


 Cite this: *RSC Adv.*, 2021, **11**, 33952

A periodic review of chemical and pharmacological profiles of Tubiechong as insect Chinese medicine

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Tubiechong, in Chinese medicine, denotes the dried female insects of *Eupolyphaga sinensis* Walker (ESW) or *Polyphaga plancyi* Bolivar (PPB). As a traditional insect-type, in medicine, it has been historically utilized to treat bruises, fractures, amenorrhea, postpartum blood stasis, lumps and relieving pain. We herein have performed a systematic survey involving the chemical and biological studies in the past decades to reveal the value of such insect resources for their development and clinical utilization. Chemical studies indicated that Tubiechong generated many active compounds, including proteins, amino acids, peptides, fatty acids, alkaloids, nucleosides, polysaccharides, fat-soluble vitamins and mineral elements. Tubiechong or its extract has a wide range of activities including anticoagulation and anti-thrombosis, anti-tumor, antioxidant, immune regulation, blood lipid regulation and hepatoprotection. Finally, a periodic mini-review was conducted to summarize such chemical and pharmacological profiles of Tubiechong medicine. The active peptides in Tubiechong are majorly focused in this review and introduced as one important aspect since there is much literature and huge investigative interest in it. Traditional medical use of the insect was also stressed in this review associating with its disease-eliminating actions by promoting blood circulation or eliminating tissue-swelling pains, which might play important roles in anticancer practices or investigation. In accordance with the modern pharmacological progress, Tubiechong and its extracts indeed exerted antitumor actions through multiple pathways, such as interfering with tumor biological behaviors (growth, apoptosis, invasion, metastasis and angiogenesis), and regulating host immune function. To some extent, this knowledge would provide a basis for further research and application of Tubiechong medicine.

 Received 15th July 2021
 Accepted 3rd October 2021

DOI: 10.1039/d1ra05425b

rsc.li/rsc-advances

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of active ingredients from traditional Chinese medicine Tubiechong material.



1 Introduction

It is determined that insects are critical sources of active agents for modern medicine. Many insects such as bees, flies, ants, cockroaches and blood-sucking insects are utilized to derive antibacterial, antitumor or antithrombotic agents. Human beings have utilized them as food, medicine, cosmetic and biochemical materials for ages for the reason that they are one of the largest resources of living organisms. In long-term practice, medicinal insects and their products have multiple applications. The most important of these is to treat or prevent various diseases, including infectious diseases, neoplasia, and miscellaneous disorders.¹ Tubiechong is an acclaimed medicinal insect, and it is a dry female carcass of the *Eupolyphaga sinensis* Walker (ESW) or *Polyphaga plancyi* Bolivar (PPB). Between the two sources, ESW is more commonly applied than PPB due to the abundant resources. ESW, belonging to the family Corydiidae (Blattodea), is widely distributed in China,² and it is a critical insect used in traditional Chinese medicine (TCM) to eliminate the stasis caused by low movement of blood in local tissue. Moreover, Tubiechong is known by the Chinese as one preferred drug to make fast healing of damaged tendons and reinforce bone strength. In TCM, Tubiechong is considered a natural health product and is expansively utilized to treat or prevent diseases, including bone injury, bruises, cancer, hepatic fibrosis, and immune-related diseases.³

Tubiechong is one of the numerous insects that have been used in TCM for a long time. This medicine not only includes many nutrients but also has high medicinal value and health benefits.⁴ Modern pharmacological studies indicate that Tubiechong has anticoagulation, antithrombotic activity, hepatoprotective and antitumor effects, *etc.*⁵ In Southeast Asia, such as in China, Thailand, India and Malaysia, due to its special flavor and healthy effects, Tubiechong has been exploited as tonic and spice recently.¹ Until now, it is still difficult to understand the therapeutic role of Tubiechong against various

diseases completely, especially the chemical basis and action mechanism. Although, some extracts have been isolated from Tubiechong and proven to have pharmacological activities, and even several pure compounds with strong activities have been purified and identified for their absolute structures.^{3,6} Many research efforts are needed along the modern pharmacologic route to explore the chemical constitutes and biological actions of Tubiechong. We will retrospectively present the traditional usage, chemical compositions and pharmacological actions of medically relevant insects in this review. It is anticipated to give an integrated viewpoint on the chemical diversity and pharmacological actions of Tubiechong from ingredient-action relation in order to avoid unilateral conclusions such as those in most previous investigations. Some content might address the potential development and application of Tubiechong agents in the future. Some anticancer products of Tubiechong were cited in this review and their possible mechanisms through interfering with carcinogenesis or cancer progression were highlighted.

2 Traditional medical opinion and clinical application

In TCM, the use of Tubiechong was documented in many well-known medicinal works, such as Shennong's Herbal Classic, Synopsis of the Golden Chamber and Compendium of Materia Medica. It was believed to have abundant capabilities, *e.g.* renewing muscles and bones, boosting the immune response and removing blood stasis to stimulate blood circulation. Because of the light of TCM theories, those diseases are closely related to Qi stagnation and blood stasis in the organs. Therefore, Tubiechong was widely employed to treat many diseases, including bruises, fractures, amenorrhea, postpartum blood stasis, lumps and it was also used to control pain.^{7,8} These insect medicines also have many folk names, including Dibiechong, Tuyuan, Diwugui, Zhechong, Zhonghuazhendibie,



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on investigating the inhibition of natural products on HSC activation.



Lingchong Wang received his B. S. in 2003, and his PhD (2008) in marine biological science from Ocean University of China. He worked for the Nanjing University of Chinese Medicine as assistant research fellow from 2008 to 2012, and then he was promoted as Associate Professor. He was a visiting scientist at the University of Hong Kong from 2018 to 2019 and worked in cooperation with Dr Yibin Feng

in herbal ingredients screen for cancer and hepatic diseases. His research interests include products development and design with new dosage form, isolation, purification and structural elucidation of active ingredients from natural resources. He has authored 12 patents and 100+ papers.



Chouchongmu and Jieguchong. In TCM, folk physicians reckoned Tubiechong to be tasting salty and cold, and possibly showing slight toxicity to patients. After oral administration of decoction of Tubiechong, some Chinese folk physicians thought that the medicine would give blood meridians and distribute it into the liver channels. Thus, Tubiechong was widely applied to treat various painful diseases associated with liver damage.

As for the traditional preparation methods, Tubiechong had different usages according to different indications, such as grinding, frying, water extraction, and rice wine extraction. Among them, the main method was extraction with water or rice wine whose alcohol content was less than 20%. Tubiechong is also an important part of many famous medicinal prescriptions and Chinese patent medicines. Expelling blood stasis decoction, also known as Xiayuxue Tang, was first recorded in the book Synopsis of the Golden Chamber, and consisted of three Chinese medicinal materials, Rhei Radix et Rhizoma, Persicae Semen, and Tubiechong. It had good clinical efficacy in treating hepatic fibrosis and cirrhosis.⁹ Similarly, a famous and classical Chinese herbal prescription, Dahuang Tubiechong pill (DHZCP), consisting of Tubiechong, Dahuang and other 10 herbs, was clinically utilized to treat hepatic diseases, gynecopathy and atherosclerosis in China for a long history.¹⁰ Chinese patent medicine is known as Huoxuezhitong capsule (HXZT) contains Tubiechong. HXZT capsule had the activities in activating the blood circulation and relieving pain, and it had been applied for osteoarthritis since 1974.¹¹ Lumbago free capsules, also known as Yaotongning (YTN) capsules, were registered TCM preparations for lumbar, leg pain, rheumatoid arthritis (RA) treatment in China for decades. YTN was composed of 11 herbs and Tubiechong was of the most important composition for manufacturing and quality control, as well as the efficiency of clinic applications.^{12,13} Another Chinese patent medicine containing Tubiechong as major ingredients belong to aitongke capsules, which was claimed as an antitumor agent and is mainly used for patients with the advanced cancers. Chinese medicine practitioners thought aitongke was capable of extenuating tumor swelling, eliminating cancer pain, reducing hydrops, lifting leukocytes and inducing pus evacuation. Aitongke prescriptions were composed of gold scorpion, Tubiechong, aspongopus, rhubarb, ginseng, *Gyrophora*, astragali and so on.¹⁴

Overall, according to TCM theoretical records and traditional clinical practice, Tubiechong was considered an effective Chinese insect medicine to treat those diseases occurring in parenchymal tissues or malignant cancers through play actions on blood vessels. This hypothesis might lay guides for modern chemical and pharmacological researches on Tubiechong.

3 Chemical constituents of Tubiechong

Many types of compounds have been found and identified from Tubiechong (ESW and/or PPB), including a variety of active proteins, amino acids, peptides, fatty acids, alkaloids,

nucleosides, polysaccharides, phenylpropanoids, pyrazines, fat-soluble vitamins, and mineral elements, and some of them can even be isolated and identified as new compounds.¹⁵

3.1 Proteins, amino acids, peptides

Tubiechong is an excellent source of high-quality proteins and is rich in amino acids.¹⁶ Amino acids, peptides, and proteins are commonly known as the major components of animal organisms, and most of them have been confirmed by modern studies as specific components of animal-derived biomass with unique nutritional and medicinal purposes. Tubiechong (ESW) has been found to contain 18 kinds of amino acids, which belong to regular amino acids and found in normal protein-constitutes. Among them, Gly, Ala, Pro, Tyr, Arg and Lys are the six most abundant amino acids.¹⁷ Another study showed that ESW had high contents of Arg, Ser and Ala residues, but little amounts of Gly and Iso residues. Moreover, the hydrophobic amino acid contents in ESW were relatively high, which accounted for nearly 46.7% of total amino acids. The abundance of hydrophobic amino acids could boost solubility in lipids and therefore enhance the activity of Tubiechong peptides product. One ESW crude peptide product named EPs was obtained by sequentially hydrolyzing with pepsin and trypsin, which was an effective antioxidant and could serve as a powerful treatment for skin photo-aging, and its further purified fractions A4 was confirmed to be more potent antioxidant activities.¹⁶ Tubiechong was also used to prepare antitumor protein products. Some researchers utilized salting-out, ultrafiltration, ion-exchange chromatography, hydrophobic chromatography, and gel filtration chromatographic techniques to purify antitumor proteins, such as EPS72 (ref. 3) and one unnamed glycoprotein product.¹⁸ EPS72 is a purified protein with 72 kDa of molecular weight and has potent anti-proliferative activities against A549 cancer cells.³ The glycoprotein has 41.3 kDa of molecular weight, and 10.5% carbohydrate content, and was capable of inhibiting Tea-8113 cells proliferation.¹⁸ EFP, a kind of fibrinolytic protein purified from ESW, is an anti-angiogenic agent. The anti-angiogenic proteins could strongly restrict the invasive growth and metastasis of malignant tumors. EFP was proved to inhibit the proliferation of MVEC (human microvascular endothelial cells).¹⁹ The subsequent study showed that EFP had an obvious antitumor effect on S180 and H₂₂ cells *in vivo*.²⁰ Tubiechong medicine is also suitable material to isolate protein products with various functions, such as fibrinolytic, plasminogen-activating, hypolipidemic and anti-microbial activities. A fibrinolytic protein named eupolytin1,²¹ was purified from ESW and was confirmed for its activity for inducing fibrinolysis and activating plasminogen. Additionally, scholars obtained a hypolipidemic peptide DP17 with 1.43 kDa of molecular weight from ESW through biomimetic enzymatic hydrolysis.²² In detail, following Table 1 summarizes all active peptide or protein products that have been prepared with Tubiechong recently and some critical information is also listed in the table.

Based on the currently available information, many protein products have been isolated from insect materials, and these,





Table 1 Summary of several active peptides or protein products isolated from Tubiechong medicine

Preparing method	Products	Identification	Characterizations	Bioactivities	Potential applications	Ref.
Hydrolyzation of ESW with pepsin and trypsin	EPs	Crude extract	With less than 3.3 kDa of Mw, fifteen amino acids (Asp, Glu, Ser, Gly, Thr, Ala, Arg, Tyr, Val, Met, Phe, Iso, Leu, Lys and Pro) in detection	Antioxidant	Protection against photo-aging skin	16
Hydrolyzation of ESW with pepsin and trypsin, gel filtration & ion exchange chromatography	A4	Purified fraction	With less Mw than EPs	Antioxidant	Protection against photo-aging of skin	23
Hydrolyzation of ESW with a trypsin protease, ultrafiltration, DEAE-cellulose chromatography	F-I, F-II, F-III	Purified fractions	No date	Antioxidant	Antioxidant agent	23
Homogenization of ESW, salting out, ultrafiltration, ion exchange, hydrophobic & gel filtration chromatographies	EPS72	Purified protein	72 kDa of Mw, a band in SDS-PAGE	Antitumor, anti-proliferative effect on A549 (IC_{50} , 18.76 $\mu\text{g mL}^{-1}$)	Protection against tumor	3
Homogenization of ESW, savage deproteinization, dialysis, DEAE-cellulose chromatography	Unnamed	Purified glycoprotein	41.3 kDa of Mw,	Antitumor, Antiproliferative effect on Tea-8113	Protection against tumor	18
Homogenization of ESW, salting out, DEAE-cellulose & Sephadex G-75 column chromatographies	EFP	Purified fibrinolytic peptide	No date	Anti-proliferative effect on MVEC, induce apoptosis and cause cell cycle arrest at S and G2/M phases	Anti-angiogenic agent	19
Homogenization of ESW, dialysis, cationic exchange, gel filtration & anionic exchange chromatographies	Eupolytin1	Purified protein	26 kDa of Mw, with IVGGSDANIEDLPYQL SFETIDYDVAVARVATPFSYSGG VQQLQVYVSPVIVSPQQCNDNYA SDPCQGDSSGPLT VGGYPGVYSNVATLR in sequence	Fibrinolytic and plasminogen-activating (PA) activities	Anti-thrombosis agent	21
Homogenization of ESW, salting out, DEAE-cellulose chromatography; PAGE electrophoresis	EFF-1, EFF-2, EFF-3	Purified protein	With 41 kDa, 32.9 kDa and 30.6 kDa of Mw, respectively	Fibrinolytic activities	Fibrinolytic enzyme	24
Homogenization of ESW; salting out; DEAE-cellulose & gel filtration chromatographies	Unnamed	Purified glycoprotein	41.3 kDa of Mw, 10.5% of carbohydrate content,	Fibrinolytic activities	Fibrinolytic enzyme	25
Water extract and alcohol precipitate, ion exchange & gel filtration chromatographies, RP-HPLC	Fraction VI	Purified fraction	3.8 kDa of Mw, 89.3% of protein content, two bands in SDS-PAGE	Fibrinolytic activities	Fibrinolytic enzyme	26



Table 1 (Contd.)

Preparing method	Products	Identification	Characterizations	Bioactivities	Potential applications	Ref.
Hydrolyzation of ESW with pepsin and trypsin, ultrafiltration & nanofiltration, macroporous resin, Sephadex G-25 column chromatography, RP-HPLC	DP17	Purified peptide	1.43 kDa of Mw, with DAVPGAGPAGCHFGAGP in sequence, 8 beta sheets and 2 alpha sheets in configuration	Lipid accumulation reducing in liver tissues, reducing blood lipids	Hypolipidemic agent	22
Hydrolyzation of ESW with pepsin and trypsin, ultrafiltration	APE	Crude extract	71.05 ± 3.10% of the protein content	Blood lipid reducing activity	Hypolipidemic agent	27
Homogenization of ESW, gel filtration chromatography, RP-HPLC	Unnamed	Purified peptide	ACDFQCWVTCRQYSIN FISARCNGDSCVCTFRT in sequence	Antimicrobial	Antimicrobial agent	28

peptides are basically assumed to have strong potentials in development and application due to their strong activities and high content, as well as structural diversity. However, further development of peptide products is limited in their separation, purification and structural identification. The combination of minimal separation rate and the diversification of components has caused a serious lack of understanding of the structure and function of macromolecular components of Tubiechong. Overall, it is worthy to develop innovative methods and strategies on functional peptides for utilizing Tubiechong materials.

3.2 Polysaccharides

Polysaccharides are macromolecules gained from natural sources and might possess diverse structures and various biological activities.⁶ Tubiechong also contained polysaccharide ingredients to contribute to its medical effects in the clinic. However, there are relatively few studies on polysaccharide constituents of Tubiechong due to their low content and difficulty in extraction. In spite of such a dilemma, a polysaccharide fraction named ESPS had been purified from ESW using ion exchange and gel chromatography. Its structure characterization and physiological effect (antitumor) were further explored. The structural determinations indicated that the mean weight (Mw) of ESPS was 21.4 kDa. The main chain of ESPS was mainly composed of $\rightarrow 4$ - α -D-Glcp-(1 \rightarrow and $\rightarrow 3$)- β -D-Galp-(1 \rightarrow . Moreover, the residues and their side chains are joined to the main chain through the O-6 atom of glucose and O-4 and O-6 atoms of Gal residue. Additionally, its monosaccharide composition included fucose (Fuc), rhamnose (Rha), xylose (Xyl), arabinose (Ara), galactose (Gal) and glucose (Glc), and their molar ratios were 3.1 : 7.4 : 9.3 : 13.9 : 26.5 : 39.7, respectively. The possible fragments constituting absolute polysaccharide structures of the ESPS are presented in Fig. 1, which could be used for further structure speculation. In evaluation, ESPS enhanced lymphocyte activity *in vitro*, mainly to the NK cells. Thereby, it could promote lymphocyte proliferation and inhibit liver cancer cell growth. Moreover, ESPS stimulated immunity and effectively inhibited H₂₂ cell growth in H₂₂-bearing mice. The results revealed that ESPS, the polysaccharide product in ESW had high anti-hepatocellular carcinoma activity and was a potential immunotherapy candidate for the treatment of liver cancer.⁶

3.3 Fatty acids

The content of fatty acids in ESW is relatively high. Among all lipids that are extractable from the insect, unsaturated fatty acids accounted for 75% of the total, and essential fatty acids linoleic acid accounted for 28.5% of unsaturated fatty acids. ESW is rich in essential fatty acids such as linolenic acid. The tonic effect of Tubiechong may be due to the essential fatty acids as nutritional ingredients in combination with the essential amino acids and some vitamins.¹ The chemical analysis uncovered the presence of 6 lipids components occurring as saturated and polyunsaturated fatty acids that accounted for 97.55% of fatty acids, indicating the high nutritional value of Tubiechong. Tubiechong oily extract consisted of a significant

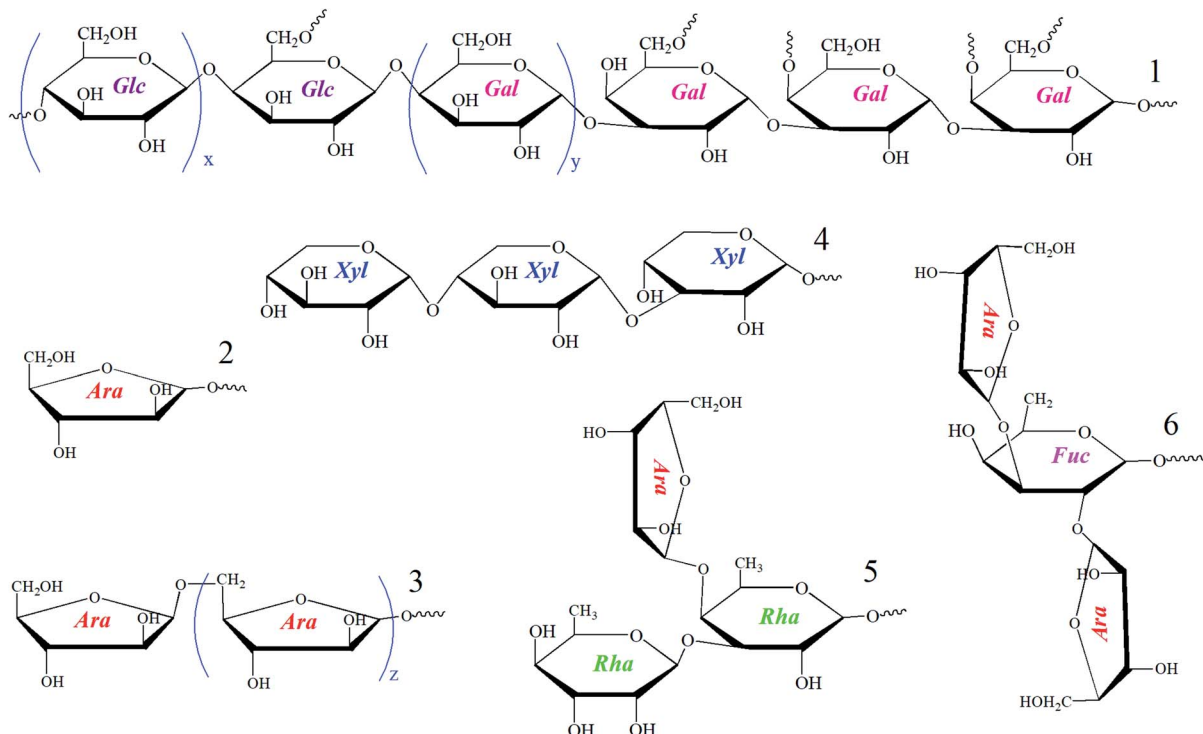
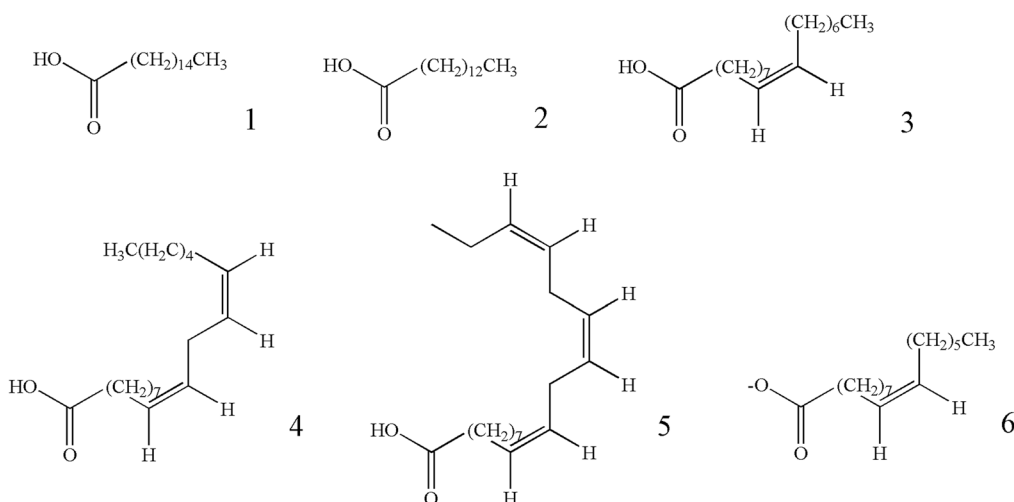


Fig. 1 Sugar chain fragments found in the structural elucidation of ESPS polysaccharide isolated from ESW.

amount of saturated and unsaturated fatty acids. The six highest content fatty acids in Tubiechong oily extract were identified as palmitic acid (21.70%), *cis*-9-oleic acid (40.78%),

cis-9,12-linoleic acid (21.86%), *cis*-9-palmitoleate (9.86%), *cis*-9,12,15-linolenate (1.69%) and myristate (1.67%). The extract containing a high amount of oleic and polyunsaturated fatty



- 1: Palmitic acid (pubmed CID: 985)
- 2: Myristic acid (pubmed CID: 11005)
- 3: Oleic acid (pubmed CID: 445639)
- 4: Linoleic acid (pubmed CID: 5280450)
- 5: Linolenic acid (pubmed CID: 5280934)
- 6: Palmitoleate (pubmed CID: 5461012)

Fig. 2 Chemical structures of 6 compounds identified as abundant fatty acids from TCM Tubiechong.



acids may have potential in treating tumors owing to oleic and polyunsaturated fatty acids produced anti-inflammatory, anti-tumor and cardioprotective effects.¹ The chemical structures of these major fatty acid molecules identified from Tubiechong extract are indicated in Fig. 2.

3.4 Nucleosides

Tubiechong contains nucleosides although there are a few literature reports on them. However, there are several reports on its metabolites and analogs. The nucleoside analogs mainly found in Tubiechong are uracil, allantoin and hypoxanthine. Among these nucleoside analogs, allantoin is even recognized as an active component and quality marker of Tubiechong by Chinese Pharmacopoeia. Allantoin can be separated from the *n*-hexane extract and *n*-butanol of extract ESW.²⁹ Moreover, the contents of uracil, xanthine, hypoxanthine and uridine from ESW were evaluated using RP-HPLC.³⁰ Another HPLC fingerprint analysis for ESW showed that there were seven characteristic peaks in the HPLC fingerprint of ESW, one of which was allantoin. The contents of allantoin were proportional to the analgesic effect of ESW.^{31,32} In addition, other scholars also isolated thymine from the ethyl acetate part of the 70% acetone extract of ESW.³³ The chemical structures of these nucleoside analogs founded in ESW are presented in Fig. 3.

3.5 Phenylpropanoids

Phenylpropanoids are an important group of natural products and are known to have multiple effects including antimicrobial, antioxidant, anti-inflammatory, antidiabetic and antitumor activities. In structure, those compounds are generally connected three straight-chain carbons with a benzene ring together. The classification includes simple phenylpropanoids, coumarins, lignans, lignins and flavonoids. Researchers recently confirmed that Tubiechong contained many types of phenylpropanoids compounds. According to their reports, at least thirteen phenylpropanoids compounds, including neolignans 3–11, 13,³⁴ isocoumarins 1, 2 (ref. 35) and a flavonoid

(genkwanin) 12,³⁶ were found in Tubiechong materials. The neolignans 3–11 and 13 were isolated from PPB and belonged to new compounds,³⁴ while isocoumarins and genkwanin were found in ESW.^{36,37} The chemical structures of these compounds are presented in Fig. 4. In addition, these insect-derived phenylpropanoids were also assayed in biological activities associated with clinic usages of Tubiechong. Most phenylpropanoids, covering compounds 1–11, possessed strong anti-inflammatory activities. It was also evidenced that the phenylpropanoids 5 and 7 could disrupt Smad activation and inhibit renal fibrosis. Genkwanin has proved to be a potential antitumor candidate in breast carcinoma therapy.³⁸ Studies by some other scholars have shown that genkwanin also has anti-inflammatory activities.^{39,40} It is common for phenylpropanoids to have such activities and there are many reports on them. For example, several important natural phenylpropanoids have significant antitumor and immunomodulatory activity on colorectal cancer in the APC (Min/+) (multiple intestinal neoplasia) mice.⁴¹ As far as the antitumor actions are concerned, phenylpropanoids could provoke apoptosis in breast cancer cells through reactive oxygen species and mitochondrial-dependent pathways.⁴² Moreover, studies demonstrated that phenylpropanoids induced cell apoptosis and thus exhibited strong cytotoxicity to liver cancer cell lines.⁴³ Apart from those, phenylpropanoids were thought to play immunostimulatory and antioxidant roles in exhibiting their antitumor abilities. Overall, the diversity and abundance of phenylpropanoids decided that Tubiechong would have huge potentials in development as therapeutics agents against inflammatory and/or cancerous diseases.

3.6 Alkaloids

Alkaloids are a large group of basic nitrogen-containing compounds that exist in organisms. Tubiechong contains a number of alkaloids. Pharmacodynamic experiments showed that the total alkaloids of Tubiechong could directly dilate blood vessels to reduce peripheral resistance and cardiac load. It could improve the tolerance of the myocardium and brain to

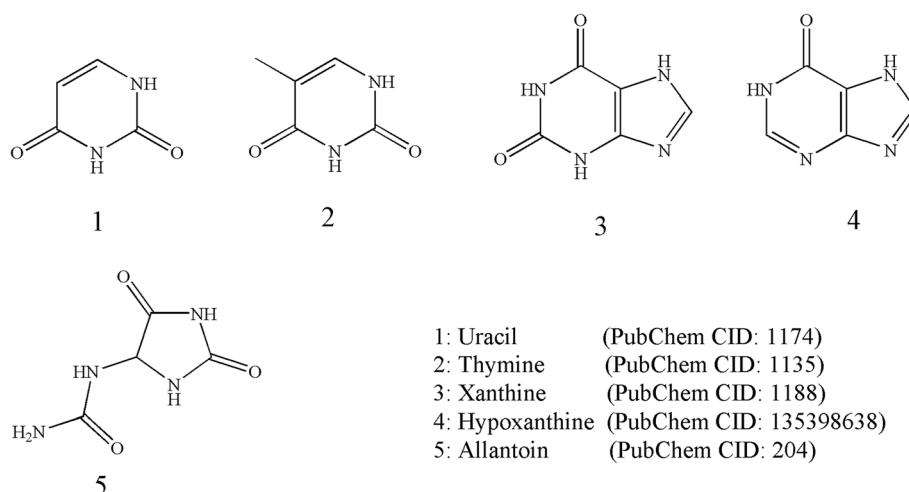
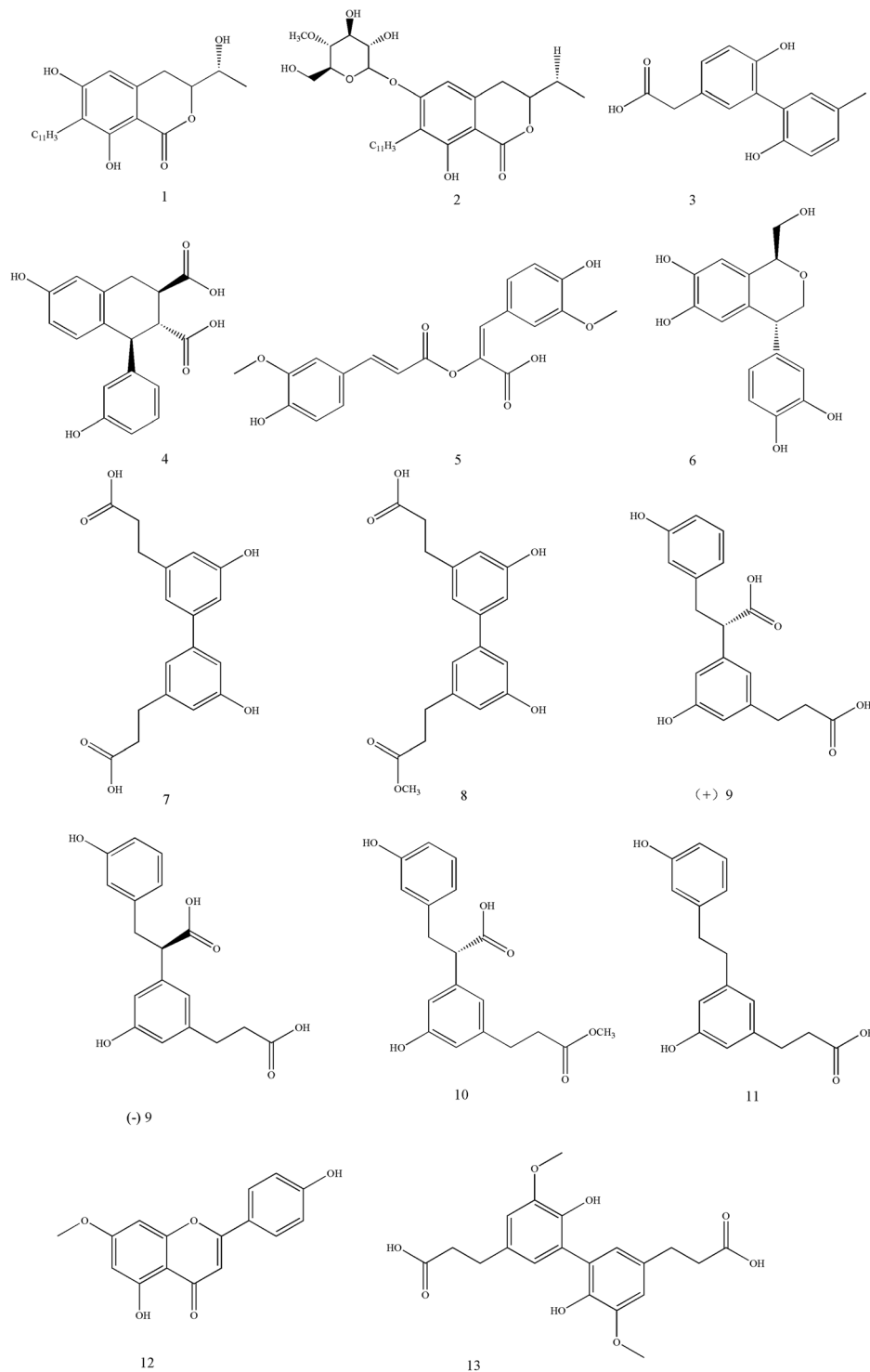


Fig. 3 Chemical structures of some nucleoside analogs detected in ESW.





1,2,3,4,5,6,7,8,9,10,11: New phenylpropanoids compounds
 12: (PubChem CID: 5281617)
 13: (PubChem CID: 11223011)
 Compounds 3-11 and 13 were from PPB
 Compounds 1,2 and 12 were from ESW

Fig. 4 Chemical structures of some phenylpropanoids detected in ESW and PPB.

ischemia and reduce the oxygen consumption of the heart and brain tissue.²⁵ Currently, twenty-one alkaloids are found in Tubiechong medicine. The chemical structures of these

molecules are presented in Fig. 5. On the one hand, about 18 alkaloids were separated from the ethanol extract of whole bodies of PPB.^{44,45} Among them, eight chemicals were known





1,2,3,4,5,6,7,8,9,10,11:New alkaloid compounds

12: (PubChem CID: 55264275)

13: (PubChem CID: 73227)

14: (PubChem CID: 3744)

15: (PubChem CID: 523151)

16: (PubChem CID: 17949580)

17: (PubChem CID: 17895110)

18: (PubChem CID: 6995640)

19: (PubChem CID: 349270380)

20: (PubChem CID: 135765825)

21: (PubChem CID: 23424618)

Compounds 1-9, 11, 12-16, 18, 19 and 21 were from PPB

Compounds 10, 17 and 20 were from ESW

Fig. 5 Chemical structures of some alkaloids identified from ESW and PPB.

alkaloids and recognized, such as alkaloids **12–16**, **18–19**, and **21**. Other ten alkaloids might be new compounds after being verified in the Pubchem database ([https://](https://pubchem.ncbi.nlm.nih.gov/)

pubchem.ncbi.nlm.nih.gov/), which indicated as compounds **1–9**, and **11**. The PPB-derived alkaloids were investigated for their renal protective activities against virus attraction and





Table 2 Pharmacological activities of Tubiechong and their extracts

Pharmacological activity	Testing substance	Tested living system/organ/cell	Dose and administration	Results	Mechanisms	Ref.
Anti-tumor activity	ESW 70% ethanol extract	Cultured A549 cells; cultured HUVECs;	0.1–0.8 mg mL ⁻¹ for 48 h	Inhibited cell proliferation (IC ₅₀ of HUVECs was 0.34 mg mL ⁻¹ ; IC ₅₀ of A549 was 0.27 mg mL ⁻¹) and migration	Inhibited the autophosphorylation of KDR, downregulate the activation of AKT and (ERK)1/2	5
	ESW 70% ethanol extract (ESWE)	Cultured MDA-MB-435s and MDA-MB-231 cells	0.4–1.6 mg mL ⁻¹ for 24 h	Inhibited cell proliferation (with 0.564 and 0.724 mg mL ⁻¹ of IC ₅₀ , respectively) and migration	Inhibition the expression of MAPK signaling and related metastasis factors	7
	ESW 70% ethanol extract (ESEE)	MDA-MB-231 xenograft mice model	200, 400 mg kg ⁻¹ for 14 days	Tumor growth inhibition		
	ESW 70% ethanol extract (ESEE)	Tumor (S180)-bearing mice	0.05–0.2 mg mL ⁻¹ for 48 h	Tumor growth inhibition, inducing G ₂ -M phase arrest	Down-regulating phosphorylation of EGFR, AKT and ERK1/2	53
	ESW 70% ethanol extract (ESWE)	SMMC-7721, BEL-7402 and HepG2 cells;	100, 200 and 400 mg kg ⁻¹ for 10 days	Inhibited cell proliferation (with 0.13 mg mL ⁻¹ , 0.14 mg mL ⁻¹ and 0.67 mg mL ⁻¹ of IC ₅₀ , respectively)	Inhibited growth and metastasis signaling (the PKC, AKT, MAPK signaling and related metastasis signaling)	54
	Polysaccharide from ESW (ESPS)	SMMC-7721 xenograft in athymic mice	400 mg kg ⁻¹ for 10 days	Tumor growth inhibition		
	ESW 95% ethanol extract (ESEE) (mainly fatty acids)	Tumor (H ₂₂)-bearing mice	31–124 mg kg ⁻¹ for 14 days	Tumor growth inhibition, promoting TNF- α and IFN- γ production and inducing apoptosis	Via increase of Bax/Bcl-2 ratio and activation of caspases-3	1
	Purified protein EPS72	Spleen lymphocytes	50, 100 and 200 μ g mL ⁻¹ for 48 h	Enhanced lymphocyte proliferation	Enhancement of lymphocyte cytotoxicity	6
	ESW 80% ethanol extract (ESWE)	NK cells	100, 200 and 400 μ g mL ⁻¹ for 48 h	Enhanced NK cytotoxicity		
	ESW 95% ethanol extract (ESE)	Tumor (H ₂₂)-bearing mice	5, 10 and 20 mg kg ⁻¹ once every two days for 15 days	Tumor growth inhibition		
	ESW 80% ethanol extract (ESWE)	A549 cells	5 and 40 μ g mL ⁻¹ for 48 h	Inhibited cell proliferation (with 18.76 μ g mL ⁻¹ of IC ₅₀), restrained cell migration and invasion	Inhibited cell adhesion to fibronectin and collagen IV, down-regulated the expression of β 1-integrin	3
	ESW 80% ethanol extract (ESWE)	PC3 cells	0.25 and 0.5 mg mL ⁻¹ for 24 h	Inhibited the growth, migration and invasion of PC3 cells	MMP-2 and MMP-9 expression inhibition	55
	ESW 95% ethanol extract (ESE)	HepG2 and SGC-7901 cells	0.1–0.535 μ g mL ⁻¹ for 48 h	Inhibited cell proliferation (with 0.90 and 0.11 μ g mL ⁻¹ of IC ₅₀ , respectively), induce HepG2 cell apoptosis	No mention	56
	Protein of ESW (EFP) containing serum	A549 cells	Serum drug from 0.73–2.90 mg kg ⁻¹ for 48 h	Tumor proliferation inhibition, inducing apoptosis	Increasing the ratio of Bax/Bcl-2	57



Table 2 (Contd.)

Pharmacological activity	Testing substance	Tested living system/organ/cell	Dose and administration	Results	Mechanisms	Ref.
	ESW fibrinolytic protein (EFP) drug serum	HepG2 and MCF-7 cells	Serum drug from 1.25 to 5 mg kg ⁻¹ for 48 h	Tumor proliferation inhibition, inhibiting angiogenesis	Down-regulating the expression of VEGF and bFGF	58
	Serum containing ESW	HepG2 cells	20% ESW serum for 72 hours	Tumor proliferation inhibition	Inducing G ₀ -G ₁ phase arrest	59
	Purified protein from ESW	Tea-8113 cells	0.010–0.0902 g mL ⁻¹ for 72 h	Tumor proliferation inhibition	No mention	18
	Neolignans from PPB	K562, A549, and Huh7 cells	2.5–40 μM for 48 h	Phenylpropanoids compound 9 inhibited cell proliferation of Huh-7 cells (with 23.2 μM and 27.1 μM of IC ₅₀ , respectively), and inhibited cell proliferation of K562, A549	No mention	34
Anti- thrombogenic and anticoagulant activities	Three kinds (EFF-1, EFF-2 and EFF-3) of fibrinolytic factors from ESW	Fibrin plate experiment <i>in vitro</i>	20 μL per well	Activated plasminogen	No mention	24
	Purified protein from ESW (eupolytin1)	Arteriovenous shunt rat models	0.06 μmol kg ⁻¹	Reduced thrombus weight	No mention	21
	Fraction from ESW (fraction VI)	Fibrin plate experiment <i>in vitro</i>	5–20 μL per well	Degraded fibrin and activated the plasminogen	No mention	26
	ESW fibrinolytic protein (EFP)	Carrageenan-induced thrombosis model mice	4.8, 16 g kg ⁻¹ d ⁻¹ for 10 days	Reduce the length of thrombus	No mention	60
Immunomodulatory activity	Decoction of ESW	Healthy mice	1.89–7.56 g kg ⁻¹ d ⁻¹ to mice for 4 weeks	Enhanced the carbon expurgatory index and phagocytic index	No mention	61
	Papain-hydrolyzed peptides of ESW	Healthy mice	0.3 g kg ⁻¹ d ⁻¹ to mice for 10 days	Increased index of thymus and spleen, enhanced the phagocytic function of macrophage and promoted the level of IL-2 in serum	No mention	62
	ESW lyophilized powder (ESL)	Immunosuppressed mice induced by cyclophosphamide	0.5, 1.0 and 2.0 g kg ⁻¹ d ⁻¹ for 14 days	Increased the immune organ index, mononuclear macrophages function and the level of NK cell	Down-regulated the phosphorylation of JNK and the Bax/Bcl-2 ratio	2
Hepatoprotective activity	ESW polypeptides	CCL4-induced chronic liver injury mice	50, 100 and 200 mg kg ⁻¹ d ⁻¹ for 6 weeks	Attenuates CCL ₄ -induced chronic liver injury in mice	Down-regulated the expression of Bax, caspase-3, α-SMA and TGF-β1	63
Anti-oxidative and anti-aging activities	Arazyme enzymatic hydrolysis peptides of ESW	D-Galactose-induced aging model mice	300 mg kg ⁻¹ d ⁻¹ for 6 weeks	Scavenge free radicals <i>in vitro</i> , reduced the content of MDA, increased the activities of CAT and GSH-Px and T-SOD in liver	No mention	64

Table 2 (Contd.)

Pharmacological activity	Testing substance	Tested living system/organ/cell	Dose and administration	Results	Mechanisms	Ref.
Polypeptide extracts of ESW		D-Galactose-induced aging model mice	0, 40, 80, 160 mg kg ⁻¹ d for 20 days	Enhanced the anti-stress and antioxidative capacity, delayed the oxidative aging	Initiating Nrf2-ARE antioxidant signaling pathway	65
Peptides from the enzymatic hydrolysate of ESW(F-I, F-II, and F-III)		Free radicals <i>in vitro</i>	F-II (10 mg mL ⁻¹ , 0.7 mg mL ⁻¹ , 1.8 mg mL ⁻¹ for O ₂ ^{•-} , OH [•] , and DPPH [•] respectively	Scavenge free radicals <i>in vitro</i>	No mention	23
Enzymatic hydrolysis products of ESW (EPs)		UV radiation-induced skin photoaging mice	Daubed with 140 μL of EPs at a dose of 25, 50 and 75 mg mL ⁻¹ per day for 8 weeks	Improved UV irradiation-induced damage of skin texture and morphology	Enhanced the activities of SOD, CAT and GPH-Px, increased the contents of HYP, and reduced the content of MDA in skin	16

tuberculosis. Surprisingly, most of them had strong activities. On the other hand, only three alkaloids had been reported in ESW, including compounds **10**, **17**, and **20**.^{33,35} Compound **10** might be a new compound and was extractable from the insect. Alkaloids **17** and **20** were isolated from the ethyl acetate extract of ESW and had some cytotoxicity to 10 types of cancer cells *in vitro*. The potential effects of those alkaloids compounds can be evidenced by many other individual pharmacological investigations. For example, some scholars have found that indole-3-glyoxylamide has anti-tumor⁴⁶ and antiviral activities.⁴⁷ Indole propionic acid has anti-inflammatory and antioxidant properties.⁴⁸ The analogs can even promote human and murine intestinal homeostasis,⁴⁹ restrain gut dysbiosis and endotoxin leakage to alleviate steatohepatitis in rats.⁵⁰ These pieces of evidence have supported strong activities and wide range of potential medical applications of Tubiechong.

3.7 Other constituents

Tubiechong also contains fat-soluble vitamins, mineral elements, steroids, pyranzines and alcohols. Vitamins play an important role in anti-oxidation and preventing fat peroxidation. Meanwhile, 4 trace element contents of Ca, Mg, Zn and Mn had also been found in ESW.⁵¹ Two steroids, including beta-sitosterol²⁹ and cholesterol³⁶ were reported in ESW along with octacosanol and butyl alcohol.²⁹ Furthermore, some pyrazines were also found in Tubiechong, including plancyprazine A isolated from PPB⁴⁵ and 2-methyl-6-(2/3/4'-trihydroxybutyl) pyrazine isolated from ESW.³³ In addition, there was a phthalide derivative isolated from ethanol extract of PPB.⁴⁵ Apart from those, some ketones were also found in Tubiechong, such as 6,8-dihydroxy-3,7-dimethyl-3,4-dihydro-1*H*-isochromen-1-one.³³ 3-Hydroxypyridine was found in Tubiechong as well.³³ There was also a lactone compound found in PPB. The lactone ring was combined with a benzene ring and glucose was connected to the benzene ring.⁴⁵

4 Pharmacological activities of Tubiechong

Modern pharmacological studies have confirmed that Tubiechong and its extracts have a wide range of pharmacological activities such as thrombolysis, anticoagulant, anti-tumor, immunomodulatory, anti-oxidative and hepatoprotective.⁵² Some important activities are summarized in Table 2.

4.1 Anti-tumor activity

According to TCM theories, the occurrence and progression of cancerous diseases are closely related to the Qi stagnation and blood stasis in the organs. Therefore, TCM physicians even believed that the medicines were very useful for the therapy and prevention of tumors if they had the capability of improving blood circulation and eliminating blood stasis.¹ Obviously, Tubiechong belongs to such medicines according to its traditional use and many anti-tumor ingredients.

First, some proteins prepared from ESW have been proved to have anti-tumor activities⁵⁸ due to their anti-angiogenesis



effect.^{19,66} At present, there are many research works paying attention to the anti-tumor effect of Tubiechong and its extracts. Some researchers have estimated the antitumor and immunomodulatory of ESEE (ESW ethanol extract) in hepatocarcinoma H₂₂ bearing mice. After implanting H₂₂ tumor cells, ICR mice were treated with ESEE for 14 consecutive days at doses of 31, 62 and 124 mg kg⁻¹. Oral administration of ESEE could inhibit tumor growth, enhance Th1 type cytokine production (TNF- α and IFN- γ) and induce apoptosis of hepatocarcinoma by increasing the ratio of Bax/Bcl-2 and activating caspases-3.¹ In addition, some people elucidated that a product extracted from ESW with 70% ethanol and named ESWE had an anti-proliferation and anti-invasion effect on breast cancer.⁷ ESWE was capable of inhibiting breast cancer growth, migration, and invasion. The mechanism underlying the above effects was that ESWE could weaken the activity of ERK1/2 and down-regulate the expression of CXCR4, MMP2, and MMP9. Another study indicated that 70% ethanol extract of ESW significantly restrained A549 cell migration in a time and dose-dependent manner and inhibited human umbilical vein endothelial cell proliferation, migration and tube formation. Furthermore, it effectively suppressed blood vessel formation in the established tissue model for angiogenesis. In addition, 70% ethanol extract of ESW was proved to inhibit the autophosphorylation of KDR, and down-regulate the subsequent activation of AKT and extracellular signal-regulated kinase (ERK)1/2 in A549 cells.⁵ Apart from these, a polysaccharide (ESPS) obtained from ESW by ion-exchange chromatography and gel chromatography was also proved to have strong antitumor activity. ESPS-enhanced lymphocyte activity *in vitro*, mainly NK cells. Thus, it could stimulate lymphocyte proliferation and inhibit liver cancer cell growth.⁶ Similarly, PPB and its extracts have anti-tumor activity as well. Some researchers have disclosed that compound

plancyol A and pyrazine compound plancypyrazine A isolated from PPB could inhibit JAK3 kinase with IC₅₀ values of 12.6 and 5.0 μ M, respectively. Besides, plancyol A expressed inhibitory activity towards DDR1 kinase with an IC₅₀ value of 4.87 μ M.⁴⁵ JAK kinase is related to cancer. Thus, these results showed their potential in anti-tumor applications. In general, Tubiechong and its extracts exert anti-tumor actions through multiple ways as shown in Fig. 6, such as interfering with tumor biological behavior (growth, differentiation, apoptosis, invasion, metastasis and angiogenesis), and regulating the host's anti-tumor response. Among all aspects, the more prominent antitumor roles of Tubiechong and its derived products lie in interfering with tumor cell proliferation, invasion and metastasis by inducing apoptosis, cell cycle arrest and inhibiting angiogenesis.

4.2 Anti-thrombotic and anticoagulant activities

The incidence of thrombotic disorders such as cerebral stroke, myocardial infarction, and venous thromboembolism is rapidly increasing throughout the world. Thrombolytic therapy is an acknowledged approach to treat these disorders. All thrombolytic agents in current clinical usage are plasminogen activators (PAs), which require plasminogen to achieve thrombolysis. Although effective, PAs uniformly increase the risk of bleeding complications, especially intracranial hemorrhage, and there is no laboratory test to avoid such bleeding.²¹ As reported in Compendium of Materia Medica, Tubiechong has been majorly applied as traditional anti-thrombosis medicine without bleeding risk for a long time.²¹ Three kinds (EFF-1, EFF-2 and EFF-3) of fibrinolytic factors from ESW were isolated by salting out, DEAE-cellulose column and preparative PAGE electrophoresis. Their activities as plasminogen activators were 171.3 U



Fig. 6 Some anti-tumor actions and mechanisms of Tubiechong and its extracts.



mg^{-1} , 234.0 U mg^{-1} and 148.5 U mg^{-1} ,²⁴ respectively. Another purified protein from ESW named eupolytin1, a bi-functional anti-thrombosis protein that not only possessed a direct-acting fibrinolytic activity but also had plasminogen-activating activity. Thus, it had anti-thrombosis activity *in vivo*. The thrombus weight was reduced to $1 \pm 0.2 \text{ mg}$ by administration of $0.06 \mu\text{mol kg}^{-1}$ of eupolytin1.²¹ In addition, a sample was obtained from ESW by water extraction and alcohol sinking, which was purified by ion-exchange column, gel filtration column and RP-HPLC, when fraction VI was obtained. Moreover, the experimental results of activity indicate that fraction VI had the ability to degrade fibrin directly and active the plasminogen.²⁶ Apart from ESW, neolignans, isolated from PPB, could play potential therapeutic roles in the inhibition of renal fibrosis by the disruption of Smad activation.³⁴ Besides, other researchers have found that alkaloid compounds **4**, **13**, and **21** might have benefits in the treatment of renal fibrosis with different and unique mechanisms through Smad or non-Smad pathways.⁴⁴

4.3 Immunomodulatory activity

Tubiechong has the immunoregulatory effect, which could contribute to its ability to develop and utilize as a health food resource.⁶⁷ Administration of the decoction of ESW in water at doses of $1.89\text{--}7.56 \text{ g kg}^{-1} \text{ d}^{-1}$ to mice for 4 weeks observably enhanced the carbon expurgatory index and phagocytic index of mice.⁶⁸ The papain-hydrolyzed peptides of ESW could elevate immune function in the mice model. *In vivo* tests demonstrated that ESW-derived peptide could significantly increase the index of the thymus and spleen of mice, enhance the phagocytic function of macrophage and promote the level of IL-2 in serum.⁶⁹ Another study elucidated that ESW lyophilized powder (ESL) had immuno-enhancement effects in immunosuppressed mice induced by cyclophosphamide. In brief, results suggested that ESL could modulate oxidative systems and innate immune cells, thereby effectively improving immune functions. ESL markedly increased the immune organ index, mononuclear macrophage function and the level of NK cells.² Moreover, many research works⁷⁰ showed that ESW could boost the immune function in rats of yin-deficiency and fire-hyperactivity syndrome with chronic blood stasis. Meanwhile, another study showed that ESW (25 g kg^{-1}) could enhance the immune function of mice since it could increase RBC-C3bR rosette rate in a mouse model with blood-deficiency. Besides, it could correct the mice body weight loss induced by cyclophosphamide and increase the spleen thymus index.⁷¹

4.4 Hepatoprotective activity

The liver is the metabolic center and participates in many important physiological functions. As the most common pathological process of various liver diseases, liver injury can develop into serious diseases such as hepatic cirrhosis, hepatic fibrosis, hepatic carcinoma, and cause harm to human health.

Research has shown that drugs that could improve blood circulation and remove blood stasis were used in model rats to treat mild chronic hepatic damage induced by CCl_4 .⁷²

Tubiechong is part of those drugs, it enters the liver meridian, and its hepatoprotective effect has been recorded from ancient times on. Tubiechong is one of the main components of DHZCP. Some reports showed that it could alleviate hepatic fibrosis by decreasing the secretion of $\text{TNF-}\alpha$ and IL-13 by the downregulation of p38 and ERK phosphorylation.⁷ Moreover, DHZCP significantly declined the levels of alanine aminotransferase (ALT), aspartate aminotransferase (AST), hyaluronic acid, laminin, type IV collagen and procollagen III, and reversed hepatic fibrosis in a rat model.⁷³ Another study suggested that DHZCP conferred protection against alcoholic liver fibrosis (ALF) injury in mice by suppressing the generation of collagen 1 (COL-1) and down-regulating apoptosis of liver cells as a result of adjusting the levels of inflammatory factors.^{74,75} These results indicated that Tubiechong was used for the treatment of liver diseases in ancient times. In addition, some researchers have investigated the antioxidative protective effects of ESW polypeptides on CCl_4 -induced chronic liver injury in mice.⁶³ No microscopic abnormalities were found in the liver cells of the normal group. In the model group, the hepatocytes showed hyperemia, necrosis and fibrosis, and the hepatic lobules had disappeared. The morphological structures of the hepatocytes in the ESW peptide group were damaged, but increasing the ESW peptide dose significantly lessened the degree of the injury, and the liver and spleen indices of these mice were significantly lower than those of the model group. Compared with the model group, the serum AST, ALT activities and the malondialdehyde (MDA) content in the livers of the mice in the ESW peptide group decreased significantly. Additionally, the antioxidant enzyme (superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GSH-Px)) activities were remarkably improved, the inflammatory factor (IL-6, TNF- α , iNOS) protein and gene expression levels were significantly decreased, and the proapoptotic factor (Bax, Bcl-2, caspase-3) and fibrosis factor (α -SMA, TGF- β) gene levels were significantly decreased and positively correlated with the ESW polypeptide dose. It suggested that ESW peptides could attenuate CCl_4 -induced chronic liver injury in mice, and these protective effects might be related to the antioxidant activities of the peptide products.

4.5 Anti-oxidative and anti-aging activities

Tubiechong and its extracts have antioxidant activity.⁷⁶⁻⁷⁸ Some scholars have disclosed that arazyme enzymatic hydrolysis peptides of ESW could scavenge hydroxyl, superoxide anion and DPPH free radicals with IC_{50} of 0.40, 8.73 mg mL^{-1} and 1.32 mg mL^{-1} , respectively. Compared to the aging model group, the peptides significantly reduced the content of MDA, increased the activities of CAT and GSH-Px and hydroxyl radical scavenging rate in both plasma and liver, and also enhanced the activity of T-SOD in liver.⁶⁴ The results of another study revealed that polypeptide extracts of ESW enhanced the anti-stress and antioxidative capacities in D-galactose-induced mouse models of oxidative aging by initiating the Nrf2-ARE antioxidant signaling pathway, therefore, delayed oxidative aging in mice.⁶⁵ Other scholars investigated the separation of antioxidant



peptides from the enzymatic hydrolysate of ESW and the antioxidant activities of peptides. Three fractions, including *F-I*, *F-II*, and *F-III*, were obtained using DEAE-Sephadex A50 chromatography, of which, the fraction *F-II* exhibited the highest $O_2^{\cdot-}$, OH^{\cdot} , and $DPPH^{\cdot}$ scavenging capabilities, reaching 86.83% (10 mg mL^{-1}), 92.28% (0.7 mg mL^{-1}), and 68.06% (1.8 mg mL^{-1}), respectively.²³

4.6 Other activities

Tubiechong also has analgesic activity and can be a treatment for arthralgia⁷⁹ and avascular necrosis of femoral head.⁸⁰ Besides, ESW could regulate blood lipids. ESW freeze-dried powder could inhibit the rise of the blood lipid of rabbits that were fed with a high-fat diet.⁸¹ Other scholars found that ESW could also reduce the cholesterol levels of hypercholesterolaemia rabbits.⁸² Another active peptide of ESW (APE) could significantly reduce the serum lipid index in model rats and improve the degree of liver pathological changes.²⁷ Another study demonstrated that ESW could raise plasma HDL-C/TC ratio and increase lecithin-cholesterol acyltransferase activity, decrease plasma HDL₃-C level and decelerate the progress of atherosclerosis to a certain degree.⁸³ In addition, ESW could also reduce the levels of blood sugar in type 2 diabetes rat models.⁸⁴ Apart from this, the substances in ESW have antibacterial activity.⁸⁵ Also, other extracts from ESW could enhance the production of polysaccharides,⁸⁶ ganoderic acid⁸⁷ and triterpenoids⁸⁸ in *Ganoderma lucidum*.

5 Summary and outlook

Tubiechong is an important insect medicine in TCM and has been extensively used to treat several illnesses. It has been used to treat bruises, fractures, amenorrhoea, postpartum blood stasis, lumps and relieving pain. In this review, we summarize the knowledge on traditional uses, chemical composition and pharmacological activities of such medicine. Our review has confirmed the diversity of its chemical composition and verified that the insects contained many active small molecular substances and macromolecular ingredients such as proteins, peptides and polysaccharides. However, gaps exist between the applications and scientific studies on Tubiechong. There is basically a lack of direct biological activity assays performed on single compounds isolated from Tubiechong. Most pharmacological studies employed crude extracts or isolated products of Tubiechong as testing samples. In a coincidence, chemical information and structural characterization of products of Tubiechong are relatively very limited as well. A lack of chemical analysis of the assessed extract is a relative insufficiency in current studies, which makes the main active ingredients to be unknown. The functional material basis is still unclear and on the consideration of its safety and effectiveness in clinical application and industrial developments. It is urgent to conduct deeper chemical-combined biological studies on the two insect medicines than before.

On the other hand, for traditional medicines, we need to give more attention to its traditional usage, including traditional

preparation methods and medical applications. Current investigations on Tubiechong were mostly focused on 70% or 95% alcohol extracts of the insect with little attention to aqueous extracts and low-concentration ethanol extracts, which were its main traditional preparation methods. Therefore, in order to ensure its efficacy, we are supposed to carry out research on Tubiechong based on following the traditional preparation methods. Understanding traditional medical applications of Tubiechong is also important for new drug discovery and development because it can make us easy to know the aim and therefore perform the pre-clinical and clinical investigations.

Tubiechong has multiple pharmacological actions and therapeutic behavior based on both traditional medicine and scientific practices, such as thrombolysis, anticoagulant, anti-tumor, immunomodulation, damage-repairing and hepatoprotection. However, pharmacological studies are performed only in animals or cell models. There is currently some suspicion regarding the data to support the therapeutic roles of Tubiechong in the clinic, and thus well-designed experiments and clinical trials are critical to confirm the safety and efficacy of the insect for human beings. Future integrated and profound studies on the chemical composition, pharmacological, and toxicological aspects of Tubiechong are essential to understand the therapeutic roles of such insect-based Chinese medicine.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

This work was mainly supported by The National Natural Science Foundation of China (no. 81673788, 81873136 (to C. L)).

References

- 1 G.-F. Ge, C.-H. Yu, B. Yu, Z.-H. Shen, D.-L. Zhang and Q.-F. Wu, *J. Ethnopharmacol.*, 2012, **141**, 178–182.
- 2 H. Liu, Y. Yan, F. Zhang and Q. Wu, *Immunol. Invest.*, 2019, **48**, 844–859.
- 3 F.-X. Wang, N. Wu, J.-T. Wei, J. Liu, J. Zhao, A.-g. Ji and X.-K. Lin, *Biochem. Cell Biol.*, 2013, **91**, 244–251.
- 4 B. Dai, Y. Zhan, J. Qi and Y. Zhang, *Environ. Toxicol. Pharmacol.*, 2014, **37**, 1177–1185.
- 5 B. Dai, J. Qi, R. Liu and Y. Zhang, *Mol. Med. Rep.*, 2014, **10**, 1590–1596.
- 6 X. Xie, W. Shen, Y. Zhou, L. Ma, D. Xu, J. Ding, L. He, B. Shen and C. Zhou, *Int. J. Biol. Macromol.*, 2020, **162**, 31–42.
- 7 Y. Zhan, H. Zhang, R. Liu, W. Wang, J. Qi and Y. Zhang, *Integr. Cancer Ther.*, 2016, **15**, 102–112.
- 8 L. Sun, Z. He, M. Zhao, R. He and Y. Feng, *Environ. Entomol.*, 2018, **40**, 23–29.
- 9 D.-Q. Zhang, Y. Xu, Y.-P. Mu, W. Liu and P. Liu, *Chin. J. Exp. Tradit. Med. Formulae*, 2018, **24**, 219–224.
- 10 Y.-H. Zhang, J.-T. Liu, B.-Y. Wen and N. Liu, *J. Ethnopharmacol.*, 2009, **124**, 125–129.



- 11 L.-J. Ju, P.-P. Hu, P. Chen, X. Xue, Z.-Q. Li, F.-Y. He, Z.-X. Qiu, J. Cheng and F. Huang, *Biomed. Pharmacother.*, 2020, **129**, 110471.
- 12 X.-F. Wang, *Chin. Herb. Med.*, 2019, **50**, 2224–2228.
- 13 L. J. Ni, N. N. Wang, L. G. Zhang, Y. Z. Guo and W. Z. Shi, *J. Ethnopharmacol.*, 2016, **179**, 420–431.
- 14 B. Xu, G. H. Zhu, J. Xia and S. H. Li, *J. Clin. Rehabil. Tissue Eng. Res.*, 2007, **11**, 2253–2256.
- 15 Y. Lu and P. Jiang, *Zhongguo Zhongyao Zazhi*, 1992, **17**, 487–489.
- 16 N. Zhang, Y. Zhao, Y. Shi, R. Chen, X. Fu and Y. Zhao, *Biomed. Pharmacother.*, 2019, **112**, 108636.
- 17 P. Cai, D. Wan, J. Xiao, S.-H. Zhang and G.-X. Cai, *Chin. Tradit. Pat. Med.*, 2011, **33**, 1645–1648.
- 18 Z. Dong-mei, L. I. Sui-jing and H. A. N. Yali, *Lishizhen Med. Mater. Med. Res.*, 2009, **20**, 778–779.
- 19 L. I. XingNuan and H. A. N. YaLi, *Acta Zool. Sin.*, 2007, **53**, 135–142.
- 20 H. Liu, Y.-L. Han, H. Ding and Z.-W. Li, *Lishizhen Med. Mater. Med. Res.*, 2010, **21**, 2140–2142.
- 21 H. Yang, Y. Wang, Y. Xiao, Y. Wang, J. Wu, C. Liu, H. Ye, F. Li, H. Yu and R. Lai, *PLoS One*, 2011, **6**, e17519.
- 22 S. Jiang, P.-P. Dong, H.-R. Li, J. Xu, H.-J. Li, Y.-Y. Yu, L. Dai, P. Gao, S.-P. Wang and J.-Y. Zhang, *China J. Chin. Mater. Med.*, 2020, **45**, 5265–5272.
- 23 M.-C. Wang, Y.-L. Jin and M.-Z. Piao, *J. Chin. Inst. Food Sci. Technol.*, 2012, **12**, 34–38.
- 24 Y.-L. Han and Z.-W. Li, *Chin. Med. Mater.*, 2006, **29**, 765–767.
- 25 Y.-L. Han and Z.-W. Li, *Chin. J. Biotechnol.*, 2006, **22**, 639–643.
- 26 S. M. Wang, X. L. Zhao, B. X. Wang, Q. L. Zhou, Z. Y. Liu, R. G. An, Z. Q. Liu and S. Y. Liu, *Chin. J. Anal. Chem.*, 2005, **33**, 1385–1388.
- 27 S.-P. Wang, S. Jiang, Y.-M. Zhao, Y. Lin, J.-Y. Zhang and L. Dai, *Chin. Pharmacol. Bull.*, 2020, **36**, 621–626.
- 28 Z.-C. Liu, K.-H. Yuan, R.-P. Zhang, X.-C. Ren, X.-L. Liu, S.-H. Zhao and D.-K. Wang, *J. Venomous Anim. Toxins*, 2016, **22**, DOI: 10.1186/s40409-016-0058-7.
- 29 Y. Lu and P. Jiang, *China J. Chin. Mater. Med.*, 1992, **17**, 487–512.
- 30 L. Liu, R. Jin and G. Xu, *China J. Chin. Mater. Med.*, 1999, **24**, 73–124.
- 31 Z. Chen, W.-T. Chen, W.-H. Luo and X.-L. Bi, *Chin. Tradit. Pat. Med.*, 2016, **38**, 1074–1077.
- 32 X.-L. Wang, Q. Li, B.-H. Li, Y. Hui, Y.-Y. Chen and K.-S. Bi, *Chin. Herb. Med.*, 2016, **47**, 1780–1784.
- 33 J.-S. Wang, Y. Nian, F.-L. Zhang and H. Li, *J. Beijing Univ. Tradit. Chin. Med.*, 2016, **39**, 850–854.
- 34 H. J. Zhu, T. Xu, Y. M. Yan, Z. C. Tu and Y. X. Cheng, *Nat. Prod. Res. Dev.*, 2020, **11**, 1–12.
- 35 H. L. Jiang, X. H. Luo, X. Z. Wang, J. L. Yang, X. J. Yao, P. Crews, F. A. Valeriote and Q. X. Wu, *Fitoterapia*, 2012, **83**, 1275–1280.
- 36 X. Q. Jin, M. M. Yan, E. X. Huang, Y. J. Xu, Y. J. Gu, D. B. Cui, S. Y. Lin and D. M. Xu, *China J. Chin. Mater. Med.*, 1993, **18**, 355–356.
- 37 H.-L. Jiang, X.-H. Luo, X.-Z. Wang, J.-L. Yang, X.-J. Yao, P. Crews, F. A. Valeriote and Q.-X. Wu, *Fitoterapia*, 2012, **83**, 1275–1280.
- 38 Y. Li, J. Hong, H. Li, X. Qi, Y. Guo, M. Han and X. Wang, *Drug Delivery*, 2017, **24**, 1491–1500.
- 39 Y. Bao, Y. W. Sun, J. Ji, L. Gan, C. F. Zhang, C. Z. Wang and C. S. Yuan, *Phytomedicine*, 2019, **63**, 153036.
- 40 Y. Gao, F. Liu, L. Fang, R. Cai, C. Zong and Y. Qi, *PLoS One*, 2014, **9**, e96741.
- 41 X. Wang, Z. J. Song, X. He, R. Q. Zhang, C. F. Zhang, F. Li, C. Z. Wang and C. S. Yuan, *Int. Immunopharmacol.*, 2015, **29**, 701–707.
- 42 A. Hematpoor, M. Paydar, S. Y. Liew, Y. Sivasothy, N. Mohebbali, C. Y. Looi, W. F. Wong, M. S. Azirun and K. Awang, *Chem.-Biol. Interact.*, 2018, **279**, 210–218.
- 43 Z. Y. Cheng, G. D. Yao, R. Guo, X. X. Huang and S. J. Song, *Bioorg. Med. Chem. Lett.*, 2017, **27**, 597–601.
- 44 H. J. Zhu, T. Xu, Y. M. Yan and Y. X. Cheng, *Bioorg. Chem.*, 2020, **104**, 104258.
- 45 H. J. Zhu, Y. M. Yan, Z. C. Tu, J. F. Luo, R. Liang, T. H. Yang, Y. X. Cheng and S. M. Wang, *Fitoterapia*, 2016, **114**, 163–167.
- 46 H. E. Colley, M. Muthana, S. J. Danson, L. V. Jackson, M. L. Brett, J. Harrison, S. F. Coole, D. P. Mason, L. R. Jennings, M. Wong, V. Tulasi, D. Norman, P. M. Lockey, L. Williams, A. G. Dossetter, E. J. Griffen and M. J. Thompson, *J. Med. Chem.*, 2015, **58**, 9309–9333.
- 47 M. J. Thompson, J. C. Louth, S. Ferrara, F. J. Sorrell, B. J. Irving, E. J. Cochrane, A. J. Meijer and B. Chen, *ChemMedChem*, 2011, **6**, 115–130.
- 48 D. A. Negatu, M. Gengenbacher, V. Dartois and T. Dick, *Front. Microbiol.*, 2020, **11**, 575586.
- 49 E. E. Alexeev, J. M. Lanis, D. J. Kao, E. L. Campbell, C. J. Kelly, K. D. Battista, M. E. Gerich, B. R. Jenkins, S. T. Walk, D. J. Kominsky and S. P. Colgan, *Am. J. Pathol.*, 2018, **188**, 1183–1194.
- 50 Z. H. Zhao, F. Z. Xin, Y. Xue, Z. Hu, Y. Han, F. Ma, D. Zhou, X. L. Liu, A. Cui, Z. Liu, Y. Liu, J. Gao, Q. Pan, Y. Li and J. G. Fan, *Exp. Mol. Med.*, 2019, **51**, 1–14.
- 51 J.-F. Chen, C. Yang and Y. Zhang, *Chin. J. Spectrosc. Lab.*, 2010, **27**, 2009–2011.
- 52 Y.-F. Yang, *Chin. Med. Mater.*, 2002, **25**, 150–152.
- 53 B. Dai, Y. Zhan, J. Qi and Y. Zhang, *Environ. Toxicol. Pharmacol.*, 2014, **37**, 1177–1185.
- 54 Y. Zhang, Y. Zhan, D. Zhang, B. Dai, W. Ma, J. Qi, R. Liu and L. He, *Sci. Rep.*, 2014, **4**, 5518.
- 55 Y. Zhang, J. Yu, H. Luo and R.-H. Dai, *Nat. Prod. Res. Dev.*, 2020, **32**, 1051–1056.
- 56 G. Ge, C. Yu and Q. Wu, *China J. Tradit. Chin. Med. Pharm.*, 2013, **28**, 826–828.
- 57 Z. Lin, Y. Han and B. Chen, *Shizhen Guoyi Guoyao*, 2013, **24**, 807–810.
- 58 F.-C. Cao, Y.-l. Han, B. Chen, M.-X. Huang, H. Ding, L. Yu and Y. Ye, *Lishizhen Med. Mater. Med. Res.*, 2011, **22**, 1827–1829.
- 59 W. Zhang, X. Zou, X.-P. Qian, L.-X. Yu and B.-R. Liu, *Tradit. Chin. New Drug Res. Clin. Pharmacol.*, 2007, **18**, 257–259.



- 60 X.-N. Li and Y. Han, *Pharmacol. Clin. Chin. Mater. Med.*, 2008, **24**, 44–46.
- 61 Q.-F. Tang, Y. Dai and X.-L. Liu, *Chin. Bull. Entomol.*, 2011, **48**, 156–159.
- 62 D. Liu, X.-N. Li, Z.-J. Qin, W. He and Y. Zhao, *Chin. Med. Mater.*, 2012, **35**, 1382–1385.
- 63 D. Liu, S. Cao, Y. Liu, J. Liu, Z. Wang, Y.-H. Zhang and H. Shen, *Acta Lab. Anim. Sci. Sin.*, 2020, **28**, 73–80.
- 64 M.-Z. Piao, M.-C. Wang and X.-D. Wang, *Food Sci.*, 2013, **34**, 242–245.
- 65 C.-G. Gu, Y.-H. Zhang, R.-R. Bai, M.-J. Tian and H. Shen, *Acta Lab. Anim. Sci. Sin.*, 2014, **22**, 66–74.
- 66 F.-C. Cao, Y.-L. Han, B. Chen, H. Liu, H. Ding, M.-X. Huang and Y. Ye, *Chin. Med. Mater.*, 2011, **34**, 676–679.
- 67 Q.-F. Tang, Y. Dai and X.-L. Liu, *Afr. J. Biotechnol.*, 2010, **9**, 8682–8686.
- 68 Q. Tang, Y. Dai and X. Liu, *Chin. J. Appl. Entomol.*, 2011, **48**, 156–159.
- 69 D. Liu, X. Li, Z. Qin, W. He and Y. Zhao, *J. Chin. Med. Mater.*, 2012, **35**, 1382–1385.
- 70 Y.-F. Yang, S.-Q. Wang, M.-J. Feng, A.-H. Ning and C.-H. Huang, *Chin. J. Cell. Mol. Immunol.*, 2005, **21**, 53–56.
- 71 Y.-F. Yang, M.-S. Peng and Y.-W. Yang, *Chin. J. Immunol.*, 2003, **19**, 686–689.
- 72 F. Xie, X. Li, K. Sun, Y. Chu, H. Cao, N. Chen, W. Wang, M. Liu, W. Liu and D. Mao, *J. Tradit. Chin. Med.*, 2001, **21**, 225–231.
- 73 H. B. Cai, X. G. Sun, Z. F. Liu, Y. W. Liu, J. Tang, Q. Liu, B. M. Ji, Y. H. Song, Y. C. Zhou, M. H. Yang and Z. P. Lv, *J. Ethnopharmacol.*, 2010, **132**, 157–164.
- 74 Y.-F. Deng, L. Qu, Y. Song and Y. Jiang, *J. Beijing Univ. Tradit. Chin. Med.*, 2015, **38**, 420–425.
- 75 W.-C. Zhong, C.-Y. Zhou, L. Gao, Z.-P. Lu and S.-H. Huang, *Chin. Tradit. Pat. Med.*, 2017, **39**, 2475–2480.
- 76 M.-R. Xie, Q.-M. Li, S.-Q. Qiu, X.-L. Qi and H. Shen, *Acta Zoonutr. Sin.*, 2018, **30**, 1726–1735.
- 77 M.-R. Xie, S.-Q. Qiu and H. Shen, *Acta Lab. Anim. Sci. Sin.*, 2016, **24**, 427–431.
- 78 M.-Z. Piao, J. Ning and F.-W. Wang, *Sci. Technol. Food Ind.*, 2010, **31**, 205–208.
- 79 G.-L. Wu, T.-Y. Li and Y.-S. Fan, *China J. Chin. Mater. Med.*, 2019, **44**, 845–848.
- 80 Z.-X. Qi, S.-Q. Li and T. Yu, *J. Clin. Rehabil. Tissue Eng. Res.*, 2013, **17**, 2597–2602.
- 81 X.-J. Bai, H.-J. Ren, Z.-R. Luo, H. Wang, Z.-Z. Zhao and X.-B. Leng, *J. Northeast Agric. Univ.*, 2014, **45**, 71–75.
- 82 Z. Zhang, E.-Z. Jiang, F.-Y. Ning, Z.-H. Du and X.-J. Bai, *J. Northeast Agric. Univ.*, 2018, **49**, 52–58.
- 83 W. Wang, J. Wang, D. Zhao, H. Liu, W. Zhou and K. Chen, *China J. Chin. Mater. Med.*, 1991, **16**, 299–320.
- 84 Z. Jing and G.-X. Xu, *J. Fourth Mil. Med. Univ.*, 2009, **30**, 465–467.
- 85 X.-M. Suo, X.-J. Lu, J.-Z. Dong, J. Li, P. Song, R. Zhao and L.-Y. Fu, *Chin. J. Biol. Control*, 2007, **23**, 64–67.
- 86 G.-Q. Liu and K.-C. Zhang, *Appl. Biochem. Biotechnol.*, 2007, **74**, 572–577.
- 87 G.-Q. Liu, X.-L. Wang, Y.-G. Zhang, Y.-H. Wu, W.-J. Han and H.-Y. Zhang, *Afr. J. Biotechnol.*, 2010, **9**, 6129–6134.
- 88 G.-Q. Liu, H.-X. Xiao, X.-L. Wang, Y. Zhao, Y.-G. Zhang and G.-P. Ren, *Appl. Biochem. Biotechnol.*, 2011, **165**, 87–97.

