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Synthetic hyaluronic acid coating preserves the phenotypes of lymphatic endothelial cells†

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Lymphatic endothelial cells (LECs) play a critical role in the formation and maintenance of the lymphatic vasculature, which is essential for the immune system, fluid balance, and tissue repair. However, LECs are often difficult to study *in vivo* and *in vitro* models that accurately mimic their behaviors and phenotypes are limited. In particular, LECs have been shown to lose their lymphatic markers over time while being cultured *in vitro*, which reflect their plasticity and heterogeneity *in vivo*. Since LECs uniquely express lymphatic vessel endothelial hyaluronan receptor-1 (*LYVE-1*), we hypothesized that surface coating with hyaluronic acid (HA) can preserve LEC phenotypes and functionalities. Dopamine conjugated hyaluronic acid (HA-DP) was synthesized with 42% degree of substitution to enable surface modification and conjugation onto standard tissue culture plates. Compared to fibronectin coating and tissue culture plate controls, surface coating with HA-DP was able to preserve lymphatic markers, such as prospero homeobox protein 1 (*Prox1*), podoplanin (*PDPN*), and *LYVE-1* over several passages *in vitro*. LECs cultured on HA-DP expressed lower levels of focal adhesion kinase (FAK) and YAP/TAZ, which may be responsible for the maintenance of the lymphatic characteristics. Collectively, the HA-DP coating may provide a novel method for culturing human LECs *in vitro* toward more representative studies in basic lymphatic biology and lymphatic regeneration.

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Introduction

Lymphatic endothelial cells (LECs) play a vital role in the immune system, serving as the gatekeepers for lymphocyte trafficking and the maintenance of immune homeostasis.^{1,2} They are also important for the removal of interstitial fluid and waste products from tissues.^{3,4} In a variety of diseases, such as cancer and chronic inflammation, the integrity of LECs is disrupted, leading to lymphatic dysfunction and immune suppression.^{5,6} To understand many of these mechanisms and pathophysiology, it is imperative to have a reliable *in vitro* culture system that can preserve the phenotype and characteristics of LECs. While prospero homeobox protein (*Prox1*) is considered as the master regulator to maintain lymphatic identity,^{7,8} other lymphatic markers are differentially expressed *in vivo*. Lymphatic capillaries express high levels of lymphatic vessel endothelial hyaluronan receptor 1 (*LYVE-1*).

Pre-collecting and collecting vessels express low level of *LYVE-1*, but high level of podoplanin.^{9,10} Hence, it is important to have a culture system that recapitulate the lymphatic phenotypes and heterogeneity observed *in vivo*.

Another important aspect of having a robust *in vitro* culture system for LECs is tissue engineering. Nowadays there have been tremendous efforts to generate lymphatic vessels using natural and synthetic materials.^{10–13} Often it is not realized that just having a more complex and physiologically relevant 3D system will not result in better tissue engineering unless rudimentary 2D culture is optimized as well. Therefore, it is important to be able to culture LECs on a coating that supports their proliferation, survival, and functional activity over extended periods of time. For decades, the isolation and growth of cells *in vitro* under controlled conditions has been one of the most utilized experimental approaches in the field of cell biology.¹⁴ As a result, various extra cellular matrix (ECM) coatings both natural and synthetic have been established.¹⁵

Given the plasticity and heterogeneity of LECs, various ECM coatings have been used to culture LECs.¹⁰ Currently, LECs are cultured on tissue culture plastics or on conventional coatings like fibronectin or collagen. Fibronectin is one of the most used ECMs for *in vitro* culture;^{16–22} it was shown that LECs adhered and proliferated differently on a fibronectin coated

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plate compared to tissue culture plastic.¹⁹ Previous study reveals that the ligand for integrin $\alpha_5\beta_1$, selectively promoted the growth of LECs through vascular endothelial growth factor receptor-3 (VEGFR-3), which is one of the transmembrane receptors responsible for lymphatic endothelial migration, survival and proliferation.^{23–25} Another recent study also shows that the adhesion and migration of LECs stimulated by vascular endothelial growth factor-C (VEGF-C) or VEGF-D are $\alpha_9\beta_1$ -dependent, which is present in some mutant version of fibronectin.^{21,26–28} Similarly, Collagen-I and Collagen-IV have been used in some studies for culturing LECs as a monolayer or as an embryoid body (EB) culture.^{22,29} Though it was observed that Collagen-IV induces migration, cell alignment, proliferation, and differentiation into mature lymphatic capillaries *in vivo*,³⁰ another study found that Collagen-IV did not favor LEC differentiation *in vitro*.³¹ Collagen-I has been used for monolayer culture^{17,22,30,32,33} of primary LECs and supporting lymphatic vessel like structure.³⁴ Laminin, among the natural ECMs, is the least utilized coating for LECs. It is reported that α_4 -laminins, such as 411 (formerly laminin-8), 421 (formerly laminin-9), and 423 (formerly laminin-14), are expressed by vascular and lymphatic endothelial cells.^{29,35,36} Overall, the outlook on using collagen or laminin as coating for culturing LEC is still in contention, while fibronectin is considered as a benchmark in this regard.

Compared to ECM proteins, the use of hyaluronic acid or hyaluronan (HA) has been shown to hold promise in culture of blood endothelial cells (ECs).^{37–39} HA is an abundant component of the ECM that binds to various receptors and influences activities of ECs. Low molecular weight (LMW) HA was proved to have the ability to interact with its receptors, such as CD44 or receptor for hyaluronan-mediated motility (RHAMM), triggering series of intracellular signal transduction and promoting angiogenesis.^{40–43} Although CD44 and RHAMM are reported as the main receptors on vascular ECs, they are mostly absent from lymphatic vessels; wherein the only known receptor for HA is LYVE-1, a homolog of CD44.^{44,45} LYVE-1 is thus likely to play a major role in the regulation of HA on biological behaviors of LECs. Indeed, it was shown in several studies that LMW HA induces lymphangiogenesis through LYVE-1 mediated signaling pathways.^{46–48} One recent study also shows that HA-binding peptide modulates EC spreading and migration through focal adhesion kinase (FAK).⁴⁹ Despite the unique expression of HA receptors by LECs, the effect of HA on the regulation of lymphatic characteristic markers has not been explored yet.

In this work, we describe the development of a novel synthetic coating based on HA to preserve the phenotypes and characteristics of LECs. We synthesized dopamine-conjugated HA (HA-DP), which can be conjugated onto the surface of tissue culture plates. Compared to other conventional ECM based coating with fibronectin, we demonstrated that HA-DP can preserve lymphatic phenotypes over several passages of LECs culture *in vitro*. Moreover, LECs cultured on HA-DP exhibited reduced FAK, which may be responsible for the maintenance of the lymphatic characteristics.

Experimental

Synthesis of dopamine-conjugated hyaluronic acid (HA-DP)

HA (1.026 g, 2.56 mmol disaccharide unit) was dissolved in 85.5 mL MES buffer (0.1 M, pH 5.5) and 28.5 mL ethanol was added. The reaction mixture was brought to equilibrium at room temperature, DMTMM (2.9 g, 4 equivalents) was added to activate HA at room temperature. After 30 minutes of incubation, dopamine (484.7 mg, 1 equivalent) was added to the reaction mixture, followed by stirring at room temperature overnight. HA-DP was purified by dialysis against deionized water at 4 °C for 4 days, lyophilized, and stored at –20 °C until use. HA-DP was characterized by ¹H NMR in D₂O showing peaks at 1.99 ppm (C=O)CH₃ in HA and 7.3–7.4 (aromatic protons in DP). The degree of substitution was calculated as 42%.

Coating of hyaluronic acid-dopamine (HA-DP)

HA-DP was coated on tissue culture plates through polymerization of dopamine under basic conditions. HA-DP was dissolved in deionized water at 5 mg mL⁻¹. Prior adding HA-DP to wells for coating, 20 μ L of 10 M NaOH was added per 1 mL of HA-DP. After incubating at 37 °C overnight, the wells were vigorously washed with deionized water and cell culture medium for cell culture.

Characterization of HA coating

HA-DP coating was characterized and quantified using toluidine blue assay, as previously described.⁵⁰ Briefly, serial dilution of free HA in DI water was prepared to create a standard curve. Then, both free HA solution and HA-DP coated plate were incubated with 1 mL Toluidine blue O (TBO, Sigma 198161-5G). In the case of free HA solution, supernatant was washed off carefully after centrifugation. For coated six-wells plate, the supernatant was aspirated directly. Both samples were washed properly with 10 mM NaOH. At this stage, a picture was taken to show TBO staining of coated wells compared to non-coated ones. Then, 50% acetic was added to the samples and incubated at room temperature for 20 minutes. Finally, the solutions were collected and ran for absorbance in microplate reader at 634 nm. Absorbance of free HA solutions of different concentration were used to create standard curve, then absorbance of HA-DP coated sample was fit to the standard curve to quantify amount of HA in the coating.

Cell culture

Human juvenile lymphatic endothelial cells (C-12216) of four donors (PromoCell, Heidelberg, Germany) were expanded and used for experiments between passages 4 and 8, as previously described.^{15,51} Briefly, LECs were maintained at 37 °C with 5% CO₂ in Endothelial Cell Growth Medium (EGM-MV2, C-22022, PromoCell). To keep the cell passaging constant throughout experiments, cells were passaged every 5 days at a 1 to 3 ratio. Human LECs were characterized for the positive expression of CD31, LYVE-1, Prox1, and podoplanin throughout the experiments. All cell lines were routinely tested for myco-



plasma contamination and were negative throughout this study.

FACS analysis

Human LECs were analyzed for lymphatic markers using flow cytometry (FACS) following standard procedure.⁵² Briefly, cells were trypsinized and centrifuged following resuspension in FACS buffer. Suspended cells (1×10^6 cells) were stained with the antibodies ($1 \mu\text{g mL}^{-1}$) for 30 minutes at room temperature: anti-LYVE-1 antibody (R&D systems, FAB20892A), anti-PDPN antibody APC (Biolegend, 337004), as well as their corresponding IgG isotype controls (ESI Table 1†). For intracellular staining, the cells were fixed and permeabilized with Foxp3/Transcription Factor Staining Buffer Set (Thermo, 00-5523-00) and then incubated with anti-Prox1 antibody FITC (Novus Biologicals, NBP1-30045AF488) for 30 minutes. The cells were washed twice and resuspended in FACS buffer for analysis. Then, the cells were analyzed using flow cytometry (BD LSR FortessaX-20) and the metadata were analyzed using FlowJo.

Gene expression

To analyze the effect of different ECM on lymphatic phenotypes, LECs were cultured on tissue culture plastic, fibronectin, and HA-DP coatings for 5 days in EGM-MV2 media. Three biological replicates ($n = 3$) were collected per condition and analyzed with real-time qRT-PCR with triplicate readings as previously described.¹⁵ RNA was reverse transcribed using a high-capacity cDNA reverse transcription kit (Thermo Fisher) according to the manufacturer's protocol. cDNA was then used with the TaqMan Universal PCR Master Mix and Gene

Expression Assays for *LYVE-1*, *Prox1*, *PDPN*, *VEGFR3*, *YAP*, *TAZ*, *MYC*, *CTGF*, and *GAPDH* (ESI Table 2†). Each sample was prepared in triplicate and the relative expression was normalized to GAPDH and analyzed using the $\Delta\Delta\text{Ct}$ method.

Immunofluorescence

To visualize the lymphatic protein expression, LECs were seeded on tissue culture plastic, fibronectin, and HA-DP coatings for 5 days. Samples were fixed with 4% paraformaldehyde, blocked with 1% BSA, permeabilized with 0.1% Triton-X, and stained for LYVE-1, Prox1, CD144, ERG and podoplanin (ESI Table 3†). To visualize focal adhesion kinase and F-actin distribution FAK antibody (Sigma, $2 \mu\text{g mL}^{-1}$) and phalloidin (Abcam, 1 : 1000) were used. Samples were rinsed twice in PBS and counterstained with DAPI (Thermo Fischer, 300 nM). All samples were imaged in Nikon AX-R confocal at 40 \times magnification.

FAK and F-actin quantification

The thresholding tool in ImageJ was used to identify the FAKs. Then analyze particles was used with size restricted to 4–20 μm to count the number of focal adhesions. Ten fluorescent images were taken per coating at 40 \times using the Nikon AXR confocal microscope and analyzed with the FIJI Directionality Plug-in.⁵³ A statistical analysis of the dispersion was performed using GraphPad Prism 9 (GraphPad Software Inc., La Jolla, CA).

Statistical analysis

Statistical analysis was performed with GraphPad Prism. For each coating condition, at least three independent experiments



Fig. 1 Schematic of experimental procedure to investigate cell–ECM interaction on HA–DP coating. The HA–DP polymer was dissolved in DI water, then added to culture plate in the presence of 10 M NaOH. The plates were incubated overnight and washed with DI water before seeding the cells. An enlarged image on top left shows interaction between HA polymer and LYVE-1 receptor, one of the key glycoprotein receptors of LECs. Lymphatic phenotypes were analysed using flow cytometry, immunofluorescent imaging, and real-time RT-PCR.



were performed with three biological replicates. Statistical comparisons were made using Student's *t* test for paired data, analysis of variance (ANOVA) for multiple comparisons, and with Tukey post hoc analysis for parametric data. Specifically, Student's *t* test was used to analyze differences between protein expression and gene expression on different coatings. Significance levels were set at the following: **P* < 0.05, ***P* < 0.01, ****P* < 0.001, *****P* < 0.0001.

Results and discussion

LECs exhibit a decrease in lymphatic markers *in vitro*

While previous studies have reported that blood endothelial cells lose their characteristics over time,^{54,55} little is known about the phenotypes of LECs during *in vitro* culture. Therefore, we aimed to characterize the phenotypes of LECs during *in vitro* culture. Human dermal LECs from single donor

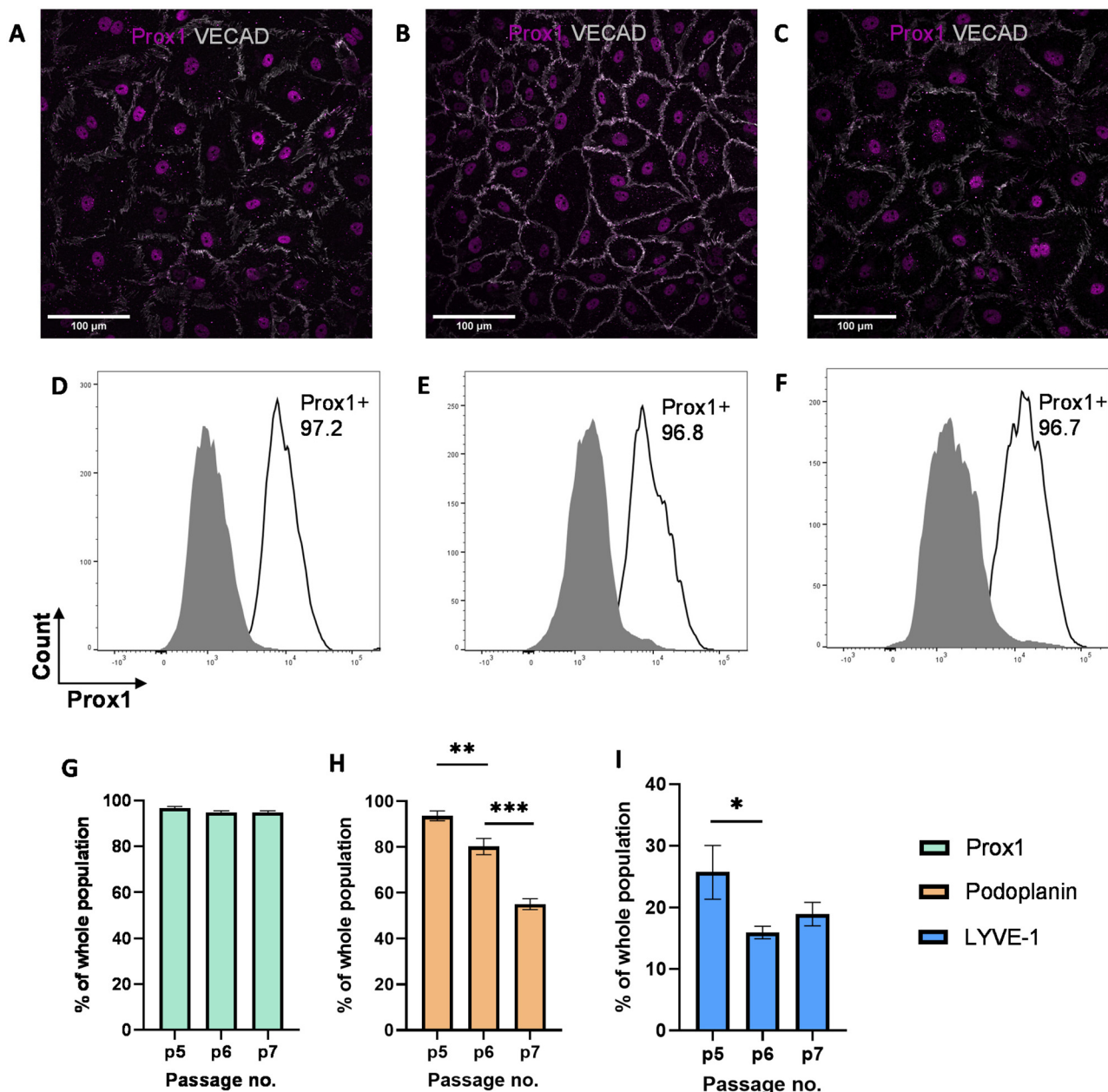


Fig. 2 Expression of lymphatic markers by LECs cultured on tissue culture plates. Representative immunofluorescent images of LECs cultured on tissue culture plates at (A) P.5, (B) P.6, and (C) P.7 stained for VE-CAD (white) and Prox-1 (magenta). Scale bars are 100 μ m. LECs cultured on tissue culture plates were analyzed for Prox-1 using flow cytometry. Representative flow cytometry analysis indicating Prox-1 expression by LECs (white histogram) at (D) P.5, (E) P.6, and (F) P.7 compared to isotype control (grey histogram). Flow cytometry data for LECs at P.5, P.6, and P.7 were analysed for percentage of (G) Prox1⁺, (H) podoplanin⁺, and (I) LYVE-1⁺ cells population. Data represents mean \pm stdev., *n* = 4 per condition. Significance levels were set at: **P* < 0.5, ***P* < 0.01, and ****P* < 0.001.



($n = 3$) were cultured on tissue culture plates from passage 5 to 7 and their lymphatic phenotypes were characterized using FACS, real-time qRT-PCR, and immunofluorescent image analysis (Fig. 1). We confirmed using immunofluorescent (Fig. 2A–C) and FACS analysis (Fig. 2D–F and ESI Fig. 1–3†) that LECs are heterogeneous in nature and consistently express Prox-1 from passage 5 to 7. Since Prox-1 is known to be the master lymphatic regulator,^{56,57} these observations confirmed that LECs maintained their lymphatic identity (Fig. 2G). However, we observed a gradual decrease in the expression of key lymphatic markers podoplanin (Fig. 2H) and LYVE-1 (Fig. 2I). Overall, these results suggest that while LECs maintained their identity in culture by expressing transcription factor Prox1, they started to lose their lymphatic phenotypes by gradual decrease in key lymphatic markers podoplanin and LYVE-1. This observation is consistent with the lymphatic heterogeneity and plasticity that can be found *in vivo*.^{58–60}

Synthesis and characterization of HA-DP

After validating the heterogeneity and plasticity of LECs, we hypothesized that the conventional coating system might not be optimized for culturing LECs. Since LECs uniquely express LYVE-1 to bind to HA and activate intracellular signaling to promote lymphangiogenesis,⁶¹ we postulated that tunable synthetic HA coating can serve as a supportive matrix for LEC culture *in vitro*. Since HA is a negatively charged polysaccharide, we have to modify HA with dopamine group to enable its conjugation to the surface of the tissue culture plate.⁶² The dopamine group was conjugated with the carboxyl group of HA to generate HA-DP polymer (Fig. 3A). ¹H-NMR indicated dopamine peaks at 7.47–7.21 ppm and *N*-acetyl groups of HA peaks at 1.99 ppm, which confirmed the conjugation of dopamine group to HA (Fig. 3B). We synthesized HA-DP polymer with various degree of substitution (DS) and discovered that HA-DP with 42% of DS to

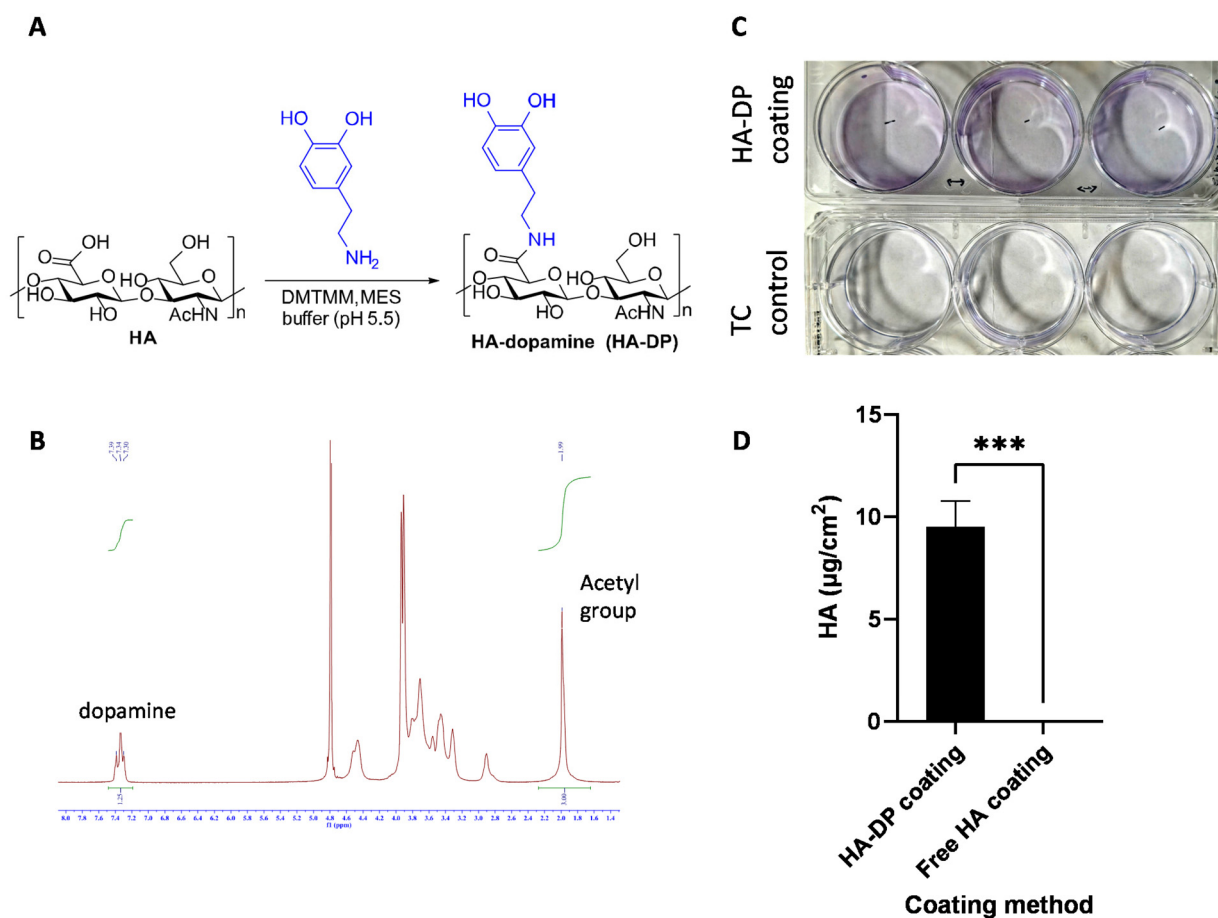


Fig. 3 Synthesis and characterization of HA-DP. (A) Schematic of chemical reaction to generate HA-DP. The dopamine was conjugated with the carboxyl group of HA using DMTMM, MES buffer (pH 5.5). (B) The conjugation of dopamine group to HA was confirmed using ¹H-NMR. The protons of the aromatic ring of DP showed peaks at 7.4–7.2 ppm and those of *N*-acetyl groups of HA showed 2.0 ppm of the spectrum of ¹H-NMR confirming an appropriate conjugation of DP to HA. ¹H-NMR spectra analysis confirms 42% of degree of substitution. (C) Representative photo taken after TBO staining and washing as described in characterization method, a control tissue culture plastic was used to show difference in purple blue stain due to staining. (D) The difference between amount of bound HA on the surface between two coating method. The left column of the graph represents the coating with HA-DP and right one represents coating with free HA solution. We found no presence of HA from free solution-based coating compared to HA-DP based coating.



be optimum for surface coating (data not shown). Therefore, HA-DP with 42% DS was used to coat tissue culture plate for our subsequent studies. Toluidine blue assay was used to quantify the amount HA-DP conjugated to the tissue culture plate. The presence of HA-DP was evident through toluidine blue assay in HA-DP coated plate compared to the control (Fig. 3C). Quantification using a standard curve revealed $9.5 \pm 0.2 \mu\text{g cm}^{-2}$ was present in the HA-DP coated plate (Fig. 3D and ESI Fig. 4†).

Preservation of LEC phenotype though HA-DP coating

To test the hypothesis that HA-DP can preserve lymphatic phenotypes, we cultured LECs on HA-DP, as well as tissue culture plate and fibronectin-coated plate as controls. LECs were ana-

lyzed for their lymphatic phenotypes using FACS and real-time qRT-PCR analysis. To evaluate lymphatic phenotypes at gene expression levels among different coating conditions, we decided to evaluate them at early passage (P.5) and late passage (P.7), where their gene expression levels were known to be different. At early passage (P.5), we noticed that LECs cultured on HA-DP expressed 1.70 \pm 0.18-fold more expression of *LYVE-1* and 2.24 \pm 0.19-fold more expression of *PDPN* compared to LECs cultured on tissue culture plate (Fig. 4A and B). LECs cultured on fibronectin coated plate expressed 1.39 \pm 0.20-fold more expression of *PDPN* compared to LECs cultured on tissue culture plate (Fig. 4B). While the expression levels of others lymphatic markers *Prox1* and *VEGFR3* were not signifi-

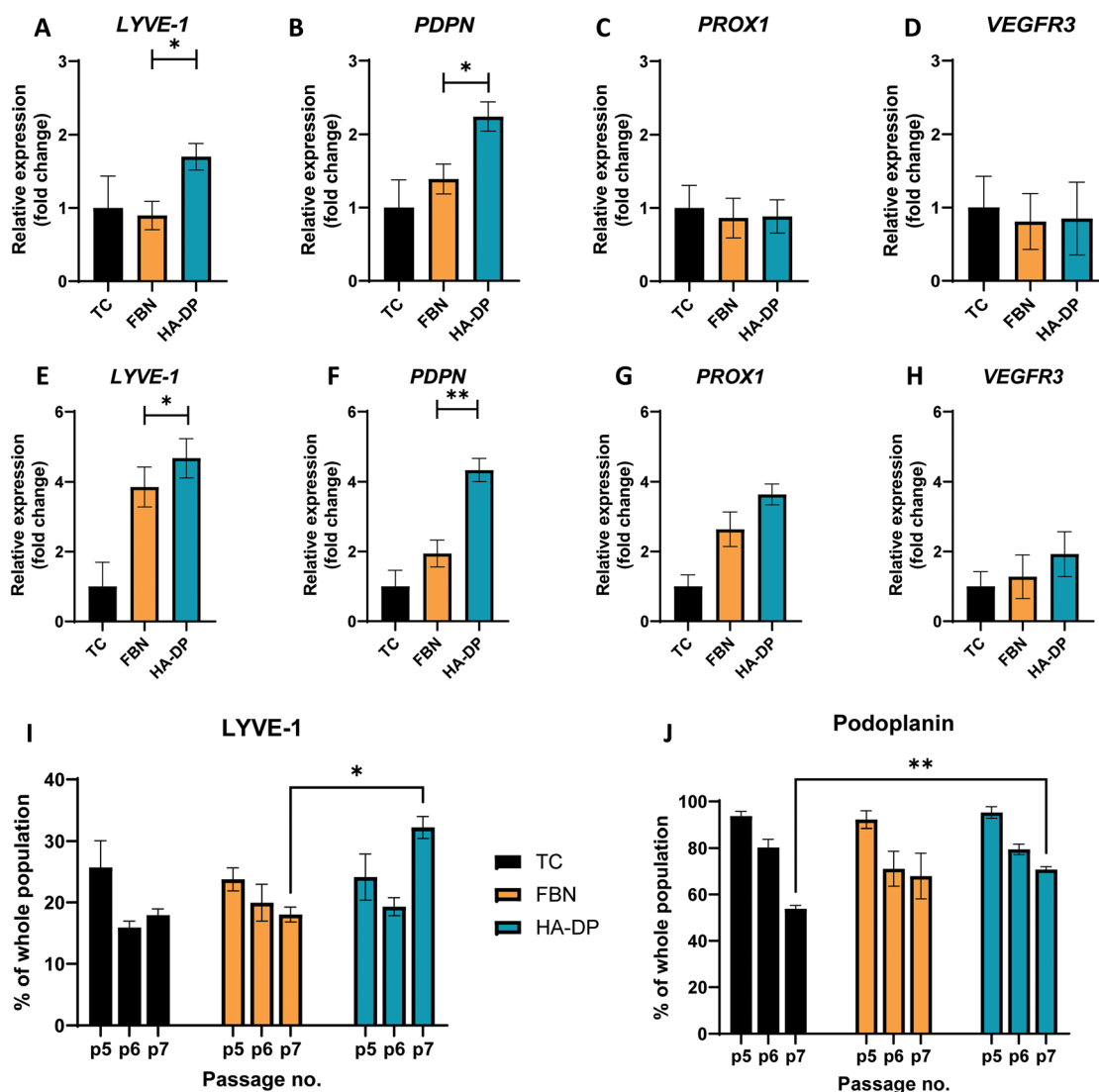


Fig. 4 Protein and gene expression of LEC cultured on different coatings. Real-time qRT-PCR for early passage (P.5) LECs cultured on tissue culture plate (black), fibronectin-coated plate (yellow), and HA-DP (blue) coated plate for (A) *LYVE-1*, (B) *PDPN*, (C) *Prox1*, and (D) *VEGFR3*. Real-time qRT-PCR for late passage (P.7) LECs cultured on tissue culture plate (black), fibronectin-coated plate (yellow), and HA-DP (blue) coated plate for (E) *LYVE-1*, (F) *PDPN*, (G) *Prox1*, and (H) *VEGFR3*. Three biological replicates ($n = 3$) were collected per condition and analysed with real-time qRT-PCR with triplicate readings. Flow cytometry analysis for LECs (P.5–7) cultured on tissue culture plate (black), fibronectin-coated plate (yellow), and HA-DP (blue) coated plate indicating percentage of cells that are (I) LYVE-1⁺ and (J) podoplanin⁺. Data represents mean \pm stdev., $n = 4$ per condition. Significance levels were set at: * $P < 0.05$ and ** $P < 0.01$.



cantly different among different coating conditions (Fig. 4C and D). At higher passage (P.7), compared to LECs cultured on tissue culture plate, LECs cultured on HA-DP express higher levels of lymphatic markers *LYVE-1* (4.67 ± 0.56 -fold), *PDPN* (4.33 ± 0.33 -fold), *Prox1* (3.64 ± 0.30 -fold), and *VEGFR3* (2.0 ± 0.64 -fold) (Fig. 4E–H). LECs cultured on fibronectin coated plate also expressed higher lymphatic markers *LYVE-1* (3.85 ± 0.57 -fold), *PDPN* (1.94 ± 0.38 -fold), *Prox1* (2.63 ± 0.49 -fold), and *VEGFR3* (1.28 ± 0.63 -fold) compared to LECs cultured on tissue culture plates (Fig. 4E–H). Overall, LECs cultured on HA-DP express higher lymphatic markers compared to LECs cultured on tissue culture plates and fibronectin coated plates. To further validate this trend at the protein level, we performed FACS analysis on LECs cultured on different coating conditions from passage 5 to 7. For LECs cultured on tissue culture plate, we noticed a decrease in the percentage of *LYVE-1*⁺ cells (Fig. 4I) and podoplanin⁺ cells (Fig. 4J) from passage 5 to 7. LECs cultured on fibronectin coated plates also demonstrated a decrease in the percentage of *LYVE-1*⁺ cells (Fig. 4I) and podoplanin⁺ cells (Fig. 4J) at a lesser extent. Interestingly, LECs cultured on HA-DP coated plates maintained the percentage of *LYVE-1*⁺ cells (Fig. 4I) and demonstrated less decrease in the percentage of podoplanin⁺ cells (Fig. 4J). Overall, these results suggested that HA-DP coating preserves lymphatic phenotypes during *in vitro* culture of LECs for at least until passage 7.

Differential expression of YAP/TAZ and FAK on fibronectin and HA-DP coated plates

Since HA-DP can preserve lymphatic phenotypes better than the standard of culture on fibronectin-coated plates, we decided to further investigate the differences in mechanotransduction among culture conditions. Difference in ECM component often induces mechanotransduction on the cells independent of matrix stiffness.⁵⁵ Though there might not be much difference in stiffness between fibronectin and HA-DP, we were interested in to investigate focal adhesion kinase (FAK) and F-actin. At low passage (P.5), we did not observe significant expression of FAK on LECs cultured on fibronectin and HA-DP (ESI Fig. 5†). However, at late passage (P.7) LECs cultured on fibronectin-coated plate express higher FAK (0.03 ± 0.004 count per μm^2) compared to LECs cultured on HA-DP (0.013 ± 0.0004 count per μm^2) (Fig. 5A and B). Similarly, F-actin density and alignment was more pronounced in LECs cultured on fibronectin compared to HA-DP (Fig. 5C and D). Since FAK can also activate the “Hippo Pathway”, we also investigated YAP/TAZ and its downstream genes *CTGF* and *MYC*. At low passage (P.5), though not significant, LECs cultured on HA-DP express lower *CTGF*, *YAP*, and *TAZ* (ESI Fig. 6†). But, at high passage (P.7), LECs cultured on HA-DP express lower *TAZ* (0.71 ± 0.1 -fold), *CTGF* (0.34 ± 0.12 -fold), and *MYC* (0.76 ± 0.07 -fold) compared to LECs cultured on fibronectin coated plates

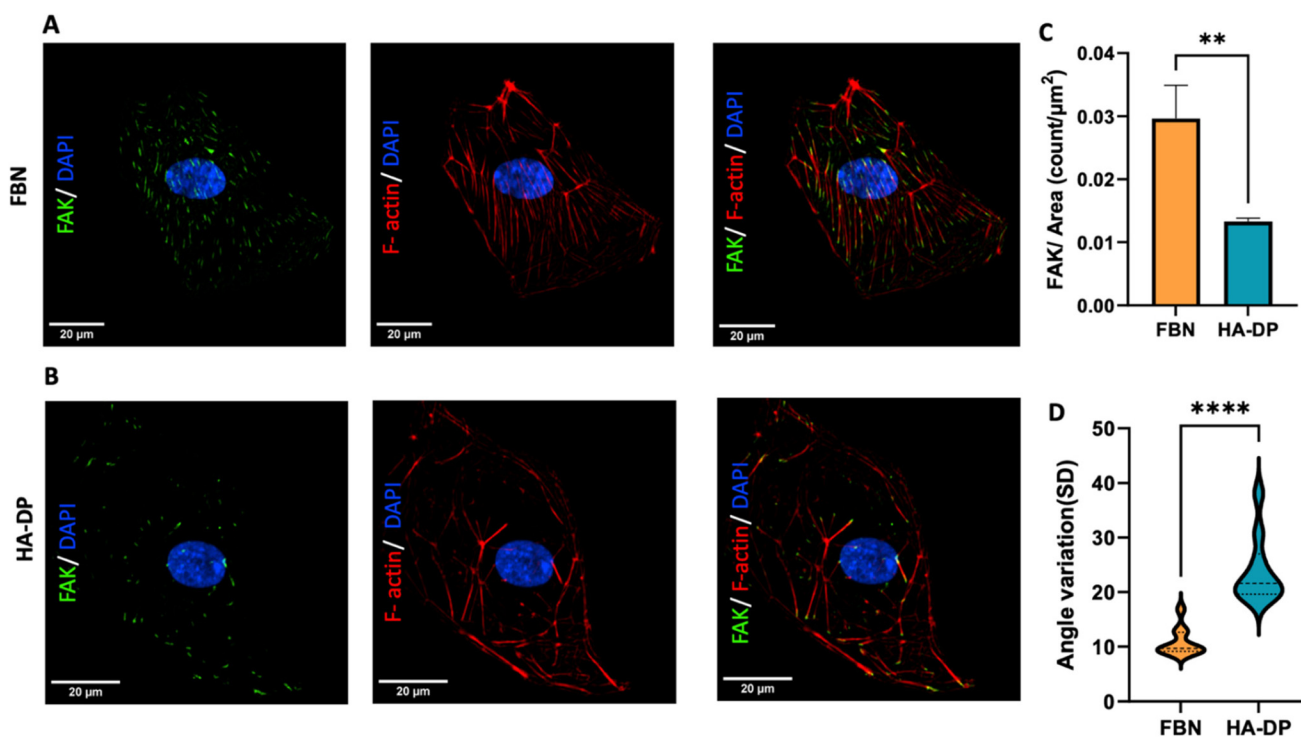


Fig. 5 Focal adhesion kinase (FAK) expression of LECs cultured on fibronectin and HA-DP coated plates. Representative immunofluorescent images of LECs (P.7) cultured on (A) fibronectin and (B) HA-DP coated plates stained for FAK (green), F-actin (red), and nuclei (blue). Scale bars are 20 μm . (C) The number of FAK per surface area was analysed for LECs cultured on fibronectin and HA-DP coated plates. (D) The orientation of F-actin was analysed using angle variation (SD) for LECs cultured on fibronectin and HA-DP coated plates. Data represents mean \pm stdev., $n = 4$ per condition. Significance levels were set at: ** $P < 0.05$ and **** $P < 0.001$.





Fig. 6 YAP/TAZ expression for LECs cultured on HA–DP and fibronectin coated plates. Real-time qRT-PCR data for (A) *CTGF*, (B) *MYC*, (C) *YAP*, and (D) *TAZ* expressed by LECs (P.7) cultured on HA–DP and fibronectin coated plates. Three biological replicates ($n = 3$) were collected per condition and analysed with real-time qRT-PCR with triplicate readings. Representative confocal z-stacks of LECs (P.7) cultured on (E) fibronectin and (F) HA–DP coated plates stained with YAP (green), TAZ (red) and nuclei (blue). Scale bars are 100 μm . Insets indicate the localization of YAP (green) and TAZ (red). Scale bars are 20 μm . Fluorescent intensity quantification demonstrates (G) an increase in cytoplasmic localization for YAP and (H) an increase in cytoplasmic localization of TAZ for LECs cultured on HA–DP compared to fibronectin. CTCF, corrected total cell fluorescence. Data represents mean \pm stdev., $n = 4$ per condition. Significance levels were set at: $**P < 0.05$, $*P < 0.05$ and $****P < 0.001$.

(Fig. 6A–D). These observations were also confirmed using immunofluorescent imaging of YAP/TAZ (Fig. 6E and F). Furthermore, quantification of the fluorescent signal indicates that culturing LECs on HA–DP led to a more cytoplasmic localization of YAP/TAZ compared to LECs cultured on fibronectin coated plates (Fig. 6G and H). Therefore, culturing LECs on HA–DP enabled cytoplasmic degradation of YAP/TAZ, which subsequently enhance transcription of lymphatic master regulator *Prox1*, including its targets *LYVE-1*, *podoplanin*, and *VEGFR3*. The differences in local and global expression of YAP/TAZ in both FBN and HA–DP explains our results and suggests that HA–DP coated plate preserve lymphatic phenotypes by downregulating FAK and YAP/TAZ pathways (Fig. 7). Overall, these observations are consistent with previous

studies that show YAP/TAZ negatively regulate *Prox1* during development and *in vitro* culture.^{15,63}

Discussion

The discovery of unique lymphatic markers, such as *Prox1*, *LYVE-1*, and *PDPN* have allowed the isolation and cultured of LECs, which paved the way for many studies for better understanding of lymphatic biology. While it is challenging to isolate and maintain murine LECs in culture, human primary LECs can be isolated from adult and juvenile tissues. Once isolated, these LECs can be cultured on regular tissue culture plate, as well as on plates coated with fibronectin or collagen.





Fig. 7 Schematic diagram depicting the role of HA–DP in preserving lymphatic phenotypes. HA–DP was able to preserve key lymphatic markers, including Prox1, LYVE-1, podoplanin, and VEGFR3. When LECs are cultured on fibronectin coated plates, YAP/TAZ enter the nucleus and bind to the PROX-1 promoter, inhibiting its transcription, including its targets, such as LYVE-1, podoplanin, and VEGFR3. In contrast, culturing LECs on HA–DP coated plates enables YAP/TAZ to undergo cytoplasmic degradation, which subsequently enhance transcription of Prox1, including its targets, such as LYVE-1, podoplanin, and VEGFR3.

While there is no agreement on the best culture condition to culture LECs *in vitro*, it has been recognized that LECs may start to lose their lymphatic expression overtime during culture.^{9,64–67} One of these investigations even mentions reduction in Prox1 as well. However, in this study, we confirmed that LECs can be cultured from P.5–7, while maintaining their Prox1 expression. But they started to lose other key lymphatic markers, such as LYVE-1 and podoplanin. Both LYVE-1 and podoplanin are surface receptor uniquely expressed by LECs. LYVE-1 is important for leukocytes trafficking and highly abundant in lymphatic capillaries.^{68,69} While podoplanin is responsible for blood-lymphatic separation and highly abundant in lymphatic pre-collecting and collecting vessels.^{51,70–72} Lymphatic capillaries are LYVE1^{high} and podoplanin^{low}, because of the abundant hyaluronic acid used by leukocytes to enter the lymphatic capillaries.^{73,74} However, it is still unclear if other matrix proteins are responsible for the pre-collecting and collecting vessels to express LYVE1^{low} and podoplanin^{high}. Therefore, understanding how different matrix proteins can be used to enrich certain population of LECs *in vitro* is important and can be used for future studies in vascular and lymphatic biology.

We demonstrated that a simple coating with HA–DP can effectively preserve key lymphatic markers over several passages *in vitro*. In fact, key lymphatic makers were maintained on high passage (P.7) of LECs cultured on HA–DP, compared to LECs cultured on fibronectin coated plates. LECs uniquely express LYVE-1, a specific receptor for HA, and provide a unique advantage for engineered matrices containing HA. Other than LYVE-1, VEGFR3 is the only receptor that can preserve lymphatic phenotypes in the presence of VEGF-C. But a growth factor independent, ECM-based interaction can be modulated through HA–DP coating, which opens many opportunities to modify culture media based on different applications. Therefore, future studies can use HA–DP to mimic

human physiology and pathology depending on the intended applications (*i.e.*, lymphatic capillaries *vs.* collecting vessels).

Moreover, we showed that LECs on HA–DP coating expressed reduced FAK and F-actin stress fibres. FAK is considered one of the key mechanotransductory components and one of the first molecules recruited to focal adhesions in response to external mechanical stimuli. FAK is also a regulator of F-actin dynamics.^{75,76} Which explains our result from Fig. 5. As cells experience more mechanical stress, more focal adhesion assembly occurs and in turn causes increased amount of F-actin stress fibre and their alignment. The FAK is a pivotal mediator of cell mechanosignaling and relays these stimuli to other mechanotransducers like YAP/TAZ.⁷⁷ Simultaneously, Hippo effectors YAP/TAZ act as mechanosensing switches whose expression is analogous to the encountered mechanotransduction.⁷⁸ All these concludes into the fact that LECs experience reduced mechanotransduction on HA–DP coating compared to fibronectin.

The “Hippo Pathway” and YAP/TAZ are critically involved in initial LEC specification, differentiation, and sprouting during early lymphatic development and in maintaining lymphatic integrity during adulthood.⁶³ Recent studies described that YAP/TAZ work as stress-mediated mechanotransducers in LECs like that of BECs but different in that YAP/TAZ regulate Prox1 transcriptional activity in LECs.^{79,80} Strikingly, lymphatic YAP/TAZ negatively regulate Prox1 transcription, and they modulate Prox1 activity and lymphatic plasticity.⁶³ Consistent to previous findings, our data also suggest that culturing LECs on HA–DP coated plates causes cytoplasmic degradation of YAP/TAZ, which subsequently enhances transcription of lymphatic master regulator Prox1, including its targets LYVE-1, PDPN, and VEGFR3.

Collectively, we demonstrated that a simple HA–DP can be used to preserve lymphatic phenotypes during *in vitro* culture of primary LECs. Mechanistically, HA–DP caused downregula-



tion of YAP/TAZ, which upregulate Prox1 and therefore maintain lymphatic phenotypes, consistent with previous findings. We hope that these results would further improve subsequent *in vitro* studies and inspire other researchers to adapt this technology for other applications (*i.e.*, culturing mouse LECs, which has been known to be very difficult to culture *in vitro*). Overall, this simple yet effective HA-DP coating may be useful for culturing human LECs *in vitro* for applications in basic lymphatic biology and lymphatic regeneration.

Author contributions

S. S., F. F., L. A., and D. H.-P. conceived the ideas, designed the experiments, interpreted the data, and wrote the manuscript. S. S., F. F., F. G., and E. H. conducted the experiments and analysed data. All authors have approved the manuscript.

Conflicts of interest

The authors have declared that no conflict of interest exists.

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References

- 1 T. Tammela and K. Alitalo, *Cell*, 2010, **140**, 460–476.
- 2 K. Alitalo and P. Carmeliet, *Cancer Cell*, 2002, **1**, 219–227.
- 3 H. Wiig and M. A. Swartz, *Physiol. Rev.*, 2012, **92**, 1005–1060.
- 4 J. B. Dixon, S. Raghunathan and M. A. Swartz, *Biotechnol. Bioeng.*, 2009, **103**, 1224–1235.
- 5 Y. Wang and G. Oliver, *Genes Dev.*, 2010, **24**, 2115–2126.
- 6 S. Liao and T. P. Padera, *Lymphatic Res. Biol.*, 2013, **11**, 136–143.
- 7 N. C. Johnson, M. E. Dillard, P. Baluk, D. M. McDonald, N. L. Harvey, S. L. Frase and G. Oliver, *Genes Dev.*, 2008, **22**, 3282–3291.
- 8 M. G. Bixel and R. H. Adams, *Genes Dev.*, 2008, **22**, 3232–3235.
- 9 M. H. Ulvmar and T. Mäkinen, *Cardiovasc. Res.*, 2016, **111**, 310–321.
- 10 L. Alderfer, E. Hall and D. Hanjaya-Putra, *Acta Biomater.*, 2021, **133**, 34–45.
- 11 L. Alderfer, A. Wei and D. Hanjaya-Putra, *J. Biol. Eng.*, 2018, **12**, 32.
- 12 S. Landau, A. Newman, S. Edri, I. Michael, S. Ben-Shaul, Y. Shandalov, T. Ben-Arye, P. Kaur, M. H. Zheng and S. Levenberg, *Proc. Natl. Acad. Sci. U. S. A.*, 2021, **118**, e2101931118.
- 13 J. S. T. Hooks, F. C. Bernard, R. Cruz-Acuna, Z. Nepiyushchikh, Y. Gonzalez-Vargas, A. J. Garcia and J. B. Dixon, *Biomaterials*, 2022, **284**, 121483.
- 14 M. A. Gimbrone Jr., R. S. Cotran and J. Folkman, *J. Cell Biol.*, 1974, **60**, 673–684.
- 15 L. Alderfer, E. Russo, A. Archilla, B. Coe and D. Hanjaya-Putra, *FASEB J.*, 2021, **35**, e21498.
- 16 A. A. A. Alghamdi, C. J. Benwell, S. J. Atkinson, J. Lambert, R. T. Johnson and S. D. Robinson, *Front. Cell Dev. Biol.*, 2020, **8**, 395.
- 17 M. Irigoyen, E. Anso, E. Salvo, J. Dotor de las Herreras, J. J. Martinez-Irujo and A. Rouzaut, *Cell. Mol. Life Sci.*, 2008, **65**, 2244–2255.
- 18 S. Lutter, S. Xie, F. Tatin and T. Mäkinen, *J. Cell Biol.*, 2012, **197**, 837–849.
- 19 T. Mäkinen, T. Veikkola, S. Mustjoki, T. Karpanen, B. Catimel, E. C. Nice, L. Wise, A. Mercer, H. Kowalski, D. Kerjaschki, S. A. Stacker, M. G. Achen and K. Alitalo, *EMBO J.*, 2001, **20**, 4762–4773.
- 20 M. Mitsi, M. M. Schulz, E. Gousopoulos, A. M. Ochsenbein, M. Detmar and V. Vogel, *PLoS One*, 2015, **10**, e0145210.
- 21 A. Sabine, Y. Agalarov, H. Maby-El Hajjami, M. Jaquet, R. Hagerling, C. Pollmann, D. Bebbber, A. Pfenniger, N. Miura, O. Dormond, J. M. Calmes, R. H. Adams, T. Mäkinen, F. Kiefer, B. R. Kwak and T. V. Petrova, *Dev. Cell*, 2012, **22**, 430–445.
- 22 E. Garrafa, L. Trainini, A. Benetti, E. Saba, L. Fezzardi, B. Lorusso, P. Borghetti, T. Bottio, E. Ceri, N. Portolani, S. Bonardelli, S. M. Giulini, G. Annibale, A. Corradi, L. Imberti and A. Caruso, *Lymphology*, 2005, **38**, 159–166.
- 23 X. Zhang, J. E. Groopman and J. F. Wang, *J. Cell Physiol.*, 2005, **202**, 205–214.
- 24 J. F. Wang, X. F. Zhang and J. E. Groopman, *J. Biol. Chem.*, 2001, **276**, 41950–41957.
- 25 T. Dietrich, J. Onderka, F. Bock, F. E. Kruse, D. Vossmeier, R. Stragies, G. Zahn and C. Cursiefen, *Am. J. Pathol.*, 2007, **171**, 361–372.
- 26 N. E. Vlahakis, B. A. Young, A. Atakilit and D. Sheppard, *J. Biol. Chem.*, 2005, **280**, 4544–4552.
- 27 E. Bazigou, S. Xie, C. Chen, A. Weston, N. Miura, L. Sorokin, R. Adams, A. F. Muro, D. Sheppard and T. Mäkinen, *Dev. Cell*, 2009, **17**, 175–186.
- 28 F. Tatin, A. Taddei, A. Weston, E. Fuchs, D. Devenport, F. Tissir and T. Mäkinen, *Dev. Cell*, 2013, **26**, 31–44.
- 29 A. M. Foskett, U. R. Ezekiel, J. P. Trzeciakowski, D. C. Zawieja and M. Muthuchamy, *Front. Physiol.*, 2011, **2**, 103.



- 30 B. Sauter, D. Foedinger, B. Sterniczky, K. Wolff and K. Rappersberger, *J. Histochem. Cytochem.*, 1998, **46**, 165–176.
- 31 T. Kono, H. Kubo, C. Shimazu, Y. Ueda, M. Takahashi, K. Yanagi, N. Fujita, T. Tsuruo, H. Wada and J. K. Yamashita, *Arterioscler., Thromb., Vasc. Biol.*, 2006, **26**, 2070–2076.
- 32 E. Garrafa, G. Alessandri, A. Benetti, D. Turetta, A. Corradi, A. M. Cantoni, E. Cervi, S. Bonardelli, E. Parati, S. M. Giuliani, B. Ensoli and A. Caruso, *J. Cell Physiol.*, 2006, **207**, 107–113.
- 33 P. S. Yoo, A. L. Mulkeen, A. Dardik and C. H. Cha, *J. Surg. Res.*, 2007, **143**, 94–98.
- 34 F. Laco, M. H. Grant and R. A. Black, *J. Biomed. Mater. Res., Part A*, 2013, **101**, 1787–1799.
- 35 T. Ishikawa, Z. Wondimu, Y. Oikawa, G. Gentilcore, R. Kiessling, S. Egyhazi Brage, J. Hansson and M. Patarroyo, *Matrix Biol.*, 2014, **38**, 69–83.
- 36 C. Schuster, M. Mildner, A. Botta, L. Nemeč, R. Rogojanu, L. Beer, C. Fiala, W. Eppel, W. Bauer, P. Petzelbauer and A. Elbe-Burger, *Am. J. Pathol.*, 2015, **185**, 2563–2574.
- 37 D. Hanjaya-Putra, V. Bose, Y. I. Shen, J. Yee, S. Khetan, K. Fox-Talbot, C. Steenbergen, J. A. Burdick and S. Gerecht, *Blood*, 2011, **118**, 804–815.
- 38 D. Hanjaya-Putra, K. T. Wong, K. Hirotsu, S. Khetan, J. A. Burdick and S. Gerecht, *Biomaterials*, 2012, **33**, 6123–6131.
- 39 M. T. Ngo and B. A. Harley, *Adv. Healthc. Mater.*, 2017, **6**, 1700687.
- 40 L. Sherman, J. Sleeman, P. Herrlich and H. Ponta, *Curr. Opin. Cell Biol.*, 1994, **6**, 726–733.
- 41 S. Ibrahim and A. Ramamurthi, *J. Tissue Eng. Regener. Med.*, 2008, **2**, 22–32.
- 42 D. C. West, I. N. Hampson, F. Arnold and S. Kumar, *Science*, 1985, **228**, 1324–1326.
- 43 P. W. Noble, *Matrix Biol.*, 2002, **21**, 25–29.
- 44 E. Turley, M. Hossain, T. Sorokan, L. Jordan and J. Nagy, *Glia*, 1994, **12**, 68–80.
- 45 A. Aruffo, I. Stamenkovic, M. Melnick, C. B. Underhill and B. Seed, *Cell*, 1990, **61**, 1303–1313.
- 46 M. Yu, H. Zhang, Y. Liu, Y. He, C. Yang, Y. Du, M. Wu, G. Zhang and F. Gao, *Exp. Cell Res.*, 2015, **336**, 150–157.
- 47 M. Wu, Y. Du, Y. Liu, Y. He, C. Yang, W. Wang and F. Gao, *PLoS One*, 2014, **9**, e92857.
- 48 C. H. Antoni, Y. McDuffie, J. Bauer, J. P. Sleeman and H. Boehm, *Front. Bioeng. Biotechnol.*, 2018, **6**, 25.
- 49 X. Pang, W. Li, L. Chang, J. E. Gautrot, W. Wang and H. S. Azevedo, *ACS Appl. Mater. Interfaces*, 2021, **13**, 25792–25804.
- 50 S. Lee, S. Kim, J. Park and J. Y. Lee, *Int. J. Biol. Macromol.*, 2020, **151**, 1314–1321.
- 51 D. P. Jeong, E. Hall, E. Neu and D. Hanjaya-Putra, *Cell. Mol. Bioeng.*, 2022, **15**, 467–478.
- 52 L. Bui, S. Edwards, E. Hall, L. Alderfer, K. Round, M. Owen, P. Sainaghi, S. Zhang, P. D. Nallathamby, L. S. Haneline and D. Hanjaya-Putra, *Commun. Biol.*, 2022, **5**, 635.
- 53 K. M. Varberg, R. O. Garretson, E. K. Blue, C. Chu, C. R. Gohn, W. Tu and L. S. Haneline, *Am. J. Physiol.: Cell Physiol.*, 2018, **315**, C502–C515.
- 54 M. Wagner, B. Hampel, D. Bernhard, M. Hala, W. Zwerschke and P. Jansen-Dürr, *Exp. Gerontol.*, 2001, **36**, 1327–1347.
- 55 J. L. Zhang, J. M. Patel and E. R. Block, *Mech. Ageing Dev.*, 2002, **123**, 613–625.
- 56 Y. K. Hong, N. Harvey, Y. H. Noh, V. Schacht, S. Hirakawa, M. Detmar and G. Oliver, *Dev. Dyn.*, 2002, **225**, 351–357.
- 57 J. T. Wagle and G. Oliver, *Cell*, 1999, **98**, 769–778.
- 58 J. Kang, J. Yoo, S. Lee, W. Tang, B. Aguilar, S. Ramu, I. Choi, H. H. Otu, J. W. Shin, G. P. Dotto, C. J. Koh, M. Detmar and Y. K. Hong, *Blood*, 2010, **116**, 140–150.
- 59 S. Lee, I. Choi and Y. K. Hong, *Semin. Thromb. Hemostasis*, 2010, **36**, 352–361.
- 60 W. Ma and G. Oliver, *Physiology*, 2017, **32**, 444–452.
- 61 D. G. Jackson, R. Prevo, S. Clasper and S. Banerji, *Trends Immunol.*, 2001, **22**, 317–321.
- 62 H. Lee, S. M. Dellatore, W. M. Miller and P. B. Messersmith, *Science*, 2007, **318**, 426–430.
- 63 H. Cho, J. Kim, J. H. Ahn, Y.-K. Hong, T. Mäkinen, D.-S. Lim and G. Y. Koh, *Circ. Res.*, 2019, **124**, 225–242.
- 64 J. W. Robering, A. Weigand, R. Pfuhlmann, R. E. Horch, J. P. Beier and A. M. Boos, *J. Cell. Mol. Med.*, 2018, **22**, 3740–3750.
- 65 S. Amatschek, E. Kriehuber, W. Bauer, B. Reininger, P. Meraner, A. Wolpl, N. Schweifer, C. Haslinger, G. Stingl and D. Maurer, *Blood*, 2007, **109**, 4777–4785.
- 66 K. L. Jablon, V. L. Akerstrom, M. Li, S. E. Braun, C. E. Norton and J. A. Castorena-Gonzalez, *Microcirculation*, 2023, **30**, e12778.
- 67 S. F. Schoppmann, A. Soleiman, R. Kalt, Y. Okubo, C. Benisch, U. Nagavarapu, G. S. Herron and S. Geleff, *Microcirculation*, 2004, **11**, 261–269.
- 68 L. A. Johnson, S. Banerji, W. Lawrance, U. Gileadi, G. Prota, K. A. Holder, Y. M. Roshorm, T. Hanke, V. Cerundolo, N. W. Gale and D. G. Jackson, *Nat. Immunol.*, 2017, **18**, 762–770.
- 69 D. G. Jackson, *Matrix Biol.*, 2019, **78–79**, 219–235.
- 70 P. R. Hess, D. R. Rawnsley, Z. Jakus, Y. Yang, D. T. Sweet, J. Fu, B. Herzog, M. Lu, B. Nieswandt, G. Oliver, T. Mäkinen, L. Xia and M. L. Kahn, *J. Clin. Invest.*, 2014, **124**, 273–284.
- 71 R. Bianchi, E. Russo, S. B. Bachmann, S. T. Proulx, M. Sesartic, N. Smaadahl, S. P. Watson, C. D. Buckley, C. Halin and M. Detmar, *Arterioscler., Thromb., Vasc. Biol.*, 2017, **37**, 108–117.
- 72 D. P. Jeong, D. Montes, H. C. Chang and D. Hanjaya-Putra, *Phys. Biol.*, 2023, **20**, 045004.
- 73 L. A. Johnson, S. Banerji, W. Lawrance, U. Gileadi, G. Prota, K. A. Holder, Y. M. Roshorm, T. Hanke, V. Cerundolo, N. W. Gale and D. G. Jackson, *Nat. Immunol.*, 2017, **18**, 762–770.
- 74 D. G. Jackson, *Matrix Biol.*, 2019, **78–79**, 219–235.
- 75 S. Y. Li, D. D. Mruk and C. Y. Cheng, *Spermatogenesis*, 2013, **3**, e25385.



- 76 M. Schober, S. Raghavan, M. Nikolova, L. Polak, H. A. Pasolli, H. E. Beggs, L. F. Reichardt and E. Fuchs, *J. Cell Biol.*, 2007, **176**, 667–680.
- 77 D. Lachowski, E. Cortes, B. Robinson, A. Rice, K. Rombouts and A. E. Del Rio Hernandez, *FASEB J.*, 2018, **32**, 1099–1107.
- 78 G. Nardone, J. Oliver-De La Cruz, J. Vrbsky, C. Martini, J. Pribyl, P. Skladal, M. Pesl, G. Caluori, S. Pagliari, F. Martino, Z. Maceckova, M. Hajduch, A. Sanz-Garcia, N. M. Pugno, G. B. Stokin and G. Forte, *Nat. Commun.*, 2017, **8**, 15321.
- 79 S. Dupont, L. Morsut, M. Aragona, E. Enzo, S. Giulitti, M. Cordenonsi, F. Zanconato, J. Le Digabel, M. Forcato, S. Bicciato, N. Elvassore and S. Piccolo, *Nature*, 2011, **474**, 179–183.
- 80 B. Cha, S. Moon and W. Kim, *BMB Rep.*, 2021, **54**, 285–294.

