



Cite this: *RSC Adv.*, 2017, 7, 51206

Stagnation point flows in analytical chemistry and life sciences

Ayoola T. Brimmo ^{ab} and Mohammad A. Qasaimeh ^{*ab}

Isolated microfluidic stagnation point flows – stagnation point flows formed within microfluidic interfaces – have come a long way as a tool for characterizing materials, trapping/manipulating micro particles, and generating confined flows and localized chemistries. Early applications of these fluidic stagnation flows focused on characterizing emulsions and polymers. However, in recent times, these flows have taken microfluidic forms to expand the range of applications to single-cell analysis, substrate patterning, and chip-integrated devices, amongst others. This article focuses on reviewing the microfluidics literature to highlight the concept behind isolated microfluidic stagnation point flows, underline different approaches of experimentally resolving them, and then look into their applications in chemistry and life sciences; with a specific focus on micro-total-analysis systems. The literature on this topic is approached historically; from the isolated fluidic stagnation point flow generated by the early four-roll mill to the currently predominating microfluidic stagnation point flows of the cross-slot device. Finally, isolated microfluidic stagnation flows produced by the relatively recent open space microfluidics – opposed jets, microfluidic probes and micropipettes – are critically analyzed and their applications are discussed. Ultimately, the goal of this article is to inform the scientific community on the constructive aspect of microfluidic stagnation flows by demonstrating their past, present, and future applicability in analytical chemistry and life sciences.

Received 10th October 2017
 Accepted 25th October 2017

DOI: 10.1039/c7ra11155j

rsc.li/rsc-advances

1. Introduction

Microfluidic stagnation is a topic that merges the field of microfluidics and hydrodynamic stagnations with a goal of developing and controlling a zero-velocity (stagnation) region within fluid interfaces. In as much as both fields are already fast evolving due to their evident advantages for analytical chemistry and biological research, their merger proposes a more telling advancement opportunity. While microfluidic devices offers an interesting avenue to develop integrated lab-on-chip devices for micro scale reagent volumes, hydrodynamic stagnations increases the resolution of this field to single micro-particles within the reagents. Such capability facilitates single cell resolutions in biological and life science studies, and this could hold the key to currently unresolved medical problems. Isolated microfluidic stagnation flow takes this concept further by generating the stagnation points away from walls/boundaries, and by using only hydrodynamic forces to manipulate them. The main advantage of this approach lay with its ability to isolate the response of trapped molecules/particles to a relatively less invasive environment – several techniques has been

used for isolating target particles (*e.g.* optical, acoustic, and magnetic tweezers), but they require the introduction of secondary fields that could modify the chemistry of trapped particles and hence interfere with the characterization process. For instance, optical tweezers could lead to secondary heating and magnetic tweezers require pre-labeling; which could both affect cell viability. Other advantages of microfluidic stagnations include: localization of chemical reactions, increased maneuverability of trapped particles, increased viability of analyzed bio-particles, and its “open-flow” nature that can accommodate the isolation of larger particle sizes.

The development of microfluidic stagnation flows is as rich in its history as it is in its physical applications. As such, this article attempts to critically review its advancement from a historical perspective – important applications are analyzed in a chronological progression in order to also highlight the concept's evolution with time. However, before we go into an in-depth analysis, the following sections present an evolution sketch of the overarching stagnation and microfluidic concepts, theoretical formulations of microfluidic stagnation point flows, and a broad perspective of applications covered in this article.

1.1 Stagnations and microfluidics

Fluidic stagnation is a phenomenon whereby a body of fluid is immobile within a certain region *i.e.* local velocity is zero. In certain type of flows, this region is limited to a dimensionless

^aDivision of Engineering, New York University Abu Dhabi, Saadiyat Island, 129188, Abu Dhabi, UAE. E-mail: maq4@nyu.edu

^bTandon School of Engineering, New York University, 6 MetroTech Center, Brooklyn, New York, USA



$$u_1 = \beta(x_1(1 + \lambda) + x_2(1 - \lambda)) \quad (3)$$

$$u_2 = -\beta(x_1(1 - \lambda) - x_2(1 + \lambda)) \quad (4)$$

This gives rise to the family of streamlines described by:

$$Z = (x_1 + x_2)^2 - \lambda(x_1 - x_2)^2 \quad (5)$$

Plotting the streamlines for the different types of flows results in the images in Fig. 2a–c. Similarly, Fig. 2d–f shows experimentally captured images of these stagnation point flow fields, specified by eqn (5).

Evidently, eqn (5) constitutes the fundamental formulation of these types of flow, and presents a mean of observing them in their theoretical forms. Applications of the stagnation point flow inherent in their structures are the focus of this review. The following sections highlight developments in the field, classified according to the stagnation point flow generation devices.

1.3 Broad perspectives

The primary function of isolated microfluidic stagnations is to trap particles in a fluid region engineered to have zero velocity gradient. Depending on the mechanism used to generate the stagnation point flow, the trapped object can be further manipulated for a wide range of practical applications in material characterization, localized chemistries, and applications of Micro-Total-Analysis Systems (μ TAS) such as: droplet deformation and break up,^{10,32–35} droplet coalescing,^{36–39} birefringence of polymers,^{31,40,41} manipulation of biological particles *in vitro*,^{42,43} and single cell analysis,⁴⁴ among others. These applications of microfluidic stagnation point flows can be

traced back to 1934, where it was used by Taylor to investigate the dynamics of stirring multi-phase fluids.¹⁰ However, amidst its long history and wide range of theoretical and experimental considerations, there still is no review that cut across the full range of isolated microfluidic stagnation point flows. Consequently, this article reviews the concept behind isolated microfluidic stagnations, the different approaches of experimentally resolving the stagnation point flow under steady and transient conditions, and then look into their applications in chemistry, material characterization and life science. In Section 2, we look into the four-roll mill as the pioneering stagnation point flow-generating device. Although, the four-roll mill cannot be categorically classified as a microfluidic device based on the aforementioned definition, advances in the design of the four-roll mill have greatly contributed to the technological development of microfluidic stagnation point flows; hence our consideration of this device is also of scientific relevance. In Section 3, we delve into the literature of the most predominantly used microfluidic device for generating stagnation point flow – the cross-slot device. Section 4 summarizes the state-of-the-art of stagnation point flows generated by modern open space microfluidic systems.⁴⁵ Finally, Section 5 presents our view on the future prospects of microfluidic stagnations and our recommendations for the field's critical success factors.

2. Four roll mill

The four-roll mill is a device that consists of four cylindrical rollers with centers positioned at the edge of a hypothetical square, rotating either in opposing or concurrent pairs.

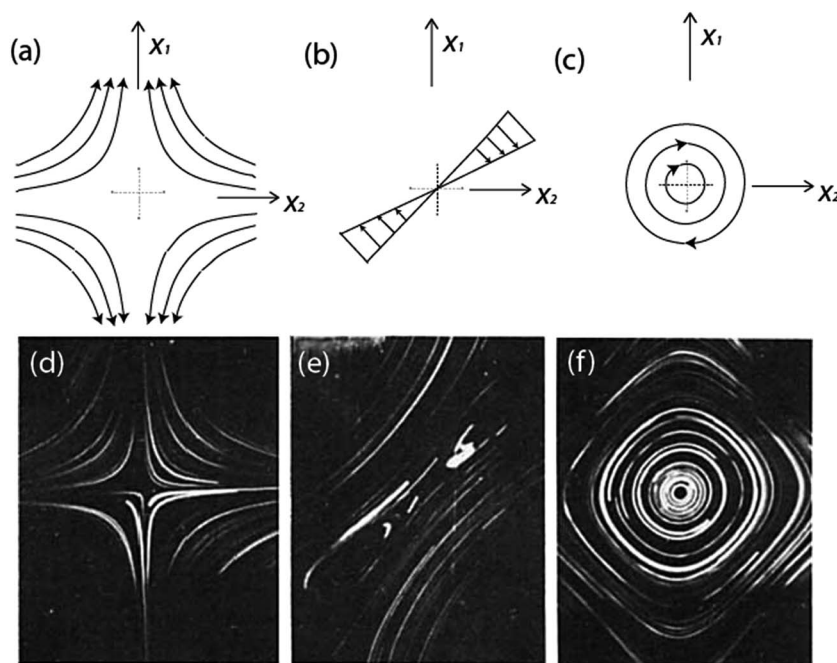


Fig. 2 Streamline schematics of stagnation point flow fields. (a) Extensional flow ($\lambda = 1$). (b) Shear flow ($\lambda = 0$). (c) Rotational flow ($\lambda = -1$). Experimental captured photographs for stagnation point flow field. (d) $\lambda = 1$. (e) $\lambda = 0$. (f) $\lambda = -1$. Type parameter varies from pure extensional flow to pure rotational flow. Reprinted with permission. Copyright© 1981 John Wiley & Sons, Inc.³¹



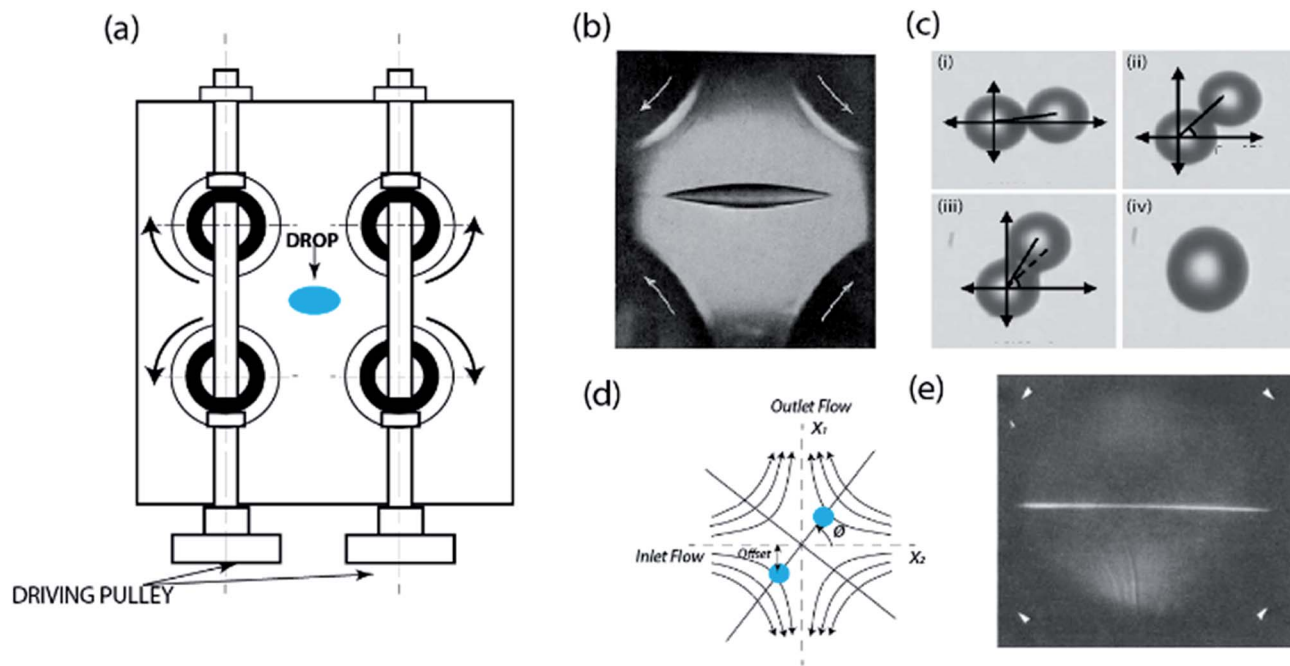


Fig. 3 The pioneering application of the stagnation point flows. (a) Schematic of the original four-roll mill for fluidic applications. The diameter of each of rolls is 2.39 cm and the gaps between the rollers are 1 cm. (b) Experimental drop elongation study when trapped within the stagnation point of an extensional flow (stagnation point flow). Sizes of the studied drops ranged from about 800–2000 μm .¹⁰ Reprinted with permission. Copyright© 1934, The Royal Society. (c) Experimental visualization of drops coalescence. Reprinted with permission. Copyright© AIP Publishing LLC. (i) contact of the two drops, (ii) thinning of film separating drops, (iii) coalescence, and (iv) daughter droplet formation of the coalescence process.³⁹ (d) Schematic of the four-roll mill generated flow field with the drop position offset.³⁹ Reprinted with permission. Copyright© AIP Publishing LLC. (e) Localized flow birefringence in the four-roll mill. Photographs showing flow birefringence for a 1.5% polyethylene oxide/water solution and velocity gradient of 6 s^{-1} . Arrow indicates orientation of rotation.⁴⁰ Reprinted with permission. Copyright© 1976 John Wiley & Sons, Inc.

Developed originally as mills for crushing materials in the 19th century,⁴⁶ the device has since evolved into fluidic applications. The fluidic four-roll mill (see Fig. 3a) produces a two-dimensional laminar flow field, and depending on the roller speed and configuration, can be used to produce the full range of stagnation point flows described in the previous section. These flows are typically visualized by mounting the setup on top of a microscope. The use of the four-roll mill to generate flow fields can be traced back to 1934 when Taylor used a “four roller” to create an extensional flow (stagnation point flow) in order to study the deformation and burst of drops in an emulsion.¹⁰ The diameter of each of his rolls was 2.39 cm and he studied drops ranging from about 800–2000 micrometers in size. As such, this specific four-roll mill cannot be categorically labeled a microfluidic technology. However, since this device evolved into miniaturized version and eventually led to the development of microfluidic stagnation point flow devices, its inclusion in this review holds important historical and technical relevance. The following sections summarize application of four-roll mill generated stagnation point flows for drop dynamics and polymer birefringence studies.

2.1 Drop deformation and break-up

The problem of establishing the break up mechanism of a single drop of fluid, and in particular, the number of different

drops a given dimension of droplet would break into, was classically highlighted and theorized by Rayleigh and others.^{47,48} Experimentally, these works were based on monitoring the breakup of drops, introduced into a pressurized bulk fluid through a hole. This technique neglects the effect of the surrounding fluid’s disruptive viscous drag on the bursting of drops, and hence reveals very little about the stirring dynamics of these fluids.^{49,50} As the principal objective of mixing devices are to increase the surface area at the interface of both phases – for increased mass and heat transfer—these studies did not produce key information intended for designing blenders, and emulsifiers.^{33,35,51} In order to circumvent these drawbacks, Taylor adapted the four-roll mill generated stagnation point flow (see Fig. 3a and b) to control the bulk fluid’s viscosity and rate of deformation through viscous drag, and monitor the corresponding interfacial surface tension.¹⁰ In Taylor’s work, a drop of fluid immiscible in water was trapped in the stagnation point of a water-based extensional flow and extended by varying the rollers’ speed until the drop eventually bursts. When the droplet is introduced into the low speed extensional flow, it takes a spherical shape, in which its surface tension exceeds the viscous and dynamic stresses of the bulk extensional flow. However, as the speed of the rollers increases, the flow’s viscous force surpasses the cohesive forces of the drop making it to extend and eventually burst.⁵² The four-roll mill allows for the



emulsion or polymer blend, which eventually affects the final product's macroscopic properties.⁷⁴ Although it has been established that drops collision do not necessarily lead to coalescence, collision is a necessary precursor for any coalescence process.^{75–77} In the event of coalescence, after collision, the thin film separating the two drops flatten-out, while the translational motion of the drops is impeded and the two drops become small enough for non-hydrodynamic attractive forces to cause rupture and then coalescence.^{38,78} Fig. 3c shows an experimentally captured time-lapse of this process.

Coalescence plays an equally important role in liquid–liquid extraction⁷⁹ and polymer blending process,⁸⁰ as much as drop break up does. Studying this process in isolation has proven to be beneficial in engineering the final properties of the resulting blend. However, compared to drop break up, relatively less experimental investigations have been carried out on coalescence due to its more complicated nature; coalescence requires two drops to collide and remain together long enough for film drainage to occur.⁸¹ This process is extremely difficult to control as such, classically, the drop collision topic has been predominantly approached from a theoretical perspective,^{82–87} with Batchelor and Green's solution (in 1972) to its creeping flow problem being a pioneer.⁸⁸ A few studies performed experiments to verify developed theories.^{89,90} However, most of these studies used Couette flow, with collision established under the influence of simple shear, and/or electric fields.^{91,92} In the typically applied Couette apparatus, drops were placed in the suspending liquid using a needle probe so that on applying shear and/or electric fields, they assumed trajectories and collided relative to the equatorial planes.^{89,93} This required a tedious drop control mechanism in order to effect collision. To ease the drop manipulation process that precedes collision, and extend the range of flows to include pure shear and extensional flows, Treathway *et al.*³⁶ and Hu *et al.*⁶² considered using the four-roll mill to generate the bulk fluid motion. Treathway *et al.* complimented previous drop collision and coalescence studies in shear flow and the buoyancy-driven flows by investigating this phenomenon in a four-roll mill generated extensional flow – manipulated by an active control scheme.³⁶ Their approach included splitting parent silicon-based Newtonian drops and trapping two equal volume daughter drops at the stagnation point of a four-roll mill generated extensional flow of castor oil, while monitoring the trajectory and collision of the droplets. This work aided the validation of existing theories for estimating trajectory, collision, and observing the dynamics of interacting spherical drops. Similarly, Yang *et al.*³⁷ used the “miniaturized” four-roll mill to study the drop coalescence phenomenon but found that the predictions of Chesters' theoretical formulation⁷⁶ did not quantitatively agree with their experimental results. For collisions when the initial position offset between the drops are relatively small, the measured time for film drainage was found to be shorter, and the critical capillary numbers were smaller than the predictions of the model (see Fig. 3d). Also, for larger initial drop offsets, experimentally measured film drainage was delayed relative to predictions by theoretical models.

Coalescence of polymer drops has also been carried out using the four-roll mill. Ha *et al.*³⁸ extended a previous study⁶² on copolymer drop coalescence; by varying the viscosity ratio and using fixed copolymer interface concentrations to ensure drop size independence. The “miniaturized” computer-controlled four-roll mill was also used to investigate the coalescence process for two equal-sized polybutadiene drops suspended in polydimethylsiloxane.^{39,94} While some studies focused on the head-on configuration,³⁹ others considered both head-on and glancing configurations.⁹⁴

In addition to providing a means of avoiding the effect of forces, stagnation point flows also brought an improved droplet control capability to the coalescing studies. Credit for this feature can be attributed to the automation efforts during droplet deformation studies. As such, early droplet deformation studies can be seen as a prerequisite the feats achieved in coalescing studies. This is the main reason that while droplet coalescing studies are technically more complicated than deformation and break-up studies, more efforts towards miniaturization were found in the coalescence literature – Borell *et al.*³⁹ and Yoon *et al.*⁹⁴ considered droplets of 10–100 μm . However, it must be noted that neither of these studies provided details about the exact size of their four-roll mills so very little can be said about the extent of miniaturization that can be achieved with the device for applications in the microscale.

2.3 Flow birefringence of polymer

Flow birefringence describes an anisotropic state of flow – a state where flow properties depend on the direction of propagation.^{95,96} Using birefringence measurements, the molecular weight distribution of a polymer blend can be calculated if the molecular weight and coil-stretch relaxation time are known.^{97,98} Investigations on this phenomenon have been predominantly performed using the Couette-type apparatus with the solution contained within the annular gap concentric cylinders.⁹⁹ However, observed polymer birefringence in these flows only slightly deform the polymer chains with a uniform intensity between the inner and outer cylinder.⁴⁰ This is in contrast to the observed marked localized birefringence when the extensional and compressional flows are used for polymer birefringence.¹⁰⁰ This revelation combined with the potential of studying birefringence in a wide range of flow types motivated the use of a four-roll mill by Crowley *et al.* for polymer birefringence.⁴⁰ In their study, birefringence of polyethylene oxide solutions (region where polymer molecules were significantly stretched) was observed to be localized at the area close to the outgoing asymptotic plane, and held at the stagnation point, as shown in Fig. 3e. More striking, localizations were observed with the four-roll mill and this was attributed to the use of extensional flows, which elongate molecules in such a way that only elements close to the “outgoing” asymptotic axis are exposed to the field for the required amount of polymer elongation time. Subsequently, the four-roll mill was used to orient and measure the deformation response of polymers to flow fields.¹⁰¹ The aim of this study was to understand and ultimately characterize flow-induced changes of molecules in polymer



solutions, which could dictate the eventual property of a polymer blend.¹⁰² These studies have also been predominantly motivated by the wide range of flow types the device offers, as early studies in this line had focused on just shear flows.⁴⁰

Using the four-roll mill generated stagnation point flow; Fuller and Leal^{30,31} found that the normalized birefringence of polystyrene dissolved in polychlorinated biphenyl (dilute solution) approached a saturation value at high velocity gradients, in purely extensional flows. Birefringence saturation was found to be consistent with the state where the chains are nearly fully extended, independent of concentration and molecular weight, and in agreement with theoretical estimates of Tanner.¹⁰³ Transient birefringence measurements in pure extensional flows revealed that birefringence went through a pronounced overshoot in time.³¹ More recently, transient extension of polymers was numerically simulated and the local strain at the stagnation point proved useful in interpreting birefringence data⁴¹ and used to experimentally investigate flow instabilities as a function of Weissenberg number.¹⁰⁴

The improved control offered by the computerized four-roll mill served as the impetus for the successful application of microfluidic stagnation point flows for birefringence studies. The highlight for this technique is that the birefringence phenomenon led to a more telling saturation – that can be interpreted as full extensions of the polymer – when confined to the plane generated by the microfluidic stagnation point. In addition to its promise as a particle extension device, this technique facilitated localized polymer examination and evolved to a classical technique for inferring the degree of polymer extension,¹⁰⁵ to become an alternative to light scattering technique that could imply deformations of only two to four times the equilibrium size.¹⁰⁶ As opposed to the averaging procedure adapted by the optical technique, the isolated stagnation point increased the resolution to single polymer molecules, which eliminated polymer–polymer interactions and polymer-induced alterations of the flow field. Such integrations ultimately led to an improved blending of polymer solutions, and to the discovery of an interesting overshoot phenomenon that is still an open topic in the field.³¹

3. Cross-slot microfluidics

In microfluidics, the cross-slot generally refers to a device with multiple channels intersecting at the center of the configuration. These devices can be used to generate microfluidic stagnation point flows of all types by concurrently injecting and withdrawing fluids through the channels. In 1979, use of the cross slot (called elongation flow cell back then) as an experimental apparatus for generating stagnation point flows came into the light as an equivalent of Taylor's four-roll mill without rollers. It served as an alternative that offered the possibility of exploiting the fine flow control in the microscale,^{107,108} which is otherwise complicated feat with the four-roll mill.¹⁰⁹

The archetypal microfluidic cross-slot consisted of four channels arranged in resemblance of the four arms of a cross, with flows injected *via* two opposite channels and concurrently aspirated out *via* the other two channels. Imaging of the

resulting flow was performed using microscopy techniques,¹¹⁰ however; X-ray techniques have been reported to provide higher spatial resolution.^{111,112} Fig. 4a shows a schematic of this device with injection carried out *via* channels in the compression axis, and aspiration *via* channels in the elongation axis. In this configuration, when all flow rates are equal, a purely extensional flow is created and the stagnation point is formed at the center.

The use of this device can be traced back to when Scrivener *et al.* applied the stagnation point flow generated by the “cross flow” apparatus to investigate the dynamics of polymer molecules during birefringence.¹⁰⁷ Although the original configuration proposed back then is still the most predominantly used today, various modifications have been made to optimize flow characteristics and extend the device's range of application. The following sections highlight key modifications of the cross-slot's configuration and trapping mechanism, report on its flow transition characteristics, and reviews its application in particle dynamics studies and DNA sequence detection.

3.1 Configurations

Over the years, several investigations have been carried out on either optimizing the typical cross-slot device or improving its range of application. In 2004, Phelan and Hudson sought a “microfluidic analog” of the four-roll mill and used a numerical approach to evaluate candidates of channel flows, which could generate the full range of linear flows.¹⁰⁹ In their follow-up studies,^{108,113} they developed a nifty asymmetric configuration consisting of six intersecting channels (see Fig. 4c(i)). This configuration was derived by combining the pure stretching capability of the prototypical configuration, with rotational conformations introduced by two channels arms; offsetting the opposing arm of the four-arm configuration. However, the asymmetric nature means that pure rotational flows cannot be obtained. To overcome this, a microfluidic channel based device that keeps the stagnation point at the center, and the flow symmetric, was demonstrated (see Fig. 4c(ii)).¹¹⁴ This allows for generation of the full range of flow types; rotational, shear and extensional. The appropriate central cavity radius and orifice size for this device was later specified through spectral boundary element numerical simulations.¹¹⁵

Haward *et al.*¹¹⁶ followed the recommendation of Alves' numerical optimization of the cross-slot flow geometry (see Fig. 4c(iii)),¹¹⁷ to facilitate an enhanced definition of the extension rate. Experiments showed that this optimized shape, provide an expansion of the inlet and outlet channels domains characterized by a nominally constant extension rate.^{116,118} Very recently, the configuration of the cross-slot evolved to a more complicated form, which constitutes six intersecting channels arranged in a symmetric manner (see Fig. 4c(iv)).^{9,119} These have been portrayed to be capable of producing an even wider range of flow patterns¹¹⁹ and multiple stagnation points.⁹

In order to minimize the fluid volumes used, the stagnation point flow generated by the cross-slot device has been combined with an oscillatory flow, to create an extensional flow oscillatory rheometer.¹²⁰ Oscillation was achieved by using four micro-piezoelectric pumps situated at the end of each slot channel



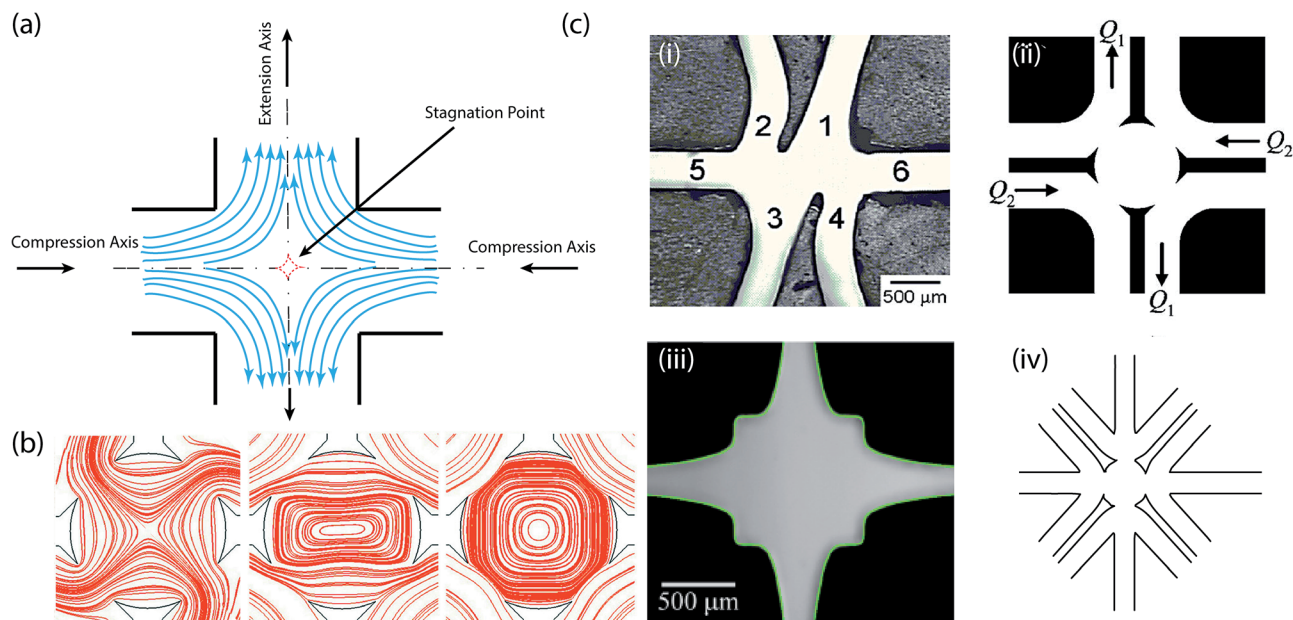


Fig. 4 The microfluidic cross-slot device. (a) Prototypical design. (b) The application of the cross-slot microfluidic device to create stagnation point flows of all types; numerically calculated flow-type streamlines.¹²⁴ Reprinted with permission. Copyright© 1976 John Wiley & Sons, Inc. (c) Geometric modification of the cross-slot microfluidic device. (i) Micrograph of the first microfluidic analog of the four-roll mill. Reprinted with permission. Copyright© AIP Publishing LLC. (ii) Schematic diagrams of a microfluidic four-roll mill devices that can generate all types of stagnation point flows shown in (b).¹²⁴ Reprinted with permission. Copyright© AIP Publishing LLC. (iii) Micrograph of an optimized cross-slot geometry.¹¹⁶ Reprinted with permission. Copyright© 2012 American Physical Society. (iv) Schematic of a six channel cross-slot device.

to provide the required repetitive flow profile. An additional advantage of this configuration lay in its clean and inert conditions that are ideal for biological samples; drawn from the enclosed cross-slot nature.¹²¹ Odell and coworker used the cross-slot configuration to measure extensional viscosity of polystyrene and hyaluronan polymer solutions.¹²⁰ This application was later extended to the characterization of ultra-dilute polymer solutions, hyaluronic acid ubiquitous in the body (synovial fluid, vitreous body of eye),¹²¹ human saliva,¹²² closely mono-disperse atactic polystyrene,¹²³ and for chaotic mixing.^{124,125} A pulsatile micro pump, based on a deflecting diaphragm, has also been recently designed and tested for the setup.¹²⁶

Furthermore, the stagnation point formed by other crossed micro channels *i.e.* the “T” and “Y” geometries, have also been used to trap particles,¹²⁷ break droplets,¹²⁸ study adsorption kinetics,¹²⁹ and visualize flow streamline for preferential real time blood plasma separation.¹³⁰ More recently, parallel flow channels have been used to develop stagnation points in separating flows, to achieve high deformation rates and ease stagnation point control,¹³¹ and stagnation points from 3D cross slot devices have been modeled.¹³² Theoretical and experimental investigations on these types of flow can be found in ref. 133–137. This review is focused on isolated stagnation point flows – stagnations generated away from walls – exclusively.

In general, advancements in the physical configuration of the cross-slot have mostly focused on miniaturization and geometrical modifications. While miniaturization efforts have been motivated by microfluidic applications, studies

have explored the potential of creating more stable and/or increased number of stagnation points by modifying the cross-slot's geometry and including opto-electronic systems. However, regardless of the modification implemented, the hydrodynamic implication has always been key; miniaturization facilitated microfluidic applications and geometrical changes had an implication of more complex flow structures.

3.2 Trapping and manipulation

Interest in trapping and manipulating micro-particles lies in their use for organizing complex structures out of micrometer-scale particles; or in other words, making order out of disorder. Such methods hold a great potential in developing complex aggregates, where micro-particles or cells of varying formulation, sizes, and shapes can be put together into new types of biological assemblies, possibly forming novel materials, in a bottom up bio-fabrication procedures.^{138–140} In doing so, a great deal of control can be exercised over the properties of assembled entity. The common route to achieving this is self-assembly,^{140–142} which could be driven by DNAs,^{143,144} particle wettability and shape,¹⁴⁵ Fisher's lock and key principle,^{146,147} and electric field actuation.^{148,149} Other existing means for trapping and manipulating micro-particles include optical,^{150,151} dielectrophoresis,^{152,153} optoelectronics,^{154,155} acoustic,^{156,157} and magnetic tweezers.^{158,159} The drawbacks of these methods are that their complicated control mechanisms often rely on multi-physics interactions, which could interact with the particle to obscure properties. However, using pure hydrodynamic forces



as traps, and potentially manipulators, for micro-particles has proved to be an attractive alternative for non-contact trapping and manipulation of single cells in suspensions.¹⁶⁰ Compared to other methods, manipulating micro- or nano-particles in this way can make the process simpler, and broader in steering range. These enable new capabilities like placement of specific quantum dots to desired chips with nanoscale precision.¹⁶¹

A wealth of fundamental studies have been carried out on demonstrating the operation and increasing the efficiency of purely hydrodynamic control of the stagnation generated by the cross-slot device, for the purpose of particle trapping, manipulation and assembly. In 2002, the stagnation point flow generated by a T-junction network, was used to study the deposition of bacteria (*Marinobacter hydrocarbonoclasticus*, *Psychrobacter* sp. and *Halomonas pacifica*, suspended in artificial seawater); in view of investigating the effect of deposition efficiency of different technologies.¹⁶² The study concluded that, in comparison to the parallel plate deposition technique, the stagnation point flow chamber has higher deposition rate and better reproducibility, but lower deposition efficiency. Such studies are relevant in examining the influence of marine coating properties on formation of biofilms on marine vessels,^{163,164} which is a serious problem in the shipping industry.^{165,166}

Schroeder *et al.* featured cross-slot configuration, together with T-, V-, and Y-junctions channel flows, for stagnation point based particle trapping device.¹⁶⁷ The system was described as a pressure controlled, particle confining tool using stagnation points generated in a channel flow, with a feedback control system – which used the instantaneous position of the trapped object to adjust the flow rates at each channel – in order to keep the particle confined for an indefinite length of time; without an optical trap, micropipette or other tethering device. They suggested applications of this stagnation point flow in biology, chemistry, physics, material science, and medical sciences.

Defining the microfluidic cross-slot as a “microfluidic four-roll mill”, Lee *et al.*¹¹⁴ used experiments and simulations to demonstrate how the device can be used to generate all types of flows; extensional, shear or rotational flows (see Fig. 4b). Their study suggested the application of the device in examining the dynamics of single cells and micro-drops for enhancing mixing efficiency.¹¹⁴ In separate studies, the device was also shown to be applicable in trapping and manipulating quantum dots¹⁶⁸ and multiple particles simultaneously.^{169,170} Subsequently, the confinement of particles as small as 100 nm, with up to 1 μm of resolution, for more than 10 minutes was demonstrated.^{171,172} The achieved resolution and extended trapping period was enabled by their active feedback control, which was programmed to guide the particle back to the set point position of stagnation (see Fig. 5a–c). Although many previous studies had managed to trap the particle for an extended period of time, the absence of this type of feedback control – a system originally suggested by Bentley³⁴ – meant that Brownian fluctuations would eventually cause the particle to escape. The control mechanism operated with an image acquisition device, a particle-tracking algorithm to determine the movement of the particle away from the set centroid position, and an active adjustment pneumatic valve for

the adjustment of fluid flow based on the position of the particle.¹⁷³ The application of this control system was only on the extensional direction, since the flow in the compressional direction is intrinsically stable.¹⁷⁴ The feedback control algorithm adjusted a valve constriction based on a linear equation relating the particle's hydrodynamic forces to its distance from the stagnation point.¹⁷⁵ To fully automate the system, calibration of the trap's response as a function of flow control valve characteristics and channel dimensions were also performed.¹⁷⁵ A detailed description of the experimental setup for generating this feedback-controlled stagnation can be found in ref. 173.

The very fine degree to which the microfluidic stagnation point can be controlled and manipulated, with the aid of the feedback control system, was demonstrated by Tanyeri *et al.*¹⁷⁶ As a proof-of-concept, the study used the microfluidic stagnation point to trap and manipulate 500 nm and 2.2 μm diameter particles in two-dimensions, with a positioning precision as small as 180 nm during confinement (see Fig. 5d and e).¹⁷⁶ The spring constant of the flow-based particle-micromanipulation technique was shown to scale linearly with particle size and viscosity of the medium, which offers the possibility for facile trapping of small nanoparticles in free solutions.¹⁷⁶ Characterization of the effect of strain rate on particle confinement and effective trap potential were further used to demonstrate the importance of strain rate on the trap performance and the minimum size of particle that can be trapped.¹⁷⁶

Shenoy *et al.*¹⁷⁴ investigated the performance of the cross-slot based stagnation trap using three different controllers – the proportional (P), proportional-integral (PI), and proportional-derivative (PD) controllers. Trap performance was quantified by examining the magnitude of particle fluctuation as functions of controller gain constant, system response time, and particle Peclet number. This revealed that the proportional and derivative controllers yield improvements in trap stability, while the integral controllers does not.

Advancements of the stagnation capability of the cross-slot setup to trap and manipulate multiple particles,⁹ and sequentially assemble particles,¹⁷⁷ have also been reported. The device for multiple particle trapping constitutes a systematic charging and discharging of fluid from a six channel cross-slot, in such a way that two stagnation points are generated (see Fig. 5f and g).⁹ Typically, the controllers used are a combination of proportional-integral-differential (PID), but this study suggests that the “integral” component does not necessarily improve trap efficiency. Development of the multiple trapping capability was achieved using the model predictive control (MPC) algorithm for a more robust control strategy, with 5–7 times increase in trap stiffness compared to previous generation traps. Using these, simultaneous confinement and manipulation of two particles to switch center of mass positions and draw shapes were achieved. Fig. 5h and i shows transient photographs of devices operation in drawing the letter I with both particles trajectory paths. Similarly, a sequential assembly algorithm was demonstrated to be applicable in sequentially joining particles, trapped in a seven-slot channel generated microfluidic stagnation point, to arrange particles in the form



Haward *et al.*¹⁸⁶ later performed cross-slot based experiments on similar solutions used by Pathak and Hudson¹⁸⁹ but used a smaller cross-slot device and presented more detailed experimental results; obtained over a much broader range of Wi . Their study also utilized a deep cross-section cross-slot, so as to obviate the effect of shear along the channel walls. Both transitions – steady asymmetric and transient unstable – were also observed in this study, but the instabilities were suggested to be influenced by the micellar fluid shear localization within the micro channels and around the re-entrant corners of the cross-slot. This claim was substantiated by the formation of lip vortices along the walls of the inlet channels, reported by Dubash and coworkers.¹⁸⁵ In a later study, Haward and McKinley¹⁸⁴ extended the rheological properties of the examined micellar fluid – by varying the zero-shear viscosity over three orders of magnitude – and investigated the resulting changes in dynamical response. The extensional properties and critical conditions for the start of flow instabilities in the fluids were shown to be reliant on the fluid formulation. By plotting Wi as a function of Re for all considered solutions, their report presented a stability diagram outlining the steady symmetric, steady asymmetric and inertia-elastic flow regimes (see Fig. 6e), which covered a wider range of Wi and Re than that presented in their previous study.¹⁸³

Arratia *et al.*¹⁸⁰ findings also motivated full sized numerical models over a wide range of flow parameters and qualitatively revealed similar transitions^{190–192} using the modeled full cross-slot geometry. It was also showed numerically that introducing a small degree of curvature to the cross-slot corners has no influence upon the transitions.¹⁸²

Although a general concession has not been reached on the origin of these transitions, based on findings mostly facilitated by the microfluidic stagnation point, it is widely accepted that they are complex phenomena, which are influenced by a combination of factors such as Wi , inertial effects, and rheological behavior of the micellar solutions.¹⁸⁵ A more detailed explanation was offered by Haward and McKinley who attributed the phenomena to extensional stresses overcoming entropic elasticity as the velocity gradient at the stagnation point exceeds the reciprocal of the characteristic relaxation time; such that Wi exceeds unity.¹⁸⁴ This can result in a significant extension and alignment of any deformable microstructural constituents of the fluid. Such stretching and orientation effects can result in significant increases in the fluid extensional viscosity and, when inertia is not significant, can give rise to purely elastic instabilities.¹⁸⁴

3.4 Particle dynamics studies

Following the pioneering work of Scriven *et al.* in 1979, the cross-slot generated extensional flow was used to demonstrate trapping and flow-induced fracture of mono-disperse atactic polystyrene (a-PS) chains in a controlled manner.^{107,193} One of the main highlights of the study is the accuracy at which these chains break at their center. This work was extended to explore the effect of chemical composition, chain length and force magnitude on the fracture mechanisms of the chains,¹⁹⁴ and the

use of the stretched macromolecule to examine molecular weight distribution, dimensions and draining characteristics chains.¹⁰⁵ During this period (1970–1990), many extensional flow based particle dynamics studies were carried out and a review of some of their experimental results can be found in the literature.¹⁹⁵ Simplified models of polymer strands in a cross flow were also used to predict birefringence behavior that portrayed good agreement with experiments.^{196,197}

Experimental use of the cross-slot generated stagnation point flow for particle dynamics studies resurfaced in the 1990's when it was used in studying single polymer dynamics and stretching.^{106,198} Perkins *et al.*¹⁰⁶ used this setup to decouple the effects of strain rate and time on the conformation and extension of single polymer molecules. Amongst other heterogeneity in polymer dynamics, they observed that polymer chains with a dumbbell shape stretched more rapidly than folded ones. This led to deductions that the shapes of stretched macromolecules (*e.g.* dumbbell, folded, coiled, kinked) is highly dependent on the initial conformation (see Fig. 7), which has been corroborated by other studies.^{199,200} In Perkins's study, the polymers were introduced through the cross-slots, and hence, were exposed to velocity gradients prior to entrapment at the stagnation point.¹⁰⁶ This methodology led to concerns relating to the thermal equilibrium state of the polymer before the inception of the elongation flow, which might have caused the observed heterogeneity. Smith *et al.*¹⁹⁸ circumvented this by initiating the cross-slot based stagnation point flow with the polymer coils at rest in the observation region and by using a higher viscosity solvent. Although heterogeneities in polymer dynamics were still observed, their investigation led to a quantitative understanding of how the initial state of the polymer can lead to elongated appearances of different conformations, and these quantitatively agreed with Brownian dynamics simulations.²⁰¹

By adapting the feedback control loop, the cross-slot based flow was used to decouple the effect of initial conformation, length, strain rate and Deborah number, on the extension dynamics of *Escherichia coli* (*E. coli*) DNA polymer chains.²⁰² Results showed that the extended conformation of the polymer depends on the conformation dependent hydrodynamic forces and deformation. The control system provided long-term stability of the polymer's physical properties, which afforded long observation duration of the molecules.

Following these groundbreaking investigations, the cross-slot generated flow was also adapted for techniques in investigating cell deformability,^{204,205} polymersome dynamics,²⁰⁶ break up of CNT bundles,²⁰⁷ stretch-coil instabilities,²⁰⁸ solution dynamics,^{110,118,209} extensional viscosity,¹²⁰ DNA stretching/compaction,^{210,211} and polymer rheology.^{123,212–216} Using this device, cell deformability measurements were achieved by harnessing cell concentrations by viscoelastic micro flows of polymer solution,²⁰⁴ CNT breakup was attained by simply increasing flow rates,²⁰⁷ drop deformation was examined as a function of confinement,²⁰⁹ and DNA compaction was triggered by introducing Na^+ and polyethylene glycol (PEG) to the DNA molecules held at the stagnation point.²¹⁰



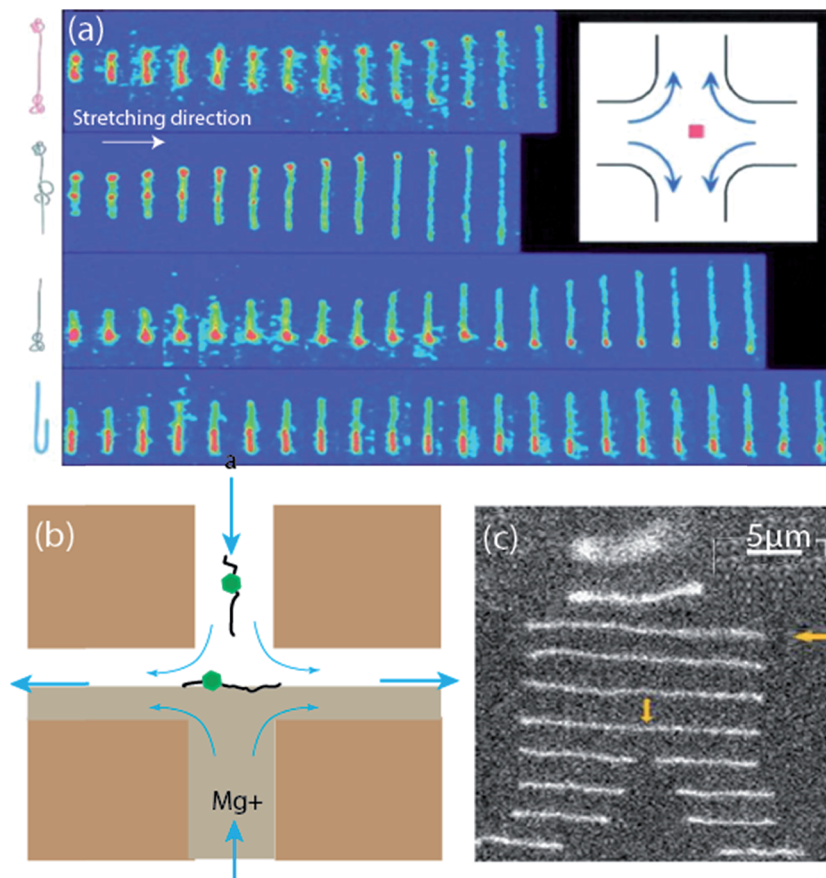


Fig. 7 Cross-slot stagnation point flow as a DNA trapping stretching device. (a) DNA elongating in the stagnation point flow showing the effects of initial polymer conformation on the time-dependent extension of single DNA molecules. Top to bottom: dumbbell, kinked, half-dumbbell, and folded conformations (sketches of molecular configurations are included on the left). Time between images is 0.13 seconds. Inset: cross-slot device and the position of the stagnation point flow.¹⁰⁶ Reprinted with permission. Copyright© 1997, The American Association for the Advancement of Science. (b) Schematic for single DNA sequence detection with stagnation point flow. Flow direction denoted by blue arrows. DNA–enzyme complexes flow via the top slot, while Mg^{2+} solution flows via the bottom slot. DNA–enzyme complex trapped at the stagnation point and elongated along the extensional axis.²⁰³ (c) Photographs of molecular configurations with non-uniform time intervals (smallest time interval, at cleavage, is 100 ms) to show the trapping, stretching and subsequent cleavage of dsDNA.²⁰³ Reprinted with permission. Copyright© 2010, Royal Society of Chemistry.

Of the aforementioned applications, one particularly interesting use of the cross-slot generated stagnation is for the precise control of droplet coalescence,²¹⁷ while generating multiple emulsions; a concept that holds a lot of promise in pharmaceuticals.²¹⁸ Wang *et al.* suggested that the simple cross-slot generated flow field is inadequate to accurately control the coalescence of multiple droplets; hence they adapted the geometry of Fig. 4c(iv) and developed numerical methodologies to investigate the rheology of multiple emulsions.^{217,219,220} In their work, they numerically demonstrated the use of cross-slot flow control techniques to precisely engineer the adhesion of single drops to form bigger drops – even to the point of dictating the angle of contact.²¹⁷ By adjusting the extensional and compressional flow towards the stagnation point, they were able to achieve location specific consecutive coalescence of multiple droplets.

The extensional flow of the oscillating cross-slot device has also been recently used to quantitatively assess cell mechanical damage in an extensional field, which is relevant in designing

practical bioreactors.²²¹ The cells were deformed at the stagnation point of the cross-slot that mimics the extensional flow produced during the bubble bursting process. This process consists of film receding, film impact and upward/downward jet formations—cell-damaging mechanics inherent in conventional agitation based bioreactors used to homogenize culturing medium.^{222–224} Although, the bioreactors flow includes shear and extensional flow components,²²⁵ the extensional component has been demonstrated to have a greater effect on cell.^{224,226} Their experiments suggest that the critical extensional stress for Chinese hamster ovary (CHO) cells is 250 Pa, which is less than the numerically estimated stress induced by bubble bursting (632.8 Pa).²²¹ This quantitatively indicates that extensional stresses in bioreactors can be critical to mechanical cell damage.

3.5 Target sequence detection

One of the earliest uses of the microfluidic cross-slot generated stagnation point flow in biomedicine was directed toward



a single-molecule genotyping assay; for detecting DNA target sequences and marker position.^{227,228} To achieve this, tagged DNA molecules were trapped at the stagnation point, elongated by the extensional flow, and directly observed with a microscope. Previous methods of doing this involve immobilizing the DNAs on surfaces, and stretching them using surface chemistries favoring the adsorption of DNA as a fluid meniscus recedes.²²⁹ However, the stagnation point flow approach potentially offers a purely hydrodynamic technique for continuous on-chip genotype assay. This potential was demonstrated by using the cross-slot to view sequences in images of trapped DNA-beads hybrids and peaked intensities signifying the position of the beads.²²⁷ Comparing the accuracy of the microfluidic technique to that of stretching on slides shows that although the slide stretching is more accurate in detecting target sequences, the stagnation point flow technique results in a 50% reduction in the detection's standard deviation.

In view of improving detection accuracy, Xu and Muller extended their earlier study by introducing enzyme pre-bound in the investigated dsDNA and then introducing Mg^{2+} required for dsDNA cleavage.²⁰³ Upon linearizing the DNA-enzyme complex at the stagnation point, cofactor Mg^{2+} is introduced and the binding location of the enzyme are determined by optical restriction mapping without surface immobilization of the DNAs (see Fig. 7b and c). The accuracy of this technique in sequence detection was found to be comparable to other single molecule techniques. In addition, the technique offers a means of simultaneously obtaining kinetic information for all DNA recognition sites in the same process.

While previous applications of the stagnation point have focused on trapping and translating the particle as a whole, particle stretching presents another functional feature of the microfluidic stagnation point flow concept. The capability of isolating the induced strain rates to that applied by the flow field – avoiding the contribution of wall shearing effects – allows for a fine control of the stretching rates. This again highlights the appeal of microfluidic stagnation point flows in this particular biological field and hence, its prospects for application in commercial integrated on-chip devices.

4. Open space microfluidics

Microfluidic systems that operate in the “open space” – without the need for the sealed channels and chambers commonly used in microfluidics – is an emerging class of fluid localization technique that isolates chemical reactions on biological samples without imposing significant “constraints”, like encapsulation, pre-processing steps, or the need for scaffolds.⁴⁴ This is achieved by eliminating walls and channels in microfluidic devices. Stagnation points have also been produced using these open microfluidic setups, and these are covered in the following section.

4.1 Opposed jets

Opposed jets, as the name implies, is a configuration of two oppositely directed fluid streams in an immersed surrounding

solution. Depending on the intended application, the jets could be made to oppose *via* impinging, or *via* suction flows to produce either compressional or extensional flows, respectively.^{230,231} The concept was originally conceived as a premixed flame stabilization technique,^{232–234} due to the uniform strain rate inherent in the flow produced by the counter jets^{235,236} – which is normally not the case for a regular flame surface.^{237,238} According to Schaffer and Cambel,²³² the stability of the flame so maintained, is ascribed to the existence of a small reaction zone around the stagnation point of the jet. The opposed jet setup was also adapted as an experimental technique for investigating the stability of diffusion flames, where the gaseous fuel and oxidant coaxial jets are opposed to produce a flame at their junction (stagnation plane).^{239–242} For a more detailed description of the early adaptations of opposed jets, one of the earliest review of the topic can be found in ref. 243. In this line of research, studies adapted the apparatus in studying non-equilibrium anomalies,²⁴⁴ chemical kinetics,^{245,246} numerical modeling approaches,²⁴⁷ and flame structures.²⁴⁸ Existence of a reaction zone at the stagnation point also serves as the stabilizing mechanism in diffusion flames,²⁴⁰ with the flame tilting to the oxidant side of the stagnation plane.^{240,249}

The flow generated by opposed jets have also been adapted in studying purely homogeneous kinetics of endothermic reactions,²⁵⁰ particularly in metal-organic chemical vapor deposition (MOCVD) of compound semiconductors.^{251,252} During the conventional MOCVD of semiconductors, growth of a thin solid film is achieved by passing a gas mixture over a heated substrate. This involves both homogeneous reactions in the gas phase near the heated substrate and surface reactions on the substrate.^{250,253} The use of opposed-jets based reactors for such studies stems from the need to decouple the effects of surface and gas-phase kinetics during MOCVD. In the opposed-jet setup, reactions can be confined near the stagnation point and away from hot surfaces by controlling the jet velocities.²⁵⁰ Furthermore, by adjusting flow rates, residence time of species and the location of the reaction zone can be adjusted, which allows for the detection of the onset of homogeneous thermal decomposition.²⁵⁰

Although the stagnation plane formed by opposed jets were originally only significant as the reference plane for rapid and effective mixing in fluid reactors,²⁵⁴ these flows have also been adapted for particle dynamics studies. Frank and co-workers adapted this apparatus for producing compressional and extensional flows (see Fig. 8a) for studying polythene birefringence in xylene solution, as a function of temperature.²⁵⁵ This concept was later highlighted as an extensional flow apparatus,^{105,256} for performing rheological changes,²⁵⁷ flow induced molecular scission,²⁵⁸ and coil-stretch transition.^{259,260} Subsequently, the Rheometrics RFX laboratory apparatus was produced using opposed nozzles through which liquid is sucked or ejected from a reservoir, as an elongation flow analyzer.²⁶¹ The device was used to make extensional viscosity measurements as a function of strain rate²⁶² and polymer solution.²⁶³

Advantages of the opposed-jet setup (see Fig. 8a) include, the ability to generate relatively large velocity gradients,²⁶⁴ and its



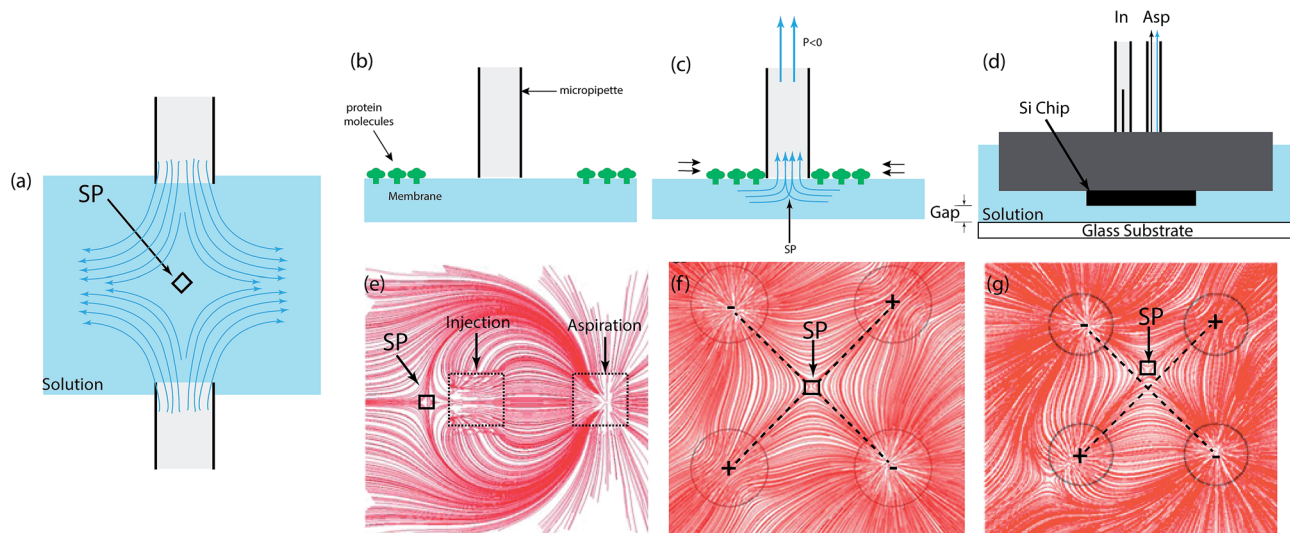


Fig. 8 Microfluidic stagnation point flows in open microfluidics. (a) Schematic drawing of an opposed-jets apparatus in “push mode,” which generates a uniaxial compressional flow along the stagnation point axis. The jets can also be inverted into “sucking mode” in order to generate a biaxial extensional flow. (b) Pipette placed above the membrane.²⁶⁷ (c) Negative hydrostatic pressure results in fluid flow into the pipette. The subsequent fluid flow causes a stagnation point to form beneath the pipette and molecules to gather and trapped at the tip of the pipette.²⁶⁷ (d) Schematic representation of the two-aperture MFP. The MFP is set close to the glass substrate and the fluid is injected and fully aspirated from the capillary tubes, which makes the injected flow hydro-dynamically confined. (e) Numerically calculated streamlines of the two-aperture MFP flow showing the confinement area and stagnation point (SP). The stagnation position (R) is defined as the distance between the center of the MFP and the stagnation point.²⁷⁵ Reprinted with permission. Copyright© 2015, Rights Managed by Nature Publishing Group. (f) Numerical deduced streamlines of the microfluidic quadrupole. The stagnation point is at the center of the flow when the both flow rates ratio (aspiration flow/injection flow) are identical. (g) Streamline of the microfluidic quadrupole demonstrating manipulation of the stagnation point's position. Varying flow rate ratio moves the stagnation point from the center in the XY plane.

applicability for a wide range of concentrations and fluids.^{264,265} However, these are overshadowed by the difficulty of obtaining high-resolution images of the flow field in the setup, and its requirement for relatively large fluid volumes.⁹⁷ Furthermore, the device's measurement of extensional viscosity were labeled as “3 times greater than the real value” due to the effect of dynamic pressure, nozzle shear and liquid inertia.²⁶⁶

4.2 Microfluidic pipettes

Microfluidic pipettes are a unique technology that use hydrodynamically confined flows to carry out a variety of complex functions like mixing, multiplexing, or gradient generation. These devices are capable of handling and dispensing solutions in cellular and sub-cellular levels, and some of their advantageous features include easy positioning, and compatibility with standard cell cultures and laboratory methods.

The use of these devices for microfluidic stagnation point flows is relatively sparse in the literature, but there are exemplar studies that highlight the simplicity of the setup that can be used for this application. A conical microfluidic pipette – in the form of a single conical tube – was demonstrated to be applicable as a trap for 5 nm large protein molecules (Streptavidin) bound to biotin receptors on a supported lipid bilayer, based on the stagnation point flow formed below the pipette's tip.²⁶⁷ In that study, this was achieved by positioning the pipette, of about 1 micrometer tip radius, above the lipid bilayer and applying a negative pressure difference to cause a net flow of fluid through the pipette and an accumulation of protein molecules

around the tip of the pipette (see Fig. 8b and c). The method was presented as a means of experimentally obtaining protein dynamics information and quantifying the intermolecular forces between molecules bound to a membrane; as long as the membrane can be approached by a pipette.²⁶⁷

Ainla *et al.* also demonstrated the use of a similar principle to deposit vesicles adhered to a film, at the stagnation point of a three-aperture on-chip multifunctional micropipette.^{268,269} In this configuration, injection is through the middle aperture while aspiration through the two outer apertures. This causes the vesicles to be trapped beneath the aspiration aperture, and upon switching the middle aperture to high aspiration, the vesicles are drawn into the channel opening. Using this multifunctional pipette, a variety of complex microfluidic processing operations, such as mixing, multiplexing, writing, erasing, functionalizing on a substrate, or gradient generation at the single cell level, can be achieved at high resolution.^{270,271} Its use in multiple-compound delivery for pharmacological screening of intact adherent cells was demonstrated by applications in activating hTRPV1 receptors in single cells, and exposing specific cells to formaldehyde/dithiothreitol-containing solutions.²⁷⁰ It was also used to electroporate single-cells,²⁷² and administer pharmacological active substances to selected areas on brain slices.^{273,274}

4.3 Microfluidic probes

The microfluidic probe (MFP) is a multi-aperture, mobile and channel-less microfluidic system, which operates by



can refer to the literature for precise setups and methodologies for the individual techniques discussed.^{10,176,267,284} In terms of applications, the use of stagnation points for performing droplet characterization studies, trapping and manipulation of cells and polymers, and investigating flow instabilities were discussed. The devices that have been used for generating these flows all lend principles from the pioneering four-roll mill.¹⁰ Early applications of the four-roll mill were inclined towards investigations on the primary mechanisms governing droplet dynamics with an aim of engineering emulsions and polymers. Today, the cross-slot deduced stagnation point flows represent the most advanced applications of microfluidic stagnations; from trapping micro-objects, to studying flow instabilities and stretching DNAs. Table 1 summarizes the features and applications of each microfluidic stagnation point flow device.

The single most important feature of the microfluidic stagnation point flow lay in its ability to incubate the analysis in an environment in interaction with only one external field – hydrodynamics of the surrounding flow. As such, by manipulating this hydrodynamic field, precise stimulus can be applied on the sample with assurance that the observed effects can be directly correlated with the applied stimuli. In the early days of this concept, the naturally instable nature of the stagnation point presented a frustration that was immediately fixed by the introduction of a feedback control system. This development unleashed the full capability of the concept, and starting from its classic macroscopic forms, it has been a consequential tool for optimizing the parameters for synthesizing emulsions, polymer blending and birefringence. However, the bulky nature and dynamic-parts of the four roll mill presented an obvious drawback towards miniaturization. The cross-slot device, often termed the microfluidic four-roll mill was developed to avert this problem. Its simple design and straightforward fabrication technique propelled several applications that later led to the successful trapping and manipulation of nanoparticles, with a very fine degree of precision. This led to one of its first biology applications in trapping and stretching DNA for detecting target sequences. However, the cross-slot is associated with its own drawbacks – akin to most of classical “closed-channel” microfluidic devices – which is its reliance on channel flows with walls/boundaries that restricts its maneuverability and might induce shear stresses on the analysis region. Although still fledgling, the advent of open space microfluidics (*i.e.* opposed jets, micropipettes, & MFPs) and the MQ mitigated these issue, and presented immense promise for the future.

The development of microfluidic stagnation point has undoubtedly come a long way. The concept has shown some promise for single cell analysis – trapping of 100 nm particles^{171,172} with very minimal wall shear stress at the stagnation point have been demonstrated.²⁸⁴ Yet, biological applications are still very limited. This sparse application in biological research can be attributed to the relatively unstable nature of its stagnation point that require complicated feedback control mechanisms, and the potential cell damage that can occur when high flow rates in channel based microfluidic chips induce significant wall shear stresses at vicinity of the stagnation point.

The MFP and micropipettes recently entered the scene of generating microfluidic stagnation points however, explicit practical applications are lacking. Majority of studies stopped at demonstrating the existence of microfluidic stagnation points,²⁸⁴ while others did not even give the generated stagnation point any considerations.^{271,282} One might now expect the adjunct open space microfluidic and MQ concepts would lead the way for microfluidic stagnation points to become fully integrated as lab-on-a-tip devices for μ TAS and new innovative biomedical applications. But to achieve this, future works must learn from the developmental phase of the concept – the absence of external fields, simple designs, and precise stagnation control are key features that cannot be compromised. While the open space microfluidic devices incubates the sample from external fields and have relatively simple designs, the feedback control mechanism adapted by previous devices might have to be significantly simplified before it can be integrated into a single miniaturized analytical tool.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

Ayoola T. Brimmo acknowledges NYUAD Global PhD Fellowship, and the authors acknowledge financial support from New York University Abu Dhabi (NYUAD).

References

- 1 R. W. Fox, A. T. McDonald and P. J. Pritchard, *Introduction to Fluid Mechanics*, John Wiley & Sons, New York, 1985, vol. 7.
- 2 G. A. Tokaty, *A History and Philosophy of Fluid Mechanics*, Courier Corporation, Mineola, New York, 1971.
- 3 C. M. Ho and S. H. Chen, Unsteady Kutta Condition of a Plunging Airfoil, in *Unsteady Turbulent Shear Flows*, Springer, Berlin, 1981.
- 4 J. D. Anderson, Ludwig Prandtl's Boundary Layer, *Phys. Today*, 2005, **58**(12), 42–48.
- 5 H. Schlichting and K. Gersten, *Boundary-Layer Theory*, Springer-Verlag, Berlin Heidelberg, 2003, vol. 8.
- 6 S. F. Hoerner, *Fluid-Dynamic Drag: Practical Information on Aerodynamic Drag and Hydrodynamic Resistance*, Hoerner Fluid Dynamics, Midland Park, NJ, US, 1965.
- 7 M. J. Lighthill, Contributions to the Theory of Heat Transfer through a Laminar Boundary Layer, *Proc. R. Soc. London, Ser. A*, 1950, **202**(1070), 359–377, DOI: 10.1098/rspa.1950.0106.
- 8 J. A. Fay and F. R. Riddell, Theory of Stagnation Point Heat Transfer in Dissociated Air, *J. Aeronaut. Sci.*, 1958, **25**(2), 73–85.
- 9 A. Shenoy, C. V. Rao and C. M. Schroeder, Stokes Trap for Multiplexed Particle Manipulation and Assembly Using Fluidics, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, 201525162.



- 10 G. I. Taylor, The Formation of Emulsions in Definable Fields of Flow, *Proc. R. Soc. London, Ser. A*, 1934, **146**(858), 0501–0523, DOI: 10.1098/rspa.1934.0169.
- 11 G. M. Whitesides, The Origins and the Future of Microfluidics, *Nature*, 2006, **442**(7101), 368–373, DOI: 10.1038/nature05058.
- 12 T. M. Squires and S. R. Quake, Microfluidics: Fluid Physics at the Nanoliter Scale, *Rev. Mod. Phys.*, 2005, **77**(3), 977–1026, DOI: 10.1103/RevModPhys.77.977.
- 13 C. J. Pipe and G. H. McKinley, Microfluidic Rheometry, *Mech. Res. Commun.*, 2009, **36**(1), 110–120, DOI: 10.1016/j.mechrescom.2008.08.009.
- 14 D. J. Beebe, G. A. Mensing and G. M. Walker, Physics and Applications of Microfluidics in Biology, *Annu. Rev. Biomed. Eng.*, 2002, **4**(1), 261–286, DOI: 10.1146/annurev.bioeng.4.112601.125916.
- 15 H. A. Stone, A. D. Stroock and A. Ajdari, Engineering Flows in Small Devices: Microfluidics toward a Lab-on-a-Chip, *Annu. Rev. Fluid Mech.*, 2004, **36**, 381–411, DOI: 10.1146/annurev.fluid.36.050802.122124.
- 16 P. Gravesen, J. Branebjerg and O. S. Jensen, Microfluidics—a Review, *J. Micromech. Microeng.*, 1993, **3**(4), 168.
- 17 H. Andersson and A. VanDenBerg, Microfluidic Devices for Cellomics: A Review, *Sens. Actuators, B*, 2003, **92**(3), 315–325.
- 18 Y. Zhang and P. Ozdemir, Microfluidic DNA Amplification—a Review, *Anal. Chim. Acta*, 2009, **638**(2), 115–125.
- 19 P. Abgrall and A. M. Gue, Lab-on-Chip Technologies: Making a Microfluidic Network and Coupling It into a Complete Microsystem—a Review, *J. Micromech. Microeng.*, 2007, **17**(5), R15.
- 20 L. Gervais, N. De Rooij and E. Delamarche, Microfluidic Chips for Point-of-Care Immunodiagnosics, *Adv. Mater.*, 2011, **23**(24), H151–H176.
- 21 S. Saleh-Lakha and J. T. Trevors, Perspective: Microfluidic Applications in Microbiology, *J. Microbiol. Methods*, 2010, **82**(1), 108–111, DOI: 10.1016/j.mimet.2010.03.022.
- 22 S. Haeberle and R. Zengerle, Microfluidic Platforms for Lab-on-a-Chip Applications, *Lab Chip*, 2007, **7**(09), 1094–1110.
- 23 A. Mukherjee and C. M. Schroeder, Microfluidic Methods for Molecular Biology, in *Microfluidic Methods in Single Cell Biology*, Switzerland, 2016, pp. 19–54.
- 24 J. Nilsson, M. Evander, B. Hammarstrom and T. Laurell, Review of Cell and Particle Trapping in Microfluidic Systems, *Anal. Chim. Acta*, 2009, **649**(2), 141–157, DOI: 10.1016/j.aca.2009.07.017.
- 25 R. R. Lagnado, N. Phanthien and L. G. Leal, The Stability of Two-Dimensional Linear Flows, *Phys. Fluids*, 1984, **27**(5), 1094–1101, DOI: 10.1063/1.864755.
- 26 R. R. Lagnado and L. G. Leal, Visualization of Three-Dimensional Flow in a Four-Roll Mill, *Exp. Fluids*, 1990, **9**, 25–32.
- 27 H. A. Barnes and J. M. Maia, Rheometry, in *Rheology, Encyclopedia of Life Support Systems (EOLSS)*, 2010, pp. 331–362.
- 28 C. W. Macosko, M. A. Ocansey and H. H. Winter, Steady Planar Extensions with Lubricated Dies, *J. Non-Newtonian Fluid Mech.*, 1982, **11**, 301–316.
- 29 J. M. Dealy, Extensional Flow of Non-Newtonian Fluids—a Review, *Polym. Eng. Sci.*, 1961, **11**(6), 433–445.
- 30 G. G. Fuller and L. G. Leal, Flow Birefringence of Dilute Polymer Solutions in Two-Dimensional Flows, *Rheol. Acta*, 1980, **19**, 580–600.
- 31 G. G. Fuller and L. G. Leal, Flow Birefringence of Concentrated Polymer Solutions in Two-Dimensional Flows, *J. Polym. Sci., Polym. Phys. Ed.*, 1981, **19**, 557–587.
- 32 F. D. Rumscheidt and S. G. Mason, Suspensions Xii. Deformation and Burst of Fluid Drops in Shear and Hyperbolic Flow, *J. Colloid Sci.*, 1961, **16**(3), 238–261.
- 33 H. P. Grace, Dispersion Phenomena in High Viscosity Immiscible Fluid Systems and Application of Static Mixers as Dispersion in Such Systems, *Chem. Eng. Commun.*, 1982, **14**, 225–277.
- 34 B. J. Bentley and L. G. Leal, A Computer-Controlled Four-Roll Mill for Investigations of Particle and Drop Dynamics in Two-Dimensional Linear Shear Flows, *J. Fluid Mech.*, 1986, **167**, 219–240.
- 35 B. J. Bentley and L. G. Leal, An Experimental Investigation of Drop Deformation and Breakup in Steady, Two-Dimensional Linear Flows, *J. Fluid Mech.*, 1986, **167**, 241–283.
- 36 D. C. Tretheway, M. Muraoka and L. G. Leal, Experimental Trajectories of Two Drops in Planar Extensional Flow, *Phys. Fluids*, 1999, **11**(5), 971–981, DOI: 10.1063/1.869969.
- 37 H. Yang, C. C. Park, Y. T. Hu and L. G. Leal, The Coalescence of Two Equal-Sized Drops in a Two-Dimensional Linear Flow, *Phys. Fluids*, 2001, **13**(5), 1087–1106, DOI: 10.1063/1.1358873.
- 38 J. W. Ha, Y. Yoon and L. G. Leal, The Effect of Compatibilizer on the Coalescence of Two Drops in Flow, *Phys. Fluids*, 2003, **15**(4), 849–867, DOI: 10.1063/1.1555803.
- 39 M. Borrell, Y. Yoon and L. G. Leal, Experimental Analysis of the Coalescence Process Via Head-on Collisions in a Time Dependent Flow, *Phys. Fluids*, 2004, **16**(11), 3945–3953.
- 40 D. G. Crowley, F. C. Frank, M. R. Mackley and R. G. Stephenson, Localized Flow Birefringence of Polyethylene Oxide Solutions in a Four-Roll Mill, *J. Polym. Sci., Polym. Phys. Ed.*, 1976, **14**, 1111–1119.
- 41 J. Feng and L. G. Leal, Transient Extension and Relaxation of a Dilute Polymer Solution in a Four-Roll Mill, *J. Non-Newtonian Fluid Mech.*, 2000, **90**(1), 117–123, DOI: 10.1016/S0377-0257(99)00052-X.
- 42 J. Deschamps, V. Kantsler, E. Segre and V. Steinberg, Dynamics of a Vesicle in General Flow, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**(28), 11444–11447, DOI: 10.1073/pnas.0902657106.
- 43 M. Levant, J. Deschamps, E. Afik and V. Steinberg, Characteristic Spatial Scale of Vesicle Pair Interactions in a Plane Linear Flow, *Phys. Rev. E: Stat., Nonlinear, Soft Matter Phys.*, 2012, **85**(5 Pt 2), 056306, DOI: 10.1103/PhysRevE.85.056306.



- 44 V. Narayanamurthy, S. Nagarajan, F. Samsuri and T. M. Sridhar, Microfluidic Hydrodynamic Trapping for Single Cell Analysis: Mechanisms, Methods and Applications., *Anal. Methods*, 2017, **9**, 3751.
- 45 G. V. Kaigala, R. D. Lovchik and E. Delamarche, Microfluidics in the "Open Space" for Performing Localized Chemistry on Biological Interfaces, *Angew. Chem., Int. Ed.*, 2012, **51**(45).
- 46 E. N. Horsford, *Report on Vienna Bread*, Government Printing Office, Washington, 1875.
- 47 L. Rayleigh, On the Instability of Jets, *Proc. Lond. Math. Soc.*, 1878, **1**(1), 4–13.
- 48 J. W. Strutt and L. Rayleigh, On the Instability of Jets, *Proc. Lond. Math. Soc.*, 1878, **10**, 4.
- 49 S. Tomotika, On the Instability of a Cylindrical Thread of a Viscous Liquid Surrounded by Another Viscous Fluid, *Proc. R. Soc. London, Ser. A*, 1935, **150**(8790), 322–337.
- 50 J. O. Hinze, Fundamentals of the Hydrodynamic Mechanism of Splitting in Dispersion Processes, *AIChE J.*, 1955, **1**(3), 289–295, DOI: 10.1002/aic.690010303.
- 51 J. W. Ha and L. G. Leal, An Experimental Study of Drop Deformation and Breakup in Extensional Flow at High Capillary Number, *Phys. Fluids*, 2001, **13**(6), 1568–1576, DOI: 10.1063/1.1358306.
- 52 J. M. Rallison, The Deformation of Small Viscous Drops and Bubbles in Shear Flows, *Annu. Rev. Fluid Mech.*, 1984, **16**(1), 45–66.
- 53 J. M. Rallison, Note on the Time-Dependent Deformation of a Viscous Drop Which Is Almost Spherical, *J. Fluid Mech.*, 1980, **98**, 625–633, DOI: 10.1017/S0022112080000316.
- 54 H. A. Stone, Dynamics of Drop Deformation and Breakup in Viscous Fluids, *Annu. Rev. Fluid Mech.*, 1994, **26**(1), 65–102, DOI: 10.1146/annurev.fluid.26.1.65.
- 55 S. Torza, R. G. Cox and S. G. Mason, Particle Motions in Sheared Suspensions Xxvii. Transient and Steady Deformation and Burst of Liquid Drops, *J. Colloid Interface Sci.*, 1972, **38**(2), 395–411.
- 56 M. V. Berry and M. R. Mackley, The Six Roll Mill: Unfolding an Unstable Persistently Extensional Flow, *Philos. Trans. R. Soc., A*, 1977, **287**(1337), 1–16.
- 57 D. L. Barthes-Biese and A. Acrivos, Deformation and Burst of a Liquid Droplet Freely Suspended in a Linear Shear Field, *J. Fluid Mech.*, 1973, **61**, 1–21.
- 58 E. J. Hinch and A. Acrivos, Steady Long Slender Droplets in Two-Dimensional Straining Motion, *J. Fluid Mech.*, 1979, **91**, 401–414.
- 59 J. M. Rallison, A Numerical Study of the Deformation and Burst of a Viscous Drop in General Shear Flows, *J. Fluid Mech.*, 1981, **109**, 465–482.
- 60 H. A. Stone and L. G. Leal, The Influence of Initial Deformation on Drop Breakup in Subcritical Time-Dependent Flows at Low Reynolds Numbers., *J. Fluid Mech.*, 1989, **206**, 223–263.
- 61 Y. Pawar and K. J. Stebe, Marangoni Effects on Drop Deformation in an Extensional Flow: The Role of Surfactant Physical Chemistry. I. Insoluble Surfactants, *Phys. Fluids*, 1996, **8**(7), 1738–1751.
- 62 Y. T. Hu, D. J. Pine and L. G. Leal, Drop Deformation, Breakup, and Coalescence with Compatibilizer, *Phys. Fluids*, 2000, **12**(3), 484–489, DOI: 10.1063/1.870254.
- 63 Y. T. Hua and A. Lips, Determination of Viscosity from Drop Deformation, *J. Rheol.*, 2001, **45**(6), 1453–1463.
- 64 Y. T. Hu and A. Lips, Transient and Steady State Three-Dimensional Drop Shapes and Dimensions under Planar Extensional Flow, *J. Rheol.*, 2003, **47**(2), 349–369, DOI: 10.1122/1.1545078.
- 65 W. J. Milliken and L. G. Leal, Deformation and Breakup of Viscoelastic Drops in Planar Extensional Flows, *J. Non-Newtonian Fluid Mech.*, 1991, **40**(3), 355–379, DOI: 10.1016/0377-0257(91)87018-S.
- 66 C. E. Chaffey, A Second-Order Theory for Shear Deformation of Drops, *J. Colloid Interface Sci.*, 1967, **24**(2), 258–269.
- 67 R. G. Cox, The Deformation of a Drop in a General Time-Dependent Fluid Flow, *J. Fluid Mech.*, 1969, **37**(3), 601–623.
- 68 N. A. Frankel and A. Acrivos, The Constitutive Equation for a Dilute Emulsion, *J. Fluid Mech.*, 1970, **44**(01), 65–78.
- 69 S. Guido and F. Greco, Drop Shape under Slow Steady Shear Flow and During Relaxation. Experimental Results and Comparison with Theory, *Rheol. Acta*, 2001, **40**(2), 176–184, DOI: 10.1007/s003970000144.
- 70 J. J. Higdon, The Kinematics of the Four-Roll Mill, *Phys. Fluids A*, 1993, **5**(1), 274–276.
- 71 P. L. Maffettone and M. Minale, Equation of Change for Ellipsoidal Drops in Viscous Flow, *J. Non-Newtonian Fluid Mech.*, 1998, **78**(2–3), 227–241, DOI: 10.1016/S0377-0257(98)00065-2.
- 72 E. D. Wetzel and C. L. Tucker, Droplet Deformation in Dispersions with Unequal Viscosities and Zero Interfacial Tension, *J. Fluid Mech.*, 2001, **426**, 199–228, DOI: 10.1017/S0022112000002275.
- 73 Y. Y. Wu, A. Z. Zinchenko and R. H. Davis, Ellipsoidal Model for Deformable Drops and Application to Non-Newtonian Emulsion Flow, *J. Non-Newtonian Fluid Mech.*, 2002, **102**(2), 281–298, DOI: 10.1016/S0377-0257(01)00183-5.
- 74 U. Sundararaj and C. W. Macosko, Drop Breakup and Coalescence in Polymer Blends: The Effects of Concentration and Compatibilization, *Macromolecules*, 1995, **28**(8), 2647–2657.
- 75 J. Qian and C. K. Law, Regimes of Coalescence and Separation in Droplet Collision, *J. Fluid Mech.*, 1997, **331**, 59–80, DOI: 10.1017/S0022112096003722.
- 76 A. K. Chesters, The Modelling of Coalescence Processes in Fluid-Liquid Dispersions: A Review of Current Understanding, *Chem. Eng. Res. Des.*, 1991, **68**(A4), 259–270.
- 77 P. R. Brazier-Smith, S. G. Jennings and J. Latham, The Interaction of Falling Water Drops: Coalescence, *Proc. R. Soc. London, Ser. A*, 1972, **326**(1566), 393–408.
- 78 S. Guido and M. Simeone, Binary Collision of Drops in Simple Shear Flow by Computer-Assisted Video Optical Microscopy, *J. Fluid Mech.*, 1998, **357**, 1–20.
- 79 T. Ban, F. Kawaizumi, S. Nii and K. Takahashi, Study of Drop Coalescence Behavior for Liquid-Liquid Extraction Operation, *Chem. Eng. Sci.*, 2000, **55**(22), 5385–5391.



- 80 J. J. Elmendorp and A. K. Van der Vegt, A Study on Polymer Blending Microrheology: Part Iv. The Influence of Coalescence on Blend Morphology Origination, *Polym. Eng. Sci.*, 1986, **26**(19), 1332–1338.
- 81 V. E. Ziegler and B. A. Wolf, Bimodal Drop Size Distributions During the Early Stages of Shear Induced Coalescence, *Polymer*, 2005, **46**(22), 9265–9273.
- 82 H. Wang, A. Z. Zinchenko and R. H. Davis, The Collision Rate of Small Drops in Linear Flow Fields, *J. Fluid Mech.*, 1994, **265**, 161–188.
- 83 M. Loewenberg and E. J. Hinch, Collision of Two Deformable Drops in Shear Flow, *J. Fluid Mech.*, 1997, **338**, 299–315, DOI: 10.1017/S0022112097005016.
- 84 A. Z. Zinchenko, M. A. Rother and R. H. Davis, A Novel Boundary-Integral Algorithm for Viscous Interaction of Deformable Drops, *Phys. Fluids*, 1997, **9**(6), 1493–1511, DOI: 10.1063/1.869275.
- 85 D. J. Jeffrey and Y. Onishi, Calculation of the Resistance and Mobility Functions for Two Unequal Rigid Spheres in Low-Reynolds-Number Flow, *J. Fluid Mech.*, 1984, **139**, 261–290.
- 86 S. Haber, G. Hetsroni and A. Solan, On the Low Reynolds Number Motion of Two Droplets, *Int. J. Multiphase Flow*, 1973, **1**(1), 57–71.
- 87 A. Z. Zinchenko, The Slow Asymmetric Motion of Two Drops in a Viscous Medium, *J. Appl. Math. Mech.*, 1980, **44**(13), 30–37.
- 88 G. K. Batchelor and J. T. Green, The Hydrodynamic Interaction of Two Small Freely-Moving Spheres in a Linear Flow Field, *J. Fluid Mech.*, 1972, **56**(2), 375–400.
- 89 P. A. Arp and S. G. Mason, The Kinetics of Flowing Dispersions: IX. Doublets of Rigid Spheres (Experimental), *J. Colloid Interface Sci.*, 1977, **61**(1), 44–61.
- 90 W. Bartok and S. G. Mason, Particle Motions in Sheared Suspensions: V. Rigid Rods and Collision Doublets of Spheres, *J. Colloid Sci.*, 1957, **12**(3), 243–262.
- 91 P. M. Adler, Interaction of Unequal Spheres: III. Experimental, *J. Colloid Interface Sci.*, 1981, **84**(2), 489–496.
- 92 P. A. Arp, R. T. Foister and S. G. Mason, Some Electrohydrodynamic Effects in Fluid Dispersions, *Adv. Colloid Interface Sci.*, 1980, **12**(4), 295–356.
- 93 C. L. Darabaner, J. K. Raasch and S. G. Mason, Particle Motions in Sheared Suspensions Xx: Circular Cylinders, *Can. J. Chem. Eng.*, 1967, **45**(1), 3–12.
- 94 Y. Yoon, M. Borrell, C. C. Park and L. G. Leal, Viscosity Ratio Effects on the Coalescence of Two Equal-Sized Drops in a Two-Dimensional Linear Flow, *J. Fluid Mech.*, 2005, **525**, 355–379, DOI: 10.1017/S0022112004002824.
- 95 W. Philippoff, Flow Birefringence and Stress, *J. Appl. Phys.*, 1956, **27**, 984.
- 96 P. G. DeGennes, Coil-Stretch Transition of Dilute Flexible Polymers under Ultrahigh Velocity Gradients, *J. Chem. Phys.*, 1974, **60**, 5030.
- 97 S. J. Haward, Microfluidic Extensional Rheometry Using Stagnation Point Flow, *Biomicrofluidics*, 2016, **10**(4), DOI: 10.1063/1.4945604.
- 98 S. J. Haward, Synovial Fluid Response to Extensional Flow: Effects of Dilution and Intermolecular Interactions, *PLoS One*, 2014, **9**(3), e92867, DOI: 10.1371/journal.pone.0092867.
- 99 R. Cerf and H. A. Scheraga, Flow Birefringence in Solutions of Macromolecules, *Chem. Rev.*, 1952, **51**(2), 185–261, DOI: 10.1021/cr60159a001.
- 100 M. R. Mackley and A. Keller, Flow Induced Polymer Chain Extension and Its Relation to Fibrous Crystallization, *Philos. Trans. R. Soc. London, Ser. A*, 1975, **278**(1276), 29–66.
- 101 P. N. Dunlap and L. G. Leal, Dilute Polystyrene Solutions in Extensional Flows: Birefringence and Flow Modification, *J. Non-Newtonian Fluid Mech.*, 1987, **23**, 5–48.
- 102 S. B. Kharchenko, J. F. Douglas, J. Obrzut, E. A. Grulke and K. B. Migler, Flow-Induced Properties of Nanotube-Filled Polymer Materials, *Nat. Mater.*, 2004, **3**(8), 564–568.
- 103 R. I. A. Tanner, Test Particle Approach to Flow Classification for Viscoelastic Fluids, *AIChE J.*, 1976, **22**(5), 910–918.
- 104 B. Liu, M. Shelley and J. Zhang, Oscillations of a Layer of Viscoelastic Fluid under Steady Forcing, *J. Non-Newtonian Fluid Mech.*, 2012, **175**, 38–43.
- 105 A. Keller and J. A. Odell, The Extensibility of Macromolecules in Solution; a New Focus for Macromolecular Science, *Colloid Polym. Sci.*, 1985, **263**(3), 181–201.
- 106 T. T. Perkins, D. E. Smith and S. Chu, Single Polymer Dynamics in an Elongational Flow, *Science*, 1997, **276**(5321), 2016–2021.
- 107 O. Scrivener, C. Berner, R. Cressely, R. Hocquart, R. Sellin and N. S. Vlachos, Dynamical Behaviour of Drag-Reducing Polymer Solutions, *J. Non-Newtonian Fluid Mech.*, 1979, **5**, 475–495.
- 108 S. D. Hudson, F. R. Phelan Jr, M. D. Handler, J. T. Cabral, K. B. Migler and E. J. Amis, Microfluidic Analog of the Four-Roll Mill, *Appl. Phys. Lett.*, 2004, **85**(2), 335–337.
- 109 F. R. Phelan and S. D. Hudson, Modeling of a Microfluidic Analog of the Four-Roll Mill for Materials Characterization, in *ANTEC Conference Proceedings Society of Plastics Engineers*, 2004, pp. 1054–1058.
- 110 S. J. Haward, A. Jaishankar, M. S. N. Oliveira, M. A. Alves and G. H. McKinley, Extensional Flow of Hyaluronic Acid Solutions in an Optimized Microfluidic Cross-Slot Device, *Biomicrofluidics*, 2013, **7**(4), 044108.
- 111 J. Penfold, E. Staples, I. Tucker, P. Carroll, I. Clayton, J. S. Cowan, G. Lawton, S. Amin, A. Ferrante and N. Ruddock, Elongational Flow Induced Ordering in Surfactant Micelles and Mesophases, *J. Phys. Chem. B*, 2006, **110**(2), 1073–1082, DOI: 10.1021/jp051122m.
- 112 M. Kisilak, H. Anderson, N. S. Babcock, M. R. Stetzer, S. H. J. Idziak and E. B. Sirota, An X-Ray Extensional Flow Cell, *Rev. Sci. Instrum.*, 2001, **72**(11), 4305–4307, DOI: 10.1063/1.1412259.
- 113 F. R. Phelan, S. D. Hudson and M. D. Handler, Fluid Dynamics Analysis of Channel Flow Geometries for Materials Characterization in Microfluidic Devices, *Rheol. Acta*, 2005, **45**(1), 59–71.
- 114 J. S. Lee, R. Dylla-Spears, N. P. Teclerian and S. J. Muller, Microfluidic Four-Roll Mill for All Flow



- Types, *Appl. Phys. Lett.*, 2007, **90**(7), 074103, DOI: 10.1063/1.2472528.
- 115 J. Wang, J. Han and D. Yu, Numerical Studies of Geometry Effects of a Two-Dimensional Microfluidic Four-Roll Mill on Droplet Elongation and Rotation, *Eng. Anal. Bound. Elem.*, 2012, **36**(10), 1453–1464.
- 116 S. J. Haward, M. S. Oliveira, M. A. Alves and G. H. McKinley, Optimized Cross-Slot Flow Geometry for Microfluidic Extensional Rheometry, *Phys. Rev. Lett.*, 2012, **109**(12), 128301, DOI: 10.1103/PhysRevLett.109.128301.
- 117 M. A. Alves, Design of a Cross-Slot Flow Channel for Extensional Viscosity Measurements, *Proceedings of the XVth International Congress on Rheology*, ed. L. G. Leal, R. H. Colby and A. J. Giacomin, American Institute of Physics, Monterey, 2008.
- 118 A. H. Motagamwala, *A Microfluidic, Extensional Flow Device for Manipulating Soft Particles*, 2013.
- 119 J. Guan, J. Liu, X. D. Li, J. Tao and J. T. Wang, Stokes Flow in a Two-Dimensional Micro-Device Combined by a Cross-Slot and a Microfluidic Four-Roll Mill, *Z. Angew. Math. Phys.*, 2015, **66**(1), 149–169, DOI: 10.1007/s00033-013-0396-z.
- 120 J. A. Odell and S. P. Carrington, Extensional Flow Oscillatory Rheometry, *J. Non-Newtonian Fluid Mech.*, 2006, **137**(1–3), 110–120, DOI: 10.1016/j.jnnfm.2006.03.010.
- 121 S. J. Haward, V. Sharma and J. A. Odell, Extensional Opto-Rheometry with Biofluids and Ultra-Dilute Polymer Solutions, *Soft Matter*, 2011, **7**(21), 9908–9921, DOI: 10.1039/c1sm05493g.
- 122 S. J. Haward, J. A. Odell, M. Berry and T. Ha, Extensional Rheology of Human Saliva, *Rheol. Acta*, 2011, **50**(11), 869–879.
- 123 S. J. Haward, J. A. Odell, Z. Li and X. F. Yuan, Extensional Rheology of Dilute Polymer Solutions in Oscillatory Cross-Slot Flow: The Transient Behaviour of Birefringent Strands, *Rheol. Acta*, 2010, **49**(6), 633–645, DOI: 10.1007/s00397-009-0420-6.
- 124 F. R. Phelan Jr, N. R. Hughes and J. A. Pathak, Chaotic Mixing in Microfluidic Devices Driven by Oscillatory Cross Flow, *Phys. Fluids*, 2008, **20**(2), 023101.
- 125 F. R. Phelan, P. Kutty and J. A. Pathak, An Electrokinetic Mixer Driven by Oscillatory Cross Flow, *Microfluid. Nanofluid.*, 2008, **5**(1), 101–118, DOI: 10.1007/s10404-007-0231-y.
- 126 R. C. van der Burgt, P. D. Anderson, J. M. den Toonder and F. N. Van De Vosse, A Microscale Pulsatile Flow Device for Dynamic Cross-Slot Rheometry, *Sens. Actuators, A*, 2014, **220**, 221–229.
- 127 A. R. Wheeler, W. R. Throdsset, R. J. Whelan, A. M. Leach, R. N. Zare, Y. H. Liao, K. Farrell, I. D. Manger and A. Daridon, Microfluidic Device for Single-Cell Analysis, *Anal. Chem.*, 2003, **75**(14), 3581–3586.
- 128 D. R. Link, S. L. Anna, D. A. Weitz and H. A. Stone, Geometrically Mediated Breakup of Drops in Microfluidic Devices, *Phys. Rev. Lett.*, 2004, **92**(5), 054503, DOI: 10.1103/PhysRevLett.92.054503.
- 129 J. C. Dijt, M. A. C. Stuart, J. E. Hofman and G. J. Fleer, Kinetics of Polymer Adsorption in Stagnation Point Flow, *Colloids Surf.*, 1990, **51**, 141–158, DOI: 10.1016/0166-6622(90)80138-T.
- 130 S. Yang, A. Undar and J. D. Zahn, A Microfluidic Device for Continuous, Real Time Blood Plasma Separation, *Lab Chip*, 2006, **6**(7), 871–880, DOI: 10.1039/b516401j.
- 131 G. Dockx, T. Verwijlen, W. Sempels, M. Nagel, P. Moldenaers, J. Hofkens and J. Vermant, Simple Microfluidic Stagnation Point Flow Geometries., *Biomicrofluidics*, 2016, **10**(4), 043506.
- 132 A. Lanzaro, D. Corbett and X. F. Yuan, Non-Linear Dynamics of Semi-Dilute Paam Solutions in a Microfluidic 3d Cross-Slot Flow Geometry, *J. Non-Newtonian Fluid Mech.*, 2017, **242**, 57–65.
- 133 M. G. Perera and K. Walters, Long-Range Memory Effects in Flows Involving Abrupt Changes in Geometry: Part I: Flows Associated with I-Shaped and T-Shaped Geometries, *J. Non-Newtonian Fluid Mech.*, 1977, **2**(1), 49–81.
- 134 T. Nishimura, K. Nakamura and A. Horikawa, Two-Dimensional Viscoelastic Flow of Polymer Solution at Channel Junction and Branch, *J. Text. Mach. Soc. Jpn.*, 1987, **33**(2), 37–45.
- 135 T. Nishimura, K. Nakamura and A. Horikawa, Two-Dimensional Viscoelastic Flow of Polymer Solution at Channel Junction and Branch, *J. Text. Mach. Soc. Jpn.*, 1985, **31**(1), 1–6.
- 136 D. M. Binding, K. Walters, J. Dheur and M. J. Crochet, Interfacial Effects in the Flow of Viscous and Elasticoviscous Liquids, *Philos. Trans. R. Soc. London, Ser. A*, 1987, **323**(1573), 449–469.
- 137 B. Thomases, M. Shelley and J. L. Thiffeault, A Stokesian Viscoelastic Flow: Transition to Oscillations and Mixing, *Phys. D*, 2011, **240**(20), 1602–1614.
- 138 H. A. Stone, Particle Assembly from Fluids, *Physics*, 2011, **4**, 17.
- 139 B. A. Parviz, D. Ryan and G. M. Whitesides, Using Self-Assembly for the Fabrication of Nano-Scale Electronic and Photonic Devices, *IEEE Trans. Adv. Packag.*, 2003, **26**(3), 233–241, DOI: 10.1109/Tadvp.2003.817971.
- 140 S. Zhang, Fabrication of Novel Biomaterials through Molecular Self-Assembly, *Nat. Biotechnol.*, 2013, **21**(10), 1171–1178.
- 141 G. M. Whitesides and B. Grzybowski, Self-Assembly at All Scales, *Science*, 2002, **295**(5564), 2418–2421, DOI: 10.1126/science.1070821.
- 142 G. M. Whitesides, J. P. Mathias and C. T. Seto, *Molecular Self-Assembly and Nanochemistry: A Chemical Strategy for the Synthesis of Nanostructures (No. Tr-45)*, CA, MA, 1991.
- 143 C. A. Mirkin, R. L. Letsinger, R. C. Mucic and J. J. Storhoff, A DNA-Based Method for Rationally Assembling Nanoparticles into Macroscopic Materials, *Nature*, 1996, **382**(6592), 607–609.
- 144 R. Häner, F. Samain and V. L. Malinovskii, DNA-Assisted Self-Assembly of Pyrene Foldamers, *Chem.–Eur. J.*, 2009, **15**(23), 5701–5708.
- 145 N. Bowden, A. Terfort, J. Carbeck and G. M. Whitesides, Self-Assembly of Mesoscale Objects into Ordered Two-



- Dimensional Arrays, *Science*, 1997, **276**(5310), 233–235, DOI: 10.1126/science.276.5310.233.
- 146 E. Fischer, Einfluss Der Configuration Auf Die Wirkung Der Enzyme, *Ber. Dtsch. Chem. Ges.*, 1894, **27**(3), 2985–2993.
- 147 S. Sacanna, W. T. Irvine, P. M. Chaikin and D. J. Pine, Lock and Key Colloids, *Nature*, 2010, **464**(7288), 575–578, DOI: 10.1038/nature08906.
- 148 O. Englander, D. Christensen, J. Kim, L. Lin and S. J. Morris, Electric-Field Assisted Growth and Self-Assembly of Intrinsic Silicon Nanowires, *Nano Lett.*, 2005, **5**(4), 705–708, DOI: 10.1021/nl050109a.
- 149 A. Winkleman, B. D. Gates, L. S. McCarty and G. M. Whitesides, Directed Self-Assembly of Spherical Particles on Patterned Electrodes by an Applied Electric Field., *Adv. Mater.*, 2005, **17**(12), 1507–1511.
- 150 A. Ashkin and J. M. Dzielic, Optical Trapping and Manipulation of Viruses and Bacteria, *Science*, 1987, **235**(4795), 1517–1520.
- 151 H. Zhang and K. K. Liu, Optical Tweezers for Single Cells., *J. R. Soc., Interface*, 2008, **5**(24), 671–690.
- 152 N. G. Green, H. Morgan and J. J. Milner, Manipulation and Trapping of Sub-Micron Bioparticles Using Dielectrophoresis, *J. Biochem. Biophys. Methods*, 1997, **35**(2), 89–102, DOI: 10.1016/S0165-022x(97)00033-X.
- 153 J. Voldman, R. A. Braff, M. Toner, M. L. Gray and M. A. Schmidt, Holding Forces of Single-Particle Dielectrophoretic Traps, *Biophys. J.*, 2001, **80**(1), 531–541, DOI: 10.1016/S0006-3495(01)76035-3.
- 154 A. T. Ohta, P. Y. Chiou, T. H. Han, J. C. Liao, U. Bhardwaj, E. R. B. McCabe, F. Q. Yu, R. Sun and M. C. Wu, Dynamic Cell and Microparticle Control Via Optoelectronic Tweezers, *J. Microelectromech. Syst.*, 2007, **16**(3), 491–499, DOI: 10.1109/Jmems.2007.896717.
- 155 M. C. Wu, Optoelectronic Tweezers, *Nat. Photonics*, 2011, **5**(6), 322–324.
- 156 J. Shi, D. Ahmed, X. Mao, S. C. Lin, A. Lawit and T. J. Huang, Acoustic Tweezers: Patterning Cells and Microparticles Using Standing Surface Acoustic Waves (Ssaw), *Lab Chip*, 2009, **9**(20), 2890–2895.
- 157 C. R. Courtney, C. E. Demore, H. Wu, A. Grinenko, P. D. Wilcox, S. Cochran and B. W. Drinkwater, Independent Trapping and Manipulation of Microparticles Using Dexterous Acoustic Tweezers, *Appl. Phys. Lett.*, 2014, **104**(15), 154103.
- 158 C. Haber and D. Wirtz, Magnetic Tweezers for DNA Micromanipulation, *Rev. Sci. Instrum.*, 2000, **71**(12), 4561–4570, DOI: .
- 159 K. C. Neuman and A. Nagy, Single-Molecule Force Spectroscopy: Optical Tweezers, Magnetic Tweezers and Atomic Force Microscopy, *Nat. Methods*, 2008, **5**(6), 491.
- 160 B. R. Lutz, J. Chen and D. T. Schwartz, Hydrodynamic Tweezers: 1. Noncontact Trapping of Single Cells Using Steady Streaming Microeddies, *Anal. Chem.*, 2006, **78**(15), 5429–5435.
- 161 R. Probst, Z. Cummins, C. Ropp, E. Waks and B. Shapiro, Flow Control of Small Objects on Chip: Manipulating Live Cells, Quantum Dots, and Nanowires, *IEEE Control Systems*, 2012, **32**(2), 26–53.
- 162 D. P. Bakker, H. J. Busscher and H. C. van der Mei, Bacterial Deposition in a Parallel Plate and a Stagnation Point Flow Chamber: Microbial Adhesion Mechanisms Depend on the Mass Transport Conditions, *Microbiology*, 2002, **148**(2), 597–603, DOI: 10.1099/00221287-148-2-597.
- 163 M. J. Dempsey, Colonisation of Antifouling Paints by Marine Bacteria, *Bot. Mar.*, 1981, **24**(4), 185–191.
- 164 J. Tsiabouklis, M. Stone, A. A. Thorpe, P. Graham, V. Peters, R. Heerlien, J. R. Smith, K. L. Green and T. G. Nevell, Preventing Bacterial Adhesion onto Surfaces: The Low-Surface energy Approach, *Biomaterials*, 1999, **20**(13), 1229–1235.
- 165 K. E. Cooksey and B. Wigglesworth-Cooksey, Adhesion of Bacteria and Diatoms to Surfaces in the Sea: A Review, *Aquat. Microb. Ecol.*, 1995, **9**(1), 87–96.
- 166 B. Little, R. Ray, P. Jones-Meehan, C. Lee and F. Mansfeld, Spatial Relationships between Marine Bacteria and Localized Corrosion on Polymer Coated Steel, *Biofouling*, 1999, **13**(4), 301–321.
- 167 C. M. Schroeder, E. S. Shaqfeh, H. P. Babcock and S. Chu, System and Method for Confining an Object to a Region of Fluid Flow Having a Stagnation Point, *US Pat.* 7,013,739, US-Patent-Appl-SN-10/841,011, March 21, 2006.
- 168 C. Ropp, Z. Cummins, R. Probst, S. Qin, J. T. Fourkas, B. Shapiro and E. Waks, Positioning and Immobilization of Individual Quantum Dots with Nanoscale Precision, *Nano Lett.*, 2010, **10**(11), 4673–4679, DOI: 10.1021/nl1029557.
- 169 M. Armani, S. Chaudhary, R. Probst and B. Shapiro, Using Feedback Control and Micro-Fluidics to Steer Individual Particles, in *18th IEEE International Conference on Micro Electro Mechanical Systems*, IEEE, 2005, pp. 855–858.
- 170 M. D. Armani, S. V. Chaudhary, R. Probst and B. Shapiro, Using Feedback Control of Microflows to Independently Steer Multiple Particles, *J. Microelectromech. Syst.*, 2006, **15**(4), 945–956, DOI: 10.1109/Jmems.2006.878863.
- 171 M. Tanyeri, E. M. Johnson-Chavarria and C. M. Schroeder, Hydrodynamic Trap for Single Particles and Cells, *Appl. Phys. Lett.*, 2010, **96**(22), 224101.
- 172 M. Tanyeri, E. M. Johnson-Chavarria and C. M. Schroeder, Hydrodynamic Trap for Single Cells and Micro- and Nanoparticles, *Biophys. J.*, 2011, **100**(3), 623a.
- 173 E. M. Johnson-Chavarria, M. Tanyeri and C. M. Schroeder, A Microfluidic-Based Hydrodynamic Trap for Single Particles, *J. Visualized Exp.*, 2011, **47**(47), e2517, DOI: 10.3791/2517.
- 174 A. Shenoy, M. Tanyeri and C. M. Schroeder, Characterizing the Performance of the Hydrodynamic Trap Using a Control-Based Approach, *Microfluid. Nanofluid.*, 2015, **18**(5–6), 1055–1066, DOI: 10.1007/s10404-014-1495-7.
- 175 M. Tanyeri, M. Ranka, N. Sittipolkul and C. M. Schroeder, A Microfluidic-Based Hydrodynamic Trap: Design and Implementation, *Lab Chip*, 2011, **11**(10), 1786–1794, DOI: 10.1039/c0lc00709a.



- 176 M. Tanyeri and C. M. Schroeder, Manipulation and Confinement of Single Particles Using Fluid Flow, *Nano Lett.*, 2013, **13**(6), 2357–2364, DOI: 10.1021/nl4008437.
- 177 T. M. Schneider, S. Mandre and M. P. Brenner, Algorithm for a Microfluidic Assembly Line, *Phys. Rev. Lett.*, 2011, **106**(9), 094503, DOI: 10.1103/PhysRevLett.106.094503.
- 178 K. D. Coventry and M. R. Mackley, Cross-Slot Extensional Flow Birefringence Observations of Polymer Melts Using a Multi-Pass Rheometer, *J. Rheol.*, 2008, **52**(2), 401–415, DOI: 10.1122/1.2836671.
- 179 D. Auhl, D. M. Hoyle, D. Hassell, T. D. Lord, O. G. Harlen, M. R. Mackley and T. C. B. McLeish, Cross-Slot Extensional Rheometry and the Steady-State Extensional Response of Long Chain Branched Polymer Melts, *J. Rheol.*, 2011, **55**(4), 875–900.
- 180 P. E. Arratia, C. C. Thomas, J. Diorio and J. P. Gollub, Elastic Instabilities of Polymer Solutions in Cross-Channel Flow, *Phys. Rev. Lett.*, 2006, **96**(14), 144502, DOI: 10.1103/PhysRevLett.96.144502.
- 181 J. Soulages, M. S. N. Oliveira, P. C. Sousa, M. A. Alves and G. H. McKinley, Investigating the Stability of Viscoelastic Stagnation Flows in T-Shaped Microchannels, *J. Non-Newtonian Fluid Mech.*, 2009, **163**(1–3), 9–24, DOI: 10.1016/j.jnnfm.2009.06.002.
- 182 G. N. Rocha, R. J. Poole, M. A. Alves and P. J. Oliveira, On Extensibility Effects in the Cross-Slot Flow Bifurcation, *J. Non-Newtonian Fluid Mech.*, 2009, **156**(1–2), 58–69, DOI: 10.1016/j.jnnfm.2008.06.008.
- 183 S. J. Haward and G. H. McKinley, Instabilities in Stagnation Point Flows of Polymer Solutions, *Phys. Fluids*, 2013, **25**(8), 083104, DOI: 10.1063/1.4818151.
- 184 S. J. Haward and G. H. McKinley, Stagnation Point Flow of Wormlike Micellar Solutions in a Microfluidic Cross-Slot Device: Effects of Surfactant Concentration and Ionic Environment, *Phys. Rev. E: Stat., Nonlinear, Soft Matter Phys.*, 2012, **85**(3), 031502.
- 185 N. Dubash, P. Cheung and A. Q. Shen, Elastic Instabilities in a Microfluidic Cross-Slot Flow of Wormlike Micellar Solution, *Soft Matter*, 2012, **8**(21), 5847.
- 186 S. J. Haward, T. J. Ober, M. S. Oliveira, M. A. Alves and G. H. McKinley, Extensional Rheology and Elastic Instabilities of a Wormlike Micellar Solution in a Microfluidic Cross-Slot Device, *Soft Matter*, 2011, **8**(2), 536–555.
- 187 C. A. Dreiss, Wormlike Micelles: Where Do We Stand? Recent Developments, Linear Rheology and Scattering Techniques, *Soft Matter*, 2007, **3**, 956–970.
- 188 P. A. Stone, S. D. Hudson, P. Dalhaimer, D. E. Discher, E. J. Amis and K. B. Migler, Dynamics of Wormlike Micelles in Elongational Flows, *Macromolecules*, 2006, **39**(20), 7144–7148, DOI: 10.1021/ma0611016.
- 189 J. A. Pathak and S. D. Hudson, Rheo-Optics of Equilibrium Polymer Solutions: Wormlike Micelles in Elongational Flow in a Microfluidic Cross-Slot, *Macromolecules*, 2006, **39**(25), 8782–8792, DOI: 10.1021/ma061355r.
- 190 M. S. N. Oliveira, F. T. Pinho, R. J. Poole, P. J. Oliveira and M. A. Alves, Purely Elastic Flow Asymmetries in Flow-Focusing Devices, *J. Non-Newtonian Fluid Mech.*, 2009, **160**(1), 31–39, DOI: 10.1016/j.jnnfm.2009.02.010.
- 191 A. M. Afonso, M. A. Alves and F. T. Pinho, Purely Elastic Instabilities in Three-Dimensional Cross-Slot Geometries, *J. Non-Newtonian Fluid Mech.*, 2010, **165**(13–14), 743–751, DOI: 10.1016/j.jnnfm.2010.03.010.
- 192 B. Thomases and M. Shelley, Transition to Mixing and Oscillations in a Stokesian Viscoelastic Flow, *Phys. Rev. Lett.*, 2009, **103**(9), 094501.
- 193 J. A. Odell, A. Keller and M. J. Miles, A Method for Studying Flow Induced Polymer Degradation-Verification of Chain Halving, *Polym. Commun.*, 1983, **24**(1), 7–10.
- 194 J. A. Odell and A. Keller, Flow-Induced Chain Fracture of Isolated Linear Macromolecules in Solution, *J. Polym. Sci., Part B: Polym. Phys.*, 1986, **24**(9), 1889–1916.
- 195 J. A. Odell, A. Keller and Y. Rabin, Flow induced Scission of Isolated Macromolecules, *J. Chem. Phys.*, 1988, **88**(6), 4022–4026.
- 196 O. G. Harlen, E. J. Hinch and J. M. Rallison, Birefringent Pipes: The Steady Flow of a Dilute Polymer Solution near a Stagnation Point, *J. Non-Newtonian Fluid Mech.*, 1992, **44**, 229–265.
- 197 O. G. Harlen, J. M. Rallison and M. D. Chilcott, High-Deborah-Number Flows of Dilute Polymer Solutions, *J. Non-Newtonian Fluid Mech.*, 1990, **34**(3), 319–349.
- 198 D. E. Smith and S. Chu, Response of Flexible Polymers to a Sudden Elongational Flow, *Science*, 1998, **281**(5381), 1335–1340.
- 199 T. Pfohl, F. Mugele, R. Seemann and S. Herminghaus, Trends in Microfluidics with Complex Fluids, *ChemPhysChem*, 2003, **4**(12), 1291–1298, DOI: 10.1002/cphc.200300847.
- 200 D. J. Mai, C. Brockman and C. M. Schroeder, Microfluidic Systems for Single DNA Dynamics, *Soft Matter*, 2012, **8**(41), 10560–10572, DOI: 10.1039/C2SM26036K.
- 201 R. G. Larson, H. Hu, D. E. Smith and S. Chu, Brownian Dynamics Simulations of a DNA Molecule in an Extensional Flow Field, *J. Rheol.*, 1999, **43**(2), 267–304, DOI: 10.1122/1.550991.
- 202 C. M. Schroeder, H. P. Babcock, E. S. Shaqfeh and S. Chu, Observation of Polymer Conformation Hysteresis in Extensional Flow, *Science*, 2003, **301**(5639), 1515–1519, DOI: 10.1126/science.1086070.
- 203 W. Xu and S. J. Muller, Exploring Both Sequence Detection and Restriction Endonuclease Cleavage Kinetics by Recognition Site Via Single-Molecule Microfluidic Trapping, *Lab Chip*, 2011, **11**(3), 435–442.
- 204 S. Cha, T. Shin, S. S. Lee, W. Shim, G. Lee, S. J. Lee, W. Shim and J. M. Kim, Cell Stretching Measurement Utilizing Viscoelastic Particle Focusing, *Anal. Chem.*, 2012, **84**(23), 10471–10477.
- 205 N. Neve, S. S. Kohles, S. R. Winn and D. C. Tretheway, Manipulation of Suspended Single Cells by Microfluidics and Optical Tweezers, *Cell. Mol. Bioeng.*, 2010, **3**(3), 213–228, DOI: 10.1007/s12195-010-0113-3.



- 206 S. Kumar, A. Shenoy and C. Schroeder, Observing Polymersome Dynamics in Controlled Microscale Flows, in *APS Meeting Abstracts*, 2015, p. 50013.
- 207 P. R. Start, S. D. Hudson, E. K. Hobbie and K. B. Migler, Breakup of Carbon Nanotube Floes in Microfluidic Traps, *J. Colloid Interface Sci.*, 2006, **297**(2), 631–636, DOI: 10.1016/j.jcis.2005.11.044.
- 208 Y. N. Young and M. J. Shelley, Stretch-Coil Transition and Transport of Fibers in Cellular Flows, *Phys. Rev. Lett.*, 2007, **99**(5), 058303, DOI: 10.1103/PhysRevLett.99.058303.
- 209 C. Ulloa, A. Ahumada and M. L. Cordero, The Effect of Confinement on the Deformation of Microfluidic Drops, *Phys. Rev. E: Stat., Nonlinear, Soft Matter Phys.*, 2014, **89**, 033004.
- 210 W. Xu and S. J. Muller, Polymer-Monovalent Salt-Induced DNA Compaction Studied Via Single-Molecule Microfluidic Trapping, *Lab Chip*, 2012, **12**(3), 647–651, DOI: 10.1039/c2lc20880f.
- 211 Y. Li, C. Brockman, D. Yates, G. McKenna, C. Schroeder, M. San Francisco, J. Kornfield and R. Anderson, Comparison of the Coil-Stretch Transition of Linear and Circular DNA in Planar Extensional Flow, in *Abstracts of Papers of the American Chemical Society*, American Chemical Society, 2014, p. 420.
- 212 Y. Li, K. W. Hsiao, C. A. Brockman, D. Y. Yates, R. M. Robertson-Anderson, J. A. Kornfield, M. J. San Francisco, C. M. Schroeder and G. B. McKenna, When Ends Meet: Circular DNA Stretches Differently in Elongational Flows, *Macromolecules*, 2015, **48**(16), 5997–6001.
- 213 Y. Zhou and C. M. Schroeder, Single Polymer Dynamics under Large Amplitude Oscillatory Extensional (Laoe) Flow, in *APS Meeting Abstracts*, Chicago, 2016, p. 7.
- 214 S. J. Haward, Buckling Instabilities in Dilute Polymer Solution Elastic Strands, *Rheol. Acta*, 2010, **49**(11–12), 1219–1225, DOI: 10.1007/s00397-010-0467-4.
- 215 V. Sharma, S. J. Haward, J. Serdy, B. Keshavarz, A. Soderlund, P. Threlfall-Holmes and G. H. McKinley, The Rheology of Aqueous Solutions of Ethyl Hydroxy-Ethyl Cellulose (Ehec) and Its Hydrophobically Modified Analogue (Hmehec): Extensional Flow Response in Capillary Break-up, Jetting (Rojer) and in a Cross-Slot Extensional Rheometer, *Soft Matter*, 2015, **11**(16), 3251–3270, DOI: 10.1039/c4sm01661k.
- 216 S. J. Haward, J. A. Odell, Z. Li and X. F. Yuan, The Rheology of Polymer Solution Elastic Strands in Extensional Flow, *Rheol. Acta*, 2010, **49**(7), 781–788.
- 217 J. T. Wang, D. M. Yu, H. F. Jing and J. Tao, Hydrodynamic Control of Droplets Coalescence in Microfluidic Devices to Fabricate Two-Dimensional Anisotropic Particles through Boundary Element Method, *Chem. Eng. Res. Des.*, 2014, **92**(11), 2223–2230, DOI: 10.1016/j.cherd.2014.02.026.
- 218 A. T. Florence and D. Whitehill, The Formulation and Stability of Multiple Emulsions, *Int. J. Pharm.*, 1982, **11**(4), 277–308, DOI: 10.1016/0378-5173(82)90080-1.
- 219 J. Wang, J. Liu, J. Han and J. Guan, Effects of Complex Internal Structures on Rheology of Multiple Emulsions Particles in 2d from a Boundary Integral Method, *Phys. Rev. Lett.*, 2013, **110**(6), 066001, DOI: 10.1103/PhysRevLett.110.066001.
- 220 J. T. Wang, J. X. Liu, J. J. Han and J. Guan, Rheology Investigation of the Globule of Multiple Emulsions with Complex Internal Structures through a Boundary Element Method, *Chem. Eng. Sci.*, 2013, **96**, 87–97, DOI: 10.1016/j.ces.2013.02.065.
- 221 Y. B. Bae, H. K. Jang, T. H. Shin, G. Phukan, T. T. Tran, G. Lee, W. R. Hwang and J. M. Kim, Microfluidic Assessment of Mechanical Cell Damage by Extensional Stress., *Lab Chip*, 2016, **16**(1), 96–103.
- 222 E. T. Papoutsakis, Fluid-Mechanical Damage of Animal Cells in Bioreactors, *Trends Biotechnol.*, 1991, **9**(1), 427–437.
- 223 Y. Chisti, Animal-Cell Damage in Sparged Bioreactors, *Trends Biotechnol.*, 2000, **18**(10), 420–432.
- 224 Y. Chisti, Hydrodynamic Damage to Animal Cells, *Crit. Rev. Biotechnol.*, 2001, **21**(2), 67–110, DOI: 10.1080/20013891081692.
- 225 M. A. Garcia-Briones and J. J. Chalmers, Flow Parameters Associated with Hydrodynamic Cell Injury, *Biotechnol. Bioeng.*, 1994, **44**(9), 1089–1098.
- 226 R. I. Tanner and R. R. Huilgol, On a Classification Scheme for Flow Fields, *Rheol. Acta*, 1975, **14**(11), 959–962.
- 227 R. Dylla-Spears, L. L. Sohn and S. J. Muller, Use of Stagnation Point Flows for DNA Trapping, Manipulation, and Target Sequence Detection, in *The XV International Congress On Rheology: The Society of Rheology 80th Annual Meeting*, 2008, pp. 959–961.
- 228 R. Dylla-Spears, J. E. Townsend, L. Jen-Jacobson, L. L. Sohn and S. J. Muller, Single-Molecule Sequence Detection Via Microfluidic Planar Extensional Flow at a Stagnation Point, *Lab Chip*, 2010, **10**(12), 1543–1549, DOI: 10.1039/b926847b.
- 229 A. Castro and J. G. Williams, Single-Molecule Detection of Specific Nucleic Acid Sequences in Unamplified Genomic DNA, *Anal. Chem.*, 1997, **69**(19), 3915–3920.
- 230 M. Champion and P. A. Libby, Reynolds Stress Description of Opposed and Impinging Turbulent Jets. Part I: Closely Spaced Opposed Jets, *Phys. Fluids A*, 1993, **5**(1), 203–216.
- 231 A. M. Dehkordi, Application of a Novel-Opposed-Jets Contacting Device in Liquid-Liquid Extraction, *Chem. Eng. Process.*, 2002, **41**(3), 251–258.
- 232 A. Schaffer and A. B. Cambel, The Effect of an Opposing Jet on Flame Stability, *Jet Propul.*, 1955, **25**(6), 284–287.
- 233 A. Schaffer and A. B. Cambel, Continued Investigations of the Opposing Jet Flameholder, *Jet Propul.*, 1956, **26**(7), 576–578.
- 234 S. S. Penner and F. Williams, Recent Studies on Flame Stabilization of Premixed Turbulent Gases, *Appl. Mech. Rev.*, 1957, **10**(6), 229–237.
- 235 G. Dixon-Lewis, Laminar Premixed Flame Extinction Limits. I Combined Effects of Stretch and Upstream Heat Loss in the Twin-Flame Unburnt-to-Unburnt Opposed Flow Configuration, in *Proceedings of the Royal Society of London A: Mathematical, Physical and Engineering Sciences London*, The Royal Society, London, 1996, pp. 1857–1884.



- 236 G. Dixon-Lewis, Laminar Premixed Flame Extinction Limits. II Combined Effects of Stretch and Radiative Loss in the Single Flame Unburnt-to-Burnt and the Twin-Flame Unburnt-to-Unburnt Opposed Flow Configurations, in *Proceedings of the Royal Society of London A: Mathematical, Physical and Engineering Sciences London*, London, 2006, pp. 349–370.
- 237 M. Matalon, On Flame Stretch, *Combust. Sci. Technol.*, 1983, **31**(3–4), 169–181.
- 238 S. M. Candel and T. J. Poinso, Flame Stretch and the Balance Equation for the Flame Area, *Combust. Sci. Technol.*, 1990, **70**(1–3), 1–15, DOI: 10.1080/00102209008951608.
- 239 A. E. Potter and J. N. Butler, A Novel Combustion Measurement Based on the Extinguishment of Diffusion Flames, *ARS J.*, 1959, **29**(1), 54–56.
- 240 A. E. Potter, S. Heimel and J. N. Butler, in *Apparent Flame Strength: A Measure of Maximum Reaction Rate in Diffusion Flames*, Symposium on Combustion (International) Elsevier, 1961, pp. 1027–1034.
- 241 V. K. Jain and H. S. Mukunda, On the Ignition and Extinction Problems in Forced Convection Systems, *Int. J. Heat Mass Transfer*, 1968, **11**(3), 491–508.
- 242 K. Seshadri, C. Trevino and M. D. Smooke, Analysis of the Structure and Mechanisms of Extinction of a Counterflow Methanol-Air Diffusion Flame, *Combust. Flame*, 1989, **76**(2), 111–132, DOI: 10.1016/0010-2180(89)90061-8.
- 243 A. Tamir and A. Kitron, Applications of Impinging-Streams in Chemical Engineering Processes—Review, *Chem. Eng. Commun.*, 1987, **50**(1–6), 241–330.
- 244 P. M. Chung, F. E. Fendell and J. F. Holt, Nonequilibrium Anomalies in the Development of Diffusion Flames, *AIAA J.*, 1966, **4**(6), 1020–1026.
- 245 F. E. Fendell, Ignition and Extinction in Combustion of Initially Unmixed Reactants, *J. Fluid Mech.*, 1965, **21**(2), 281–303, DOI: 10.1017/S0022112065000186.
- 246 S. A. Safvi and T. J. Mountziaris, Gas Phase Decomposition Kinetics of Moppe Precursors in a Counterflow Jet Reactor, in *Chemical Perspectives of Microelectronic Materials III (Mat. Res. SOC. Symp. Proc.)*, ed. C. R. Abernathy, 1993.
- 247 V. K. Jain and H. S. Mukunda, The Extinction Problem in an Opposed Jet Diffusion Flame with Competitive Reactions, *Combust. Sci. Technol.*, 1969, **1**(2), 105–117.
- 248 A. Linan, The Asymptotic Structure of Counterflow Diffusion Flames for Large Activation Energies, *Acta Astronaut.*, 1974, **1**(7), 1007–1039.
- 249 D. B. Spalding, Theory of Mixing and Chemical Reaction in the Opposed-Jet Diffusion Flame, *ARS J.*, 1961, **31**(6), 763–771.
- 250 S. A. Safvi and T. J. Mountziaris, A New Reactor Studies for Purely Homogeneous Kinetic of Endothermic Reactions, *AIChE J.*, 1994, **40**(9), 1535–1548.
- 251 J. L. Zilko, Metallo-Organic Cvd Technology and Equipment, in *Handbook of Thin-Film Deposition Processes and Techniques*, ed. K. K. Shuegraf, Noyes Publications, Park Ridge, NJ, 1988, pp. 84–101.
- 252 H. O. Pierson, *Handbook of Chemical Vapor Deposition: Principles, Technology and Applications*, 1999.
- 253 P. Burgraaf, *The Status of Mocvd Technology*, Semiconductor International, 1993, pp. 80–83.
- 254 Z. G. Sun, W. F. Li and H. F. Liu, Stagnation Point Offset of Two Opposed Jets, *Ind. Eng. Chem. Res.*, 2010, **49**(12), 5877–5883.
- 255 F. C. Frank, A. Keller and M. R. Mackley, Polymer Chain Extension Produced by Impinging Jets and Its Effect on Polyethylene Solution, *Polymer*, 1971, **12**(7), 467–473.
- 256 M. R. Mackley, Flow Singularities, Polymer Chain Extension and Hydrodynamic Instabilities, *J. Non-Newtonian Fluid Mech.*, 1978, **4**(1), 111–136.
- 257 A. J. Muller, J. A. Odell and A. Keller, Elongational Flow and Rheology of Monodisperse Polymers in Solution, *J. Non-Newtonian Fluid Mech.*, 1988, **30**(2–3), 99–118, DOI: 10.1016/0377-0257(88)85018-3.
- 258 J. A. Odell, A. J. Muller, K. A. Narh and A. Keller, Degradation of Polymer Solutions in Extensional Flows, *Macromolecules*, 1990, **23**(12), 3092–3103.
- 259 S. P. Carrington, J. P. Tatham, J. A. Odell and A. E. Saez, Macromolecular Dynamics in Extensional Flows: 1. Birefringence and Viscometry, *Polymer*, 1997, **38**(16), 4151–4164.
- 260 S. P. Carrington, J. P. Tatham, J. A. Odell and A. E. Saez, Macromolecular Dynamics in Extensional Flows: 2. The Evolution of Molecular Strain, *Polymer*, 1997, **38**(18), 4595–4607.
- 261 R. K. Prud'homme and G. G. Warr, Elongational Flow of Solutions of Rodlike Micelles, *Langmuir*, 1994, **10**(10), 3419–3426.
- 262 C. G. Hermansky and D. V. Boger, Opposing-Jet Viscometry of Fluids with Viscosity Approaching That of Water, *J. Non-Newtonian Fluid Mech.*, 1995, **56**(1), 1–14.
- 263 S. L. Ng, R. P. Mun, D. V. Boger and D. F. James, Extensional Viscosity Measurements of Dilute Solutions of Various Polymers, *J. Non-Newtonian Fluid Mech.*, 1996, **65**(2–3), 291–298, DOI: 10.1016/0377-0257(96)01463-2.
- 264 G. G. Fuller, C. A. Cathey, B. Hubbard and B. E. Zebrowski, Extensional Viscosity Measurements for Low-Viscosity Fluids, *J. Rheol.*, 1987, **31**(3), 235–249.
- 265 C. A. Cathey and G. G. Fuller, Uniaxial and Biaxial Extensional Viscosity Measurements of Dilute and Semi-Dilute Solutions of Rigid Rod Polymers, *J. Non-Newtonian Fluid Mech.*, 1988, **30**(2–3), 303–316, DOI: 10.1016/0377-0257(88)85030-4.
- 266 P. Dontula, M. Pasquali, L. E. Scriven and C. C. W. Macosko, Can Extensional Viscosity Be Measured with Opposed-Nozzle Devices?, *Rheol. Acta*, 1997, **36**(4), 429–448.
- 267 P. Jonsson, J. McColl, R. W. Clarke, V. P. Ostanin, B. Jonsson and D. Klennerman, Hydrodynamic Trapping of Molecules in Lipid Bilayers, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**(26), 10328–10333, DOI: 10.1073/pnas.1202858109.



- 268 A. Ainla, G. D. M. Jeffries, R. Brune, O. Orwar and A. Jesorka, A Multifunctional Pipette, *Lab Chip*, 2012, **12**, 1255–1261.
- 269 A. Ainla, I. Gozen, B. Hakonen and A. Jesorka, Lab on a Biomembrane: Rapid Prototyping and Manipulation of 2d Fluidic Lipid Bilayer Circuits, *Sci. Rep.*, 2013, **3**, 2734, DOI: 10.1038/srep03317.
- 270 A. Ainla, E. T. Jansson, N. Stepanyants, O. Orwar and A. Jesorka, A Microfluidic Pipette for Single-Cell Pharmacology, *Anal. Chem.*, 2010, **82**(11), 4529–4536, DOI: 10.1021/ac100480f.
- 271 A. Ainla, E. T. Jansson, N. Stepanyants, O. Orwar and A. Jesorka, A Multi Purpose Microfluidic Pipette for Single Cell Analysis, in *14th International Conference on Miniaturized Systems for Chemistry and Life Sciences*, Groningen, Netherlands, 2010, pp. 932–934.
- 272 A. Ainla, S. J. Xu, N. Sanchez, G. D. M. Jeffries and A. Jesorka, Single-Cell Electroporation Using a Multifunctional Pipette, *Lab Chip*, 2012, **12**(22), 4605–4609, DOI: 10.1039/c2lc40563f.
- 273 A. Ahemaiti, A. Ainla, G. D. Jeffries, H. Wigström, O. Orwar, A. Jesorka and K. Jardemark, A Multifunctional Pipette for Localized Drug Administration to Brain Slices, *J. Neurosci. Methods*, 2013, **219**(2), 292–296.
- 274 A. Ahemaiti, H. Wigstrom, A. Ainla, G. D. M. Jeffries, O. Orwar, A. Jesorka and K. Jardemark, Spatial Characterization of a Multifunctional Pipette for Drug Delivery in Hippocampal Brain Slices, *J. Neurosci. Methods*, 2015, **241**, 132–136, DOI: 10.1016/j.jneumeth.2014.12.017.
- 275 M. Safavieh, M. A. Qasaimeh, A. Vakil, D. Juncker and T. Gervais, Two-Aperture Microfluidic Probes as Flow Dipole: Theory and Application, *Sci. Rep.*, 2015, **5**, 11943.
- 276 C. M. Perrault, M. A. Qasaimeh and D. Juncker, The Microfluidic Probe: Operation and Use for Localized Surface Processing, *J. Visualized Exp.*, 2009, **28**, e1418, DOI: 10.3791/1418.
- 277 T. Gervais, M. Safavieh, M. A. Qasaimeh and D. Juncker, Systematic Analysis of Microfluidic Probe Design and Operation, in *36th Annual International Conference of the IEEE in Engineering in Medicine and Biology Society (EMBC)*, 2014, pp. 1567–1570.
- 278 D. Juncker, H. Schmid and E. Delamarche, Multipurpose Microfluidic Probe, *Nat. Mater.*, 2005, **4**(8), 622–628.
- 279 N. Ostromohov, M. Bercovici and G. V. Kaigala, Delivery of Minimally Dispersed Liquid Interfaces for Sequential Surface Chemistry, *Lab Chip*, 2016, **15**, 3015–3023.
- 280 R. D. Lovchik, G. V. Kaigala, M. Georgiadis and E. Delamarche, Micro-Immunohistochemistry Using a Microfluidic Probe, *Lab Chip*, 2012, **12**(6), 1040–1043, DOI: 10.1039/c2lc21016a.
- 281 A. Queval, N. R. Ghattamaneni, C. M. Perrault, R. Gill, M. Mirzaei, R. A. McKinney and D. Juncker, Chamber and Microfluidic Probe for Microperfusion of Organotypic Brain Slices, *Lab Chip*, 2010, **10**(3), 326–334, DOI: 10.1039/b916669f.
- 282 A. Sarkar, S. Kolitz, D. A. Lauffenburger and J. Han, Microfluidic Probe for Single-Cell Analysis in Adherent Tissue Culture, *Nat. Commun.*, 2014, **5**, 3421, DOI: 10.1038/ncomms4421.
- 283 A. Kashyap, J. Autebert, E. Delamarche and G. V. Kaigala, Selective Local Lysis and Sampling of Live Cells for Nucleic Acid Analysis Using a Microfluidic Probe, *Sci. Rep.*, 2016, **6**, 29579.
- 284 M. A. Qasaimeh, T. Gervais and D. Juncker, Microfluidic Quadrupole and Floating Concentration Gradient, *Nat. Commun.*, 2011, **2**, 464.
- 285 J. Autebert, A. Kashyap, R. D. Lovchik, E. Delamarche and G. V. Kaigala, Hierarchical Hydrodynamic Flow Confinement: Efficient Use and Retrieval of Chemicals for Microscale Chemistry on Surfaces, *Langmuir*, 2014, **30**(12), 3640–3645, DOI: 10.1021/la500875m.
- 286 J. Autebert, J. F. Cors, D. P. Taylor and G. V. Kaigala, Convection-Enhanced Biopatterning with Recirculation of Hydrodynamically Confined Nanoliter Volumes of Reagents, *Anal. Chem.*, 2016, **88**(6), 3235–3242.
- 287 S. Mao, Y. Zhang, W. Zhang, H. Zeng, H. Nakajima, J. M. Lin and K. Uchiyama, Convection-Diffusion Layer in an “Open Space” for Local Surface Treatment and Microfabrication Using a Four-Aperture Micro Chemical Pen, *ChemPhysChem*, 2017, **18**(17), 2357–2363.
- 288 H. Shiku, T. Yamakawa, Y. Nashimoto, Y. Takahashi, Y.-s. Torisawa, T. Yasukawa, T. Ito-Sasaki, M. Yokoo, H. Abe, H. Kambara and T. Matsue, A Microfluidic Dual Capillary Probe to Collect Messenger Rna from Adherent Cells and Spheroids, *Anal. Biochem.*, 2009, **385**, 138–142.
- 289 T. Kang, J. Han and K. S. Lee, Concentration Gradient Generator Using a Convective–Diffusive Balance, *Lab Chip*, 2008, **8**(7), 1220–1222.
- 290 E. Kolaczowska and P. Kubes, Neutrophil Recruitment and Function in Health and Inflammation, *Nat. Rev. Immunol.*, 2003, **13**(3), 159–175.
- 291 T. M. Keenan and A. Folch, Biomolecular Gradients in Cell Culture Systems, *Lab Chip*, 2008, **8**(1), 34–57, DOI: 10.1039/b711887b.
- 292 D. Irimia, Microfluidic Technologies for Temporal Perturbations of Chemotaxis, *Annu. Rev. Biomed. Eng.*, **15**(12), 259–284.
- 293 M. A. Qasaimeh, M. Astolfi, M. Pyzik, S. Vidal and D. Juncker, Neutrophil Dynamics During Migration in Microfluidic Concentration Gradients, in *Proceeding of the 40th Annual Northeast Bioengineering Conference (NEBEC)*, Boston, USA, April 25–27, 2014, pp. 1–2.
- 294 M. A. Qasaimeh, M. Astolfi, M. Pyzik, S. Vidal and D. Juncker, Neutrophils Migrate Longer Distances in Moving Microfluidic Concentration Gradients Compared to Static Ones, in *The 17th International Conference on Miniaturized Systems for Chemistry and Life Sciences*, Freiburg, Germany, October 2013, pp. 2007–2009.
- 295 M. A. Qasaimeh, S. G. Ricoult and D. Juncker, Microfluidic Probes for Use in Life Sciences and Medicine, *Lab Chip*, 2013, **13**(1), 40–50, DOI: 10.1039/c2lc40898h.



- 296 M. A. Qasaimeh, R. Stefavieh and D. Juncker, The Generation of Biochemical Gradients in a Microfluidic Stagnant Zone, in *Proceedings of MMB 2009, The fifth International Conference on Microtechnologies in Medicine and Biology*, Quebec, QC, Canada, April 2009, pp. 62–63.
- 297 A. T. Brimmo and M. A. Qasaimeh, Microfluidic Probes and Quadrapoles: A New Era of Open Microfluidics, *IEEE Nanotechnol. Mag.*, 2017, **11**(1), 20–31.
- 298 M. A. Qasaimeh, S. G. Ricoult and D. Juncker, Microfluidic probes to process surfaces, cells, and tissues, *Selected Topics in Nanomedicine (Regenerative Medicine, Artificial Cells and Nanomedicine)*, World Science Publisher/Imperial College Press, 2013, ch. 11, pp. 257–280.
- 299 M. Chachivili, Y. L. Zhang and J. A. Frangos, G Protein-Coupled Receptors Sense Fluid Shear Stress in Endothelial Cells, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**(42), 15463–15468, DOI: 10.1073/pnas.0607224103.
- 300 N. Li Jeon, H. Baskaran, S. K. W. Dertinger, G. M. Whitesides, L. Van De Water and M. Toner, Neutrophil Chemotaxis in Linear and Complex Gradients of Interleukin-8 Formed in a Microfabricated Device, *Nat. Biotechnol.*, 2002, **20**(8), 826–830.
- 301 J. A. Fay, Theory of Stagnation Point Heat Transfer in Dissociated Air, *J. Aerosp. Sci.*, 1958, **25**(2), 73–85.
- 302 G. K. Batchelor, *An Introduction to Fluid Dynamics*, Cambridge university press, 2000.

