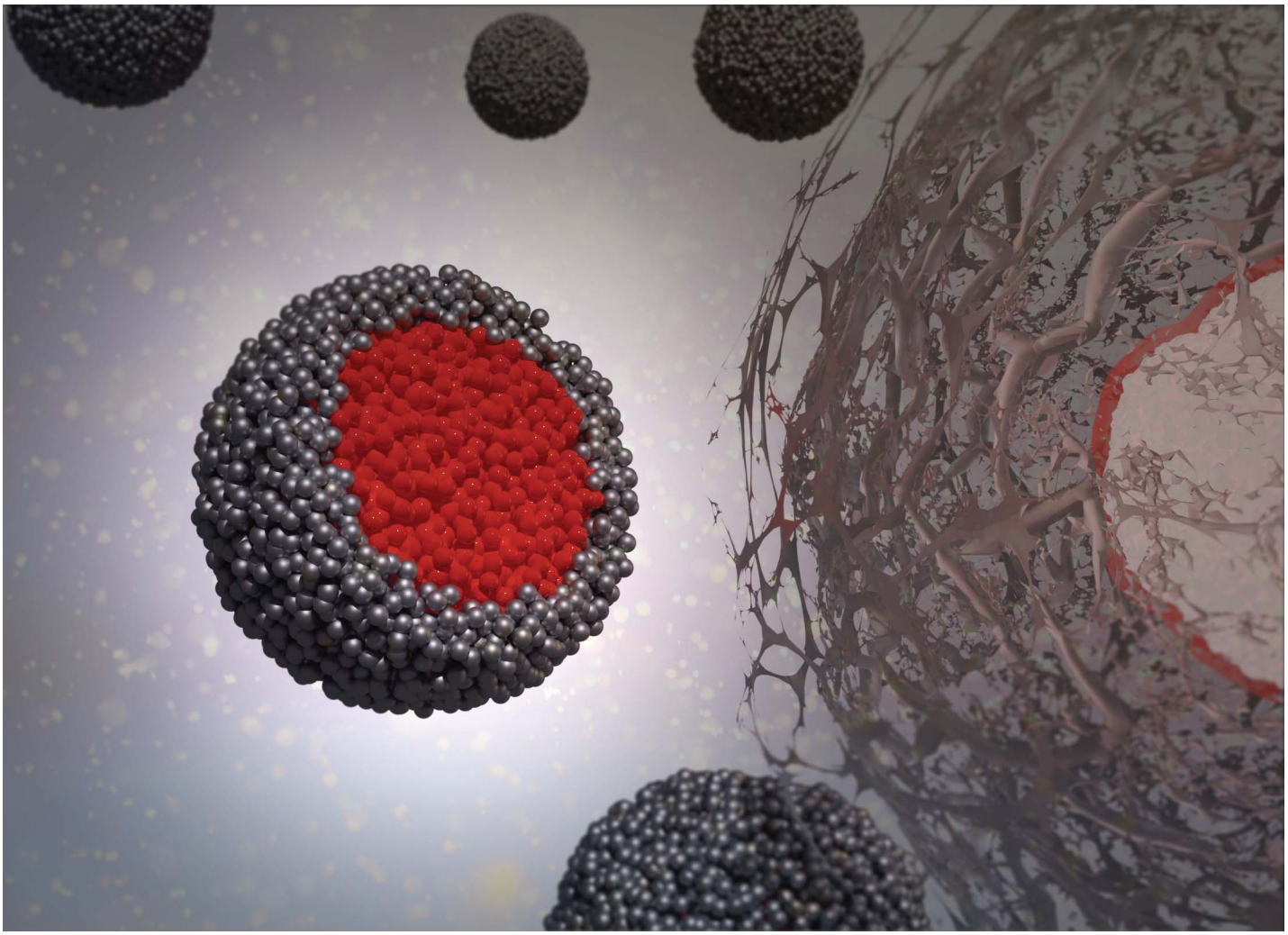

Showcasing research from Assistant Professor Asma Ghazzy's laboratory, Pharmacological and Diagnostic Research Centre, Faculty of Pharmacy, Al-Ahliyya Amman University, Amman, Jordan.

Magnetic iron oxide-based nanozymes: from synthesis to application

This comprehensive review article provides a thorough overview of Iron Oxide Nanozymes (IONzymes), magnetic nanoparticles that mimic natural enzyme activities. Spotting their remarkable stability, magnetic properties, and biocatalytic capabilities, moreover, the article demonstrates various synthesis methods, including chemical, physical, and biological processes. This review also discusses the current applications of IONzymes in biomedicine, environmental fields, and the potential promising applications.

As featured in:

See Asma Ghazzy *et al.*, *Nanoscale Adv.*, 2024, **6**, 1611.

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Magnetic iron oxide-based nanozymes: from synthesis to application

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Iron oxide nanozymes (IONzymes) are a class of magnetic nanoparticles that mimic the enzymatic activity of natural enzymes. These particles have received significant attention in recent years due to their unique properties, such as high stability, tunable magnetic responsiveness, and ability to act as biocatalysts for various chemical reactions. In this review, we aim to provide an overview of the production methods of magnetic nanozymes, including chemical, physical, and biological synthesis. The structure and design of magnetic nanozymes are also discussed in detail, as well as their applications in various fields such as biomedicine and environmental science. The results of various studies and the latest advances in the field of magnetic nanozymes are also discussed. This review provides valuable insights into the current state of magnetic nanozymes and highlights their potential for further development and application in various fields.

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1. Introduction

The advancement of nanotechnology over the past two decades has highlighted new prospects in a wide range of industries due to the extraordinary qualities and distinct structure of nanomaterials.[1,2] The structure of nanomaterials is divided into three layers (surface, shell, and core) where functional groups, including metal ions, tiny compounds, surfactants, and polymers, distinguish one layer from another.[3,4] In general, the core is the nanoparticles (NPs), which can bond with various structures and macromolecules such as composites, metal organic frameworks (MOFs), polymers, and carbon nanotubes. This diversity enhances their unique properties in terms of size, shape, composition, and structural framework, which require optimization through synthesis procedures.[5–8]

Metallic nanoparticles represent a corner stone in the preparation of nanomaterials.[9,10] Various metal oxides such as FeO,

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Hamdi Nsairat

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NiO, ZnO, CuO, AgO, TiO, SnO, and WO have unlimited applications in the medical sector (drug delivery, cancer treatment, and tissue repair), environment (qualitative and quantitative analysis of pollutants and toxins, water purification, and photodegradation), energy nanogenerators, electronics, catalysis, and mechanical and textile industries.[11–17]

Iron oxide is one of the best biocompatible inorganic nanoparticles, and it has remarkable microscopic physical properties including superparamagnetism, low susceptibility to oxidation, firmness in liquid solution, extended blood half-life, and flexible surface chemistry.[18–22] Also, from an application point view, iron oxide NPs have high sustainability and superior properties in comparison to natural substances such as enzymes, which have drawbacks including high cost of isolation and purification, limited thermostability, and small pH window, which disrupts the enzyme activity upon handling, storage, and transportation.[23,24]

Artificial enzymes have replaced real enzymes in many applications for decades due to their stability and low cost. Metal complexes, cyclodextrins, polymers, dendrimers, and biomolecules have been studied to replicate enzyme activities and structures. Due to the rapid advancement of nano-studies and

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Dr Rana Said is an Associate Professor at Al-Ahliyya Amman University. She is specialist in sample preparation methods for the measurement of drugs and metabolites in biological samples to provide information that plays an important role in toxicokinetic and pharmacokinetic studies, and in therapeutic drug monitoring (TDM). She was always curious about drugs and how they work in the human body. She loved her chemistry and biological classes at school given that they were related to understanding more about drugs. She worked at a pharmaceutical company for a short time, and she travelled to the UK to attend a Master's Pharmaceutical Program, and at the end, she did her research project at one of the biggest pharmaceutical companies in the UK, AstraZeneca. Subsequently, she left for Sweden and joined a pharmacology department to get a PhD in developing new drugs analysis methods for pharmacokinetic studies. It was not easy to do this considering that this university is ranked number 4 in the world and a lot of work was required.

Obada A. Sibai

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Aseel AbuRuman

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Alaa S. Shraim

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the exceptional properties of nanomaterials, several nanomaterials have shown enzyme-like functions. Moreover, nanozymes are popular because of their ease of manufacture, storage, isolation, and exceptional outcomes. In this respect, IONzymes can be used effectively to mimic natural enzymes and applied in several environmental applications, such as degradation of antibiotics and adsorption of dyes, in the food industry and biomedical, biosensing, cosmetics, and bioengineering.[25–32]

In this review, we highlight the methods for the synthesis of IONzymes and the current advances in the development of their applications. We discuss several nanomaterials that have been studied to imitate various types of enzymes in order to highlight the advancement in the area of nanomaterial-based artificial enzymes. We discuss their synthetic methods, processes, and applications in several domains, such as biosensing and immunoassays, as well as pollution elimination. We also outline techniques, such as several green, chemical, and physical methods, to produce iron oxide nanozymes.

2. Synthesis approaches of IONzymes

The methods commonly used to produce metal oxide nanoparticles are often applied in the creation of IONzymes, especially when they consist primarily of two magnetic nanoparticles, namely, magnetite (Fe_3O_4) and maghemite (γ-Fe_2O_3).[33]

The synthesis of IONzymes is accomplished using different chemical, physical, and biological techniques. Co-precipitation, evaporative decomposition of solution (EDS), aerosol, ultrasonic, sol–gel synthesis, micro-emulsion methods, reverse micelles, flow injection, solid-state reaction, spraying, and hydrothermal/solvothermal processes are typically used in chemical synthesis.[34] The physical methods include milling, grinding, pyrolysis, and thermal ablation, as illustrated in Fig. 1. Also, the "Green Approach" has recently attracted significant consideration due to its eco-friendly nature and sustainability, which can be conducted using algae, bacteria, fungi, and plants.

It is critical to distinguish between the general synthesis of iron oxide nanoparticles and specific processes that provide the characteristics of an enzyme in the production of IONzymes. Several conventional methods are effective in generating iron oxide nanoparticles, including co-precipitation, thermal breakdown, and hydrothermal synthesis. These steps must be performed to expose the properties of nanozymes. For instance, surface functionalization is essential to provide the ability to use enzymes.

Gao *et al.* (2007) demonstrated that the peroxidase-like activity of iron oxide nanoparticles can be significantly increased by adding specific functional groups to their surface. Both the size and structure of nanoparticles play a key role in determining their enzymatic activity.[35] Another study demonstrated that an increase in the surface area to volume ratio of smaller iron oxide nanoparticles leads to higher catalytic efficacy. This characteristic resembles the active regions of natural enzymes.[36] In addition, the crystalline structure of IONzymes influences their catalytic activity. Wei and co-workers showed that the intrinsic catalase-like, oxidase, and peroxidase activities of magnetite (Fe_3O_4) nanoparticles are attributed to their spinel structure.[37]

The synthesis method influences the stability and specificity of IONzymes prior to catalytic activity. Research in a related study indicated that the value of adding stabilizing chemicals during the synthesis of IONzymes can improve the lifespan of their catalytic activity and their thermal stability.[38] This study emphasized that the functional characteristics of nanozymes are stable with time, stressing the importance of stabilization in the synthesis process.

Additionally, the effectiveness and selectivity of nanozymes can be altered by doping them with different metals towards particular substrates, hence expanding their applicative potential, as demonstrated by Zhang *et al.*[39] These features demonstrate the significance of carefully choosing the methods for the synthesis of IONzymes to induce enzymatic activity and particle formation. This approach is consistent with the principles of biomimetic design given that it features both the functional and physical attributes of the nanoparticles. This illustrates the intricate nature of enzymes found in biological systems.

2.1. Chemical synthesis

The chemical production of nanoparticles is the most typical technique. However, the key challenges in this type of procedure include particle dispersion, clumping, and size uniformity. Additionally, chemical-based procedures involve the use of solvents such potassium bitartrate, sodium dodecyl sulfate, sodium borohydride, and hydrazine, all of which are detrimental to the environment given that they produce unpleasant waste flows. Herein, we focus on the four most popular methods, as listed in Table 1.

Afnan Al hunaiti

Dr Afnan Al-Hunaiti, an Associate Professor at the University of Jordan, is a seasoned academic with a comprehensive background in chemistry and catalysis. She earned her PhD from the University of Helsinki in 2015, focusing on the oxidation of fine chemicals through iron-based and metal-free catalysis. With over a decade of experience, Dr Afnan has held positions at the University of Petra and conducted research at the University of Helsinki. Dr Afnan's dedication extends to impactful publications in peer-reviewed journals, showcasing her commitment to advancing scientific knowledge in fields such as environmental inorganic chemistry and atmospheric catalysis. As an Associate Professor, Dr Afnan's multifaceted contributions underscore her significant role in advancing the understanding in the fields of catalysis and environmental science, making her a respected figure in the academic community.

2.1.1. Co-precipitation. Massart developed a chemical co-precipitation approach for the large-scale synthesis of hydrophilic IONzymes.[44] This reaction is performed in aqueous solution; therefore, the product is water-dispersed and may be directly employed for diverse applications without complicated ligand exchange procedures. The co-precipitation procedure to manufacture Fe_3O_4 involves the hydrolysis and condensation of ferrous and ferric ions in aqueous solution in the pH range of 8–14, as shown in eqn (1).[18]

$$Fe^{2+} + 2Fe^{3+} + 8OH^- \rightarrow Fe_3O_4 + 4H_2O \quad (1)$$

Co-precipitation relies on the pH, reaction temperature, ion concentration, ionic strength, salt type, and alkali used. However, the application of co-precipitation to create magnetite NPs is a challenging process, and the reaction conditions must be tightly regulated.[45,46]

Another factor is the molar ratio of ferrous/ferric, which affects the physical and magnetic properties of NPs. When 1 ferrous : 1 ferric is used, there is larger magnetization saturation than other ratios.[47,48] Similar research has been conducted to create various synthesis techniques employing the chemical co-precipitation process to produce stable, homogeneous, smaller-sized, crystalline particles.[49]

A method involving co-precipitation in flow chemistry, combined with an *in situ* synchrotron X-ray diffraction (XRD) technique, was devised to "freeze" the transient reaction states through steady-state operation. This technique showed appealing findings, as follows:

(i) Five seconds after mixing, the only crystalline phase was the inverse spinel framework of magnetite/maghemite.

(ii) The particle size increased slightly, and solid phase development (owing to particle growth) was completed within 2 min.

(iii) The mixing conditions did not affect the XRD pattern.

(iv) During co-precipitation, the diffraction peaks widened, indicating the presence of smaller coherently scattered regions (Fig. 2).[50,51]

The co-precipitation chemical technique can be used to produce functional materials, as shown in Table 2. Chen *et al.* discovered a new co-precipitation approach to generate ferumoxytol, a therapeutically relevant magnetic nanoparticle with

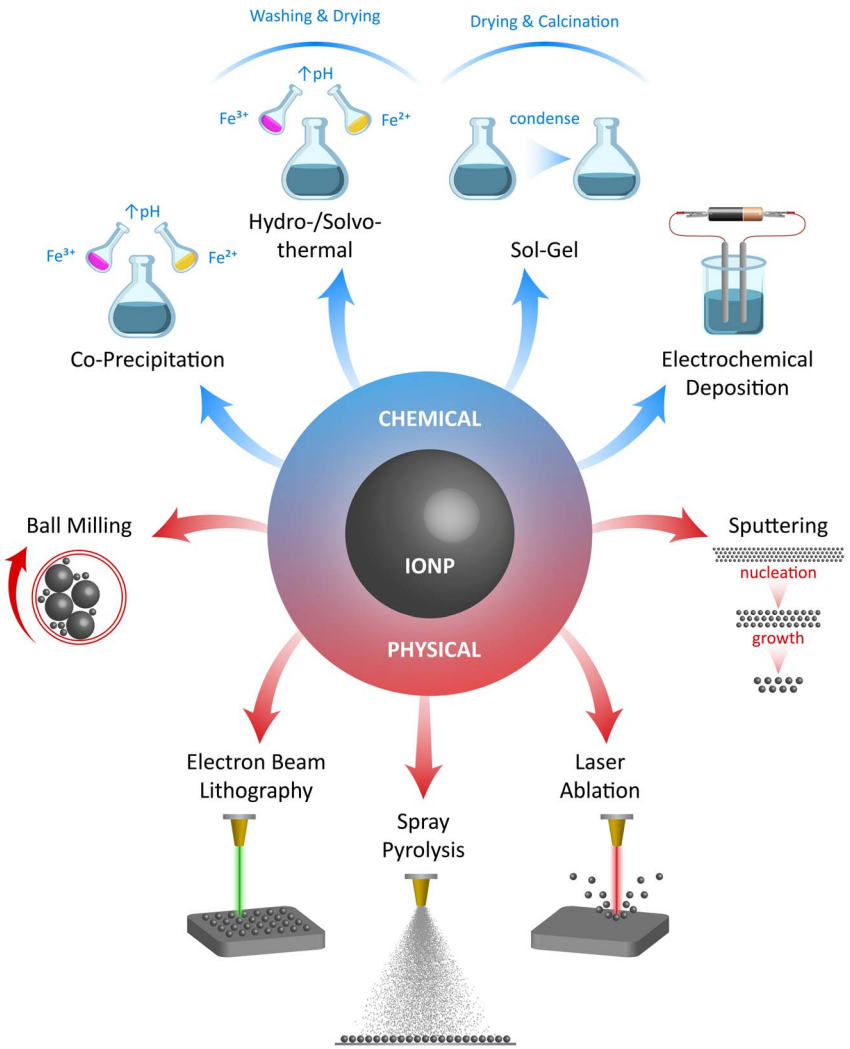

Fig. 1 Chemical and physical techniques for the synthesis of IONzymes.

Table 1 The most common chemical techniques employed for the synthesis of IONzymes

Synthetic technique	Advantage	Disadvantage	Ref.
Co-precipitation	Water-dispersed Environmentally friendly Efficient and economical Simple procedure Rapid particle formation	Multi-variable dependence Toxic liquid waste Requires trained person for maintenance and regeneration Toxic liquid waste	40
Hydrothermal/ solvothermal	Producing highly crystalline nanocrystals Well-controlled dimensions Combined with microwaves and magnetic fields improves reproducibility and quality	High temperature and pressure range Anti-corrosion autoclave material Relatively costly reactors Mass production is not possible	41
Electrochemical deposition	Short formation time Simple apparatus Uniformly coated on complicated geometries Control of film thickness and morphology	Stable solvent media High electrophoretic mobility	42
Sol–gel synthesis	Simplicity of the process Uniform composition and high purity High production efficiency Production of intricately shaped optical components Controlling homogeneous products Capacity to use the product with unique structures such as fibers and aerogels	Wear resistance reduced Weak bonding strength Hard to regulate porosity and permeability	43

γ-Fe_2O_3 as the core. The magnetization of ferumoxytol is the greatest recorded to date, reaching 104–105 emu g^{-1}, and its crystal structure has been substantially improved.[52] Superparamagnetic IONzymes were produced with a limited size distribution, and their magnetic susceptibility, coercivity, remanence, and saturation magnetization at 5–300 K were analyzed.[53]

2.1.2. Hydrothermal and solvothermal. The technique employed for the synthesis of IONzymes can alter the primary properties of the generated IONzymes. The solvothermal and hydrothermal processes are the most effective chemical ways to create nanomaterials, specifically nanocrystals with precise dimension control.[55] The suggested process begins with the formation of nuclei from the solute molecules, which subsequently undergo significant growth during heating, leading to the formation of the final crystal structure (Fig. 3). The reaction rate increases together with crystallinity.[56] Highly crystalline iron oxide nanoparticles with a size in the range of 14 and 25 nm were produced in a pressure-resistant reactor at 473 K.[57] However, this method requires costly reactors.[58]

Many advances have contributed to a deeper understanding and improved this technique (Table 3). A novel strategy was

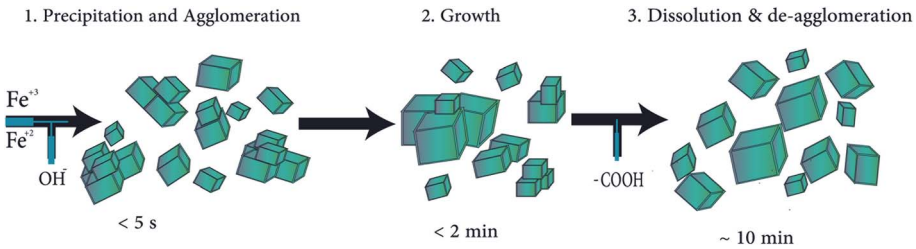

Fig. 2 Suggested particle mechanism during the co-precipitation procedure: (1) particles are precipitated and agglomerated within 5 s, (2) agglomerated pieces grow over the next 2 or 3 min and (3) addition of neutralization solution causes particles to de-agglomerate within 10 min.

Table 2 Representative iron oxides obtained through co-precipitation procedures

Compound/property	Particle size (nm)	Morphology	Magnetization (emu g^{-1})	Ref.
Ferumoxytol	7.1	Spherical	104–105	52
Temperature-dependent particles	11.22	Spherical	64–72	53
(Zn–Mn)-co-doped	10–13	Spherical	81	54

demonstrated to control the carbon chain length of the iron(III) carboxylate precursors and the amount of reaction solvent in the solvothermal synthesis of FeO nanocrystals.[59] Additionally, deep eutectic solvents with hydrated mixtures have been applied to solvothermal approaches for the preparation of functional nanomaterials.[51] A study presented the first *in situ* and static structural analysis of the production of iron oxide (hematite) nanoparticles in a deep eutectic solvent (DES) of choline chloride : urea.[60] González-Rivera *et al.* created a quick and easy approach. By directly applying microwave radiation in the solvothermal reactor with the aid of a coaxial antenna, the synthesis was thermally initiated, accelerated, and controlled. This method was perfectly regulated in short synthesis periods utilizing the phosphorylated nanoreactor.[61]

2.1.3. Electrochemical deposition. The electrophoretic deposition (EPD) technique involves the use of charged particles that move and are deposited on the surface of a conductive electrode to create thin or thick coatings and films. A broad range of fine powder, composite particles, colloidal metals, and ceramics can be produced *via* EPD.[64] EPD is one of several solution methods employing colloidal NPs, which has recently emerged as a successful method for producing dense and durable NP films. The relationship in EPD systems between colloidal NPs and the organic solvent has been studied using hexane, toluene, and chloroform in various solvent ratios to examine the charge formation function of the solvent in EPD systems (10 : 0, 7 : 3, 5 : 5, 3 : 7, and 0 : 10). The NP layer gets thicker and rougher as the toluene to hexane ratio increases. Alternatively, the film thickness is dramatically reduced when the chloroform to hexane ratio increases.[65]

The electrophoretic deposition approach was used to produce a bioactive coating, such as hydroxyapatite-iron oxide-chitosan (HA-CS) with varying amounts of Fe_3O_4 (1, 3, and 5 wt%) and porous morphology.[13] Another methodology demonstrated potential for generating thick magnetic nanocomposites for on-chip power components by incorporating iron oxide nanoparticles into a mold, and subsequently performing electro-infiltration of nickel through the porous film. The resulting magnetic saturation of the nanocomposites was measured at 473 kA m^{-1}, which is intermediate between the magnetic saturation values of iron oxide nanoparticles and nickel.[66] Also, the formulated and cost-effective coating method can enhance the surface characteristics and hemocompatibility for biomedical applications, resulting in decreased contact angle values and hydrophilic nature. In one study, the Ti–13Nb–13Zr alloy was electrophoretically coated with Bioglass (BG),

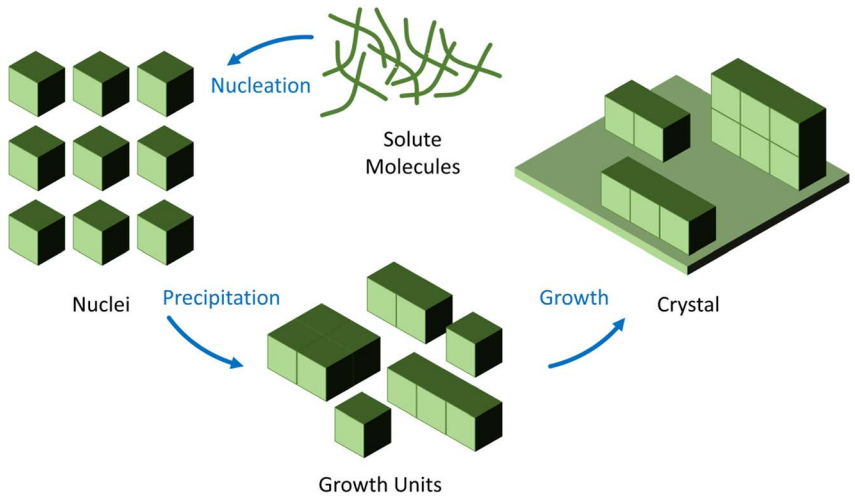

Fig. 3 Schematic representation of crystal growth mechanisms under hydrothermal/solvothermal conditions.

Table 3 Representative iron oxide nanozymes obtained through advanced hydrothermal/solvothermal procedures

Variable/technique	Starting materials	Solvent	Particle size (nm)	Morphology	Ref.
Deep eutectic-solvothermal	Iron(III) nitrate nonahydrate	1 : 2 : 10 choline chloride : urea : water	5–9	Oblate spheroid	51 and 60
Size-controlled facile solvothermal method	$FeCl_3 \cdot 6H_2O$ NaAc polyvinyl pyrrolidone (PVP)	Ethylene glycol (EG) diethylene glycol (DEG)	23	Spherical	33
Ligands and solvent composition	$FeCl_3 \cdot 6H_2O$ sodium carboxylate	2 : 1 water : ethanol	25	Cubic	59
Oxidation-precipitation solvothermal process	$FeCl_2 \cdot 4H_2O$	Deionized water	33	Spherical	62
Microwave solvothermal treatment	$FeCl_3 \cdot 6H_2O$ sodium citrate	Ethylene glycol (EG)		Irregular	63
Magnetothermally-responsive nanocarriers	$FeCl_2 \cdot 4H_2O$	Ethylene glycol (EG) urea	50 nm diameter, 250 nm length	Tubular shape	61

hydroxyapatite (HA), and iron oxide particles (FeO), which improved the stability of the suspension.[67]

2.1.4. Sol–gel synthesis. Using the sol–gel method, a gel-like network is generated, incorporating both liquid and solid phases. Also, by selecting the appropriate complexing agent, concentration, type of chemical additives, and temperature settings, it is possible to control the crystallinity, shape, and magnetic characteristics of IONzymes.[68,69] Apparently, the annealing temperature plays a central role in this method, and the outcome shows that the crystalline Fe_2O_3 nanoparticles[70] and the dielectric properties are enhanced.[71] Additionally, this technique can be used to create products with efficient physical characteristics, such as low UV absorption and thermal expansion coefficient and high optical transparency.[72]

The crystalline structure, composition, purity, magnetism, and morphology of iron oxide nanomaterials can be enhanced by optimizing some variables or combining techniques (Table 4). One technique is optimizing the precursor-to-solvent (P/S) ratio for three iron oxide phases (α-Fe_2O_3, Fe_3O_4, and γ-Fe_2O_3) to tune the structural and magnetic properties *via* sol–gel synthesis.[73] Another method combines microwave radiation and aluminum doping in iron oxide thin films, which controls the structural transitions of the iron oxide thin film. A study demonstrated a γ-Fe_2O_3 to Fe_3O_4 transition at 6–10 wt% Al with increasing saturation magnetization of the films from 251.3 emu cm^{-3} to 405.6 emu cm^{-3}.[74]

2.2. Physical genesis

Physical techniques such as mechanical milling, grinding, and thermal ablation are all expensive given that they consume a significant amount of energy. Furthermore, another significant drawback of this strategy is its exceedingly low output yield.

2.2.1. Ball milling. Ball milling is a shear-force-dominated method, which is also known as mechanical alloying, ultrafine grinding, and nanosizing in the literature. It is one of the most widely used industrial processes, in which the particle size is continuously reduced by impact and attrition. Metal balls, often made of zirconia (ZrO_2) or steel, serve as the grinding medium, while a spinning shell generates centrifugal force. By regulating the milling variables, such as ball-to-powder ratio (B/P), milling time, milling rpm, starting weight, and ball diameter, the excessive compression force that may harm the crystalline characteristics of nanomaterials can be reduced. Table 5 demonstrates examples of two states of milling that can be initiated, *i.e.*, dry ball milling (DBM) and wet ball milling (WBM). The solid-state mechanical size reduction process known as ball milling transforms iron precursors into MNPs (magnetic nanoparticles). To speed up milling and prevent the agglomeration of the created nanoparticles, solvents or excess salt can be added.[78]

Also, different mechanical ball milling techniques can be applied, such as conventional ball milling. Specifically, larger particles collide with steel balls or the interior wall of the tank to produce ultrafine particles, while high-energy ball milling uses a specialized grinding machine to synthesize a nano-spinel-type ferrite by mechanically alloying the initial materials.[79] However, the significant drawbacks of ball milling are pollution of the steel ball, the potential chemical and mechanical amorphization of the crystals, the high power used, and the prolonged milling period.[79,80]

2.2.2. Electron beam lithography. The use of electron beam lithography or electron beam deposition to apply either an exposure-sensitive resist material or high-purity iron material to a substrate can be employed of the synthesis of MNPs. This process produces MNPs by evaporating the first iron precursors onto the resist pattern, and then removing the resist through a lift-off procedure. Alternatively, the nanopatterns can be etched onto a functional substrate to produce MNPs.[78,84]

2.2.3. Laser ablation. Laser ablation is a method that involves irradiating a solid material placed under a thin layer with a laser beam.[80,85] When the solid material is placed at the bottom of a cell containing a liquid,[86] the technique is referred

Table 4 Representative iron oxide nanozymes obtained through sol–gel procedures

Variable/technique	Starting materials	Solvent	Particle size (nm)	Morphology	Additional property	Ref.
Agitation time	Fe $(NO_3)_3 \cdot 9H_2O$ Ba $(NO_3)_2$	Absolute ethanol	10 nm	Spherically	Purity > 75%	75
Carbonization method	$FeCl_3 \cdot 6H_2O$/ rosin	Deoxygenated H_2O/ ethanol	<50 nm	Varies with different rosins: $FeCl_3$	Enhances interfacial reactivity	76
Non-hydrolytic	Anhydrous $FeCl_3$	Anhydrous ethanol	202–373 Å	Rod-shaped	Homogeneous dispersion	77

Table 5 Iron oxide-based nanomaterials prepared *via* ball milling process

Milling process	Equipment/ball properties	Milling agent/solvents and conditions	Characteristics	Ref.
WBM	Planetary ball mill	DI water, 4 h, 500 rpm	High adsorption capacities of Cr(IV) $q_e = 48.1$ mg g^{-1}	34
DBM	Iron balls with 1.5 cm diameter	30 h and 90 h	31.48 emu g^{-1} and 37.80 emu g^{-1}	81
DBM	Steel balls with 8 mm diameter	25 rpm, 60 min	Particle size = 45 nm	82
DBM	Planetary ball mill	30 min, 320 rpm	$M_s = 20.45$ emu g^{-1}	83

to as "laser ablation synthesis in solution (LASiS)". In this case, various lasers can be employed, including Nd:YAG, Ti:sapphire, and copper vapor lasers,[80,87] allowing precise control of the phase composition, size, and shape of the particles, thus producing nanoparticles with an average diameter of approximately 15 nm.[87] Although laser ablation can quickly generate MNPs when exposed to a laser for short periods, this method has a low production rate.[88] Also, prolonged laser ablation leads to the formation of an excessive number of nanoparticles, which remain suspended in the colloidal solution and obstruct the laser beam. This causes the laser energy to be absorbed by the previously formed nanoparticles, rather than the target surface, resulting in a decreased ablation rate.[80]

2.2.4. Sputtering. Sputtering is a process that entails bombarding the surface of a bulk material with high-energy particles, such as noble gas ion beams, to remove atoms from the surface. This method produces nanoparticles with the same composition as the target material and is more cost-effective than electron beam lithography.[80] However, the choice of sputtering gas, such as helium, neon, argon, krypton, and xenon, can impact the surface morphology, texture, and optical properties of the resulting nanoparticles.[89,90]

2.2.5. Aerosol spray pyrolysis. Aerosol spray pyrolysis is a scalable and cost-effective physical synthesis process.[91] In this method, nanoparticle precursors are transmitted into a heated reactor in the form of small droplets suspended in a vapor, obtaining MNPs with a spherical morphology, narrow particle size distribution, and no agglomeration.[80] However, there are still challenges to be addressed, such as difficulty in controlling the homogeneous pore sizes and inner structure of the particles.[91,92]

Spray pyrolysis is a chemical vapor deposition (CVD) process used to prepare nanomaterials, which have a consistent particle diameter compared to the traditional nanomaterials.[93,94] A precursor solution of metallic salts is used to create an aerosol in the spray pyrolysis process. The produced solution droplets (aerosol) undergo several stages, as follows: (1) solvent evaporation from the droplet surfaces, (2) drying, (3) annealing, (4) production of microporous particles with a defined phase structure, (5) creation of solid particles, and (6) sintering of solid parts. Fig. 4 shows these steps starting from precursors to nanozyme formation.[95]

Several studies were conducted to investigate the influence of different substrate temperatures,[96,97] sampling techniques,[98] presence of chloride ion,[99] and other dispersion parameters[100] on the pyrolysis process. Interestingly, highly porous ternary NiCoFe oxide nanomesh with a two-dimensional shape and quasi-single-crystalline (QSC) property was created using a practical molten-salt-protected pyrolysis method.

The NiCoFe oxide nanomesh possessed high stability, low over-potential, high current density, and excellent oxygen evolution reaction performance with increased intrinsic activity. A quick pyrolysis technique shielded by molten salt (MS, 53% KNO_3, 7% $NaNO_3$, and 40% $NaNO_2$) was carried out at 300 °C to produce mild dehydration to form mixed metal oxides with retained morphology and minimum particle sintering.[101] Another study employed the phase-selective laser-induced breakdown spectroscopy technique to investigate the production of FeO particles along the axial centerline of the spray in an external mixing spray flame pyrolysis reactor, under different precursor solutions. The addition of 2-ethylhexanoic acid to the precursors was examined and significant changes in the evolution of the atomic emission spectra were observed. These changes enabled the differentiation between the gas-to-particle and droplet-to-particle routes *in situ*.[102]

2.3. Biosynthesis

Green synthesis, which involves the use of plants, microbes, and other biological materials, has gained significant attention as a safe, sustainable, and biologically acceptable method for the synthesis of metal oxide nanoparticles, such as iron oxide nanoparticles (IONPs) (Fig. 5). IONPs have attracted particular interest due to their magnetic properties, which allow them to be easily separated from the reaction mixture using an external magnetic field. Biomaterials such as plants, fungi, bacteria, and algae can be used in green synthesis to produce IONPs with a size in the range of 1 to 100 nm and a variety of shapes, including cubic, tetragonal crystalline, spherical, cylindrical, elliptical, octahedral, orthorhombic, hexagonal rods, nanospheres, and quasi spherical. In addition to synthesizing IONPs,

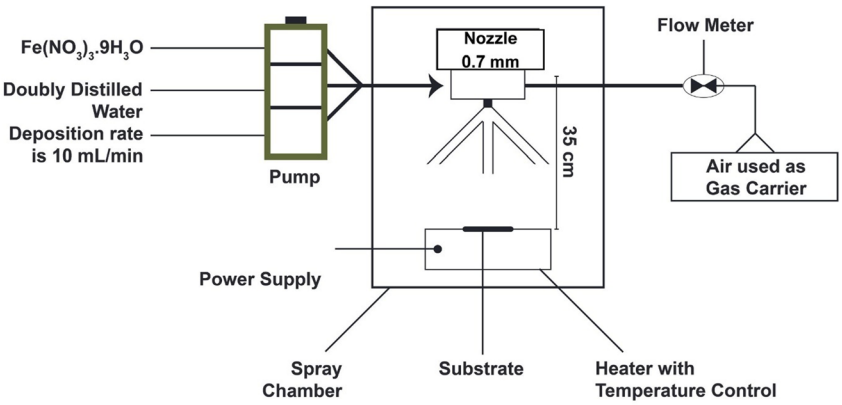

Fig. 4 Setup of the spray pyrolysis technique used for synthesis IONzymes thin film at various temperatures.

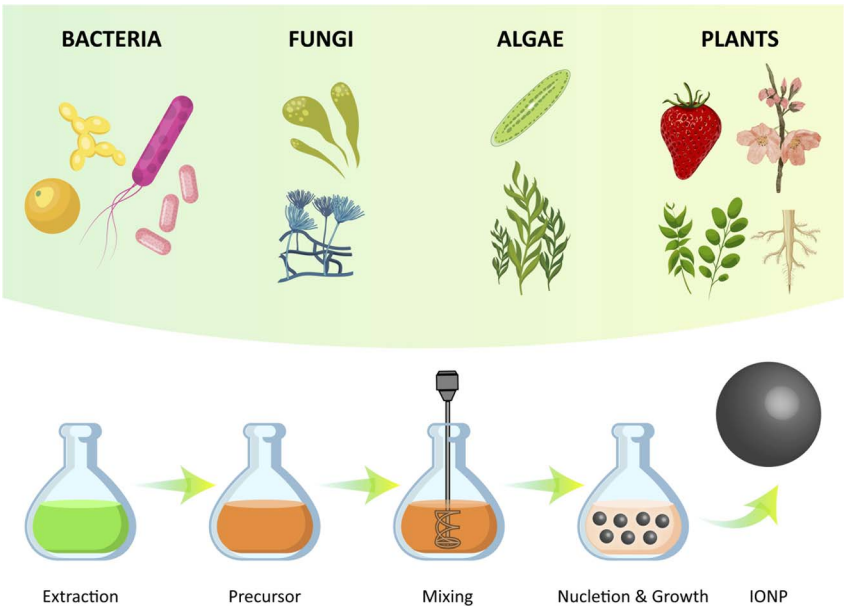

Fig. 5 Biosynthesis of IONzymes using different green sources.

these biomaterials can also act as reducing agents, capping agents, stabilizing agents, and fabricating agents in the green synthesis of nanoparticles.[103]

It is important to highlight the diverse biological pathways employed by various organisms. For instance, *Plumeria obtusa* leaves were employed for the biofabrication of well-defined, crystalline INPs *via* an eco-friendly, cost-effective, and surfactant-free technique. These nanoscale particles displayed potent antimicrobial and antioxidant activity, while remaining non-toxic to red blood cells. This green synthesis presented a potential strategy for the synthesis of sustainable nanomedicines against microbial infections.[104] A study revealed the eco-friendly biosynthesis of IONPs from *Penicillium* spp. fungal filtrate. The extracellular strategy starts by the reduction of $FeCl_3$, with the protein from *Penicillium* spp. playing a pivotal role in capping and stabilizing the IONPs. The characterization of the IONPs showed that they were spherical with high stability. The IONPs exhibited powerful antibacterial and antioxidant activities, making them potential alternatives to antimicrobial and anticancer agents in biomedical applications.[105]

A notable example is the use of bacterial extracellular polymeric substances (EPS) as reducing and stabilizing compounds during the bio-mediated production of metal nanoparticles for multifunctional applications, such as a new bacterium, *E. faecalis*_RMSN6. The EPS was extracted from *E. faecalis* and used for producing highly stable IONPs. This study aimed to assess the effectiveness of the synthesized Fe_3O_4 NPs as adsorbents for removing Cr(VI) metal ions from aqueous solutions. Furthermore, an *in vitro* toxicity analysis using bacterial EPS was conducted to evaluate the potential adverse effects of the synthesized Fe_3O_4 NPs.[106]

Shalaby *et al.* presented a green synthesis method for the preparation of recyclable IONPs utilizing *Spirulina platensis* microalgae. This study highlighted the efficient adsorptive removal of cationic and anionic dyes for water treatment applications. The environmentally friendly synthesis method not only contributes to the sustainable formation of nanomaterials but also exhibits the recyclability of the synthesized IONPs.[107]

2.3.1 Biosynthesis of IONPs using plants. Plants are widely available, easy to handle, and relatively inexpensive materials that can be used for the synthesis of various types of nanoparticles.[108] Different parts of plants, such as their roots, leaves, seeds, flowers, fruits, peels, petals, and whole plants, can be utilized in the biosynthesis process because they contain various biomolecules, such as amino acids, carbohydrates, terpenoids, flavonoids, saponins, proteins, and nitrogenous compounds, which can act as reducing agents, stabilizers, redox mediators, and capping agents in the synthesis of nanoparticles[109–116] (Table 6).

2.3.2 Biosynthesis of IONPs using fungi. The synthesis of iron oxide nanoparticles using fungal species has several advantages, including ease of scaling up the process, use of economical raw materials for growth, high biomass-forming capacity of fungi, simplicity of the downstream processing steps, low toxicity of the residue, and economic feasibility of the process.[180–182] Fungal species also have superior tolerance and bioaccumulation properties, which can aid in the synthesis of metal nanoparticles.[93] The relationship between microorganisms and metals has been thoroughly researched and applied in various biological processes, including bioleaching, heavy metal removal, and bioremediation.[183] In these processes, microorganisms can accumulate and extract metals through the release of enzymes or other mechanisms. These interactions have practical applications in fields such as biotechnology, environmental science, and metallurgy (Table 7).[116,184]

2.3.3 Biosynthesis of IONPs using bacteria. Prokaryotes, which are simple organisms without a defined nucleus or

Table 6 Biosynthesis of iron oxide nanoparticles using plants

Name of the plant	Biomaterial used	Iron precursor used	Size	Shape	Application	Ref.
Hibiscus rosa-sinensis	Dried petals	Ferric chloride (25 mM) and ferrous chloride (25 mM) (2 : 1)	65 nm	Spinel	Biscuit fortification	117
Carica papaya	Dried leaves	$FeCl_3 \cdot 6H_2O$ (0.1 M), NaOH (1 M)	2.159 nm	Not uniform (agglomerated particles)	Antibacterial	118
Psidium guajava	Leaves	$FeCl_3 \cdot 6H_2O$	1–5 nm	Spherical	Antibacterial	119
Citrus	Fresh leaves	Iron chloride (0.1 mM)	15–80 nm	Spherical	Antibacterial	120
Malus pumila (apple)	Peels	$FeCl_2 \cdot 4H_2O$ (20 mM), $FeCl_3$.6 H_2O (40 mM), NaOH (1 M)	50–100 nm	Elliptical and spherical	Decolorization of dye	121
Citrus paradisi	Peels	$FeCl_3 \cdot 6H_2O$ (6 mM)	28–32 nm	Spherical	Antioxidant	122
Syzygium cumini	Leaves	$FeCl_3$ (0.010 mol L^{-1})	40–52 nm	Spherical	Antibacterial, antifungal, aflatoxin B1 adsorption	123
Juglans regia	Dried green husk	$FeCl_3 \cdot 6H_2O$ (97%), $FeCl_2 \cdot 4H_2O$ (99%), NaOH (2 M)	12.6 nm	Cubic	Cytotoxic assay	124
Pyrus sinkiangensis Yu	Peels	$FeSO_4 \cdot 7H_2O$ (0.1 M)	10–90 nm	Irregularly shaped	Cr(IV) removal	125
Cymbopogon citratus	Leaves	$FeCl_3 \cdot 6H_2O$ (0.26 M), $FeCl_3 \cdot 6H_2O$ (0.52 M), Na_2CO_3 (0.75 M)	9 ± 4 nm	Irregular cubic	Nanotoxicological	126
Laurus nobilis	Leaves	$FeCl_3 \cdot 6H_2O$ (0.1 M)	8.03 ± 8.99 nm	Spherical	Antimicrobial	127
Hyphaene thebaica	Fruits	$FeH_{12}N_2O_{12}$ (5 g)	5–10 nm	Quasi-spherical and cuboidal	Antimicrobial, antioxidant, and antiviral	128
Solanum lycopersicum	Leaves	$FeSO_4 \cdot 5H_2O$ (0.1 M)	200–800 nm	Flower	Antibacterial and anticancer	129
Lawsonia inermis	Leaves	$FeSO_4$ (0.01 M)	2 μm	Hexagonal	Antimicrobial	130
Ficus carica	Fruit	($F_2Cl_3 \cdot 6H_2O$) (100 mL)	11–29 nm	Spherical	Antimicrobial	131
Rhamnus Triquetra	Leaves	Ferric acetate (3 g)	∼21 nm	Spherical	Antimicrobial, antioxidant, anticancer, antileishmanial, brine shrimp cytotoxicity	132
Trigonella foenumgraecum	Leaves	$FeCl_3$ (1 M)	27.91–40.94 nm	Grain	Antibacterial	133
Tomato	Fruits	$FeCl_3$ (1 M)	48.18–77.54 nm	Semispherical	Antibacterial	133
Grapes	Fruits	$FeCl_3$ (16.2 g)	49–50 nm	Cubic	Antibacterial	134
Moringa oleifera	Leaves	$FeCl_3$ (0.5 M)	15.01 ± 6.03 nm	Rod-like	Antibacterial	135
Withania coagulans	Berries	$FeCl_3 \cdot 6H_2O$, $FeCl_2 \cdot 4H_2O$ (1 : 2 M)	16 ± 2–18 ± 2 nm	Rods	Photocatalytic degradation and antimicrobial	136
Citrullus colocynth	Pulp Seed	$FeCl_3$ (0.5 M)	12–45 nm 6–15 nm	Spherical	Antimicrobial	137
Durian rind	Peels	Ferrous sulfate (0.05 M)	10 nm	Spherical	Antibacterial	138
Borassus flabellifer	Seed coat	Ferric chloride (0.2 M), ferrous sulphate (0.1 M) (2 : 1)	30–200 nm	Hexagonal	Antimicrobial, antioxidant	139
Citrus sinensis	Peels	Ferric chloride (1 mM)	97.5 nm	—	Antibacterial	140
Thymbra spicata	Leaves	$FeSO_4 \cdot 7H_2O$ (0.1 M)	120.3–17 nm 93.9 nm	Spherical	Antibacterial, antibiofilm, and antioxidant	141
Cocos nucifera L.	Pulps	0.502 g of $FeCl_3$	90–95 nm	Husked rice shape	Antibacterial and anticancer	142
Euphorbia herita	Leaves	Ferrous sulfate (0.1 M), ferric chloride (0.1 M)	25–80 nm	Cavity like	Antimicrobial	143
Camellia sinensis L	Grinded waste of pruned teas	$FeSO_4 \cdot 5H_2O$ (0.1 M), NaOH (0.5 M)	28.5 nm	Regular spherical	Antioxidant	144
Gundelia tournefortii L	Leaves	$FeCl_3 \cdot 6H_2O$ (2 M), $FeSO_4 \cdot 7H_2O$ (1 M), NaOH (1 M)	29.9 nm	Spherical	Remove crystal violet, malachite green, and safranin dyes from prepared aqueous solutions	145

Table 6 (*Contd.*)

Name of the plant	Biomaterial used	Iron precursor used	Size	Shape	Application	Ref.
Aegle marmelos	Leaves	Ferric nitrate (90 mL)	—	Agglomerated	Antimicrobial and antifungal	146
Alstonia scholaris	Leaves	$FeCl_2 \cdot 4H_2O$ (0.5 M) $MgCl_2 \cdot 6H_2O$ (0.5 M)	8.14–13.4 nm	Cubic	Antimicrobial, antioxidant and larvicidal	147
Polyalthia longifolia	Leaves	$FeCl_2 \cdot 4H_2O$ (0.5 M) $MgCl_2 \cdot 6H_2O$ (0.5 M)	8.14–13.4 nm	Cubic	Antimicrobial, antioxidant and larvicidal	147
Coffee	Seed	Fe^{2+} and Fe^{3+} (1 : 1)	23.2–37.5 nm	Cubic	Antibacterial	148
Brassica oleracea var. *Capitata sub.var. rubra*	Peels	Iron(III) chloride (10 mM)	675 ± 25 nm	Agglomerated	Anticancer	149
Zingiber officinale	Root	Ferric chloride (0.1 M)	5.10 nm	Nanocube	Antimicrobial	150
Artemisia	Leaves	$FeCl_3$ (0.01, 0.04, 0.07, and 0.1 M)	24.67–34.28 nm	Cubical	Antioxidant	151
Garcinia mangostana	Peels	$FeCl_3 \cdot 6H_2O$ and $FeCl_2 \cdot 4H_2O$ at a molar ratio of 2 : 1	13.42 ± 1.58 nm	Spherical	Anticancer	152
Chlorophytum comosum	Leaves	$FeCl_3 \cdot 6H_2O$ (0.1 M)	<100 nm	Spherical	Methyl orange dye degradation and antimicrobial	153
Mikania mikrantha	Leaves	$FeSO_4 \cdot 7H_2O$ (5 mmol) and $FeCl_3$ (10 mmol)	20.27 nm	Rhomboidal	Antimicrobial	154
Garlic	Peels	$FeCl_3$ (1 M)	24–44 nm	Irregular	Degrade methylene blue dye	155
Onion	Peels	$FeCl_3$ (1 M)	29–32 nm	Nanofiber	Degrade methylene blue dye	155
Ficus carica	Leaves	$FeCl_3 \cdot 6H_2O$ (0.01 M), NaOH (0.1 M)	43–57 nm	Agglomerated and are multiform	Antioxidant	156
Celosia argentea	Leaves	Ferric nitrate (0.1 M)	5–10 nm	Spherical	Antibiofilm, antioxidant, anti-inflammatory, antidiabetic, and larvicidal activities	157
Plumeria obtusaobtuse	Leaves	$Fe(C_2H_3O_2)_2$ (3 mM)	50 nm	Spheroidal	Antimicrobial, antioxidant	104
Camellia sinensis	Leaves	$FeCl_3$ (10 mM)	13 nm	Cubical	Antioxidant, antimicrobial	158
Persimmon	Fruits	$FeCl_3$ (0.04 M), NaOH (1 M)	30–60 nm	Spherical	Antibacterial and anticancer	159
Punica granatum	Peels	$FeCl_3$ (0.1 M)	17.8 ± 6.5 nm	Cubical	Enzyme mimicking peroxidase, catalase, and superoxide dismutase	160
Buddleja lindleyana	Leaves	Fe $(SO_4)_3 \cdot 6H_2O$ (1 g), $AgNO_3$ (0.1 g)	25 and 174 nm	Triangular and spheroidal	Antimicrobial	161
Hibiscus rosa sinensis	Flowers	$FeCl_2 \cdot 4H_2O$ (1 mM)	51 nm	Tetragonal	Antibacterial	162
Allium cepa	Peel	Ferric chloride (250 mL)	42.78 1 nm	—	Memory-enhancing agent	163
Centaurea alba	Leaves	$FeCl_3 \cdot H_2O$ (0.001 M)	10–52 nm	Spherical	Anti-atherosclerotic and antioxidant	164
Peltophorum pterocarpum	Leaves	$FeSO_4 \cdot 7H_2O$ (0.1 M)	0.085 to 0.2 μm	Irregular	Photocatalytic and catalytic removal of organic pollutants	165
Psidium guajava Linn	Leaves	$FeCl_3$ (1 M), NaOH (1 N)	80.3–99.1 nm	Spherical	Antimicrobial, antioxidant	166
Hylocereus undantus	Fruits	Ferric sulphate and ferrous sulphate (2 : 1)	10–15 nm	Spherical	—	167
Nigella sativa	Seeds	$FeCl_3$ (1 M) and $FeCl_2$ (2 M)	31.45 nm	Spherical	Antimicrobial	168
Mentha spicata	Leaves	$FeCl_3$ (0.4 M)	21–82 nm	Circular or rod	Antimicrobial	169
Cassia auriculata	Flowers	$FeCl_3 \cdot 6H_2O$ (0.1 M)	15–35 nm	Spherical	Photocatalytic degradation and larvicidal effect	170
Melia azedarach	Flowers	Ferrous sulphate (20 mM), NaOH (0.1 M)	231.43 ± 5.21 nm	Spherical	Antimicrobial, antioxidant	171
Echinochloa frumentacea	Grains	$Fe(NO_3)$ (0.1 M)	20–40 nm	Rectangular and triangular	Pharmaceutical, agricultural, targeted drug delivery and biomedical applications	172
Pimenta dioica	Leaves	$FeSO_4$ (0.1 M)	5–15 nm	Spherical	Anticancer	173

Table 6 (*Contd.*)

Name of the plant	Biomaterial used	Iron precursor used	Size	Shape	Application	Ref.
Banana	Peels	$FeCl_3 \cdot 6H_2O$ (2.16 g) CH_3COONa (6.56 g)	44–58 nm	Cubic and agglomerated	Nondestructive technique (NDT) applications	174
Amla	Seeds	$FeCl_3$ (0.01 M)	4–5 nm	Spherical	Removal of toxic dyes	175
Centaurea solstitialis	Leaves	$FeCl_3$ (0.1 M)	—	Spherical	Antimicrobial activity and dye decolorization	176
Eucalyptus globulus	Leaves	$Fe(NO_3)_3 \cdot 9H_2O$ (0.1 M)	2.34 ± 0.53 nm	Spherical	Removal of heavy metals from agricultural soil	177
Galega officinalis	Leaves	$FeCl_3 \cdot 6H_2O$ (40 mM), $FeCl_2 \cdot 4H_2O$ (20 mM) (2 : 1 M ratio)	41.9 ± 1.00 nm	Spherical	Toxicity assessment in plants and aquatic model organisms	178
Coriandrum sativum L.	Leaves	$FeSO_4$ (0.01 mM)	163.5 nm	Spherical	—	179

membrane-bound organelles, have been extensively studied as a model system in the field of nanotechnology due to their widespread presence, fast doubling time, ability to grow under challenging conditions, and the fact that they can be cultivated using inexpensive and straightforward media.[191,192] The application of this system is considered an effective method for synthesizing nanoparticles with a range of shapes, sizes, structural frameworks, and physical and chemical properties through the reduction of metal ions using reductase enzymes, which allow microorganisms to accumulate and detoxify

Table 7 Biosynthesis of iron oxide nanoparticles using fungi

Fungal strain	Biomaterial used	Iron precursor used	Size (nm)	Shape	Applications	Ref.
Aspergillus niger BSC-1	Cell-free filtrate	($FeCl_3 \cdot 6H_2O$) and ferrous sulfate ($FeSO_4 \cdot 7H_2O$) in 2 mM:1 mM	20–40 nm	Orthorhombic	Cr(VI) removal	182
Penicillium spp.	Cell-free filtrate	$FeCl_3$ (3 mM)	3.31–10.69 nm	Spherical	Antimicrobial, antioxidant	105
Chaetomium cupreum	Fungal biomass	$FeSO_4$ (2 g) and NaOH (1.20 g)	25 nm	Spherical	Anticancer	185
Chitosan	—	($FeCl_3 \cdot 6H_2O$), ($FeSO_4 \cdot 4H_2O$)	200–600 nm	Spherical	Postharvest disease inhibition in fruit	186
Penicillium roqueforti	Fungal biomass	Ferric chloride hexahydrate (10^{-3} M) and ferrous chloride tetrahydrate (10^{-3} M)	5–16 nm	Spherical	Antimicrobial	187
Lichen Ramalina sinensis	—	Fe^{2+}/Fe^{3+} (100 mL)	31.74–53.91 nm	Spherical	Antimicrobial	188
Pleurotus florida	—	Ferric chloride (1 M)	100 nm	Spherical	Antimicrobial	189
Penicillium commune	—	$FeCl_3$ (1 mM), $FeSO_4$ (1 mM)	30–50 nm	Spherical	Cleaning gel	190
Bacillus megaterium	—	$FeCl_3$ (1 mM), $FeSO_4$ (1 mM)	40–60 nm	Cubic	Cleaning gel	190
Fusarium oxysporum	—	$FeCl_3$ (1 mM), $FeSO_4$ (1 mM)	20–50 nm	Quasi-spherical	Cleaning gel	190

Table 8 Biosynthesis of iron oxide nanoparticles using bacteria

Bacteria	Salt	Size	Shape	Applications	Ref.
Bacillus subtilis	$FeCl_3$ (2 mM)	12–32 nm	Spherical	Cytotoxicity assay	196
Bacillus subtilis	$FeCl_3$ (2 mM)	3.6 nm	Spherical	—	196
Proteus vulgaris	$FeCl_3$ (3 mM)	20–30 nm	Spherical	Antibacterial, antioxidant	197
Enterococcus faecalis	$FeCl_2 \cdot 4H_2O$ (0.1 M), $FeCl_3 \cdot 6H_2O$ (0.2 M)	15.4 nm	Cubical, hexagonal, brick, and irregular	Heavy metal removal and cytotoxic	106
Enterococcus faecalis	FeCl-.6H–O (1 M)	48.77–55.55 nm	Cubic	Antibiofilm	198
Bacillus coagulans	$FeCl_3 \cdot 6H_2O$, $FeCl_2 \cdot 4H_2O$ (2 : 1 M)	15.18 nm	Cubic	Antibacterial	199
Aeromonas hydrophila	$FeCl_2$ (5 mmol), $FeCl_3$ hexahydrate (10 mmol)	8–12 nm	Spherical	Antibacterial	200

Table 9 Biosynthesis of iron oxide nanoparticles using algae

Algae	Biomaterial used	Iron precursor salt	Size	Shape	Applications	Ref.
Spirulina platensis	Powder	$FeCl_3 \cdot 6H_2O$ (from 0.1 to 0.6 M)	<10 nm	Slightly irregular and rounded	Adsorptive removal of cationic and anionic dyes	107
Sargassum vulgare (*Phaeophyceae*)	Powder	$FeCl_3$ (0.1 M)	22.73 nm	Nanospheres	Antibiofilm	204
Ulva fasciata (*Chlorophyceae*)	Powder	$FeCl_3$ (0.1 M)	28.41 nm	Nanospheres	Antibiofilm	204
Jania rubens (*Rhodophceae*)	Powder	$FeCl_3$ (0.1 M)	27.78 nm	Nanospheres	Antibiofilm	204
Sargassum crassifolium	Dried powder	($FeCl_3$: $FeCl_2$) (0.1 : 0.05 and 0.02 : 0.01)	40–215 nm	Quasi-spherical	—	205
Chlorella vulgaris	Powder	$FeSO_4 \cdot 7H_2O$ and $Fe(NO_3)_3 \cdot 9H_2O$	4.855, 5.702 and 3.614 nm	Amorphous biochar	Adsorbent for dye removal	206
Aegagropila linnaei	Powder	$FeSO_4 \cdot 7H_2O$ (0.01 mol)	100–150 nm	Geomorphic	Adsorption and Fenton-like reaction	207
Ulva lactuca	Powder	$FeCl_3 \cdot 6H_2O$ (28 mM), $FeSO_4 \cdot 7H_2O$ (14 mM)	50–80 nm	Spherical	Adsorptive removal of Pb(II) from heavy metal bearing water	208
Ulva prolifera	Dried-refrigerated powder	$FeSO_4 \cdot 7H_2O$ (0.1 M)	41.23 nm	Spherical	As(III) removal	209

metals.[193] This process involves the use of metal salts as precursors in the reaction and has been used to synthesize metallic nanoparticles.[116,194,195] Table 8 presents some types of bacteria used to produce IONzymes.

2.3.4 Biosynthesis of IONPs using algae. Algae, which include both microalgae (single-celled organisms) and macroalgae or seaweeds (multi-celled organisms), are used in the field of nanotechnology for the synthesis of various types of metallic nanoparticles, such as gold, silver, palladium, iron, and copper (Table 9).[201,202] Similar to plants and bacteria, algae also produce a range of biomolecules, including proteins, fats, carbohydrates, peptides, alkaloids, terpenes, macrolides, cell wall polysaccharides, glycoproteins (containing functional groups such as carbonyl, hydroxyl, carboxyl, and sulfonate), and enzymes, which play a key role in the reduction, capping, fabrication, and stabilization of nanoparticles.[201–203] The use of algae in the production of nanoparticles is considered a safe, simple, cost-effective, and environmentally friendly approach.[116]

3. Structure and design

Functionalized polymeric MNPs exhibit certain distinguished features for drug delivery in terms of effectiveness and efficiency compared to traditional oral and intravenous techniques. This is because they can control the particle size, morphology, and surface charge,[210–212] which enhance the drug delivery and release by joining with other molecules such as antibodies, proteins, and ligands. This can help reduce the side effects associated with chemotherapy, radiotherapy, and surgery.[213] MNPs used for biomedical purposes are often composed of metals such as iron and iron oxide, which can possess a variety of morphologies.

Iron oxide MNPs with nanocrystalline magnetite (Fe_3O_4) cores are preferred for biomedical applications because of their biocompatibility (they swiftly decompose into non-toxic iron and oxygen elements *in vivo*), biodegradability,[214–216] and ease of manufacturing.[135,217] Iron oxide nanoparticles (FeO-NPs) are classified into two categories, *i.e.*, superparamagnetic iron oxide nanoparticles (SPIONs) and ultra-small superparamagnetic iron oxide nanoparticles (USPIONs). These two classes have different relaxometry properties and mean hydrodynamic sizes.[214] SPIONs are made of iron oxide cores with average diameters in the range of 3–20 nm and composed of agglomerates with a hydrodynamic diameter of more than 50 nm.[214] Therefore, any spherical FeO-NPs with a diameter equal to or less than 20 nm will exhibit SPION behavior, which can be used to facilitate targeted drug delivery in the treatment of oncological diseases (Fig. 6).

Magnetite has a face-centered cubic (FCC), closed packing cubic, and inverse spinel structure with the ferric (Fe^{3+}) ion occupying all the tetrahedral (T_h) sites and both ferric (Fe^{3+}) and ferrous (Fe^{2+}) ions occupying the octahedral (O_h) sites. It has attracted significant attention due to the hopping of electrons between Fe^{2+} and Fe^{3+} ions in its octahedral lattice at ambient temperature, as well as its low toxicity (Fig. 6). This is because the iron oxide core of magnetite degrades to low molecular weight iron, making it a useful material in biomedical applications and an effective carrier for drug delivery to target locations, avoiding the negative effects of oral and intravenous drug delivery.[218–227] This is due to the unique properties of magnetite, such as its biocompatibility, lack of toxicity, targeting ability, biodegradability, chemical stability, stable dispersion, and magnetic stability.[228–232]

Due to the anisotropic dipolar attraction and high surface energies of FeO-NPs, large surface-to-volume ratio unmodified FeO-NPs tend to form clusters of large aggregates, which can reduce their surface area to volume ratio and decrease their effectiveness. Additionally, FeO-NPs are prone to oxidation in air, which can lead to the loss of magnetization because of their chemical activity from the oxidation of ferrous (Fe^{2+}) to ferric

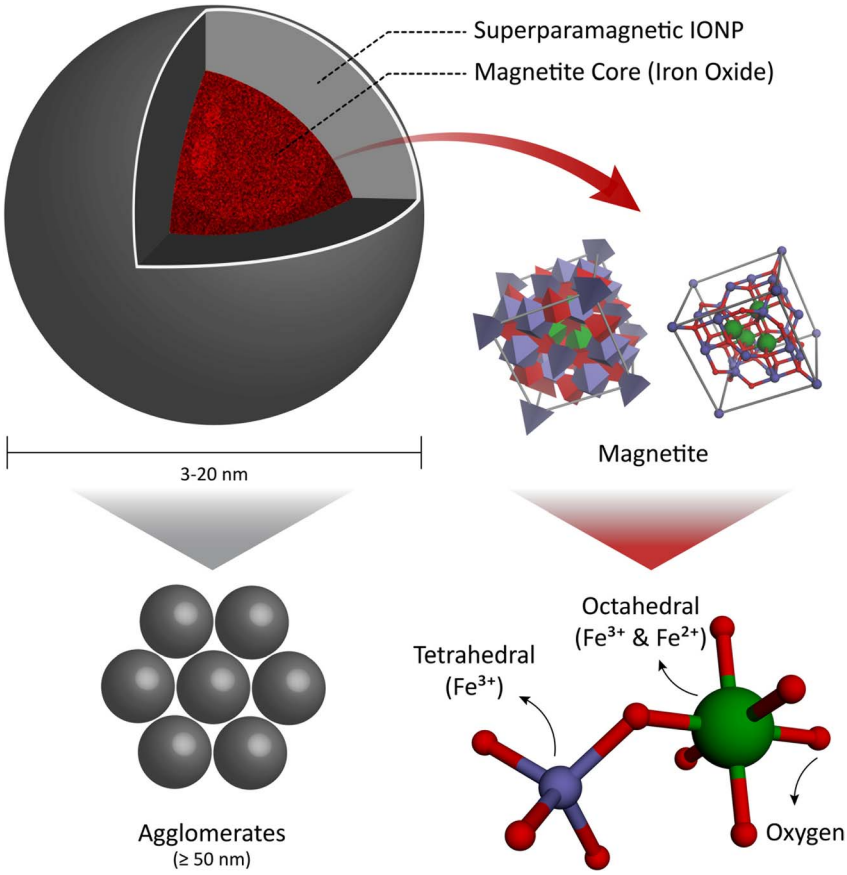

Fig. 6 (a) Superparamagnetic iron oxide nanoparticles (SPIONs). (b) Face-centered cubic (FCC) closed packing, with Fe^{3+} in the tetrahedral sites and Fe^{2+} occupying the octahedral sites.

(Fe^{3+}) ions. Thus, to avoid this oxidation, experiments with FeO-NPs are often conducted under dry conditions.[232–234]

4. Characteristics and applications of IONzymes

Nanomaterials have established numerous novel applications to improve human health, ranging from diagnosis to therapeutic effects, control, and monitoring environment pollution, together with improving the chemical industry.[235–237] Iron oxide nanomaterials have versatile applications that are not limited to their magnetic properties. IONzymes are considered one of the most representative nanozymes being explored for their kinetics and catalytic properties.[238] IONzymes have several benefits in real applications, particularly in biomedicine. In this section (Fig. 7), we focus on the biomedical and environmental applications of IONzymes.

4.1. Characteristics of IONzymes

Recently, diverse nanomaterials with enzyme-like actions were discovered with catalytic properties such as the natural oxidoreductase enzyme family as artificial enzymes or enzyme mimics.[239–243] Recently, researchers have utilized IONzymes in numerous innovative biomedical applications due to their enzyme-like activities.[244–246] In addition, the features of IONzymes are not limited to catalytic activity, where they are also widely applied as biosensors, biomarkers, and in immunoassay approaches.[247–249] Here, we intend to highlight the importance of IONzymes in biomedical applications.

4.1.1. Enzymatic-like characteristics. In 2007, it was reported for the first time that iron oxide nanomaterials display enzymatic-like characteristics. Gao *et al.* stated that FeO-NPs showed basic peroxidase (POD)-like activity, with catalytic behavior similar to horseradish POD (HRP).[250] Since then, IONzymes and their typical POD and catalase (CAT)-like activities have attracted attention because they have been proven to work under physiological conditions like natural enzymes, including the same substrate, pH, and temperature. Moreover, they follow similar kinetics and pathways as conventional enzymes.[251–255] IONzymes are stated to mimic the peroxidase and catalase enzymes. Both enzymes have a porphyrin heme as a cofactor in their active site and utilize hydrogen peroxide as the substrate. Also, both enzymes play a crucial role in avoiding cellular oxidative damage in aerobically respiring creatures by forming free radicals and oxygen.[240,256,257]

4.1.1.1. IONzymes mimic POD activity. POD-like activity was verified for both Fe_2O_3 and Fe_3O_4 IONzymes, which catalyzed a colorimetric reaction including hydrogen peroxide (H_2O_2) utilizing the same optimal conditions as HRP at the

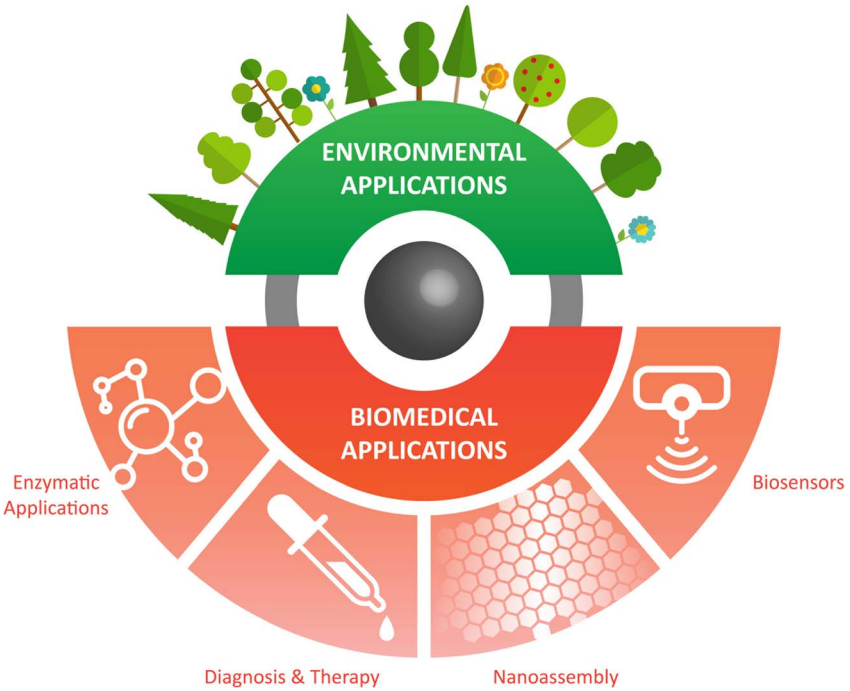

Fig. 7 Applications of IONzymes.

physiological temperature and in acidic media.[258] In addition, IONzymes can function over several substrates, including 3,3′,5,5′-tetramethylbenzidine (TMB), *o*-phenylenediamine (OPD), 3,3′-diaminobenzidine (DAB), 2,2′-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid) (ABTS),[250,259,260] polydopamine,[261] terephthalic acid (TA),[262] luminol, and benzoic acid.[263] Moreover, IONzymes could peroxidize biomolecules such as proteins, nucleic acids, sugars,[264] and lipids.[265]

Furthermore, the enzymatic activity of IONzymes similar to natural PODs can be affected by several natural effectors. ATP, ADP, AMP,[265–267] and DNA are the main activators to improve the POD-like activity of IONzymes by involving them in the electron transfer mechanism.[268,269] Free radical quenchers as sodium azide, ascorbic acid, hypotaurine, and catecholamines were found to decrease the POD activity of IONzymes[270,271] by delaying the affinity of the substrate to IONzymes more than quenching the free radicals.[238]

4.1.1.2. IONzymes mimic catalase activity. IONzymes have been reported to exhibit catalase-like activity *via* H_2O_2 decomposition under neutral and high pH conditions. As previously described for POD and proven by Chen *et al.*, pH plays a significant role in the effectiveness of the H_2O_2 decomposition rate.[259]

4.1.1.3. Kinetics of IONzymes. IONzymes, as POD and CAT enzymes, follow Michaelis–Menten behavior.[272] The apparent affinity of a substrate to the enzyme (KM) value for H_2O_2 was higher for IONzymes compared to the native HRP, indicating that IONzymes have a lower affinity to H_2O_2 than HRP by nearly 41-fold.[250] Alternatively, the KM value for TMB against IONzymes was lower than that of HRP, indicating that the IONzymes have a higher affinity to TMB than the natural enzyme.[273] Given that IONzymes have an abundance of iron ions, this increases their POD activity by around 40-times compared to HRP.[250]

The rate of the CAT activity depends on the O_2 production rate in the solution. IONzymes also adopt Michaelis–Menten kinetics for the CAT reaction.[259] The volumetric measurement of oxygen gas is influenced by many other parameters, such as temperature and O_2 diffusion and can be attained by a volumetric bar-chart chip.[274]

4.1.1.4. Mechanism of action of IONzymes. IONzymes show a catalytic mechanism similar to HRP, where they react with H_2O_2 to form hydroxyl free radicals ($^{\bullet}OH$) as an intermediate state like the POD enzyme state. $^{\bullet}OH$ captures H^+ from the hydrogen donor such as TMB.[265] Interestingly, the produced $^{\bullet}OH$ does not have reaction specificity and can bind to any hydrogen donors, leading to a wide range of applications.[275]

During the activity of IONzymes, two types of free radicals are produced, *i.e.*, $^{\bullet}OH$ and hydroperoxyl ($HO_2^{\bullet}$) radicals. The ferryl ion (FeO^{2+}) that is typically formed in POD catalysis is not detected in the POD-like IONzyme activity but produced in the CAT-like IONzyme activity.[276,277] Moreover, IONzymes have two iron types, *i.e.*, Fe^{2+} and Fe^{3+}, where Fe^{2+} ions may play a major role in their catalytic POD-like activity.[250]

The POD-like activity arises also from the integral nanoparticles rather than free iron ions, and thus the IONzyme mechanism performance includes kinetic procedures involving substrate binding, surface reaction, and product release, displaying similar enzymatic kinetics.[278,279] Furthermore, IONzymes can be utilized as an exceptional carrier to load other enzymatic functionalities on their surface. For example, glucose oxidase (GOx) can form a new nano-complex by GOx catalyzing glucose to produce hydrogen peroxide, which in turn can be catalyzed by IONzymes.[280]

4.2. Biomedical applications of IONzymes

4.2.1. Immunoassay, diagnosis, therapy, and biomarker detection. Based on the superiority of IONzymes over HRP, they can be used as an alternative to HRP in the enzyme-linked immunosorbent assay (ELISA) and other associated molecular detection procedures through the conjugation of antibodies.[281,282] Based on the superparamagnetism of IONzymes, they can be used to enhance antigen detection at low concentrations.[283] Gao *et al.* developed chitosan-modified magnetic nanoparticles (CS-MNPs) as additional enzymes in conventional ELISA configurations, with 1 ng mL^{-1} detection limit for a carcinoembryonic antigen (CEA).[281] Similar approaches were adapted to detect other antigens or pathogens, containing immunoglobulin G (IgG), hepatocellular carcinoma biomarker Golgi protein 73 (GP73),[284] human chorionic gonadotropin (HCG),[285] mycoplasma pneumonia,[286] *Vibrio cholerae*, rotavirus,[287] cancer cells with human epidermal growth factor receptor 2 (HER2),[287,288] and epidermal growth factor receptor (EGFR).[289] Daun *et al.* developed an iron oxide nanozyme-strip to sense Ebola virus (EBOV) with a detection limit as low as 1 ng mL^{-1} for EBOV glycoprotein.[290] Moreover, the surface of IONzymes was covered with streptavidin to attain signal amplification *via* IONzyme catalysis by Thiramanas *et al.* to sense *Vibrio cholerae* with a sensitivity of 10^3 CFU mL^{-1} in drinking and tap water.[291] Moreover, Zhang *et al.* established a colorimetric aptasensor for the determination of thrombin by employing chitosan-modified Fe_3O_4 (MNPs). The results exhibited that the thrombin absorption values improved in a concentration-dependent manner with a linear range from 1 to 100 nM.[292] Based on the aptamer conjugated to the IONzymes, Fe_3O_4 NPs with an aptamer-based immunosorbent assay (NAISA) were developed for aflatoxin B1 (AFB1) recognition with better operation and separation. The aptamer was implemented to diagnose AFB1, and this method showed a limit of detection of 5 pg mL^{-1} (Fig. 8).[293]

Magnetoferritin NPs (M-HFn) are a certain type of IONzymes that are linked to the recombinant human heavy-chain ferritin (HFn) protein shell, which binds to transferrin receptor 1 (TfR1) overexpressed in most tumor cells.[294] This approach enables tumor diagnosis by utilizing the POD/CAT functionality of the Fe_3O_4 core to yield a color reaction, which can be utilized to visualize cancer tissues. This strategy can differentiate cancerous cells from normal cells with a sensitivity of 98% and specificity of 95%.[238,295] IONzymes and their POD activity can be used in bio-distribution studies. Based on this technique, Zhuang reported that dextran-coated Fe_3O_4 NPs were confined in the liver, spleen, and lungs more than the kidney, lymph nodes, and thymus (Fig. 9A).[295]

In addition, IONzymes showed a therapeutic effect on tumor cells and against bacterial growth by catalyzing H_2O_2 to produce toxic radicals.[296,297] To increase the intracellular H_2O_2 concentration, H_2O_2 was directly injected, or an enzyme was merged to generate H_2O_2. The former showed a significant inhibition efficacy against a mouse model bearing subcutaneous HeLa tumors.[296] Ferumoxytol was utilized with a low concentration of H_2O_2 to fight oral biofilms and avoid dental decay. Ferumoxytol, carboxymethyl dextran-coated IONzymes could catalyze the decomposition of H_2O_2 to hydroxyl radicals (Fig. 9B).[298] However, due to the increased toxicity of the H_2O_2 injection option, incorporating an H_2O_2-producing enzyme is considered an efficient and safer choice. For example, Huo *et al.* reported that Fe_3O_4 NPs and GOx co-entrapped in mesoporous silica NPs could be used for tumor catalytic therapy.[299]

Moreover, iron-based NPs can cause generate sufficient reactive oxygen species (ROS) to induce apoptosis in tumor cells into (ferroptosis).[300,301] For example, Fe^{3+} is reduced to Fe^{2+} by the overexpressed glutathione (GSH) in tumor tissues, leading

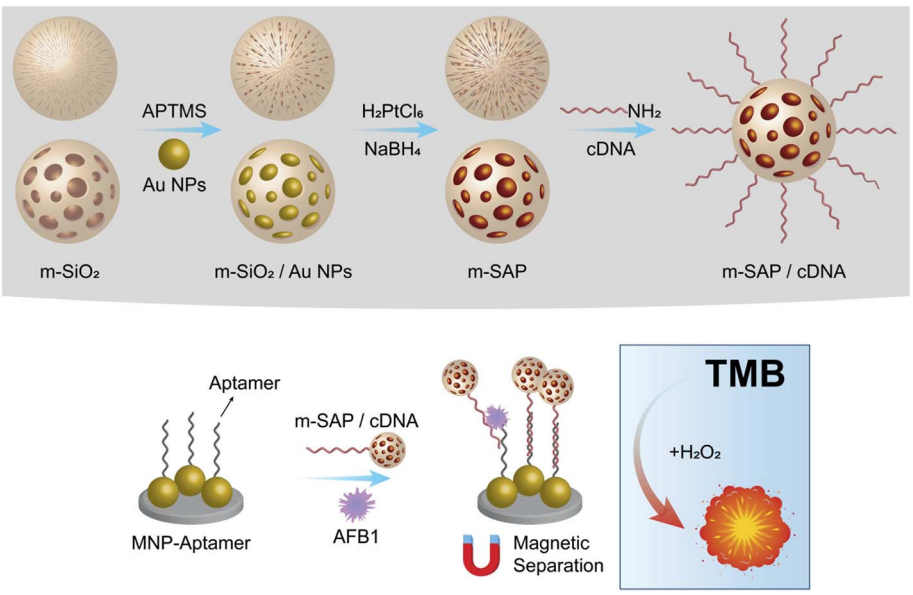

Fig. 8 Schematic presentation of nanozyme and aptamer-based immunosorbent assay (NAISA): (A) preparation of m-SAP/cDNA and (B) construction of NAISA for AFB1 detection.

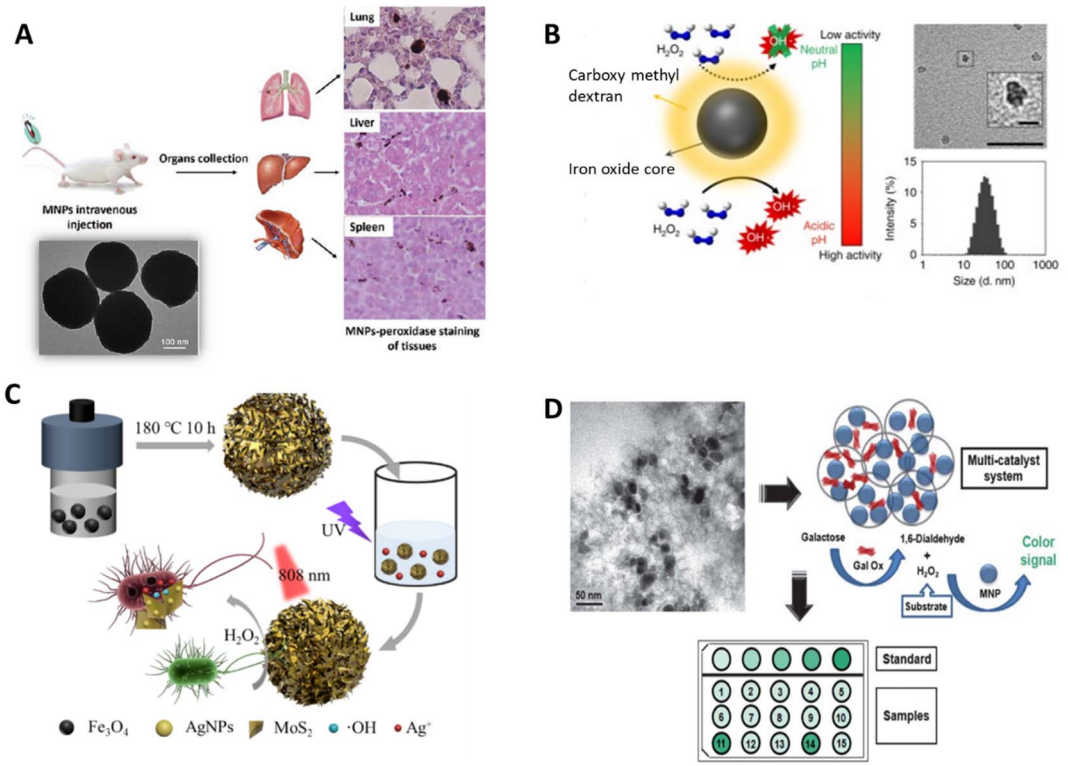

Fig. 9 Schematic illustration of (A) investigation of dextran-coated Fe_3O_4 NPs in the liver, spleen, and lungs[295] and (B) pH-dependent catalytic activity of ferumoxytol. (Insets) negative stain TEM of ferumoxytol (Scale bar: 50 nm and 10 nm for close image) and hydrodynamic diameter measurements.[298] (C) Recoverable peroxidase-like Fe_3O_4@MoS_2-Ag nanozyme with enhanced antibacterial ability.[303] (D) Multi-catalyst system for the quantification of galactose, entrapping both MNPs and Gal Ox in mesocellular silica.[316]

to the promotion of ROS production and resultant tumor ferroptosis.[257,302] In addition, Wei *et al.* developed Fe_3O_4@MoS_2-Ag IONzymes that showed a good antibacterial effect against *E. coli* (~69.4%) by the generated ROS through POD-like activity and released Ag^+ (Fig. 9C).[303] Furthermore, Wang *et al.* conveyed that a new cobalt-doped Fe_3O_4 (Co@Fe_3O_4) IONzyme exhibited better POD activity and a 100-fold higher affinity to H_2O_2 than Fe_3O_4 nanozymes to generate ROS for kidney tumor catalytic therapy *in vitro* and *in vivo*, presenting a potential novel avenue for tumor nanozyme catalytic treatment.[304] Similarly, Sun *et al.* improved highly toxic ROS levels from iron oxide core–shell mesoporous silica nanocarrier-mediated Fenton reactions for cancer therapy.[305] Furthermore, Li *et al.* prepared an H_2O_2-responsive POD and CAT-mimic PtFe@Fe_3O_4 IONzyme, which displaced a 99.8% anti-tumor rate for deep pancreatic cancer in collaboration with photothermal treatment.[306] In a recent study, the application of adenosine triphosphate disodium salt (ATP) as a synergistic agent increased the generation of OH radicals and restored the antibacterial activity of Fe_3O_4 IONzymes over the full pH range against both Gram-positive (*B. subtilis*) and Gram-negative (*E. coli*) bacterial strains in the presence of H_2O_2 at a pH of around 7.0.[307]

4.2.2. Enzyme-IONzyme nanoassembly. This approach utilizes IONzymes loaded with oxidase enzymes to enable the fast colorimetric detection of biomolecules. The natural enzyme usually produces H_2O_2 as an intermediate, which is catalyzed by the activity of the POD like-IONzymes.[308]

Glucose oxidase is the main enzyme assembled with IONzymes for the detection of glucose.[309,310] Firstly, GOx catalyzes glucose to produce H_2O_2, which is then catalyzed by IONzymes, and a color signal can be formed related to the amount of glucose with a detection limit of 0.5 to 3 μM.[311–314] Other oxidases and esterases can also be utilized in this approach such as cholesterol oxidase for cholesterol detection,[315,316] galactose oxidase (Gal Ox) for galactose (Fig. 9D),[316] alcohol oxidase (AlOx) for alcohol,[317] and acetylcholine esterase (AChE) for acetylcholine (ACh).[265]

4.2.3. Biosensors. Several IONzymes have been used by researchers for the development of IONzyme-based biosensors for biomedical applications. IONzyme-based biosensors are based on the mimicking activity of IONzymes and can be categorized into three main groups: POD, oxidase, and CAT mimics.[318] Table 10 summarizes the reported IONzyme-based biosensors together with their enzyme-simulating activities and sensing mechanism.

4.3. Environmental applications of IONzymes

Due to their high catalytic activity, stability, and multifunctionality, IONzymes have shown an increasingly wide range of applications in the biomedical, agricultural, and environmental fields.[329–331] Given that IONzymes possess intrinsic POD and CAT properties and follow a Fenton and/or Haber–Weiss reaction mechanism (including $^{\bullet}OH/HO_2^{\bullet}$), they can be utilized

Table 10 IONzyme-based biosensors and their type of enzyme-mimicking activities and sensing procedures

IONzymes	Enzyme-mimicking activities	Biotarget	Biosensor type	Ref.
Fe_3O_4	POD	Ebola virus	Colorimetric	319
Fe_3O_4-Pt/core–shell	POD	Human chorionic gonadotropin (hCG)	Colorimetric (paper-based strip)	320
Fe-MOF-Au NPs	POD	*Salmonella enteritidis*	Colorimetric immunosensor	321
Fe_3O_4	POD	Glucose	Colorimetric	322
Fe_3O_4	POD	*Listeria monocytogenes*	Colorimetric	323
Fe_3O_4	POD	Prostate-specific antigen	Photoelectrochemical (PEC) immunoassay	324
Fe_3O_4	POD	Micro RNA	Electrochemical	325
Fe_3O_4	POD	Hepatitis B virus surface antigen (preS1)	Colorimetric, immunoassay	250
Fe@PCN-224 NPs	POD	Glucose	Colorimetric	326
Fe_3O_4@C	POD	Platelet-derived growth factor BB (PDGF-BB)	Colorimetric	327
Fe_3O_4/CoFe-LDH hybrid	POD	Ascorbic acid	Colorimetric	328

for the degradation of organic pollutants by combining free radical production with the magnetic characteristics of iron oxide.[332] Moreover, the catalytic activity IONzymes can be used for environmental monitoring, for example, detecting H_2O_2 in rainwater and measuring heavy metals in environmental samples. The environmental applications of IONzymes are considered suitable for numerous environmental conditions, relatively easy and cheap, and can be simply applied to the screening of pesticides, organophosphorus compounds, and other ingredients. IONzymes can determine pollutants indirectly when they enable a target to undergo an alteration in chemical properties and react with the colorimetric sensor to be detected.[333]

A histidine-modified Fe_3O_4 IONzyme offered an easy, inexpensive approach to detect Ag^+ with a detection limit of 18 fg mL^{-1}.[334] 4-Chloro-1-naphthol was utilized as a substrate, in which the Fe_3O_4 IONzyme POD enzyme activity was activated in the presence of Ag^+, which produced the insulating precipitation of benzo-4-chlorohexadienone. The insulating products attenuated the photocurrent signal, reflecting the presence of Ag^+. Guo *et al.* developed an excellent colorimetric selective method for the detection of Hg^{2+} based on the stimulus of the intrinsic oxidase-like catalytic activity of Ag–$CoFe_2O_4$/rGO NPs *via* a one-pot microwave-assisted reaction, which can oxidize 3,3′,5,5′-tetramethylbenzidine (TMB) to yield a light-blue product.[335]

Recently, IONzymes have been established as anti-microbial for environmental treatments. IONzymes effectively inactivate viruses (IAVs) *via* envelope lipid peroxidation and destruction of the integrity of neighboring proteins, including hemagglutinin, neuraminidase, and matrix protein. Furthermore, IONzymes possess a broad-spectrum antiviral activity against 12 subtypes of IAVs 244 (H1–H12).[336]

In the treatment of organic pollutants in water, ferromagnetic chitosan IONzymes (MNP@CTS), which have superior catalytic activity and exceptional POD activity, were produced for the degradation of phenol. MNP@CTS removed over 95% of phenol from an aqueous solution within 5 h under the optimum conditions (pH range of 2–10).[337]

Huo *et al.* showed that IONzymes can enhance the performance of plants under unfavorable conditions such as abiotic stresses. They studied the effect of a 25 ppm IONzyme dose on *Eucalyptus tereticornis* against a high salinity concentration of 300 mM NaCl. The IONzymes showed a separate biochemical change in superoxide dismutase, malondialdehyde concentration, and total soluble sugar and proline content, which are biomarkers that circumvent the stress response and synergistically improve the activity of CAT and POD enzymes.[338]

Recently, Fe_3O_4-TiO_2/reduced graphene oxide (Fe_3O_4-TiO_2/rGO) NPs with hydrogen peroxide activity and photocatalytic efficiency were designed for the colorimetric detection of atrazine pesticides, which can cause long-term negative effects because of their persistence. TMB was used as the substrate compound with a detection limit of 2.98 μg L^{-1}.[339] Moreover, the POD-like activity of IONzymes was utilized in water purification in another study.[340–343] This designates the promising application potential of Fe_3O_4 IONzymes in water treatment and quality analysis.

4. Future scope and drawbacks of IONzymes

The future potential as and present limitations of the applications of IONzymes must be considered for their development. IONzymes have enormous potential for use in the environmental and biomedical fields. Employing IONzymes in targeted drug delivery systems and improved diagnostics is one field of research and development that has great potential.[344] Thorek *et al.* suggested that IONzymes may revolutionize magnetic resonance imaging (MRI) by improving the imaging contrast and specificity. However, there are a few issues and disadvantages that need to be resolved. One of the main challenges is still their long-term biocompatibility and toxicity, particularly for *in vivo* applications.[345]

Although iron oxides are considered to be biocompatible in general, Szalay *et al.* proposed that further research is needed to determine their long-term impacts in biological systems.[346] Another serious obstacle is the synthesis of IONzymes on a large scale for industrial use. Researchers emphasized that the shift from laboratory-scale production to large-scale manufacturing frequently leads to instabilities in particle size and enzyme activity, which can hinder their practical implementation.[346]

One more crucial element is the stability of IONzymes in physiological settings. Thus, enhancing their stability through surface modifications, while maintaining their enzymatic activity under various pH and temperature conditions, is crucial for their effective use in biological systems.[38] In addition, achieving high specificity in the catalytic action of IONzymes remains a major research goal. As noted by Zhang and co-workers, tailoring nanozymes to exhibit enzyme-like specificity is a complex but vital aspect for their application in both the medical and environmental fields. Enabling the actual deployment of IONzymes requires addressing these obstacles *via* inventive research and technological developments.[39] To fully utilize the promise of IONzymes in a variety of applications, future research should focus on improving their biocompatibility, scalability, stability, and specificity.[346]

5. Conclusion

In conclusion, the study of IONzymes has seen significant advances in recent years. IONzymes can be synthesized using chemical, physical, and biological techniques and offer unique advantages for various applications, including biomedical and environmental purposes. IONzymes have been explored for their enzymatic properties and used in enzyme mimicry, immunoassays, diagnosis, therapy, and biomarker detection. Thus, the versatile nature of IONzymes, combined with their biocompatibility and biodegradability, make them a promising area for continued research and development.

Author contributions

Ghazzy A, conceptualization, writing – original draft, writing, review & editing, supervision; Nsairat H, writing – original draft; Said R, writing – original draft, writing, review & editing; Sibai1 O, writing – original draft, resources; AbuRuman A, writing – original draft; Shraim A, visualization & editing; Al hunaiti A, conceptualization.

Conflicts of interest

There are no conflicts to declare.

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