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A heterologous expression platform in *Aspergillus nidulans* for the elucidation of cryptic secondary metabolism biosynthetic gene clusters: discovery of the *Aspergillus fumigatus* sartorypyrone biosynthetic pathway†

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Aspergillus fumigatus is a serious human pathogen causing life-threatening Aspergillosis in immunocompromised patients. Secondary metabolites (SMs) play an important role in pathogenesis, but the products of many SM biosynthetic gene clusters (BGCs) remain unknown. In this study, we have developed a heterologous expression platform in *Aspergillus nidulans*, using a newly created genetic dereplication strain, to express a previously unknown BGC from *A. fumigatus* and determine its products. The BGC produces sartorypyrones, and we have named it the *spy* BGC. Analysis of targeted gene deletions by HRESIMS, NMR, and microcrystal electron diffraction (MicroED) enabled us to identify 12 products from the *spy* BGC. Seven of the compounds have not been isolated previously. We also individually expressed the polyketide synthase (PKS) gene *spyA* and demonstrated that it produces the polyketide triacetic acid lactone (TAL), a potentially important biorenewable platform chemical. Our data have allowed us to propose a biosynthetic pathway for sartorypyrones and related natural products. This work highlights the potential of using the *A. nidulans* heterologous expression platform to uncover cryptic BGCs from *A. fumigatus* and other species, despite the complexity of their secondary metabolomes.

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Introduction

Aspergillosis is the spectrum of diseases caused by *Aspergillus* species that colonize the bronchial trees of patients.¹ It is estimated that there are over 300 000 cases of life-threatening Aspergillosis in humans annually, with a mortality rate of 30–80%.^{2–5} Invasive Aspergillosis (IA) is one of the most common causes of death in severely immunocompromised patients, with

mortality rates up to 40% to 50% in patients with acute leukemia and recipients of hematopoietic stem cell transplantations.

Aspergillus fumigatus is by far the most common species causing IA (70%).⁵ The World Health Organization “WHO fungal priority pathogens list to guide research, development and public health action 2022” classified *A. fumigatus* as one of four species in the critical priority group, the most urgent need category.⁶ Secondary metabolites (SMs) in *A. fumigatus* contribute to its unique ability to survive in the environment and establish itself in human hosts.^{7,8} SMs are small molecules produced by fungi, prokaryotes and plants that are not strictly required for growth or reproduction, but they confer a selective advantage to the producing organism. In *A. fumigatus*, SMs play important roles in iron and copper uptake and homeostasis, for example.^{8,9} *A. fumigatus* also produces SMs that kill or inhibit competitors and these activities are often important to pathogenesis. Gliotoxin, for one example of several, protects *A. fumigatus* against amoebae in the environment and against T lymphocytes and the macrophage immune response in pathogenesis. Genetic elimination of gliotoxin production results in attenuation of virulence.^{10–12}

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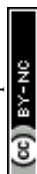
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SMs in filamentous fungi are synthesized by enzymes encoded by genes organized in contiguous biosynthetic gene clusters (BGCs).¹³ The genes of the BGCs are coordinately regulated, and the BGCs often contain genes encoding transcription factors that drive the expression of the other genes of the BGC. SM BGCs vary significantly across species and strains of *Aspergillus*.¹⁴ The two most studied isolates of *A. fumigatus* are Af293 and CEA10.^{15,16} Bioinformatics indicate that the *A. fumigatus* Af293 strain contains 34 predicted SM BGCs, and the CEA10 derivative strain A1163 has 33 predicted SM BGCs. The SM BGCs of the two strains are largely but not completely overlapping, with three BGCs unique to Af293 and two unique to CEA10 making a total of 36 BGCs. The products of only 19 of these BGCs have been determined, leaving, prior to this study, 17 to be determined.^{17–19}

In fungi, many SM BGCs are not expressed under normal laboratory growth conditions, whereas others are expressed at high levels, resulting in a “forest” of SM peaks in LC/MS traces. To determine the SMs produced by cryptic BGCs, one needs to find a way to activate their expression. Once they are expressed, the SMs they produce must be identified and purified from among the forest of metabolites produced by the fungus. Various approaches have been developed in the model fungus *Aspergillus nidulans* to activate SM production.^{20–31} In principle, some of these approaches could be transferred to *A. fumigatus* to elucidate its secondary metabolome, but it would require a great deal of time and effort to develop them. Perhaps more seriously, metabolite profiles from *A. fumigatus* grown under laboratory conditions are quite complex due to high levels of production of many compounds such as fumigaclavine C, fumiquinazoline, and monomethyl-sulochrin and their pathway intermediates.^{32–38} Therefore, identifying minor compounds is difficult due to the interference of these major metabolites, and determining the BGC responsible for any minor compound is equally difficult. In *A. nidulans*, we have developed a strategy we call “genetic dereplication” that facilitates new compound discovery. We have deleted eight BGCs, in their entirety, that produce abundant SMs (>244 000 bp deleted), thereby reducing the complexity of SM profiles such that novel compounds are more easily detected.²⁶ Eliminating highly expressed biosynthetic pathways may also increase pools of SM precursors such as acetyl-CoA and malonyl-CoA for pathways expressed at low levels. Because developing the genetic dereplication approach for *A. fumigatus* would be a lengthy process, we are developing methods to express *A. fumigatus* BGCs in *A. nidulans* and determine their products.

In this study, we report the expression of a heretofore cryptic *A. fumigatus* secondary metabolite biosynthesis pathway in *A. nidulans* and the determination of the products of the BGC. Using a genetic dereplication strain to reduce the SM background, we refactored the target BGC genes into *A. nidulans*, placing each gene under the control of an inducible promoter. The heterologously expressed SMs were purified, and the structures were determined by NMR and electron cryo-microscopy (cryoEM) microcrystal electron diffraction (MicroED). We found that the BGC encodes a family of meroterpenoids called sartorypyrones that have never been

reported in *A. fumigatus*. Two of the compounds are novel while two have been found in the closely related nonpathogenic fungus *Neosartorya fischeri* (KUFC 6344 and FO-5897).^{39,40} We generated deletant strains that were each missing one gene of the biosynthetic pathway. Some of these strains accumulated chemically stable intermediates and shunt products in sufficient amounts for complete structural characterization, resulting in the identification of a total of eight additional, related compounds. We also individually expressed the non-reducing polyketide synthase (NR-PKS) gene of the BGC and demonstrated that triacetic acid lactone (TAL) is the product of the PKS. TAL is a potentially important biorenewable platform chemical.⁴¹ Combining these data with further bioinformatic analysis, we propose a biosynthetic pathway for sartorypyrones. Importantly, the approach reported in this study, and variations thereof, can be used to elucidate other *A. fumigatus* cryptic secondary metabolism BGCs.

Results

Analysis of a potential gene cluster for production of polyketide diterpenoids (PK-DT) in *A. fumigatus*

One of the *A. fumigatus* Af293 SM BGCs with unknown products attracted our interest because it contains a non-reducing polyketide synthase (NR-PKS) gene (Afu8g02350), a prenyltransferase gene (Afu8g02410) and a terpene cyclase (TC) gene (Afu8g02390), suggesting that the SM product of this gene cluster is an unidentified meroterpenoid. As annotated by Inglis *et al.*, this BGC also contains an acetyltransferase gene (Afu8g02360), a flavin-containing monooxygenase (FMO) gene (Afu8g02380), a geranylgeranyl pyrophosphate synthase (GGPPS) gene (Afu8g02400), a hypothetical protein (Afu8g02420), and an alcohol dehydrogenase (Afu8g02430).⁴² A C-4 methyl sterol oxidase (Afu8g02440) is adjacent. In all, the putative BGC is greater than 24 kbp in length. Synteny analysis in FungiDB reveals that the genes within the putative BGC are conserved and collinear between *A. fumigatus* isolates Af293 (Afu8g02350–Afu8g02440) and A1163 (AFUB_084240–AFUB_084150).⁴³ Bioinformatic analysis of this BGC using BLASTp showed that genes Afu8g02350–Afu8g02410 are homologous to genes in the *sre* cluster, which encodes the biosynthetic pathway for sartorypyrone A and D in *Aspergillus felis* 0260, and the *cle* cluster, which encodes the biosynthetic pathway for chevalone E in *Aspergillus versicolor* 0312 (Table 1).⁴⁴

Heterologous expression strategy of the Afu8g02350 BGC in *A. nidulans*

Our heterologous expression platform has two major elements. One is the use of an updated genetic dereplication strain we have developed, LO11098, and related strains for expression. We have engineered this strain for heterologous SM expression by removing eight of the most highly expressed BGCs which lowers the SM background, making expressed metabolites easier to identify and purify. We have also incorporated seven selectable markers in the strain to facilitate multiple sequential gene transfers. The strain also carries a deletion of the *nkua*



Table 1 Putative function of genes within the *A. fumigatus spy* cluster (the percentages represent the aligned regions in BLASTp searches)

<i>A. fumigatus spy</i> cluster	<i>A. felis sre</i> cluster	Similarity/identity (%)	<i>A. versicolor cle</i> cluster	Similarity/identity (%)	Putative function
Afu8g02350 (<i>spyA</i>)	<i>sre6</i>	89/82	<i>cle1</i>	59/43	Polyketide synthase
Afu8g02360 (<i>spyB</i>)	<i>sre5</i>	86/79	—	—	Acetyltransferase
Afu8g02380 (<i>spyC</i>)	<i>sre4</i>	90/83	<i>cle3</i>	73/59	FAD-dependent monooxygenase
Afu8g02390 (<i>spyD</i>)	<i>sre3</i>	77/63	<i>cle7</i>	56/37	Terpene cyclase
Afu8g02400 (<i>spyE</i>)	<i>sre2</i>	91/87	<i>cle6</i>	81/70	Geranylgeranyl pyrophosphate synthase
Afu8g02410 (<i>spyF</i>)	<i>sre1</i>	83/76	<i>cle5</i>	68/57	Prenyltransferase
Afu8g02420	—	—	—	—	Hypothetical protein
Afu8g02430	—	—	—	—	Alcohol dehydrogenase
Afu8g02440	—	—	—	—	C-4 methyl sterol oxidase
No homolog	—	—	<i>cle2</i>	—	P450 monooxygenase
No homolog	—	—	<i>cle4</i>	—	P450 monooxygenase

gene which dramatically reduces non-homologous recombination. The second element is an elaboration of multiplex fusion PCR procedures we have developed⁴⁵ to rapidly create large linear molecules for transformation with multiple genes under control of inducible promoters.

To express our target BGC we used two approaches. In the first approach, we refactored the entire target BGC in *A. nidulans*, placing each gene under control of the inducible *alcA* or *aldA* promoter (Fig. 1A, B and S1†). The *alcA* and *aldA* promoters are short (300–400 bp), strongly induced by a variety of alcohols, aldehydes and ketones, and repressed by glucose and certain other carbon sources.^{46–48} Their shortness and relatively low homology reduce the probability of recombination of these promoters with each other or the native *alcA* or *aldA* loci during transformation. Our second approach was to reconstruct the entire BGC intact in *A. nidulans* and use global regulators of secondary metabolism to activate expression of the BGC. The BGC as predicted by Inglis *et al.* consisted of Afu8g2350–Afu8g02430.⁴² We also included Afu8g02440 in case the BGC was larger than predicted (Fig. 1A).

Heterologous expression of the Afu8g02350 (*spy*) BGC in *A. nidulans* reveals that it produces sartorypyrones

A key in correctly refactoring BGCs is identification of the correct start codon for each gene. BLASTp searches with the amino acid sequence of the putative acetyl transferase, Afu8g02360, as predicted from the FungiDB annotation (<https://fungidb.org/fungidb/app>),⁴³ suggested that the annotation of this gene might be incorrect, and the correct start site might be 593 base pairs downstream. The second start site also had a better Kozak consensus sequence. We, therefore, refactored this gene in two ways inserting the *alcA* promoter upstream of the FungiDB annotated start codon (start site 1) and, separately, in front of the downstream putative start codon (start site 2).

The putative BGC genes were refactored in two steps. Five genes were refactored at the *yA* locus (Fig. 1B). Three transforming fragments were first created by fusion PCR. A fragment carrying ~1 kb of DNA flanking the *yA* gene as well as the *pyrG* gene from *A. fumigatus* (*AfpyrG*) was fused to the *alcA* promoter [*alcA*(p)] and a portion of the Afu8g02350 gene from *A.*

fumigatus. A second fragment constructed by fusion PCR contained an overlapping region of Afu8g02350 as well as two additional genes under control of the *aldA* promoter [*aldA*(p)] and *alcA*(p) and a portion of the *Aspergillus terreus pabaA* gene (*AtpabaA*), which was used as a selectable marker. The third fragment contained an overlapping fragment of *AtpabaA*, two additional genes under control of *alcA*(p) and *aldA*(p) and ~1 kb of *yA* 3' flank. Upon transformation with the three fragments, homologous recombination resulted in the replacement of the *yA* locus with five genes, each under control of *alcA*(p) or *aldA*(p). Note that a functional copy of *AtpabaA* is not formed unless fragments 2 and 3 recombine correctly. Four additional genes were placed under control of *alcA* or *aldA* promoters and inserted at the *wA* locus using essentially the same procedure (Fig. S1†). Using fusion PCR to make transforming fragments, we were, thus, able to refactor the entire >24 kb BGC in two transformations.

Refactored strains were cultivated in lactose minimal medium (LMM), which is non-repressing for the *alcA* and *aldA* promoters and induced with methyl-ethyl-ketone (MEK). Extracts from the culture medium and mycelia were analyzed separately by LC/MS for the presence of new metabolites. In comparison with the *A. nidulans* control strain, which lacked the *A. fumigatus* BGC genes, compounds 1–4 were detected specifically in the refactored strains (Fig. 1C). The strains in which Afu8g02360 was refactored by fusing *alcA*(p) at start site 1 (LO11784–LO11793), were not able to produce compounds 3 and 4, however, in comparison with the start site 2 refactored strains (LO11794–LO11803) (Fig. S2†). This indicated to us that the FungiDB annotation of the acetyl transferase is incorrect. Refactoring Afu8g02360 at start site 2 results in a functional acetyl transferase and production of 3 and 4, the acetylated final products of the BGC. Data with deletion strains (below) support this conclusion.

Yields of compounds from the initial strains were inadequate to allow determination of the structures of 1–4. To increase yields, we deleted the *A. nidulans agsB* gene. *agsB* encodes α -1,3-glucan synthase and *agsB* deletants cause hyphae to disperse in liquid culture.⁴⁹ We hypothesized that deleting *agsB* would allow denser growth and increased metabolite yields, and this proved to be the case. An *agsB* Δ , Afu8g02360



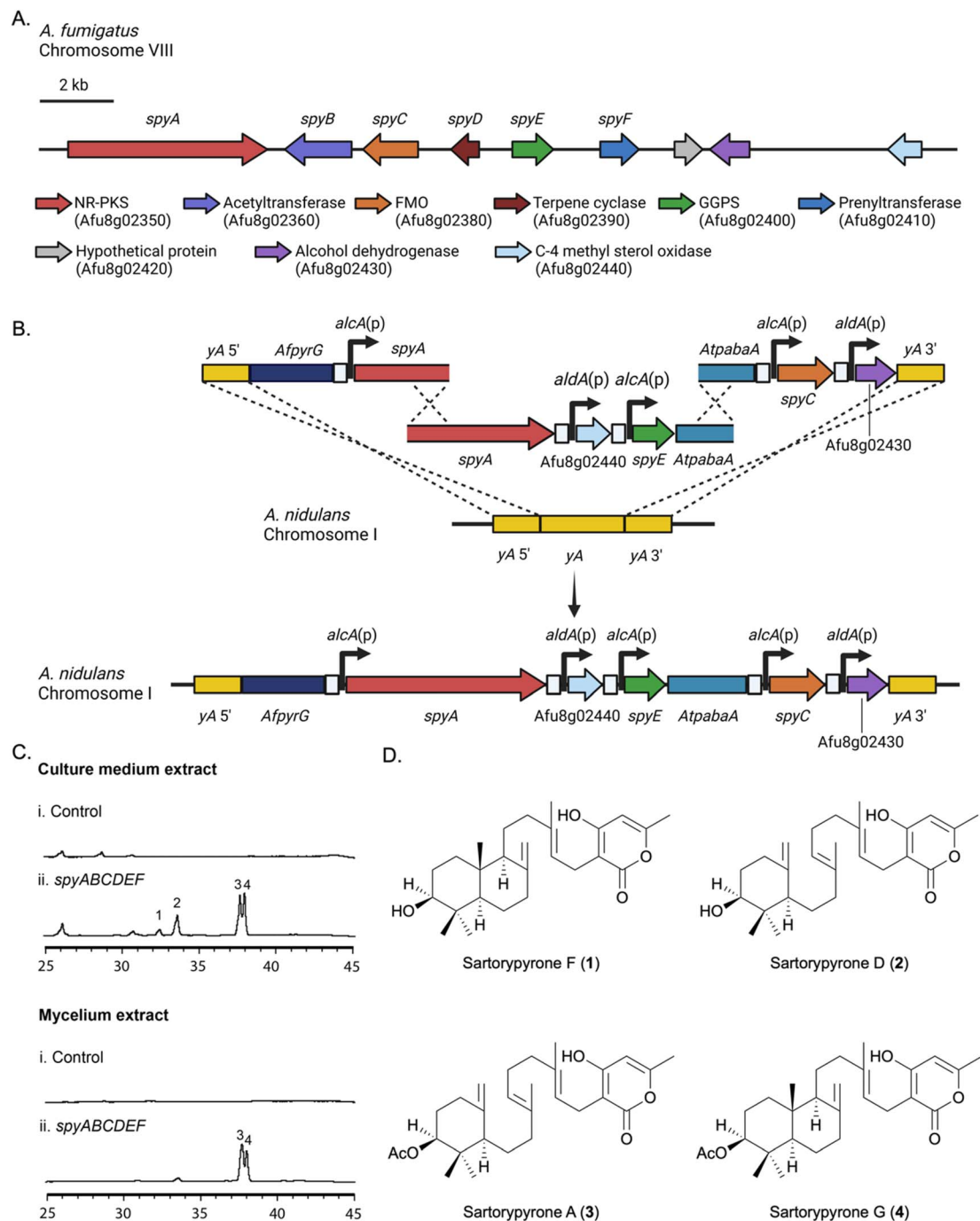


Fig. 1 (A) The *spy* BGC from *A. fumigatus*. (B) Refactoring the *spyA*, *spyE*, *spyC*, *Afu8g02430* and *Afu8g02440* genes at the *yA* locus in *A. nidulans*. (C) HPLC profiles of culture medium and mycelium extracts from *A. nidulans* transformants. (D) Structures of compounds 1–4. *Afu8g02440* (putative C-4 methyl sterol oxidase) and *Afu8g02430* (putative alcohol dehydrogenase) were also refactored at *yA* as shown, but deletion experiments proved them not to be components of the *spy* BGC.

start site 2 refactored strain (LO11839) was subjected to large-scale cultivation and compounds 1–4 were isolated by flash chromatography and semi-preparative HPLC. Spectroscopic analysis (NMR, MS, and UV-Vis data) allowed us to determine that compounds 2 and 3 are sartorypyrone D and sartorypyrone A, respectively, which were previously identified in *Neosartorya fischeri* strains KUFC-6344 and FO-5897 (Fig. 1D and Tables S3

and S4†).^{39,40} (Note: *N. fischeri* is a homotypic synonym of *Aspergillus fischeri* and both designations are used in the scientific literature.) Sartorypyrone D has also been produced synthetically in *Aspergillus oryzae* by expressing chevalone E BGC genes from *A. versicolor* along with a terpene cyclase from *A. felis*.⁴⁴ Because it produces sartorypyrones we have designated the *Afu8g02350* SM BGC as the *spy* BGC. Compounds 1 and 4 are



new to science and were structurally elucidated by 1D and 2D NMR spectra (Fig. 1D, for detailed structural elucidation, see ESI† – detailed structural characterizations of new compounds). Furthermore, the relative configuration was confirmed by MicroED analysis on compound **1** (Fig. 2, S53 and Table S13†). Taking the above spectroscopic analyses together, compounds **1** and **4** are new bicyclic meroditerpenoids, which we have named sartorypyrone F and sartorypyrone G, respectively.

In our second heterologous expression approach, we reconstructed the entire target BGC in *A. nidulans* and attempted to use global regulators of secondary metabolism to activate its expression. We amplified portions of the target BGC and used PCR to fuse them to sequences flanking the *yA* (yellow spore color) gene and to selectable markers (*A. terreus pyrG* [*AtpyrG*] and *A. fumigatus pyroA* [*AfpyroA*]) (Fig. S3†). This created two fragments carrying a total of six genes. Afu8g02420 was on both fragments. Upon co-transformation (Fig. S3A–D†), the two fragments recombined homologously with the *yA* flanking sequences and the Afu8g02420 sequence on the two fragments recombined homologously with each other. The result is a replacement of the *yA* coding sequence with six genes from the target cluster. A second transformation with two additional fragments (Fig. S3A and E–G†) resulted in the reconstruction of the entire target BGC in *A. nidulans*. The second transformation also removes the *AfpyroA* selectable marker. Including primers, the entire sequence transferred into *A. nidulans* was 25 832 bp.

To potentially upregulate expression of the BGC in the reconstructed BGC strain, we separately deleted *mcrA*, a negative regulator of secondary metabolism,²⁹ and we placed two positive regulators of secondary metabolism, *laeA* and *llmG* under control of the inducible *alca* promoter [*alca*(p)*laeA* and *alca*(p)*llmG*].^{50,51} We also created pairwise combinations of the *mcrA* deletion (*mcrA*Δ), *alca*(p)*laeA* and *alca*(p)*llmG*. Initial metabolite profiles (not shown) revealed that while deletion of *mcrA* upregulated secondary metabolite production, it created a sufficiently high background that identification of products of the target BGC wasn't feasible. Overexpression of *laeA* or *llmG*, however, did not create an excessive background, nor did overexpression of these two positive regulators in tandem. Co-overexpression of *laeA* and *llmG* did not yield strong new peaks that could easily be ascribed to the reconstructed BGC.



Fig. 2 MicroED structure of compound **1**. The relative stereochemistry of compound **1** was confirmed by MicroED.

However, knowing the masses of the sartorypyrones produced by the refactored cluster allowed us to unambiguously detect sartorypyrone A (**3**) and sartorypyrone G (**4**) in the *laeA*, *llmG* overexpressing strain by mass spectrometry in extracted ion chromatogram (EIC) mode (Fig. S3B†), while they were not detectable in control strains. This result provides a useful confirmation that **3** and **4** are the final products of the BGC.

Identification, purification, and structural elucidation of the intermediates and shunt products from single-gene-deleted mutant strains

In order to determine the roles of the individual genes in the BGC, we generated a series of single gene deletion strains (Table S1†) from the completely refactored strain LO11911. LO11911 is LO11839 engineered to give even higher yields by replacing the promoter of the *alcR* gene with the strong constitutive *gpdA* promoter.⁵² The deletion strains were verified by multiple diagnostic PCR reactions. They were cultivated under the same conditions as were used to produce sartorypyrones in the refactored strain (*i.e.*, under inducing conditions) and their SM profiles were examined by LC/MS. The results showed that the final products, **3** and **4**, were not produced in the *spyA–F* single gene deletion strains (Fig. 3). Detectable intermediates or shunt products accumulated, however, in *spyB–E* single gene deletion strains (Fig. 3). The metabolite profiles of mutants missing Afu8g02420, Afu8g02430, or Afu8g02440 did not show significant differences from their parent strain indicating that they were not involved in the biosynthesis of sartorypyrones (Fig. S4†). These data are consistent with transcription data available at FungiDB indicating that these three genes are regulated independently of the *spy* genes.

Deleting the PKS gene, *spyA* (Afu8g02350), which is expected to catalyze the first metabolic step in the pathway, dramatically reduced production of the products of the pathway. However, compounds **3** and **4** were produced in very small amounts (more than a 100-fold reduction relative to the strain with *spyA* present, as determined by EIC), suggesting that an alternative mechanism exists in *A. nidulans* for the production of small amounts of the polyketide produced by *SpyA* (Fig. S5†). Based on their domain structures, two *A. nidulans* PKS genes were candidates for producing the same compound as *SpyA*, AN6448, the PKS in the cichorine BGC and AN12440, the product of which is unknown.⁵³ However, deletion of these genes along with *spyA* left a very small residuum of **3** and **4**. These PKSs, therefore are not responsible for the production of trace amounts of **3** and **4** in the absence of *spyA* (Fig. S5†).

The *spyB* (Afu8g02360) gene is predicted to encode an acetyltransferase, and the mutant missing the *spyB* gene was unable to produce the acetylated compounds **3** and **4**. Instead, the unacetylated derivatives, compounds **2** and **1**, respectively, accumulated in the culture medium and, at lower concentrations, in mycelia. We detected substantially more of compounds **1** and **2** than in the parental strain with the intact refactored BGC (more than a 100-fold increase as determined by EIC). These data confirm that the protein encoded by *spyB* is an acetyl transferase and, as indicated above, that start site 2 is correct.



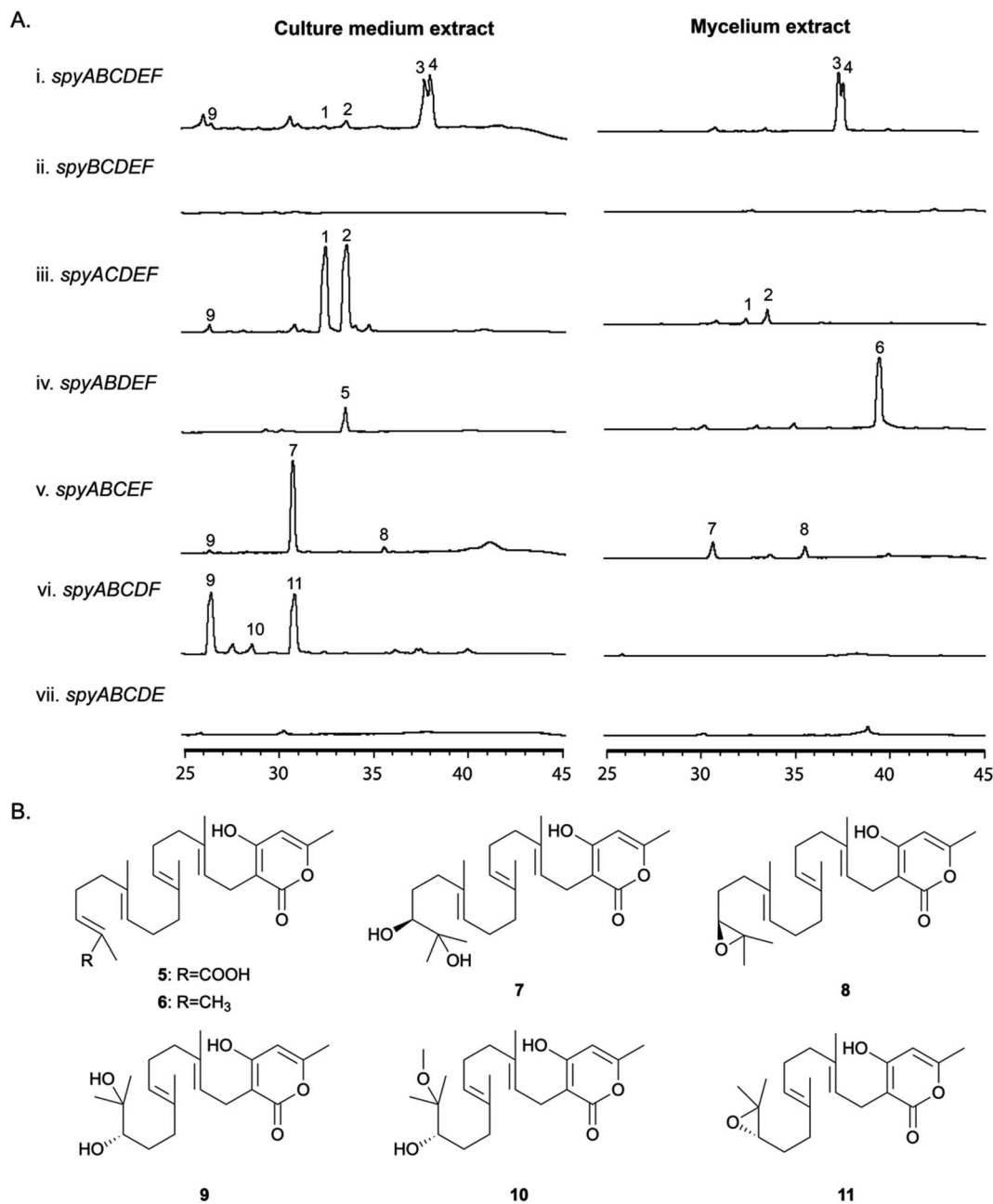


Fig. 3 (A) HPLC profiles of culture medium and mycelium extracts from *A. nidulans* transformants, which carry individual gene deletions from the completely refactored *spy* strain LO11911 (Afu8g02420, Afu8g02430, Afu8g02440 were included). (B) Structures of compounds 5–11.

In the mutant missing the *spyC* (Afu8g02380) gene, we isolated two prenylated polyketides 5 and 6, suggesting that the *spyC* gene codes for an FAD-dependent monooxygenase as the terminal olefin in the diterpene moiety was incapable of epoxidation. The analysis of ¹H, ¹³C, and 2D NMR confirm compound 6 is a known intermediate, geranylgeranyl-triacetate lactone (Table S7[†]),⁴⁴ while compound 5 is a new carboxylate shunt product of 6 (Fig. 3B, for detailed structural elucidation see ESI[†]).

The *spyD* (Afu8g02390) gene is predicted to encode a terpene cyclase. In the mutant lacking the *spyD* gene, we detected two uncyclized prenylated polyketides 7 and 8. The ¹H, ¹³C, and 2D

NMR analysis confirmed compound 7 is sartorypyrone E, which has been previously characterized in the literature (Table S8[†]),⁵⁴ while compound 8 is the epoxide-containing intermediate in the biosynthetic pathway of sartorypyrones (Table S9 and Fig. S33–S37[†]). Although other studies have proposed the hypothetical biosynthetic intermediate structure of compound 8,^{44,55} our study provides the first evidence of its existence as a metabolite of the producing fungus. We named this new compound epoxygeranylgeranyl-triacetate lactone. The epoxide in compound 8 is relatively unstable and thus significantly more of the hydroxylated shunt product compound 7 is generated.





Fig. 4 HPLC profiles of acidified induced culture medium extracts from an *A. nidulans* strain lacking *spyA*, and transformants which carry inducible *spyA* alone or carry the entire inducible *spy* BGC with *spyF* deleted (*spyABCDE*). An authentic TAL standard HPLC profile was included for comparison.

Compound **9** was present at low levels in the culture medium of the strain with the full refactored BGC and in the *spyB* and *spyD* deletants, but it accumulated at much higher levels in the *spyE* (*Afu8g02400*) deletant. **10** and **11** also accumulated in the *spyE* deletant but were not detected by LC/MS in other strains. Large scale isolation and characterization *via* NMR spectroscopy showed that metabolites **9**–**11** were produced by utilizing farnesyl pyrophosphate (FPP) instead of geranylgeranyl pyrophosphate (GGPP) (Table S10–S12 and Fig. S38–S52†), consistent with *spyE* encoding a GGPS that converts FPP to GGPP. The production of these compounds suggests that the prenyl-transferase and FMO have a broad substrate tolerance and can utilize both GGPP and FPP as substrates. Furthermore, the relative configurations of these compounds were deduced to be identical to their analogs with the GGPP moiety because of their biosynthetic relationships. We named these new compounds dihydroxyfarnesyl-triacetate lactone (**9**), 17-methoxy-16-



Fig. 5 Proposed biosynthetic pathway for **3**, **4**, and related shunt products. The pathway bifurcates to **3**–**4** and related shunt products, which are separated by a dashed line. Brackets indicate hypothetical parts of the pathway. The following domains in the NR-PKS SpyA are indicated: SAT: starter unit-ACP transacylase; KS: ketosynthase; AT: acyl transferase; PT: product template; ACP: acyl carrier protein. SpyA is shown with acetyl-CoA conjugated to the SAT domain.



hydroxyfarnesyl-triacetate lactone (**10**) and epoxyfarnesyl-triacetate lactone (**11**).

In the mutant missing the *spyF* (Afu8g02410) gene, which encodes a putative prenyltransferase, all the peaks were abolished, as expected if SpyF is required for prenylation of triacetic acid lactone (TAL) for production of compounds **6** and **11**, but we did not detect the polyketide products. Matsuda *et al.* showed that co-expression in *A. oryzae* of the *A. felis sre3* terpene cyclase with the *A. versicolor* chevalone E BGC genes encoding the PKS, prenyl transferase, geranylgeranyl pyrophosphate synthase and FAD-dependent monooxygenase produced sartorypyrone D. The proposed biosynthetic pathway suggested that the polyketide TAL was the likely product of the chevalone E BGC PKS. TAL was not detected, however, when the chevalone E BGC NR-PKS Cle1 was expressed in *A. oryzae*.⁴⁴ Similarly, we deduced that TAL is the likely polyketide product of the SpyA NR-PKS, because compound **6** is a geranylgeranylated derivative of TAL. Our proposed biosynthetic pathway predicts that TAL should accumulate in the *spyF* deletion strain, but we did not initially detect TAL in this deletant. We therefore constructed an *A. nidulans* strain (LO12091) in which the SpyA NR-PKS, alone, is expressed under control of the inducible *alcA* promoter. Cultivation and direct analysis of the induced culture medium of the *spyA*-expressing strain compared with that of a control strain lacking *spyA* did not initially reveal any polyketide products of SpyA. In order to extract acidic phenolic polyketide compounds, we lowered the pH value of the culture medium filtrate and subsequently extracted the acidified medium by ethyl acetate. Acidification of the culture medium from the *spyA*-expressing strain, but not the control strain lacking *spyA*, revealed a new peak, and we were able to confirm that the peak is TAL (**12**) by comparing it to a TAL standard (Fig. 4). Therefore, SpyA is a TAL synthase. Armed with this information, we acidified culture medium from a *spyF* deletant and were able to detect **12**. The deletant strains in addition to the complete refactored strain allowed us to isolate, and characterize by NMR, all the intermediates in the sartorypyrone biosynthetic pathway (Fig. 5).

Discussion

Secondary metabolites contribute to the reproductive success and pathogenicity of fungi, but many of the secondary metabolites produced by the BGCs of the serious fungal pathogen *A. fumigatus* are unknown. We have used a combination of heterologous expression and natural products chemistry to determine the products of a heretofore cryptic *A. fumigatus* meroterpenoid BGC, which we have named the *spy* BGC. The *spy* BGC harbors homologs of all the genes in the sartorypyrone A (*sre*) BGC in *A. felis*. Several sartorypyrones have been isolated from *Aspergillus* species, but this study is, to our knowledge, the first to show the biosynthesis of this class of compounds from an *A. fumigatus* BGC. The isolation of compounds **1–4** from the refactored strain and the cluster reconstructed strain suggested that a single terpene cyclase in the pathway is able to produce both *trans*-decalin meroterpenoids and monocyclic meroterpenoids. Promiscuous behaviors of terpene cyclases have been reported in other studies.^{56,57}

We created a set of targeted gene deletions that have allowed us to elucidate roles of the *spy* genes in the biosynthesis of compounds **3** and **4**. Cultivation of deletant strains allowed us to identify and fully characterize twelve compounds from the pathway, seven of which have not been reported previously. Based on the intermediates and shunt products isolated from the mutant strains, we were able to propose a biosynthetic pathway for the sartorypyrones produced by the *spy* BGC (Fig. 5). First, compound **12** is generated by the NR-PKS SpyA using one molecule of acetyl-CoA and two molecules of malonyl-CoA. As SpyA lacks a thioesterase (TE) domain, **12** is likely generated through self-release from SpyA by spontaneous lactonization. The prenyltransferase SpyF then conjugates GGPP to **12** to form compound **6**, for which the pathway-specific GGPS SpyE is required to provide GGPP. Subsequently, compound **6** is epoxidized at the terminal olefin by the FMO SpyC, followed by cyclization of the terpenoid component catalyzed by the terpene cyclase SpyD. SpyD exhibits promiscuous activity, resulting in the formation of bicyclic sartorypyrone F (**1**) and monocyclic sartorypyrone D (**2**). While both compounds are generated by direct terminating deprotonation, the degrees of polyene cyclization are different (path a and b). Finally, the last step of the biosynthesis involves the acetylation of the meroterpenoids by the acetyltransferase SpyB to produce compound **3** and compound **4**. In the absence of GGPP (*i.e.* in the mutant lacking *spyE*), SpyF is able to catalyze addition of FPP to **12** to produce shunt products **9–11**. If *spyE* is intact, GGPP is produced, and we hypothesize that SpyF favors the addition of GGPP over addition of FPP such that little of the shunt products accumulate (although a small amount of **9** was detected). In addition, by driving conversion of FPP to GGPP, SpyE may reduce the available levels of FPP, thereby further reducing production of **9–11**. As to why **9** is detected in strains with SpyE but **10** and **11** are not, **11** is likely to be somewhat unstable which may reduce its levels and **11** appears to break down to **9** in preference to **10** such that little **10** is produced even in *spyE* deletants. A very small amount of **3** and **4** are produced in the *spyA* deletant, which indicates that **12** is also produced by an unknown minor mechanism. Note, however, that **12** is one of four products produced when the *A. terreus* PKS gene ATEG_00145.1 is expressed in *A. nidulans*.⁵⁸ The major product of ATEG_00145.1 is a pentaketide, but some triketide (**12**) and tetraketide is produced. It is likely that the PT domain in ATEG_00145.1 that controls product length is not stringent, allowing release of shorter products in addition to the main product. It is possible that an endogenous *A. nidulans* PKS produces a small amount of **12** as an early release product similar to ATEG_00145.1.

Several additional points are worth making. First, we were able to refactor the entire *A. fumigatus* BGC as annotated by Inglis *et al.*,⁴² and an additional gene, in two transformations using transforming fragments created by fusion PCR, and we were easily able to detect metabolites produced by the BGC in our reduced SM background *A. nidulans* genetic dereplication strain. This demonstrates the efficiency and practicality of our heterologous expression system for elucidating the products of cryptic BGCs from *A. fumigatus* and potentially from other fungi. Second, in the native BGC, promoters are likely of different strengths, optimized by evolution to produce the final products of the



biosynthetic pathway. In our refactored BGC, all promoters are strong and this likely results in the accumulation of intermediate compounds such as **1** and **2**. This can be advantageous in elucidating the biosynthetic pathway. Furthermore, we demonstrated the feasibility of MicroED for elucidating structures of SMS produced in our heterologous expression system. Our strategy of reconstructing the target BGC in *A. nidulans* and up-regulating by manipulating global regulators of secondary metabolism, while very feasible, was of minor value with the sartorypyrone BGC, only providing a confirmation of the final products of the BGC. However, nearly all of the remaining cryptic BGCs of *A. fumigatus* harbor putative transcription factors that are predicted to drive expression of the genes of the BGCs. Reconstructing *A. fumigatus* BGCs in *A. nidulans* and upregulating the transcription factors of the BGCs could prove to be a very efficient approach toward elucidating the remaining *A. fumigatus* BGCs.

Finally, TAL is a potentially very valuable biorenewable platform chemical, and efforts are underway to produce it efficiently by a number of routes. The most well-known route is *via* type III PKSs, such as 2-pyrone synthase (2-PS), which catalyze decarboxylative Claisen condensation with one acetyl-CoA molecule as the initial unit and two malonyl-CoA molecules as extension units, followed by spontaneous lactonization to generate TAL.^{59,60} Some research has employed heterologous expression of 2-PS in various organisms, such as *Escherichia coli*, *Yarrowia lipolytica*, and *Saccharomyces cerevisiae* to achieve a higher production yield.^{61–64} Alternatively, polyketoacyl-CoA thiolases (PKTs) were shown to be capable of producing TAL using acetyl-CoA as both initial and extension units.⁶⁵ In this study, we have now demonstrated that the NR-PKS SpyA is also able to form TAL. Although our yields of TAL are not large, our system has not been optimized for TAL production, and there are many routes to much higher yields. Unambiguous identification of a type I NR-PKS that makes TAL herein potentially opens the way to more efficient and commercially viable TAL production.

Conclusions

In summary, we have deciphered an unknown SM BGC in the human pathogen *A. fumigatus* using a heterologous expression approach. The *spy* BGC consists of six contiguous genes involved in the biosynthesis of the sartorypyrones. By integrating data obtained from bioinformatic analysis and intermediates or shunt products isolated from the individual gene deletion mutants, we were able to propose a biosynthetic pathway for this family of compounds. Our approach of refactoring the entire gene cluster in the dereplicated *A. nidulans* host system provides us with a straightforward way to dissect the biosynthetic pathway. This work provides an appealing demonstration that the *A. nidulans* heterologous expression platform can be used for the elucidation of cryptic BGCs in *A. fumigatus* and other species.

Data availability

All experimental procedures and characterization data are available in the ESI.† The NMR spectra data are available in the Natural Products Magnetic Resonance Database (NP-MRD).

Author contributions

C. C. C. W. and B. R. O. supervised the project. S. L. and C. E. O. performed all the experiments. S. L. completed data analysis. C. B. J. and R. B. T. contributed to the design of the experiments. S. L., Y. C. and C. L. contributed to the elucidation of the structures. The MicroED data collection and processing was performed by C. G. J. and P. M. S. with the help of H. M. N.; B. R. O. and R. B. T. contributed to data interpretation. The initial manuscript was written by S. L. with the help of R. B. T., C. C. C. W. and B. R. O. and all authors participated in reviewing and editing.

Conflicts of interest

The authors declare no conflicts of interest.

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References

- 1 D. A. Stevens, R. B. Moss, V. P. Kurup, A. P. Knutsen, P. Greenberger, M. A. Judson, D. W. Denning, R. Cramer, A. S. Brody, M. Light, M. Skov, W. Maish and G. Mastella, and participants in the Cystic Fibrosis Foundation Consensus Conference, *Clin. Infect. Dis.*, 2003, **37**, S225–S264.
- 2 K. Pianalto and J. Alspaugh, *J. Fungi*, 2016, **2**, 26.
- 3 F. Bongomin, S. Gago, R. Oladele and D. Denning, *J. Fungi*, 2017, **3**, 57.
- 4 T. R. T. Dagenais and N. P. Keller, *Clin. Microbiol. Rev.*, 2009, **22**, 447–465.
- 5 J.-P. Latgé and G. Chamilos, *Clin. Microbiol. Rev.*, 2019, **33**, e00140.
- 6 WHO fungal priority pathogens list to guide research, development and public health action, <https://www.who.int/publications/i/item/9789240060241>, accessed June 27, 2023.
- 7 J. L. Steenwyk, M. E. Mead, S. L. Knowles, H. A. Raja, C. D. Roberts, O. Bader, J. Houburken, G. H. Goldman, N. H. Oberlies and A. Rokas, *Genetics*, 2020, **216**, 481–497.
- 8 N. Raffa and N. P. Keller, *PLoS Pathog.*, 2019, **15**, e1007606.
- 9 S. M. Leal, S. Roy, C. Varechon, S. deJesus Carrion, H. Clark, M. S. Lopez-Berges, A. diPietro, M. Schrettl, N. Beckmann, B. Redl, H. Haas and E. Pearlman, *PLoS Pathog.*, 2013, **9**, e1003436.
- 10 J. A. Sugui, J. Pardo, Y. C. Chang, K. A. Zarembek, G. Nardone, E. M. Galvez, A. Müllbacher, J. I. Gallin, M. M. Simon and K. J. Kwon-Chung, *Eukaryotic Cell*, 2007, **6**, 1562–1569.



- 11 S. Spikes, R. Xu, C. K. Nguyen, G. Chamilos, D. P. Kontoyiannis, R. H. Jacobson, D. E. Ejzykowicz, L. Y. Chiang, S. G. Filler and G. S. May, *J. Infect. Dis.*, 2008, **197**, 479–486.
- 12 R. A. Cramer, M. P. Gamcsik, R. M. Brooking, L. K. Najvar, W. R. Kirkpatrick, T. F. Patterson, C. J. Balibar, J. R. Graybill, J. R. Perfect, S. N. Abraham and W. J. Steinbach, *Eukaryotic Cell*, 2006, **5**, 972–980.
- 13 N. P. Keller, G. Turner and J. W. Bennett, *Nat. Rev. Microbiol.*, 2005, **3**, 937–947.
- 14 A. E. Barber, T. Sae-Ong, K. Kang, B. Seelbinder, J. Li, G. Walther, G. Panagiotou and O. Kurzai, *Nat. Microbiol.*, 2021, **6**, 1526–1536.
- 15 W. C. Nierman, A. Pain, M. J. Anderson, J. R. Wortman, H. S. Kim, J. Arroyo, M. Berriman, K. Abe, D. B. Archer, C. Bermejo, J. Bennett, P. Bowyer, D. Chen, M. Collins, R. Coulsen, R. Davies, P. S. Dyer, M. Farman, N. Fedorova, N. Fedorova, T. V. Feldblyum, R. Fischer, N. Fosker, A. Fraser, J. L. García, M. J. García, A. Goble, G. H. Goldman, K. Gomi, S. Griffith-Jones, R. Gwilliam, B. Haas, H. Haas, D. Harris, H. Horiuchi, J. Huang, S. Humphray, J. Jiménez, N. Keller, H. Khouri, K. Kitamoto, T. Kobayashi, S. Konzack, R. Kulkarni, T. Kumagai, A. Lafton, J.-P. Latgé, W. Li, A. Lord, C. Lu, W. H. Majoros, G. S. May, B. L. Miller, Y. Mohamoud, M. Molina, M. Monod, I. Mouyna, S. Mulligan, L. Murphy, S. O'Neil, I. Paulsen, M. A. Peñalva, M. Perlea, C. Price, B. L. Pritchard, M. A. Quail, E. Rabinowitsch, N. Rawlins, M.-A. Rajandream, U. Reichard, H. Renauld, G. D. Robson, S. R. de Córdoba, J. M. Rodríguez-Peña, C. M. Ronning, S. Rutter, S. L. Salzberg, M. Sanchez, J. C. Sánchez-Ferrero, D. Saunders, K. Seeger, R. Squares, S. Squares, M. Takeuchi, F. Tekaia, G. Turner, C. R. V. de Aldana, J. Weidman, O. White, J. Woodward, J.-H. Yu, C. Fraser, J. E. Galagan, K. Asai, M. Machida, N. Hall, B. Barrell and D. W. Denning, *Nature*, 2005, **438**, 1151–1156.
- 16 N. D. Fedorova, N. Khaldi, V. S. Joardar, R. Maiti, P. Amedeo, M. J. Anderson, J. Crabtree, J. C. Silva, J. H. Badger, A. Albarraq, S. Angiuoli, H. Bussey, P. Bowyer, P. J. Cotty, P. S. Dyer, A. Egan, K. Galens, C. M. Fraser-Liggett, B. J. Haas, J. M. Inman, R. Kent, S. Lemieux, I. Malavazi, J. Orvis, T. Roemer, C. M. Ronning, J. P. Sundaram, G. Sutton, G. Turner, J. C. Venter, O. R. White, B. R. Whitty, P. Youngman, K. H. Wolfe, G. H. Goldman, J. R. Wortman, B. Jiang, D. W. Denning and W. C. Nierman, *PLoS Genet.*, 2008, **4**, e1000046.
- 17 J. Romsdahl and C. C. C. Wang, *Med. Chem. Commun.*, 2019, **10**, 840–866.
- 18 M. C. Stroe, T. Netzker, K. Scherlach, T. Krüger, C. Hertweck, V. Valiante and A. A. Brakhage, *eLife*, 2020, **9**, e52541.
- 19 D. Kalb, T. Heinekamp, G. Lackner, D. H. Scharf, H.-M. Dahse, A. A. Brakhage and D. Hoffmeister, *Appl. Environ. Microbiol.*, 2015, **81**, 1594–1600.
- 20 J. W. Bok, Y.-M. Chiang, E. Szweczyk, Y. Reyes-Dominguez, A. D. Davidson, J. F. Sanchez, H.-C. Lo, K. Watanabe, J. Strauss, B. R. Oakley, C. C. C. Wang and N. P. Keller, *Nat. Chem. Biol.*, 2009, **5**, 462–464.
- 21 A. A. Brakhage and V. Schroeckh, *Fungal Genet. Biol.*, 2011, **48**, 15–22.
- 22 Y.-M. Chiang, S.-L. Chang, B. R. Oakley and C. C. Wang, *Curr. Opin. Chem. Biol.*, 2011, **15**, 137–143.
- 23 M. Ahuja, Y.-M. Chiang, S.-L. Chang, M. B. Praseuth, R. Entwistle, J. F. Sanchez, H.-C. Lo, H.-H. Yeh, B. R. Oakley and C. C. C. Wang, *J. Am. Chem. Soc.*, 2012, **134**, 8212–8221.
- 24 A. A. Soukup, Y.-M. Chiang, J. W. Bok, Y. Reyes-Dominguez, B. R. Oakley, C. C. C. Wang, J. Strauss and N. P. Keller, *Mol. Microbiol.*, 2012, **86**, 314–330.
- 25 J. Yaegashi, B. R. Oakley and C. C. C. Wang, *J. Ind. Microbiol. Biotechnol.*, 2014, **41**, 433–442.
- 26 Y.-M. Chiang, M. Ahuja, C. E. Oakley, R. Entwistle, C. Zutz, C. C. C. Wang and B. R. Oakley, *Angew. Chem., Int. Ed.*, 2016, **55**, 1662–1665.
- 27 H.-H. Yeh, M. Ahuja, Y.-M. Chiang, C. E. Oakley, S. Moore, O. Yoon, H. Hajovsky, J.-W. Bok, N. P. Keller, C. C. C. Wang and B. R. Oakley, *ACS Chem. Biol.*, 2016, **11**, 2275–2284.
- 28 J. Macheleidt, D. J. Mattern, J. Fischer, T. Netzker, J. Weber, V. Schroeckh, V. Valiante and A. A. Brakhage, *Annu. Rev. Genet.*, 2016, **50**, 371–392.
- 29 C. E. Oakley, M. Ahuja, W.-W. Sun, R. Entwistle, T. Akashi, J. Yaegashi, C.-J. Guo, G. C. Cerqueira, J. Russo Wortman, C. C. C. Wang, Y.-M. Chiang and B. R. Oakley, *Mol. Microbiol.*, 2017, **103**, 347–365.
- 30 M. F. Grau, R. Entwistle, Y.-M. Chiang, M. Ahuja, C. E. Oakley, T. Akashi, C. C. C. Wang, R. B. Todd and B. R. Oakley, *ACS Chem. Biol.*, 2018, **13**, 3193–3205.
- 31 L. K. Caesar, N. L. Kelleher and N. P. Keller, *Fungal Genet. Biol.*, 2020, **144**, 103477.
- 32 K. A. O'Hanlon, L. Gallagher, M. Schrettl, C. Jöchl, K. Kavanagh, T. O. Larsen and S. Doyle, *Appl. Environ. Microbiol.*, 2012, **78**, 3166–3176.
- 33 I. A. Unsöld and S.-M. Li, *ChemBioChem*, 2006, **7**, 158–164.
- 34 P. Wiemann, B. E. Lechner, J. A. Baccile, T. A. Velk, W.-B. Yin, J. W. Bok, S. Pakala, L. Losada, W. C. Nierman, F. C. Schroeder, H. Haas and N. P. Keller, *Front. Microbiol.*, 2014, **5**, 530.
- 35 B. D. Ames, X. Liu and C. T. Walsh, *Biochemistry*, 2010, **49**, 8564–8576.
- 36 B. D. Ames and C. T. Walsh, *Biochemistry*, 2010, **49**, 3351–3365.
- 37 K. Throckmorton, F. Y. Lim, D. P. Kontoyiannis, W. Zheng and N. P. Keller, *Environ. Microbiol.*, 2016, **18**, 246–259.
- 38 A. Blachowicz, N. Raffa, J. W. Bok, T. Choera, B. Knox, F. Y. Lim, A. Huttenlocher, C. C. C. Wang, K. Venkateswaran and N. P. Keller, *mBio*, 2020, **11**, e03415–e03419.
- 39 A. Eamvijarn, N. M. Gomes, T. Dethoup, J. Buaruang, L. Manoch, A. Silva, M. Pedro, I. Marini, V. Roussis and A. Kijjoa, *Tetrahedron*, 2013, **69**, 8583–8591.
- 40 S. Kaifuchi, M. Mori, K. Nonaka, R. Masuma, S. Ōmura and K. Shiomi, *J. Antibiot.*, 2015, **68**, 403–405.
- 41 M. Chia, T. J. Schwartz, B. H. Shanks and J. A. Dumesic, *Green Chem.*, 2012, **14**, 1850–1853.



- 42 D. O. Inglis, J. Binkley, M. S. Skrzypek, M. B. Arnaud, G. C. Cerqueira, P. Shah, F. Wymore, J. R. Wortman and G. Sherlock, *BMC Microbiol.*, 2013, **13**, 91.
- 43 B. Amos, C. Aurrecochea, M. Barba, A. Barreto, E. Y. Basenko, W. Bažant, R. Belnap, A. S. Blevins, U. Böhme, J. Brestelli, B. P. Brunk, M. Caddick, D. Callan, L. Campbell, M. B. Christensen, G. K. Christophides, K. Crouch, K. Davis, J. DeBarry, R. Doherty, Y. Duan, M. Dunn, D. Falke, S. Fisher, P. Flicek, B. Fox, B. Gajria, G. I. Giraldo-Calderón, O. S. Harb, E. Harper, C. Hertz-Fowler, M. J. Hickman, C. Howington, S. Hu, J. Humphrey, J. Iodice, A. Jones, J. Judkins, S. A. Kelly, J. C. Kissinger, D. K. Kwon, K. Lamoureux, D. Lawson, W. Li, K. Lies, D. Lodha, J. Long, R. M. MacCallum, G. Maslen, M. A. McDowell, J. Nabrzyski, D. S. Roos, S. S. C. Rund, S. W. Schulman, A. Shanmugasundram, V. Sitnik, D. Spruill, D. Starns, C. J. Stoeckert, S. S. Tomko, H. Wang, S. Warrenfeltz, R. Wieck, P. A. Wilkinson, L. Xu and J. Zheng, *Nucleic Acids Res.*, 2022, **50**, D898–D911.
- 44 W.-G. Wang, L.-Q. Du, S.-L. Sheng, A. Li, Y.-P. Li, G.-G. Cheng, G.-P. Li, G. Sun, Q.-F. Hu and Y. Matsuda, *Org. Chem. Front.*, 2019, **6**, 571–578.
- 45 E. Szewczyk, T. Nayak, C. E. Oakley, H. Edgerton, Y. Xiong, N. Taheri-Talesh, S. A. Osmani and B. R. Oakley, *Nat. Protoc.*, 2006, **1**, 3111–3120.
- 46 M. Flipphi, M. Mathieu, I. Cirpus, C. Panozzo and B. Felenbok, *J. Biol. Chem.*, 2001, **276**, 6950–6958.
- 47 R. B. Waring, G. S. May and N. R. Morris, *Gene*, 1989, **79**, 119–130.
- 48 B. Felenbok, *J. Biotechnol.*, 1991, **17**, 11–17.
- 49 A. Yoshimi, M. Sano, A. Inaba, Y. Kokubun, T. Fujioka, O. Mizutani, D. Hagiwara, T. Fujikawa, M. Nishimura, S. Yano, S. Kasahara, K. Shimizu, M. Yamaguchi, K. Kawakami and K. Abe, *PLoS One*, 2013, **8**, e54893.
- 50 N. Keller, J. Bok, D. Chung, R. M. Perrin and E. Keats Shwab, *Med. Mycol.*, 2006, **44**, 83–85.
- 51 M. F. Grau, R. Entwistle, C. E. Oakley, C. C. C. Wang and B. R. Oakley, *ACS Chem. Biol.*, 2019, **14**, 1643–1651.
- 52 P. J. Punt, N. D. Zegers, M. Busscher, P. H. Pouwels and C. A. M. J. J. van den Hondel, *J. Biotechnol.*, 1991, **17**, 19–33.
- 53 J. F. Sanchez, R. Entwistle, D. Corcoran, B. R. Oakley and C. C. C. Wang, *Med. Chem. Commun.*, 2012, **3**, 997–1002.
- 54 S. Bang, J. H. Song, D. Lee, C. Lee, S. Kim, K. S. Kang, J. H. Lee and S. H. Shim, *J. Agric. Food Chem.*, 2019, **67**, 1831–1838.
- 55 K. Kanokmedhakul, S. Kanokmedhakul, R. Suwannatrai, K. Soyong, S. Prabpai and P. Kongsaree, *Tetrahedron*, 2011, **67**, 5461–5468.
- 56 M.-C. Tang, H.-C. Lin, D. Li, Y. Zou, J. Li, W. Xu, R. A. Cacho, M. E. Hillenmeyer, N. K. Garg and Y. Tang, *J. Am. Chem. Soc.*, 2015, **137**, 13724–13727.
- 57 L. Barra and I. Abe, *Nat. Prod. Rep.*, 2021, **38**, 566–585.
- 58 Y.-M. Chiang, C. E. Oakley, M. Ahuja, R. Entwistle, A. Schultz, S.-L. Chang, C. T. Sung, C. C. C. Wang and B. R. Oakley, *J. Am. Chem. Soc.*, 2013, **135**, 7720–7731.
- 59 S. Eckermann, G. Schröder, J. Schmidt, D. Strack, R. A. Edrada, Y. Helariutta, P. Elomaa, M. Kotilainen, I. Kilpeläinen, P. Proksch, T. H. Teeri and J. Schröder, *Nature*, 1998, **396**, 387–390.
- 60 L. Zhu, M. Pietiäinen, J. Kontturi, A. Turkkelin, P. Elomaa and T. H. Teeri, *New Phytol.*, 2022, **236**, 296–308.
- 61 Y. Li, S. Qian, R. Dunn and P. C. Cirino, *J. Ind. Microbiol. Biotechnol.*, 2018, **45**, 789–793.
- 62 J. Yu, J. Landberg, F. Shavarebi, V. Bilanchone, A. Okerlund, U. Wanninayake, L. Zhao, G. Kraus and S. Sandmeyer, *Biotechnol. Bioeng.*, 2018, **115**, 2383–2388.
- 63 K. A. Markham, C. M. Palmer, M. Chwatko, J. M. Wagner, C. Murray, S. Vazquez, A. Swaminathan, I. Chakravarty, N. A. Lynd and H. S. Alper, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, 2096–2101.
- 64 J. Cardenas and N. A. Da Silva, *Metab. Eng.*, 2014, **25**, 194–203.
- 65 Z. Tan, J. M. Clomburg, S. Cheong, S. Qian and R. Gonzalez, *Nat. Catal.*, 2020, **3**, 593–603.

