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Lipid vesicle-based molecular robots

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A molecular robot, which is a system comprised of one or more molecular machines and computers, can execute sophisticated tasks in many fields that span from nanomedicine to green nanotechnology. The core parts of molecular robots are fairly consistent from system to system and always include (i) a body to encapsulate molecular machines, (ii) sensors to capture signals, (iii) computers to make decisions, and (iv) actuators to perform tasks. This review aims to provide an overview of approaches and considerations to develop molecular robots. We first introduce the basic technologies required for constructing the core parts of molecular robots, describe the recent progress towards achieving higher functionality, and subsequently discuss the current challenges and outlook. We also highlight the applications of molecular robots in sensing biomarkers, signal communications with living cells, and conversion of energy. Although molecular robots are still in their infancy, they will unquestionably initiate massive change in biomedical and environmental technology in the not too distant future.

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Introduction

Machines equipped to perform specific actions have disengaged humans from exhausting labor, helping us accomplish tasks that would be impossible with only human hands. The revolution in mechanization began in the second half of the 18th century and has led to previously unimaginable improvements in humankind's quality of life. During the past 40 years, this revolution has led to a new generation of machines with smaller sizes, pushing the boundaries of applications. In the field of organic chemistry, a breakthrough molecular catenane with two interlocked rings was reported by Jean-Pierre Sauvage in the early 1980s,¹ subsequently inspiring other scientists to develop molecular-scale machines with complicated functions such as rotaxanes,² motors,³ and

nanocars.⁴ In 2016, three pioneers of molecular machines were awarded the Nobel Prize in Chemistry, reflecting the recognition of the broad impact of molecular machines.

The concept of molecular machines has motivated research activity in a novel field called "molecular robots".⁵ According to the Cambridge dictionary, a robot is "a machine controlled by a computer that is used to perform jobs automatically". By this definition, a molecular robot is a system composed of molecular-scale machines and computers that are used to execute tasks automatically. A living cell could be considered as one such miraculous robot produced by nature. With DNA serving as computers to provide solutions and proteins working as machines to perform specific functions, a living cell performs sophisticated tasks independent of human control. Taking inspiration from living cells, the ultimate goal of the field of molecular robots is to artificially construct an automated system capable of solving problems at the molecular level using molecular machines and computers.^{6,7}

A molecular robot always includes some or all of the following: a body, sensors, computers, and actuators (Fig. 1). Pioneers in the field have applied hydrogels as the body of molecular robots,⁵ however, the lack of a barrier between the embeddings and environment can lead to undesired leakage. Lipid vesicles, which are comprised of lipid membranes separating an inner lumen from the outer solution, provides an alternative that could fully meet this problem. The size of the lipid vesicles can be tailored from nano- to micro-meter in diameter. For molecular robots, the micro-sized vesicles,

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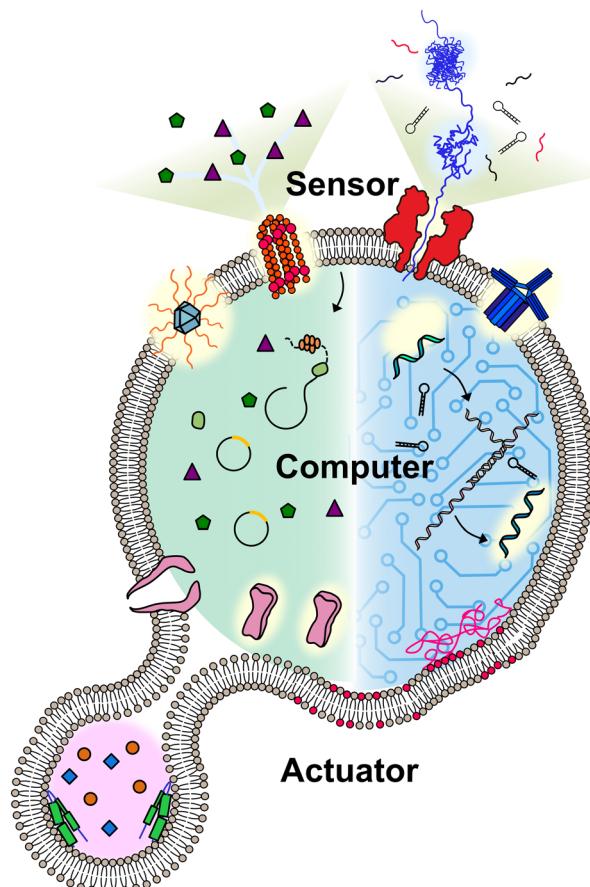


Fig. 1 Conceptual illustration of a molecular robot including: a body to encapsulate molecular machinery, sensors to collect signals, computers to make decisions, and actuators to perform the tasks.

so-called giant unilamellar vesicles (GUVs), are more desirable due to the demand for sufficient internal volume to house the sensors, computers, and actuators.^{8,9} Molecular robots require sensors in order to detect signals in the environment. This can be accomplished by ion channels or nanopores, which punch holes in lipid membranes. Nanopores can act as a signal filter, selectively transporting molecular signals based on their size or charge.^{10,11} Once the signals are transported, they can be processed and translated by molecular computing machinery. DNA computing, pioneered by L. Adleman in 1994,¹² has evolved in recent decades into computers applicable in molecular robots, with the benefit of their capability to perform multiple parallel computations.^{12,13} An alternative choice for the computing machinery is cell-free protein synthesis (CFPS), enabling the output of proteins *in vitro* in response to the input of DNA.¹⁴ Actuators for molecular robots, which include DNA nanostructures,¹⁵ peptides,¹⁶ and proteins,¹⁷ convert signals to achieve physical movements like deformation or propulsion.

Although constant progress has been made on the underlying technology, studies into prototyping molecular

robots do not have a long history. In 2014, Nishimura *et al.* incorporated CFPS into GUVs, and then, in the presence of amino acids, GFP synthesis was performed as an output.¹⁸ In 2017, Sato *et al.* developed GUVs equipped with actuators containing DNA clutches and microtubule motors.¹⁵ Once light irradiation was applied, the clutch was engaged and the shape of GUVs underwent sequential changes. These pioneering studies provide a clear path towards molecular robots with increasingly complex functions.

It should be noted that molecular robots sometimes share related technologies with the field of artificial cells. The goal of constructing artificial cells is to mimic the function of living cells, while molecular robots place great value on developing engineering applications that could help humans to perform tasks in micro or nano dimensions. There is therefore a drive to engineer molecular robots with functions exceeding those of living cells (for details on artificial cells please refer to previous reviews^{19,20}). This review focuses on the recent progress and challenges in the core technologies of molecular robots and explores emerging directions. We also highlight some applications of molecular robots such as molecular sensing, communications with living cells, and energy conversion. Finally, we close with a discussion on potential research trends in this field during the coming decade.

Body of molecular robots

Molecular robots typically require encapsulation within a compartment, which acts as a boundary, separating the interior from the exterior environment. Various types of compartments have been employed to date, including lipid membranes,²¹ hydrogels,²² block co-polymers,²³ DNA droplets,²⁴ and coacervates,²⁵ each offering distinct advantages and limitations. Some researchers have explored the formation of hybrid chassis by combining different compartment types, leveraging the advantages associated with each constituent part. For instance, coacervate or DNA/hydrogel systems interfaced with lipid membranes can be combined to enhance functionality.^{26–28}

Among these compartment types, lipid vesicles are the most commonly used for several reasons. Firstly, they are biomimetic, closely resembling biological membranes from a chemical and morphological perspective. This characteristic enables facile incorporation of membrane-bound molecular machinery, including membrane proteins, nanopores, and receptors, thereby imparting specific functionalities into the membranes.^{29,30} For example, the controlled flux of cargo molecules in response to stimuli,^{30,31} which can be used to mediate responses in living cells.³²

Furthermore, lipid vesicles are chemically inert and highly efficient at compartmentalizing large charged molecules from the surrounding environment, creating a chemically distinct internal environment. This feature enables researchers to exploit the diversity of lipid building blocks, both synthetic and biological, to create functional membranes with diverse



behaviors. Examples include membranes capable of self-assembling into tissue-like structures,³³ membranes that can disassemble and reassemble in response to physicochemical cues to reshuffle material between them,³⁴ and membranes that release cargo triggered by light, temperature fluctuations, magnetic fields, or biomarkers.³¹ Such versatility opens up exciting possibilities for molecular robotics and targeted drug delivery systems, among other applications.

Vesicles can be classified primarily based on their size and lamellarity.³⁵ GUVs have a diameter of approximately 2 μm and above (making them cell-sized vesicles), comprising a single lipid membrane (as opposed to multi-layered onion-like structures known as multilamellar vesicles). Other types of vesicles include small and large unilamellar vesicles, which fall into the sub-micron size range, as well as multi-vesicular vesicles (multisomes). Additionally, there have been intriguing examples of hybrid structures, where vesicles of different types are assembled into more architecturally complex arrangements, such as nested³⁶ or layered geometries.³⁷ A schematic of the different architectures it is now possible to generate microfluidic techniques and principles in biomembrane engineering is shown in Fig. 2.

In molecular robotics, GUVs are often the go-to architecture due to their resolvability by optical microscopy. Moreover, they allow for efficient encapsulation of large and highly charged building blocks (DNA, proteins, nanoparticles, and even whole organelles/cells),^{38,39} which becomes more problematic when using vesicles in the sub-micron regime. For instance, to encapsulate the biomolecular components required for transcription and translation, precise stoichiometries of plasmids, ribosomes, polymerases, and around 40 other molecular components must be co-encapsulated.⁴⁰ Achieving such precision becomes statistically improbable in lower-size regimes. GUVs thus

provide a more suitable environment for accommodating these complex biomolecular systems.

Vesicle generation strategies

To generate GUVs, various methods are available, with several comprehensive reviews published in this area.^{41–43} When evaluating the different methods, it is essential to consider the relevant Key Performance Indicators (KPIs), which include:

(i) **Encapsulation efficiency.** The method's efficacy in encapsulating molecules may vary based on the type of molecule involved. In molecular robotics applications, achieving a high encapsulation efficiency for large, charged species (such as DNA, proteins, nanoparticles, *etc.*) is crucial, as they contribute to the system's functionality.

(ii) **Monodispersity and control over size.** The ability to produce vesicles with well-defined dimensions is of paramount importance, as size directly influences downstream applications, and consistent sizes ensure consistent performance.

(iii) **Control of architecture and lipid content.** Ideally, the method should offer precise control over the lipid composition of the vesicle membrane, ensuring uniformity, and yielding unilamellar membranes (or membranes of a defined multi-lamellarity).^{44,45}

(iv) **Generation throughput.** For practical applications, it is advantageous to produce significant quantities of vesicles within a reasonable time frame to meet industrial demands efficiently.

(v) **Presence of impurities.** Some applications, particularly those related to fundamental biological studies, benefit from vesicle membranes free of any impurities and oil.

Considering these KPIs when assessing the different GUV generation methods will aid researchers in selecting the most

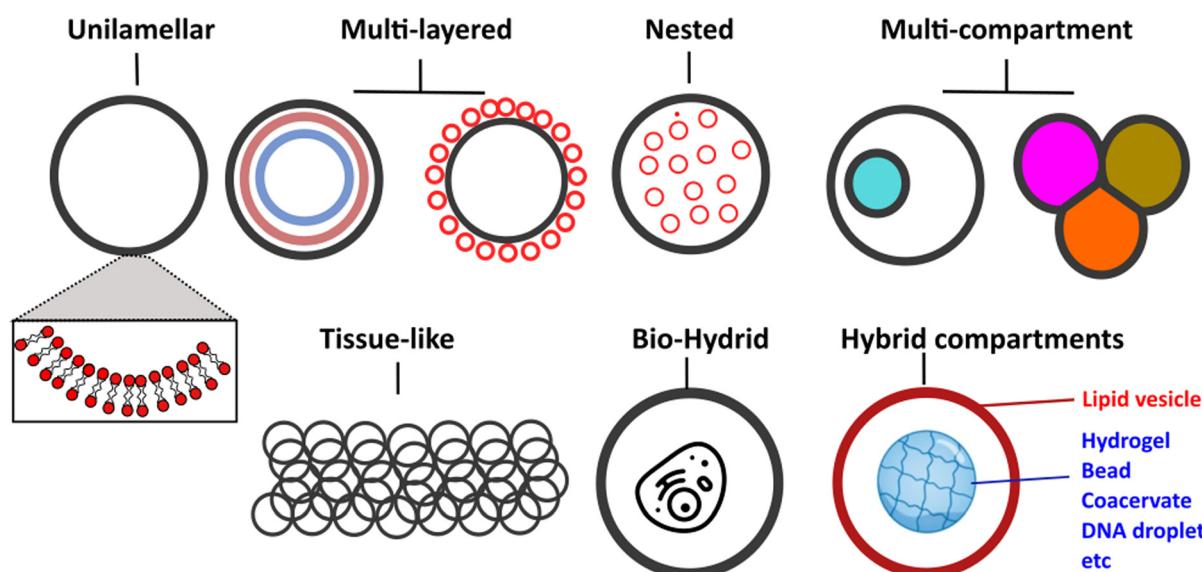


Fig. 2 Schematic showing examples of different vesicle architectures that can be controllably manufactured using microfluidic techniques.



appropriate technique for their specific applications, ensuring optimal performance and reliable outcomes.

Classical methods of generating GUVs include gentle hydration, gel-assisted swelling, and electroformation.³⁵ These techniques have played a pivotal role in revolutionizing our understanding of membrane biophysics phenomena. They provided crucial insights into the mechanical properties of membranes, the influence of lipid composition, the coupling between mechanics and membrane protein activity, as well as the principles governing phase behavior and coexistence of domains (lipid rafts) in cell membranes.⁴⁶ However, for molecular robotics applications, these methods have proven largely unsuitable due to their limitations. They suffer from poor encapsulation efficiency, uncontrolled production, and yield polydisperse populations concerning size, architecture, cargo, and lipid composition.

In the past two decades, innovations have led to the emergence of a new class of vesicle fabrication strategies based on water/oil emulsion technology^{41,47} (Fig. 3(a)). These methods rely on the formation of water/oil droplets stabilized by lipid monolayers. When a second lipid-stabilized water/oil interface is deposited on top of these droplets (*i.e.*, when they are transferred from a bulk oil phase to a water phase), it templates the formation of a second monolayer, resulting in the formation of a lipid membrane. These techniques are sometimes referred to as emulsion transfer or phase transfer methods.

The simplest version of this method involves the manual formation of a water/oil emulsion through pipetting and/or

agitation, followed by phase transfer under gravity, often with the aid of a centrifuge. This process can be repeated multiple times to yield multi-layered arrangements.⁴⁵ This method is highly adaptable for translation into a microfluidic format, where devices can be created using soft-lithography,^{48,49} glass capillaries,⁵⁰ or 3D printing.⁵¹ On-chip droplet production and transfer across a water/oil interface can be achieved using microfabricated pillars⁵² or flow-based approaches.⁵³ Another approach involves using bespoke instrumentation to enable the passage of monodisperse droplets through a continuously rotating horizontal column at the water/oil interface, a method known as cDICE (continuous droplet interface crossing encapsulation), depicted in⁵⁴ Fig. 3(b).

An alternative technique employs microfluidics to form a lipid-stabilized water/oil/water double emulsion. Upon evaporation or de-wetting of the intermediate oil phase, this process leads to the formation of GUVs.^{49,55} The most commonly used version of this method employs octanol to promote de-wetting and is therefore referred to as octanol-assisted liposome assembly (OLA) (Fig. 3(c)).

Advantages of microfluidics

The use of microfluidics enables high-throughput manufacturing of monodisperse (<5% C.V.) GUVs with defined sizes (*ca.* 5–200 μm diameter) and remarkably high encapsulation efficiency (approaching 100%).⁴¹ In the context

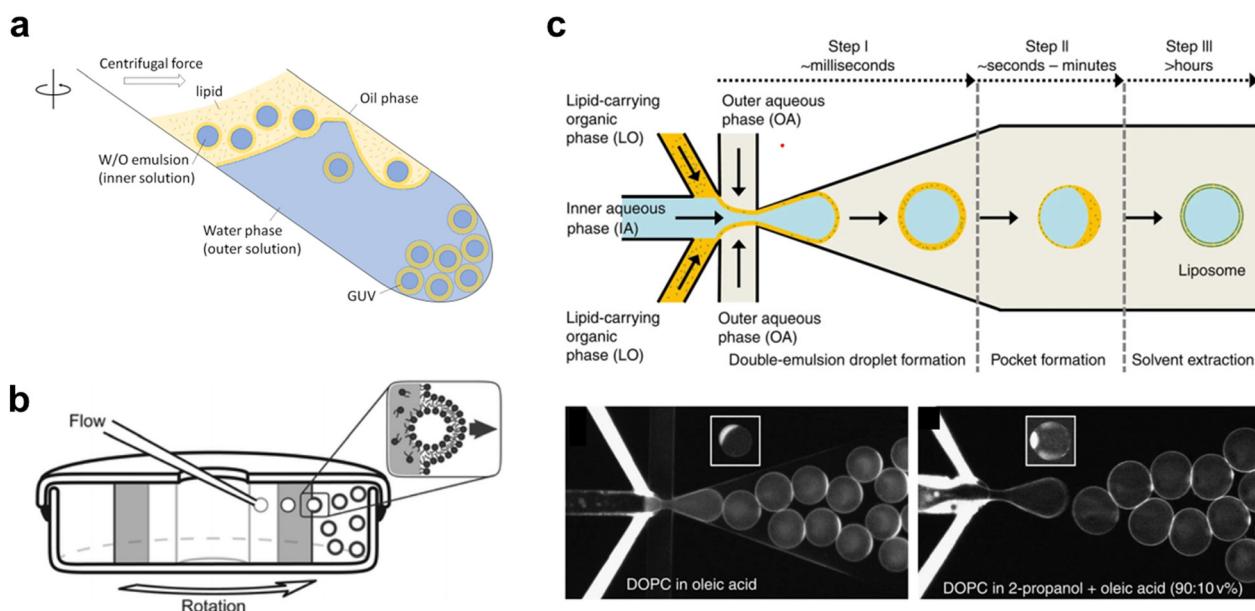


Fig. 3 Examples of different techniques used to generate Giant Unilamellar Vesicles (GUVs). **a.** Emulsion phase transfer, where lipid stabilised w/o droplets are taken through a w/o column through a centrifugal force, resulting in the formation of a second monolayer, and hence the establishment of a lipid bilayer vesicle.³⁵⁴ **b.** Continuous droplet interface crossing encapsulation (cDICE) involving instrumentation which houses concentric layers of fluids with different densities, where aqueous droplets that emerge from a capillary are driven by centrifugal force through multiple lipid interfaces.³⁵⁵ **c.** Octanol-assisted liposome assembly (OLA) method of generating lipid vesicles on-chip. This method involves generating a lipid stabilised w/o/w double emulsion followed by extraction of the intermediate oil phase, leaving behind a lipid membrane.³⁵⁶ Figures reproduced with permission from: a, ref. 354, copyright 2023, MDPI; b, ref. 355, copyright 2016, the Royal Society of Chemistry; c, ref. 356, copyright 2016, Nature Publishing Group.



of creating, manipulating, and analysing molecular robotic systems based on vesicle structures, other lab-on-chip microfluidic modules come into play. For instance, microstructure traps have been employed to capture arrays of hundreds of vesicles, facilitating high-throughput experimentation and extended analysis.⁵⁶ These traps have been coupled with valves to control the perfusion of materials for permeation studies. Purification modules have also been included to remove unencapsulated material and debris from the surroundings.⁴⁸

Different chip geometries can be utilized to generate multi-compartment architectures, such as vesicles-in-vesicles and arrangements with compartments side-by-side.^{50,57} By functionalizing compartments with enzymes, DNA programs, nanoparticles, and CFPS, various features like signalling cascades, trans-compartment communication, division, and spatial segregation of reactions have been achieved within these microrobots.^{19,20} Moreover, linking up thousands of vesicles through adhesive membrane patches has led to the creation of tissue-like structures with self-healing properties.³³ Related technologies include opto-fluidic methods that employ optical trapping and laser technologies for the on-demand fusion of compartments. These techniques trigger biochemical reactions in femtoliter (pL/fL) reaction vessels, study architectural transformations induced by lipid mixing,⁵⁸ and manipulate raft-like domains on the vesicle surface.⁵⁹

Sensors of molecular robots

In living cells, membrane receptors, transporters, and ion channels work as sensors to help cells respond to chemical and physical stimuli. Such functionalities have also been exploited to implement sensing capabilities in molecular robots. In particular, nanopores forming stable nanoscale openings across lipid membranes have been shown to mediate transport of large molecules, enabling their detection by the molecular robots.⁷ The opening and closing of nanopores (gating) can be regulated by environmental stimuli (e.g., pH, light, temperature, osmotic pressure), further enhancing the sensing capabilities of the molecular robots. Various materials have been utilized to assemble nanopores including proteins,⁶⁰ peptides,⁶¹ DNA,¹¹ and synthetic materials.⁶² In the current section, we will mainly focus on the characteristics, differences, and recent progress of nanopore assembly using different building materials, and we will also describe some unique approaches that introduce membrane receptors to GUVs. (For more details on the introduction of membrane receptors to GUVs, we direct readers to a review by Tosaka *et al.*⁶³).

Protein nanopores

Protein nanopores are the most widely-used type of nanopore, both in planar lipid membrane systems and lipid vesicle systems. When reconstituted in planar membrane systems, nanopores are powerful tools for single molecule

detection. The target molecules are detected by recording the change in ionic current output as the molecules pass through nanopores under a constant voltage input. Since the detection sensitivity of nanopore sensing largely depends on the size and structure of the pores, a wide range of proteins have been investigated for the detection of specific targets.^{60,64} In 1996, α -hemolysin (α HL), with a pore size of 1.4 nm at the restriction, which is compatible with the translocation of single-stranded DNA (ssDNA), was initially employed by Kasianowicz *et al.* for detecting polynucleotides.⁶⁵ In 2010, *Mycobacterium smegmatis* porin A (MspA) was found to be suitable for discriminating individual nucleotide bases.⁶⁶ The above pioneering works finally led to the release of the first commercial nanopore DNA sequencer for general use in 2015 by Oxford Nanopore Technologies, using the *E. coli* curli transport channel, CsgG.⁶⁷ Following the achievement of nanopore DNA sequencing (Fig. 4(a)), single-molecule analysis of folded proteins^{68–72} and amino acid sequences in proteins^{73–75} are now underway. Advances such as the detection of single amino acid mutations in peptides using aerolysin from *Aeromonas hydrophila*,⁷⁶ sequential readout of peptides using MspA,^{77,78} and identification of digested protein fragments using fragaceatoxin C from *Actinia fragacea* (FraC)⁷⁹ have been reported in recent years.

Toward the realization of molecular robots, enormous efforts have been made to construct a protein nanopore-lipid vesicle system. Similar to planar lipid membrane systems, the transport of molecules in the lipid vesicle system is highly dependent on the size of the protein nanopores. In addition to conventional studies using protein nanopores of different sizes to transport fluorophores, there are reports on the use of protein nanopores to transport molecules into lipid vesicles, including: the transmission of pH information by a voltage-dependent anion channel (VDAC, 2.5–3 nm) from yeast mitochondria,⁸⁰ the light-/pH-dependent release of fluorescent molecules by a mechanosensitive channel of large conductance (MscL, 3 nm) from *E. coli*,^{81,82} the transport of ATP⁸³ and UTP⁸⁴ by α HL (1.4 nm) from *Staphylococcus aureus*, and the translocation of T7 RNA polymerase (98 kDa) by streptolysin O (SLO, 25–30 nm) from group A, C, and G streptococci⁸⁵ (Fig. 4(b)).

Although numerous protein nanopores with pore diameters ranging from 1.2 to 42 nm have been reported,⁶⁰ the investigation of novel protein nanopores remains intriguing due to the need to expand the toolbox for new target molecules. In 2023, the Maglia group reported the detection of proteins of different sizes using an α -helical pore-forming toxin derived from *Yersinia enterocolitica*, YaxAB.⁸⁶ It consists of a dimer of YaxA and YaxB, forming assemblies of varying sizes, ranging from 8 to 12 dimeric subunits. The characteristic feature of this pore is its huge conical shape with 15/3.5 nm *cis/trans* openings (Fig. 4(c)). Notably, it is the largest protein nanopore ever used for the analysis of folded proteins in planar membrane systems, with the ability to capture a wide range of proteins (from 35–125



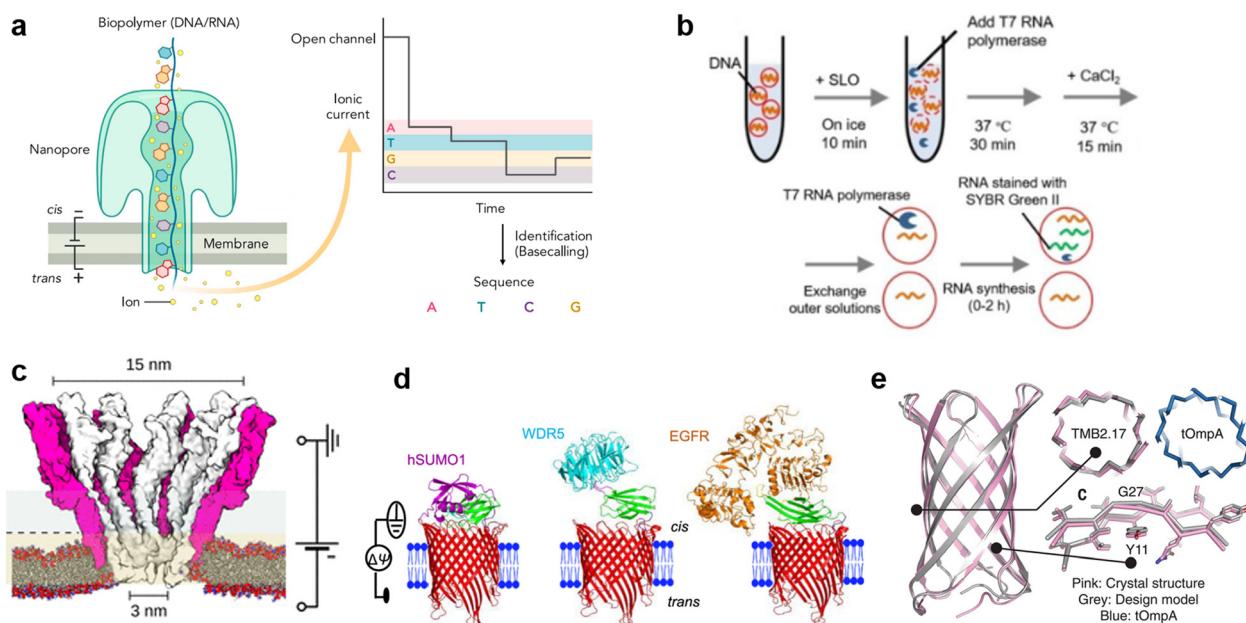


Fig. 4 Protein nanopores for transmitting information. **a**, Illustration of the mechanism of nanopore DNA sequencing.⁷⁵ **b**, Transport of T7 RNA polymerase by the SLO nanopore in a lipid vesicle system.⁸⁵ **c**, Molecular surface illustration of the YaxAB nanopore.⁸⁶ **d**, Protein detection with a monobody containing tFhuA nanopores.⁹⁴ **e**, A comparison of the crystal structure (pink) and the designed structure (gray) of a *de novo*-designed transmembrane eight-strand β -barrel.⁹⁹ Figures reproduced with permission from: a, ref. 75, copyright 2021, Elsevier; b, ref. 85, copyright 2021, Wiley-VCH; c, ref. 86, copyright 2023, American Chemical Society; d, ref. 94, copyright 2023, Nature Publishing Group; e, ref. 99, copyright 2021, AAAS.

kDa). YaxAB exhibits a greater electroosmotic flow under voltage-applied conditions compared to existing nanopores, which aids the entrapment of proteins within the nanopore. Using YaxAB, the group successfully discriminated C-reactive protein (CRP, 125 kDa), hemoglobin (HG, 64 kDa), streptavidin (SA, 53 kDa), and bovine thrombin (BT, 35 kDa) from their mixture. Moreover, CRP was detected at clinically relevant concentrations ($>2.5 \text{ mg L}^{-1}$) in 160-fold diluted depleted human serum.

It is also challenging to detect molecules that are larger than the pore, or to selectively discriminate molecules of a similar size and hydrodynamic radius. Binding of target-specific moieties such as partial antibody domains,⁸⁷ aptamers (single-stranded nucleic acids with the ability to bind to specific molecules),^{88,89} biotin (for the avidin-biotin complex),⁹⁰ inhibitory peptides,⁹¹ protein receptors,⁹² and gold nanoclusters,⁹³ have been employed to achieve the detection of large molecules,^{87,88,90,91} or selective discrimination.^{89,92,93} Since the configuration of the detection system changes depending on the recognition element used, it has been necessary to engineer a distinct system for each target protein. Recently, Movileanu and colleagues reported a general system for the sensitive detection of target proteins.⁹⁴ In their study, the tFhuA nanopore, a β -barrel scaffold of ferric hydroxamate uptake component A (FhuA) from *E. coli*, is covalently attached to a monobody, a recombinant protein based on the fibronectin type III (FN3) domain (Fig. 4(d)). The advantages of monobodies are that they can be selected from a broad range

of variants against numerous target proteins, and their relatively small hydrodynamic radius that does not interfere with pore opening, unlike the larger antibody-derived proteins. Using the monobody-conjugated tFhuA, the authors detected human small ubiquitin-related modifier 1 (hSUMO1), WD40 repeat protein5 (WDR5). Moreover, the group achieved the identification of the 20 nM-epidermal growth factor receptor (EGFR), which is a 180 kDa prognostic protein biomarker associated with lung, colon, and breast cancer, in 5% (v/v) fetal bovine serum (FBS), suggesting the potential applications of molecular robots in nanomedicine.

In recent years, in addition to using natural membrane proteins, attempts have been made to fabricate nanopores through *de novo* design. This approach uses computer-aided protein design to create proteins with tailored shapes and functionalities.⁹⁵⁻⁹⁸ *De novo* design allows the fabrication of nanopores with arbitrary structures that do not exist in nature, thereby expanding the range of detectable molecules. Conventionally, *de novo* design of transmembrane proteins is considered difficult due to the unclear relationship between membrane insertion, folding mechanisms and the corresponding amino acid sequences. In 2021, Vorobieva *et al.* achieved the first *de novo* design of an eight-strand transmembrane β -barrel⁹⁹ through a complex optimization process including: (i) the refinement of the loop structure and adjustment of hydrophobicity, (ii) backbone design and sequence optimization using Rosetta,¹⁰⁰ (iii) additional design guidelines appropriate for the membrane environment, and (iv) exhaustive expression trials. The



expressed structure was confirmed to be highly consistent with the designed structure (1.1 Å backbone RMSD over all residues) (Fig. 4(e)). Single-molecule detection by this protein nanopore has not yet been reported, probably due to the smaller pore size compared to natural nanopores. However, with the recent popularization of *de novo* protein design tools,^{100–102} the development of protein nanopores suitable for molecular robotics applications with better molecular selectivity, transport capacity, stability, and durability are expected in the future.

Peptide nanopores

As with protein nanopores, peptides that assemble into pores in the membrane are also attractive as sensors in molecular robotics. Compared with protein nanopores, peptide nanopores are more tractable, since stable peptide nanopores can be designed based on naturally occurring sequences or by *de novo* design, and peptides with the 30–40 residues required for membrane-spanning can be synthesized in large quantities by chemical synthesis. Originally, antibacterial peptides (AMPs) such as gramicidin and alamethicin were known to assemble to form pores in bacterial membranes, exhibiting antibacterial activity^{61,103–106} (Fig. 5(a)). The AMP nanopores have been reported to enable permeation of small molecules such as fluorophores,^{107–111} but it has proven

difficult to transport molecules with larger molecular weights.^{112–114} In recent years, attempts have been made to design more stable and monodispersely sized peptide nanopores compared to AMP nanopores.

One approach to obtaining stable peptide nanopores is the redesign of the transmembrane regions of natural membrane proteins.^{115,116} The advantage of this approach is that no explicit complex design principles are required, and a certain degree of stability is expected. Redesigned peptide nanopores have demonstrated that such transmembrane peptides stably form pores with a certain number of monomers, opening the field of peptide nanopores and providing promising nanopore-forming peptides to date. Recently, the Mahendran group reported that DpPorA, which is a D-amino acid-substituted pPorA peptide derived from the porin PorACj of *Corynebacterium jeikeium*, could form a stable monodisperse octameric pore^{116–118} (Fig. 5(b)). Compared to conventional helix barrels, DpPorA is characterized by very large conductance and is highly cation-selective, so that cyclodextrins and nonaarginine (R9) can translocate under applied voltage in a planar membrane system, while there is negligible interaction with anionic cyclodextrins and oligopeptides. Notably, whilst their L counterparts are readily degraded by proteinase K, DpPorA is highly resistant to protease K treatment, indicating promising potential for use as a stable sensor in molecular robots in biological environments.

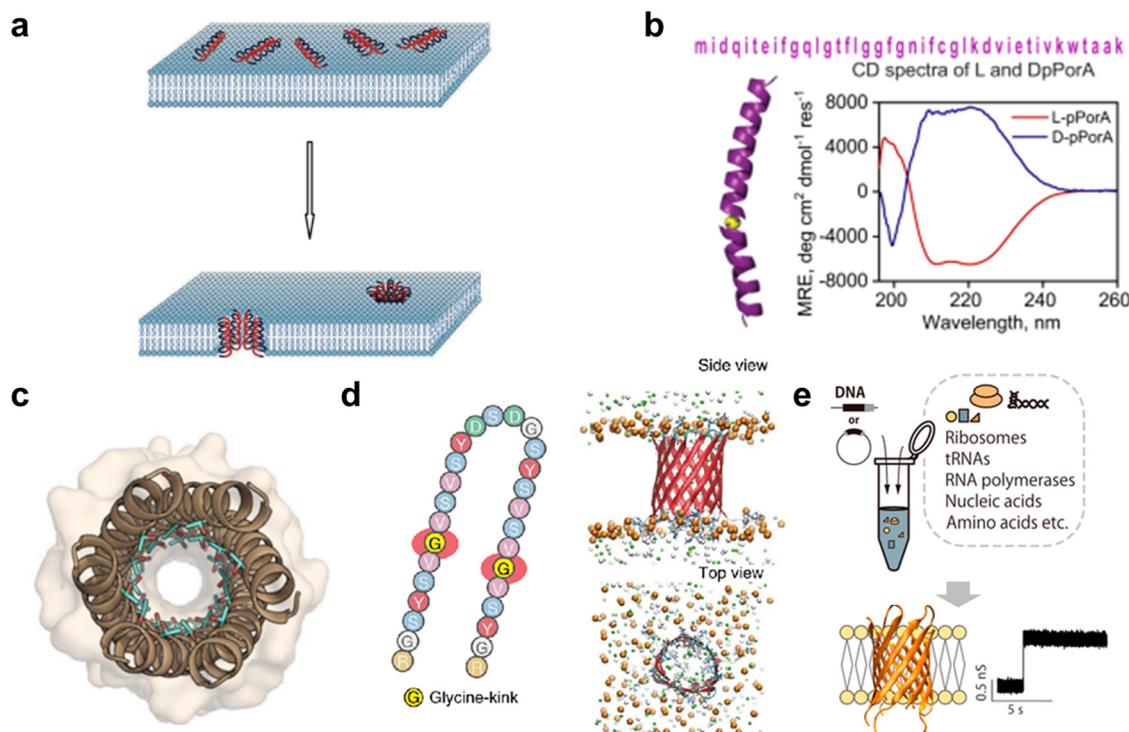


Fig. 5 Peptide nanopores for transmitting information. **a**, Formation of nanopores in lipid membranes by alamethicin.³⁵⁷ **b**, DpPorA peptide sequence and CD spectra for DpPorA (blue) and LpPorA (red).¹¹⁸ **c**, X-ray crystal structure of the nanopore formed by the *de novo*-designed α -helix peptide.³⁵⁸ **d**, SVG28 sequence and simulated pore structure.¹²² **e**, Utilizing CFPS to synthesize variants of SVG28.¹²³ Figures reproduced with permission from: a, ref. 357, copyright 2014, Hindawi Publishing Corporation; b, ref. 118, copyright 2022, Nature Publishing Group; c, ref. 358, copyright 2022, American Chemical Society; d, ref. 122, copyright 2023, Nature Publishing Group; e, ref. 123, copyright 2023, American Chemical Society.



Similar to protein nanopores, another approach to obtain peptide nanopores is *via de novo* peptide design. Several *de novo*-designed α -helix peptides have been reported in recent years,^{119,120} as α -helices allow for the evaluation of inter-helix stability and the precise control of packing interfaces. In 2021, Woolfson and colleagues reported the *de novo* design of an α -helix peptide that forms stable, monodispersely sized pores.¹²⁰ They began by designing a water-soluble α -helical barrel by introducing polar residues on the lumen of the α -helical bundles of pentameric or higher order (Fig. 5(c)), to support the helical interface and the passage of water molecules, resulting in a parallel hexamer water-soluble pore. By incorporating hydrophobic leucine and membrane interface-preferring tryptophan residues into the outer surface of the water-soluble pore, peptides that form monodisperse pores in the membrane were achieved. To further improve the inter-helix packing, computational scoring was used to fabricate possibly hexameric nanopores with a longer lifetime and fewer open–close states (gating). Although single-molecule detection has not been attempted, probably due to the small pore size, the group has demonstrated the fabrication of water-soluble helical barrels from 5 to 9-mers by changing the residue size at specific positions on the helical wheel,¹²¹ leading to the possibility of pore size expansion and greater molecular transport ability.

In 2022, Shimizu *et al.* reported the *de novo* design of β -hairpin peptides SV28 and SVG28, which form transmembrane β -barrel nanopores following four simple design principles: (i) alternating hydrophilic and hydrophobic transmembrane residues, (ii) introduction of membrane interface-anchoring residues, (iii) introduction of charged residues for membrane insertion by electrophoresis, and (iv) introduction of a glycine kink to reduce the strain of the β -barrel (only for SVG28)¹²² (Fig. 5(d)). SVG28 forms almost monodisperse heptameric pores under optimized conditions and can discriminate between L-PLL (molecular weight 30 000–70 000) and S-PLL (molecular weight 10 000). In addition, SVG28 detects cationic peptides more efficiently than the commonly used α HL nanopore due to its terminal charge. Moreover, Fujita *et al.* have found that a hydrophilic variant of SVG28 can be synthesized using CFPS, which retained the pore-forming ability¹²³ (Fig. 5(e)), promising easy access to peptide nanopores for molecular robotics. Furthermore, we showed that SV28, an antecedent of SVG28, can form a wide range of pore sizes from 1.7 to 6.3 nm and can translocate the G4 structure of DNA when the pore size is sufficiently large. Therefore, we believe that it is possible to find SVG28 variant sequences that form nanopores with a suitable size for molecular transport. Future efforts are required not only to design assembled pore structures but also to add vestibule regions¹²⁴ and lining structures¹²⁵ that stabilize the pore structure.

DNA nanopores and receptors

DNA nanotechnology presents a unique tool for mimicking the function of biological nanopores as sensors for molecular

robots, due to its advantages in design flexibility and programmability. Nanopores formed by DNA nanotechnology, so-called DNA nanopores, can be realized by DNA origami strategies which use a long single-stranded scaffold DNA and hundreds of staple DNA strands,^{126,127} as well as DNA nanostructures strategies which use few short DNA strands.^{128,129} To embed the DNA nanopores into lipid membranes, the DNA nanopores carry hydrophobic molecules such as cholesterol and tocopherol so that they can interact with the hydrophobic membrane core. From the first two groundbreaking DNA nanopores that transported ions through planar lipid membranes,^{130,131} in 2016, Krishnan *et al.* then reported the successful transport of small dye molecules through lipid membranes of GUVs using square-shaped DNA nanopores with a 4.2 nm-wide channel lumen¹³² (Fig. 6(a)). Since then, several DNA nanopores that function on GUV membranes have been reported.^{133–136}

The current strategies to improve DNA nanopores constructed by DNA origami mainly focus on two areas: expanding the pore diameter,^{133–137} and adding unique functions for selective transportation.^{133,134,137} A larger pore not only allows larger molecules to be transported, but also improves the efficiency of transport. In 2021, a 35 nm-wide DNA nanopore was reported with the capability of transporting dextran with a molecular weight of up to 250 kDa¹³⁵ (Fig. 6(b)). Furthermore, in 2022, Xing *et al.* developed DNA nanopores with different polygonal shapes and sizes that permit the transport of antibodies.¹³⁸ Meanwhile, adding functions such as control over gating can provide the ability to selectively transport molecules. To date, various methods for controlling the gating have been demonstrated, including: modifying the pore lumen with PEG molecules,¹³³ changing the number of lid-like DNA strands on top of the pore¹³⁴ (Fig. 6(c)), and reversible opening and closing with key DNA strands¹³⁷ (Fig. 6(d)).

While DNA origami is advantageous in constructing pores with large diameters, their versatility and stability is hindered by the complicated design procedure. In contrast, DNA nanopores based on DNA nanostructures have generated great interest due to their simpler design strategy. To date, DNA-nanostructure-based nanopores with six-helix^{11,139–141} (Fig. 6(e)) or eight-helix¹⁴² bundle structures, and wireframes of triangular¹⁴³ and square¹⁴⁴ shapes have been reported to function on GUV membranes. The relatively small lumen size provides the merit of charge-selective molecular transport, since negatively-charged DNA molecules have strong electrostatic interactions with charged cargo molecules^{11,139} (Fig. 6(f)). Moreover, similar approaches have been described to control the gating of DNA nanopores with inputs including specific molecules,¹⁴⁵ temperature,¹⁴⁶ light,¹⁴⁷ and mechanical stimuli.¹⁴⁸

Receptor-mimicking DNA nanostructures, which do not puncture a pore but simply span the lipid membrane, are also potential sensors for molecular robots. Unlike DNA nanopores, receptor-mimicking DNA nanostructures do not suffer from leakage issues, and hence can achieve signal



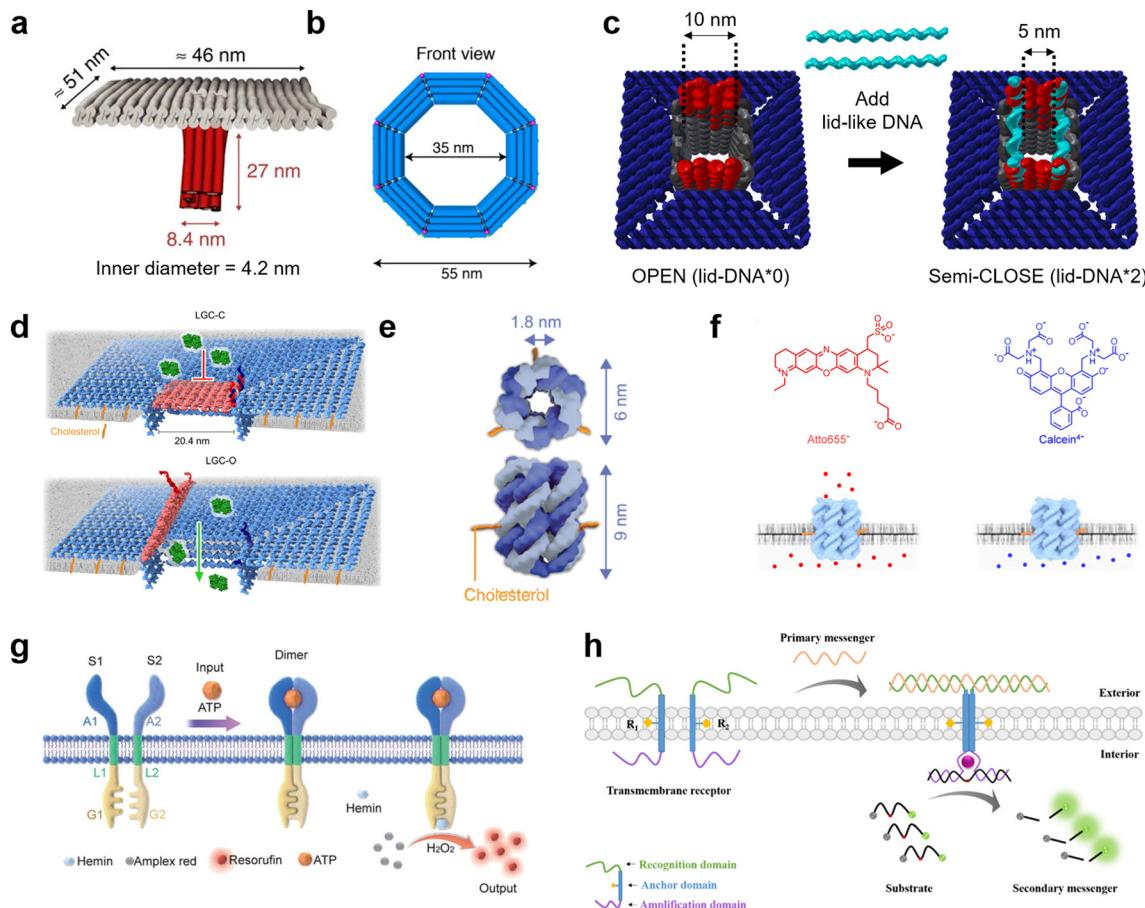


Fig. 6 DNA nanopores and receptors for transmitting information. **a**, First reported DNA origami nanopore that transported dye molecules across the lipid membranes of GUVs.¹³² **b**, DNA origami nanopore with the largest inner diameter.¹³⁵ **c**, DNA origami nanopore for size-selective transport, controlled by changing the number of lid-like DNA strands on top of the pore.¹³⁴ **d**, DNA origami nanopore that reversibly opens and closes upon addition of key DNA strands.¹³⁷ **e**, DNA nanopore composed of six DNA strands.³⁵⁹ **f**, Charge selective transport of dye molecules by a six-helix DNA nanopore.¹³⁹ **g**, Receptor-mimicking DNA nanostructures capable of detecting ATP and lysozyme.¹⁴⁹ **h**, Receptor-mimicking DNA nanostructures capable of detecting DNA strands.¹⁵⁰ Figures reproduced with permission from: a, ref. 132, copyright 2016, Nature Publishing Group; b, ref. 135, copyright 2021, American Chemical Society; c, ref. 134, copyright 2021, Royal Chemical Society; d, ref. 137, copyright 2023, Nature Publishing Group; e, ref. 359, copyright 2023, Wiley-VCH; f, ref. 139, copyright 2021, American Chemical Society; g, ref. 149, copyright 2021, Royal Chemical Society; h, ref. 150, copyright 2021, American Chemical Society.

transduction with high accuracy. Typical receptor-mimicking DNA nanostructures include two transmembrane DNA with a recognition domain, an anchor domain, and an amplification domain. When the signal inputs arrive at the GUV outer membrane, the two transmembrane DNA hybridize with each other and hence enter close spatial proximity. The amplification domain consequently forms a G-quadruplex structure or complement strands, resulting in the generation of further chemical reactions inside the GUVs. To date, receptor-mimicking DNA nanostructures capable of detecting ATP,¹⁴⁹ lysozyme¹⁴⁹ (Fig. 6(g)), and DNA messengers¹⁵⁰ (Fig. 6(h)) have been reported.

All the above reports have been achieved during the past decade. Thus, the construction of DNA nanopores and receptor-mimicking DNA nanostructures with more complicated functions is expected to accelerate in the next decade. For DNA nanopores, current studies perform molecular transport using simple diffusion, whereas active

transport of molecules is highly desired. For receptor-mimicking DNA nanostructures, systems that can detect multiple molecules and generate multiple reactions simultaneously are yet to be realized.

Synthetic channels

Synthetic supramolecular complexes have emerged as a type of pore capable of functioning in lipid membranes.¹⁵¹ These pores are always smaller than 1 nm in diameter and thus only transport ions, similar to ion channels in living cells. Previous studies have referred to these pores as “nanopores” or “nanochannels”, but without clear criteria. In a distinction from the nanopores that can transport both ions and larger molecules, in this review, we define pores based on synthetic supramolecular complexes as “synthetic channels”. Thanks to the wide chemical repertoire, synthetic channels with different functions have been generated to date. A



comprehensive introduction to this field is beyond the scope of this review, so we will focus on the recent advances in this field and discuss how they can impact the field of molecular robots (for a full survey of synthetic channels, we direct the readers to previous reviews^{31,152–154}).

Selective ion transport is the most well-established function for synthetic channels and is a highly desired function for molecular robots because it affords the opportunity for strict regulation of information input. Similar to natural ion channels,¹⁵⁵ the selectivity of synthetic channels is highly dependent on both the pore lumen size and the type of functional groups in the pore lumen. Starting from several ground-breaking works in 1990s,^{156–158} synthetic channels that can selectively transport K⁺,¹⁵⁹ Cl⁻,¹⁶⁰ I⁻,¹⁶¹ and H₂O¹⁶² have been reported. Currently, efforts are focusing on constructing channels that can selectively transport smaller cations.^{163–165} In 2023, Zhang *et al.* reported a Li⁺-selective synthetic channel using self-assembled aromatic molecules

with well-adjusted lumen size and coordination numbers¹⁶³ (Fig. 7(a)). The group found that the channel, with a 1.8 Å-diameter lumen and four ion coordination sites inside the cavity, exhibits higher transport selectivity for Li⁺ over Na⁺, achieving Li⁺/Na⁺ selectivity ratios ranging in value from 15.1 to 23.0. Meanwhile, activities in developing new types of anion-selective channels continue unabated.^{161,166,167} In 2020, Roy *et al.* reported a fully hydrogen-bonded, helically-folded aromatic foldamer-based synthetic channel, with a central lumen rendered by the many methyl groups¹⁶¹ (Fig. 7(b)). The methyl groups provide a positive electrostatic potential, resulting in a selectivity towards anions over cations. Moreover, I⁻ was observed to be more favourably transported than Cl⁻ due to having fewer hydrogen bonds than Cl⁻ (I⁻/Cl⁻ selectivity ratio was around 11).

Responsivity to external stimuli is another essential function for synthetic channels and would provide the ability for remote-control in molecular robots. Light is one of the

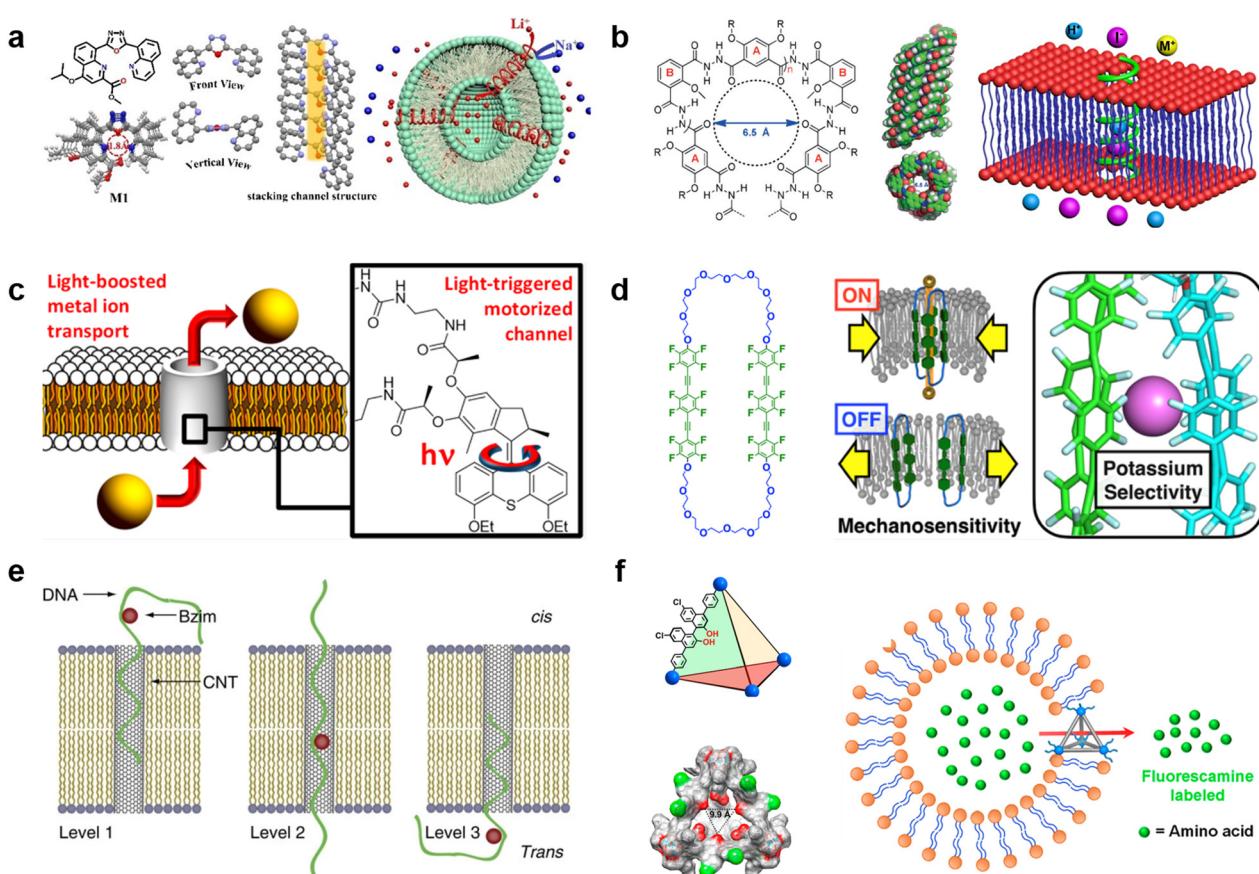


Fig. 7 Synthetic channels for transmitting information. **a**, Chemical structures of aromatic molecules (left). Single crystal structure of aromatic molecules and its linearly self-assembling channel structure (central). Illustration of selective transmembrane transport of Li⁺ (right).¹⁶³ **b**, Chemical structure of foldamer (left). Side and top view of the foldamer-based synthetic channel. Illustration of selective transmembrane transport of I⁻ (right).¹⁶¹ **c**, Schematic showing light-driven transport of metal ions and an enlarged view of the molecular motor.¹⁷⁰ **d**, Chemical structure of amphiphilic cyclophanes with perfluorinated aromatic units (left) and its response to mechanical stress with selective transport of K⁺.¹⁷² **e**, Illustration of benzoimidazole (Bzim)-modified 5hmC-containing DNA2 translocation through a SWCNT.¹⁸¹ **f**, Illustration and single-crystal X-ray structure of the tetrahedral MOCs (left). Transport process of amino acids through the tetrahedral MOCs (right).¹⁸⁷ Figures reproduced with permission from: a, ref. 163, copyright 2023, Wiley-VCH; b, ref. 161, copyright 2020, Wiley-VCH; c, ref. 170, copyright 2021, American Chemical Society; d, ref. 172, copyright 2022, American Chemical Society; e, ref. 181, copyright 2013, Nature Publishing Group; f, ref. 187, copyright 2021, American Chemical Society.



most widely used stimuli due to having high biocompatibility and ease of spatio-temporal control. So far, light-responsive synthetic channels which are irreversibly¹⁶⁸ and reversibly¹⁶⁹ photo-controlled have been developed, and recent studies focus predominantly on reversible photo-control. In 2021, Wang *et al.* reported a light-driven synthetic channel that can actively transport alkali ions¹⁷⁰ (Fig. 7(c)). To achieve active transport, the group included a light-driven rotary motor around the channel. Under light irradiation, the motor provides thermal energy to the channel to help overcome the activation barrier necessary to translocate ions between macrocycles, leading to an increase in Na^+ transport activity (up to 400%). In living cells, mechanical stress is a major stimulus involved in the regulation of structure and function. In the cytoplasmic membrane, ion channels called mechano-sensitive channels (MSCs) sense mechanical stress and enhance substance influx/efflux as an output. Synthetic channels that mimic the function of MSCs to respond to mechanical stress are now attracting great attention. The first mechano-sensitive synthetic channel was reported by Muraoka *et al.* in 2017.¹⁷¹ Using a channel consisting of repeating oligo-(ethylene glycol) (OEG) and aromatic units, with subunits that destabilize the pore structure, the ion transport activity was observed to decrease with expanding membrane tension. In 2022, the group further extended the concept to develop a synthetic channel with potassium selectivity by using amphiphilic cyclophanes with perfluorinated aromatic units¹⁷² (Fig. 7(d)), opening up the possibility to process multiple signals with synthetic channels. Other synthetic channels responding to stimuli including voltage^{173,174} and the presence of ligands^{175,176} have also been reported in recent years.

Over the past decades, many efforts have been made to construct synthetic channels employing macrocyclic molecules,¹⁷⁷ foldamers,¹⁷⁸ and π -stack architectures.¹⁷¹ Nevertheless, the synthesis of conventional synthetic channels varies case-by-case, making it challenging to prepare channels with different functions using a general protocol, and hence restricting their use in molecular robots. Carbon nanotubes (CNTs), which are all-carbon hollow nanobarrels with a diameter in the range of 0.8–2 nm,⁶² are considered as a versatile synthetic channel that can be easily prepared. Benefiting from a larger size than conventional synthetic channels, previous reports have shown that CNTs can transport not only ions and water molecules,¹⁷⁹ but also larger molecules such as amino acids¹⁸⁰ and ssDNAs.¹⁸¹ Importantly, Liu *et al.*, utilized CNTs to selectively detect benzoimidazole (Bzim)-modified 5-hydroxymethylcytosine (5hmC) in ssDNA¹⁸¹ (Fig. 7(e)), opening the door to the use of CNT-based molecular robots for the diagnosis of gene mutations. Moreover, a recent study by Marcotte *et al.* has shown that CNTs enable mechanically-activated ionic transport that depends quadratically on the applied pressure,¹⁸² making CNTs a potential candidate as mechanical stress sensor.

Metal-organic polyhedra (MOPs), also known as metal-organic cages (MOCs), have emerged as another popular

option in the synthetic channel family.¹⁸³ MOPs/MOCs are nano-sized cages constructed by linking transition metal clusters with organic ligands, hence benefiting from the ability to control functionality by choosing appropriate metal clusters and ligands from a wide variety of options.¹⁸⁴ The first report that studied the interaction between MOPs/MOCs with lipid membranes was by Jung *et al.* in 2008, and their results suggested that MOPs/MOCs synthesized from 5-dodecoxybenzene-1,3-dicarboxylic acid ($5\text{-OC}_{12}\text{H}_{25}\text{-mBDCH}_2$) and $\text{Cu}(\text{CH}_3\text{CO}_2)_2\text{H}_2\text{O}$ have transport selectivity dependent on cation size.¹⁸⁵ In 2017, Kawano *et al.* synthesized rhodium metal-organic polyhedrals (RhMOPs) and demonstrated their multiple conductance states, which are proposed to result from reversible geometry switching of the RhMOPs, between square and triangular geometry of the apertures exposed to the aqueous phase in the lipid membranes.¹⁸⁶ In 2021, Li *et al.* developed tetrahedral MOCs with hydrogen bonding units inside the cage cavity using six chiral BINOL-derived ligands and four $n\text{-Bu}_3\text{-Cp}_3\text{Zr}_3$ clusters, and demonstrated their ability to transport amino acids¹⁸⁷ (Fig. 7(f)). Importantly, the chiral cavity enables enantiospecific recognition of amino acids, which is a rarely-reported function for conventional synthetic channels. The endless potential combinations of metal clusters and ligands make MOPs/MOCs sensors for molecular robots an exciting prospect with limitless possibilities.

Computers of molecular robots

Molecular robots go hand-in-hand with biological computers, where biological computers take available inputs and translate them into appropriate outputs. Thanks to advances in synthetic chemistry and biology, biologically derived molecules such as DNA and proteins have become readily available, leading to the development of biological computers such as DNA computing systems and CFPS.^{188,189} In this section, we will briefly explain the characteristics and reveal the research trends in DNA computing and CFPS.

DNA computing

Due to the remarkable programmability of DNA molecular behavior based on sequence-dependent hybridization, enzymatic reactions, and strand displacement reactions, DNA computing has emerged as a promising candidate for the computational machinery of molecular robots. The genesis of DNA computing can be traced back to Adleman's pioneering work: massively parallel computation using artificially sequence-designed DNA.¹² He encoded the nodes and paths in the Hamiltonian path problem onto different ssDNA to execute hybridization-based parallel exploration of the correct Hamiltonian path. By exploiting DNA self-assembly, this methodology allowed large-scale computations to be performed with low energy consumption. Besides the subsequent implementation of mathematical computations (satisfiability problem,^{190,191} maximal clique problem,¹⁹¹ etc.), Benenson *et al.* constructed DNA-based finite automata



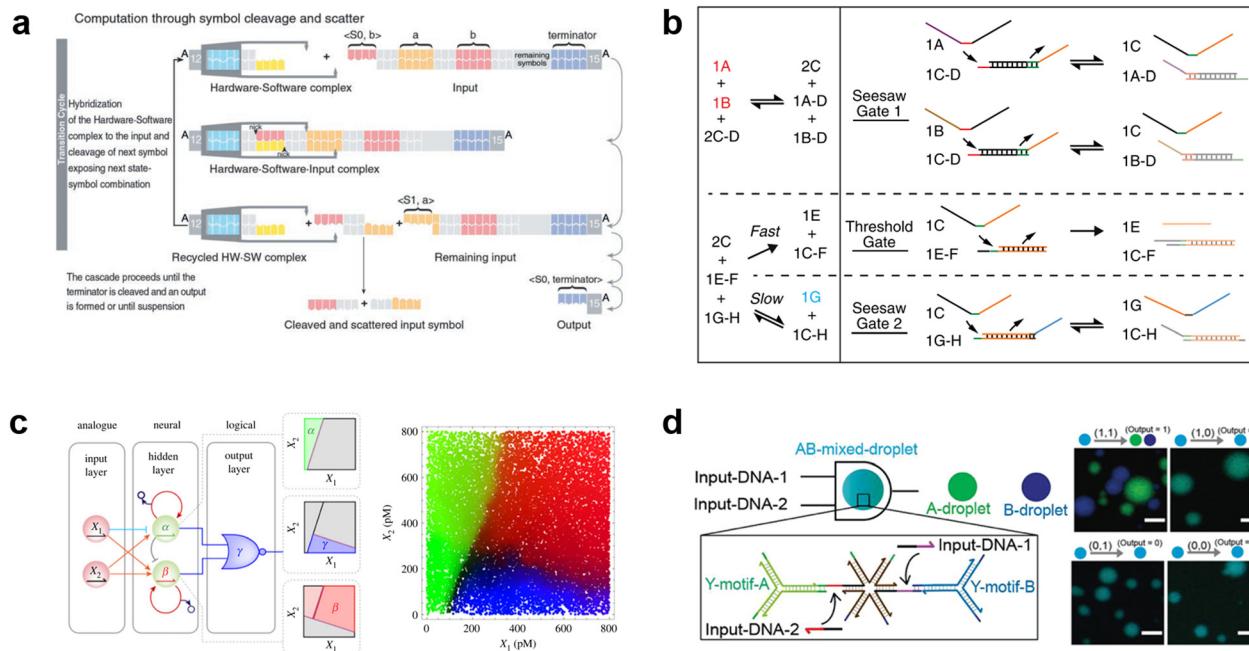


Fig. 8 DNA computing for signal processing. **a**, Suggested mechanism of operation of the automaton by DNA-based state transition using the restriction nuclease FokI and ligase (right).¹⁹³ **b**, ‘Seesawing’, ‘thresholding’, and ‘reporting’ DNA reactions.³⁶⁰ **c**, The decision architecture (left) of the neural network. Fluorescence levels of α , β and γ , measured in approximately 25 000 droplets (right).³⁶¹ **d**, Illustration of the AND gate operation using DNA droplets (left) and CLSM images for droplet phase separation corresponding to four input patterns (right). Scale bars: 10 μm .²¹² Figures reproduced with permission from: a, ref. 193, copyright 2003, National Academy of Sciences; b, ref. 360, copyright 2012, Elsevier; c, ref. 361, copyright 2023, Royal Chemical Society; d, ref. 212, copyright 2022, Wiley-VCH.

with two states using programmed DNA sticky/blunt ends, restriction nuclease, and ligase¹⁹² (Fig. 8(a)). The two states in the automata run at a rate of 109 transitions per second in an input-responsive manner, being the prototype of the smallest biocomputer¹⁹³ as certified by Guinness World Records. Originating from the above single-information processing systems, the development of DNA computing has recently steered towards multiplex information processing.

As a multi-input responsive module, logic gate operations were adopted due to the inherent simplicity in harnessing binary information represented by ‘0’ and ‘1’. Stojanovic *et al.* first demonstrated a DNA-based logic gate using deoxyribozymes (DNAzymes).¹⁹⁴ In their pioneering approach, two ssDNA were defined as inputs and a DNAzyme was employed as a computational module to catalytically cleave DNA in a logically controlled manner, producing a distinct output that was detected *via* fluorescence readout. Based on this principle, they successfully implemented NOT, AND, and XOR gates. This was followed by other groups that reported the construction of OR, NOR, and NAND gates using DNAzyme/ribozyme-based methodology.^{195,196}

Having developed basic logic gates, the next phase of the research effort focussed on cascading the individual gates towards the construction of DNA circuits. DNA strand displacement has emerged as a pivotal technology for implementing cascades, providing a more permissive operating framework than the use of DNAzymes/ribozymes that require strict experimental conditions.¹⁹⁷ In strand

displacement reactions, an input ssDNA reacts with a double-stranded DNA (dsDNA), where it selectively binds to an exposed single-stranded region. This binding event leads to the displacement and release of a pre-existing hybridized ssDNA component. In 2006, Seelig *et al.* proposed a way to cascade DNA logic gates using strand displacement reactions.¹⁹⁸ In their approach, the output ssDNA released by the strand displacement reaction in the first layer gate was exploited as a subsequent input for the next layer gates. By programming short oligonucleotides to bind/release and incorporating fluorophore/quencher modification, AND, OR, and NOT gates were cascaded without any enzymatic reactions. The same group then extensively applied this strand displacement principle to construct a larger-scale circuit consisting of AND and OR gates using 130 DNA strands, with a combination of ‘seesawing’, ‘thresholding’, and ‘reporting’ DNA reactions¹⁹⁹ (Fig. 8(b)). Importantly, the group demonstrated the circuit’s ability to compute the floor of the square root of a four-bit binary number, enabling digital logic networks to be compiled into DNA-based implementations. Moreover, circuits with unique functions, for example, DNA circuits with enhanced reaction speed, responsiveness, and robustness using the cationic polymer, poly(L-lysine)-graft-dextran,²⁰⁰ in addition to temporal DNA circuits that can respond to both the presence and history of a molecular environment,²⁰¹ and pH-responsive switchable DNA circuits²⁰² have all been reported. Through such cascading, scaling, and additional functionalization, DNA

circuits can potentially attain the capabilities of reliable computation, precise error correction, and automated circuit compilers, ultimately achieving a computational behaviour reminiscent of electronic computers in wet biological environments.

Besides the binary information processing as described above, what types and level of complex information can DNA computing handle? One intriguing topic is the construction of neural networks, mathematical models that mimic the interconnected neurons of the human brain, using DNA molecules. Qian *et al.* demonstrated the translation of arbitrary linear threshold circuits (neural network model) into strand displacement cascades as a Hopfield network consisting of four interconnected artificial DNA-based neurons.²⁰³ By setting the proper weights and thresholds for each neuron, the network was able to 'remember' a series of binary patterns, opening up possibilities for embedding 'learning' into DNA computations. Following this remarkable demonstration, several types of neural networks with training processes were constructed such as winner-take-all neural networks that recognized 9 patterns from 100-bit inputs,²⁰⁴ convolutional neural networks that recognized 32 categories from 144-bit inputs,²⁰⁵ and oscillation-based reservoir computing as a type of recurrent neural network.²⁰⁶ Focusing on the practical applications of DNA-based neural networks, decision-making functionality is useful for clinical diagnosis in healthcare. Okumura *et al.* recently proposed nonlinear decision-making using DNA-based enzymatic neurons with adjustable weights and biases. Combining a hidden layer with two linear classifiers and a logical layer with a NOR gate, the network correctly partitioned the concentration space of the two cancerous miRNAs (X1 and X2) as inputs into three nonlinearly separable regions using distinct fluorescence (α (green), β (red) γ (blue))²⁰⁷ (Fig. 8(c)). As represented in this report, oligonucleotide biomarkers, such as miRNA, can be used as inputs in DNA computation. Diagnostic applications are therefore an emerging trend in the development of medical DNA computation,^{208,209} holding the promise of substantial impacts on the advancement of medical molecular robotics. Furthermore, the above DNA-based neural network architectures offer the fascinating prospect of endowing molecular robots with 'intelligence', including the realisation of intricate human-brain-like functionalities from perception to memory and consciousness, using DNA.

In addition to the liquid-phase DNA computations described so far, other computational methods using DNA droplets/condensates have also attracted attention. For instance, the Takinoue group developed DNA droplets based on liquid–liquid phase separation of Y-motif DNA consisting of three ssDNA, whose sticky ends enabled selective and exclusive fusion of the droplets.^{210,211} Based on this DNA droplet technology, the group further developed computational DNA droplets with AND gate functionality, where the mixed droplet separated into two distinct droplets only in the presence of two input DNA (= input (1, 1))²¹² (Fig. 8(d)). Moreover, using cholesterol modification as an

alternative to DNA sticky ends, the Di Michele group established DNA condensates with a responsive core-shell structure²¹³ and with internal domains induced by reaction-diffusion waves.²¹⁴ These technologies can be integrated with GUVs-based molecular robots as components with input-responsive computational capabilities.

Cell-free protein synthesis (CFPS)

The current developments in the field of molecular robots have focused on using DNA as the output for the computational machinery. On the other hand, the diverse building blocks of proteins make them an attractive alternative output, capable of increased information relay compared with DNA. To this end, another promising tool by which molecular robots respond to external stimuli and convert these into signals or outputs is CFPS. CFPS is capable of synthesizing proteins in a test tube *via* transcription and translation from DNA, without the use of living cells.^{215,216} CFPS can be divided into two types: a reconstituted system and a cell-extract-based system. A reconstituted system is a CFPS where components essential for protein synthesis are highly purified individually and reconstituted in a test tube to carry out the reactions, while the cell-extract-based system is essentially a crude cell extract that contains not only the protein translation machinery but also additional molecules involved in other reactions. A reconstituted system has the advantage of less contamination by factors unrelated to protein synthesis and the possibility of tailoring the composition of the reaction on demand. For these reasons, not all but many of the molecular robots consisting of CFPS reported to date use a reconstituted system, the PURE system,¹⁴ with its high adjustability.

Input signals for controlling the fate of molecular robots consisting of CFPS can be divided into two major categories: 1) chemical and 2) physical signals. In response to the input of various chemical signals, intravesicular CFPS has been shown to synthesize proteins, most often reporter proteins. Kamat's group has developed a sensor that expresses luciferase in response to nitrate.²¹⁷ In the bacterial two-component sensing system, NarX–NarL, the binding of nitrate to the sensor domain of NarX induces phosphorylation of NarL by the kinase domain of NarX. The phosphorylated NarL acts as a transcriptional activator and downstream reporter proteins are expressed (Fig. 9(a)). The authors reconstituted the NarX–NarL system inside GUVs and showed not only the sensing of nitrate but also that of nickel, iron, and vancomycin is possible, by only modifying the sensor module. In addition, the same group has recently shown the use of a fluoride-responsive riboswitch that allows detection of the presence of fluoride in the environment by coupling it to the expression of reporter proteins.²¹⁸ While the examples above use previously reported or natural response elements, Dwidar *et al.* started by creating a novel riboswitch. They created a histamine-responsive riboswitch using SELEX. The riboswitch was encoded upstream of a



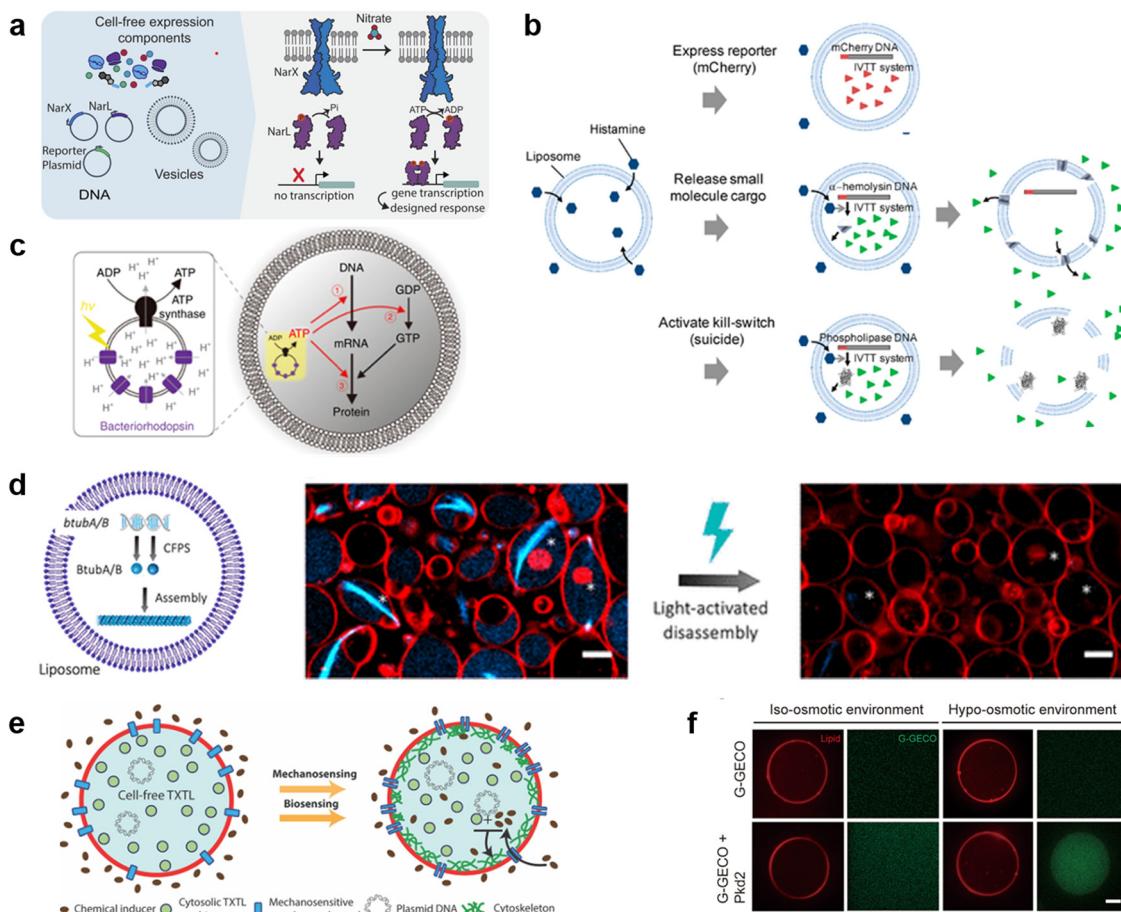


Fig. 9 Cell-free protein synthesis system (CFPS) DNA for signal processing. **a**, Using a two-component sensing system (NarX–NarL) to sense various substances.²¹⁷ **b**, Controlling the fate of GUVs with a histamine-responsive riboswitch.²¹⁹ **c**, Synthesis of a reporter protein using light irradiation of ATP synthase and bacteriorhodopsin-containing GUVs.²²³ **d**, Deforming GUVs from elliptical to spherical shape by degrading CFPS-synthesized BtubA/B formed microtubules into monomers using light irradiation. Scale bars: 5 μm .²²⁴ **e**, Expression of a bacterial cytoskeletal protein with GUVs loaded with CFPS and the mechanosensitive protein MscL by exposing the GUVs to hypo-osmotic solution.²²⁵ **f**, Activating the CFPS-synthesized ion channel Pkd2 with osmotic stress on the membrane to enhance the influx of calcium ions. Scale bars: 10 μm .²²⁶ Figures reproduced with permission from: a, ref. 217, copyright 2023, National Academy of Sciences; b, ref. 219, copyright 2019, American Chemical Society; c, ref. 223, copyright 2020, Nature Publishing Group; d, ref. 224, copyright 2021, American Chemical Society; e, ref. 225, copyright 2019, American Chemical Society; f, ref. 226, copyright 2022, American Society for Cell Biology.

reporter protein, pore-forming protein, or phospholipase, which allowed the GUVs to show fluorescence, release small chemicals, or self-destruct, respectively²¹⁹ (Fig. 9(b)). Other examples of chemical signalling control over molecular robots include membrane deformation of GUVs by external supply of FtsZ synthesized by CFPS,²²⁰ a fusion of two different vesicles by calcium and polyethylene glycol to initiate protein synthesis inside GUVs,²²¹ and driving an in-chip and lipid-compartmentalized protein synthesis using CFPS by supplying an energetic substance to the outside of the lipid membrane.²²²

In addition to chemical signalling, light and osmotic stress have been used as physical input signals. Kuruma's group synthesized F0F1 ATP synthase and bacteriorhodopsin using CFPS and incorporated both into small lipid vesicles to prepare energy-generating proteoliposome (PL). Upon light irradiation, bacteriorhodopsin created a proton gradient across the

membrane, which then was used by ATP synthase for the phosphorylation of ADP to ATP. The PL encapsulated inside the GUVs was used as an ATP regeneration system and using the generated ATP, GFP synthesis was performed with CFPS²²³ (Fig. 9(c)). Danelon's group showed the synthesis of BtubA/B microtubule formation inside of GUVs, which ultimately deformed GUVs from spherical to elliptical shapes. Light irradiation cleaved the microtubules into monomers and restored the spherical shape of GUVs²²⁴ (Fig. 9(d)). When MscL, a mechanosensitive protein, was synthesized using CFPS inside GUVs and osmotic stress was applied, MscL changed its conformation and opened a pore, resulting in the influx of an inducer which then triggered the expression of a bacterial cytoskeletal protein²²⁵ (Fig. 9(e)). Another example synthesized Pkd2, an ion channel, inside GUVs and upon an increase in osmotic stress, an influx of Ca^{2+} ions was observed *via* the binding of the ion with its indicator G-GECO²²⁶ (Fig. 9(f)).

As described above, molecular robots, in response to chemical and physical inputs, have exhibited various outputs using CFPS as a computer. For chemical input signals, the permeability of the chemicals plays an important role. Membrane-permeable chemicals can directly affect the CFPS inside the GUVs, whilst membrane-impermeable chemicals require receptors or transporters to transmit the chemical signal to the intravesicular CFPS. Several examples have been reported,^{217,227} however, since many input signalling molecules are charged and have little membrane permeability, the development of a sensing domain using receptors or transporters is a future challenge. For physical inputs, temperature shift, pH change, redox state, and electron transfer are yet to be realized. As for the outputs, many studies are limited to the expression of reporter proteins. These are sufficient as proof-of-concept experiments or sensor developments. However, more sophisticated output including motility and cooperative interaction with extravehicular environments are yet to be realized.

Actuators of molecular robots

Molecular robots that achieve complicated tasks are desired to exhibit specific physical movements such as deformation and propulsion. So far, we have demonstrated the power of DNA and proteins as molecular tools to construct sensors and computers for molecular robots. In a similar vein, DNA and proteins also provide a unique path forward in building actuators for molecular robots.^{228,229} This section aims to give a brief overview of the strategies to actuate molecular robots.

DNA nanotechnology

DNA nanotechnology enables exquisite control over the structure of self-assembled macromolecular and nanoscale motifs.²³⁰ DNA nanodevices are however far from static and have been engineered to reconfigure, change shape, and move in response to a wide array of stimuli.^{230,231} From the ground-breaking examples of DNA tweezers¹⁹⁷ and walkers,²³² to the origami crank-sliders and joints demonstrated by the Castro group,²³³ to the bioinspired rotors built by the Dietz and Simmel groups,^{234,235} DNA nanomachines and nano-actuators have demonstrated an unparalleled ability to control different types of motion at the nanoscale. Furthermore, these nanodevices can be actuated through a variety of different stimuli, from strand displacement²³⁶ to the species and concentration of cations,^{237,238} to changes in pH,²³⁷ light exposure,²³⁹ and enzymatic action.²⁴⁰

As discussed in the section on DNA nanopores, DNA nanostructures can be mechanically coupled to lipid membranes using lipophilic anchors, typically cholesterol or tocopherol.^{241,242} This coupling unlocks vast opportunities to engineer both the morphology and dynamic responses of GUV-based microrobots and artificial cells through membrane-anchored DNA devices that imitate the functions of membrane proteins.

Membrane adhesion is among the most basic functions mediated by cell-surface receptors, underpinning a plethora of biological processes, including motility, tissue formation, mechanosensing, and endocytosis. Exploiting the selectivity of base-pairing interactions, synthetic, membrane-anchored DNA linkers have been used to induce and program adhesion between lipid membranes²⁴² (Fig. 10(a)), starting with the seminal works of Höök and coworkers,^{243,244} Boxer and coworkers^{245,246} and Beales and Vanderlick^{247,248} (Fig. 10(b)). Parolini *et al.* have then demonstrated the DNA-mediated assembly of thermoresponsive synthetic tissues²⁴⁹ and, leveraging toehold-exchange reactions, established control over the kinetics of tissue formation.²⁵⁰

Membrane fusion is another ubiquitous mechanism that living cells use to manipulate the structure and composition of their lipid membranes, manifesting, for instance, in neurotransmitter release from synaptic vesicles and in the invasion by enveloped viruses. Dynamic DNA nanostructures have been designed to replicate the response of extant fusogenic protein machinery, notably SNARE proteins.²⁵¹ Höök and co-workers first demonstrated that zipper-like DNA constructs anchored to membranes can induce fusion by bringing the membranes into molecular proximity,²⁵² as shown in Fig. 10(c). The efficiency of fusion has been observed to depend on nanostructure design, for instance the presence of non-binding DNA spacers between the lipophilic anchors and the zipping domains, as well as on the number and chemistry of the hydrophobic anchors and the surface density of the DNA-zippers.^{253,254} Later designs have included dsDNA “tendrils”, where the fusogenic zipping action is mediated by four-way branch migration, which were found to improve fusion efficiency and facilitate its modulation with soluble DNA strands²⁵⁵ (Fig. 10(d)). Membrane composition can also significantly influence fusion efficiency,^{252,253} which increases substantially in the presence of conical lipid species (*e.g.* DOPE)²⁵⁵ and for phase-separated membranes.²⁵⁶ Fusogenic DNA constructs have been applied to engineer complex behaviors in artificial cell systems, for instance triggering CFPS²⁵⁶ (Fig. 10(e)) and mediating intricate fusion pathways that mimic cell differentiation.²⁵⁷ Such nanodevices have also been shown to facilitate fusion between synthetic and biological cells for the purpose of intra-cellular delivery²⁵⁸ or to generate “hybrid” cells.²⁵⁹

Similar to smaller nanostructures, DNA origami can be linked to membranes *via* multiple hydrophobic moieties.²⁶⁰ Owing to their larger size, stiffness, and precisely programmable 3D geometry, membrane-bound origami can be used to influence the morphology of synthetic lipid membranes, mimicking membrane-sculpting proteins. Czogalla *et al.* first demonstrated that, for sufficiently high coverage, polymerizable, brick-like DNA origami can induce large-scale deformation in GUVs, which were found to lose their equilibrium spherical shape.²⁶⁰ Franquelim *et al.* later showed that GUVs decorated with convex origami imitating natural bar-domain proteins, acquired shallow invaginations upon osmotic deflation.²⁶¹ In turn, functionalization with



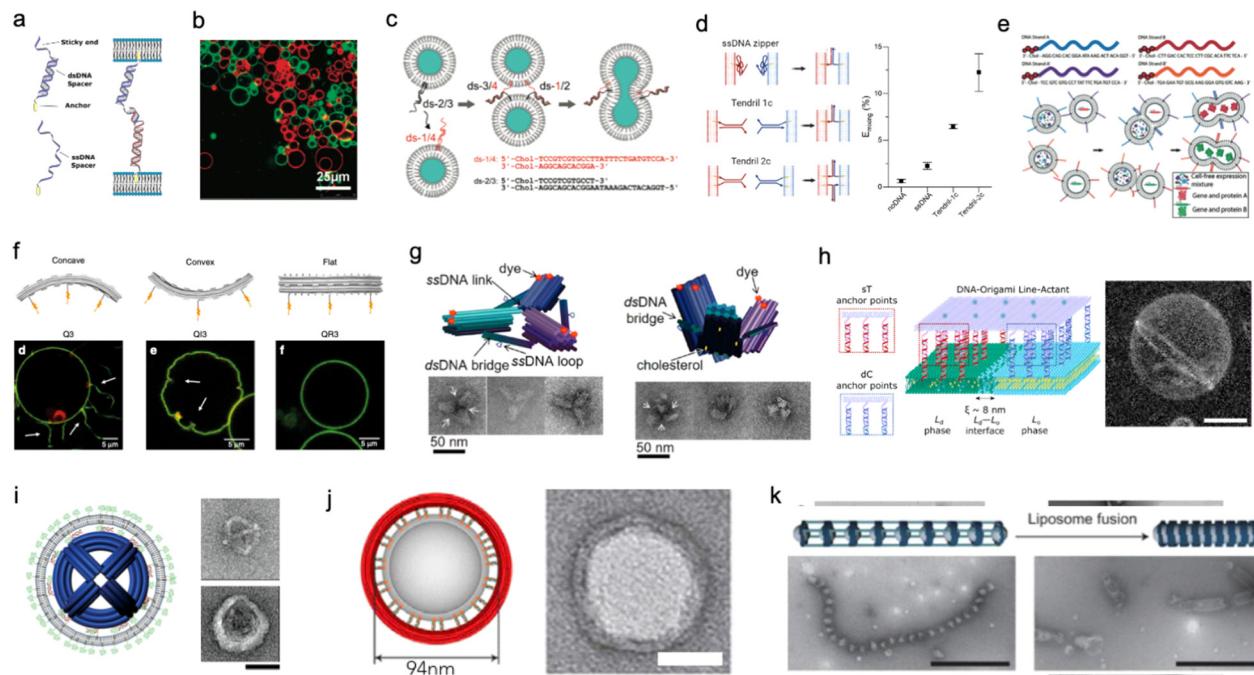


Fig. 10 DNA nanostructures for actuating lipid membranes. **a**, Schematic of DNA linkers anchored to membranes and mediating adhesion between lipid vesicles. Linkers may feature a double-stranded DNA spacer (top) or be fully single-stranded (bottom).²⁴² **b**, Confocal micrograph of GUVs adhering due the action of single-stranded DNA linkers (right), adapted from ref. 247. **c**, DNA “zipper” constructs mediating lipid vesicle fusion.²⁵² **d**, Comparison of fusion efficiency (right) from various DNA zipper and “tendril” constructs (left).²⁵⁵ **e**, Cascades of lipid vesicle fusion reactions mediated by DNA zippers trigger CFPS.²⁵⁶ **f**, Curved DNA origami (top) influence the morphology of GUVs as determined with confocal microscopy (bottom).²⁶¹ **g**, Clathrin-like DNA origami triskelia of controllable curvature (top) and their TEM images (bottom).²⁶² **h**, DNA origami line-actants (left) accumulate at the interface between liquid ordered and liquid disordered domains in phase-separated GUVs (confocal projection, right) thanks to the phase-selectivity of double cholesterol (dC) and single tocopherol (sT) anchors. Scale bar: 10 μ m.²⁷¹ **i**, Virus-like DNA particle obtained by templating the formation of a lipid vesicle around a spherical DNA origami decorated with lipids (left) and TEM images of a bare (top right) and lipid-enveloped (bottom right) origami. Scale bar: 50 nm.²⁷² **j**, Lipid vesicles of controlled size templated by lipid-modified DNA-origami rings. Images are TEM micrographs of the constructs shown on the immediate left. Scale bars: 50 nm.³⁶² **k**, Liposomes captured by dynamic DNA origami arrays. Scale bars: 100 nm.³⁶³ Figures reproduced with permission from: **a**, ref. 242, copyright 2019, IOP Publishing; **b**, ref. 247, copyright 2007, American Chemical Society; **c**, ref. 252, copyright 2008, American Chemical Society; **d**, ref. 255, copyright 2022, Royal Chemical Society; **e**, ref. 256, copyright 2019, Wiley-VCH; **f**, ref. 261, copyright 2018, Nature Publishing Group; **g**, ref. 262, copyright 2019, American Chemical Society; **h**, ref. 271, copyright 2023, American Chemical Society; **i**, ref. 272, copyright 2014, American Chemical Society; **j**, ref. 362, copyright 2017, Elsevier; **k**, ref. 363, copyright 2020, Wiley-VCH.

concave origami induced tubulation, which became more prominent for increasing origami coverage²⁶¹ (Fig. 10(f)). Following a similar principle, Journot *et al.* designed convex DNA-origami “triskelia” resembling clathrin proteins. The origami triskelia could be polymerized to form a two-dimensional network, producing membrane deformation reminiscent of clathrin-coated invaginations²⁶² (Fig. 10(g)).

Membrane-adhesion and the polymerization state of DNA nanostructures can be externally controlled through a variety of different mechanisms, including strand displacement reactions,^{263,264} pH-responsive non-canonical DNA constructs,²⁶⁵ light-responsive azobenzene-modified DNA linkers,²⁶⁶ and by modulating ionic composition and membrane phase.²⁶⁷ These tools open up vast opportunities for controlling dynamic reconfiguration in lipid-based microrobots and engineering biomimetic pathways for environmental adaptation.

The differences in affinity that different lipophilic anchors display for co-existing lipid phases also offer a useful

mechanism for programming dynamic responses in DNA-functionalised synthetic membranes.^{268,269} Rubio-Sánchez *et al.* considered phase-separated GUVs with liquid-ordered (Lo) and liquid disordered (Ld) domains and decorated them with DNA nanostructures using both double-cholesterol (dC) and single-tocopherol (sT) anchors, which display a preference for Lo and Ld, respectively.²⁷⁰ The authors demonstrated that connecting and disconnecting selected anchors *via* strand displacement could induce the redistribution of the DNA nanostructures on the surface of the GUVs – a simple example of directed cargo transport. Specifically, linking dC and removing sT would cause the devices to accumulate in the Lo phase, while removing the dC and bringing back the sT would trigger migration to Ld.²⁷⁰ In a subsequent contribution, Rubio-Sánchez *et al.* noted that DNA origami plates functionalized with both dC and sT, arranged at opposite ends of the plate, showed the tendency to accumulate at the line-interface between Lo and Ld (Fig. 10(h)). These DNA-origami line-actants (DOLAs) were

shown to reversibly stabilize small lipid domains against coalescence, establishing a route to control the surface patterning of GUVs. DOLAs were also shown to provide control over a biomimetic membrane-fission pathway, whereby budding-off of DOLA-stabilised domains could be induced upon de-activating the line-actants under hyperosmolar conditions²⁷¹ (Fig. 10(h)).

When the overall size of the lipid vesicles one seeks to engineer approaches the nanoscale, DNA origami can be used to directly template the formation of vesicles with precisely controlled size and shape. This concept was first proposed by Perrault and Shih, who used spherical, lipid-modified DNA-origami to template the formation of monodisperse lipid vesicles (diameter ~76 nm)²⁷² (Fig. 10(i)). The lipid vesicles would form around the origami following lipid addition and removal of an initially present surfactant. The lipid envelope was found to protect the DNA nanostructure from enzymatic degradation – a useful feature for envisaged *in vivo* deployment.²⁷² A similar strategy has been pursued by Yang *et al.* using DNA-origami rings of various diameters that, thanks to a lipid-modified inner edge, could template the growth of size-controlled lipid vesicles in a Saturn-ring-like geometry²⁷³ (Fig. 10(j)). Zhang *et al.* later demonstrated that similar DNA-origami exoskeletons could be polymerized, generating arrays of lipid vesicles with controlled inter-membrane distance, or tubular vesicles²⁷⁴ (Fig. 10(k)). The devices could also be actuated to induce lipid vesicle fusion into and bending of the lipid tubes.²⁷⁴ In addition to providing control over lipid architectures, lipid vesicle-templating DNA origami has also been applied to precisely position molecules onto membranes and study reaction kinetics with absolute control over stoichiometry.²⁷⁵

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Protein and peptides

Owing to their high affinity to lipid membranes, proteins and peptides act as promising actuators in molecular robots, in the same way that they deform or propel living cells. Deformation could allow molecular robots to overcome complex terrains. Various proteins/peptides have been reported to deform GUVs.^{17,276–283} Membrane deformation can be classified into two broad categories: outward

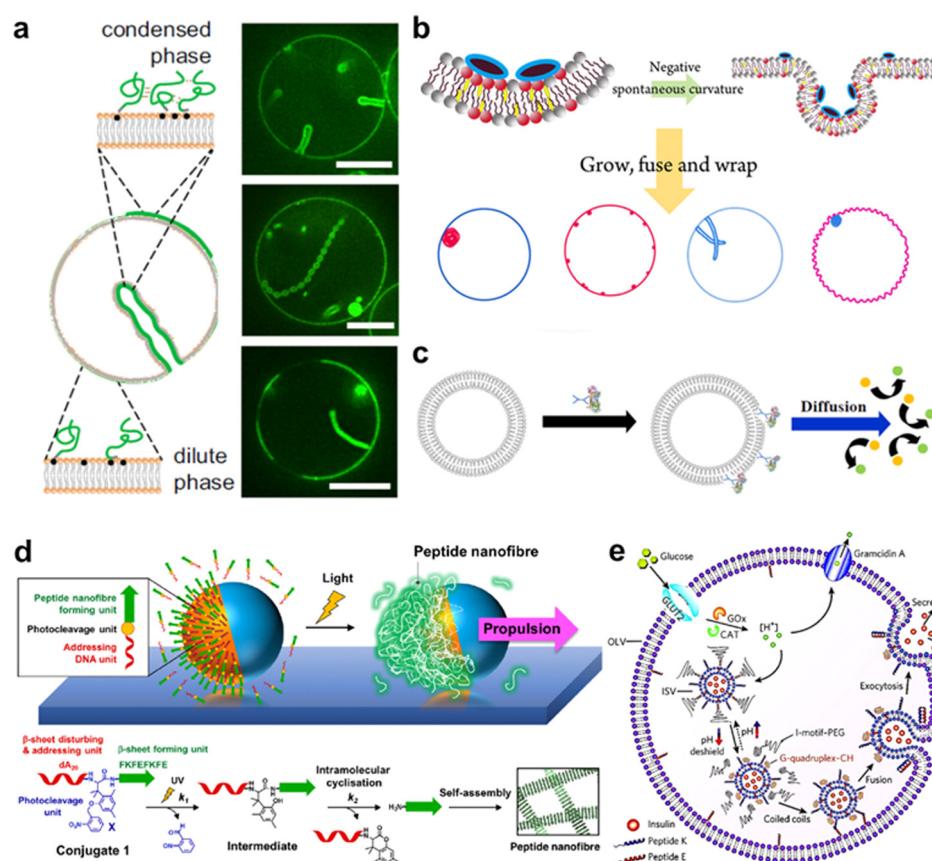


Fig. 11 Proteins and peptides for actuating lipid membranes. **a**, GUV deformation triggered by phase separation of FUS LC adhered on the lipid membrane. Scale bars: 10 μm .²⁷⁹ **b**, GUV deformation triggered by *de novo* peptides.²⁸⁵ **c**, Propulsion of ATPase-coated GUVs.²⁸⁶ **d**, Light-induced propulsion of a phase-separated GUVs driven by local peptide nanofibre growth (top) and the photocleavage reaction of a DNA-peptide conjugate (bottom).²⁸⁹ **e**, Illustration of the biochemical process inside OLVs. Vesicle fusion is triggered by the interaction between peptide K and peptide E.³⁶⁴ Figures reproduced with permission from: a, ref. 279, copyright 2021, National Academy of Sciences; b, ref. 285, copyright 2018, American Chemical Society; c, ref. 286, copyright 2019, American Chemical Society; d, ref. 289, copyright 2018, Nature Publishing Group; e, ref. 364, copyright 2021, Royal Chemical Society.



deformation and inward deformation. Inward deformation is considered more energetically unfavorable because it leads to membrane expansion.²⁸⁴ As a consequence, most of the previous studies focus on outward deformation. However, inward deformation is also an attractive route towards molecular robots with increased functionality, such as endocytosis-like membrane fission. In 2021, Yuan *et al.* reported inward bending of the membrane of phase-separated GUVs of membrane-bondable N-terminal low-complexity domain of fused in sarcoma (FUS LC) proteins²⁷⁹ (Fig. 11(a)). Meanwhile, Yu *et al.* designed a *de novo* amphiphilic bola-type peptide composed of lysine and leucine that regulates inward budding of lipid membranes²⁸⁵ (Fig. 11(b)).

Propulsion could help molecular robots to reach their various destinations. One strategy to perform propulsion is to make use of the diffusive movement of enzymes. In 2019, Ghosh *et al.* showed that GUVs with ATPase-tagged lipid membranes exhibited enhanced mobility in the presence of ATP.²⁸⁶ In 2023, Jin *et al.* demonstrated that horseradish peroxidase (HRP)-decorated Janus GUVs undergo directional motion in the presence of H₂O₂, while such enhanced mobility is absent from homogeneously enzyme-decorated GUVs²⁸⁷ (Fig. 11(c)). Another attempt reported by Kurakazu *et al.* involved GUV propulsion using motor protein flagella isolated from *Chlamydomonas*.²⁸⁸ Interestingly, the mobility was further enhanced when the GUVs were coated with annexin. On the other hand, there are no reports of peptide-based GUV propulsion, with the need for peptides to convert chemical energy to kinetic energy.

Morphological change with a combination of proteins/peptides and DNA is also of great interest owing to the high programmability and versatility of DNA. A groundbreaking work by Sato *et al.* reported a system using kinesin-DNA complexes to deform GUVs. The group showed that kinesin can be attached to lipid membranes using a light-triggered DNA displacement reaction, leading to a continuous shape change of the GUVs driven by microtubule sliding on the membrane.¹⁵ Meanwhile, Inaba *et al.* have developed a system in which propulsion is achieved by partial growth of peptide nanofibers on a phase-separated membrane.²⁸⁹ In their system, photocleavable peptide-DNA complexes are specifically conjugated to the lipid-disordered membrane. Upon exposure to light, the peptides were released from the membrane and locally self-assembled into nanofibers, resulting in autonomous GUVs propulsion (Fig. 11(d)).

Unique applications have been realised when combining protein/peptide actuators with multi-compartment architecture, such as vesicles-in-vesicles. Chen *et al.* engineered 'artificial beta cells (A β Cs)' that sense glucose in solution and release insulin by membrane fusion.²⁹⁰ They first prepared insulin-encapsulated inner small vesicles (ISVs) with a membrane composed of lipids, peptide (peptide K), and pH-sensitive polyethylene glycol (PEG) conjugates which detach from the membrane at a low pH environment. A mixture of ISVs, glucose oxidase, and

catalase are then encapsulated in outer large vesicles (OLVs) containing a lipid membrane with peptide E (complementary to peptide K) and glucose transporters. When glucose was transported from the outside to the inside of the OLVs, a low pH environment was generated due to the glucose oxidase and catalase reaction. The pH decrease triggered PEG detachment from the membrane, and consequently, the unshielded peptide K formed coiled coils with peptide E, promoting the fusion of two vesicles and the release of insulin (Fig. 11(e)). We anticipate future research to develop in the use of proteins/peptides to form complex structures, fine-tuning of morphology changes, and cooperative interaction with each other.

Applications of molecular robots

Robots that can extend the reach of human engineering capability in the microscale are in high demand. Compared to micro-sized robots using alloys,^{291,292} oxide minerals,^{293,294} and ceramics,²⁹⁵ molecular robots are biocompatible and biodegradable due to their use of biological materials. This also affords versatility in biological environments. At the same time, the large toolbox of sensors, computers, and actuators, as mentioned above, enables them to execute multiple and complicated tasks which are challenging for alternatives such as hydrogel-based microswimmers.^{296,297} Additionally, the leakage issues for such microswimmers are resolved in molecular robots owing to the lipid membrane functionality as a semipermeable barrier. Accordingly, molecular robots have exhibited great potential in various applications as a promising alternative to conventional microrobots. In this section, we will provide some representative examples of emerging state-of-the-art applications of molecular robots.

Detection of molecules

Small toxic molecules (≤ 1000 daltons), for example, heavy metals and organochlorine, are well-known pollutants in water. Though spectroscopic methods such as inductively coupled plasma atomic emission spectroscopy and mass spectrometry are powerful for molecular detection, the requirement of real-time monitoring of the local molecular concentrations cannot be met with such methods in principle.²⁹⁸ On the other hand, molecular robots with sensors and computers may be able to fill this gap. Previous studies have reported that GUVs with nanopores are useful for the detection of K⁺,⁵⁶ Cu²⁺,²⁹⁹ and Cl⁻.³⁰⁰ Recently, a unique strategy using GUVs with a DNA nanopore and circuit offered by Peng *et al.* suggested switchable detection of Sr²⁺ (ref. 141) (Fig. 12(a)). In the presence of ATP, the DNA nanopore switches from closed to open, which enhances the influx of environmental Sr²⁺. The Sr²⁺ sequentially triggers a set of confined downstream cascade reactions to output both a fluorescence signal and switching of the nanopores back to the closed state. Another pioneering work that uses CFPS inside GUVs to detect fluoride was demonstrated by Boyd



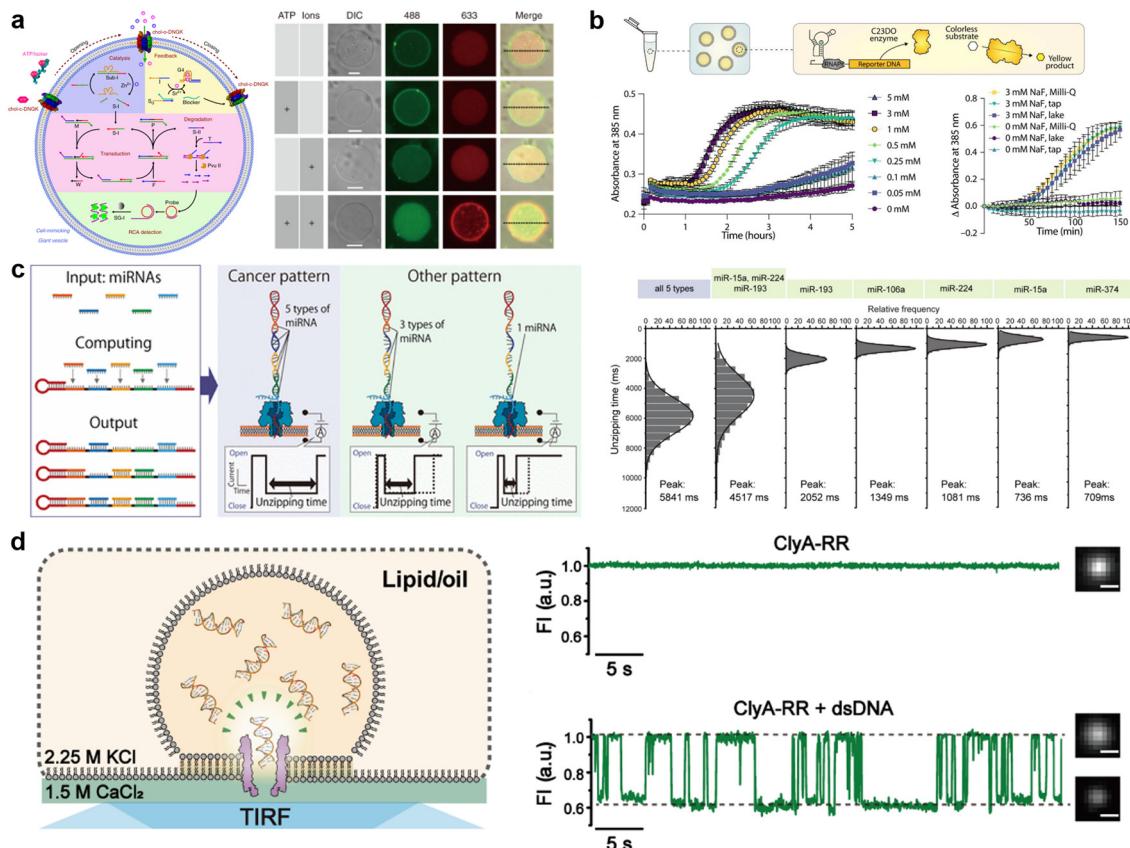


Fig. 12 Molecular robots for detection of molecules. **a**, Detection of Sr^{2+} by GUVs with a DNA nanopore and circuit (left), as observed by the increase of 488 fluorescent signals from SG-I in the presence of ATP and Sr^{2+} (right).¹⁴¹ **b**, Illustration of the GUVs designed to detect sodium fluoride (top) and its absorbance over time in response to *in vitro* samples (bottom left) and real-world samples (bottom right).²¹⁸ **c**, Using nanopores and DNA computing to detect five types of miRNAs (left) from histograms of the unzipping time of each miRNA pattern (right).³⁰⁴ **d**, Illustration of DiffusiOptoPhysiology (DOP) method (left). Comparison of the signal of Fluo-8 with/without dsDNA in the solution (right).³¹⁰ Figures reproduced with permission from: a, ref. 141, copyright 2020, Nature Publishing Group; b, ref. 218, copyright 2023, AAAS; c, ref. 304, copyright 2022, American Chemical Society; d, ref. 310, copyright 2019, AAAS.

*et al.*²¹⁸ In their system, the membrane-permeable sodium fluoride (NaF) initializes the riboswitch-mediated synthesis of catechol (2,3)-dioxygenase (C23DO), which subsequently catalyzes the conversion of its colorless substrate, catechol, to the yellow-colored 2-hydroxymuconate semialdehyde, constituting a colorimetric response (Fig. 12(b)). Importantly, they successfully showed the possibility of detecting fluoride in real-world samples from Lake Michigan and the Evanston, IL municipal tap water supply. Though the sensitivity of molecular robots remains inferior to that of spectroscopic methods, they provide the opportunity to probe in detail the concentration of small molecules in a biological environment. We infer that research into other novel applications, for example, real-time monitoring of the pollutants in aquatic life, is also underway.³⁰¹

Biomarkers such as DNA, microRNAs, and peptides are another essential target for molecular robots. So far, we have reported that nanopores have a strong synergy with DNA computing to allow for highly sensitive detection.^{13,302–304} In our most recent study, we proposed a system for pattern recognition of five types of miRNAs overexpressed in bile

duct cancer (BDC) with an α HL nanopore, diagnostic DNA, and a planar lipid membrane³⁰⁴ (Fig. 12(c)). With our system, we succeeded in the label-free detection of miRNA expression patterns from the plasma of BDC patients at sub-femtomolar concentrations. On the other hand, detecting peptides using nanopores is more challenging due to peptides' relatively small size and heterogeneous charge compared with polynucleotides. During the last decades, academic efforts have been invested into designing mutant nanopores^{305,306} or optimizing solution conditions^{307,308} to address the above problems. However, there remains a long way before the realisation of peptide detection with amino acid resolution. Meanwhile, biomarker detection with GUV-based molecular robots also likely faces an arduous path to development. Liu *et al.* reported GUVs that detect environmental DNA messengers,¹⁵⁰ while Barba-Bon *et al.* have used GUVs to monitor the transport of cell-penetrating peptides.³⁰⁹ Nevertheless, the sensitivity and amount of information conferred using fluorescence-based detection is several times lower compared with electrical recording technology. Recently, the Huang group has reported an electrode-free

nanopore sensing system by a so-called DiffusiOptoPhysiology (DOP) method, which optically monitors fluorescence emission resulting from diffusive binding of Ca^{2+} with its indicator dye Fluo-8 through a nanopore on a planar lipid membrane by total internal reflection fluorescence, and detected dsDNA^{310,311} (Fig. 12(d)). Accordingly, the combination of GUV-based molecular robots with new technology (for example, single-molecule localization microscopy³¹² and stimulated emission depletion microscopy³¹³) is worth exploring to facilitate higher levels of detection sensitivity.

Communication with living cells

Engineering communication between molecular robots and living cells would provide novel strategies to address issues in fields such as drug development or bioremediation. Communication is a process of signal production and transmission. Attempts have been made towards both DNA computing^{256,314,315} and CFPS^{316,317} for signal processing.

The use of such systems can greatly improve the complexity and programmability of signalling, endowing molecular robots with the potential of performing complicated tasks. For instance, Adamala *et al.* constructed two populations of lipid vesicles that contain different genetic circuits called “sensor liposomes” and “reporter liposomes”.³¹⁷ When the membrane-permeable arabinose (Ara) was applied to the bulk solution, α HL expression started in the sensor liposomes, consequently enabling the release of the non-membrane-permeable activator β -D-1-thiogalactopyranoside (IPTG). The IPTG was then recognized by the reporter liposomes, triggering the output of luciferase (Fig. 13(a)).

Signal transmission of molecular robots can be accomplished by nanopores including α HL,³¹⁷⁻³²⁰ melittin,^{321,322} and perfringolysin O (PFO).³¹⁶ Importantly, the nanopores not only facilitate the transmission of signals but can also cause leakage problems of other encapsulated molecules. For this reason, heptakis(2,3,6-tri-O-methyl)-beta-cyclodextrin (TRIMEB), which is noncovalently bound into the lumen of α HL, is sometimes introduced to prevent the

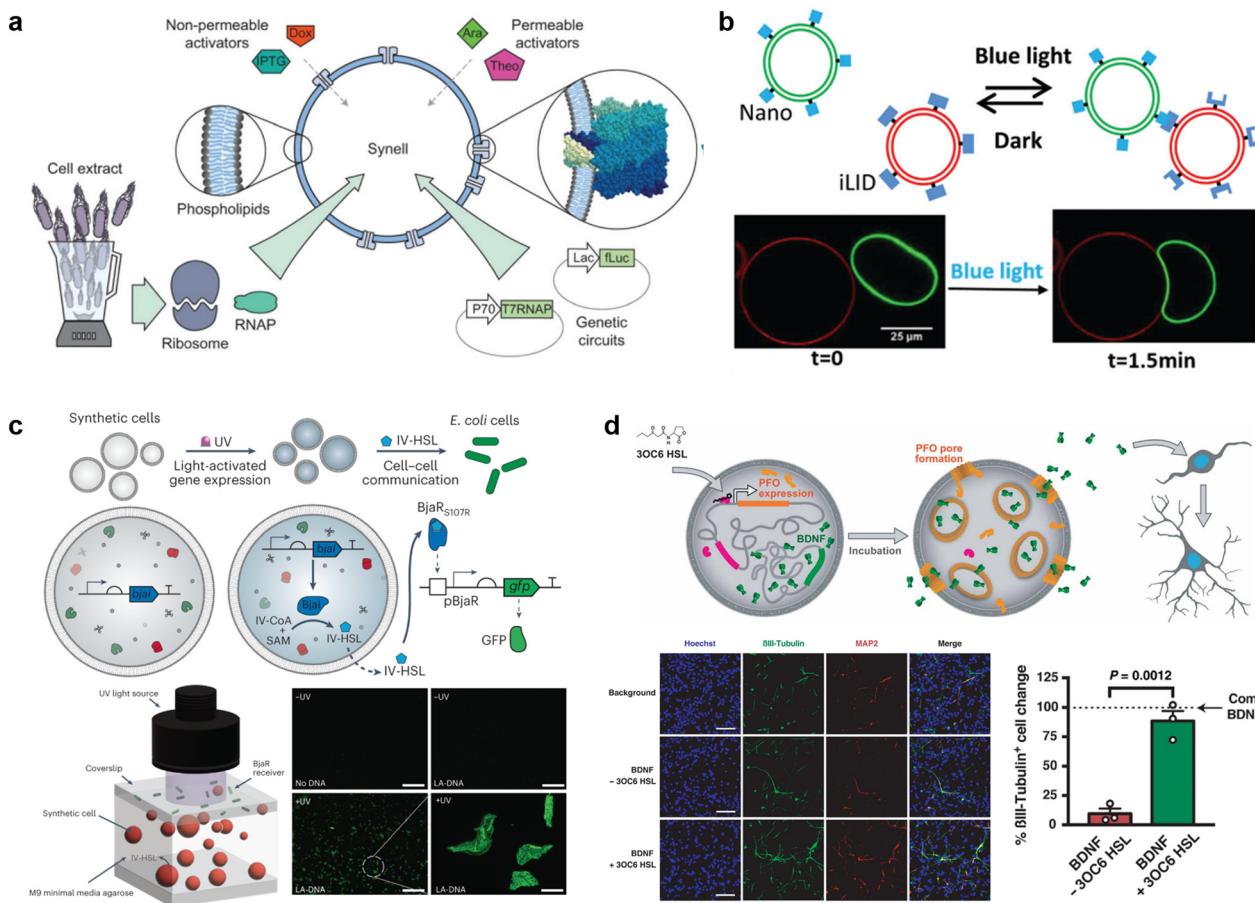


Fig. 13 Molecular robots for communication with living cells. **a**, Modular design of genetic circuit interactions within and between GUVs.³⁶⁵ **b**, Controlling the distance between GUVs with light-oxygen-voltage 2 protein. Scale bar = 25 μm .³²⁶ **c**, Schematic representation of communication between GUVs with bacteria (top). GFP is expressed only when the GUVs are exposed to UV light (bottom). Scale bar = 200 μm . Scale bar (bottom right) = 20 μm .³²⁹ **d**, Schematic representation of communication between GUVs with neural stem cells (top). GUVs function to enhance stem cell differentiation, as revealed by the increase in the percentage of bIII-tubulin-overexpressing neurons (bottom). Scale bar = 50 μm .³¹⁶ Figures reproduced with permission from: a, ref. 365, copyright 2020, Royal Chemical Society; b, ref. 326, copyright 2019, Wiley-VCH; c, ref. 329, copyright 2023, Nature Publishing Group; d, ref. 316, copyright 2020, AAAS.



leakage of encapsulated content.^{319,320} Meanwhile, considering the approximation diffusion equation $t = L^2/2D$ (here, t is the elapsed time since signal transmission began, L is the distance from the GUVs to the target, and D is the diffusion coefficient), the distance between the GUVs also plays an important role in signal transmission. To this end, significant advances have been made in recent years to engineer membrane adhesion with DNA nanostructures³²³ and proteins.^{319,324,325} Notably, the Wegner group showed that membrane adhesion could be controlled with the use of an improved light-induced dimer protein based on the light-oxygen-voltage 2 (LOV2) domain from *Avena sativa*, opening up a new horizon for designing efficient communication networks^{319,325,326} (Fig. 13(b)).

The above progress has paved the way for the realization of communication between molecular robots and living cells. Quorum sensing, which is the communication behavior of cells to detect and respond to cell population density, has been extended to molecular robots and bacterial populations.^{327–329} Remarkably, Smith *et al.* reported light-activated GUVs that could perform quorum-sensing-based communication with bacteria³²⁹ (Fig. 13(c)). The system is achieved by installing the T7 promoters of DNA templates with photocleavable biotinylated (PCB) and monovalent streptavidin (mSA) that could impede T7 RNA polymerase from binding to the T7 promoter. When exposed to UV light, the mSA was liberated, resulting in the expression of the BjaI enzyme followed by the production of *N*-isovaleryl-L-homoserine lactone (IV-HSL) in GUVs. As the bacterial populations contained a gene circuit highly sensitive to IV-HSL, they responded to the IV-HSL signal with resultant GFP expression. This work sets the stage for the development of a remote-controlled molecular robot for communication with living cells. Meanwhile, communication between molecular robots and mammalian cells provides another area of interest. A remarkable work by Kaneda *et al.* reported that calcien dye could be transferred from Connexin43 (Cx43)-expressing GUVs to cultured Cx43-expressing U2OS cells through the gap junction.³³⁰ Recently, Toparlak *et al.* demonstrated GUVs that can drive mammal neural differentiation.³¹⁶ The GUVs contained transcription-translation machinery and DNA templates that coded for brain-derived neurotrophic factor (BDNF), LuxR, and PFO. In the presence of *N*-3-oxohexanoyl homoserine lactone (3OC6 HSL)-responsive transcriptional repressor LuxR, PFO was expressed and assembled into nanopores in lipid membranes, enabling the release of BDNF. After 19-day coincubation with mouse embryonic stem cell-derived neural stem (mNS) cells, the group successfully observed the capability of GUVs to enhance the neural differentiation and maturation, with an increase in pan-neuronal and mature neuronal markers β III-tubulin and microtubule-associated protein 2 (MAP2) (Fig. 13(d)). We anticipate that future efforts will be geared toward adapting multiple molecular robots and living cell populations to build communication networks with higher complexity.

Conversion of energy

The operation of sensors, computers, and actuators in molecular robots requires energy sources, hence ATP is sometimes encapsulated in GUVs or added to the external solution.^{141,316} Moreover, such molecular robots may also provide solutions to global energy issues. In a living organism, energy production is powered by a transmembrane proton gradient, meaning that the membrane-spanning ATP synthase turns ADP into ATP only when there is a proton gradient.³³¹ In thylakoids, this proton gradient is caused by a light energy-driven proton pump. Accordingly, it has attracted the interest of scientists to incorporate ATP synthase and proton pumps into GUVs to build a self-fuelling system. A groundbreaking work in this field was reported by Lee *et al.* in 2018.³³² The group built an artificial organelle containing ATP synthases and two light-driven proton pumps, photosystem II (PSII) and proteorhodopsin (PR), and encapsulated it into GUVs. The PSII is a proton pump that can be activated by red light, whereas PR proton pumping is mainly initiated by green light. Moreover, due to the pH-dependent bidirectional proton-pumping ability of PR, PR and PSII work in conjunction to increase the proton gradient at low pH, but counteract the action of PSII at high pH. This makes it possible to facilitate or impede ATP synthesis with exposure to different colors of light. The ATP-dependent polymerization of cytoskeletal proteins was used to confirm ATP synthesis. As expected, the red light facilitated and the green light impeded the growth of cytoskeletal proteins, responding to the operation of PR and PSII. Moreover, the polymerized cytoskeletal proteins showed the ability to deform GUVs with an adjusted lipid composition (Fig. 14(a)). A similar idea was demonstrated by Berhanu *et al.*²²³ Instead of cytoskeletal protein reaction, successful ATP synthesis was monitored by GFP expression in a CFPS.

In eukaryotic cells, energy conversion mainly occurs in the mitochondria. Li *et al.* encapsulated mitochondria extracted from C6 glioma cells into GUVs and showed ATP synthesis with the addition of pyruvate which is a trigger molecule to stimulate mitochondria to produce ATP³³³ (Fig. 14(b)). Though this work paves the way to develop mitochondria-based self-fuelling systems in GUVs, it remains difficult to construct an artificial mitochondrion from scratch, due to the complexity of the citric acid cycle and the electron transport chain. Another fascinating work demonstrated by Hicks *et al.* inserted carbon nanotubes in GUVs to mimic voltage-dependent anion-selective channels, which were able to transfer electrons *via* redox upon a change in potential.³³⁴ The group showed that a 1.5 V voltage stimulation triggers the reduction of gold chloride to solid gold deposits on the carbon nanotube terminus (Fig. 14(c)). This work opens up the potential for nanopore transistors, able to convert electrical energy to chemical energy in molecular robots. Subsequent attempts in this field focus on improving the energy conversion efficiency to accelerate the development of molecular robots towards practical applications.



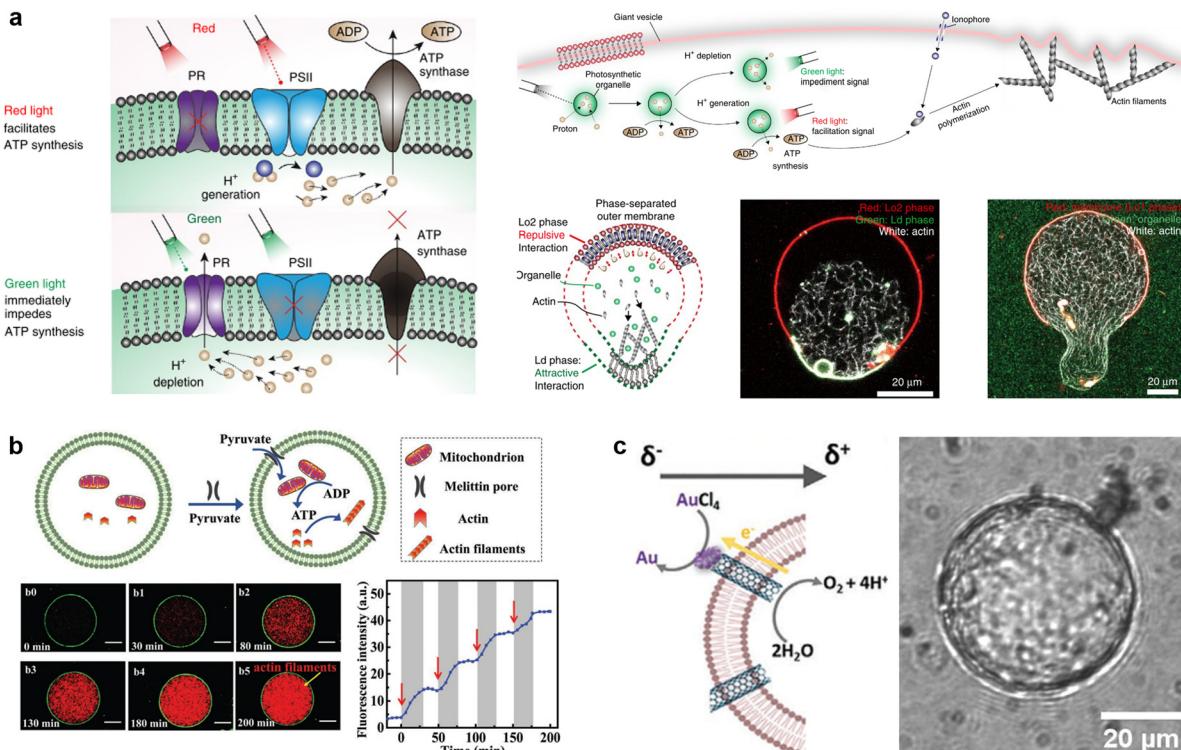


Fig. 14 Molecular robots for conversion of energy. **a**, Facilitating or impeding the ATP synthesis with different light color exposure. (left) Optical stimulation couples ATP synthesis with ATP-dependent actin polymerization and morphological change of the GUVs (top right). Confirmation of the formed cytoskeletal proteins and their impact on membrane deformation with microscopy (bottom right). Scale bar = 20 μm .^{366,367} **b**, Mitochondrion-containing GUVs' response to pyruvate addition (top). Time-dependent process of cytoskeletal proteins polymerization triggered by the addition of pyruvate (bottom). Scale bar = 10 μm .³³³ **c**, Schematic showing polarization of carbon nanotubes within lipid membranes, triggering the reduction of gold chloride to solid gold deposits (left). Observed deposition of gold on GUVs (right). Scale bar = 20 μm .³³⁴ **a**, ref. 366, copyright 2023, Frontiers Media and ref. 367, copyright 2020, Wiley-VCH; **b**, ref. 333, copyright 2022, Wiley-VCH; **c**, ref. 334, copyright 2022, Wiley-VCH.

Conclusion

The multi-functionality, programmability, and controllability of building materials for molecular robots engender molecular robots with superior potential to other types of microrobots. However, constructing molecular robots is not an easily achieved task, as it requires not only parts with high quality but also great synergy between each part. In this review, we have introduced several recent advances in the core parts of molecular robots. Looking to the future, what should be the direction of development for these core parts?

The lifetime of a GUV is reported to be up to several months,³³⁵ in contrast to the 10 to 20 years' average lifetime of industrial robots,³³⁶ reflecting a great hurdle in the successful implementation of molecular robots. Adding a second component to the lipid membranes, for example, polymers^{337,338} or surfactants,³³⁹ could make a giant leap in developing GUVs with higher stability. However, these impurities would also change the permeability of lipid membranes and impair the functionality of the sensors, computers, and actuators. A comprehensive list of such components with their advantages and drawbacks will have to be obtained using both experimental and *in silico*

approaches. In addition, while the water/oil emulsion technology-based strategy is the leading contender for GUV fabrication, adding polymers and surfactants may change the interfacial tension between water and oil, making GUVs fabrication difficult. Consequently, new GUVs fabrication methods must be continuously explored.

For nanopores, future improvements should be directed towards increasing their homogeneity and imparting selectivity. So far, we have shown that each of the building materials for nanopores provides unique benefits: proteins are highly compatible with lipid membranes, peptides are easy to synthesize, DNA offers high programmability, and synthetic materials offers strong operability. Accordingly, combining these materials has gained great interest as a unique strategy to achieve homogeneity and selectivity. Recent reports have demonstrated methods to tune the diameter of protein^{340,341} and peptide^{342,343} nanopores using DNA nanostructures/origami as a scaffold. Such DNA nanostructures could also impart selectivity to the protein/peptide nanopores, whilst the incorporation of synthetic materials is also worth exploring.

Building a molecular computer that can perform tasks in an intelligent way like a human being is the ultimate goal for

molecular robotics. The first step towards intelligence is the ability to learn. Neural networks are the backbone of deep learning algorithms, and a neural network capable of 144-bit pattern recognition with a two-layer implementation function has already been achieved using DNA computing.²⁰⁵ However, research into DNA computers with the capability to learn remains in the early phase due to the challenges in scalability (this is discussed in detail in a survey by Nagipogu *et al.*³⁴⁴). Consequently, new methods to address the scalability problems are highly desired. On the other hand, computers based on CFPS are still in their infancy. Whilst arithmetic operation in digital computers is based on a combination of logic gates, the use of protein logic gates is rarely reported. One difficult issue is the limited pool of building blocks due to the complicated interaction between proteins. Recently, Chen *et al.* reported a *de novo* design of three-input protein logic gates, giving careful consideration to the interaction between each building block.³⁴⁵ Rapid advances in *de novo* design will bring out the full potential of CFPS as computers for molecular robots, and the great diversity in the types of functional outputs of CFPS may lead to new breakthroughs in computational intelligence.

Despite the significant progress in actuating lipid membranes with DNA nanostructures/origamis and proteins/peptides, most of the reported methods rely on uncontrollable thermal fluctuation-dependent molecular processes, hence restricting their operability. Applying a controllable external field may help meet this challenge, as achievements in the actuation of DNA nanostructures/origamis with both electric³⁴⁶ and magnetic fields³⁴⁷ have progressed in recent years. The incorporation of electrochemically, magnetically, or acoustically sensitive chemical groups would allow development of increasingly superior actuators for molecular robots.

Improving the cooperation between sensors, computers, and actuators is the key to realizing the potential of molecular robots. Previous molecular robots mainly utilize open-loop control systems, meaning that the output generated by computers or actuators does not affect the input from sensors. Though the simplicity of such an open-loop control system offers many benefits, its poor accuracy limits the reliability of molecular robots. In conventional mechanical engineering, closed-loop control systems are known to improve accuracy, and we argue that the same rule-of-thumb should be valid for molecular robots. Only a limited number of molecular robots utilizing closed-loop systems have been reported. In one example, Peng *et al.* developed a molecular robot that could detect Sr^{2+} with a DNA computer while regulating the Sr^{2+} input from DNA nanopores¹⁴¹ (this is also discussed in the section on Applications of Molecular Robots). Usage of sensors, computers, and actuators that can precisely control the function of each other is expected to become mainstream in molecular robots' development.

Another challenge that needs to be addressed is how to maintain molecular robots' sustainability. The working

principles of current molecular robots follow the minimization of free energy, limiting the lifetime of their functionalities. Looking ahead toward a versatile molecular robot, the ability to sustain non-equilibrium conditions where energy and materials are continuously supplied is highly desirable. As we discussed earlier, the introduction of a self-fuelling system may solve the energy supply issue (readers are referred to the section on Applications of Molecular Robots for details). On the other hand, the requirement to supply new material can be addressed by utilizing membrane fusion. Zhao *et al.* have demonstrated that CFPS in GUVs can be triggered by the fusion of GUVs and large unilamellar vesicles encapsulating DNA plasmids.³⁴⁸ Enabling sustained out-of-equilibrium operation represents a critical step towards the deployment of molecular robots for practical application.

Looking at the success of currently reported applications of molecular robots, how can these applications be further expanded in combination with other technologies? Wearable sensors, which are electronic devices that can be comfortably worn, provide promising approaches to monitor physiological information in real-time.³⁴⁹ To provide sensing selectivity and specificity, biological systems such as living engineered bacteria³⁵⁰ and CFPS³⁵¹ have been incorporated into wearable sensors. Compared with previous systems, molecular robots can combine both the functionality of living organisms and the durability of CFPS, making them a strong candidate for new wearable sensors. Another likely trend will be using molecular robots in the atmosphere to perform tasks such as monitoring atmospheric pollution or delivery of specific substances. While this may seem to run contrary to the principle of molecular robots, due to the absence of aqueous media, we consider the soap bubbles as a potential solution to address this issue because they are stable in the atmosphere and can trap molecules in their wall. Inspired by some interesting studies that used soap bubbles to detect methamphetamine in aerosols³⁵² or to pollinate flowers,³⁵³ we expect that such soap bubble-based molecular robots incorporating sensors, computers, and actuators can be realized in the near future.

In conclusion, the molecular robot is the fruit of numerous technological progresses in biophysics, biochemistry, and systems engineering. With an increasing number of reports on new findings in these areas, we believe the development of molecular robots will be further accelerated, opening the door to a new era in robotics.

Author contributions

Z. P. and R. K. conceptualized the article. Z. P. prepared the initial draft of section: Introduction, Applications of Molecular Robots, and Conclusions. Y. E. prepared the draft of section Body of Molecular Robots. Z. P., S. I., M. Y., and S. F. prepared the draft of section: Sensors of Molecular Robots. T. M., S. T., G. F., and F. K. prepared the draft of section: Computers of Molecular Robots. L. D. M., Z. P. and K. I.



prepared the draft of section: Actuators of Molecular Robots. Z. P., H. S. and A. C. edited the manuscript with expert guidance, suggested edits, and supervision from R. K.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 C. O. Dietrich-Buchecker, J. P. Sauvage and J. P. Kintzinger, Une nouvelle famille de molécules : les metallo-catenanes, *Tetrahedron Lett.*, 1983, **24**, 5095–5098.
- 2 P. L. Anelli, N. Spencer and J. Fraser Stoddart, A Molecular Shuttle, *J. Am. Chem. Soc.*, 1991, **113**, 5131–5133.
- 3 N. Kourumura, R. W. J. Zijlstra, R. A. van Delden, N. Harada and B. L. Feringa, Light-driven monodirectional molecular rotor, *Nature*, 1999, **401**, 152–154.
- 4 T. Kudernac, *et al.*, Electrically driven directional motion of a four-wheeled molecule on a metal surface, *Nature*, 2011, **479**, 208–211.
- 5 M. Hagiya, A. Konagaya, S. Kobayashi, H. Saito and S. Murata, Molecular robots with sensors and intelligence, *Acc. Chem. Res.*, 2014, **47**, 1681–1690.
- 6 R. Kawano, Synthetic Ion Channels and DNA Logic Gates as Components of Molecular Robots, *ChemPhysChem*, 2018, **19**, 359–366.
- 7 K. Shoji and R. Kawano, Recent advances in liposome-based molecular robots, *Micromachines*, 2020, **11**, 788.
- 8 D. T. Chiu, *et al.* Chemical transformations in individual ultrasmall biomimetic containers, *Science*, 1999, **283**, 1892–1895.
- 9 T. M. Hsin and E. S. Yeung, Single-molecule reactions in liposomes, *Angew. Chem., Int. Ed.*, 2007, **46**, 8032–8035.
- 10 N. An, A. M. Fleming, E. G. Middleton and C. J. Burrows, Single-molecule investigation of G-quadruplex folds of the human telomere sequence in a protein nanocavity, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, 14325–14331.
- 11 J. R. Burns, A. Seifert, N. Fertig and S. Howorka, A biomimetic DNA-based channel for the ligand-controlled transport of charged molecular cargo across a biological membrane, *Nat. Nanotechnol.*, 2016, **11**, 152–156.
- 12 L. M. Adleman, Molecular computation of solutions to combinatorial problems, *Science*, 1994, **266**, 1021–1024.
- 13 S. Takiguchi and R. Kawano, Nanopore decoding for a Hamiltonian path problem, *Nanoscale*, 2021, **13**, 6192–6200.
- 14 Y. Shimizu, *et al.* Cell-free translation reconstituted with purified components, *Nat. Biotechnol.*, 2001, **19**, 751–755.
- 15 Y. Sato, Y. Hiratsuka, I. Kawamata, S. Murata and S. I. M. Nomura, Micrometer-sized molecular robot changes its shape in response to signal molecules, *Sci. Robot.*, 2017, **2**, eaal3735.
- 16 P. Drücker, *et al.* Membrane deformation and layer-by-layer peeling of giant vesicles induced by the pore-forming toxin pneumolysin, *Biomater. Sci.*, 2019, **7**, 3693–3705.
- 17 J. Steinkühler, *et al.* Controlled division of cell-sized vesicles by low densities of membrane-bound proteins, *Nat. Commun.*, 2020, **11**, 905.
- 18 K. Nishimura, *et al.* Identification of giant unilamellar vesicles with permeability to small charged molecules, *RSC Adv.*, 2014, **4**, 35224–35232.
- 19 B. C. Buddingh' and J. C. M. Van Hest, Artificial Cells: Synthetic Compartments with Life-like Functionality and Adaptivity, *Acc. Chem. Res.*, 2017, **50**, 769–777.
- 20 C. Guindani, L. C. da Silva, S. Cao, T. Ivanov and K. Landfester, Synthetic Cells: From Simple Bio-Inspired Modules to Sophisticated Integrated Systems, *Angew. Chem., Int. Ed.*, 2022, **61**, e202110855.
- 21 Y. Lu, G. Allegri and J. Huskens, Vesicle-based artificial cells: materials, construction methods and applications, *Mater. Horiz.*, 2022, **9**, 892–907.
- 22 I. N. Westensee, *et al.* Mitochondria encapsulation in hydrogel-based artificial cells as ATP producing subunits, *Small*, 2021, **17**, e2007959.
- 23 C. Martino, *et al.* Protein expression, aggregation, and triggered release from polymersomes as artificial cell-like structures, *Angew. Chem., Int. Ed.*, 2012, **51**, 6416–6420.
- 24 M. Walczak, *et al.* A Synthetic Signalling Network Imitating the Action of Immune Cells in Response to Bacterial Metabolism, *Adv. Mater.*, 2023, **2301562**, e2301562.
- 25 T. Y. Dora Tang, D. Van Swaay, A. DeMello, J. L. Ross Anderson and S. Mann, *In vitro* gene expression within membrane-free coacervate protocells, *Chem. Commun.*, 2015, **51**, 11429–11432.
- 26 T. Y. Dora Tang, *et al.* Fatty acid membrane assembly on coacervate microdroplets as a step towards a hybrid protocell model, *Nat. Chem.*, 2014, **6**, 527–533.
- 27 Y. Sato and M. Takinoue, Capsule-like DNA Hydrogels with Patterns Formed by Lateral Phase Separation of DNA Nanostructures, *JACS Au*, 2022, **2**, 159–168.
- 28 T. Lu, S. Javed, C. Bonfio and E. Spruijt, Interfacing Coacervates with Membranes: From Artificial Organelles and Hybrid Protocells to Intracellular Delivery, *Small Methods*, 2023, 2300294.
- 29 M. A. Boyd and N. P. Kamat, Designing Artificial Cells towards a New Generation of Biosensors, *Trends Biotechnol.*, 2021, **39**, 927–939.

30 Z. A. Manzer, *et al.* Cell-Free Synthesis of a Transmembrane Mechanosensitive Channel Protein into a Hybrid-Supported Lipid Bilayer, *ACS Appl. Bio Mater.*, 2021, **4**, 3101–3112.

31 M. J. Langton, Engineering of stimuli-responsive lipid-bilayer membranes using supramolecular systems, *Nat. Rev. Chem.*, 2021, **5**, 46–61.

32 I. Gispert, *et al.* Stimuli-responsive vesicles as distributed artificial organelles for bacterial activation, *Proc. Natl. Acad. Sci. U. S. A.*, 2022, **119**, e2206563119.

33 A. A. Gu, P. Tran, A. Prindle, N. P. Kamat and J. Steinkühler, Remodeling of lipid-foam prototissues by network-wide tension fluctuations induced by active particles, *bioRxiv*, 2023, preprint, DOI: [10.1101/2023.06.15.545042](https://doi.org/10.1101/2023.06.15.545042).

34 R. Rubio-Sánchez, *et al.* Thermally Driven Membrane Phase Transitions Enable Content Reshuffling in Primitive Cells, *J. Am. Chem. Soc.*, 2021, **143**, 16589–16598.

35 P. Walde, K. Cosentino, H. Engel and P. Stano, Giant Vesicles: Preparations and Applications, *ChemBioChem*, 2010, **11**, 848–865.

36 E. Altamura, P. Albanese, F. Mavelli and P. Stano, The Rise of the Nested Multicompartment Model in Synthetic Cell Research, *Front. Mol. Biosci.*, 2021, **8**, 750576.

37 G. Zubaite, J. W. Hindley, O. Ces and Y. Elani, Dynamic Reconfiguration of Subcompartment Architectures in Artificial Cells, *ACS Nano*, 2022, **16**, 9389–9400.

38 Y. Elani, Interfacing Living and Synthetic Cells as an Emerging Frontier in Synthetic Biology, *Angewandte Chemie*, 2021, **133**, 5662–5671.

39 E. Altamura, *et al.* Chromatophores efficiently promote light-driven ATP synthesis and DNA transcription inside hybrid multicompartment artificial cells, *Proc. Natl. Acad. Sci. U. S. A.*, 2021, **118**, 6416–6420.

40 K. Nishimura, S. Tsuru, H. Suzuki and T. Yomo, Stochasticity in gene expression in a cell-sized compartment, *ACS Synth. Biol.*, 2015, **4**, 566–576.

41 D. Van Swaay and A. Demello, Microfluidic methods for forming liposomes, *Lab Chip*, 2013, **13**, 752–767.

42 T. Robinson, Microfluidic Handling and Analysis of Giant Vesicles for Use as Artificial Cells: A Review, *Adv. Biosyst.*, 2019, **3**, 1800318.

43 Y. Ai, R. Xie, J. Xiong and Q. Liang, Microfluidics for Biosynthesizing: from Droplets and Vesicles to Artificial Cells, *Small*, 2020, **16**, e1903940.

44 T. Ip, Q. Li, N. Brooks and Y. Elani, Manufacture of Multilayered Artificial Cell Membranes through Sequential Bilayer Deposition on Emulsion Templates, *ChemBioChem*, 2021, **22**, 2275–2281.

45 S. Matosevic and B. M. Paegel, Layer-by-layer cell membrane assembly, *Nat. Chem.*, 2013, **5**, 958–963.

46 R. Dimova, Giant Vesicles and Their Use in Assays for Assessing Membrane Phase State, Curvature, Mechanics, and Electrical Properties, *Annu. Rev. Biophys.*, 2019, **48**, 93–119.

47 S. Pautot, B. J. Frisken and D. A. Weitz, Production of unilamellar vesicles using an inverted emulsion, *Langmuir*, 2003, **19**, 2870–2879.

48 R. Tivony, M. Fletcher, K. Al Nahas and U. F. Keyser, A Microfluidic Platform for Sequential Assembly and Separation of Synthetic Cell Models, *ACS Synth. Biol.*, 2021, **10**, 3105–3116.

49 S. Deshpande and C. Dekker, On-chip microfluidic production of cell-sized liposomes, *Nat. Protoc.*, 2018, **13**, 856–874.

50 N.-N. Deng, M. Yelleswarapu and W. T. S. Huck, Monodisperse Uni- and Multicompartment Liposomes, *J. Am. Chem. Soc.*, 2016, **138**, 7584–7591.

51 M. S. Friddin, Y. Elani, T. Trantidou and O. Ces, New Directions for Artificial Cells Using Prototyped Biosystems, *Anal. Chem.*, 2019, **91**, 4921–4928.

52 S. Matosevic and B. M. Paegel, Stepwise synthesis of giant unilamellar vesicles on a microfluidic assembly line, *J. Am. Chem. Soc.*, 2011, **133**, 2798–2800.

53 K. Karamdad, R. V. Law, J. M. Seddon, N. J. Brooks and O. Ces, Preparation and mechanical characterisation of giant unilamellar vesicles by a microfluidic method, *Lab Chip*, 2015, **15**, 557–562.

54 M. Abkarian, E. Loiseau and G. Massiera, Continuous droplet interface crossing encapsulation (cDICE) for high throughput monodisperse vesicle design, *Soft Matter*, 2011, **7**, 4610–4614.

55 H. C. Shum, *et al.* Dewetting-induced membrane formation by adhesion of amphiphile-laden interfaces, *J. Am. Chem. Soc.*, 2011, **133**, 4420–4426.

56 M. Fletcher, *et al.* DNA-Based Optical Quantification of Ion Transport across Giant Vesicles, *ACS Nano*, 2022, **16**, 17128–17138.

57 N. N. Deng, M. Yelleswarapu, L. Zheng and W. T. S. Huck, Microfluidic assembly of monodisperse vesosomes as artificial cell models, *J. Am. Chem. Soc.*, 2017, **139**, 587–590.

58 A. Vivek, G. Bolognesi and Y. Elani, Fusing artificial cell compartments and lipid domains using optical traps: A tool to modulate membrane composition and phase behaviour, *Micromachines*, 2020, **11**, 388.

59 M. S. Friddin, G. Bolognesi, A. Salehi-Reyhani, O. Ces and Y. Elani, Direct manipulation of liquid ordered lipid membrane domains using optical traps, *Commun. Chem.*, 2019, **2**, 6.

60 H. Watanabe, *et al.* Analysis of Pore Formation and Protein Translocation Using Large Biological Nanopores, *Anal. Chem.*, 2017, **89**, 11269–11277.

61 K. A. Brogden, Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria?, *Nat. Rev. Microbiol.*, 2005, **3**, 238–250.

62 J. Geng, *et al.* Stochastic transport through carbon nanotubes in lipid bilayers and live cell membranes, *Nature*, 2014, **514**, 612–615.

63 T. Tosaka and K. Kamiya, Function Investigations and Applications of Membrane Proteins on Artificial Lipid Membranes, *Int. J. Mol. Sci.*, 2023, **24**, 7231.

64 A. Tada, N. Takeuchi, K. Shoji and R. Kawano, Nanopore Filter: A Method for Counting and Extracting Single DNA Molecules Using a Biological Nanopore, *Anal. Chem.*, 2023, **95**, 9805–9812.



65 J. J. Kasianowicz, E. Brandin, D. Branton and D. W. Deamer, Characterization of individual polynucleotide molecules using a membrane channel, *Proc. Natl. Acad. Sci.*, 1996, **93**, 13770–13773.

66 I. M. Derrington, *et al.* Nanopore DNA sequencing with MspA, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 16060–16065.

67 M. Jain, H. E. Olsen, B. Paten and M. Akeson, The Oxford Nanopore MinION: delivery of nanopore sequencing to the genomics community, *Genome Biol.*, 2016, **17**, 239.

68 S. Zhang, *et al.* Bottom-up fabrication of a proteasome-nanopore that unravels and processes single proteins, *Nat. Chem.*, 2021, **13**, 1192–1199.

69 G. Huang, *et al.* PlyAB Nanopores Detect Single Amino Acid Differences in Folded Haemoglobin from Blood, *Angew. Chem.*, 2022, **61**, e202206227.

70 M. Miyagi, S. Takiguchi, K. Hakamada, M. Yohda and R. Kawano, Single polypeptide detection using a translocon EXP2 nanopore, *Proteomics*, 2022, **22**, e2100070.

71 D. Vikraman, R. Krishnan, R. Satheesan, A. D. Das and K. R. Mahendran, Electrostatic Filtering of Polypeptides Through Membrane Protein Pores, *Chem. – Asian J.*, 2022, **17**, e202200891.

72 K. B. Jeong, *et al.* Single-molecule fingerprinting of protein-drug interaction using a funneled biological nanopore, *Nat. Commun.*, 2023, **14**, 1461.

73 N. Varongchayakul, J. Song, A. Meller and M. W. Grinstaff, Single-molecule protein sensing in a nanopore: a tutorial, *Chem. Soc. Rev.*, 2018, **47**, 8512–8524.

74 L. Restrepo-Pérez, C. Joo and C. Dekker, Paving the way to single-molecule protein sequencing, *Nat. Nanotechnol.*, 2018, **13**, 786–796.

75 K. Motone, N. Cardozo and J. Nivala, Herding cats: Label-based approaches in protein translocation through nanopore sensors for single-molecule protein sequence analysis, *iScience*, 2021, **24**, 103032.

76 H. Ouldali, *et al.* Electrical recognition of the twenty proteinogenic amino acids using an aerolysin nanopore, *Nat. Biotechnol.*, 2020, **38**, 176–181.

77 H. Brinkerhoff, A. S. W. Kang, J. Liu, A. Aksimentiev and C. Dekker, Multiple rereads of single proteins at single-amino acid resolution using nanopores, *Science*, 2021, **374**, 1509–1513.

78 S. Yan, *et al.* Molecule Ratcheting Motion of Peptides in a *Mycobacterium smegmatis* Porin A (MspA) Nanopore, *Nano Lett.*, 2021, **21**, 6703–6710.

79 F. L. R. Lucas, R. C. A. Versloot, L. Yakovlieva, M. T. C. Walvoort and G. Maglia, Protein identification by nanopore peptide profiling, *Nat. Commun.*, 2021, **12**, 5795.

80 K. Takagi, T. Ohgita, T. Yamamoto, Y. Shinohara and K. Kogure, Transmission of external environmental pH information to the inside of liposomes *via* pore-forming proteins embedded within the liposomal membrane, *Chem. Pharm. Bull.*, 2016, **64**, 432–438.

81 A. Koçer, M. Walko, W. Meijberg and B. L. Feringa, Chemistry: A light-actuated nanovalve derived from a channel protein, *Science*, 2005, **309**, 755–758.

82 A. Koçer, A remote controlled valve in liposomes for triggered liposomal release, *J. Liposome Res.*, 2007, **17**, 219–225.

83 S. Deshpande, *et al.* Spatiotemporal control of coacervate formation within liposomes, *Nat. Commun.*, 2019, **10**, 1800.

84 V. Noireaux and A. Libchaber, A vesicle bioreactor as a step toward an artificial cell assembly, *Proc. Natl. Acad. Sci. U. S. A.*, 2004, **101**, 17669–17674.

85 G. Tsuji, T. Sunami, M. Oki and N. Ichihashi, Exchange of Proteins in Liposomes through Streptolysin O Pores, *ChemBioChem*, 2021, **22**, 1966–1973.

86 S. Straathof, *et al.* Protein Sizing with 15 nm Conical Biological Nanopore YaxAB, *ACS Nano*, 2023, **17**, 13685–13699.

87 X. Zhang, *et al.* Specific Detection of Proteins by a Nanobody-Functionalized Nanopore Sensor, *ACS Nano*, 2023, **17**, 9167–9177.

88 D. Rotem, L. Jayasinghe, M. Salichou and H. Bayley, Protein detection by nanopores equipped with aptamers, *J. Am. Chem. Soc.*, 2012, **134**, 2781–2787.

89 M. Soskine, *et al.* An engineered ClyA nanopore detects folded target proteins by selective external association and pore entry, *Nano Lett.*, 2012, **12**, 4895–4900.

90 M. Fahie, C. Chisholm and M. Chen, Resolved single-molecule detection of individual species within a mixture of anti-biotin antibodies using an engineered monomeric nanopore, *ACS Nano*, 2015, **9**, 1089–1098.

91 H. Xie, O. Braha, L. Q. Gu, S. Cheley and H. Bayley, Single-molecule observation of the catalytic subunit of cAMP-dependent protein kinase binding to an inhibitor peptide, *Chem. Biol.*, 2005, **12**, 109–120.

92 A. K. Thakur and L. Movileanu, Real-time measurement of protein–protein interactions at single-molecule resolution using a biological nanopore, *Nat. Biotechnol.*, 2019, **37**, 96–104.

93 M. L. Ghimire, *et al.* Detection and Characterization of Small Cysteine-Containing Peptides with Cluster-Modified Nanopore Sensing, *ACS Nano*, 2022, **16**, 17229–17241.

94 M. Ahmad, *et al.* A generalizable nanopore sensor for highly specific protein detection at single-molecule precision, *Nat. Commun.*, 2023, **14**, 1374.

95 N. H. Joh, *et al.* De novo design of a transmembrane zn^{2+} -transporting four-helix bundle, *Science*, 2014, **346**, 1520–1524.

96 P. S. Huang, S. E. Boyken and D. Baker, The coming of age of de novo protein design, *Nature*, 2016, **537**, 320–327.

97 J. Dou, *et al.* De novo design of a fluorescence-activating β -barrel, *Nature*, 2018, **561**, 485–491.

98 A. H. W. Yeh, *et al.* De novo design of luciferases using deep learning, *Nature*, 2023, **614**, 774–780.

99 A. A. Vorobieva, *et al.* De novo design of transmembrane β barrels, *Science*, 2021, **371**, eabc8182.

100 N. Koga, *et al.* Principles for designing ideal protein structures, *Nature*, 2012, **491**, 222–227.



101 J. Dauparas, *et al.* Robust deep learning-based protein sequence design using ProteinMPNN, *Science*, 2022, **378**, 49–56.

102 J. L. Watson, *et al.* De novo design of protein structure and function with RFdiffusion, *Nature*, 2023, **620**, 1089–1100.

103 G. A. Woolley and B. A. Wallace, Membrane Biology Topical Review Model Ion Channels: Gramicidin and Alamethicin, *J. Membr. Biol.*, 1992, **129**, 109–136.

104 D. Peter Tieleman, B. Hess and M. S. P. Sansom, Analysis and Evaluation of Channel Models: Simulations of Alamethicin, *Biophys. J.*, 2002, **83**, 2393–2407.

105 H. Watanabe and R. Kawano, Channel current analysis for pore-forming properties of an antimicrobial peptide, magainin 1, using the droplet contact method, *Anal. Sci.*, 2016, **32**, 57–60.

106 N. Saigo, K. Izumi and R. Kawano, Electrophysiological Analysis of Antimicrobial Peptides in Diverse Species, *ACS Omega*, 2019, **4**, 13124–13130.

107 A. Schmidtchen, *et al.* Boosting antimicrobial peptides by hydrophobic oligopeptide end tags, *J. Biol. Chem.*, 2009, **284**, 17584–17594.

108 N. B. Last and A. D. Miranker, Common mechanism unites membrane poration by amyloid and antimicrobial peptides, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 6382–6387.

109 K. D. Saint Jean, *et al.* Effects of Hydrophobic Amino Acid Substitutions on Antimicrobial Peptide Behavior, *Probiotics Antimicrob. Proteins*, 2018, **10**, 408–419.

110 É. Forde, G. Shafiy, D. Fitzgerald-Hughes, A. A. Strömstedt and M. Devocelle, Action of antimicrobial peptides and their prodrugs on model and biological membranes, *J. Pept. Sci.*, 2018, **24**, e3086.

111 F. Ben Trad, *et al.* Electrochemiluminescence Imaging of Liposome Permeabilization by an Antimicrobial Peptide: Melittin, *Chem. Biomed. Imaging*, 2023, **2023**, 58–65.

112 S. Rex, Pore formation induced by the peptide melittin in different lipid vesicle membranes, *Biophys. Chem.*, 1996, **58**, 75–85.

113 A. S. Ladokhin, M. E. Selsted and S. H. White, Sizing Membrane Pores in Lipid Vesicles by Leakage of Co-Encapsulated Markers: Pore Formation by Melittin, *Biophys. J.*, 1997, **72**, 1762–1766.

114 J. M. Rausch, J. R. Marks, R. Rathinakumar and W. C. Wimley, β -Sheet Pore-Forming Peptides Selected from a Rational Combinatorial Library: Mechanism of Pore Formation in Lipid Vesicles and Activity in Biological Membranes, *Biochemistry*, 2007, **46**, 12124.

115 K. R. Mahendran, *et al.* A monodisperse transmembrane α -helical peptide barrel, *Nat. Chem.*, 2016, **9**(5), 411–419.

116 S. Krishnan, *et al.* Autonomously Assembled Synthetic Transmembrane Peptide Pore, *J. Am. Chem. Soc.*, 2019, **141**, 2949–2959.

117 S. K. R, *et al.* Designed alpha-helical barrels for charge-selective peptide translocation, *Chem. Sci.*, 2021, **12**, 639–649.

118 S. K. R, *et al.* Assembly of transmembrane pores from mirror-image peptides, *Nat. Commun.*, 2022, **13**, 5377.

119 C. Xu, *et al.* Computational design of transmembrane pores, *Nature*, 2020, **585**, 129–134.

120 A. J. Scott, *et al.* Constructing ion channels from water-soluble α -helical barrels, *Nat. Chem.*, 2021, **13**, 643–650.

121 W. M. Dawson, *et al.* Coiled coils 9-to-5: rational de novo design of α -helical barrels with tunable oligomeric states, *Chem. Sci.*, 2021, **12**, 6923–6928.

122 K. Shimizu, *et al.* De novo design of a nanopore for single-molecule detection that incorporates a β -hairpin peptide, *Nat. Nanotechnol.*, 2022, **17**, 67–75.

123 S. Fujita, I. Kawamura and R. Kawano, Cell-Free Expression of De Novo Designed Peptides That Form β -Barrel Nanopores Article, *ACS Nano*, 2023, **17**, 31.

124 M. D. Peraro and F. G. Van Der Goot, Pore-forming toxins: Ancient, but never really out of fashion, *Nat. Rev. Microbiol.*, 2016, **14**, 77–92.

125 A. Niitsu, J. W. Heal, K. Fauland, A. R. Thomson and D. N. Woolfson, Membrane-spanning α -helical barrels as tractable protein-design targets, *Philos. Trans. R. Soc., B*, 2017, **372**, 20160213.

126 P. W. K. Rothmund, Folding DNA to create nanoscale shapes and patterns, *Nature*, 2006, **440**, 297–302.

127 S. M. Douglas, *et al.* Self-assembly of DNA into nanoscale three-dimensional shapes, *Nature*, 2009, **459**, 414–418.

128 J. Chen and N. C. Seeman, Synthesis from DNA of a molecule with the connectivity of a cube, *Nature*, 1991, **350**, 631–633.

129 F. Zhang, J. Nangreave, Y. Liu and H. Yan, Structural DNA nanotechnology: State of the art and future perspective, *J. Am. Chem. Soc.*, 2014, **136**, 11198–11211.

130 M. Langecker, *et al.* Synthetic lipid membrane channels formed by designed DNA nanostructures, *Science*, 2012, **338**, 932–936.

131 J. R. Burns, *et al.* Lipid-bilayer-spanning DNA nanopores with a bifunctional porphyrin anchor, *Angew. Chem., Int. Ed.*, 2013, **52**, 12069–12072.

132 S. Krishnan, *et al.* Molecular transport through large-diameter DNA nanopores, *Nat. Commun.*, 2016, **7**, 12787.

133 R. P. Thomsen, *et al.* A large size-selective DNA nanopore with sensing applications, *Nat. Commun.*, 2019, **10**, 5655.

134 S. Iwabuchi, I. Kawamata, S. Murata and S. I. M. Nomura, A large, square-shaped, DNA origami nanopore with sealing function on a giant vesicle membrane, *Chem. Commun.*, 2021, **57**, 2990–2993.

135 A. Fragasso, *et al.* Reconstitution of Ultrawide DNA Origami Pores in Liposomes for Transmembrane Transport of Macromolecules, *ACS Nano*, 2021, **15**, 12768–12779.

136 K. Jahnke, *et al.* DNA Origami Signaling Units Transduce Chemical and Mechanical Signals in Synthetic Cells, *Adv. Funct. Mater.*, 2023, 2301176.

137 S. Dey, *et al.* A reversibly gated protein-transporting membrane channel made of DNA, *Nat. Commun.*, 2022, **13**, 232–240.

138 Y. Xing, A. Dorey, L. Jayasinghe and S. Howorka, Highly shape- and size-tunable membrane nanopores made with DNA, *Nat. Nanotechnol.*, 2022, **17**, 708–713.



139 T. Diederichs, *et al.* Principles of Small-Molecule Transport through Synthetic Nanopores, *ACS Nano*, 2021, **15**, 16194–16206.

140 S. Ikarashi, *et al.* DNA Nanopore-Tethered Gold Needle Electrodes for Channel Current Recording, *ACS Nano*, 2023, **17**, 10598–10607.

141 R. Peng, *et al.* DNA-based artificial molecular signaling system that mimics basic elements of reception and response, *Nat. Commun.*, 2020, **11**, 978.

142 L. Chen, S. Liang, Y. Chen, M. Wu and Y. Zhang, Destructing the Plasma Membrane with Activatable Vesicular DNA Nanopores, *ACS Appl. Mater. Interfaces*, 2020, **12**, 96–105.

143 Q. Yang, *et al.* A Cascade Signaling Network between Artificial Cells Switching Activity of Synthetic Transmembrane Channels, *J. Am. Chem. Soc.*, 2021, **143**, 232–240.

144 P. Chidchob, *et al.* Spatial Presentation of Cholesterol Units on a DNA Cube as a Determinant of Membrane Protein-Mimicking Functions, *J. Am. Chem. Soc.*, 2019, **141**, 1100–1108.

145 C. Lanphere, P. M. Arnott, S. F. Jones, K. Korlova and S. Howorka, A Biomimetic DNA-Based Membrane Gate for Protein-Controlled Transport of Cytotoxic Drugs, *Angew. Chem.*, 2021, **60**, 1903–1908.

146 P. M. Arnott and S. Howorka, A Temperature-Gated Nanovalve Self-Assembled from DNA to Control Molecular Transport across Membranes, *ACS Nano*, 2019, **13**, 3334–3340.

147 D. Offenbartl-Stiegert, A. Rottensteiner, A. Dorey and S. Howorka, A Light-Triggered Synthetic Nanopore for Controlling Molecular Transport Across Biological Membranes, *Angew. Chem.*, 2022, **61**, e202210886.

148 O. Mendoza, *et al.* A tensegrity driven DNA nanopore, *Nanoscale*, 2017, **9**, 9762–9769.

149 H. Chen, *et al.* Controlled dimerization of artificial membrane receptors for transmembrane signal transduction, *Chem. Sci.*, 2021, **12**, 8224–8230.

150 G. Liu, *et al.* DNA-Based Artificial Signaling System Mimicking the Dimerization of Receptors for Signal Transduction and Amplification, *Anal. Chem.*, 2021, **93**, 13807–13814.

151 L. E. Bickerton and M. J. Langton, Controlling transmembrane ion transport *via* photo-regulated carrier mobility, *Chem. Sci.*, 2022, **13**, 9531–9536.

152 F. De Riccardis, I. Izzo, D. Montesarchio and P. Tecilla, Ion transport through lipid bilayers by synthetic ionophores: Modulation of activity and selectivity, *Acc. Chem. Res.*, 2013, **46**, 2781–2790.

153 N. Sakai and S. Matile, Synthetic ion channels, *Langmuir*, 2013, **29**, 9031–9040.

154 A. Mondal, M. Ahmad, D. Mondal and P. Talukdar, Progress and prospects toward supramolecular bioactive ion transporters, *Chem. Commun.*, 2023, **59**, 1917–1938.

155 Y. Zhou, J. H. Morais-Cabral, A. Kaufman and R. Mackinnon, Chemistry of ion coordination and hydration revealed by a K⁺ channel-Fab complex at 2.0 Å resolution, *Nature*, 2001, **414**, 43–48.

156 Y. Tanaka, Y. Kobuke and M. Sokabe, A Non-Peptidic Ion Channel with K⁺ Selectivity, *Angew. Chem., Int. Ed. Engl.*, 1995, **34**, 693–694.

157 A. Nakano, Q. Xie, J. V. Mallen, L. Echegoyen and G. W. Gokel, Synthesis of a Membrane-Insertable, Sodium Cation Conducting Channel: Kinetic Analysis by Dynamic ²³Na NMR, *J. Am. Chem. Soc.*, 1990, **112**, 1287–1289.

158 V. E. Carmichael, *et al.* Biomimetic Ion Transport: A Functional Model of a Unimolecular Ion Channel, *J. Am. Chem. Soc.*, 1989, **111**, 767–769.

159 D. Qiao, *et al.* Synthetic Macrocycle Nanopore for Potassium-Selective Transmembrane Transport, *J. Am. Chem. Soc.*, 2021, **143**, 15975–15983.

160 W. L. Huang, X. D. Wang, Y. F. Ao, Q. Q. Wang and D. X. Wang, An Artificial Single Molecular Channel Showing High Chloride Transport Selectivity and pH-Responsive Conductance, *Angew. Chem., Int. Ed.*, 2023, **62**, e202302198.

161 A. Roy, *et al.* Polyhydrazide-Based Organic Nanotubes as Efficient and Selective Artificial Iodide Channels, *Angew. Chem., Int. Ed.*, 2020, **59**, 4806–4813.

162 J. Shen, *et al.* Fluorofoldamer-Based Salt- and Proton-Rejecting Artificial Water Channels for Ultrafast Water Transport, *Nano Lett.*, 2022, **22**, 4831–4838.

163 L. Zhang, C. Zhang, X. Dong and Z. Dong, Highly Selective Transmembrane Transport of Exogenous Lithium Ions through Rationally Designed Supramolecular Channels, *Angew. Chem.*, 2023, **62**, e202214194.

164 J. Shen, *et al.* Sulfur-Containing Foldamer-Based Artificial Lithium Channels, *Angew. Chem.*, 2023, **78712**, e202305623.

165 S. Qi, *et al.* Unimolecular Transmembrane Na⁺ Channels Constructed by Pore-Forming Helical Polymers with a 2.3 Å Aperture, *CCS Chem.*, 2022, **4**, 1850–1857.

166 Y. Li, *et al.* Metal-organic macrocycles with tunable pore microenvironments for selective anion transmembrane transport, *Mater. Chem. Front.*, 2022, **6**, 1010–1020.

167 R. Sharma, A. Vijay, S. Chattopadhyay, A. Mukherjee and P. Talukdar, Self-assembled anion channel formation by bis(1,3-propanediol)-linked meta-dipropynylbenzene-based small molecules, *Chem. Commun.*, 2023, **59**, 3602–3605.

168 C. Bao, M. Ma, F. Meng, Q. Lin and L. Zhu, Efficient synthetic supramolecular channels and their light-deactivated ion transport in bilayer lipid membranes, *New J. Chem.*, 2015, **39**, 6297–6302.

169 T. Liu, *et al.* Light-controlled ion channels formed by amphiphilic small molecules regulate ion conduction *via* cis-trans photoisomerization, *Chem. Commun.*, 2013, **49**, 10311–10313.

170 W. Z. Wang, *et al.* Light-Driven Molecular Motors Boost the Selective Transport of Alkali Metal Ions through Phospholipid Bilayers, *J. Am. Chem. Soc.*, 2021, **143**, 15653–15660.

171 T. Muraoka, *et al.* Mechano-Sensitive Synthetic Ion Channels, *J. Am. Chem. Soc.*, 2017, **139**, 18016–18023.



172 K. Sato, *et al.* Supramolecular Mechanosensitive Potassium Channel Formed by Fluorinated Amphiphilic Cyclophane, *J. Am. Chem. Soc.*, 2022, **144**, 11802–11809.

173 R. Sasaki, K. Sato, K. V. Tabata, H. Noji and K. Kinbara, Synthetic Ion Channel Formed by Multiblock Amphiphile with Anisotropic Dual-Stimuli-Responsiveness, *J. Am. Chem. Soc.*, 2021, **143**, 1348–1355.

174 S. P. Zheng, J. J. Jiang, A. van der Lee and M. Barboiu, A Voltage-Responsive Synthetic Cl-Channel Regulated by pH, *Angew. Chem., Int. Ed.*, 2020, **59**, 18920–18926.

175 T. Muraoka, *et al.* A synthetic ion channel with anisotropic ligand response, *Nat. Commun.*, 2020, **11**, 2924.

176 K. Komaki, *et al.* Cu(II)-Triggered Ion Channel Properties of a 2,2'-Bipyridine-Modified Amphotericin B, *ACS Appl. BioMater.*, 2023, **6**, 828–835.

177 X. Wei, *et al.* Persistent Organic Nanopores Amenable to Structural and Functional Tuning, *J. Am. Chem. Soc.*, 2016, **138**, 2749–2754.

178 C. Lang, *et al.* Biomimetic Transmembrane Channels with High Stability and Transporting Efficiency from Helically Folded Macromolecules, *Angew. Chem., Int. Ed.*, 2016, **55**, 9723–9727.

179 R. H. Tunuguntla, *et al.* Enhanced water permeability and tunable ion selectivity in subnanometer carbon nanotube porins, *Science*, 2017, **357**, 792–796.

180 W. Peng, *et al.* High-resolution discrimination of homologous and isomeric proteinogenic amino acids in nanopore sensors with ultrashort single-walled carbon nanotubes, *Nat. Commun.*, 2023, **14**, 2662.

181 L. Liu, C. Yang, K. Zhao, J. Li and H. C. Wu, Ultrashort single-walled carbon nanotubes in a lipid bilayer as a new nanopore sensor, *Nat. Commun.*, 2013, **4**, 2989.

182 A. Marcotte, T. Mouterde, A. Niguès, A. Siria and L. Bocquet, Mechanically activated ionic transport across single-digit carbon nanotubes, *Nat. Mater.*, 2020, **19**, 1057–1061.

183 K. Yang, H. A. Kotak and C. J. E. Haynes, Metal-organic ion transport systems, *Coord. Chem. Rev.*, 2022, **470**, 214705.

184 S. Lee, H. Jeong, D. Nam, M. S. Lah and W. Choe, The rise of metal-organic polyhedra, *Chem. Soc. Rev.*, 2021, **50**, 528–555.

185 M. Jung, *et al.* Synthetic Ion Channel Based on Metal-Organic Polyhedra, *Angew. Chem., Int. Ed.*, 2008, **47**, 5755–5757.

186 R. Kawano, *et al.* Metal-Organic Cuboctahedra for Synthetic Ion Channels with Multiple Conductance States, *Chem.*, 2017, **2**, 393–403.

187 Y. Li, *et al.* Artificial Biomolecular Channels: Enantioselective Transmembrane Transport of Amino Acids Mediated by Homochiral Zirconium Metal-Organic Cages, *J. Am. Chem. Soc.*, 2021, **143**, 20939–20951.

188 T. Nakakuki, K. Murayama and H. Asanuma, DNA Concentration Regulator That can be Driven for a Long Time, *New Gener. Comput.*, 2022, **40**, 681–702.

189 T. Y. Wei and W. C. Ruder, Engineering control circuits for molecular robots using synthetic biology, *APL Mater.*, 2020, **8**, 101104.

190 R. J. Lipton, DNA solution of hard computational problems, *Science*, 1995, **268**, 542–545.

191 Q. Ouyang, P. D. Kaplan, S. Liu and A. Libchaber, DNA solution of the maximal clique problem, *Science*, 1997, **278**, 446–449.

192 Y. Benenson, *et al.* Programmable and autonomous computing machine made of biomolecules, *Nature*, 2001, **414**, 430–434.

193 Y. Benenson, R. Adart, T. Paz-Elizur, Z. Livneh and E. Shapiro, DNA molecule provides a computing machine with both data and fuel, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, **100**, 2191–2196.

194 M. N. Stojanovic, T. E. Mitchell and D. Stefanovic, Deoxyribozyme-based logic gates, *J. Am. Chem. Soc.*, 2002, **124**, 3555–3561.

195 R. Penchovsky and R. R. Breaker, Computational design and experimental validation of oligonucleotide-sensing allosteric ribozymes, *Nat. Biotechnol.*, 2005, **23**, 1424–1433.

196 S. Bi, Y. Yan, S. Hao and S. Zhang, Colorimetric logic gates based on supramolecular DNAzyme structures, *Angew. Chem., Int. Ed.*, 2010, **49**, 4438–4442.

197 B. Yurke, A. J. Turberfield, A. P. Mills, F. C. Simmel and J. L. Neumann, A DNA-fuelled molecular machine made of DNA, *Nature*, 2000, **406**, 605–608.

198 G. Seelig, D. Soloveichik, D. Y. Zhang and E. Winfree, Enzyme-free nucleic acid logic circuits, *Science*, 2006, **314**, 1585–1588.

199 L. Qian and E. Winfree, Scaling up digital circuit computation with DNA strand displacement cascades, *Science*, 2011, **332**, 1196–1201.

200 J. Wang, H. Raito, N. Shimada and A. Maruyama, A Cationic Copolymer Enhances Responsiveness and Robustness of DNA Circuits, *Small*, 2023, 2304091.

201 A. P. Lapteva, N. Sarraf and L. Qian, DNA Strand-Displacement Temporal Logic Circuits, *J. Am. Chem. Soc.*, 2022, **144**, 12443–12449.

202 M. Qi, *et al.* Reconfigurable DNA triplex structure for pH responsive logic gates, *RSC Adv.*, 2023, **13**, 9864–9870.

203 L. Qian, E. Winfree and J. Bruck, Neural network computation with DNA strand displacement cascades, *Nature*, 2011, **475**, 368–372.

204 K. M. Cherry and L. Qian, Scaling up molecular pattern recognition with DNA-based winner-take-all neural networks, *Nature*, 2018, **559**, 370–388.

205 X. Xiong, *et al.* Molecular convolutional neural networks with DNA regulatory circuits, *Nat. Mach. Intell.*, 2022, **4**, 625–635.

206 X. Liu and K. K. Parhi, Reservoir Computing Using DNA Oscillators, *ACS Synth. Biol.*, 2022, **11**, 780–787.

207 S. Okumura, *et al.* Nonlinear decision-making with enzymatic neural networks, *Nature*, 2022, **610**(7932), 496–501.

208 P. Zhou, *et al.* Dual-AND Logic Gate-Based Strip Assay for Amplified Detection of Four miRNAs and Diagnosis of Lung Cancer, *Anal. Chem.*, 2023, **95**, 1280–1286.

209 C. Zhang, *et al.* Logical Analysis of Multiple Single-Nucleotide-Polymorphisms with Programmable DNA



Molecular Computation for Clinical Diagnostics, *Angew. Chem., Int. Ed.*, 2022, **61**, e202117658.

210 Y. Sato, T. Sakamoto and M. Takinoue, Sequence-based engineering of dynamic functions of micrometer-sized DNA droplets, *Sci. Adv.*, 2020, **6**, eaba3471.

211 Y. Sato and M. Takinoue, Capsule-like DNA Hydrogels with Patterns Formed by Lateral Phase Separation of DNA Nanostructures, *JACS Au*, 2022, **2**, 159–168.

212 J. Gong, N. Tsumura, Y. Sato and M. Takinoue, Computational DNA Droplets Recognizing miRNA Sequence Inputs Based on Liquid–Liquid Phase Separation, *Adv. Funct. Mater.*, 2022, **32**, 2202322.

213 M. Walczak, *et al.* Responsive core-shell DNA particles trigger lipid-membrane disruption and bacteria entrapment, *Nat. Commun.*, 2021, **12**, 4743.

214 A. Leathers, *et al.* Reaction-Diffusion Patterning of DNA-Based Artificial Cells, *J. Am. Chem. Soc.*, 2022, **144**, 17468–17476.

215 Y. Shimizu, T. Kanamori and T. Ueda, Protein synthesis by pure translation systems, *Methods*, 2005, **36**, 299–304.

216 A. D. Silverman, A. S. Karim and M. C. Jewett, Cell-free gene expression: an expanded repertoire of applications, *Nat. Rev. Genet.*, 2020, **21**, 151–170.

217 J. A. Peruzzi, N. R. Galvez and N. P. Kamat, Engineering transmembrane signal transduction in synthetic membranes using two-component systems, *Proc. Natl. Acad. Sci. U. S. A.*, 2023, **120**, e2218610120.

218 M. A. Boyd, W. Thavarajah, J. B. Lucks and N. P. Kamat, Robust and tunable performance of a cell-free biosensor encapsulated in lipid vesicles, *Sci. Adv.*, 2023, **9**, eadd6605.

219 M. Dwidar, *et al.* Programmable Artificial Cells Using Histamine-Responsive Synthetic Riboswitch, *J. Am. Chem. Soc.*, 2019, **141**, 11103–11114.

220 E. Godino and C. Danelon, Gene-Directed FtsZ Ring Assembly Generates Constricted Liposomes with Stable Membrane Necks, *Adv. Biol.*, 2023, **7**, 2200172.

221 Y. Uwaguchi, K. Fujiwara and N. Doi, Switching ON of Transcription-Translation System Using GUV Fusion by Co-supplementation of Calcium with Long-Chain Polyethylene Glycol, *ChemBioChem*, 2021, **22**, 2319–2324.

222 Z. Izri, D. Garenne, V. Noireaux and Y. T. Maeda, Gene Expression in on-Chip Membrane-Bound Artificial Cells, *ACS Synth. Biol.*, 2019, **8**, 1705–1712.

223 S. Berhanu, T. Ueda and Y. Kuruma, Artificial photosynthetic cell producing energy for protein synthesis, *Nat. Commun.*, 2019, **10**, 1325.

224 J. Kattan, A. Doerr, M. Dogterom and C. Danelon, Shaping Liposomes by Cell-Free Expressed Bacterial Microtubules, *ACS Synth. Biol.*, 2021, **10**, 2447–2455.

225 J. Garamella, S. Majumder, A. P. Liu and V. Noireaux, An Adaptive Synthetic Cell Based on Mechanosensing, Biosensing, and Inducible Gene Circuits, *ACS Synth. Biol.*, 2019, **8**, 1913–1920.

226 A. Poddar, *et al.* Membrane stretching activates calcium permeability of a putative channel Pkd2 during fission yeast cytokinesis, *Mol. Biol. Cell*, 2022, **33**, ar134.

227 H. Soga, *et al.* *In vitro* membrane protein synthesis inside cell-sized vesicles reveals the dependence of membrane protein integration on vesicle volume, *ACS Synth. Biol.*, 2014, **3**, 372–379.

228 T. Nitta, Y. Wang, Z. Du, K. Morishima and Y. Hiratsuka, A printable active network actuator built from an engineered biomolecular motor, *Nat. Mater.*, 2021, **20**, 1149–1155.

229 M. Akter, *et al.* Cooperative cargo transportation by a swarm of molecular machines, *Sci. Robot.*, 2022, **7**, eabm0677.

230 N. C. Seeman and H. F. Sleiman, DNA nanotechnology, *Nat. Rev. Mater.*, 2017, **3**, 17068.

231 M. Deluca, Z. Shi, C. E. Castro and G. Arya, Dynamic DNA nanotechnology: Toward functional nanoscale devices, *Nanoscale Horiz.*, 2020, **5**, 182–201.

232 P. Yin, H. Yan, X. G. Daniell, A. J. Turberfield and J. H. Reif, A unidirectional DNA walker that moves autonomously along a track, *Angew. Chem., Int. Ed.*, 2004, **43**, 4906–4911.

233 A. E. Marras, L. Zhou, H. J. Su and C. E. Castro, Programmable motion of DNA origami mechanisms, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, 713–718.

234 M. Vogt, *et al.* Storage of mechanical energy in DNA nanorobotics using molecular torsion springs, *Nat. Phys.*, 2023, **19**, 741–751.

235 A. K. Pumm, *et al.* A DNA origami rotary ratchet motor, *Nature*, 2022, **607**, 492–498.

236 F. C. Simmel, B. Yurke and H. R. Singh, Principles and Applications of Nucleic Acid Strand Displacement Reactions, *Chem. Rev.*, 2019, **119**, 6326–6369.

237 S. He, Z. Ge, X. Zuo, C. Fan and X. Mao, Dynamic regulation of DNA nanostructures by noncanonical nucleic acids, *NPG Asia Mater.*, 2021, **13**, 42.

238 J. L. Mergny and D. Sen, DNA quadruple helices in nanotechnology, *Chem. Rev.*, 2019, **119**, 6290–6325.

239 J. Ryssy, *et al.* Corrigendum to: Light-Responsive Dynamic DNA-Origami-Based Plasmonic Assemblies, *Angew. Chem., Int. Ed.*, 2022, **61**, e202210394.

240 E. Del Gross, A. M. Dallaire, A. Vallée-Bélisle and F. Ricci, Enzyme-Operated DNA-Based Nanodevices, *Nano Lett.*, 2015, **15**, 8407–8411.

241 Q. Shen, M. W. Grome, Y. Yang and C. Lin, Engineering Lipid Membranes with Programmable DNA Nanostructures, *Adv. Biosyst.*, 2020, **4**, 1900215.

242 B. M. Mognetti, P. Cicuta and L. Di Michele, Programmable interactions with biomimetic DNA linkers at fluid membranes and interfaces, *Rep. Prog. Phys.*, 2019, **82**, 116601.

243 J. J. Benkoski and F. Höök, Lateral mobility of tethered vesicle-DNA assemblies, *J. Phys. Chem. B*, 2005, **109**, 9773–9779.

244 S. R. Tabaei, P. Jönsson, M. Brändén and F. Höök, Self-assembly formation of multiple DNA-tethered lipid bilayers, *J. Struct. Biol.*, 2009, **168**, 200–206.

245 M. Chung, R. D. Lowe, Y. H. M. Chan, P. V. Ganesan and S. G. Boxer, DNA-tethered membranes formed by giant vesicle rupture, *J. Struct. Biol.*, 2009, **168**, 190–199.



246 B. Van Lengerich, R. J. Rawle and S. G. Boxer, Covalent attachment of lipid vesicles to a fluid-supported bilayer allows observation of DNA-mediated vesicle interactions, *Langmuir*, 2010, **26**, 8666–8672.

247 P. A. Beales and T. Kyle Vanderlick, Specific binding of different vesicle populations by the hybridization of membrane-anchored DNA, *J. Phys. Chem. A*, 2007, **111**, 12372–12380.

248 P. A. Beales and T. K. Vanderlick, DNA as membrane-bound ligand-receptor pairs: Duplex stability is tuned by intermembrane forces, *Biophys. J.*, 2009, **96**, 1554–1565.

249 L. Parolini, *et al.* Volume and porosity thermal regulation in lipid mesophases by coupling mobile ligands to soft membranes, *Nat. Commun.*, 2015, **6**, 5948.

250 L. Parolini, J. Kotar, L. Di Michele and B. M. Mognetti, Controlling Self-Assembly Kinetics of DNA-Functionalized Liposomes Using Toehold Exchange Mechanism, *ACS Nano*, 2016, **10**, 2392–2398.

251 J. Han, K. Pluhackova and R. A. Böckmann, The multifaceted role of SNARE proteins in membrane fusion, *Front. Physiol.*, 2017, **8**, 244875.

252 G. Stengel, R. Zahn and F. Höök, DNA-induced programmable fusion of phospholipid vesicles, *J. Am. Chem. Soc.*, 2007, **129**, 9584–9585.

253 G. Stengel, L. Simonsson, R. A. Campbell and F. Höök, Determinants for membrane fusion induced by cholesterol-modified DNA zippers, *J. Phys. Chem. B*, 2008, **112**, 8264–8274.

254 Y. H. M. Chan, B. Van Lengerich and S. G. Boxer, Effects of linker sequences on vesicle fusion mediated by lipid-anchored DNA oligonucleotides, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 979–984.

255 M. Paez-Perez, I. A. Russell, P. Cicuta and L. Di Michele, Modulating membrane fusion through the design of fusogenic DNA circuits and bilayer composition, *Soft Matter*, 2022, **18**, 7035–7044.

256 J. A. Peruzzi, M. L. Jacobs, T. Q. Vu, K. S. Wang and N. P. Kamat, Barcoding Biological Reactions with DNA-Functionalized Vesicles, *Angew. Chem., Int. Ed.*, 2019, **58**, 18683–18690.

257 N. J. Gaut, *et al.* Programmable Fusion and Differentiation of Synthetic Minimal Cells, *ACS Synth. Biol.*, 2022, **11**, 855–866.

258 L. Sun, *et al.* Guiding protein delivery into live cells using DNA-programmed membrane fusion, *Chem. Sci.*, 2018, **9**, 5967–5975.

259 S. Takamori, P. Cicuta, S. Takeuchi and L. Di Michele, DNA-assisted selective electrofusion (DASE) of *Escherichia coli* and giant lipid vesicles, *Nanoscale*, 2022, **14**, 14255–14267.

260 A. Czogalla, *et al.* Amphipathic DNA Origami Nanoparticles to Scaffold and Deform Lipid Membrane Vesicles, *Angew. Chem., Int. Ed.*, 2015, **54**, 6501–6505.

261 H. G. Franquelim, A. Khmelinskaia, J. P. Sobczak, H. Dietz and P. Schwille, Membrane sculpting by curved DNA origami scaffolds, *Nat. Commun.*, 2018, **9**, 811.

262 C. M. A. Journot, V. Ramakrishna, M. I. Wallace and A. J. Turberfield, Modifying Membrane Morphology and Interactions with DNA Origami Clathrin-Mimic Networks, *ACS Nano*, 2019, **13**, 9973–9979.

263 J. W. Conway, *et al.* Dynamic behavior of DNA cages anchored on spherically supported lipid bilayers, *J. Am. Chem. Soc.*, 2014, **136**, 12987–12997.

264 J. K. D. Singh, *et al.* Binding of DNA origami to lipids: Maximizing yield and switching *via* strand displacement, *Nucleic Acids Res.*, 2021, **49**, 10835–10850.

265 K. Jahnke, *et al.* Proton gradients from light-harvesting *E. coli* control DNA assemblies for synthetic cells, *Nat. Commun.*, 2021, **12**, 3967.

266 Y. Suzuki, M. Endo, Y. Yang and H. Sugiyama, Dynamic assembly/disassembly processes of photoresponsive DNA origami nanostructures directly visualized on a lipid membrane surface, *J. Am. Chem. Soc.*, 2014, **136**, 1714–1717.

267 D. Morzy, *et al.* Cations Regulate Membrane Attachment and Functionality of DNA Nanostructures, *J. Am. Chem. Soc.*, 2021, **143**, 7358–7367.

268 P. A. Beales and T. Kyle Vanderlick, Partitioning of membrane-anchored DNA between coexisting lipid phases, *J. Phys. Chem. B*, 2009, **113**, 13678–13686.

269 Y. Sato, *et al.* Environment-Dependent Self-Assembly of DNA Origami Lattices on Phase-Separated Lipid Membranes, *Adv. Mater. Interfaces*, 2018, **5**, 1800437.

270 R. Rubio-Sánchez, S. E. Barker, M. Walczak, P. Cicuta and L. D. Michele, A Modular, Dynamic, DNA-Based Platform for Regulating Cargo Distribution and Transport between Lipid Domains, *Nano Lett.*, 2021, **21**, 2800–2808.

271 R. Rubio-Sánchez, B. M. Mognetti, P. Cicuta and L. Di Michele, DNA-Origami Line-Actants Control Domain Organization and Fission in Synthetic Membranes, *J. Am. Chem. Soc.*, 2023, **145**, 2.

272 S. D. Perrault and W. M. Shih, Virus-inspired membrane encapsulation of DNA nanostructures to achieve *in vivo* stability, *ACS Nano*, 2014, **8**, 5132–5140.

273 Y. Yang, *et al.* Self-assembly of size-controlled liposomes on DNA nanotemplates, *Nat. Chem.*, 2016, **8**, 476–483.

274 Z. Zhang, Y. Yang, F. Pincet, M. C. Llaguno and C. Lin, Placing and shaping liposomes with reconfigurable DNA nanocages, *Nat. Chem.*, 2017, **9**, 653–659.

275 V. Maingi, *et al.* Digital nanoreactors to control absolute stoichiometry and spatiotemporal behavior of DNA receptors within lipid bilayers, *Nat. Commun.*, 2023, **14**, 1532.

276 S. Tanaka, K. Takiguchi and M. Hayashi, Repetitive stretching of giant liposomes utilizing the nematic alignment of confined actin, *Commun. Phys.*, 2018, **1**, 18.

277 F. C. Keber, *et al.* Topology and dynamics of active nematic vesicles, *Science*, 2014, **345**, 1135–1139.

278 T. Furusato, *et al.* De Novo Synthesis of Basal Bacterial Cell Division Proteins FtsZ, FtsA, and ZipA Inside Giant Vesicles, *ACS Synth. Biol.*, 2018, **7**, 953–961.

279 F. Yuan, *et al.* Membrane bending by protein phase separation, *Proc. Natl. Acad. Sci. U. S. A.*, 2021, **118**, e2017435118.



280 T. Litschel, *et al.* Reconstitution of contractile actomyosin rings in vesicles, *Nat. Commun.*, 2021, **12**, 2254.

281 C. Gray, S. A. Tatulian, S. A. Wharton and L. K. Tamm, Effect of the N-terminal glycine on the secondary structure, orientation, and interaction of the influenza hemagglutinin fusion peptide with lipid bilayers, *Biophys. J.*, 1996, **70**, 2275–2286.

282 M. Mally, J. Majhenc, S. Svetina and B. Žekš, The response of giant phospholipid vesicles to pore-forming peptide melittin, *Biochim. Biophys. Acta, Biomembr.*, 2007, **1768**, 1179–1189.

283 T. Takahashi, *et al.* Multiple membrane interactions and versatile vesicle deformations elicited by melittin, *Toxins*, 2013, **5**, 637–664.

284 N. Uchida, *et al.* Endocytosis-Like Vesicle Fission Mediated by a Membrane-Expanding Molecular Machine Enables Virus Encapsulation for In Vivo Delivery, *J. Am. Chem. Soc.*, 2023, **145**, 6210–6220.

285 Q. Yu, *et al.* Inward Budding and Endocytosis of Membranes Regulated by de Novo Designed Peptides, *Langmuir*, 2018, **34**, 6183–6193.

286 S. Ghosh, *et al.* Motility of Enzyme-Powered Vesicles, *Nano Lett.*, 2019, **19**, 6019–6026.

287 H. Jin, J. Cui and W. Zhan, Enzymatic Janus Liposome Micromotors, *Langmuir*, 2023, **39**, 4198–4206.

288 T. Kurakazu, M. Takinoue, K. Kuribayashi-Shigetomi and S. Takeuchi, Flagella-driven liposomes: Liposomes actuated by attached flagella, *14th International Conference on Miniaturized Systems for Chemistry and Life Sciences 2010, MicroTAS 2010*, 2010, vol. 1, pp. 220–222.

289 H. Inaba, *et al.* Light-induced propulsion of a giant liposome driven by peptide nanofibre growth, *Sci. Rep.*, 2018, **8**, 6243.

290 Z. Chen, *et al.* Synthetic beta cells for fusion-mediated dynamic insulin secretion, *Nat. Chem. Biol.*, 2018, **14**, 86–93.

291 M. Han, *et al.* Submillimeter-scale multimaterial terrestrial robots, *Sci. Robot.*, 2022, **7**, eabn0602.

292 M. S. Kim, H. T. Lee and S. H. Ahn, Laser Controlled 65 Micrometer Long Microrobot Made of Ni-Ti Shape Memory Alloy, *Adv. Mater. Technol.*, 2019, **4**, 1900583.

293 Q. Liu, *et al.* Micrometer-sized electrically programmable shape-memory actuators for low-power microrobotics, *Sci. Robot.*, 2021, **6**, eabe6663.

294 M. Urso, M. Ussia and M. Pumera, Breaking Polymer Chains with Self-Propelled Light-Controlled Navigable Hematite Microrobots, *Adv. Funct. Mater.*, 2021, **31**, 2101510.

295 K. W. Gyak, *et al.* Magnetically Actuated SiCN-Based Ceramic Microrobot for Guided Cell Delivery, *Adv. Healthcare Mater.*, 2019, **8**, 1900739.

296 H. Ceylan, *et al.* 3D-Printed Biodegradable Microswimmer for Theranostic Cargo Delivery and Release, *ACS Nano*, 2019, **13**, 3353–3362.

297 Y. Choi, *et al.* Photopatterned microswimmers with programmable motion without external stimuli, *Nat. Commun.*, 2021, **12**, 4724.

298 L. A. Malik, A. Bashir, A. Qureashi and A. H. Pandith, Detection and removal of heavy metal ions: a review, *Environ. Chem. Lett.*, 2019, **17**, 1495–1521.

299 K. Górecki, *et al.* Microfluidic-Derived Detection of Protein-Facilitated Copper Flux Across Lipid Membranes, *Anal. Chem.*, 2022, **94**, 11831–11837.

300 H. Valkenier, N. López Mora, A. Kros and A. P. Davis, Visualization and quantification of transmembrane ion transport into giant unilamellar vesicles, *Angew. Chem., Int. Ed.*, 2015, **54**, 2137–2141.

301 K. Koiwai, *et al.* Single-cell RNA-seq analysis reveals penaeid shrimp hemocyte subpopulations and cell differentiation process, *Elife*, 2021, **10**, e66954.

302 M. Hiratani and R. Kawano, DNA Logic Operation with Nanopore Decoding to Recognize MicroRNA Patterns in Small Cell Lung Cancer, *Anal. Chem.*, 2018, **90**, 8531–8537.

303 M. Hiratani, M. Ohara and R. Kawano, Amplification and Quantification of an Antisense Oligonucleotide from Target microRNA Using Programmable DNA and a Biological Nanopore, *Anal. Chem.*, 2017, **89**, 2312–2317.

304 N. Takeuchi, M. Hiratani and R. Kawano, Pattern Recognition of microRNA Expression in Body Fluids Using Nanopore Decoding at Subfemtomolar Concentrations, *JACS Au*, 2022, **2**, 1829–1838.

305 Y. Lu, X. Y. Wu, Y. L. Ying and Y. T. Long, Simultaneous single-molecule discrimination of cysteine and homocysteine with a protein nanopore, *Chem. Commun.*, 2019, **55**, 9311–9314.

306 K. Sun, *et al.* Single-Molecule Interaction of Peptides with a Biological Nanopore for Identification of Protease Activity, *Small Methods*, 2020, **4**, 1900892.

307 U. Lamichhane, *et al.* Peptide translocation through the mesoscopic channel: Binding kinetics at the single molecule level, *Eur. Biophys. J.*, 2013, **42**, 363–369.

308 G. Huang, K. Willems, M. Soskine, C. Wloka and G. Maglia, Electro-osmotic capture and ionic discrimination of peptide and protein biomarkers with FraC nanopores, *Nat. Commun.*, 2017, **8**, 935.

309 A. Barba-Bon, *et al.* Fluorescence Monitoring of Peptide Transport Pathways into Large and Giant Vesicles by Supramolecular Host-Dye Reporter Pairs, *J. Am. Chem. Soc.*, 2019, **141**, 20137–20145.

310 Y. Wang, *et al.* Electrode-free nanopore sensing by DiffusiOptoPhysiology, *Sci. Adv.*, 2019, **5**, eaar3309.

311 X. Du, *et al.* Microscopic Screening of Cyclodextrin Channel Blockers by DiffusiOptoPhysiology, *Anal. Chem.*, 2021, **93**, 14161–14168.

312 M. Lelek, *et al.* Single-molecule localization microscopy, *Nat. Rev. Methods Primers*, 2021, **1**, 39.

313 G. Vicidomini, P. Bianchini and A. Diaspro, STED super-resolved microscopy, *Nat. Methods*, 2018, **15**, 173–182.

314 Y. Lyu, *et al.* Constructing Smart Protocells with Built-In DNA Computational Core to Eliminate Exogenous Challenge, *J. Am. Chem. Soc.*, 2018, **140**, 6912–6920.

315 A. Joesaar, *et al.* DNA-based communication in populations of synthetic protocells, *Nat. Nanotechnol.*, 2019, **14**, 369–378.



316 D. Toparlak, *et al.* Artificial cells drive neural differentiation, *Sci. Adv.*, 2020, **6**, eabb4920.

317 K. P. Adamala, D. A. Martin-Alarcon, K. R. Guthrie-Honea and E. S. Boyden, Engineering genetic circuit interactions within and between synthetic minimal cells, *Nat. Chem.*, 2017, **9**, 431–439.

318 B. C. Buddingh', J. Elzinga and J. C. M. van Hest, Intercellular communication between artificial cells by allosteric amplification of a molecular signal, *Nat. Commun.*, 2020, **11**, 1652.

319 T. Chakraborty and S. V. Wegner, Cell to Cell Signaling through Light in Artificial Cell Communities: Glowing Predator Lures Prey, *ACS Nano*, 2021, **15**, 9434–9444.

320 G. Bolognesi, *et al.* Sculpting and fusing biomimetic vesicle networks using optical tweezers, *Nat. Commun.*, 2018, **9**, 1882.

321 V. Maffeis, *et al.* Clustering of catalytic nanocompartments for enhancing an extracellular non-native cascade reaction, *Chem. Sci.*, 2021, **12**, 12274–12285.

322 X. Wang, *et al.* Chemical communication in spatially organized protocell colonies and protocell/living cell micro-arrays, *Chem. Sci.*, 2019, **10**, 9446–9453.

323 S. F. Shimobayashi, *et al.* Direct measurement of DNA-mediated adhesion between lipid bilayers, *Phys. Chem. Chem. Phys.*, 2015, **17**, 15615–15628.

324 O. A. Amjad, B. M. Mognetti, P. Cicuta and L. Di Michele, Membrane Adhesion through Bridging by Multimeric Ligands, *Langmuir*, 2017, **33**, 1139–1146.

325 A. Heidari, *et al.* Orthogonal Light-Dependent Membrane Adhesion Induces Social Self-Sorting and Member-Specific DNA Communication in Synthetic Cell Communities, *Small*, 2023, **19**, 2206474.

326 T. Chakraborty, S. M. Bartelt, J. Steinkühler, R. Dimova and S. V. Wegner, Light controlled cell-to-cell adhesion and chemical communication in minimal synthetic cells, *Chem. Commun.*, 2019, **55**, 9448–9451.

327 R. Lentini, *et al.* Two-Way Chemical Communication between Artificial and Natural Cells, *ACS Cent. Sci.*, 2017, **3**, 117–123.

328 Y. Ding, L. E. Contreras-Llano, E. Morris, M. Mao and C. Tan, Minimizing Context Dependency of Gene Networks Using Artificial Cells, *ACS Appl. Mater. Interfaces*, 2018, **10**, 30137–30146.

329 J. M. Smith, D. Hartmann and M. J. Booth, Engineering cellular communication between light-activated synthetic cells and bacteria, *Nat. Chem. Biol.*, 2023, 1138–1146.

330 M. Kaneda, *et al.* Direct formation of proteo-liposomes by *in vitro* synthesis and cellular cytosolic delivery with connexin-expressing liposomes, *Biomaterials*, 2009, **30**, 3971–3977.

331 M. Yoshida, E. Muneyuki and T. Hisabori, ATP synthase - A marvellous rotary engine of the cell, *Nat. Rev. Mol. Cell Biol.*, 2001, **2**, 669–677.

332 K. Y. Lee, *et al.* Photosynthetic artificial organelles sustain and control ATP-dependent reactions in a protocellular system, *Nat. Biotechnol.*, 2018, **36**, 530–535.

333 C. Li, *et al.* Reversible Deformation of Artificial Cell Colonies Triggered by Actin Polymerization for Muscle Behavior Mimicry, *Adv. Mater.*, 2022, **34**, 2204039.

334 J. M. Hicks, *et al.* Electric Field Induced Biomimetic Transmembrane Electron Transport Using Carbon Nanotube Porins, *Small*, 2021, **17**, e2102517.

335 Ž. Pandur, I. Dogsa, M. Dular and D. Stopar, Liposome destruction by hydrodynamic cavitation in comparison to chemical, physical and mechanical treatments, *Ultrason. Sonochem.*, 2020, **61**, 104826.

336 D. Karastoyanov and S. Karastanov, Reuse Industrial Robots Reuse Robots Reuse Industrial Industrial, *IFAC-Pap.*, 2018, **51**, 44–47.

337 S. Khan, M. Li, S. P. Muench, L. J. C. Jeukens and P. A. Beales, Durable proteo-hybrid vesicles for the extended functional lifetime of membrane proteins in bionanotechnology, *Chem. Commun.*, 2016, **52**, 11020–11023.

338 A. Peyret, E. Ibarboure, J. Meins and S. LeCOMMANDOUX, Asymmetric Hybrid Polymer- Lipid Giant Vesicles as Cell Membrane Mimics, *Anal. Chem.*, 2022, **94**, 6311–6317.

339 D. W. Jeong, H. Jang, S. Q. Choi and M. C. Choi, Enhanced stability of freestanding lipid bilayer and its stability criteria, *Sci. Rep.*, 2016, **6**, 38158.

340 Q. Shen, *et al.* Functionalized DNA-Origami-Protein Nanopores Generate Large Transmembrane Channels with Programmable Size-Selectivity, *J. Am. Chem. Soc.*, 2023, **145**, 1292–1300.

341 A. Henning-Knechtel, J. Knechtel and M. Magzoub, NAR breakthrough article DNA-Assisted oligomerization of pore-forming toxin monomers into precisely-controlled protein channels, *Nucleic Acids Res.*, 2017, **45**, 12057–12068.

342 E. Spruijt, S. E. Tusk and H. Bayley, DNA scaffolds support stable and uniform peptide nanopores, *Nat. Nanotechnol.*, 2018, **13**, 739–745.

343 A. Fennouri, *et al.* Tuning the Diameter, Stability, and Membrane Affinity of Peptide Pores by DNA-Programmed Self-Assembly, *ACS Nano*, 2021, **15**, 11263–11275.

344 R. T. Nagipogu, D. Fu and J. H. Reif, A survey on molecular-scale learning systems with relevance to DNA computing, *Nanoscale*, 2023, **15**, 7676–7694.

345 Z. Chen, *et al.* De novo design of protein logic gates, *Science*, 2020, **84**, 78–84.

346 E. Kopperger, *et al.* A self-assembled nanoscale robotic arm controlled by electric fields, *Science*, 2018, **359**, 296–301.

347 S. Lauback, *et al.* Real-time magnetic actuation of DNA nanodevices *via* modular integration with stiff micro-levers, *Nat. Commun.*, 2018, **9**, 1446.

348 J. Zhao, *et al.* Mimicking Cellular Metabolism in Artificial Cells: Universal Molecule Transport across the Membrane through Vesicle Fusion, *Anal. Chem.*, 2022, **94**, 3811–3818.

349 H. C. Ates, *et al.* End-to-end design of wearable sensors, *Nat. Rev. Mater.*, 2022, **7**, 887–907.

350 X. Liu, *et al.* Stretchable living materials and devices with hydrogel-elastomer hybrids hosting programmed cells, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **114**, 2200–2205.



351 P. Q. Nguyen, *et al.* Wearable materials with embedded synthetic biology sensors for biomolecule detection, *Nat. Biotechnol.*, 2021, **39**, 1366–1374.

352 K. J. Vannoy, N. E. Tarolla, P. J. Kauffmann, R. B. Clark and J. E. Dick, Detecting Methamphetamine in Aerosols by Electroanalysis in a Soap Bubble Wall, *Anal. Chem.*, 2022, **94**, 6311–6317.

353 X. Yang and E. Miyako, Soap Bubble Pollination, *iScience*, 2020, **23**, 101188.

354 Y. Zhang, H. Obuchi and T. Toyota, A Practical Guide to Preparation and Applications of Giant Unilamellar Vesicles Formed *via* Centrifugation of Water-in-Oil Emulsion Droplets, *Membranes*, 2023, **13**, 440.

355 M. C. Blosser, B. G. Horst and S. L. Keller, CDICE method produces giant lipid vesicles under physiological conditions of charged lipids and ionic solutions, *Soft Matter*, 2016, **12**, 7364–7371.

356 S. Deshpande, Y. Caspi, A. E. C. Meijering and C. Dekker, Octanol-assisted liposome assembly on chip, *Nat. Commun.*, 2016, **7**, 10447.

357 T. Ebenhan, O. Gheysens, H. G. Kruger, J. R. Zeevaart and M. M. Sathekge, Antimicrobial Peptides: Their Role as Infection-Selective Tracers for Molecular Imaging, *BioMed Res. Int.*, 2014, **2014**, 867381.

358 R. Qing, *et al.* Protein Design: From the Aspect of Water Solubility and Stability, *Chem. Rev.*, 2022, **122**, 14085–14179.

359 Y. Xing, A. Rottensteiner, J. Ciccone and S. Howorka, Functional Nanopores Enabled with DNA, *Angew. Chem., Int. Ed.*, 2023, **62**, e202303103.

360 A. J. Hockenberry and M. C. Jewett, Synthetic *in vitro* circuits, *Curr. Opin. Chem. Biol.*, 2012, **16**, 253–259.

361 M. Takinoue, DNA droplets for intelligent and dynamical artificial cells: from the viewpoint of computation and non-equilibrium systems, *Interface Focus*, 2023, **13**, 20230021.

362 P. Wang, T. A. Meyer, V. Pan, P. K. Dutta and Y. Ke, The Beauty and Utility of DNA Origami, *Chem.*, 2017, **2**, 359–382.

363 D. Yang, C. Zhou, F. Gao, P. Wang and Y. Ke, DNA-Guided Assembly of Molecules, Materials, and Cells, *Adv. Intell. Syst.*, 2020, **2**, 1900101.

364 V. Mukwaya, S. Mann and H. Dou, Chemical communication at the synthetic cell/living cell interface, *Commun. Chem.*, 2021, **4**, 161.

365 J. Luan, D. Wang and D. A. Wilson, Leveraging synthetic particles for communication: From passive to active systems, *Nanoscale*, 2020, **12**, 21015–21033.

366 P. Albanese, F. Mavelli and E. Altamura, Light energy transduction in liposome-based artificial cells, *Front. Bioeng. Biotechnol.*, 2023, **11**, 1161730.

367 E. Rideau, F. R. Wurm and K. Landfester, Self-Assembly of Giant Unilamellar Vesicles by Film Hydration Methodologies, *Adv. Biosyst.*, 2019, **3**, 1800324.

