red and NIR emission with a maximum at 775 nm in a dilute

DMSO solution. The emission intensity decreased gradually with the increment of the PBS fraction from 0 to 60%, due to the twisted intramolecular charge transfer (TICT) effect. With higher PBS fractions, the emission is recovered with a much higher intensity and a blue-shift with a maximum at 665 nm. This fluorescence enhancement is attributed to the restriction of intramolecular motions, which is activated by the formation of nanoparticles of AIE-Red in aqueous media. This is different to the case of AIE-Green, which was almost non-emissive in the dilute DMSO solution (Fig. 1D). The emission intensity increases gradually as the PBS fraction increases from 0 to 80%. Then, a sharp intensity increase is noted as the PBS fraction goes up from 80% to 99%. As shown in Fig. S9,<sup>†</sup> the average hydrodynamic sizes of AIE-Red and AIE-Green in solutions of PBS/DMSO (99/1, v/v) are 160 and 400 nm, respectively, measured by dynamic light scattering (DLS). Owing to the AIE properties, both AIE-Red and AIE-Green can be used in high concentrations with reliable fluorescent signals. The fluorescence quantum yields (QYs) of AIE-Red and AIE-Green in PBS are 1.8% and 5.2%, respectively, which are measured with reference to 4-(dicyanomethylene)-2-methyl-6-(*p*-dimethylaminostyryl)-4*H*-pyran (DCM). After addition of 10% fetal bovine serum (FBS) to mimic the living organism system, their QYs increased to 8.1% and 7.2%, respectively. The increased QYs in FBS solution are beneficial for cellular system tracking. The pyridine moiety in AIE-Green could be protonated in acidic lysosomes, which may affect the emission of AIE-Green. Therefore, the pH-dependent fluorescence spectra of AIE-Red and AIE-Green were measured (Fig. S10<sup>†</sup>). AIE-Red exhibits constant fluorescence, while AIE-Green exhibits gradually decreased fluorescence along with pH reduction. The fluorescence decrease of AIE-Green is ascribed to the partial protonation of the pyridine moiety. As the emission wavelength remains unchanged, it will not affect the multicolor tracking by single wavelength excitation. Additionally, the high concentration of these AIE probes leads to their enhanced photostability. As can be seen in Fig. 2, AIE-Red and AIE-Green are far more resistant to photobleaching as compared to the commercial mitochondria and lysosome trackers. Signals from MitoTracker® Green FM (Mito Tracker) disappeared almost





Fig. 2 Relative fluorescence intensities of AIE-Red (A, red), Mito Tracker (A, green), AIE-Green (C, green) and Lyso Tracker (C, red) under continuous scanning; confocal images of HeLa cells stained with AIE-Red (B, lower panel), Mito Tracker (B, upper panel), AIE-Green (D, lower panel) and Lyso Tracker (D, upper panel) after being scanned various times. The scale bar is 20  $\mu\text{m}$ .

completely at the 60<sup>th</sup> scan, while signals from AIE-Red remained stable (Fig. 2A and B). Similarly, the signal loss of LysoTracker® Red DND-99 (Lyso Tracker) was 82% after 60 scanning cycles, while the signal loss of AIE-Green was only 8% (Fig. 2C and D). Both AIE-Red and AIE-Green also have good biocompatibility even at high concentrations (Fig. S11<sup>†</sup>). Since AIE-Red is reported as a photosensitizer, which can produce toxic reactive oxygen species (ROS) upon white light irradiation, the cytotoxicity of AIE-Red under confocal laser irradiation is also studied. As shown in Fig. S12,<sup>†</sup> upon continuous irradiation of AIE-Red incubated HeLa cells by a confocal laser for 10 min, the cell morphology hardly changed, indicating negligible cytotoxicity. Its biocompatibility was further confirmed using a live/dead assay after continuous confocal laser irradiation for 10 min. In these images, the green signals from fluorescein diacetate (FDA) indicate living cells, and the lack of red signals from propidium iodide (PI) indicates that there are no dead cells. The HeLa cell viability remained above 80% after treatment with AIE-Red or AIE-Green at 15  $\mu\text{M}$  for 24 h. The high photostability and excellent biocompatibility of AIE-Red or AIE-Green are highly attractive for monitoring cellular bioprocesses. As commercial organelle targeting dyes are commonly used at a very low concentration, we also compared the fluorescence intensities of AIE-Red and AIE-Green to the commercial Mito Tracker and Lyso Tracker at different concentrations (Fig. S13<sup>†</sup>). Mito Tracker (150 nM) has a sharp emission at around 525 nm in PBS, while AIE-Red (150 nM) is almost non-emissive. The negligible emission of AIE-Red at 150 nM is ascribed to its good water solubility and lack of AIE activation. After addition of 10% FBS to mimic the living organism system, the emission of AIE-Red (150 nM) is dramatically enhanced to the same level as that

of Mito Tracker (150 nM) due to AIE activation by interaction with proteins. When the concentration of AIE-Red is increased to 5  $\mu\text{M}$ , the emissions in PBS and FBS-containing PBS are both stronger than those of Mito Tracker (150 nM). AIE-Green (150 nM) has a much stronger emission at 525 nm than Lyso Tracker (150 nM), which is quite weak with a maximum at 590 nm in both PBS and FBS-containing PBS. The emissions of AIE-Green at 5  $\mu\text{M}$  are even stronger. These results indicate that AIE-Red has almost the same fluorescence intensity as Mito Tracker at 150 nM in the FBS-mimicked living organism system, and AIE-Green has a much stronger fluorescence intensity than Lyso Tracker at 150 nM. However, AIE-Red and AIE-Green at 5  $\mu\text{M}$  still have very good biocompatibility. The usage of AIE-Red and AIE-Green at high concentration is aimed at high photostability for continuous scanning.

### Multicolor imaging of organelles and mitophagy monitoring

We subsequently studied the mitochondrion specificity of AIE-Red and lysosome specificity of AIE-Green in HeLa cells by the colocalization of both probes with commercial organelle dyes. As shown in Fig. 3A, AIE-Red and Mito Tracker were simultaneously incubated with HeLa cells for 15 min at 37 °C. The red emission signals from AIE-Red overlapped with the green signals from Mito Tracker very well, with a Pearson correlation coefficient of 0.94, indicating the outstanding targeting ability of AIE-Red towards the mitochondria of living HeLa cells. Similarly, the lysosome specificity of AIE-Green was well confirmed by its colocalization with Lyso Tracker, with a Pearson correlation coefficient of 0.95 (Fig. 3B). AIE-Red and AIE-Green also exhibit organelle targeting abilities toward other cell





Fig. 3 Confocal images of HeLa cells after incubation with AIE-Red (5  $\mu\text{M}$ ), Mito Tracker (150 nM) and the overlay image (A); AIE-Green (5  $\mu\text{M}$ ), Lyso Tracker (150 nM) and the overlay image (B); overlay image of HeLa cells after incubation with AIE-Red (5  $\mu\text{M}$ ), AIE-Green (5  $\mu\text{M}$ ) and Hoechst 33342 (150 nM) (C). The scale bar is 20  $\mu\text{m}$ .

lines such as MCF-7 and MDA-MB-231 cells. As shown in Fig. S14,<sup>†</sup> the colocalizations with commercial organelle targeting dyes are good in MCF-7 and MDA-MB-231 cells. The Pearson correlation coefficients of **AIE-Red** with Mito Tracker are 0.73 and 0.92 for MCF-7 and MDA-MB-231 cells, respectively. The Pearson correlation coefficients of **AIE-Green** with Lyso Tracker are 0.81 and 0.82 for the MCF-7 and MDA-MB-231 cells, respectively. We then investigated the distinguishable emission colors of both probes based on the 405 nm excitation wavelength inside the HeLa cells. As shown in Fig. 3C, the organelle images with highly distinguishable emission colors were obtained inside the HeLa cells after simultaneous incubation with **AIE-Red**, **AIE-Green** and Hoechst 33342. The networks of mitochondria were clearly shown by the red signals while the morphology and distribution of lysosomes were portrayed well through green dots. These two distinguishable emission colors under the same visual field of the living cells afford the monitoring of mitophagy using only a single excitation laser. The blue nucleus signals originate from the commercial Hoechst 33342, which can also be excited by the 405 nm laser.

With the advantages of distinguishable emission colors upon single wavelength excitation and high specificity to mitochondria and lysosomes, **AIE-Red** and **AIE-Green** were further utilized to monitor the mitophagy process in real time. In these experiments, rapamycin, which can promote the mitophagy process, was incubated with HeLa cells pre-treated with **AIE-Red** and **AIE-Green**. Subsequently, the confocal images of rapamycin-incubated HeLa cells were continuously collected under 405 nm excitation. As shown in Fig. 4A, after rapamycin treatment for 1 min, the red signals from the mitochondria hardly overlapped with the green signals from the lysosomes, with a Pearson correlation coefficient of 0.23. This indicates that the occurrence of the mitophagy process was not notable. As time went on, the Pearson correlation coefficient increased gradually, specifically, to 0.36 at 30 min and 0.46 at 60 min, indicating that the occurrence of the mitophagy process became more pronounced. Meanwhile, the evident increase in the amount and intensity of the yellow signals with a prolonged rapamycin treatment, as noted in the enlarged images, provide visual evidence of the mitophagy process (Fig. 4D–F). During



Fig. 4 Overlay confocal images of AIE-Red- and AIE-Green-stained HeLa cells after treatment with rapamycin (50  $\mu\text{g mL}^{-1}$ ) at 1 min (A), 30 min (B), and 60 min (C) and the enlarged images at 1 min (D), 30 min (E) and 60 min (F). The scale bars are 20  $\mu\text{m}$  for (A–C) and 5  $\mu\text{m}$  for (D–F).

these experiments, all fluorescent signals were collected upon excitation with a 405 nm laser, which is more convenient as compared to conventional methods.

#### Organelle imaging in zebrafish

Since the zebrafish is a transparent and versatile *in vivo* model, multicolor imaging of different cellular organelles in zebrafish larvae was also explored. After soaking the larvae in the **AIE-Red**

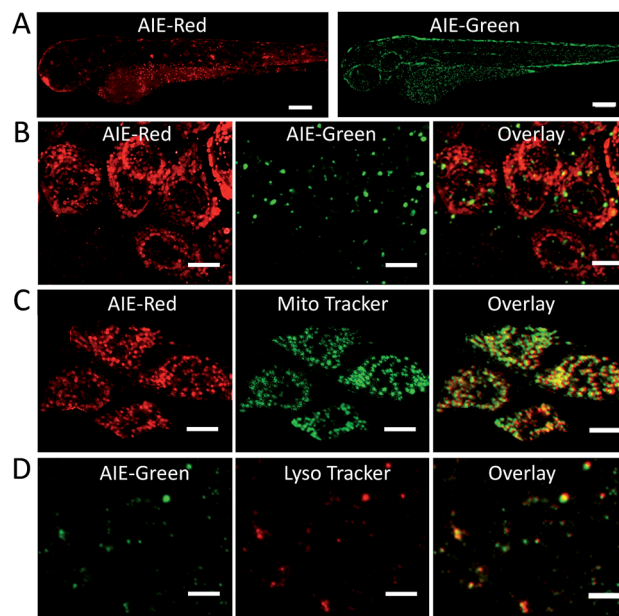


Fig. 5 (A) Confocal images of zebrafish larvae after soaking with AIE-Red (5  $\mu\text{M}$ ) and AIE-Green (5  $\mu\text{M}$ ) for 30 min. The scale bar is 30  $\mu\text{m}$ ; (B) enlarged images of zebrafish larvae skin cells. The scale bar is 10  $\mu\text{m}$ . Enlarged photos of confocal images for zebrafish larvae after incubation with AIE-Red (5  $\mu\text{M}$ ) and Mito Tracker (150 nM) and the overlay picture (C); AIE-Green (5  $\mu\text{M}$ ), Lyso Tracker (100 nM) and the overlay picture (D). The scale bar is 10  $\mu\text{m}$ .



