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Electrochemical hydrogen sulfide biosensors

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The measurement of sulfide, especially hydrogen sulfide, has held the attention of the analytical community due to its unique physiological and pathophysiological roles in biological systems. Electrochemical detection offers a rapid, highly sensitive, affordable, simple, and real-time technique to measure hydrogen sulfide concentration, which has been a well-documented and reliable method. This review details up-to-date research on the electrochemical detection of hydrogen sulfide (ion selective electrodes, polarographic hydrogen sulfide sensors, etc.) in biological samples for potential therapeutic use.

1. Introduction

Hydrogen sulfide (H₂S) is well known as a colorless, flammable, and water soluble gas with the distinctive smell of rotten eggs and has received attention as a toxic gas and as an environmental hazard for many decades.¹ Recent research has found that H₂S is a common constituent of mammalian

tissues and cells, notably in the brain, heart, gastrointestinal tract, vascular smooth muscles, and endothelial cells, acting as a messenger much like nitric oxide (NO) and carbon monoxide (CO).^{2–5} Its important roles in mammalian systems prompt us to explore a fast and sensitive detection method to monitor its concentration in biological samples for fully understanding the effects of its possible physiological and pathophysiological roles, as well as its potential therapeutic use.^{6–8} Due to the widespread interest in its biological functions, detection methods for sulfide have evolved from simple colorimetric assays to the more recently used techniques, including high pressure liquid chromatography (HPLC) and gas chromatography (GC),^{9–11} monobromobimane (MBB),^{12–15} fluorescence probe,⁸ and electrochemical sensors (ion sensitive electrodes and polarographic H₂S sensors, etc.).^{16,17} Certainly, each technique has its advantages and limitations. Chromatography based methods, which have been used to

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chemical species, including nitric oxide, hydrogen peroxide, hydrogen sulfide, and oxygen.



measure both plasma and tissue H₂S sensitivity, however, do not have the capability of real-time measuring in biological samples and require hypoxic or anoxic conditions.^{10,11,18} In a typical MBB assay, H₂S reacts with two molecules of monobromobimane to form the fluorescent sulfide-dibimane, followed by HPLC assay with fluorescence detection. This method requires relatively expensive equipment, and its time consuming process results in unknown reactions in blood samples, thus interfering in the detection of sulfide and making real-time measurements of H₂S impossible.^{12–15} The methylene-blue method is one of the most commonly used spectrometric methods for measuring plasma H₂S, which involves the reaction of H₂S with *N,N*-dimethyl-*p*-phenylenediamine sulfate to produce the light-absorbing methylene blue. Nevertheless, this direct method lacks sensitivity at low concentrations (<1 μM) and requires long incubation periods (>30 min), making it unsuitable for continuous measurement of H₂S under real-time, physiological conditions.^{19,20} Some other spectroscopic methods offer high sensitivity and selectivity for H₂S.^{21–25} For example, it is noteworthy that the most recently developed sulfide-sensitive fluorescent probes enable H₂S measurement in cell and blood samples at low concentrations.^{14,26} In addition, the greatest limitations for fluorescent indicators (*i.e.* response time and sample damage) leave significant room for improvement in the future.

Chemical and biological methods with several advantages have become one of the most active areas of analytical research in recent years. Electrochemical biosensors, as one of the most common methods, have been widely used to detect biomolecules such as glucose,²⁷ nitric oxide,^{28,29} ascorbic acid,³⁰ uric acid,³¹ dopamine,³² and hydrogen peroxide.³³ Electrochemical biosensing is also an effective method to measure the H₂S concentration in different kinds of samples with distinct advantages of high sensitivity, high selectivity, low-detection limit, stability, long lifetime, wide linear range, fast

response time, biocompatibility, reproducibility, and simple operation.^{18,34–37} These advantages have attracted a number of researchers to develop new electrochemical methods for determining sulfide species in inorganic and organic forms.^{18,34–37} The present review has made an effort to detail the most recent research on ion sensitive electrodes, polarographic H₂S sensors, and enzyme based electrodes for H₂S concentration detection in biological samples.

2. Background perspective of H₂S

2.1 H₂S in chemistry

Hydrogen sulfide is a weak acid with pK_{a1} = 6.6, and pK_{a2} = 13.8 at 25 °C.³⁸ H₂S aqueous solution is a mixture of H₂S, HS⁻, S²⁻ and dissociates (H₂S ↔ H⁺ + HS⁻ ↔ 2H⁺ + S²⁻). However, there is only a trace of S²⁻ in normal biological samples (pH ~ 7) because pK_{a2} is higher than 13. H₂S is strongly harmful, toxic, and flammable when released into the atmosphere either as a pollutant from the chemical industry or as a by-product of biological activity. The electric potential between H₂S and O₂ is really high, making it extremely favorable for oxidation reactions, so H₂S can be oxidized to sulfur dioxide (SO₂), sulfates, and elemental sulfur. Chemical industry oxidation (H₂S + 2O₂ → SO₄²⁻ + 2H⁺) which is catalyzed by metals or metal oxides, is one of the main sources of acid rain. Moreover, H₂S is also harmful in industrial production: fabrication of fuel cells requires the control of H₂S at ppb levels in order to extend the lifetime of the proton exchange membrane, while in the food industry, H₂S needs to be monitored at a trace level to guarantee the quality of products.

2.2 H₂S function in a biological system

Recent studies have established that H₂S is an important signalling molecule of comparable importance to NO and CO in



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materials, electrochemical biosensors, and supramolecular assembly.

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mammalian tissues and cells. The production of H₂S can occur *via* two pathways: enzymatic and non-enzymatic. Much emphasis has been placed on the enzymatic production of H₂S, which is formed from cysteine by H₂S synthases, namely cystathionine β-synthase (CBS), cystathionine γ-lyase (CSE) and 3-mercaptopyruvate sulfurtransferase (MST).³⁹ Non-enzymatic production of H₂S occurs through glutathione, inorganic and organic polysulfides, and elemental sulfur.^{18,39} Apart from the free state, H₂S can react with several different biochemical molecules to establish different bio-available pools, such as acid labile sulfides (Fe–S complexes and persulfides) and bound sulfide forms (polythionates, polysulfides, thiosulfate, thiosulfonates, bisorganylpolysulfanes, monoarylthio-sulfonates, and elemental sulfur). However, less is known about the catabolism of H₂S in the body. One possible catabolic pathway is the rapid oxidation to thiosulfate (S₂O₃²⁻) by mitochondria and subsequent conversion to the stable product sulfate (SO₄²⁻) in cells.⁴⁰ H₂S is more soluble in lipids than in water, which makes it easier to travel into and out of cell membranes. As one of the messenger molecules throughout the body, H₂S can modulate the neuronal transmission, can relax smooth muscles, and is involved in inflammation together with NO and CO.⁷ Several cardiovascular diseases are also related to its function in cardiovascular physiology and pathophysiology.⁴¹ Potential therapeutic uses in ischemia/reperfusion (I/R) injury have been examined extensively in the heart, brain, lungs, and liver.^{42,43} H₂S produces anti-inflammatory effects in neurocytes, which may contribute to atherosclerosis by various mechanisms depending on the disease stage.⁴⁴ Whereas H₂S is toxic to the human body at high levels; it may cause unconsciousness and death in humans at concentrations above 500 ppm.⁴⁵

2.3 Electrochemical oxidation of H₂S

A typical three-electrode configuration, consisting of a working electrode, a reference electrode, and a counter electrode, is employed to detect H₂S concentration from different kinds of sources. Hydrogen sulfide can be oxidized at the working electrode with the transfer of two electrons (H₂S = S + 2H⁺ + 2e⁻, E⁰ = 0.171 V vs. Ag/AgCl) and produce a current. With the transfer of more electrons, it can be further oxidized into sulfite (SO₃²⁻) and sulfate (SO₄²⁻).⁴⁶ Electrochemical H₂S sensors can be divided into liquid and solid electrolyte sensors based on its electrolytes. Based on their working principles, these sensors can be further divided into amperometric and potentiometric,³⁷ which are two of the most common techniques employed to detect H₂S concentration in biological samples. Specifically, an amperometric method can monitor the very low redox current (*e.g.* pA's) produced by the oxidation of H₂S over time at a fixed (poise) voltage potential. This method has a fast response time of only a few seconds, high sensitivity, and provides the quantitative measurement of minute changes in H₂S concentration.

3. Electrochemical detection of H₂S

Electrochemical sensors have several advantages that make them an attractive method for real-time detection of H₂S in biological samples due to their high sensitivity, high selectivity, low detection limit, fast response time, miniaturization capabilities, and the fact that they do not require additional chemical reagents. Solid-electrolyte sensors (such as solid-polymer electrolyte (SPE)-Pt and metal oxide based sensor) are commonly applied for quantitative analysis of hydrogen sulfide gas in the chemical industry.^{47–49} Ion sensitive electrodes (ISEs) and amperometric (polarographic) H₂S sensors (PHSSs) are two well-known devices that have been applied to detect H₂S concentration in biological samples.

3.1. Ion sensitive electrodes (ISEs) for H₂S detection

Ion sensitive based electrodes are one of the most commonly used methods to measure concentrations of H₂S in plasma with a detection range of 1–100 μM. This method is easy to obtain and operate. It is sensitive towards sulfide anions and allows for dynamic detection of sulfide concentration within one minute. Common Ag/Ag₂S ISEs are simply prepared by dipping a silver wire into saturated sodium sulfide solution. The silver wire slowly reacts with the S²⁻ ions, forming a thin and dark layer of Ag₂S on the surface of the silver wire. The Ag₂S layer can serve as a protection membrane preventing the interference of the samples. Carefully checking the electrode surface is necessary to make sure that the thin layer of Ag₂S without silver is exposed to the surface. Otherwise, there will be a loss of selectivity due to the silver portion of the electrode being responsive to redox changes in the solution.⁵⁰ This method was first reported in plasma H₂S concentration measurement by Mason *et al.*, but without the details of the procedure.⁵¹ Khan *et al.* made a full detailed procedure of



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Table 1 Measurement of the concentration of H₂S by using an ISE in biological samples

Standard	Plasma or blood H ₂ S concentration	Samples	Applications	Ref.
Na ₂ S	10	Rat blood	Sulfide oxidation	51
Na ₂ S	2	Ewes and steers blood		52
NaHS		Rat blood	K _{ATP} channel	53
NaHS	38	Trout plasma	Smooth muscle	57
H ₂ S	34	Human plasma	Chronic obstructive pulmonary disease	58
S ²⁻	51.7	Human plasma	Coronary heart disease	59
NaHS	47	Rat plasma	K _{ATP} channels	60
S ²⁻	65.7	Children plasma	Hypertension	61
H ₂ S	39.7	Rat plasma	Balloon injury	62
S ²⁻	46	Rat plasma	L-Arginine/NO pathway	63
H ₂ S	21	Rat plasma	Lung injury	64
Na ₂ S	23.8	Rat plasma	Metabolic function	65
NaHS	40	Rat plasma	Hypertension	54
NaHS	90	Rainbow trout plasma	Adrenergic stress response	66
NaHS	57.7	Rat plasma	Atherosclerotic lesions	44
H ₂ S		Rat plasma	Pancreatic insulin metabolism	67
NaHS or H ₂ S		Human plasma	Respiratory infections	68
NaHS or H ₂ S	30.8	Rat plasma	Airway inflammation	69
NaHS		Rat plasma	S-sulfhydration	70
NaHS	1.75 nM mg ⁻¹	Rat cellular H ₂ S	Methylglyoxal function	71
NaHS		Leaf supernatant	Gasotransmitter	72
NaHS		Leaf supernatant	Autophagy, and transcriptional modulator	73
		Rat liver	Dietary restriction	74

plasma H₂S concentration measurement.⁵² In brief, sulfide antioxidant buffer was added into blood samples at a ratio of 1:1. The electrode was rinsed with distilled water and then placed in sample solutions. Next, the value was recorded when a stable reading was displayed. Finally, H₂S concentration was calculated by the standard S²⁻ solution.⁵² Based on this method, commercial sulfide sensitive electrodes (Model 9616, Orion Research, Beverly, MA) were widely used in serum H₂S concentration detection.⁵³ Another commercial ISE, known as ArrowH₂STM developed by Lazar Research Laboratories (Los Angeles, CA), is a small ion electrode. This sensor can measure volumes down to 10 microliters with a one-step sample prep-performance directly in the micro container, which obviously prevented H₂S loss from the sample and increased accuracy. In addition, unlike other methods, this micro-electrode measurement method has no known interferents and has a detection limit as low as 100 nM.⁵⁴ A variety of applications of the ISE-based H₂S sensor have been listed in Table 1. However, using Ag/Ag₂S electrodes detection of H₂S has several problems, such as the requirement of an alkaline environment favoring the S²⁻ equilibrium, operating in a low concentration range, and maintaining the connection between the electrolyte bridge and the reference electrode.^{18,36} Whitfield *et al.* have demonstrated that S²⁻ levels of trout plasma and BSA placed in alkaline buffer increased by 1 mM over a period of 12 hours, which makes its application to biological samples questionable and requires consideration of strong chemical pretreatment of samples.^{18,55,56} Another disadvantage of this method is the requirement for the electrode to be reconditioned and calibrated daily due to formation of Ag₂S on the electrode surface.

3.2. Polarographic H₂S sensors (PHSSs)

Although it is clear that H₂S plays an important physiological role in mammalian systems, the effects of H₂S under physiological conditions remain largely undefined without the benefit of real-time H₂S measurements. An amperometric H₂S sensor offers a reliable real-time method and has shown promise in detecting the concentration of H₂S in whole blood and mammalian tissues, without altering the sample as summarized in Table 2.³⁶ This was first developed by Jeroschewski *et al.*, based on a standard Clark-type oxygen electrode, and used for H₂S measurement in aquatic environments with a detection limit as low as 2 μM.⁷⁵ Kraus and Doeller modified the polarographic oxygen sensor, enabling it for the real-time measurement of H₂S gas from biological samples with a lower detection limit in the nM range under anoxic conditions.⁷⁶ In short, macro PHSSs ($L = 25$ mm, $D = 12$ mm) were designed originally from a polarographic oxygen sensor (POS, model 2120, Orbisphere, Geneva, Switzerland) with platinum wires (1 and 0.5 mm diameter) as the anode and cathode, respectively. The PHSS housing was machined from polyether ether ketone (PEEK). The electrolyte was held in the sensor tip reservoir with a two-layer membrane made of an H₂S-permeable membrane (MEM-213, 25 μm thick, WPI, Inc.), cemented (Silicone Adhesive RTV 167, GE, Waterford, NY) to a 25 μm thick H₂S-impermeable membrane. The H₂S-permeable membrane allows the permeation of H₂S through a hole ($D = 0.5$ mm) into an alkaline K₃Fe(CN)₆ internal electrolyte solution. H₂S was dissociated into HS⁻ after crossing the permeable membrane and then chemically oxidized to SO while simultaneously reducing Fe(CN)₆³⁻ to Fe(CN)₆⁴⁻. The Fe(CN)₆⁴⁻ is then electroche-



Table 2 Measurement of the concentration of H₂S by using a PHSS in biological samples

Mode	Standard	Sample	Applications	Related disease	Ref.
ISO-H ₂ S-2	Na ₂ S	Mussel (gill tissue)	Metabolic pathways		76
	Na ₂ S	Rat organ homogenate	Sulfide production and consumption		34
	Na ₂ S	Human blood	Consumption of garlic		77
ISO-H ₂ S-2	H ₂ S	Rat tissues	Vasoactivity and O ₂ dependent		78
ISO-H ₂ S-2	H ₂ S	Mice	Preservation of mitochondrial function	Ischemia-reperfusion injury	42
	Na ₂ S	Cow, sea lions lung	O ₂ dependent H ₂ S metabolism	Hypoxic pulmonary vasodilation	79
ISO-H ₂ S-2		RAW264.7 macrophages	H ₂ S donor (GYY4137) function	Inflammatory	80
NS-ISO-H ₂ S PF100H		Spinacia oleracea seedlings	Modulating photosynthesis		81
ISO-H ₂ S-2	NaHS	Diabetic rat plasma	Reduce ROS formation	Diabetic	82
ISO-H ₂ S-100		Peripheral blood lymphocytes	AKT/GSK3β signal pathway	Systemic lupus erythematosus	83
ISO-H ₂ S-2		Rat plasma	S-Propargyl-Cysteine Formulation	Heart failure	84
ISO-H ₂ S-100	NaHS	Mitochondria (rat liver), mouse hepatoma cells	Intramitochondrial H ₂ S-producing pathway		85
	Na ₂ S	Rats tissue, Bovine lung and heart, sea lamprey	Oxygen sensing		86
ISO-H ₂ S-2		Micelle solutions	H ₂ S delivery		87
ISO-H ₂ S-2	Na ₂ S	Extracellular H ₂ S	Up-regulate nicotinamide phosphoribosyltransferase	Cancer cells recover	88
ISO-H ₂ S-100	Na ₂ S	Rat plasma	K _{ATP} channel expression	Hypertension	89
ISO-H ₂ S-2	Na ₂ S	Rats (serum, renal slice culture, renal tissue)	CBS/H ₂ S pathway	Hypertension	90

mically oxidized back to Fe(CN)₆³⁻ (0.1–0.2 V vs. SCE) on the surface of the platinum electrode, producing a current proportional to H₂S concentration in biological samples. Kraus's group developed another miniature PHSS based on the original design of the macro one. This miniature PHSS utilizes a cylindrical platinum anode that was electrically insulated along the side from a coiled platinum cathode. The core was housed in a 2 mm outside diameter stainless steel cylindrical sleeve and a 25 μm thick H₂S-permeable membrane to cover the tip. The reservoir between the sleeve and core was filled with the ferricyanide electrolyte solution. The end of the sensor, contained within the sleeve ($L = 50$ mm), was fitted into a port of a PEEK stopper (contained two sensor ports and an injection port). Whitfield *et al.* made further modifications of PHSS. In brief, this design of the PHSS consisted of a heat-sealed platinum wire and a platinum wire cathode coiled around the capillary tube. The whole assembly was then inserted into an outer glass case containing a solution of 50 mM potassium ferricyanide in pH 10, 0.5 M sodium carbonate buffer. An O-ring was seated in a groove in the outer glass case to hold in place a 25 μm thick silicone polycarbonate membrane (SSP-M213; Specialty Silicone Products, Inc., Ballston Spa NY). The 2 mm diameter PHSS (response time <5 s, detection limit <5 nM, sensitivity = 2 pA nM⁻¹) described above is also available from World Precision Instruments (model ISO-H₂S-2, Sarasota, FL, USA), which was connected to a multichannel Free Radical Analyzer (TBR4100, WPI Inc., Sarasota, FL, USA).³⁴ Moreover, this company developed an ultra-micro PHSS (ISO-H₂S-100-CXX, WPI, Inc., Sarasota, FL, USA) with a diameter of only 100 μm, which can obviously reduce the damage of the samples and offers several advantages,

including requiring no sleeves or filling solutions, durability for long-term use, rapid response time, and a bigger linear range. Compared to an ISE based H₂S sensor, this type of sensor detects H₂S without an external electrical potential, has a simple structure, good reproducibility, a short response time, and less containment of noble metals. However, a greater disadvantage of the liquid electrolyte sensors is that they leak easily, are prone to dry up, and have large residual current due to impurities in the solution.

3.3 Enzyme based electrochemical H₂S biosensors

Enzyme-based electrochemical biosensors have become more practical than other electrochemical detection methods due to their high sensitivity and selectivity in recent years. An enzyme-inhibition biosensor depends on both the concentration of the target analyte in the sample and the type of inhibited enzyme, which are commonly used for measurement of the concentration of inhibitory compounds.⁹¹ The common design of this type of H₂S sensor involves an electrochemical transducer and an enzyme as the recognition element. Early in the 1990s, cytochrome oxidase-based inhibited enzyme electrodes were studied by researchers with a detection limit concentration of H₂S down to 1 ppm in the gas phase. However, this electrode responds to other target analytes (HCN, and azide ion), making its application to biological samples questionable and developing a need for chemical pretreatment of samples.⁹² Horseradish peroxidase is stable and has a number of potential inhibitors in homogeneous solutions. Therefore, it has been widely used as an inhibited enzyme electrode by researchers for H₂S detection. Colloidal gold-immobilized horseradish peroxidase for H₂S detection has been reported by



Zhao *et al.*, but it seems more sensitive to cyanide and thiourea than sulfide. In addition, its detection limit is only 8 μM which is not enough for analyzing environmental samples.⁹³ Yang *et al.* developed an amperometric horseradish peroxidase inhibition H_2S biosensor based on a thiolate self-assembled monolayer (SAM), which has a much lower detection limit (0.3 μM). Nevertheless, the selectivity of SAM-based biosensors is poor for many common cations which greatly limits their application for analysis of environmental water.⁹⁴ Based on the method Yang *et al.* developed, Liu *et al.* reported a multi-layer film-modified biosensor *via* layer-by-layer (LBL) biospecific binding of concanavalin A and horseradish peroxidase on the surface of the gold electrode, which shows no interference of the common anions and cations in environmental water.⁹⁵ Coprinus cinereus peroxidase immobilized on a screen printed electrode was reported by Savizi *et al.* This was used as an enzyme inhibition H_2S biosensor which has a detection limit of 0.3 μM (linear response in the range of 1.09–16.3 μM) and a response time of 43 s.⁴⁷ Another enzyme inhibition H_2S biosensor, ascorbate oxidase, was immobilized on a Clark electrode and protected with a nylon membrane. This electrode can detect H_2S at a low detection limit and prevent interference of main cations and anions.⁹⁶ The enzyme based H_2S biosensor mentioned above shows great advantages of selectivity and sensitivity. However, one of the most regrettable characteristics of the enzyme inhibition-based biosensors is the different inhibition degrees caused by different inhibitors. Moreover, this kind of sensor also highly depends on the pH concentration due to its inhibition on the activity of enzyme, making it difficult to apply to *in vivo* H_2S detection.

4. Calibration of the H_2S electrode

Calibration of an H_2S sensor is important in order to ensure the exact experimental results. In general, the PHSS was calibrated with stepwise additions of H_2S stock solution at the pH and temperature of the subsequent experiment. Detection of the H_2S content in the sample was calculated from the calibration curve of current (pA) *vs.* concentration of H_2S . H_2S stock solution was freshly prepared by directly bubbling distilled water with pure H_2S gas (Matheson, Newark, CA) to make the saturated H_2S solution (0.09 M at 30 °C). The H_2S stock solution was diluted to different concentrations into cell culture medium and the pH of the medium was adjusted to 7.4. Stock solutions of H_2S can also be conveniently made with the salt, $\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$ (Sigma-Aldrich). This method avoids the need for a special gas-handling apparatus required for pure H_2S gas, and the additional micromolar sodium, as Na_2S , dissociates to form Na^+ and the three sulfide species H_2S , HS^- and S^{2-} in pH dependent ratios.⁷⁶ The calibration of the ISE based H_2S sensor should be carried out daily. Four 10 ml aliquots of 10 ppm S^{2-} standard solution were pipetted into 50 ml beakers. Electrodes were immersed in the solution and then measurements were recorded once the reading became stable (approx. 1 min). Then, 1 ml 100 ppm S^{2-} standard solu-

tion was immediately added and the second reading was recorded after 1 min. Finally, the resulting difference in the potential was used to calculate the exact concentration of the standardizing solution by applying the Nernst equation.⁵²

5. Characterization of H_2S electrodes

An excellent H_2S electrochemical sensor can be characterized by high sensitivity, high selectivity, low-detection limit, stability, long lifetime, wide linear range, fast response time, biocompatibility, and reproducibility.¹⁷ The sensitivity of the electrochemical H_2S sensor depends on the reactive surface, electrode material, the assisted membrane material, and pH of the solution. The measurement of the S^{2-} form of sulfide species using an ISE-based H_2S sensor requires an alkaline environment. A PHSS is also highly dependent on solution pH, and has a sensitivity of 5.5 nA μM^{-1} at pH = 6 where H_2S is approximately 90% of the total sulfide.¹⁷ However, high sensitivity does not imply a low detection limit; the low detection limit of the sensor can make it more practical for a wide variety of samples. The detection limit of a PHSS is dependent on the pH, in anoxic 10 mM PBS at pH 7.0 and 37 °C it was near 10 nM, while the detection limit of an ISE sensor could be 100 nM; both these sensors are sufficient for biological applications.^{34,55} Selectivity is usually controlled by the applied voltage and the selective layer provided by each kind of H_2S sensor. The acceptable voltage can reduce the noise signal level of other biological matrices oxidized on the electrode. An Ag/Ag₂S ISE H_2S electrode with a permissible Ag₂S membrane specifically allows S^{2-} ions to migrate across the thin layer which can effectively reduce interference from other biological matrices. The membranes of the PHSS are cemented together with silicone cement, which can prevent the interference from O^{2-} , NO, and millimolar concentrations of hydrogen peroxide, sulfite, and sulfate. An enzyme-based H_2S sensor with a special inhibited enzyme as the recognition element can make it just respond to H_2S .¹⁷ Linear range and response time are important before performing any measurement for accurate results, as a wide linear range means that the sensor can be applied in additional sample detection. A typical commercially available H_2S sensor from World Precision Instruments (PHSS, Sarasota, FL, USA) has a concentration range from 0.3 to 200 μM , which is much wider than that of ISE and enzyme based H_2S sensors. However, the amperometric signal loses linearity at high H_2S concentrations. The PHSS typically takes less than 10 s to reach 90% of the new signal in response to H_2S changes by addition or removal. Sensor stability, long lifetime, and reproducibility are important parameters in order to make sure that there is no sensitivity loss. PHSS has a loss of H_2S sensitivity and an elevated background current after several calibrations compared to earlier ones, these sensors require membrane or electrolyte replacement to ensure their sensitivity. To determine the precision of the PHSS response, 5 to 10 successive 2 μL injections of a 10 mM anoxic stock were made into a 2 mL anoxic chamber at pH 7.3 and 37 °C, yield-



ing 10 μM H_2S at each step, these stepwise injections produced a stepwise increase in current that differed by less than $\pm 2\%$ for each injection. Response accuracy was determined by periodically sampling the chamber solution and determining the H_2S concentration with 2-PDS. Calibration of the PHSS was within $\pm 1\%$ of the 2-PDS assay. Biocompatibility is also extremely important when measuring H_2S *in vivo*; the reactivity between the tissue and the electrochemical components must be minimal in order to verify there is no contribution to tissue inflammation. Since the PHSS electrochemical components are separated from sample solution by a selective membrane, there is no indication of altered biological processes.

6. Biological application of H_2S electrochemical sensors

Detection of sulfide in biosamples has gained significant attention from analytical and biological applications; currently, several techniques have been used to measure H_2S concentration in biological samples and biological sources like blood, serum, tissue, and breath.¹⁸ The continuous refinement of these techniques has improved detection specificity and lowered threshold limitations. An electrochemical method is still an effective and fast way to measure H_2S and its derivatives in biological samples.

6.1. Electrochemical detection of H_2S in plasma and blood

Detection of H_2S in plasma (or blood) has significantly enhanced the understanding of pathophysiological conditions and predicted clinical outcomes.³⁶ Sulfide species exist with an approximate ratio of 20% H_2S , 80% HS^- , and 0% S^{2-} under physiological conditions at 37 $^\circ\text{C}$ and pH 7.4. Similar to NO and CO, H_2S is lipophilic and permeates plasma membranes freely.¹⁹ Almost all studies reported that sulfide concentration in plasma (or blood) was between 10 and 300 μM by using the MBB methods, fluorescence probes, or ISE based methods.³⁷ However, MBB and fluorescence methods require practically irreversible derivative reactions and take free sulfide out of the sample, which induces liberating sulfide from some of the reversibly bound sulfide complexes. An ISE H_2S sensor is the most common method employed in plasma H_2S detection, which was first reported in the detection of rat plasma H_2S concentration to study the role of sulphide and sulphide oxidation in the copper molybdenum antagonism in 1987.⁵¹ Khan *et al.* made a full detailed procedure to rapidly estimate H_2S concentration in blood using a sulfide ion electrode (Model 94-16A).⁵² With the properties of great availability and easy operation, ISE H_2S electrochemical sensors have been widely used to detect plasma (or blood) H_2S concentration in rat or trout blood to study its biological function as shown in Table 1. However, one of the major problems that existed in detection of sulfide in plasma (or blood) is to overcome the interference of other thiol species (GSH, protein cys derivatives *etc.*). Therefore, relatively high blood sulfide concentrations were detected using ISE based H_2S sensors because the

required alkaline condition of these sensors released some sulfide from a proposed sulfide–biomolecule.⁹⁷ Free blood sulfide levels are much lower than the total species of sulfide. In order to overcome interference of biomolecules and other problems that ISE H_2S sensors face, the PHSS was developed with a sulfide-selective membrane to protect the electrode material from sample solution (section 3.2). Olson's research reported that the ISE may not accurately measure free sulfide in the plasma due to its rapid consumption by blood.⁵⁶ Furthermore, H_2S consumption was also O_2 dependent which was governed by the balance between constitutive production and oxidation.⁵⁶ Wintner *et al.* also detected low free sulfide levels in animal blood using the MBB method; they measured a similar rate of added sulfide loss in human plasma samples with the MBB method and a polarographic sensor.¹³

6.2. Electrochemical detection of H_2S in tissues and mitochondria

In order to better understand the biological roles of sulfide, it is necessary to build robust technologies that allow dynamic detection of endogenous sulfide in organs, tissues, or *in vivo*. However, dynamic measurements of sulfide in tissue samples are more problematic than that in the plasma (or blood) due to continuous enzymatic sulfide production and consumption processes. PHSSs, fluorescent probes, and MBB methods are shown to be able to detect sulfide levels in organs and tissues. Among these three methods, PHSS is well documented and is a reliable method to detect H_2S levels in tissues or organs as reported by Kraus's groups and the Olson laboratory.^{34,76,98} These sensors can be employed for real-time free H_2S detection with a low detection limit of 10 nM, and with great advantages of good reproducibility, short response, and a long linear range during detection without significant damage to the samples (section 3.2). With this approach, the production of H_2S was found in almost every organ system, such as trout gill and heart, bovine heart and lung, and rat and seal lung.^{17,18,34,76,78} In short, the detection of H_2S production using a PHSS in animal organ homogenate (or cells) was demonstrated by adding the substrate L-cysteine (L-cys) and the cofactor pyridoxal phosphate (PLP) for the enzymes cystathionine- β -synthase (CBS) and cystathionine γ lyase (CGL). These were used to dilute homogenate supernatant in a respirometer chamber using 5 μM O_2 , and the rise in the PHSS signal was recorded on a periodic basis. H_2S production rate was determined at the initial steepest slope of each trace. Recent studies on H_2S function suggested that the metabolism of H_2S could serve as an oxygen sensor in many cell types in various species. PHSS provided the first direct measurements showing that there is an inverse relationship between O_2 and H_2S , thus making it a potential oxygen sensor for both the pulmonary and systemic systems.^{18,56,98–100} The detection of H_2S consumption was exhibited by mitochondria isolated from the animal tissues (an inhabitant of sulfide-rich intertidal sediment) in an O_2^- dependent manner. Under air-equilibrated conditions, it was observed that mitochondrial H_2S consumption led to simultaneous increase in O_2 consumption, causing



some investigators to use H₂S-stimulated O₂ consumption as an indirect and largely qualitative measure of H₂S consumption. Using the PHSS to measure H₂S consumption as well as O₂ consumption provided a dynamic metabolic stoichiometry of H₂S to O₂ consumption; this demonstrated that H₂S can be consumed in the absence of O₂. This detailed assessment of mitochondrial H₂S metabolism provided evidence of specific enzymatic steps and insight into mammalian H₂S metabolism.

6.3. H₂S in modulation of plant physiological processes

H₂S is not just emerging as a third messenger molecule after nitric oxide (NO) and carbon monoxide (CO) involvement in modulation of physiological processes in animals systems,^{2–5} the role of H₂S metabolism in a plant system has also attracted attention in recent years. The positive effect of H₂S is also being identified as it is an important endogenous signalling molecule or bioregulator involved in many vital aspects of plant growth and responses against abiotic stresses.^{101,102} However, the metabolic pathways of H₂S at different developmental stages of plants under various environmental perturbations is unclear. The electrochemical method with several advantages has become an important detection method to explore the H₂S physiological processes in plants in recent years. Chen *et al.* used an electrochemical method (WPI, Inc., Sarasota, FL, USA) to detect the H₂S in spinacia oleracea seedlings, which provides an effective way to understand its further roles in physiological processes of photosynthesis such as promoting chloroplast biogenesis, photosynthetic enzyme expression, and thiol redox modification.⁸¹ Garcia-Mata *et al.* measured endogenous H₂S in leaf supernatant using a micro sulfide ion electrode (LIS-146AGSCM, Lazar Research Laboratories, Los Angeles, CA, USA) to study its functions of stomatal closure and participation in an ATP-binding cassette dependent signaling in guard cells.⁷² This method was also reported by Alvarez *et al.* to understand its specific role of sulfide as a general repressor of autophagy and a transcriptional modulator in Arabidopsis.⁷³

6.4 Electrochemical detection of H₂S for diagnoses and clinical therapeutic uses

The ultimate purpose of our study of H₂S metabolism in bacteria, plants, invertebrates, and fungi is to evaluate our understanding of the important role of H₂S in regulating human body function. Directly measuring H₂S concentration in human plasma (or tissue) provides an effective way to explore its biological role and its potential therapeutic uses. Once a statistically significant difference between patients and normal control subjects is observed, we can conclude that H₂S can be used as a biomarker for treatment decisions.^{36,39} An electrochemical H₂S sensor is a fast and effective method to detect H₂S in human plasma in order to explore the relationship between H₂S concentration and several diseases such as chronic obstructive pulmonary disease, coronary heart disease, hypertension, *etc.* Significantly higher H₂S concentration in serum was observed in patients with stable chronic obstructive pulmonary disease (COPD) than patients with acute exacerbation of COPD using an ISE H₂S sensor (Model 9616, Orion Research, Beverly, MA).^{58,103}

In addition, the study of age-matched control objects suggested that H₂S can be used as a surrogate marker with the need for antibiotics.^{58,68,103} A lower plasma H₂S concentration was observed in coronary heart disease (CHD) patients compared with angiographically normal control subjects by measuring H₂S concentration in 40 patients with CHD and 17 angiographically normal patients using a sulfide-sensitive electrode.⁵⁹ H₂S concentration was also reported related to essential hypertension; metabolic imbalance of homocysteine (Hcy) was markedly increased and the final product of this pathway (plasma H₂S) was significantly decreased in hypertensive children compared with normal subjects using an ionic meter (PXS-270, Shanghai Leici, China).^{61,104} Zhang *et al.* explored the predictive value of plasma H₂S in differentiating between vasovagal syncope (VVS) and postural orthostatic tachycardia syndrome (POTS) in children (PXS-270, Shanghai Leici, China). Plasma H₂S levels were significantly higher in children with VVS and POTS; the increased erythrocyte H₂S production may be involved in the pathogenesis of VVS in children.^{105,106} Han *et al.* found that the abnormal activation of the AKT/GSK3 β signal pathway in lymphocytes from systemic lupus erythematosus (SLE) patients plays an important role in the pathogenesis of the disease (ISO-H₂S-100, WPI). Benavides *et al.* explored human RBCs converting garlic-derived organic polysulfides into H₂S (measured in real time using a novel polarographic H₂S sensor), suggesting that the major beneficial effects (cardiovascular diseases *etc.*) of garlic rich diets are mediated by the biological production of H₂S from garlic-derived organic polysulfides.⁷⁷ All the evidence has indicated that electrochemical detection is becoming a leading method to detect H₂S for diagnoses and clinical therapeutic uses.

7. Conclusions

The enormous activity in the field of H₂S biosensors is a reflection of the major clinical importance of this topic. An electrochemical method provides an effective, direct, and responsive real-time detection of the concentration of H₂S without significant damage to living tissue. This approach is suitable for real-time detection of the concentration of H₂S in different kinds of biological samples, and has significantly enhanced the understanding of H₂S physiology functions. As researchers continue to unravel the complex biological roles of H₂S and develop therapies based on H₂S, the need for highly sensitive, highly selective, reproducible, biocompatible, and accurate H₂S measurement devices will increase in the future. Finally, the increasing clarification regarding H₂S in cells and tissues will lead to a better understanding of its role in the biological system. Due to the unique properties of electrochemical H₂S, the future of H₂S pathophysiology and chemical biology will be much clearer, and more research should be devoted to the detection of H₂S in blood and tissues to meet the increasing requirement of *in vivo* studies and clinical diagnoses.



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Notes and references

- V. Lantto and J. Mizsei, *Sens. Actuators, B*, 1991, **5**, 21–25.
- H. Kimura, *Mol. Neurobiol.*, 2002, **26**, 13–19.
- D. Boehning and S. H. Snyder, *Annu. Rev. Neurosci.*, 2003, **26**, 105–131.
- K. Abe and H. Kimura, *J. Neurosci.*, 1996, **16**, 1066–1071.
- R. Hosoki, N. Matsuki and H. Kimura, *Biochem. Biophys. Res. Commun.*, 1997, **237**, 527–531.
- C. Szabo, *Nat. Rev. Drug Discovery*, 2007, **6**, 917–935.
- H. Kimura, *Amino Acids*, 2011, **41**, 113–121.
- Y. Zhao, T. D. Biggs and M. Xian, *Chem. Commun.*, 2014, **50**, 11788–11805.
- T. Ubuka, *J. Chromatogr. Biomed. Appl.*, 2002, **781**, 227–249.
- J. Furne, A. Saeed and M. D. Levitt, *Am. J. Physiol.: Regul., Integr. Comp. Physiol.*, 2008, **295**, R1479–R1485.
- M. D. Levitt, M. S. Abdel-Rehim and J. Furne, *Antioxid. Redox Signaling*, 2011, **15**, 373–378.
- X. Shen, C. B. Pattillo, S. Pardue, S. C. Bir, R. Wang and C. G. Kevil, *Free Radicals Biol. Med.*, 2011, **50**, 1021–1031.
- E. A. Wintner, T. L. Deckwerth, W. Langston, A. Bengtsson, D. Leviten, P. Hill, M. A. Insko, R. Dumpit, E. VandenEckart, C. F. Toombs and C. Szabo, *Br. J. Pharmacol.*, 2010, **160**, 941–957.
- M. Nishida, T. Sawa, N. Kitajima, K. Ono, H. Inoue, H. Ihara, H. Motohashi, M. Yamamoto, M. Suematsu, H. Kurose, A. van der Vliet, B. A. Freeman, T. Shibata, K. Uchida, Y. Kumagai and T. Akaike, *Nat. Chem. Biol.*, 2012, **8**, 714–724.
- X. Shen, G. K. Kolluru, S. Yuan and C. G. Kevil, in *Methods Enzymol.*, ed. C. Enrique and P. Lester, Academic Press, 2015, vol. 554, pp. 31–45.
- X. Hu and B. Mutus, *Rev. Anal. Chem.*, 2013, **32**, 247–256.
- D. W. Kraus, J. E. Doeller and X. Zhang, in *Electrochemical Sensors, Biosensors and their Biomedical Applications*, ed. J. Wang, X. Zhang and J. Huangxian, Academic Press, San Diego, 2008, pp. 213–235.
- G. K. Kolluru, X. Shen, S. C. Bir and C. G. Kevil, *Nitric Oxide*, 2013, **35**, 5–20.
- K. R. Olson, *Biochim. Biophys. Acta*, 2009, **1787**, 856–863.
- K. R. Olson, *Antioxid. Redox Signaling*, 2012, **17**, 32–44.
- A. R. Lippert, E. J. New and C. J. Chang, *J. Am. Chem. Soc.*, 2011, **133**, 10078–10080.
- C. Liu, J. Pan, S. Li, Y. Zhao, L. Y. Wu, C. E. Berkman, A. R. Whorton and M. Xian, *Angew. Chem., Int. Ed.*, 2011, **50**, 10327–10329.
- H. Peng, Y. Cheng, C. Dai, A. L. King, B. L. Predmore, D. J. Lefer and B. Wang, *Angew. Chem., Int. Ed.*, 2011, **50**, 9672–9675.
- Y. Qian, J. Karpus, O. Kabil, S. Y. Zhang, H. L. Zhu, R. Banerjee, J. Zhao and C. He, *Nat. Commun.*, 2011, **2**, 495.
- K. Sasakura, K. Hanaoka, N. Shibuya, Y. Mikami, Y. Kimura, T. Komatsu, T. Ueno, T. Terai, H. Kimura and T. Nagano, *J. Am. Chem. Soc.*, 2011, **133**, 18003–18005.
- V. S. Lin, W. Chen, M. Xian and C. J. Chang, *Chem. Soc. Rev.*, 2015, **44**, 4596–4618.
- J. Wang, *Chem. Rev.*, 2008, **108**, 814–825.
- T. Xu, N. Scafa, L.-P. Xu, L. Su, C. Li, S. Zhou, Y. Liu and X. Zhang, *Electroanalysis*, 2014, **26**, 449–468.
- F. Bedioui and S. Griveau, *Electroanalysis*, 2013, **25**, 587–600.
- A. Malinauskas, R. Garjonyt, R. Mazeikien and I. Jureviciut, *Talanta*, 2004, **64**, 121–129.
- D. Lakshmi, M. J. Whitcombe, F. Davis, P. S. Sharma and B. B. Prasad, *Electroanalysis*, 2011, **23**, 305–320.
- M. K. Zachek, A. Hermans, R. M. Wightman and G. S. McCarty, *J. Electroanal. Chem.*, 2008, **614**, 113–120.
- W. Chen, S. Cai, Q. Q. Ren, W. Wen and Y. D. Zhao, *Analyst*, 2012, **137**, 49–58.
- J. E. Doeller, T. S. Isbell, G. Benavides, J. Koenitzer, H. Patel, R. P. Patel, J. R. Lancaster Jr., V. M. Darley-Usmar and D. W. Kraus, *Anal. Biochem.*, 2005, **341**, 40–51.
- C. E. Paulsen and K. S. Carroll, *Chem. Rev.*, 2013, **113**, 4633–4679.
- K. R. Olson, E. R. DeLeon and F. Liu, *Nitric Oxide*, 2014, **41**, 11–26.
- S. K. Pandey, K.-H. Kim and K.-T. Tang, *TrAC, Trends Anal. Chem.*, 2012, **32**, 87–99.
- E. Bitziou, M. B. Joseph, T. L. Read, N. Palmer, T. Mollart, M. E. Newton and J. V. Macpherson, *Anal. Chem.*, 2014, **86**, 10834–10840.
- B. D. Paul and S. H. Snyder, *Nat. Rev. Mol. Cell Biol.*, 2012, **13**, 499–507.
- M. Gubern, M. Andriamihaja, T. Nubel, F. Blachier and F. Bouillaud, *FASEB J.*, 2007, **21**, 1699–1706.
- D. J. Polhemus and D. J. Lefer, *Circ. Res.*, 2014, **114**, 730–737.
- J. W. Elrod, J. W. Calvert, J. Morrison, J. E. Doeller, D. W. Kraus, L. Tao, X. Jiao, R. Scalia, L. Kiss, C. Szabo, H. Kimura, C.-W. Chow and D. J. Lefer, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 15560–15565.
- C. K. Nicholson and J. W. Calvert, *Pharmacol. Res.*, 2010, **62**, 289–297.
- Y. Wang, X. Zhao, H. Jin, H. Wei, W. Li, D. Bu, X. Tang, Y. Ren, C. Tang and J. Du, *Arterioscler., Thromb., Vasc. Biol.*, 2009, **29**, 173–179.



- 45 R. J. Reiffenstein, W. C. Hulbert and S. H. Roth, *Annu. Rev. Pharmacol. Toxicol.*, 1992, **32**, 109–134.
- 46 A. V. Kroll, V. Smorchkov and A. Y. Nazarenko, *Sens. Actuators, B*, 1994, **21**, 97–100.
- 47 I. S. P. Savizi, H.-R. Kariminia, M. Ghadiri and R. Roosta-Azad, *Biosens. Bioelectron.*, 2012, **35**, 297–301.
- 48 J.-L. Chang, G.-T. Wei, T.-Y. Chen and J.-M. Zen, *Electroanalysis*, 2013, **25**, 845–849.
- 49 Y. Wang, H. Yan and E. F. Wang, *Sens. Actuators, B*, 2002, **87**, 115–121.
- 50 G. Li, B. J. Polk, L. A. Meazell and D. W. Hatchett, *J. Chem. Educ.*, 2000, **77**, 1049.
- 51 J. Mason, C. J. Cardin and A. Dennehy, *Res. Vet. Sci.*, 1978, **24**, 104–108.
- 52 S. U. Khan, G. F. Morris and M. Hidiroglou, *Microchem. J.*, 1980, **25**, 388–395.
- 53 W. Zhao, J. Zhang, Y. Lu and R. Wang, *EMBO J.*, 2001, **20**, 6008–6016.
- 54 G. Yang, L. Wu, B. Jiang, W. Yang, J. Qi, K. Cao, Q. Meng, A. K. Mustafa, W. Mu, S. Zhang, S. H. Snyder and R. Wang, *Science*, 2008, **322**, 587–590.
- 55 N. L. Whitfield, E. L. Kreimier, F. C. Verdial, N. Skovgaard and K. R. Olson, *Am. J. Physiol.: Regul., Integr. Comp. Physiol.*, 2008, **294**, R1930–R1937.
- 56 K. R. Olson, M. J. Healy, Z. Qin, N. Skovgaard, B. Vulesevic, D. W. Duff, N. L. Whitfield, G. Yang, R. Wang and S. F. Perry, *Am. J. Physiol.: Regul., Integr. Comp. Physiol.*, 2008, **295**, R669–R680.
- 57 R. A. Dombkowski, M. J. Russell and K. R. Olson, *Am. J. Physiol.: Regul., Integr. Comp. Physiol.*, 2004, **286**, R678–R685.
- 58 Y. H. Chen, W. Z. Yao, B. Geng, Y. L. Ding, M. Lu, M. W. Zhao and C. S. Tang, *Chest*, 2005, **128**, 3205–3211.
- 59 H. L. Jiang, H. C. Wu, Z. L. Li, B. Geng and C. S. Tang, *Di Yi Jun Yi Da Xue Xue Bao*, 2005, **25**, 951–954.
- 60 E. Distrutti, L. Sediari, A. Mencarelli, B. Renga, S. Orlandi, E. Antonelli, F. Roviezzo, A. Morelli, G. Cirino, J. L. Wallace and S. Fiorucci, *J. Pharmacol. Exp. Ther.*, 2006, **316**, 325–335.
- 61 L. Chen, S. Ingrid, Y. G. Ding, Y. Liu, J. G. Qi, C. S. Tang and J. B. Du, *Chin. Med. J.*, 2007, **120**, 389–393.
- 62 Q. H. Meng, G. Yang, W. Yang, B. Jiang, L. Wu and R. Wang, *Am. J. Pathol.*, 2007, **170**, 1406–1414.
- 63 B. Geng, Y. Cui, J. Zhao, F. Yu, Y. Zhu, G. Xu, Z. Zhang, C. Tang and J. Du, *Am. J. Physiol.: Regul., Integr. Comp. Physiol.*, 2007, **293**, R1608–R1618.
- 64 T. Li, B. Zhao, C. Wang, H. Wang, Z. Liu, W. Li, H. Jin, C. Tang and J. Du, *Exp. Biol. Med.*, 2008, **233**, 1081–1087.
- 65 G. P. Volpato, R. Searles, B. Yu, M. Scherrer-Crosbie, K. D. Bloch, F. Ichinose and W. M. Zapol, *Anesthesiology*, 2008, **108**, 659–668.
- 66 S. F. Perry, B. McNeill, E. Elia, A. Nagpal and B. Vulesevic, *Am. J. Physiol.: Regul., Integr. Comp. Physiol.*, 2009, **296**, R133–R140.
- 67 L. Wu, W. Yang, X. Jia, G. Yang, D. Duridanova, K. Cao and R. Wang, *Lab. Invest.*, 2009, **89**, 59–67.
- 68 Y.-H. Chen, W.-Z. Yao, J.-Z. Gao, B. Geng, P.-P. Wang and C.-S. Tang, *Respirology*, 2009, **14**, 746–752.
- 69 Y.-H. Chen, R. Wu, B. Geng, Y.-F. Qi, P.-P. Wang, W.-Z. Yao and C.-S. Tang, *Cytokine*, 2009, **45**, 117–123.
- 70 A. K. Mustafa, M. M. Gadalla, N. Sen, S. Kim, W. Mu, S. K. Gazi, R. K. Barrow, G. Yang, R. Wang and S. H. Snyder, *Sci. Signaling*, 2009, **2**, ra72.
- 71 T. Chang, A. Untereiner, J. Liu and L. Wu, *Antioxid. Redox Signaling*, 2010, **12**, 1093–1100.
- 72 C. Garcia-Mata and L. Lamattina, *New Phytol.*, 2010, **188**, 977–984.
- 73 C. Alvarez, I. Garcia, I. Moreno, M. E. Perez-Perez, J. L. Crespo, L. C. Romero and C. Gotor, *Plant Cell*, 2012, **24**, 4621–4634.
- 74 C. Hine, E. Harputlugil, Y. Zhang, C. Ruckenstein, B. C. Lee, L. Brace, A. Longchamp, J. H. Treviño-Villarreal, P. Mejia, C. K. Ozaki, R. Wang, V. N. Gladyshev, F. Madeo, W. B. Mair and J. R. Mitchell, *Cell*, 2015, **160**, 132–144.
- 75 P. Jeroschewski, C. Steuckart and M. Kühn, *Anal. Chem.*, 1996, **68**, 4351–4357.
- 76 D. W. Kraus and J. E. Doeller, *J. Exp. Biol.*, 2004, **207**, 3667–3679.
- 77 G. A. Benavides, G. L. Squadrito, R. W. Mills, H. D. Patel, T. S. Isbell, R. P. Patel, V. M. Darley-Usmar, J. E. Doeller and D. W. Kraus, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 17977–17982.
- 78 J. R. Koenitzer, T. S. Isbell, H. D. Patel, G. A. Benavides, D. A. Dickinson, R. P. Patel, V. M. Darley-Usmar, J. R. Lancaster, Jr., J. E. Doeller and D. W. Kraus, *Am. J. Physiol.: Heart Circ. Physiol.*, 2007, **292**, H1953–H1960.
- 79 K. R. Olson, N. L. Whitfield, S. E. Bearden, J. St Leger, E. Nilson, Y. Gao and J. A. Madden, *Am. J. Physiol.: Regul., Integr. Comp. Physiol.*, 2010, **298**, R51–R60.
- 80 M. Whiteman, L. Li, P. Rose, C. H. Tan, D. B. Parkinson and P. K. Moore, *Antioxid. Redox Signaling*, 2010, **12**, 1147–1154.
- 81 J. Chen, F. H. Wu, W. H. Wang, C. J. Zheng, G. H. Lin, X. J. Dong, J. X. He, Z. M. Pei and H. L. Zheng, *J. Exp. Bot.*, 2011, **62**, 4481–4493.
- 82 K. Suzuki, G. Olah, K. Modis, C. Coletta, G. Kulp, D. Gero, P. Szoleczky, T. Chang, Z. Zhou, L. Wu, R. Wang, A. Papapetropoulos and C. Szabo, *Proc. Natl. Acad. Sci. U. S. A.*, 2011, **108**, 13829–13834.
- 83 Y. Han, F. Zeng, G. Tan, C. Yang, H. Tang, Y. Luo, J. Feng, H. Xiong and Q. Guo, *Cell. Physiol. Biochem.*, 2013, **31**, 795–804.
- 84 C. Huang, J. Kan, X. Liu, F. Ma, B. H. Tran, Y. Zou, S. Wang and Y. Z. Zhu, *PLoS One*, 2013, **8**, e69205.
- 85 K. Modis, C. Coletta, K. Erdelyi, A. Papapetropoulos and C. Szabo, *FASEB J.*, 2013, **27**, 601–611.
- 86 K. R. Olson, E. R. DeLeon, Y. Gao, K. Hurley, V. Sadauskas, C. Batz and G. F. Stoy, *Am. J. Physiol.: Regul., Integr. Comp. Physiol.*, 2013, **305**, R592–R603.
- 87 U. Hasegawa and A. J. van der Vlies, *MedChemComm*, 2015, **6**, 273–276.



- 88 E. A. Ostrakhovitch, S. Akakura, R. Sanokawa-Akakura, S. Goodwin and S. Tabibzadeh, *Exp. Cell Res.*, 2015, **330**, 135–150.
- 89 Y. Sun, Y. Huang, R. Zhang, Q. Chen, J. Chen, Y. Zong, J. Liu, S. Feng, A. D. Liu, L. Holmberg, D. Liu, C. Tang, J. Du and H. Jin, *J. Mol. Med.*, 2015, **93**, 439–455.
- 90 P. Huang, S. Chen, Y. Wang, J. Liu, Q. Yao, Y. Huang, H. Li, M. Zhu, S. Wang, L. Li, C. Tang, Y. Tao, G. Yang, J. Du and H. Jin, *Nitric Oxide*, 2015, **46**, 192–203.
- 91 M. D. Luque de Castro and M. C. Herrera, *Biosens. Bioelectron.*, 2003, **18**, 279–294.
- 92 W. John Albery, A. E. G. Cass, B. P. Mangold and Z. X. Shu, *Biosens. Bioelectron.*, 1990, **5**, 397–413.
- 93 J. Zhao, R. W. Henkens and A. L. Crumbliss, *Biotechnol. Prog.*, 1996, **12**, 703–708.
- 94 Y. Yang, M. Yang, H. Wang, J. Jiang, G. Shen and R. Yu, *Sens. Actuators, B*, 2004, **102**, 162–168.
- 95 L. Liu, Z. Chen, S. Yang, X. Jin and X. Lin, *Sens. Actuators, B*, 2008, **129**, 218–224.
- 96 N. Z. M. Poor, L. Baniyadi, M. Omid, G. Amoabediny, F. Yazdian, H. Attar, A. Heydarzadeh, A. S. H. Zarami and M. H. Sheikhha, *Enzyme Microb. Technol.*, 2014, **63**, 7–12.
- 97 P. Nagy, Z. Pálincás, A. Nagy, B. Budai, I. Tóth and A. Vasas, *Biochim. Biophys. Acta*, 2014, **1840**, 876–891.
- 98 K. R. Olson, *J. Exp. Biol.*, 2008, **211**, 2727–2734.
- 99 K. R. Olson, R. A. Dombkowski, M. J. Russell, M. M. Doellman, S. K. Head, N. L. Whitfield and J. A. Madden, *J. Exp. Biol.*, 2006, **209**, 4011–4023.
- 100 K. R. Olson and N. L. Whitfield, *Antioxid. Redox Signaling*, 2010, **12**, 1219–1234.
- 101 Z.-G. Li, M. Gong, H. Xie, L. Yang and J. Li, *Plant Sci.*, 2012, **185–186**, 185–189.
- 102 L.-Y. Hu, S.-L. Hu, J. Wu, Y.-H. Li, J.-L. Zheng, Z.-J. Wei, J. Liu, H.-L. Wang, Y.-S. Liu and H. Zhang, *J. Agric. Food Chem.*, 2012, **60**, 8684–8693.
- 103 Y.-H. Chen, W.-Z. Yao, Y.-L. Ding, B. Geng, M. Lu and C.-S. Tang, *Pulm. Pharmacol. Ther.*, 2008, **21**, 40–46.
- 104 M. Zheng, Q. Zeng, X. Q. Shi, J. Zhao, C. S. Tang, N. L. Sun and B. Geng, *Chin. Med. J.*, 2011, **124**, 3693–3701.
- 105 F. Zhang, X. Li, C. Stella, L. Chen, Y. Liao, C. Tang, H. Jin and J. Du, *J. Pediatr.*, 2012, **160**, 227–231.
- 106 J. Yang, H. Li, T. Ochs, J. Zhao, Q. Zhang, S. Du, Y. Chen, P. Liu, Y. Wang, X. Feng, C. Zhang, C. Tang, J. Du and H. Jin, *J. Pediatr.*, 2015, **166**, 965–969.

