

FDA-approved poly(ethylene glycol)–protein conjugate drugs

Steevens N. S. Alconcel,^a Arnold S. Baas^{*b} and Heather D. Maynard^{*a}

Received 21st January 2011, Accepted 20th March 2011

DOI: 10.1039/c1py00034a

PEGylation or covalent attachment of poly(ethylene glycol) improves the pharmacokinetic properties of protein drugs. *In vivo* circulation lifetimes are increased and dosages are decreased, resulting in improved patient quality of life. PEG may be attached to proteins using a variety of different chemical reactions. This review discusses currently available FDA-approved PEGylated protein drugs, their intended use and target, and the PEG attachment chemistry utilized.

Introduction to PEGylation

PEGylation, or covalent attachment of poly(ethylene glycol) (PEG) to proteins, was introduced by Abuchowski and coworkers in 1977.¹ They reported on the superior immunogenic properties of bovine serum albumin (BSA)–PEG conjugates compared to the unmodified biomolecule. In later work Abuchowski and coworkers were able to demonstrate that PEGylation resulted in enhanced circulation lifetimes, as well as reduced immunogenicities relative to native proteins.² Since that time, many researchers have demonstrated that PEGylation improves pharmacological properties of proteins. As a result, several

FDA-approved protein–polymer drugs currently exist, and many others are currently being studied in clinical trials.³ This review focuses on the synthesis and mode of action of protein–PEG conjugates that are already FDA-approved for use to treat diseases in humans.

The main objective of using polymers in drug delivery is to stabilize and improve the therapeutic activity of the standalone drug.^{4–6} The non-ionic, hydrophilic PEG provides a steric shield for conjugates from recognition by the patient's immune system and effectively increases the size of the biomolecule,⁷ thereby reducing clearance from the bloodstream.⁸ By increasing the half-life of the protein drug, the dosage frequency is reduced.⁹ Since most protein drugs need to be injected, this result is significant. Increased duration of pharmacological activity, reduction of toxic side effects, and increased quality of life due to controlled, timed release are several positive results attributed to PEGylation. The detailed pharmacokinetics of PEGylated drugs have been recently reviewed.¹⁰ The dosage schedule and

^aDepartment of Chemistry & Biochemistry and the California NanoSystems Institute, University of California, 607 Charles E. Young Dr East, Los Angeles, CA, 90095, USA. E-mail: maynard@chem.ucla.edu

^bRonald Reagan UCLA Medical Center, Division of Cardiology, 757 Westwood Plaza, Los Angeles, CA, 90095, USA. E-mail: abaas@mednet.ucla.edu



Steevens N. S. Alconcel

Steevens N. S. Alconcel received a BS in Chemistry in 2007 from the University of Southern California, where he worked with Professor G. K. Surya Prakash on modification of Strecker and Mitsunobu reactions to synthesize fluorinated analogues. He is currently working toward his PhD in organic chemistry at the University of California, Los Angeles. Steevens is a trainee in the NIH-sponsored Chemistry–Biology Interface Training Program. His

research focuses on the synthesis of protein–polymer conjugates for drug delivery.



Arnold S. Baas

Arnold S. Baas received his BA from Baylor University in 1985 and his MD from UT Southwestern in 1989. He was resident, followed by chief resident at the University of Michigan before training as a fellow in cardiology at the University of Washington. Baas was a clinical assistant professor there before moving to UCLA to the David Geffen School of Medicine and Division of Cardiology. Baas' interests are focused on general cardiology, cardiac transplantation, mechanical circulatory

support, cardiomyopathy, atherosclerosis, cardiac rehabilitation, and cardiac imaging.

biological activity for each of the FDA-approved protein-PEG drugs are described in this review.

PEGylation may also decrease parent drug activity by changing conformation, sterically interfering, and altering electrostatic binding properties.³ Notably, the shape and number of PEG chains conjugated to a protein affects the stability and efficacy of the drug conjugate. Chain location is also critical to activity, as nonspecific PEGylation near an active site or to a region of a protein that causes a conformational change deleteriously affects the activity of the intended drug.⁹ As of today, there are well-established methods of PEGylation available,⁷ with a large array of different linking chemistries used.^{3,4,6,7,11,12} There has been a lot of recent effort placed on strategies that result in the site-specific conjugation.¹³ However, for many FDA-approved drugs, non-specific attachment strategies are used; these are described below and summarized in Table 1.

FDA-approved protein–polymer drugs

I. Enzymes

Adagen® (mPEG per adenosine deaminase). Adagen (pegademase bovine) is a modified enzyme drug sold by Enzon. The drug is used for the treatment of severe combined immunodeficiency disease (SCID) and was approved for such use in 1990. An autosomal recessive genetic disorder, SCID, is caused by adenosine deaminase deficiency. Native adenosine deaminase catalyzes the deamination of adenosine and deoxyadenosine. Lack of this natural enzyme results in lymphocyte toxicity leading to a severe reduction in immunity. It is usually fatal in children unless the patient is kept in protective isolation or undergoes a bone marrow transplant. As an alternative, Adagen is administered every 7 days intramuscularly. Adagen is not a cure for the disease; it is a replacement therapy that must be repeated for the rest of the patient's life.^{14,15} The dosing schedule is as follows. 10 U kg⁻¹, 15 U kg⁻¹, and 20 U kg⁻¹ for the first three doses, and the weekly maintenance dose is 20 U kg⁻¹.¹⁶

Initially, PEG was synthesized by reacting mPEG (monomethoxy-PEG) of molecular weight (M_w) 5000 with cyanuric chloride, forming PEG dichlorotriazine. In this approach, one of the two remaining chlorines on PEG dichlorotriazine is displaced by nucleophilic amino acid units such as lysine, serine, tyrosine,

cysteine, and histidine. The remaining chlorine is not as electrophilic, but may react to cause crosslinking of the protein. This is a nonspecific attachment process so multiple units of PEG are attached to the protein. Depending on the pH of the buffer used in the reaction, a small measure of control can be obtained to conjugate to specific nucleophilic amino acid units.^{1,14} This PEG was utilized at first by the company to study the drug, but is not what is used in the FDA-approved formulation.

The FDA-approved protein drug is synthesized using PEG succinimidyl succinate.⁷ This compound is prepared by reacting M_w 5000 mPEG with succinic anhydride. The resulting carboxylic acid is activated with *N*-hydroxysuccinimide (NHS) by carbodiimide coupling. The NHS group is displaced by nonspecific reaction with nucleophilic amino acid side chains.¹⁷

Oncaspar® (mPEG-L-asparaginase). Oncaspar (pegaspargase) by Enzon is classified as an antineoplastic drug used to treat acute lymphoblastic leukemia and was approved in 1994.^{17–20} It consists of a PEGylated L-asparaginase which is responsible for the hydrolysis of asparagine to aspartic acid. L-Asparaginase depletes asparagine, an amino acid essential for tumor growth. Native L-asparaginase^{18,21} has been shown to cause hypersensitivity reactions and shock. Oncaspar is an improvement upon the native enzyme. Phase I studies with PEGylated L-asparaginase showed increased serum half-life (357 h) relative to the native enzyme (20 h) with fewer cases of hypersensitivity.²² The drug can be delivered either intramuscularly or intravenously. The recommended dose is 2500 IU m⁻² and should be administered at the most every 14 days. If the volume administered is greater than 2 mL for intramuscular administration, multiple injection sites must be used.

The protein–polymer conjugate is also synthesized by the use of PEG succinimidyl succinate. The NHS group is displaced by nucleophilic amino acid units such as lysine, serine, cysteine, tyrosine, and histidine. The PEG ester and thioesters are highly susceptible to hydrolysis and thus modification occurs primarily at the amines forming amides.^{7,17}

Krystexxa® (pegloticase). Krystexxa (pegloticase) by Savient is a recombinant tetrameric urate oxidase used for the treatment of chronic gout refractory to conventional therapy that was FDA approved in 2010.²³ It consists of a PEGylated mammalian urate oxidase (uricase). This enzyme is responsible for the degradation of uric acid to allantoin and hydrogen peroxide.²⁴ Humans lack uricase and produce uric acid, which in high levels result in hyperuricemia.²⁵ Native urate oxidase has been used; but high concentrations are required, potentially causing serious allergic reactions or anaphylaxis. In addition, native uricase has a very short circulating half-life and is associated with inherent immunogenicity.²⁴ The PEGylated enzyme has an increased serum half-life (154–331 h) relative to the native enzyme (4 h). It also has reduced immunogenicity thereby minimizing allergic reactions.^{26,27}

The protein–polymer conjugate is synthesized by the use of a PEG *p*-nitrophenyl carbonate ester. The *p*-nitrophenol is displaced by primary amine lysine side chains, forming carbamates, which are subject to slow hydrolysis under mildly basic conditions.²⁸ Each subunit of urate oxidase contains a total of 28–29 lysines but only approximately 12 are surface-accessible in the native tetrameric form of the complete enzyme. In fact, some of



Heather D. Maynard

Heather D. Maynard received her BS degree from UNC-Chapel Hill in chemistry and an MS from UCSB in Materials Science. Maynard's PhD in chemistry was awarded in 2000 from Caltech, and she was an American Chemical Society postdoctoral fellow at the ETH Switzerland from 2000–2002. In 2002, Maynard moved to UCLA where she is now an Associate Professor in the Department of Chemistry and Biochemistry. Her interests include polymer synthesis, biohybrid materials, and surface modification.

of 20 000. To synthesize this linker (Fig. 1), monomethoxy PEG is reacted with *N,N'*-disuccinimidyl carbonate, resulting in PEG *N*-succinimidyl carbonate. Linking the PEG is accomplished by a one pot reaction with the side chain and *N*-terminal amine groups of lysine, forming a bis-carbamate.³² Finally, the carboxylic acid is activated with NHS, giving the branched PEG chain linker capable of forming stable amide bonds with 11 possible lysine residues. Four main sites of PEGylation have been identified, Lys 31, 121, 131, and 134. The reaction can be manipulated to favor formation of the monosubstituted conjugate by limiting the amount of PEG chain linker used in the conjugation step. The monosubstituted conjugate is then purified using ion-exchange chromatography, with 95–99% monoPEGylated conjugates.³⁰

PEG-INTRON® (peginterferon alfa-2b). PEG-INTRON by Schering-Plough (peginterferon alfa-2b) is also used to treat chronic hepatitis C alone or in conjunction with ribavirin. It was approved for use in 2001. Interferon alfa-2a and -2b half-lives are under 12 hours, so three times weekly subcutaneous injections are required. As a result, the quality of life of the patient is reduced.^{33–35} Monomethoxy-PEG-linked interferon has a sustained serum half-life (48–72 h) relative to the native protein (7–9 h).³⁴ The recommended dosage for standalone PEG-INTRON therapy is subcutaneous administration of 1 $\mu\text{g kg}^{-1}$ per week for 52 weeks on the same day of the week.³⁵

PEG-INTRON is a covalent conjugate of interferon alfa-2b linked to a single unit of M_w 12 000 PEG. Monomethoxy-PEG *N*-succinimidyl carbonate is subject to nucleophilic attack from several possible amino acid residues.³⁶ Histidines, serines, and lysines are possible PEG conjugation sites. In fact, there are 14 identified sites of PEGylation. The major isomer (47%) of PEG-INTRON is PEGylated at His 34.³⁴ The His 34 conjugate is most active, perhaps because of its location in the protein sequence which is not involved in receptor recognition.^{37,38} In addition, the His 34 conjugate slowly hydrolyzes and releases interferon alfa-2b over time.³⁷

III. Antibodies

Cimzia® (peg-anti-TNF α antibody fragment). Cimzia (certolizumab pegol) is a therapeutic monoclonal antibody drug used for the treatment of Crohn's disease and rheumatoid arthritis,

approved for use in 2008 and 2009, respectively. Tumor necrosis factor alpha (TNF α) is a pro-inflammatory cytokine involved in systemic inflammation, and is able to induce apoptotic cell death. As a result, it can be responsible for chronic inflammatory diseases. The antibody is designed and directed against TNF α , neutralizing the soluble and membrane-bound TNF α .³⁹ The recommended dose for treating Crohn's disease is 400 mg delivered subcutaneously in two injections of 200 mg initially and at week 2 and 4. Following this is a regimen of 400 mg every four weeks. The recommended dose for treating rheumatoid arthritis is 400 mg delivered subcutaneously in two injections of 200 mg initially and at week 2 and 4, followed by 200 mg administration every other week.^{40–42}

The active substance of Cimzia sold by UCB is certolizumab pegol consisting of a recombinant Fab' antibody fragment obtained and purified from *E. coli* expression. The fragment is covalently bound to a M_w 40 000 branched PEG chain through a *C*-terminal cysteine reaction with maleimide.⁴³ The synthesis of the branched PEG chain linker is as described for PEG-INTRON (*vide supra*). An additional step is included at the end, as an amino-maleimide linker is coupled to the lysine carboxylic acid.⁴⁴

IV. Growth factors

Neulasta® (pegfilgrastim). Neulasta (pegfilgrastim) is a modified granulocyte colony-stimulating factor (G-CSF) drug used to manage febrile neutropenia. It was approved for use in 2002 and is sold by Amgen. Febrile neutropenia is a condition found in patients with abnormally low counts of neutrophil granulocytes, usually developed during chemotherapy. PEGylated G-CSF artificially stimulates production of the neutrophil granulocytes. The PEGylated protein has an increased serum half-life (42 h) relative to the unmodified G-CSF (3.5–3.8 h). This allows for a single cycle dose that is as effective as daily doses of native G-CSF.^{45–48} The recommended dose of Neulasta is a single subcutaneous administration of 6 mg once-per-chemotherapy cycle. It should not be given within 14 days before and 24 days after administration of chemotherapeutics.⁴⁹

Neulasta is a PEGylated G-CSF synthesized by covalent attachment of M_w 20 000 monomethoxy-PEG aldehyde by reductive amination with the *N*-terminal methionine residue of the filgrastim protein. Targeting the *N*-terminal residue is



Fig. 1 Synthesis of the branched PEG used to prepare PEGASYS.

accomplished by exploiting the lower pK_a of this amine compared to the other amines such as lysines. Aldehyde-containing monomethoxy-PEG forms an imine with the *N*-terminal methionine residue and is subsequently reduced with sodium cyanoborohydride (Fig. 2).^{45,50}

Mircera® (continuous erythropoiesis receptor activator or methoxy polyethylene glycol-epoetin beta). Hoffmann–LaRoche's Mircera was approved in 2007. It is a PEGylated continuous erythropoietin (EPO) receptor activator (CERA)⁵¹ used to treat renal anemia in patients with chronic kidney disease (CKD). The PEGylated CERA protein combats anemia by stimulating erythropoiesis, or red blood cell growth, through interaction with the EPO receptor. Previously the disease was managed by blood transfusions as often as every two to three weeks or by recombinant EPO, administered up to three times weekly.⁵² The increased serum half-life of the PEGylated drug (134–139 h) compared to EPO (7–20 h) has reduced the frequency of administration required for the standalone protein.⁵³ The recommended dosage for patients who have not already started treatment with a non-PEGylated erythropoiesis stimulating agent (ESA) is single subcutaneous or intravenous administration of $0.6 \mu\text{g kg}^{-1}$ once every two weeks. Dosage should maintain hemoglobin levels between 10 and 12 g dL^{-1} . After the optimal hemoglobin concentration is achieved, Mircera should be administered once monthly at $1.2 \mu\text{g kg}^{-1}$. In the case of patients currently undergoing treatment with ESAs, Mircera may be administered once every two weeks or monthly to those patients where hemoglobin has previously been stabilized.⁵⁴

The PEG–CERA conjugate is synthesized by attachment of an NHS activated monomethoxy-PEG butanoic acid to Lys 46 and 52 on EPO.^{52,53,55}

Somavert® (pegvisomant). Somavert (pegvisomant) is a growth hormone (GH) receptor antagonist used for the treatment of acromegaly and approved for use in 2003. Acromegaly is a chronic metabolic disorder caused by excess concentration of human growth hormone (hGH) in the body. Excess hGH causes the body tissues to enlarge, creating conditions such as

disfigurement and premature death. This GH receptor antagonist enables the dimerization of the receptor at the cell surface, but does not allow the receptor to undergo the conformational change necessary for activation of the signaling pathway to occur.^{56–58} Although PEGylation reduces the activity of the GH receptor antagonist, immunogenicity and the rate of clearance from the body are greatly reduced, making it an effective drug against acromegaly.^{59,60} The recommended dosage for patients begins with a 40 mg loading dose administered subcutaneously. The patient will subcutaneously self-administer 10 mg of Somavert daily. Serum IGF-I concentrations should be measured every four to six weeks, with adjustments to the dosage of Somavert in 5 mg increments depending on the elevation or decline of insulin growth factor-1 (IGF-I) levels.^{61,62}

The pegvisomant conjugate sold by Pfizer is synthesized by covalent attachment of four to six M_w 5000 PEG units *via* NHS displacement as described for Adagen (*vide supra*) to several lysine residues available on hGH antagonist B2036, as well as the *N*-terminal phenylalanine residue. Possible sites of attachment include lysines 38, 41, 70, 115, 120, 140, 145, and 158, and phenylalanine 1.^{60,63–65}

Conclusions and outlook

Covalent attachment of PEG to proteins produces conjugates with significantly improved pharmacokinetic properties compared to the unmodified proteins. Specifically, PEG reduces clearance, slows enzymatic degradation and provides an effective shield from the immune system. Proteins must be administered by subcutaneous or intravenous injection, and longer half-lives lead to fewer injections. As a result there are a number of FDA-approved PEG–protein conjugates that have been reviewed herein with many other PEGylated drugs currently in phase I, II, and III trials that were not described.^{3,6,43} There are several chemistries that are utilized in FDA-approved PEG drugs, including amidation, reductive amination, and Michael addition, although most target multiple residues.

Nonspecific reaction of PEG with proteins leads to heterogeneous mixtures of multiple PEGs attached at different sites. This can lead to reductions in protein bioactivity. As a result, there has been a lot of effort placed on developing specific modification chemistries. An obvious choice is covalent attachment to free cysteines, as described above for Cimzia. The free cysteines can be naturally occurring or recombinantly placed away from the active site of a protein. PEGs with activated disulfide, maleimide, and vinyl sulfone end groups have been utilized for this purpose. In addition, proteins have either been modified chemically or produced recombinantly with artificial amino acids to present groups that react chemoselectively with PEGs.^{67,68} The field of incorporating a non-canonical amino acid and subsequent modification with PEG is still in the early stages of investigation. However, examples of chemistries that have been utilized are oxime bond formation and azide/alkyne click chemistries.^{69–71} These strategies produce homogeneous conjugates that retain bioactivity and are likely to play an increasing role in the protein–drug area.

PEG is classified to be Generally Regarded As Safe (GRAS) by the FDA and has been shown to be removed by renal clearance. Human studies show urinary excretion of PEG units up to

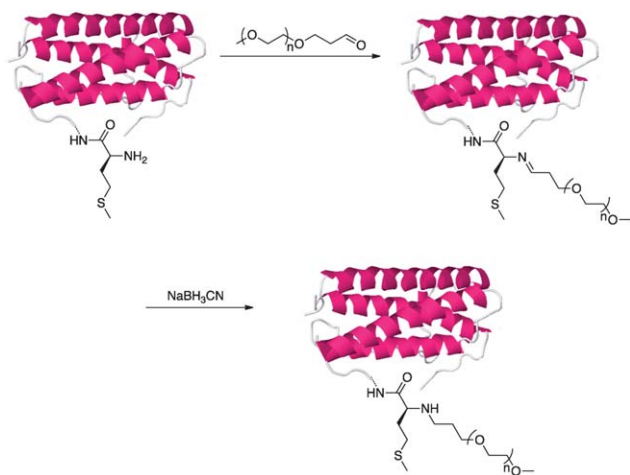


Fig. 2 Synthesis of Neulasta by imine formation and reductive amination with sodium cyanoborohydride.

20 kDa, with up to 190 kDa PEG seen excreted in mouse and rat studies.^{3,66} However, there are potential limitations of PEG including its non-degradability in the body, and these have been recently reviewed.³ It has been discussed that PEG of molecular weights below 40–60 kDa are required to minimize accumulation in the liver⁷² making it advantageous to utilize smaller branched PEGs, cleavable by hydrolysis, instead of larger, nondegradable linear PEGs to form the conjugates. Branched PEG chains may provide improved shielding from the immune system, reducing immunogenicity and antigenicity further, thus minimizing the size of the PEG required. In recent years controlled radical polymerization (CRP) techniques have been increasingly utilized to synthesize these branched PEG polymers. In particular, atom transfer radical polymerization (ATRP)^{73–75} and reversible addition–fragmentation chain transfer (RAFT) polymerization^{76–78} have been employed to make end chain reactive branched PEGs^{12,79,80} and to form bioconjugates directly by polymerization from modified proteins.^{81,82} In fact, recent studies have exploited the latter approach to polymerize branched poly(ethylene glycol) methyl ether methacrylate (PEGMA) *in situ* from therapeutic proteins such as recombinant human growth hormone (rh-GH)⁸³ and an intein fusion protein.⁸⁴ Importantly, this grafting from approach retains protein activity in many cases and eliminates the need for postpolymerization conjugation. These strategies lead to conjugates that in some cases are superior to their PEGylated analogs.⁸⁵ Recent reviews have summarized CRP approaches to bioconjugation.^{3,79,80,86} It is anticipated that CRPs will be increasingly utilized in the future to synthesize protein–polymer drugs.

Acknowledgements

The authors appreciate the National Institutes of Health (NCI R21 CA 137506-02) for funding. S.N.S.A. thanks the NIH sponsored Chemistry and Biology Interface (CBI) Training Program (2T32GM008496-16) for a fellowship.

References

- 1 A. Abuchowski, T. Vanes, N. C. Palczuk and F. F. Davis, *J. Biol. Chem.*, 1977, **252**, 3578–3581.
- 2 A. Abuchowski, J. R. McCoy, N. C. Palczuk, T. Vanes and F. F. Davis, *J. Biol. Chem.*, 1977, **252**, 3582–3586.
- 3 K. Knop, R. Hoogenboom, D. Fischer and U. S. Schubert, *Angew. Chem., Int. Ed.*, 2010, **49**, 6288–6308.
- 4 J. M. Harris, N. E. Martin and M. Modi, *Clin. Pharmacokinet.*, 2001, **40**, 539–551.
- 5 R. Duncan, *Nat. Rev. Drug Discovery*, 2003, **2**, 347–360.
- 6 R. Duncan, *Nat. Rev. Cancer*, 2006, **6**, 688–701.
- 7 M. J. Roberts, M. D. Bentley and J. M. Harris, *Adv. Drug Delivery Rev.*, 2002, **54**, 459–476.
- 8 G. Pasut and F. M. Veronese, *Isr. J. Chem.*, 2010, **50**, 151–159.
- 9 P. Caliceti and F. M. Veronese, *Adv. Drug Delivery Rev.*, 2003, **55**, 1261–1277.
- 10 C. S. Fishburn, *J. Pharm. Sci.*, 2008, **97**, 4167–4183.
- 11 J. M. Harris and R. B. Chess, *Nat. Rev. Drug Discovery*, 2003, **2**, 214–221.
- 12 M. A. Gauthier and H. A. Klok, *Chem. Commun.*, 2008, 2591–2611.
- 13 K. L. Heredia and H. D. Maynard, *Org. Biomol. Chem.*, 2007, **5**, 45–53.
- 14 S. Davis, A. Abuchowski, Y. K. Park and F. F. Davis, *Clin. Exp. Immunol.*, 1981, **46**, 649–652.
- 15 B. E. Bax, M. D. Bain, L. D. Fairbanks, A. D. B. Webster and R. A. Chalmers, *Br. J. Haematol.*, 2000, **109**, 549–554.
- 16 M. S. Hershfield, R. H. Buckley, M. L. Greenberg, A. L. Melton, R. Schiff, C. Hatem, J. Kurtzberg, M. L. Markert, R. H. Kobayashi, A. L. Kobayashi and A. Abuchowski, *N. Engl. J. Med.*, 1987, **316**, 589–596.
- 17 F. F. Davis, T. Van Es and N. C. Palczuk, *US Pat.*, 4179337, 1979.
- 18 H. F. Oettgen, L. J. Old, E. A. Boyse, H. A. Campbell, F. S. Philips, B. D. Clarkson, L. Tallal, R. D. Leeper, M. K. Schwartz and J. H. Kim, *Cancer Res.*, 1967, **27**, 2619.
- 19 L. J. Ettinger, J. Kurtzberg, P. A. Voute, H. Jurgens and S. L. Halpern, *Cancer*, 1995, **75**, 1176–1181.
- 20 P. A. Dinndorf, J. Gootenberg, M. H. Cohen, P. Keegan and R. Pazdur, *Oncologist*, 2007, **12**, 991–998.
- 21 J. D. Broome, *Nature*, 1961, **191**, 1114.
- 22 D. H. Ho, N. S. Brown, A. Yen, R. Holmes, M. Keating, A. Abuchowski, R. A. Newman and I. H. Krakoff, *Drug Metab. Dispos.*, 1986, **14**, 349–352.
- 23 N. Schlesinger, U. Yasothan and P. Kirkpatrick, *Nat. Rev. Drug Discovery*, 2011, **10**, 17–18.
- 24 M. S. Hershfield, J. S. Sundry, N. J. Ganson and S. J. Kelly, in *PEGylated Protein Drugs: Basic Science and Clinical Applications*, 2009, pp. 217–227.
- 25 M. R. Sherman, M. G. P. Saifer and F. Perez-Ruiz, *Adv. Drug Delivery Rev.*, 2008, **60**, 59–68.
- 26 J. S. Sundry, M. A. Becker, H. S. B. Baraf, A. Barkhuizen, L. W. Moreland, W. Huang, R. W. Waltrip, A. N. Maroli, Z. Horowitz and Pegloticase Phase 2 Study Investigators, *Arthritis Rheum.*, 2008, **58**, 2882–2891.
- 27 J. S. Sundry, N. J. Ganson, S. J. Kelly, E. L. Scarlett, C. D. Rehrig, W. Huang and M. S. Hershfield, *Arthritis Rheum.*, 2007, **56**, 1021–1028.
- 28 L. D. Williams, M. S. Hershfield, S. J. Kelly, M. G. P. Saifer and M. R. Sherman, *US Pat.*, 7723089, 2010.
- 29 M. W. Fried, M. L. Shiffman, K. R. Reddy, C. Smith, G. Marinos, F. L. Goncalves, D. Haussinger, M. Diago, G. Carosi, D. Dhumeaux, A. Craxi, A. Lin, J. Hoffman and J. Yu, *N. Engl. J. Med.*, 2002, **347**, 975–982.
- 30 K. R. Reddy, M. W. Modi and S. Pedder, *Adv. Drug Delivery Rev.*, 2002, **54**, 571–586.
- 31 S. Zeuzem, S. V. Feinman, J. Rasenack, E. J. Heathcote, M. Y. Lai, E. Gane, J. O'Grady, J. Reichen, M. Diago, A. Lin, J. Hoffman and M. J. Brunda, *N. Engl. J. Med.*, 2000, **343**, 1666–1672.
- 32 J. M. Harris, F. M. Veronese, P. Caliceti and O. Schiavon, *US Pat.*, 5932462, 1999.
- 33 M. P. Manns, J. G. McHutchison, S. C. Gordon, V. K. Rustgi, M. Shiffman, R. Reindollar, Z. D. Goodman, K. Koury, M. H. Ling and J. K. Albrecht, *Lancet*, 2001, **358**, 958–965.
- 34 Y. S. Wang, S. Youngster, M. Grace, J. Bausch, R. Bordens and D. F. Wyss, *Adv. Drug Delivery Rev.*, 2002, **54**, 547–570.
- 35 R. M. Bukowski, C. Tendler, D. Cutler, E. Rose, M. M. Laughlin and P. Statkevich, *Cancer*, 2002, **95**, 389–396.
- 36 C. W. Gilbert and M. Park-Cho, *US Pat.*, 5951974, 1999.
- 37 G. Pasut, in *PEGylated Protein Drugs: Basic Science and Clinical Applications*, 2009, pp. 205–216.
- 38 M. Grace, S. Youngster, G. Gitlin, W. Sydor, L. Xie, L. Westreich, S. Jacobs, D. Brassard, J. Bausch and R. Bordens, *J. Interferon Cytokine Res.*, 2001, **21**, 1103–1115.
- 39 A. Nesbitt, G. Fossati, M. Bergin, P. Stephens, S. Stephens, R. Foulkes, D. Brown, M. Robinson and T. Bourne, *Inflammatory Bowel Dis.*, 2007, **13**, 1323–1332.
- 40 W. J. Sandborn, B. G. Feagan, S. Stoinov, P. J. Honiball, P. Rutgeerts, D. Mason, R. Bloomfield, S. Schreiber and P. S. Investigators, *N. Engl. J. Med.*, 2007, **357**, 228–238.
- 41 S. Schreiber, P. Rutgeerts, R. N. Fedorak, M. Khaliq-Kareemi, M. A. Kamm, M. Boivin, C. N. Bernstein, M. Staun, O. O. Thomsen, A. Innes and C. D. P. C. D. S. Grp, *Gastroenterology*, 2005, **129**, 807–818.
- 42 S. Schreiber, M. Khaliq-Kareemi, I. C. Lawrance, O. O. Thomsen, S. B. Hanauer, J. McColm, R. Bloomfield, W. J. Sandborn and P. S. Investigators, *N. Engl. J. Med.*, 2007, **357**, 239–250.
- 43 F. M. Veronese and A. Mero, *BioDrugs*, 2008, **22**, 315–329.
- 44 D. S. Athwal, D. Brown, A. N. C. Weir, A. G. Poplewell, A. P. Chapman and D. J. King, *US Pat.*, 7012135, 2006.
- 45 G. Molineux, *Curr. Pharm. Des.*, 2004, **10**, 1235–1244.
- 46 C. L. Vogel, M. Z. Wojtukiewicz, R. R. Carroll, S. A. Tjulandin, L. J. Barajas-Figueroa, B. L. Wiens, T. A. Neumann and L. S. Schwartzberg, *J. Clin. Oncol.*, 2005, **23**, 1178–1184.

- 47 M. D. Green, H. Koelbl, J. Baselga, A. Galid, V. Guillem, P. Gascon, S. Siena, R. I. Lalisang, H. Samonigg, M. R. Clemens, V. Zani, B. C. Liang, J. Renwick, M. J. Piccart and International Pegfilgrastim 749 Study Group, *Ann. Oncol.*, 2003, **14**, 29–35.
- 48 F. A. Holmes, J. A. O'Shaughnessy, S. Vukelja, S. E. Jones, J. Shogan, M. Savin, J. Glaspy, M. Moore, L. Meza, I. Wiznitzer, T. A. Neumann, L. R. Hill and B. C. Liang, *J. Clin. Oncol.*, 2002, **20**, 727–731.
- 49 K. Welte, J. Gabrielove, M. H. Bronchud, E. Platzer and G. Morstyn, *Blood*, 1996, **88**, 1907–1929.
- 50 O. B. Kinstler, N. E. Gabriel, C. E. Farrar and R. B. DePrince, *US Pat.*, 5824784, 1998.
- 51 F. Locatelli, G. Villa, A. L. M. de Francisco, A. Albertazzi, H. J. Adrogoe, F. C. Dougherty and U. Beyer, *Curr. Med. Res. Opin.*, 2007, **23**, 969–979.
- 52 I. C. Maccougall and K. U. Eckardt, *Lancet*, 2006, **368**, 947–953.
- 53 I. C. Maccougall, R. Robson, S. Opatrna, X. Liogier, A. Pannier, P. Jordan, F. C. Dougherty and B. Reigner, *Clin. J. Am. Soc. Nephrol.*, 2006, **1**, 1211–1215.
- 54 J. Hertel, H. Locay, D. Scarlata, L. Jackson, R. Prathikanti and P. Audhya, *Am. J. Nephrol.*, 2006, **26**, 149–156.
- 55 I. C. Maccougall, P. Bailon, N. Tare, W. Pahlke and J. Pill, *J. Am. Soc. Nephrol.*, 2003, **14**, 769A.
- 56 W. Y. Chen, D. C. Wight, T. E. Wagner and J. J. Kopchick, *Proc. Natl. Acad. Sci. U. S. A.*, 1990, **87**, 5061–5065.
- 57 J. J. Kopchick and W. Y. Chen, *US Pat.*, 5958879, 1999.
- 58 J. J. Kopchick and W. Y. Chen, *US Pat.*, 6583115, 2003.
- 59 R. J. M. Ross, K. C. Leung, M. Maamra, W. Bennett, N. Doyle, M. J. Waters and K. K. Y. Ho, *J. Clin. Endocrinol. Metab.*, 2001, **86**, 1716–1723.
- 60 S. Pradhananga, I. Wilkinson and R. J. M. Ross, *J. Mol. Endocrinol.*, 2002, **29**, 11–14.
- 61 P. J. Trainer, W. M. Drake, L. Katznelson, P. U. Freda, V. Herman-Bonert, A. J. van der Lely, E. V. Dimaraki, P. M. Stewart, K. E. Friend, M. L. Vance, G. M. Besser, J. A. Scarlett, M. O. Thorner, C. Parkinson, A. Klibanski, J. S. Powell, A. L. Barkan, M. C. Sheppard, M. Maldonado, D. R. Rose, D. R. Clemmons, G. Johannsson, B. A. Bengtsson, S. Stavrou, D. L. Kleinberg, D. M. Cook, L. S. Phillips, M. Bidlingmaier, C. J. Strasburger, S. Hackett, K. Zib, W. F. Bennett and R. J. Davis, *N. Engl. J. Med.*, 2000, **342**, 1171–1177.
- 62 A. J. van der Lely, R. K. Hutson, P. J. Trainer, G. M. Besser, A. L. Barkan, L. Katznelson, A. Klibanski, V. Herman-Bonert, S. Melmed, M. L. Vance, P. U. Freda, P. M. Stewart, K. E. Friend, D. R. Clemmons, G. Johannsson, S. Stavrou, D. M. Cook, L. S. Phillips, C. J. Strasburger, S. Hacker, K. A. Zib, R. J. Davis, J. A. Scarlett and M. Thorner, *Lancet*, 2001, **358**, 1754–1759.
- 63 G. Fuh, B. C. Cunningham, R. Fukunaga, S. Nagata, D. V. Goeddel and J. A. Wells, *Science*, 1992, **256**, 1677–1680.
- 64 B. C. Cunningham, H. B. Lowman, J. A. Wells, R. G. Clark, K. Olson and G. G. Fuh, *US Pat.*, 5849535, 1998.
- 65 B. C. Cunningham, H. B. Lowman, J. A. Wells, R. G. Clark, K. Olson and G. G. Fuh, *US Pat.*, 6057292, 2000.
- 66 R. Webster, E. Didier, P. Harris, N. Siegel, J. Stadler, L. Tilbury and D. Smith, *Drug Metab. Dispos.*, 2007, **35**, 9–16.
- 67 R. E. Connor and D. A. Tirrell, *Polym. Rev.*, 2007, **47**, 9–28.
- 68 T. S. Young and P. G. Schultz, *J. Biol. Chem.*, 2010, **285**, 11039–11044.
- 69 G. G. Kochendoerfer, S. Y. Chen, F. Mao, S. Cressman, S. Traviglia, H. Y. Shao, C. L. Hunter, D. W. Low, E. N. Cagle, M. Carnevali, V. Gueriguian, P. J. Keogh, H. Porter, S. M. Stratton, M. C. Wiedeke, J. Wilken, J. Tang, J. J. Levy, L. P. Miranda, M. M. Crnogorac, S. Kalbag, P. Botti, J. Schindler-Horvat, L. Savatski, J. W. Adamson, A. Kung, S. B. H. Kent and J. A. Bradburne, *Science*, 2003, **299**, 884–887.
- 70 R. A. Scheck, M. T. Dedeo, A. T. Lavarone and M. B. Francis, *J. Am. Chem. Soc.*, 2008, **130**, 11762–11770.
- 71 B. Parrish, R. B. Breitenkamp and T. Emrick, *J. Am. Chem. Soc.*, 2005, **127**, 7404–7410.
- 72 G. Pasut and F. M. Veronese, *Prog. Polym. Sci.*, 2007, **32**, 933–961.
- 73 K. Matyjaszewski, T. E. Patten and J. H. Xia, *J. Am. Chem. Soc.*, 1997, **119**, 674–680.
- 74 V. Coessens, T. Pintauer and K. Matyjaszewski, *Prog. Polym. Sci.*, 2001, **26**, 337–377.
- 75 K. Matyjaszewski and J. H. Xia, *Chem. Rev.*, 2001, **101**, 2921–2990.
- 76 G. Moad, E. Rizzardo and S. H. Thang, *Aust. J. Chem.*, 2005, **58**, 379–410.
- 77 G. Moad, E. Rizzardo and S. H. Thang, *Polymer*, 2008, **49**, 1079–1131.
- 78 G. Moad, E. Rizzardo and S. H. Thang, *Acc. Chem. Res.*, 2008, **41**, 1133–1142.
- 79 R. M. Broyer, G. N. Grover and H. D. Maynard, *Chem. Commun.*, 2011, **47**, 2212–2226.
- 80 G. N. Grover and H. D. Maynard, *Curr. Opin. Chem. Biol.*, 2011, **14**, 818–827.
- 81 D. Bontempo and H. D. Maynard, *J. Am. Chem. Soc.*, 2005, **127**, 6508–6509.
- 82 J. Q. Liu, V. Bulmus, D. L. Herlambang, C. Barner-Kowollik, M. H. Stenzel and T. P. Davis, *Angew. Chem., Int. Ed.*, 2007, **46**, 3099–3103.
- 83 J. P. Magnusson, S. Bersani, S. Salmaso, C. Alexander and P. Caliceti, *Bioconjugate Chem.*, 2010, **21**, 671–678.
- 84 W. P. Gao, W. G. Liu, T. Christensen, M. R. Zalutsky and A. Chilkoti, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 16432–16437.
- 85 V. Depp, A. Alikhani, V. Grammer and B. S. Lele, *Acta Biomater.*, 2009, **5**, 560–569.
- 86 B. Le Droumaguet and J. Nicolas, *Polym. Chem.*, 2010, **1**, 563–598.