Green Chemistry



View Article Online

CRITICAL REVIEW

Check for updates

Cite this: Green Chem., 2024, 26, 42

High-value utilization of lignin: construction of an intelligent release system for targeting the delivery of pesticides

Yitong Wang, ^(D)^a Xiaona Yu,^a Shuaishuai Ma,^a Shuling Cao, ^(D)^b Xufeng Yuan,^a Wanbin Zhu^{a,b} and Hongliang Wang ^(D)*^{a,b}

The utilization of biomass for the production of agricultural green inputs is regarded as a crucial strategy for achieving low-carbon development in agriculture, while also fully harnessing the potential of renewable resources. Pesticides, as a vital agricultural input, often encounter issues pertaining to inefficient usage, resulting in significant environmental pollution and economic losses. As a main component of lignocellulosic biomass, lignin has become one of the most appealing biopolymers for the construction of advanced pesticide delivery systems. This review aims to provide a thorough summary of the advancements in lignin-based controlled release formulations (LCRFs) for the precise delivery of pesticides. The research in this field has experienced rapid growth in the past five years, making it an important area of study. Common LCRFs are introduced, and the factors influencing the release of active ingredients (Als) within different LCRFs are analyzed. Special emphasis is placed on intelligent-responsive LCRFs, encompassing an overview of the existing formulations and an exploration of their potential application scenarios and development strategies. It is crucial to promote innovation in pesticide formulations based on the actual demands of agricultural production. We hope this review will stimulate the high-value utilization of lignin and the green development of plant protection technologies.

Received 11th September 2023, Accepted 22nd November 2023 DOI: 10.1039/d3gc03434h rsc.li/greenchem

Introduction

Plant diseases are regarded as one of the most significant restrictions to agricultural productivity. According to statistics, globally, pests and diseases cause an average crop loss of 21.5% for wheat, 30.0% for rice, 22.6% for maize, 17.2% for potatoes, and 21.4% for soybeans.¹ Global economic loss from plant diseases brought on by bacteria, fungi, nematodes, and viruses exceeds \$220 billion annually.2 Pesticides have an important role in improving global crop yields, relieving hunger, and ensuring adequate food quality. Humans have used drugs to prevent and control agricultural diseases since ancient times. However, to date, the use of pesticides still poses some pressing issues. One of the most common phenomena is that during pesticide spraying, off-target losses occur due to evaporation, photolysis, splashing, and bouncing. Studies have shown that during the actual production process, harmful organisms were exposed to only 0.1% of the amount of pesticide applied.³ Furthermore, most farmers rely on agrochemicals to control pests and abuse pesticides to improve agricultural product yields.^{4,5} The misuse of pesticides not only generates financial losses but also causes severe risks to the environment and non-target creatures.^{6–8} The aim of maintaining high crop yields while reducing the use of pesticides remains a great challenge.

As an increasingly valued technology, pesticide-controlled release involves specific techniques to control the release rate, quantity, and pathway of pesticides in media, such as soil, air, water bodies, and plants. Compared with traditional pesticide application, the use of a pesticide-controlled release system (CRS) reduces issues associated with chemical drift, leaching, volatilization, and degradation, thus enhancing the pesticide utilization efficiency and reducing occupational exposure risks, as well as minimizing detrimental effects on ecosystems and non-target organisms.^{9,10} Pesticide-controlled release formulation (CRF) is a pesticide dosage form that can realize the gradual release of active ingredients (AIs) from processed products. Among the numerous parameters that influence CRS, the selection of carrier materials for the formulation plays a crucial role in determining the performance of the delivery system. The ideal pesticide carrier material should not react with the core contents and should possess appropriate characteristics, including mechanical strength, solubility, fluidity,

^aCenter of Biomass Engineering/College of Agronomy and Biotechnology,

China Agricultural University, Beijing, 100193, P. R. China.

E-mail: Hlwang@cau.edu.cn

^bSanya Institute of China Agricultural University, Sanya, 572025, P. R. China

emulsifying property, permeability, and stability.¹¹ Various inorganic materials such as clay,¹² silica,¹³ metals/metal oxides,¹⁴ synthetic polymers, such as polyethylene glycol,¹⁵ polyurea,¹⁶ and polyhydroxyalkanoates,¹⁷ and natural organic polymer materials, such as alginates,¹⁸ chitosan,¹⁹ and lignin,²⁰ have been explored for CRF carriers. Inorganic materials have advantages, such as high surface area, high drug delivery efficiency, and stable physical and chemical properties, but are usually monofunctional and difficult to degrade after entering the natural environment. Synthetic polymers have the characteristics of low cost and stable performance, but they largely rely on fossil resources. With the continued depletion of fossil resources, society needs to shift towards a more sustainable development mode, promoting the use of bioresources to replace fossil ones.²¹

In recent years, natural polymers have emerged as promising additives and carriers for constructing environmentallyfriendly and high-performance CRFs because of their easy access, low prices, and biodegradability. Lignin, in particular, is a highly compelling subject that continues to garner increasing interest in this field. As one of the primary components of lignocellulose, lignin, after cellulose, stands as the second most abundant renewable bioresource. Additionally, it is the only natural aromatic polymer that is obtainable in substantial quantities from terrestrial plants.^{22,23} The pulp & paper industry generates a significant amount of lignin (over 50 million tons per year) as by-products,²⁴ most of which have not been effectively utilized. The valorization of lignin through the conversion into aromatic chemicals and advanced fuels has emerged as an appealing pathway. However, due to the low selectivity of desired products, achieving profitability from this process can be challenging. On the other hand, utilizing lignin as a substitute for synthetic polymers in various applications as functional materials proves to be a more lucrative option. In recent years, lignin has been widely tested for the preparation of industrial polymeric materials, such as polyurethanes, phenolic, and epoxy resins, exhibiting comparable performance to synthetic materials while offering cost reduction.²⁵ However, to fully unlock the potential of lignin, an exciting avenue worth exploring lies in utilizing lignin for the production of high-value materials, particularly micro/nanomaterials, which find widespread applications in medical and agricultural fields. The development of lignin-based controlled release formulations (LCRFs) of pesticides is now emerging as a promising strategy, which is not only beneficial for agricultural production, but also for the high-value utilization of biomass.

There has been an increasing number of reports on LCRFs. As a pesticide carrier material, lignin has the following advantages: (1) from the standpoint of formulation functionality, lignin contains a variety of functional groups that can be chemically modified (*e.g.*, hydroxyalkylation, esterification, and amination) to tailor LCRFs for different agricultural scenarios.²⁶ (2) As for pesticide utilization efficiency, the phenolic, ketonic, and chromophoric structures in lignin impart the biopolymer UV resistance and antioxidant properties, which can reduce the degradation of pesticides under natural conditions

and prolong their effectiveness.^{27,28} (3) From an environmental protection standpoint, lignin is non-toxic, harmless, and naturally degradable without exhibiting cytotoxicity.^{29,30} (4) Considering crop growth, lignin degradation can transform into humus, thereby enhancing soil fertility.³⁰ Moreover, the benzene ring, phenolic hydroxyl groups, and methoxy groups in lignin endow it with natural antibacterial activity,^{31,32} which can alleviate the damage caused by pathogenic bacteria to crops and be beneficial to their growth and development. Based on the multiple advantages, lignin has been designed for various LCRFs in recent years, with intelligent ones being the most striking. Intelligent LCRFs possess the capability to convert various stimuli into predetermined outputs, thereby altering or activating specific functional characteristics of the system. This is more in line with the trend of precision agriculture (Fig. 1). There are several excellent reviews on lignin conversion to materials, including those of smart ones used as sensors, biomedical systems, and shape-programmable materials.^{26,33,34} However, a comprehensive summary and analysis of the most recent advancements in the utilization of lignin as intelligent materials, specifically focusing on applications in the targeted delivery of pesticides in the past five years, has yet to be presented. Therefore, this review offers a summary of recently published papers on intelligent-responsive LCRFs, introducing typical systems, application scenarios, development strategies, and summarizing their trigger factors and release characteristics. The highlight of this study is the emphasis on the response patterns of lignin to various environmental stimuli, revealing the potential of lignin as an intelligent material used in agriculture. We hope this review will stimulate the high-value utilization of lignin and the green development of plant protection technologies.

Typical LCRF systems

As compared to conventional burst-release formulations, sustained release systems gradually and consistently release AIs over an extended period of time, ensuring the long-lasting control of harmful organisms. By maintaining suitable concentrations of AIs around the target, sustained release technology allows for a reduction of environmental concerns associated with its implementation. This is achieved by incorporating AIs into specialized matrices or formulations. Since the late 20th century, attempts have been made to utilize lignin as the initial matrix for CRF.^{35,36} The source,³⁵ structure,³⁷ molecular weight,³⁸ and concentration³⁷ of lignin can all impact the release rate of AIs. Lignin macromolecules in various aggregation states form a variety of LCRF systems (Fig. 2). Some typical LCRF systems and their functional characteristics are summarized below.

Lignin-based granules

The lignin-AIs granular complex is the earliest system to be used for the sustained release of pesticides. The preparation method is relatively simple, involving the direct mixing of pes-



Fig. 1 Development of lignin-based controlled release systems: from sustained release to intelligent response of active ingredients.



Fig. 2 Different types of lignin-based controlled release systems. (In the figure, the granule system depicts the macrostructure, while the other systems illustrate the microstructure.)

ticides and lignin, followed by heating in a molten state for a certain period of time and subsequent cooling. Upon cooling, a glassy matrix is formed, from which the pesticide is released through diffusion. When preparing granules, the solubility parameters of the pesticide and lignin need to be considered. Only when the solubility parameter of the pesticide is similar to that of lignin can favorable compatibility be achieved.³⁵ The size of granules is typically within the millimeter range, and it is the primary factor affecting the release rate. As the average granule size increases, it implies a longer distance for the AIs diffusion from the center of the matrix to the external surface, resulting in a slower release rate.³⁹ Researchers have developed regression equations between the granule size and t_{50} (the time required for 50% of the AIs to be released), which indicates that the release rate of AIs from the granules can be

tuned by regulating the particle size.^{39,40} However, due to only a portion of AIs having acceptable compatibility with lignin, as well as the potential loss of AIs during heating and melting, the application of granules is significantly restricted.

Lignin-based micro- or nanoparticles

Micro- and nanoparticles currently represent the most extensively studied LCRF system. Lignin-based micro- or nanoparticles typically possess adjustable sizes and shapes, with AIs being captured, encapsulated, or adhered to the surface of these nanoparticles.¹⁰ Special structures such as porous⁴¹ and hollow⁴² LCRF particle systems have also been reported. The anti-solvent method is commonly used for the preparation of micro- and nanoparticles, and the choice of solvent can serve as a means to adjust particle size. In the same solvent, an increase in lignin concentration leads to a reduction in particle diameter, but this reduction does not affect the encapsulation efficiency and drug loading of the system.^{43,44} Similar to granules, the release of AIs is mainly determined by their particle size, with smaller sizes resulting in faster release.^{45,46} Reducing the particle size can help to enhance the application performance of formulations (*e.g.*, better dispersion at the foliar surface and better penetration into target organisms),⁴⁷ but an excessively small particle size may alter the enrichment and fate of pesticides in the environment. Therefore, it is necessary to control the particle size to be within a range that can maintain the advantages of the small particle size, while mitigating risks.⁴⁸

Lignin-based micro- or nanocapsules

Micro- or nanocapsules refer to egg-like structures with a coreshell architecture, which can encapsulate solid, liquid, or even gaseous AIs.⁶³ Capsules can effectively control the release behavior of AIs, and protect them from degradation, as well as isolate harmful components from non-target organisms. In LCRFs, lignin often acts as the shell of the capsules. Unlike particle systems, which regulate release rates by adjusting the particle sizes, capsule systems modulate release by adjusting the thickness of the capsule shell. Increasing the thickness of the shell (or adding more layers) slows down the release. However, thicker shells typically result in a slight compromise in drug loading and encapsulation efficiency of the system.^{64,65} It should be noted that compared to particles, capsules usually exhibit a lag-time effect on the release of AIs (i.e., AIs usually take more time to release from the capsule) due to the physical separation between the AIs and the shell material.66

Lignin-based emulsions

An emulsion refers to a mixture of oil phase, water phase, and surfactant in varying proportions.⁶⁷ It is also frequently used for encapsulation and the controlled release of sensitive chemicals. Due to the amphiphilic nature of lignin molecules,²⁵ they can be partially or completely used as alternative surfactants after slight modification. Consequently, compared to particles and capsules, the use of lignin-based emulsion systems reduces the cost and potential environmental issues associated with the extensive use of surfactants. When all surfactants are replaced with lignin solid particles in an emulsion system, a specific emulsion called a lignin-based Pickering emulsion is formed.⁶⁸ The release of AIs in emulsion systems is closely related to the stability of the emulsion, which is regulated by the content and amphiphilicity of lignin particles.^{59,60} Insufficient lignin at the oil-water interface hinders the formation of a stable emulsion, while an excessive content leads to aggregation and affects the emulsification performance. Lignin particles with excessive hydrophilicity or lipophilicity fail to stabilize well at the oil-water interface, requiring modification to achieve suitable amphiphilicity. Generally, higher stability of the emulsion results in a slower release of AIs.^{59,60}

Lignin-based coacervates

In the current LCRFs system, aqueous-phase coacervates stand out as the only system that can be formed without the use of organic solvents, making them highly environmentally friendly.⁶² Lignin-based coacervates exhibit a rigid honeycomb-like network structure, providing a higher encapsulation efficiency for both hydrophilic and hydrophobic AIs. Simultaneously, it is precisely these network structures that can be entangled with the micro/nanostructures of the superhydrophobic foliage, resulting in superior wetting and antisplash performance when the coacervate droplets are highspeed sprayed on leaf surfaces.⁶² The development of ligninbased coacervates is still in its nascent stage, but holds tremendous potential for various applications.

The applications of these aforementioned systems for the controlled release of pesticides are summarized in Table 1. It is important to note that there is no single exclusive and one universal lignin type that is best suited for the preparation of LCRFs. Thus, it should be selected based on the desired properties of the final products. For instance, sodium lignosulfonate is often employed as pesticide dispersants due to its favorable water solubility and polyanionic properties. Kraft lignin has low solubility under acidic conditions, and can be used to prepare lignin nanoparticles through acid precipitation method.⁶⁹ Meanwhile, the pursuit of excessively prolonged release times should be avoided, and choices should be made based on specific agricultural scenarios to prevent the loss of maximum therapeutic effects.

Intelligent response for LCRF: trigger factors and release characteristics

Due to the complex and ever-changing environmental conditions and diseases in agricultural production, the release performance of AIs from CRFs is susceptible to external influences. As a result, this often leads to a deviation in the release rate from the original design, which would weaken the benefits of the original formulation and reduce its efficacy. Therefore, there is a growing desire for AIs to exhibit targeted responsiveness to diseases or infestation occurrences. The controlled release of pesticides with an environmental intelligentresponse is another breakthrough in plant protection technology after CRS. Based on the differences in the release sources of trigger signals, response factors can be broadly categorized into biogenic responses (e.g., pH response, enzyme response, redox response) and environmental-source responses (e.g., temperature response, photo response, CO₂ response). Strategies for developing these two types of responsive formulations should have their own emphasis. Biogenic-responsive LCRFs should focus on the ability of the formulation to effectively penetrate pests or plants, because only after entering the body can the AIs be released by the trigger signal in the organism. It has been demonstrated that pesticide nanocarriers, such as lignin-based nanoparticles, can effectively penetrate

÷.
7:1
÷
6
52
ğ
20
3
Ę
0 U
bei
0
ail
Ψ
Ę,
led
)aC
Ы
M
Ă
i3
02
ė
q
en
N
ŭ
53
on
ģ
she
bli
Pa

Table 1 Typical lignin-based controlled release formulations

Ref.	s 49 ting tse	of 50	лс Л	ç		51		44	-	_	of 52		53	ç	42		tion 43	tion	Ļ	4c	in 55		56	6	46		for 57		58	ht 50	32	
Application properties	Drug release followed the Higuchi mode. 58% azadirachtin wa released in 550 h The encapsulated azadirachtin remained at 100% after irradia by sunlight for 192 h Lignin as a biosorbent in alginate was used to control the relev	rate and stabilize the botanical insecticide Reduced leaching of herbicides, thus mitigating the pollution	groundwater The cumulative release amounts ranged from 56 97% to 87 33	in 62 h The retention rates ranged from 46.67% to 63.41% after 50 h	ultraviolet irradiation Enhanced nlant surface adhesion and scourring resistance	Drug release followed the Fickian diffusion. The cumulative	The decomposition rate was 66% after 24 h of UV radiation	Drug release followed the Fickian diffusion. The cumulative	release amounts ranged from 22.4% to 34% in 72 h The retention rates of abscisic acid ranged from 34% to 75% i	60 h Hicklin official in motorine alon coollines from colinity choose	Highly enecuve in protecting necessed ings from saminy success Drug release followed the non-Ficklan diffusion. The amount	Als released was 49.4% within 60 n The degradation amount was 10% within 10 h	The cumulative release range was 45% after 5 days	High stability and encapsulation efficiency	The cumulative release amount within 120 n was 84.10%. The decomposition rate was less than 41% after 72 h of UV	irradiation	The cumulative release range was 35.5% at 72 h Greater than 75% of AIs could be retained after 60 h of irradia	Thermal stability has been improved, leading to a good inhibit	ut seeu gemmanon ann urought resistance penomiance	The cumulative release amount within /2 n was //% More than 85% of Als could be preserved after 96 h of UV	The cumulative release amounts ranged from 73.9% to 83.6%	40 h	Atter /U n 01 UV Irradiation, 88.9–93./% 01 AIS were retained The AIs remained at 15.7% by the end of the second week	High stability	The cumulative release proportion reached 96.6% at 4 h Evcellent control against soilborne finneal diseases and lower	residual risk	Drug release followed the Fickian diffusion. The time required	More than 50% remained after being exposed to UV light	irradiation for 30 min The 85% cumulative release amount took 72 h	The retention rate was up to 72% after being exposed to UV lig	As a wood preservative, it exhibited enhanced antifungal	resistance
Drug loading (%) Encapsulation efficiency (%)	2.81 99.5	9.24-14.27	85.64-99.41		a	а	а	20.66 - 26.15	61.97-78.31		62.6	93.3	a	77-97	16.80 61.49		/8	>70		42.34-03.19 85.31-96.25	11.2 - 20.6		61.2-/6.4 5.6	56.1	α >00		93.3-96.7	49.8-87.6	a	а	a	a
AI classes	Plant-derived insecticide	Herbicide	Insectivide and	acaricide		Fungicide		Plant growth	regulator		Insecticide		Fungicide		Insecuciae and acaricide	- 7	Plant growth regulator	0	Tuccoticide and	insecucide and acaricide	Insecticide and	acaricide	Funoricide	ann gun 1	Fungicide		Herbicide		Insecticide and	acaricide	Fungicide)
Active ingredients (AIs)	Azadirachtin	Metribuzin and	chloridazon Avermectin			Difenoconazole		Abscisic acid			Emamectin benzoate		Prothioconazole and	pyraclostrobin	Avermecun		Abscisic acid		A second of the	Avermecun	Avermectin		Dronieonazole	TOPROTISTON	Pyraclostrobin		Picloram		Avermectin		Propiconazole	
Raw materials	Kraft lignin, alginate	Kraft lignin, ethylcellulose, polvethylene glycol	Sodium lienosulfonate	cetyltrimethylammonium bromide (CTAB)		Benzoic acid esterified lignin sulfonates		Alkaline lignin, cationic surfactant			Alginate, chitosan, sodium lignosulfonate		Methacrylated lignin sulfonate		Lignin-based azo polymer		Alkalı lıgnın, cetyltrimethylammonıum bromide (CTAB)		Output and a limit minute of a limit	Quaternary ammonum ngnin, soulum dodecyl benzenesulfonate (SDBS)	Alkali lignin, crosslinker		Kraft lienin surfactants	Main against accaries	Sodium lignosulfonate		Chitosan, sodium lignosulfonate		Alkali lionin nolvurea		Kraft lignin)
Systems	Granules		Darticles																		Capsules											

The intelligent response of AIs in CRFs needs the support of intelligent materials. Although overlooked by many, lignin is actually a kind of intelligent material. The following section will illustrate the release triggers and mechanisms of intelligent-responsive LCRFs from the nature of lignin in combination with environmental changes in agricultural production.

pH response

Triggers of pH response in agricultural scenarios. The pH features of pests, bacterial colonies, and plants themselves offer a basis for setting triggers. The digestive tracts of different insect species exhibit specific pH values under long-term evolution due to the nutrients they feed on. Most lepidop-teran¹³ and coleopteran⁷¹ insects have relatively higher alka-line pH in their midguts. The intestinal lumen of *Drosophila* is alkali, while the central "stomach" of the midgut is naturally acidic.⁷² The pH value of different plant tissues also varies. Most plant tissues are weakly acidic, but phloem is weakly alkaline.^{73,74} Many plants are more vulnerable to phloem pathogens, most notably the Citrus Huanglongbing caused by *Candidatus* Liberibacter asiaticus.⁷⁵ The specific alkalinity of plant phloem can act as a trigger for pH-responsive pesticide release.

The interaction of environmental stress with the crop might also result in specific pH values. When some pathogenic bacteria infect the host, they secrete acidic chemicals like citric acid and gluconic acid, thus acidifying the host. It has been found that the process of environmental acidification caused by pathogenic bacteria facilitates the enhancement of their pathogenicity.⁷⁶ In response to biotic or abiotic stress, crops also release acidic root exudates, which are related to the initiation of plant defense mechanisms.⁷⁷

pH is the most representative and widely used stimulus. When designing intelligent materials, it is vital to examine not only the sensitivity of the material to the response, but also its specific usage scenarios to achieve the transition from laboratory to farmland.

Release characteristics of the LCRFs response to pH. Alkali lignin shows different solubility under different pH conditions, which is an important reason why lignin can be applied as a pH-responsive intelligent material. In an alkaline aqueous solution, acidic groups such as phenolic hydroxyl and carboxyl groups of alkali lignin are deprotonated and negatively charged. The electrostatic repulsion between these groups stretches the macromolecular network of lignin, causing it to dissolve in the solution (Fig. 3a).⁵⁹ The predomi-

Published on 23 noviembre 2023. Downloaded by Fail Open on 23/07/2025 9:17:13.

				Drug loading (%) Encapsulation		e
systems	Raw materials	Active ingredients (Als)	AI classes	efficiency (%)	Application properties	Ret.
Emulsions	Phenolated alkali lignin	Avermectin	Insecticide and acaricide	52–72 86–95	The 300 h-cumulative release rates reached about 45% The retention rate after UV irradiation for 48 h was 78.65% Hich viscosity for reduced drouler hounce	59
	Amino acid-modified enzymatic hydrolysis lignin, surfactants	Avermectin	Insecticide and acaricide	a a	Drug released followed the non-Fickian diffusion. 100% of Als were released after 6000 min The hishest residual level was 80.1% after UV radiation for 72 h	60
	Cellulolytic enzyme lignin or alkali lignin or organosolv lignin	Curcumin	Drug	a	The remaining curcumin content was greater than 70% after 36 h UV radiation	61
Coacervates	Sodium lignosulfonate, cationic surfactants	Abscisic acid	Plant growth regulator	a 40–90	More than 40% of Als remained after 72 h UV radiation Improved deposition performance on superhydrophobic leaves and high drought resistance of crops	62
¹ Not mentio	ned.					

This journal is © The Royal Society of Chemistry 2024



Fig. 3 pH-Responsive LCRFs: (a) mechanism of alkali-responsive LCRFs mediated by the solubility of lignin. Adapted with permission from ref. 59. Copyright 2022 from Elsevier. (b) Effects of pH on the release performance of avermectin (AVM) from AVM@(CH + SL)5. Adapted with permission from ref. 64. Copyright 2023 from Royal Society of Chemistry. (c) The pH stability of lignin-based microcapsules samples. Adapted from ref. 79. (d) Influence of the environmental pH on the release behavior of lignin-based emulsion. Adapted with permission from ref. 60. Copyright 2021 from American Chemical Society. (e) Release curve of avermectin nano-delivery system (Av-NDs) in 70% methanol with different pH values. Adapted with permission from ref. 80. Copyright 2021 from Springer Nature.

nant pH-responsive LCRFs commonly observed are alkalineresponsive releases. In the study by Wu et al.,41 porous lignin microspheres with bifunctional adsorption and controlled release were prepared by an inverse suspension copolymerization method. The lignin microspheres loaded with 2,4-dichlorophenoxyacetic acid showed an explosive release at pH = 11 with a release rate of 81.95% within 4 h. Conversely, it was only 25.78% at pH = 2 and achieved almost no release under neutral conditions. Under alkaline conditions, the microcapsules created by our group⁶⁴ through the layer-by-layer assembly of chitosan and lignin also displayed notable release characteristics (Fig. 3b). In addition to the solubility of lignin, the weaker interactions between the two materials under alkaline conditions were responsible for the abrupt release of AIs.⁶⁴ Plant phloem diseases, lepidopteran and coleopteran pests with alkaline intestinal tracts can be well prevented and controlled with alkali-responsive LCRFs.

There have been scattered reports on acid-responsive release, acid/alkali-responsive release, and neutral-responsive release by using lignin as intelligent material. The "cation– π "

effect is primarily responsible for the theory underlying the release of AIs in acidic environments.^{78,79} Lignin, as an aromatic polymer, interacts with a large amount of H^+ (and other possible cations) in acidic solutions, favoring the formation of extended conformations, leading to the disassembly of LCRF and the release of AIs (Fig. 3c).^{78,79} Chen *et al.*⁶⁰ developed a lignin-based emulsion for encapsulating avermectin (AVM), where the release of AVM under neutral conditions was relatively low (Fig. 3d). This is because the carboxylic acid groups in lignin ionize in a medium with a pH of 7.5, generating a strong electrostatic repulsion that maintains the stability of the emulsion.⁶⁰ Microspheres with the highest release rate of AVM under neutral conditions have also been reported (Fig. 3e).⁸⁰

The pH value also influences the intermolecular forces within the LCRF, thereby controlling the release of AIs.^{81,82} Furthermore, grafting pH-sensitive polymers onto lignin is an excellent strategy for preparing pH-responsive LCRFs.^{83,84}

Enzyme response

Triggers of enzyme response in agricultural scenarios. Enzymatic triggers play a pervasive role in agricultural production, rendering their application in intelligent agrochemical delivery highly beneficial. This strategy capitalizes on the exceptional efficiency and specificity of enzymatic reactions. Pests, plant pathogenic fungi, and bacteria can all create a variety of enzymes to gain nutrients from the outside world. Phytophagous insects possess a diverse array of digestive enzymes in their gastrointestinal tract. These enzymes can be categorized into distinct groups based on their digestive targets, such as amylase, protease, lipase, and cellulase.^{85–87} In addition, during the process of infecting host plants, phytopathogenic fungi and bacteria secrete a series of cell walldegrading enzymes such as pectinase, cellulase, glycosidase, and xylanase, which are conducive to the invasion, colonization, and expansion of pathogens.^{88,89} Research studies have reported on the use of zein or α-cyclodextrin as carriers in CRF that can be triggered for release by proteases or α -amylase in the gut of insects.^{90,91}

Various enzymes with lignin-degrading functions have been recognized, including lignin peroxidase, manganese peroxidase, versatile peroxidase, laccase, and others.⁹² Lignindegrading enzymes catalyze the cleavage of a series of lignin linkages, and play an essential role in the efficient utilization of lignin.^{93,94} Among them, laccase is a digestive enzyme that is present in the gut of lepidopteran species,⁹⁵ and is also abundant in fungus secretions.⁹⁶ Taking advantage of the fact that lignin can be degraded by laccases, many enzyme-responsive LCRFs have been designed, which will be discussed in detail below.

Release characteristics of the LCRFs response to enzymes. In enzyme-responsive drug-loading systems, two mechanisms are typically used in design applications. The first mechanism entails the conversion of prodrugs into active pharmaceutical compounds facilitated by specific enzymes. This transformation enables the desired therapeutic effects, and is commonly observed in medical delivery systems.⁹⁷ The second mechanism involves the degradation of controlled-release matrix by specific enzymes, thereby achieving targeted drug delivery. Most of the reported mechanisms for enzyme-responsive LCRF are of the latter kind. Specifically, enzymes destroy the drugloading system by degrading lignin itself or lignin grafts, thereby leading to the release of AIs.

In enzyme-responsive LCRFs, the most commonly used triggering enzymes are laccases. Wurm and coauthors have conducted several years of research to develop pesticide formulations for the treatment of the grapevine trunk disease Esca.⁹⁸⁻¹⁰⁰ Esca is a very contagious and devastating fungal disease caused by a fungus that secretes lignin-degrading enzymes like laccase. A combination of miniemulsion polymerization and solvent evaporation was used to produce lignin nanocarriers. The fungicides encapsulated by these nanocarriers were released only when infected, while no leakage of AIs was detected during storage or in the absence of infection (Fig. 4a and c). Zhang et al. have successfully developed lignin-based nanogel¹⁰¹ and nanocapsule⁴⁸ suspensions as a means to combat lepidopteran pests. These formulations display selective reactivity towards the endogenous laccases present in these pests. In the latter research study, the authors



Fig. 4 Enzyme-responsive LCRFs: (a) *in vitro* investigations of lignin nanocarriers (NC). Pyr stands for pyraclostrobin. Adapted with permission from ref. 98. Copyright 2019 from Wiley. (b) Effects of laccase on the release performance of avermectin (AVM) from (CH + SL)20@PE. Adapted with permission from ref. 65. Copyright 2023 from Wiley. (c) Concept of the nanocarriers-mediated drug delivery. Adapted with permission from ref. 98. Copyright 2019 from Wiley. (d) Schematic diagram of pH/laccase-responsive release of AVM-loaded microcapsules (CH + SL)n@PE in pests. Adapted with permission from ref. 65. Copyright 2023 from Wiley.

ingeniously introduced polysaccharides into the nanocapsule suspension to optimize the drug delivery system, thereby conferring responsiveness to both pectinases and cellulases present within the digestive tracts of pests. *In vivo* experiments have conclusively shown the accelerated release of AIs from the carrier composed of lignin and polysaccharides.⁴⁸

Researchers often combine enzyme response with pH response, and these dual-responsive LCRFs exhibit enhanced targeted lethality towards lepidopteran pests. Zhang *et al.*¹⁰² have established lignin nanoparticles with core–shell structures through electrostatic self-assembly, which accelerates the release of lambda-cyhalothrin under both alkaline conditions and the presence of laccases, with the latter triggering a more pronounced release. Our group⁶⁵ has utilized the alternating deposition of chitosan and lignosulfonate on an alkali lignin-based Pickering emulsion to form microcapsules. Similarly, these microcapsules display dual responsiveness to pH and laccase. The shell thickness of the microcapsule can be adjusted through the deposition process, thereby enabling the tunable release behavior of the AIs (Fig. 4b and d).

In addition to degrading lignin itself, the enzymatic degradation of lignin-grafted materials can be used to prepare enzyme-responsive LCRFs.¹⁰³ The high efficiency of the enzymatic reaction enables the rapid release of AIs when necessary, minimizing the negative "lag-time effect".

Temperature response

Triggers of temperature response in agricultural scenarios. The aforementioned reaction triggers are derived from the characteristics of crops, pests, and weeds, as well as their interactions, which exhibit strong response signals. In contrast, ambient temperature, being a climatic variable, does not experience significant fluctuations in a short period, yet some elements are highly sensitive to even minuscule changes of temperature. On the one hand, the severity of disease, pest, and weed damage is affected by temperature. A rise in ambient temperature may cause weeds to germinate earlier and increase the severity of weed damage. Large temperature fluctuations will also promote some weed germination.¹⁰⁴ For diseases, each "plant-pathogen" interaction has an optimal temperature range. For example, 15 °C is the optimum temperature for cyst nematodes to infect potatoes,¹⁰⁵ and rice blast is prone to epidemics at 24-26 °C and below.¹⁰⁶ Temperature can influence disease incidence by impacting the behavior of vector insects as well.¹⁰⁷ On the other hand, the infestation of diseases, pests, and weeds can cause temperature changes. Symptoms of scab disease on apple leaves appear to be associated with disruption of the apple cuticle by V. inaequalis conidiophores, resulting in a localized reduction in leaf temperature.¹⁰⁸ For wheat stripe rust, irrespective of the crop growth stages, the canopy mean temperature increases with the disease progression.¹⁰⁹ In comparison, changes in foliar temperature caused by crop infestation by pathogenic microorganisms such as fungi and viruses are more significant. Based on this change, researchers have built remote sensing technology to detect plant diseases non-destructively.^{110,111}

When designing temperature-responsive LCRFs, two temperature values of lignin deserve special attention: the glass transition temperature (T_g) and the critical solution temperature. Lignin itself exhibits a high T_g , ranging from 90 °C to 145 °C, depending on the source and isolation process of lignin.²⁶ By tuning T_g , lignin can be designed as different shape memory materials for smart applications, including vitrimer materials.¹¹² The critical solution temperature is generally divided into the upper critical solution temperature (LCST). Limited by the relatively high T_g , researchers tend to exploit the critical solution temperature of lignin for the development of intelligent LCRFs.

Currently, most temperature-responsive polymers are LCSTtype. Lignin-based LCST intelligent materials are primarily prepared by grafting classical temperature-sensitive polymers.¹¹³ The research on UCST is relatively less extensive compared to the widespread studies on LCST. Alcohol/water solvent mixtures are considered as promising solvents for various polymers to obtain UCST behavior.114 Related works have looked at the UCST behavior of lignin in ethanol/water mixed solvents, pointing out that it is the result of the combined action of the hydrogen bond and hydrophobic force of lignin.¹¹⁵ High temperature breaks the hydrogen bonds between lignin molecules, while also reducing the polarity of the ethanol/water mixed solvent. Consequently, the hydrophobic force of the solvent on lignin is stronger than that between lignin, resulting in lignin solvation. Thus, lignin slowly dissolves during the heating process in ethanol/water, changing from two-phase to singlephase, and the UCST behavior occurs (Fig. 5a).¹¹⁵ However, the UCST behavior of lignin is solvent-dependent, and it does not exhibit UCST behavior in pure water.116

Release characteristics of the LCRFs response to temperature. Lin et al.¹¹⁵ revealed the UCST behavior of lignin in ethanol/water mixtures, and applied it to the controlled release of pesticides. The anti-solvent method was used to prepare enzymatic hydrolysis lignin microspheres loaded with AVM, and the lignin retained its UCST behavior after microsphere formation via microstructure regulation. Experiments revealed that at 25 °C, only 20% of avermectin was released. However, at 50 °C, this value was up to 90% (Fig. 5b). In fact, this research can be optimized further because agricultural production under natural conditions rarely reaches 50 °C. Shen et al.¹¹⁷ fabricated lignin nanospheres for encapsulating AVM. Under xenon lamp-replicated solar radiation, the nanosphere solution undergoes photothermal conversion, rapidly heating up. Consequently, the structure of lignin nanospheres gradually swelled and even dissolved, releasing AVM in an ethanol/ water solution (Fig. 5c). Experimental evidence demonstrates that these nanospheres exhibit better pest control efficacy at 32 °C compared to 12 °C.

It has been reported that graft polymerization of lignin with NIPAAm can form thermosensitive hydrogels.^{118,119} The LCST of NIPAAm is close to the natural temperature, which is approximately 32 °C.¹²⁰ Furthermore, the LCST of the lignin–NIPAAm copolymer can be flexibly adjusted according to the



Fig. 5 Temperature-responsive LCRFs: (a) mechanism of UCST behavior of enzymatic hydrolysis lignin (EHL) in 50% (v/v) ethanol/water. (b) Drug release of avermectin-loaded EHL microspheres at different temperatures. Adapted with permission from ref. 115. Copyright 2021 from American Chemical Society. (c) The process from encapsulation to the release of avermectin (AVM) by lignin nanospheres. Adapted with permission from ref. 117. Copyright 2022 from Royal Society of Chemistry.

lignin content. This adaptability enables the copolymer to effectively respond to diverse agricultural application scenarios, explaining its growing popularity in recent years. However, the NIPAAm monomer exhibits cytotoxicity.¹²¹ Thus, its effects on crops, non-target organisms, and the environment must be carefully investigated in order to develop green and safe temperature-sensitive LCRFs.

Promising and underexploited intelligent LCRFs: potentials and ideas

The microenvironment of crop growth is closely linked and mutually influenced by disease occurrence. In addition to the aforementioned environmental triggering factors, light intensity, redox potential, gas concentration, and other factors are key components of the microenvironment for crop growth (Fig. 6). Understanding the variation patterns between the two can help develop the spatiotemporal targeting LCRFs. It should be pointed out that these response triggers may have been explored in other fields. However, there are few publications on the LCRFs. Lignin can be used to create ligninbased intelligent materials through grafting or crosslinking with responsive polymers. Additionally, its wide availability and rich functional groups make the biopolymer a promising candidate for exploring its own potential as an intelligent material. The following section will delve into the application



Fig. 6 Various intelligent lignin-based controlled release formulations (LCRFs) activated by different stimuli used in agricultural production.

potential and research directions of intelligent LCRFs activated by other stimuli.

content.¹¹⁷ The inherent excellent optical properties of lignin can revitalize photo-responsive LCRFs.

Photo-responsive LCRFs

Photo-responsive intelligent systems for the controlled release of pesticides have been widely reported in recent years, with a wide range of application settings.^{122,123} In one aspect, photosynthesis is a unique and vital physiological and biochemical process in green plants. By utilizing photosynthesis-inhibiting herbicides that only release when the weeds are exposed to sunlight, the effectiveness of the pesticide application can be greatly enhanced.¹⁹ In another aspect, light trapping technology for pest control is one of the main methods used in physical pest management. By combining photo-responsive pesticide delivery systems with light-trapping devices, pests can be attracted and eradicated simultaneously. Photo-responsive CRF also holds great application prospects in greenhouses equipped with lighting devices.

The molecular structure of lignin contains numerous aromatic rings and conjugated functional groups, resulting in strong conjugation and π - π interactions between molecules. These characteristics grant lignin a range of unique optical properties, including aggregation-induced luminescence, UV absorption, and sustainable photothermal conversion, among others.^{124,125} Several reports have explored the coupling of lignin with photosensitive groups,¹²⁶⁻¹²⁸ some of which revealed its potential for preparing photo-responsive materials.¹²⁶ However, its application in drug delivery is relatively scarce.

Photo-responsive LCRFs can be further converted into temperature-responsive LCRFs due to the photothermal conversion action of lignin,¹²⁴ as introduced in the preceding

Redox-responsive LCRFs

Many factors influence redox reactions in agricultural output. For crops, most abiotic and biotic stresses induce changes in cellular redox homeostasis, and increased the generation and accumulation of reactive oxygen species (ROS).¹²⁹ Correspondingly, pests and pathogens have evolved various strategies to evade plant defense systems. Among them, some insects maintain the reducing state of the midgut by secreting antioxidants like glutathione.¹³⁰ It has been found that glutathione accumulates in many plant–pathogen interactions to minimize the adverse impacts of ROS on cells.¹³¹

Utilizing redox reactions to trigger the release of AIs in the context of drug delivery systems is a well-established approach. A promising strategy to induce this response involves incorporating disulfide bonds within the carriers. Glutathione, present in the environment, can act as a "scissor" to cleave the disulfide bonds in pesticide carriers, acting as an intelligent "switch" to release AIs. It has been reported that disulfide bonds are inserted into lignin modified with polyetheramine as an adhesive,¹³² while LCRFs with disulfide bonds have not been widely reported.

Lignin possesses antioxidant properties due to the capacity of its phenolic groups to scavenge free radicals.¹³³ When lignin undergoes oxidative reactions, some covalent bonds will break and its molecular weight will decrease.¹³⁴ In fact, the enzymatic degradation mechanism of lignin is also the process of oxidative degradation.^{93,94} By employing the characteristic feature of lignin that is prone to decomposition in the presence of oxidative conditions caused by ROS generated from crops under stress, there is promising potential for the development of redox-responsive LCRFs. This novel approach shows great potential in enhancing crop resilience in the face of stressful conditions.

Gas-responsive LCRFs

Present studies in the field of gas-responsive systems predominantly concentrate on CO₂, and CO₂ triggers are of interest because of their cost-effectiveness, widespread availability, ecological and biological compatibility, as well as the ease with which they can be eliminated.¹³⁵ CO₂ is also a key factor influencing agricultural production. In the storage of grains, elevated levels of CO2 indicate the presence of mold spoilage or pest activity within the grain.¹³⁶ If fungicides/pesticides can respond to the ambient CO₂ concentration, it will effectively ensure the safety of grain storage. Besides pesticides, other gas-responsive agrochemicals are also worth considering. CO₂ is a necessary raw material in agricultural photosynthesis. CO2 fertilization, a process that stimulates plant growth by increasing CO₂ concentration,¹³⁷ has been widely used in greenhouse cultivation. Studies have shown that the growth of plant biomass is limited by low levels of nitrogen and phosphorus when CO₂ fertilization is implemented.¹³⁸ Therefore, nitrogen or phosphorus fertilizers can be applied together with CO₂ fertilization to enhance its efficiency. The development of CO₂responsive nitrogen and phosphorus fertilizers could reduce the labor-intensive nature of greenhouse horticulture.

 CO_2 -switchability is induced by the presence of CO_2 -functional groups such as amidine, tertiary amine, imidazole, guanidine, or carboxylic acid, and lignin is advantageous for grafting these groups to elicit the response.¹³⁵ DEAEMA-grafted lignin nanoparticles can be easily dispersed in water by CO_2 bubbling, and swiftly precipitated out by N₂ bubbling. Lignin-based Pickering emulsions made with these particles can switch between emulsified and demulsified states by bubbling different gases, which has great potential for drug delivery applications.¹³⁹ Moreover, CO_2 dissolved in water produces carbonic acid, so the gas response initiated by CO_2 can be further turned into a pH response.

Challenges coupled and future outlook

In order to better capitalize on the remarkable features of LCRFs and better serve the need of actual agricultural production, the selection of pesticide formulations is crucial. In recent years, there have been numerous reports on novel and functional LCRFs, including seed coating,¹⁴⁰ trunk injection,^{98–100} liquid mulching film,¹⁴¹ controlled-release adsorbent,⁴¹ and super-spreading agent.⁶² These advancements have expanded the application scenarios of lignin-based CRS. Pesticide formulations are currently evolving towards broad functionality, labor-saving efficiency, and enhanced pesticide utilization (Fig. 7).



Fig. 7 Versatile lignin-based controlled release system.

In recent years, the widespread application of plant protection unmanned aerial vehicle (UAV) has seen rapid progress in hardware and flight control systems. However, there exists a significant deficiency in the corresponding aerial spraying agents. Compared to conventional ground spraying, UAV application is characterized by low volume, high concentration, fine mist, elevated operational altitude, and extended spraying distance. These factors make it more susceptible to issues like evaporation, photolysis, and splashing, leading to ineffective application and higher frequency of drug damage accidents. LCRFs have the potential to reduce unintended losses when applying pesticides through conventional spraying methods, making them valuable assets in aerial crop protection. Numerous studies support this claim by demonstrating that certain LCRF systems have high viscosity, which promotes better adhesion of the agents to leaf surfaces.^{59,101} Additionally, some LCRFs form a topological structure with the wax layer on foliage surfaces, making them more resistant to rainfall.⁵¹ It is important to acknowledge that emerging LCRF systems based on lignin-based coacervates possess exceptional wetting and spreading properties of their own. These properties largely arise from the internal disordered network structure of coacervates and the entanglement of micro/nano structures on the surfaces of leaves, as previously mentioned.62 Overall, the use of LCRFs can enhance the precise application characteristics of intelligent-responsive systems, ultimately leading to improved efficiency in delivering pesticides to the target areas. In addition to aerial spray adjuvant, looking ahead, pesticide formulations combining drugs and fertilizers are a promising labor-saving agent. Researchers should drive pesticide formulation innovation based on practical agricultural production needs, promoting sustainable and environmentally friendly practices in the field of agriculture.

The research and development of LCRFs is still under continuous exploration, with several pressing issues awaiting resolution: (1) to enhance the reproducibility and repeatability of LCRF products, there is a need for an economical, ecofriendly, and efficient pretreatment technique, which would extract structurally ideal and stable lignin from pollutants or lignocellulose. (2) Although some intelligent-responsive LCRFs have been developed, further validation under field conditions is essential. This validation aims to confirm whether this responsiveness truly synchronizes with pest infestations, adapting to the intricate and ever-changing realities of agricultural production. (3) Tracing the remaining amounts of AIs in LCRF is crucial. This information is vital for farmers to determine when to replenish drugs. However, there is a lack of reports on this aspect in the existing literature. Comprehensive laboratory investigations are a necessary prerequisite for the commercial application of new products.

Before LCRFs can be widely commercialized, it is essential to subject them to rigorous toxic evaluation. The toxicity of new products often stands as a primary concern for the public. Being a natural substance, the basic ecotoxicity of lignin is not a problem as such. The problem may arise from necessary lignin modifications during the production process and the addition of surfactants or other substances. Indeed, many studies have evaluated the impact of LCRFs on seed vitality,¹⁴² plant growth,142 as well as their acute toxicity to natural insect predators and aquatic organisms^{48,65,102,143} under laboratory conditions. All of these studies concluded that lignin-based agrochemical encapsulation systems do not pose negative ecological impacts, and even partially exhibit enhanced safety quotient. Nonetheless, despite these findings, larger-scale studies of the long-term effects of LCRFs on ecosystems under actual field conditions should be necessary. Additionally, the occupational chronic toxicity of LCRFs to agricultural workers, and their potential accumulation in human cells through the food chain deserve attention.

Another pertinent issue pertaining to the commercial utilization of LCRFs is their costs. Given the critical nature of agriculture as a primary sector, responsible companies should prioritize the provision of affordable products to farmers in order to safeguard their livelihoods. Simultaneously, these companies should also bear the negative external costs associated with pesticides, which encompass the contamination of drinking water, loss of biodiversity, and potential effects on human health.¹⁴⁴ Fortunately, the conversion from lignin to high-value materials is known to be commercially competitive.²⁵ Substantial economic returns not only facilitate the successful transition of products from laboratory settings to actual production plants, but also drive the continuous innovation and improvement of factory-produced goods. Therefore, it is imperative for companies, research institutions, and all relevant stakeholders to collaboratively address these issues, and enable LCRFs to make significant contributions to the overall well-being of humanity.

Conclusions

Lignin polymers, due to their cost-effectiveness, green carbon footprint, tunable properties, and resistance to UV radiation, are considered ideal raw materials for constructing CRFs. As

summarized in this review, numerous innovative LCRF systems have been developed and refined, resulting in improved effectiveness and environmental friendliness. The development of intelligent-responsive LCRFs is considered a noteworthy advancement in the field of plant protection, as it contributes to the progress of precision agriculture and lowcarbon development. In this system, AIs are released based on the timing and location of disease occurrence, thereby increasing pesticide utilization efficiency while minimizing its harmful impact on the environment and non-target organisms. Many of the examples discussed in this review demonstrate that the sensitivity to various environmental stimuli allows lignin to be employed as an intelligent material in the construction of environmentally responsive LCRFs. In summary, LCRFs are a new pesticide formulation that is flourishing, experiencing a leap from sustained release to intelligent response, and will do much more in the foreseeable future.

Author contributions

Yitong Wang: conceptualization, writing – original draft. Xiaona Yu: conceptualization, writing – review & editing. Shuaishuai Ma: writing – review & editing. Shuling Cao: writing – review & editing. Xufeng Yuan: supervision. Wanbin Zhu: funding acquisition, supervision. Hongliang Wang: conceptualization, writing – review & editing, funding acquisition, supervision.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the National Key R&D Program of China (2021YFA0716700), National Natural Science Foundation of China (22278422), the 2115 Talent Development Program of China Agricultural University Fund (1011-00109018), and Beijing Innovation Team of the Modern Agricultural Research System (BAIC08-2023-FQ02).

Notes and references

- M. Carvajal-Yepes, K. Cardwell, A. Nelson, K. A. Garrett, B. Giovani, D. G. O. Saunders, S. Kamoun, J. P. Legg, V. Verdier, J. Lessel, R. A. Neher, R. Day, P. Pardey, M. L. Gullino, A. R. Records, B. Bextine, J. E. Leach, S. Staiger and J. Tohme, *Science*, 2019, **364**, 1237–1239.
- 2 S. He and K. M. Creasey Krainer, *Mol. Plant*, 2020, 13, 933–934.

- 3 Y. Zhang, B. Y. Liu, K. X. Huang, S. Y. Wang, R. L. Quirino, Z. X. Zhang and C. Q. Zhang, *ACS Appl. Mater. Interfaces*, 2020, **12**, 37607–37618.
- 4 M. F. A. Jallow, D. G. Awadh, M. S. Albaho, V. Y. Devi and B. M. Thomas, *Sci. Total Environ.*, 2017, 574, 490–498.
- 5 X. Guo, C. Wang and F. Zhang, *Front. Agric. Sci. Eng.*, 2022, **9**, 511–522.
- 6 A. I. Sankoh, R. Whittle, K. T. Semple, K. C. Jones and
 A. J. Sweetman, *Environ. Int.*, 2016, 94, 458–466.
- 7 A. Sharma, A. Shukla, K. Attri, M. Kumar, P. Kumar,
 A. Suttee, G. Singh, R. P. Barnwal and N. Singla, *Ecotoxicol. Environ. Saf.*, 2020, 201, 110812.
- 8 A. Wang, A. Padula, M. Sirota and T. J. Woodruff, *Fertil. Steril.*, 2016, **106**, 905–929.
- 9 E. V. R. Campos, J. L. de Oliveira, L. F. Fraceto and B. Singh, *Agron. Sustainable Dev.*, 2015, **35**, 47–66.
- A. Singh, N. Dhiman, A. K. Kar, D. Singh, M. P. Purohit,
 D. Ghosh and S. Patnaik, *J. Hazard. Mater.*, 2020, 385, 121525.
- 11 T. Li, D. Teng, R. Mao, Y. Hao, X. Wang and J. Wang, J. Biomed. Mater. Res., Part A, 2019, 107, 2371–2385.
- 12 A. Chevillard, H. Angellier-Coussy, V. Guillard, N. Gontard and E. Gastaldi, *J. Hazard. Mater.*, 2012, **205–206**, 32–39.
- 13 Y. Gao, Y. Zhang, S. He, Y. Xiao, X. Qin, Y. Zhang, D. Li, H. Ma, H. You and J. Li, *Chem. Eng. J.*, 2019, 364, 361–369.
- 14 J. Tang, G. Ding, J. Niu, W. Zhang, G. Tang, Y. Liang, C. Fan, H. Dong, J. Yang, J. Li and Y. Cao, *Chem. Eng. J.*, 2019, **359**, 225–232.
- 15 B. Liu, J. Zhang, C. Chen, D. Wang, G. Tian, G. Zhang, D. Cai and Z. Wu, J. Agric. Food Chem., 2021, 69, 6981–6988.
- 16 D. Zhang, B. Li, X. Zhang, Z. Zhang, W. Wang and F. Liu, J. Agric. Food Chem., 2016, 64, 2841–2846.
- 17 S. V. Prudnikova, A. N. Boyandin, G. S. Kalacheva and A. J. Sinskey, *J. Polym. Environ.*, 2013, 21, 675–682.
- 18 S. Kumar, G. Bhanjana, A. Sharma, M. C. Sidhu and N. Dilbaghi, *Carbohydr. Polym.*, 2014, **101**, 1061–1067.
- 19 Z. Ye, J. Guo, D. Wu, M. Tan, X. Xiong, Y. Yin and G. He, *Carbohydr. Polym.*, 2015, **132**, 520–528.
- 20 Y. Li, M. Zhou, Y. Pang and X. Qiu, *ACS Sustainable Chem.* Eng., 2017, 5, 3321–3328.
- 21 R. Tao, C. You, Q. Qu, X. Zhang, Y. Deng, W. Ma and C. Huang, *Environ. Sci.: Nano*, 2023, **10**, 351–371.
- 22 H. Wang, Y. Pu, A. Ragauskas and B. Yang, *Bioresour. Technol.*, 2019, **271**, 449–461.
- 23 J. Xu, C. Li, L. Dai, C. Xu, Y. Zhong, F. Yu and C. Si, *ChemSusChem*, 2020, **13**, 4284–4295.
- 24 D. S. Bajwa, G. Pourhashem, A. H. Ullah and S. G. Bajwa, *Ind. Crops Prod.*, 2019, **139**, 111526.
- 25 X. Yu, B. Yang, W. Zhu, T. Deng, Y. Pu, A. Ragauskas and H. Wang, *Ind. Crops Prod.*, 2023, **200**, 116824.
- 26 A. Moreno and M. H. Sipponen, *Mater. Horiz.*, 2020, 7, 2237–2257.
- 27 Z. Sun, B. Fridrich, A. de Santi, S. Elangovan and K. Barta, *Chem. Rev.*, 2018, **118**, 614–678.
- 28 Y. Zhang and M. Naebe, ACS Sustainable Chem. Eng., 2021, 9, 1427–1442.

- 29 M. Tortora, F. Cavalieri, P. Mosesso, F. Ciaffardini, F. Melone and C. Crestini, *Biomacromolecules*, 2014, **15**, 1634–1643.
- 30 M. Tuomela, M. Vikman, A. Hatakka and M. Itävaara, *Bioresour. Technol.*, 2000, **72**, 169–183.
- 31 W. Yang, E. Fortunati, D. Gao, G. M. Balestra, G. Giovanale, X. He, L. Torre, J. M. Kenny and D. Puglia, ACS Sustainable Chem. Eng., 2018, 6, 3502–3514.
- 32 R. C. Andeme Ela, S. H. Chipkar, T. L. Bal, X. Xie and R. G. Ong, ACS Sustainable Chem. Eng., 2021, 9, 2684– 2692.
- 33 D. Kai, M. J. Tan, P. L. Chee, Y. K. Chua, Y. L. Yap and X. J. Loh, *Green Chem.*, 2016, 18, 1175–1200.
- 34 S. Sugiarto, Y. Leow, C. L. Tan, G. Wang and D. Kai, *Bioact. Mater.*, 2022, 8, 71–94.
- 35 J. V. Cotterill, R. M. Wilkins and F. T. Da Silva, J. Controlled Release, 1996, 40, 133–142.
- 36 M. Fernández-Pérez, E. González-Pradas, M. D. Ureña-Amate, R. M. Wilkins and I. Lindup, *J. Agric. Food Chem.*, 1998, 46, 3828–3834.
- 37 J. Cao, R. H. Guenther, T. L. Sit, S. A. Lommel, C. H. Opperman and J. A. Willoughby, *Cellulose*, 2016, 23, 673–687.
- 38 J. Rao, A. N. Chandrani, A. Powar and S. Chandra, *Green Chem. Lett. Rev.*, 2021, 14, 204–220.
- 39 M. Fernández-Pérez, M. Villafranca-Sánchez, F. Flores-Céspedes and I. Daza-Fernández, *Carbohydr. Polym.*, 2011, 83, 1672–1679.
- 40 F. J. Garrido-Herrera, I. Daza-Fernández, E. González-Pradas and M. Fernández-Pérez, *J. Hazard. Mater.*, 2009, 168, 220–225.
- 41 H. Wu, L. Gong, X. Zhang, F. He and Z. Li, *Chem. Eng. J.*, 2021, **411**, 128539.
- 42 Y. Deng, H. Zhao, Y. Qian, L. Lü, B. Wang and X. Qiu, *Ind. Crops Prod.*, 2016, 87, 191–197.
- 43 J. Yin, H. Wang, Z. Yang, J. Wang, Z. Wang, L. Duan, Z. Li and W. Tan, *J. Agric. Food Chem.*, 2020, 68, 7360–7368.
- 44 J. Yin, M. Quan, Z. Wang, J. Wang, Z. Yang, L. Duan, Z. Li, Q. X. Li, H. Wang and W. Tan, *Ind. Crops Prod.*, 2022, **177**, 114573.
- 45 R. Peng, D. Yang, X. Qiu, Y. Qin and M. Zhou, Int. J. Biol. Macromol., 2020, 151, 421–427.
- 46 J. Luo, D.-x. Zhang, T. Jing, G. Liu, H. Cao, B.-x. Li, Y. Hou and F. Liu, *Chem. Eng. J.*, 2020, **394**, 124854.
- 47 S. Kumar, M. Nehra, N. Dilbaghi, G. Marrazza,
 A. A. Hassan and K.-H. Kim, *J. Controlled Release*, 2019, 294, 131–153.
- 48 D. Zhang, R. Wang, C. Ren, Y. Wang, B. Li, W. Mu, F. Liu and Y. Hou, ACS Appl. Mater. Interfaces, 2022, 14, 41337– 41347.
- 49 F. Flores-Céspedes, G. P. Martínez-Domínguez, M. Villafranca-Sánchez and M. Fernández-Pérez, *J. Agric. Food Chem.*, 2015, 63, 8391–8398.
- 50 F. Flores-Céspedes, I. Daza-Fernández, M. Villafranca-Sánchez, M. Fernández-Pérez, E. Morillo and T. Undabeytia, *J. Hazard. Mater.*, 2018, 343, 227–234.

- 51 W. Liang, J. Zhang, F. R. Wurm, R. Wang, J. Cheng, Z. Xie, X. Li and J. Zhao, *Int. J. Biol. Macromol.*, 2022, 220, 472– 481.
- 52 G. Huang, G. Qian, Y. Yan, D. Xu, C. Xu, L. Fu and B. Lin, *React. Funct. Polym.*, 2020, **146**, 104429.
- 53 S. Beckers, S. Peil and F. R. Wurm, ACS Sustainable Chem. Eng., 2020, 8, 18468–18475.
- 54 Y. Li, D. Yang, S. Lu, S. Lao and X. Qiu, *J. Agric. Food Chem.*, 2018, **66**, 3457-3464.
- 55 Y. Pang, Y. Sun, Y. Luo, M. Zhou, X. Qiu, C. Yi and H. Lou, *Ind. Crops Prod.*, 2021, **167**, 113468.
- 56 R. C. Andeme Ela, M. Tajiri, N. K. Newberry, P. A. Heiden and R. G. Ong, ACS Sustainable Chem. Eng., 2020, 8, 17299–17306.
- 57 X. Wang and J. Zhao, J. Agric. Food Chem., 2013, 61, 3789– 3796.
- 58 Y. Pang, X. Li, S. Wang, X. Qiu, D. Yang and H. Lou, *React. Funct. Polym.*, 2018, **123**, 115–121.
- 59 X. Yu, S. Chen, W. Wang, T. Deng and H. Wang, *J. Cleaner Prod.*, 2022, **339**, 130769.
- 60 K. Chen, S. Yuan, D. Wang, Y. Liu, F. Chen and D. Qi, *Langmuir*, 2021, 37, 12179–12187.
- 61 H. Zhang, F. Yue, S. Hu, H. Qi and F. Lu, Int. J. Biol. Macromol., 2023, 228, 178–185.
- 62 J. Wang, Y. Fan, H. Wang, J. Yin, W. Tan, X. Li, Y. Shen and Y. Wang, *Chem. Eng. J.*, 2022, **430**, 132920.
- 63 T. He, X. Xu, B. Ni, H. Lin, C. Li, W. Hu and X. Wang, Angew. Chem., Int. Ed., 2018, 57, 10148– 10152.
- 64 X. Yu, J. Wang, X. Li, S. Ma, W. Zhu and H. Wang, *Mater. Adv.*, 2023, 4, 1089–1100.
- 65 X. Yu, X. Li, S. Ma, Y. Wang, W. Zhu and H. Wang, *Adv. Funct. Mater.*, 2023, **33**, 2214911.
- 66 B. D. Mattos, B. L. Tardy, W. L. E. Magalhães and O. J. Rojas, *J. Controlled Release*, 2017, 262, 139–150.
- 67 P. Fernandez, V. André, J. Rieger and A. Kühnle, *Colloids Surf.*, A, 2004, 251, 53–58.
- 68 J. Wu and G.-H. Ma, Small, 2016, 12, 4633-4648.
- 69 M. Stanisz, Ł. Klapiszewski, M. N. Collins and T. Jesionowski, *Mater. Today Chem.*, 2022, **26**, 101198.
- 70 O. E. Mendez, C. E. Astete, R. Cueto, B. Eitzer, E. A. Hanna, F. Salinas, C. Tamez, Y. Wang, J. C. White and C. M. Sabliov, *J. Agric. Food Res.*, 2022, 7, 100259.
- 71 T. Lemke, U. Stingl, M. Egert, M. W. Friedrich and A. Brune, *Appl. Environ. Microbiol.*, 2003, **69**, 6650–6658.
- 72 C. T. J. Ferguson, A. A. Al-Khalaf, R. E. Isaac and O. J. Cayre, *PLoS One*, 2018, 13, e0201294.
- 73 D. Mendoza Cozatl, E. Butko, F. Springer, J. Torpey, E. Komives, J. Kehr and J. Schroeder, *Plant J.*, 2008, 54, 249–259.
- 74 B. J. Shelp, J. Exp. Bot., 1987, 38, 1619–1636.
- 75 C.-Y. Huang, K. Araujo, J. N. Sánchez, G. Kund, J. Trumble, C. Roper, K. E. Godfrey and H. Jin, *Proc. Natl. Acad. Sci. U. S. A.*, 2021, **118**, e2019628118.
- 76 D. Prusky, J. L. McEvoy, R. Saftner, W. S. Conway and R. Jones, *Phytopathology*, 2004, 94, 44–51.

- I. Lager, O. Andréasson, T. L. Dunbar, E. Andreasson, M. A. Escobar and A. G. Rasmusson, *Plant, Cell Environ.*, 2010, 33, 1513–1528.
- 78 L. Zongo, H. Lange and C. Crestini, *ACS Omega*, 2019, 4, 6979–6993.
- 79 M. Sgarzi, M. Gigli, C. Giuriato and C. Crestini, *Materials*, 2022, **15**, 1857.
- 80 D. Mo, X. Li, Y. Chen, Y. Jiang, C. Gan, Y. Zhang, W. Li, Y. Huang and J. Cui, *Sci. Rep.*, 2021, **11**, 23248.
- 81 J. Cui, D. Mo, Y. Jiang, C. Gan, W. Li, A. Wu, X. Li, J. Xiao, Q. Hu, H. Yuan, R. Lu and Y. Huang, *Ind. Eng. Chem. Res.*, 2019, 58, 19741–19751.
- 82 G. Huang, Y. Deng, Y. Zhang, P. Feng, C. Xu, L. Fu and B. Lin, *Chem. Eng. J.*, 2021, **403**, 126342.
- 83 D. Yiamsawas, W. Kangwansupamonkon and S. Kiatkamjornwong, *Eur. Polym. J.*, 2021, 145, 110241.
- 84 N. Chen, L. A. Dempere and Z. Tong, ACS Sustainable Chem. Eng., 2016, 4, 5204–5211.
- 85 G. Tokuda, in Advances in Insect Physiology, ed. R. Jurenka, Academic Press, Okinawa, 2019, vol. 57, pp. 97–136.
- 86 M. Hafeez, X. W. Li, J. M. Zhang, Z. J. Zhang, J. Huang, L. K. Wang, M. M. Khan, S. K. Shah, G. M. Fernandez-Grandon and Y. B. Lu, *Insect Sci.*, 2021, 28, 611–626.
- 87 A. Filipović, M. Mrdaković, L. Ilijin, A. Grčić, D. Matić, D. Todorović, M. Vlahović and V. Perić-Mataruga, *Comp. Biochem. Physiol., Part C: Toxicol. Pharmacol.*, 2021, 249, 109123.
- 88 C. P. Kubicek, T. L. Starr and N. L. Glass, *Annu. Rev. Phytopathol.*, 2014, **52**, 427–451.
- 89 K. Hématy, C. Cherk and S. Somerville, *Curr. Opin. Plant Biol.*, 2009, **12**, 406–413.
- 90 R. A. Monteiro, M. C. Camara, J. L. de Oliveira, E. V. R. Campos, L. B. Carvalho, P. L. de Freitas Proença, M. Guilger-Casagrande, R. Lima, J. do Nascimento, K. C. Gonçalves, R. A. Polanczyk and L. F. Fraceto, *J. Hazard. Mater.*, 2021, **41**7, 126004.
- 91 A. E. Kaziem, Y. Gao, Y. Zhang, X. Qin, Y. Xiao, Y. Zhang,
 H. You, J. Li and S. He, *J. Hazard. Mater.*, 2018, 359, 213–221.
- 92 C. Weng, X. Peng and Y. Han, *Biotechnol. Biofuels*, 2021, 14, 84.
- 93 G. Janusz, A. Pawlik, U. Świderska-Burek, J. Polak, J. Sulej, A. Jarosz-Wilkołazka and A. Paszczyński, *Int. J. Mol. Sci.*, 2020, 21, 966.
- 94 G. Janusz, A. Pawlik, J. Sulej, U. Świderska-Burek, A. Jarosz-Wilkołazka and A. Paszczyński, *FEMS Microbiol. Rev.*, 2017, 41, 941–962.
- 95 Z. Liu, H. Wang and C. Xue, J. Integr. Agric., 2018, 17, 2310–2319.
- 96 Z. Lu, J. Deng, H. Wang, X. Zhao, Z. Luo, C. Yu and Y. Zhang, *Environ. Microbiol.*, 2021, 23, 1256–1274.
- 97 R. Walther, J. Rautio and A. N. Zelikin, *Adv. Drug Delivery Rev.*, 2017, **118**, 65–77.
- 98 J. Fischer, S. J. Beckers, D. Yiamsawas, E. Thines, K. Landfester and F. R. Wurm, *Adv. Sci.*, 2019, 6, 1802315.

- 99 T. O. Machado, S. J. Beckers, J. Fischer, B. Müller,
 C. Sayer, P. H. H. de Araújo, K. Landfester and
 F. R. Wurm, *Biomacromolecules*, 2020, 21, 2755–2763.
- 100 S. Peil, S. J. Beckers, J. Fischer and F. R. Wurm, *Mater. Today Bio*, 2020, 7, 100061.
- 101 D. Zhang, R. Wang, H. Cao, J. Luo, T. Jing, B. Li, W. Mu,
 F. Liu and Y. Hou, *Colloids Surf.*, B, 2022, 209, 112166.
- 102 D. Zhang, J. Du, R. Wang, J. Luo, T. Jing, B. Li, W. Mu, F. Liu and Y. Hou, Adv. Funct. Mater., 2021, 31, 2102027.
- 103 Y. Jiang, Y. Chen, D. Tian, F. Shen, X. Wan, L. Xu, Y. Chen,
 H. Zhang, J. Hu and F. Shen, *Soft Matter*, 2020, 16, 9083–9093.
- 104 V. Nichols, N. Verhulst, R. Cox and B. Govaerts, *Field Crops Res.*, 2015, **183**, 56–68.
- 105 L. M. Jones, A. K. Koehler, M. Trnka, J. Balek, A. J. Challinor, H. J. Atkinson and P. E. Urwin, *Global Change Biol.*, 2017, 23, 4497–4507.
- 106 J. Qiu, J. Xie, Y. Chen, Z. Shen, H. Shi, N. I. Naqvi, Q. Qian, Y. Liang and Y. Kou, *Mol. Plant*, 2022, **15**, 723–739.
- 107 A. C. Velásquez, C. D. M. Castroverde and S. Y. He, *Curr. Biol.*, 2018, 28, R619–R634.
- 108 E. C. Oerke, P. Fröhling and U. Steiner, *Precis. Agric.*, 2011, 12, 699–715.
- 109 R. N. Singh, P. Krishnan, V. K. Singh and K. Banerjee, *Ecol. Inform.*, 2022, **71**, 101774.
- 110 J. Zhang, Y. Huang, R. Pu, P. Gonzalez-Moreno, L. Yuan, K. Wu and W. Huang, *Comput. Electron. Agric.*, 2019, 165, 104943.
- 111 V. Singh, N. Sharma and S. Singh, *Artif. Intell. Agric.*, 2020, 4, 229–242.
- 112 S. Zhang, T. Liu, C. Hao, L. Wang, J. Han, H. Liu and J. Zhang, *Green Chem.*, 2018, **20**, 2995–3000.
- 113 R. Liu, T. Ding, P. Deng, X. Yan, F. Xiong, J. Chen and Z. Wu, *Int. J. Biol. Macromol.*, 2022, **194**, 358–365.
- 114 Q. Zhang and R. Hoogenboom, *Chem. Commun.*, 2015, 51, 70–73.
- 115 Y. A. Lin, Y. X. Pang, Z. P. Li, M. S. Zhou, H. M. Lou and X. Q. Qiu, *ACS Sustainable Chem. Eng.*, 2021, **9**, 15634–15640.
- 116 Y. Qian, Y. H. Deng and X. Q. Qiu, *Adv. Mater. Res.*, 2012, **550–553**, 1321–1326.
- 117 F. Shen, S. Wu, M. Huang, L. Zhao, J. He, Y. Zhang, S. Deng, J. Hu, D. Tian and F. Shen, *Green Chem.*, 2022, 24, 5242–5254.
- 118 P. Jiang, Y. Cheng, S. Yu, J. Lu and H. Wang, *Polymers*, 2018, **10**, 1109.
- 119 P. Jiang, X. R. Sheng, S. Yu, H. M. Li, J. Lu, J. H. Zhou and H. S. Wang, *Sci. Rep.*, 2018, 8, 14450.
- 120 A. Halperin, M. Kroger and F. M. Winnik, *Angew. Chem., Int. Ed.*, 2015, **54**, 15342–15367.
- 121 A. S. Wadajkar, B. Koppolu, M. Rahimi and K. T. Nguyen, J. Nanopart. Res., 2009, 11, 1375–1382.
- 122 X. Xu, B. Bai, H. Wang and Y. Suo, ACS Appl. Mater. Interfaces, 2017, 9, 6424–6432.
- 123 C. Chen, G. Zhang, Z. Dai, Y. Xiang, B. Liu, P. Bian, K. Zheng, Z. Wu and D. Cai, *Chem. Eng. J.*, 2018, 349, 101–110.

- 124 X. Jin, X. Li, X. Liu, L. Du, L. Su, Y. Ma and S. Ren, *Int. J. Biol. Macromol.*, 2023, **228**, 528–536.
- 125 J. Li, W. Liu, X. Qiu, X. Zhao, Z. Chen, M. Yan, Z. Fang, Z. Li, Z. Tu and J. Huang, *Green Chem.*, 2022, 24, 823–836.
- 126 Y. Deng, Y. Liu, Y. Qian, W. Zhang and X. Qiu, *ACS Sustainable Chem. Eng.*, 2015, **3**, 1111–1116.
- 127 H.-Y. Tse, S.-C. Cheng, C. S. Yeung, C.-Y. Lau, W.-H. Wong, C. Dong and S.-Y. Leu, *Green Chem.*, 2019, 21, 1319–1329.
- 128 H.-Y. Tse, C. S. Yeung, C. Y. Lau, M. Y. Cheung, J. Guan, M. K. Islam, P. T. Anastas and S.-Y. Leu, *Green Chem.*, 2022, 24, 2904–2918.
- 129 M. A. Farooq, A. K. Niazi, J. Akhtar, Saifullah, M. Farooq, Z. Souri, N. Karimi and Z. Rengel, *Plant Physiol. Biochem.*, 2019, **141**, 353–369.
- 130 J. Dong, W. Chen, J. Feng, X. Liu, Y. Xu, C. Wang, W. Yang and X. Du, *ACS Appl. Mater. Interfaces*, 2021, **13**, 19507–19520.
- 131 G. Noctor, A. Mhamdi, S. Chaouch, Y. I. Han, J. Neukermans, B. Marquez-Garcia, G. Queval and C. H. Foyer, *Plant, Cell Environ.*, 2012, 35, 454–484.
- 132 W. Liu, C. Fang, F. Chen and X. Qiu, *ChemSusChem*, 2020, 13, 4691–4701.
- 133 T. Dizhbite, G. Telysheva, V. Jurkjane and U. Viesturs, *Bioresour. Technol.*, 2004, **95**, 309–317.
- 134 B. Jiang, Y. Zhang, H. Zhao, T. Guo, W. Wu and Y. Jin, Int. J. Biol. Macromol., 2019, 139, 21–29.
- 135 A. Darabi, P. G. Jessop and M. F. Cunningham, *Chem. Soc. Rev.*, 2016, **45**, 4391–4436.
- 136 A. Müller, M. T. Nunes, V. Maldaner, P. C. Coradi, R. S. D. Moraes, S. Martens, A. F. Leal, V. F. Pereira and C. K. Marin, *Rice Sci.*, 2022, 29, 16–30.
- 137 C. Huntingford and R. J. Oliver, Nature, 2021, 600, 224-225.
- 138 C. Terrer, R. B. Jackson, I. C. Prentice, T. F. Keenan, C. Kaiser, S. Vicca, J. B. Fisher, P. B. Reich, B. D. Stocker, A. Hungate, J. Peñuelas, I. McCallum, в. N. A. Soudzilovskaia, L. A. Cernusak, A. F. Talhelm, K. Van Sundert, S. Piao, P. C. D. Newton, M. J. Hovenden, D. M. Blumenthal, Y. Y. Liu, C. Müller, K. Winter, C. B. Field, W. Viechtbauer, C. J. Van Lissa, M. R. Hoosbeek, M. Watanabe, T. Koike, V. O. Leshyk, H. W. Polley and O. Franklin, Nat. Clim. Change, 2019, 9, 684-689.
- 139 Y. Qian, Q. Zhang, X. Qiu and S. Zhu, *Green Chem.*, 2014, 16, 4963–4968.
- 140 Y. Gu, Y. Pang, D. Yang, Y. Qian, H. Lou and M. Zhou, *Ind. Crops Prod.*, 2022, **182**, 114877.
- 141 D. Tian, J. Zhang, J. Hu, M. Huang, L. Zhao, Y. Lei, J. Zou, S. Zhang and F. Shen, *Chem. Eng. J.*, 2023, **452**, 139383.
- 142 T. Kacsó, E. A. Hanna, F. Salinas, C. E. Astete, E. Bodoki, R. Oprean, P. P. Price, V. P. Doyle, C. A. R. Bonser, J. A. Davis and C. M. Sabliov, *Appl. Nanosci.*, 2022, 12, 1557–1569.
- 143 N. Zhao, L. Zhu, M. Liu, L. He, H. Xu and J. Jia, *J. Agric. Food Chem.*, 2023, **71**, 3790–3799.
- 144 N. G. Shakhramanyan, U. A. Schneider and B. A. McCarl, *Clim. Change*, 2013, **117**, 711–723.