


Cite this: *RSC Adv.*, 2022, 12, 35469

Adsorption of cationic surfactant as a probe of the montmorillonite surface reactivity in the alginate hydrogel composites†

Danielle Silva do Nascimento, Mariana Etcheverry, Angie E. Orduz, Carolina V. Waiman  and Graciela P. Zanini *

Adsorption of a cationic surfactant allowed to probe the surface reactivity of montmorillonite encapsulated in a composite of alginate hydrogels (A-MMT). Dodecylbenzyltrimethylammonium chloride (BAC-12) was the surfactant used for these studies. BAC-12 is part of the widely used surfactant mixture known as benzalkonium chloride. XRD showed that up to three different types of basal spacing (d_{001}) were present within the composite indicating that as the concentration of adsorbed BAC-12 increases, populations with different adsorption conformational arrangements are present, even unexpanded clay remains. From the SEM-EDS spectra it is observed that the clay is distributed in the whole composite. In addition, the effect of the presence of cationic and anionic biocides on BAC-12 adsorption was studied. Cationic biocides such as tetradecylbenzyltrimethylammonium chlorides (BAC-14) and paraquat (PQ) show a competitive behavior for the clay adsorption sites at BAC-12 low concentration indicating an electrostatic adsorption mechanism. However, the presence of anionic contaminants such as 2,4-D and metsulfuron methyl do not affect surfactant adsorption. In all scenarios is observed an abrupt increase of BAC-12 adsorbed amount reaching values higher than the clay CEC suggesting strong tail–tail interactions. This occurs at concentrations 10 times lower than the CMC of BAC-12 promoted by clay encapsulation in the composite. In these composites the alginate does not affect the surface reactivity of the clay, but the formation of the hydrogel allows it to be easily extracted from aqueous media which makes it an interesting material with a potential use in water remediation.

Received 21st November 2022
Accepted 7th December 2022

DOI: 10.1039/d2ra07405b

rsc.li/rsc-advances

Introduction

Adsorption is one of the most important technologies used to retain pollutants for water remediation and purification purposes. The development of future adsorbents depends on the knowledge of the role of each component, the interactions between adsorbate and adsorbent and/or the adsorption mechanisms.

Composites formed by inorganic solids and biopolymers are among the materials that have been studied for water pollutant retention.^{1–4} Regarding the inorganic solids, the clays as montmorillonite (MMT) are commonly applied for cationic pollutant retention due to these clays show mainly structural negative charges.⁵ Besides, MMT is ubiquitous in nature, nontoxic, environmental-friendly and low cost.^{6,7} It has large specific surface area, exhibits high cation exchange capacity and excellent adsorbent ability.^{8–12} Since clay forms stable suspensions, the incorporation of MMT into biopolymer matrices

simplifies the separation procedures compared to the use of natural clays.¹³ Alginate is a biopolymer widely used in synthesizing materials with different shapes due to its ability to form hydrogels in the presence of multivalent cations such as Ca^{2+} .^{14–18} In addition, MMT improves the mechanical properties of alginate hydrogels.

Alginate-montmorillonite beads (A-MMT) are composites that consist of spheres around 3 mm in diameter which have been used as adsorbents for the removal of heavy metals,¹⁹ dyes^{20,21} and herbicides from aqueous media.²² However, most of the information available in the literature reports the percentage of adsorption, the maximum amount adsorbed and the isotherm fittings and not the interactions of the contaminants with the material components.

Among the pollutants widely discharged into the environment are the surfactants. These substances are commonly used in personal care products, hospital and household cleaners, laundry detergent, oil recovery industries, agriculture and nanotechnology.^{23–25} After use, residual surfactants can reach surface waters or groundwater, and end up dispersed in soil, water or sediments.^{26,27}

There are different types of surfactants; however, the most used are the ionic ones.²⁴ In this regard, benzalkonium chloride

INQUISUR (UNS-CONICET), Departamento de Química, Universidad Nacional del Sur, Av. Alem 1253, B8000CPB-Bahía Blanca, Argentina. E-mail: gzanini@uns.edu.ar

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d2ra07405b>



(BAC) consists of a group of cationic surfactants that are highly effective as antimicrobial agents, thereby ranking as one of the most popular antiseptics and disinfectants in the world.^{28,29} In terms of chemistry, BAC is a mix of alkylbenzyltrimethylammonium chlorides having different alkyl chain lengths (C8–C18). Most of the biocidal activity is associated with the C12–C14 derivatives, which are the main constituents of the mix.^{30,31} In addition, BAC is an active ingredient in common disinfectants recommended for use as prophylaxis protocol for viruses such as SARS-CoV-2.³² In consequence, it has been noted that after their use, BACs accumulate in public facilities, transportation, hospital effluents, wastewater treatment plants, and even common households.³³

Due to their molecular structure, BACs interact with negatively charged surfaces as MMT.^{34–38} The interaction between cationic surfactant and clay minerals such as MMT is well documented but lately most of these studies have focused on the synthesis of organoclays in order to modify clay surface reactivity.^{39–41} However, the interest of our research is to study these surfactants as contaminants dissolved in water.

Although composites formed by alginate and clay have been studied as adsorbents of pollutants dissolved in water, there is no information on their use in surfactant adsorption and even less on such adsorption is affected by other cationic and anionic substances. In addition, there is a lack of information on the adsorption process inside the material.

The novel purpose of this work is to study the surface reactivity of the clay inside the bead through the adsorption of BAC-12. In addition, the work evaluates how this adsorption process is affected by the presence of other biocides (cationic and anionic) and by the encapsulation of the clay in a bead-like composite.

Experimental

Materials and methods

Sodium alginate biopolymer (CAS: 9005-38-3, $M_w = 231.5 \text{ g mol}^{-1}$) was purchased from Fluka. Na-montmorillonite (MMT, 99.4% purity) was obtained from Lago Pellegrini (Rio Negro, Argentina). The CEC of clay is 0.91 mEq g^{-1} and this clay was fully described by Lombardi *et al.*⁴² According to these authors, the MMT structural formula is $[(\text{Si}_{3.87}\text{Al}_{0.13})(\text{Al}_{1.44}\text{Fe}^{3+}_{0.27}\text{Mg}_{0.29})\text{O}_{10}(\text{OH})_2]\text{Na}^{+}_{0.42}$ where Na^{+} is the exchangeable cation.^{34,43}

Surfactants BAC-12 and BAC-14 were purchased from Fluka (CAS: 139-07-1; 139-08-2, 99.0% purity). For the studies, 5 mM stock solutions at $\text{pH } 6.0 \pm 0.2$ of BAC-12 and BAC-14 were prepared in 0.01 M NaCl. The critical micellar concentration (CMC) was determined by dynamic light scattering with a Zetasizer ZS90 (Malvern, UK) by Zanini *et al.* obtaining values around 3.8 mM and 1.18 mM for BAC-12 and BAC-14, respectively (0.01 M NaCl).²⁸

Herbicides. Metsulfuron methyl, MM (CAS: 74223-64-6, 99.3% purity, pK_a : 3.3), 2,4-dichlorophenoxyacetic acid, 2,4-D (CAS: 94-75-7, 99.3% purity, pK_a : 2.98) and paraquat, PQ (CAS: 75365-73-0, 98.0% purity) were purchased from Riedel-de Haën, Sigma-Aldrich. The 10 mM stock solutions at $\text{pH } 6.0 \pm 0.2$ were prepared in 0.01 M NaCl. $\text{pH } 6.0 \pm 0.2$ was adjusted for all

solutions using small volumes of HCl and/or NaOH solutions (1 M). pH measurements were performed by using a Cole Parmer Model 59003-25 pH meter with an Alpha® model PY-41 electrode.

The molecular structures of all contaminants are shown in Table S1 of the ESI†. Considering the pK_a values of 2,4-D and MM their anionic species are predominant at working pH; on the other hand, BAC-12, BAC-14 and PQ are quaternary ammonium salts, therefore they only form cations in aqueous solution.⁴⁴

Calcium chloride $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ (CAS: 10035-04-8, 99.0% purity) was supplied by Sigma-Aldrich Company. All other chemicals were analytical grade.

Preparation alginate-clay composite beads

Briefly, 100 mL of a 1% (w/v) alginate solution was mixed with 4 g of MMT and stirred at room temperature. This mixture was added drop by drop through a micropipette tip using a peristaltic pump to a reservoir containing 0.1 M CaCl_2 under moderate magnetic stirring. Each drop produces a spherical hydrogel which was referred to as an A-MMT composite bead. The beads were left in this solution for 30 min to harden. A-MMT were separated and rinsed with distilled water and stored in NaCl 0.1 M until use. Simultaneously, alginate beads were synthesized following the same steps without the addition of clay, these hydrogels were named AA beads. Fig. S1 (ESI†) shows digital photographs of A-MMT and AA beads.

Characterization

Fourier transform infrared (FTIR) spectra were recorded between $4000\text{--}400 \text{ cm}^{-1}$ using KBr pellets by Nicolet Nexus 470 FTIR spectrometer equipped with a DTGS detector. Thermogravimetric analysis (TGA) was carried out with the aid of STDQ600 Thermal Analysis system operated in air. Each powder sample was placed in a ceramic crucible. A constant heating rate of $10 \text{ }^\circ\text{C min}^{-1}$ was applied from 20 to $1000 \text{ }^\circ\text{C}$. The internal structure of the lyophilized beads was examined using a scanning electron microscopy (SEM-EDS) LEO microscope model EVO 40. Samples were analyzed at 10.0 kV. A-MMT composite beads were washed with doubly distilled water, ground to powder and dried at $30 \text{ }^\circ\text{C}$ for analysis by powder X-ray diffraction (XRD). XRD patterns of A-MMT beads and A-MMT beads with BAC-12 adsorbed were performed. Powder samples X-ray diffraction patterns data were collected using a PANalytical Empyrean 3 diffractometer with a Ni-filtered CuK_α radiation ($\lambda = 1.54 \text{ \AA}$) and a PIXcel3D detector. It was operated at a voltage of 45 kV and a current of 40 mA. Diffractograms were collected using a continuous scan mode with slit FDS of $1/16$ and FDS IBASS of $1/8^\circ$. The measurement conditions were step size: 0.0131303; time per step: 480 s; scan speed: 0.041683 and run time: 30 min for the 2θ range $2^\circ \leq 2\theta \leq 10^\circ$. The d -values of clay incorporated in the composite with and without BAC-12 adsorbed were calculated by using Bragg's equation ($n\lambda = 2d \sin \theta$).



Adsorption experiments

Adsorption experiments were performed by batch equilibration method. Briefly, four wet beads were placed in 15 mL polypropylene centrifuge tubes with 10 mL of surfactant and/or herbicide solutions. The initial concentrations (C_i) were from 0.025 to 0.625 mEq L⁻¹ in 0.01 M NaCl. Then, the tubes were shaken during 6 h at room temperature and pH was kept at 6.0 ± 0.2 using small volumes of 1 M HCl and NaOH solutions. Preliminary kinetic experiments showed that this period (6 h) is enough to reach equilibrium adsorption. BAC-12 adsorbed as a function of time is shown in Fig. S2 (ESI[†]). All experiments were carried out at room temperature (25 ± 1 °C) and 0.01 M NaCl.

The adsorption experiments of BAC-12 on A-MMT were also performed at pH 4.0 ± 0.2 , 6.0 ± 0.2 and 8.0 ± 0.2 (see Fig. S3, ESI[†]). Since no significant changes were observed at the pH studied, it was selected pH 6.0 ± 0.2 for the rest of the experiments.

After equilibration reached, the beads were easily removed leaving a clear supernatant. Beads were kept for further analysis and the supernatants were withdrawn for surfactants and/or herbicides quantification.

For comparative purposes, BAC-12 adsorption onto MMT aqueous suspension was performed following experiments described in our previous work.³⁴

The concentrations of PQ, 2,4-D, MM, BAC-12 and BAC-14, in the supernatant were determined by UV-vis spectroscopy using an Agilent 8453 UV-vis diode array spectrophotometer. PQ, 2,4-D and MM were quantified at 258, 283 and 233 nm respectively, and BAC-12 and BAC-14 were quantified using the dye method proposed by Few *et al.*⁴⁵

The amounts of surfactant and herbicide adsorbed on A-MMT (Q_{ads} , in mEq g⁻¹) were calculated using the following equation:

$$Q_{\text{ads}} = \frac{(C_0 - C_{\text{eq}})V}{W}$$

where: C_0 and C_{eq} (mEq L⁻¹) are the pollutants initial and equilibrium concentrations respectively; V is the volume of the solution (L); and W is the weight of the dry beads (g). To determine W , ten groups of four dry beads were weighted. The average W obtained was 0.0050 ± 0.0002 grams.

Adsorption isotherms of BAC-12 in presence of BAC-14 (BAC-12:BAC-14), PQ (BAC-12:PQ), 2,4-D (BAC-12:2,4-D) and MM (BAC-12:MM) on A-MMT were performed as described above. In these experiments the adsorbates were added simultaneously, and proper volumes of stock solutions were mixed to obtain the concentration ratio (1 : 1), pH was kept at 6.0 ± 0.2 , the ionic strength was 0.01 M NaCl and the initial concentration range studied in these binary systems were from 0.025 to 0.625 mEq L⁻¹. The isotherms were performed in triplicate.

Results and discussion

Characterization of beads

Infrared spectroscopy. The FTIR spectra revealed the functional groups of alginate beads (AA), MMT clay and A-MMT

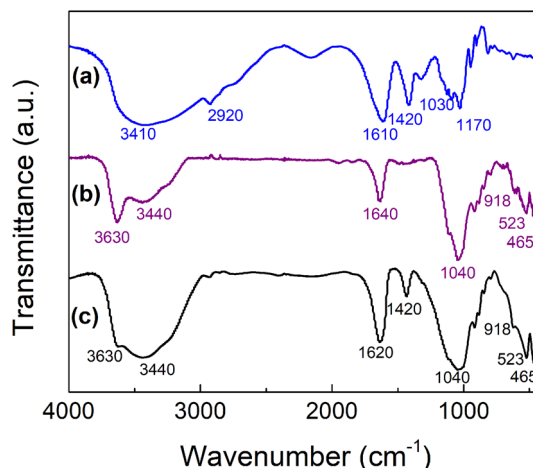


Fig. 1 FTIR spectra of (a) AA bead (blue line), (b) MMT (purple line) and (c) A-MMT bead (black line).

beads (Fig. 1). As can be seen in Fig. 1a the FTIR spectrum of AA shows absorption bands at 3410 cm⁻¹ (OH stretching), 2920 cm⁻¹ (CH₂ stretching), 1610 cm⁻¹ (COO⁻ asymmetric stretching), and 1420 cm⁻¹ (COO⁻ symmetric stretching). The bands that occur at 1170–1130 cm⁻¹ can be attributed to the presence of O–C–O bonds typical of polysaccharide rings.^{14,46} Fig. 1b shows the characteristic bands for MMT, the band at 3630 cm⁻¹ corresponds to the stretching mode of OH band of Si–OH. The broad band near 3440 cm⁻¹ due to –OH stretching for interlayer absorbed H₂O and the band at 1640 cm⁻¹ due to –OH deformation of H₂O. The band observed at 1040 cm⁻¹ is attributed to the asymmetric stretching vibrations of Si–O–Si, the band at 918 cm⁻¹ is attributed to Al–Al–OH bending vibrations. Furthermore, the band at 523 cm⁻¹ corresponds to Si–O–Al deformation and the band at 465 cm⁻¹ corresponds to bending vibrations of Si–O–Si.^{47,48} The FTIR spectrum of A-MMT beads (Fig. 1c) shows that the bands at 1610 and 1420 cm⁻¹ corresponding to COO⁻ stretching of AA are shifted to higher wavenumbers occurring at 1620 and 1422 cm⁻¹ respectively.^{22,49} According to Amarasinghe *et al.*, this behavior could be attributed to the electrostatic interaction of the carboxyl group of alginate with the MMT edge positive charge.⁵⁰ Alginate interacts with clay edge hydroxyl groups, which brings about many contact sites and formed a three-dimensional network.^{50,51}

Thermal analysis. TGA analyses of AA beads, MMT clay and A-MMT beads were performed to determine the thermal stability of the beads over a given heating profile (Fig. S4, ESI[†]). TGA and DTG curves show a weight-decreasing pattern as a function of temperature and the maximum temperature needed for the complete thermal degradation of the sample, respectively. Generally, the thermal decomposition of polysaccharides exhibits three main thermal events as can be seen in Fig. S4a.† The first event that can be seen in the range of 40 to 160 °C is associated with the loss of physically absorbed water and removal of structural water.⁵² The second event occurs in the temperature range of 250–350 °C due to depolymerization, decomposition of alginate carbon chains and formation to intermediates products of sodium carbonate (Na₂CO₃). At the



end, the third event involves the decomposition of the Na_2CO_3 at 550°C .^{53,54}

As can be seen in Fig. S4b,† MMT clay shows two steps. The first step at $30\text{--}100^\circ\text{C}$ is attributed to the loss of free water and the second weight loss at $550\text{--}700^\circ\text{C}$ is due to the loss of structural OH .⁵⁵

For A-MMT beads, the curves show an initial weight loss up to 150°C suggesting that corresponds to the dehydration (Fig. S4c†). DTG curve shows that the thermal behavior of the A-MMT beads seems to be a combination of the behavior of AA and MMT.²²

From Fig. S4† analysis it can be estimated that the residual weight percentages are 19.66, 67.97 and 84.15% for AA beads, A-MMT beads and MMT respectively after heating at 1000°C . From these results, it can be concluded that A-MMT beads exhibit an improvement of thermal stability compared with AA beads.

SEM analysis. The morphologies of AA and A-MMT composite beads were investigated by scanning electron

microscopy. Fig. 2 shows the SEM images of the beads. Fig. 2a, c and e (left side) show the cross-section of lyophilized AA beads at different magnification. The interior of the bead has thin walls that form holes giving rise to a structure known in the literature as “egg box”.⁵⁶ Fig. 2b, d and f (right side) show the cross-section of lyophilized A-MMT beads, it is observed that the inner structure of the bead is similar to the AA beads indicating that alginate is responsible for the structure of the bead inner holes.

The main difference between both composites can be seen by comparing the micrographs at higher magnification (Fig. 2e vs. 2f). Fig. 2e shows thin walls and smooth surfaces^{14,22} while Fig. 2f shows a rough surface.⁵⁷ The presence and distribution of MMT in the beads was confirmed by energy dispersive spectroscopy (EDS). Fig. S5 (ESI†) shows the EDS elemental mapping of A-MMT, the presence of elements such as Si and Al are observed both outside and inside the bead. Since these elements are the main constituents of clay, it can be assumed that MMT is distributed throughout the composite.



Fig. 2 SEM micrographs of cross-section of lyophilized beads at different magnifications AA bead (a) bar = $200\ \mu\text{m}$, (c) bar = $100\ \mu\text{m}$, (e) bar = $20\ \mu\text{m}$, A-MMT bead, (b) bar = $200\ \mu\text{m}$ (d) bar = $200\ \mu\text{m}$, and (f) bar = $20\ \mu\text{m}$.



XRD analysis. Fig. 3a shows the clay diffraction patterns within A-MMT with and without BAC-12 adsorbed (Q_{ads} from 0 to 0.91 mEq g^{-1}) and Fig. 3b shows a simple scheme that helps to understand the behavior of the clay in the composite as the surfactant is adsorbed. The roman numbers in Fig. 3a correspond to those in Fig. 3b.

A-MMT composite beads XRD pattern (Fig. 3a-I) shows a peak at $2\theta = 7.08^\circ$ corresponding to a clay basal spacing $d_{001} = 12.48 \text{ \AA}$. Comparing this value with $d_{001} = 12.3 \text{ \AA}$ of the MMT,³⁴ it is observed that the biopolymer increases only by $0.18 \text{ \AA}$ the laminar spacing of the clay. This indicates that the alginate does not fully intercalate between the layers but, as shown by FTIR spectra, the carboxyl groups could interact with the edge sites of the clay thus slightly increasing the d_{001} without modifying its layered structure.³⁴ Zhang *et al.* reported a similar increase in basal space for montmorillonite included in this type of composites.⁵⁸

Fig. 3a from II to V shows that the peak is shifted to lower angles as the adsorbed amount of BAC-12 increases indicating that the basal spacing increases due to the clay expansion by the intercalation of the surfactant. These experimental data are represented in a simple scheme in Fig. 3b. Table S2 (ESI†) shows the 2θ angle and its respective d_{001} for all the adsorbed amount of BAC-12 studied. Fig. 3a-II shows two peaks, one of them at $2\theta = 7.08^\circ$ and the other at $2\theta = 6.19^\circ$ corresponding to 12, 48 $\text{\AA}$ and 14.27 $\text{\AA}$ of d_{001} respectively. The presence of two peaks indicates that within the bead, BAC-12 is adsorbed on certain sites in the clay interlayer and does not have access to other ones, probably in the core of the bead. It is known from cationic dyes adsorption on A-MMT beads that adsorption occurs by dye

migration from bead surface to inner bead and that the extent of this migration depends on the initial concentration.⁵⁹ Therefore, at low concentrations of BAC-12 adsorbed two clay populations with different interlaminar spacing are observed. Fig. 3a-III shows three peaks at $2\theta = 7.08^\circ$, $2\theta = 6.14^\circ$ and $2\theta = 5.23^\circ$ corresponding to a basal spacing of 12.48, 14.27 and 16.89 $\text{\AA}$, respectively. This indicates that there are three clay populations present with different interlaminar spacing, two of them correspond to clay with adsorbed BAC-12 in different molecular arrangement and there are still interlaminar clay surfaces that have not been reached by the surfactant at these concentrations.^{35,60} The increase in interlayer spacing from 12.48 $\text{\AA}$ to 16.89 $\text{\AA}$ is similar to those found for BAC-12 adsorption on montmorillonite in aqueous suspension for similar adsorbed amount. Remarkably, this indicates that clay as part of the composite does not modify its surface reactivity.^{34,38} Fig. 3a-IV shows two peaks around $2\theta = 7.10^\circ$ and $2\theta = 5.07^\circ$ corresponding to a basal spacing around 12.48 $\text{\AA}$, and 17.3 $\text{\AA}$, respectively demonstrating that exist two clay populations. Fig. 3a-V shows only one peak at 7.10° . This indicates that there is a unique clay population and that every interlaminar spaces contain BAC-12 intercalated with the same conformation.

Adsorption isotherms

BAC-12 adsorption onto beads on A-MMT composite. Fig. 4 shows the adsorption isotherms of BAC-12 on MMT (blue diamonds) and on A-MMT composites beads (red squares). It can be observed that the adsorption isotherm on MMT increases abruptly at low concentrations and reached a plateau at 0.91 mEq g^{-1} which correspond to the cation exchange capacity (CEC) of the clay. This behavior is typical of an interaction mechanism with a strong electrostatic character.^{34,61–63} The BAC-12 adsorption isotherm on A-MMT shows that the surfactant maximum adsorbed amount overcomes CEC of the clay at the same initial concentrations studied, indicating the presence of

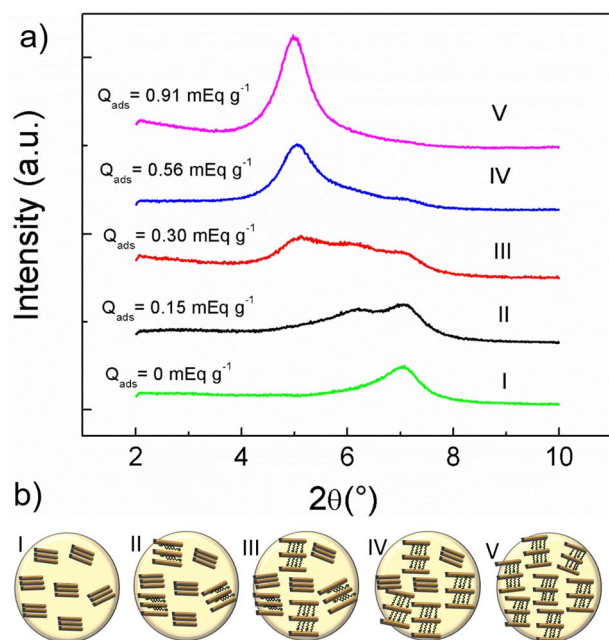


Fig. 3 (a) X-ray diffraction patterns of montmorillonite as a component of A-MMT beads with different amounts of BAC-12 adsorbed, (b) scheme showing montmorillonite expansion as a component of A-MMT beads.



Fig. 4 Adsorption isotherms of BAC-12 on MMT (blue diamonds) and of BAC-12 on A-MMT composites (red squares). Dashed line denotes the CEC value. The inset show regions (i), (ii), and (iii) for adsorption isotherm of BAC-12 on A-MMT composite.



other interactions besides the electrostatic one. The isotherm of BAC-12 on A-MMT can be analyzed by three defined regions that can be better visualized in the inset of the figure. In region (i) ($Q_{\text{ads}} = 0-0.30 \text{ mEq g}^{-1}$), the surfactant-clay interactions are predominantly electrostatic. In region (ii) ($Q_{\text{ads}} = 0.30-0.47 \text{ mEq g}^{-1}$), the experimental results seem to reach a plateau lower the CEC value of MMT possibly due to there are still some remaining interlaminal MMT surfaces that the surfactant was not able to access as indicated by the diffractogram patterns (Fig. 3). In region (iii) (Q_{ads} higher than 0.47 mEq g^{-1}) it can be seen an abrupt rise in the amount adsorbed that exceeds the CEC of the clay at C_{eq} around 0.1 mEq g^{-1} (C_i from 0.38 to 0.625 mM). This behavior often occurs due to bilayer formation and/or micellization as a consequence of surfactant tail-tail interactions.

It is noteworthy that although the CMC of the BAC-12 in aqueous solution is 3.8 mM the formation of bilayers or micelles on A-MMT composite beads occurs at lower initial concentration (C_i from 0.38 to 0.625 mM). Undoubtedly the clay encapsulated within A-MMT composite bead has a significant effect on the micellization of BAC-12.

Simultaneous adsorption of BAC-12 and BAC-14 on A-MMT composite. Fig. 5 shows BAC-12, BAC-14 and a mixture of BAC-12:BAC-14 ($1:1$) adsorption isotherms on A-MMT composite. The experimental results are clearly similar to that discussed for BAC-12 adsorbed on A-MMT (Fig. 4 red squares).

It is also observed in Fig. 5 that a strong tail-tail interaction occurs at equilibrium concentration around $C_{\text{eq}} = 0.10 \text{ mEq L}^{-1}$ corresponding to C_i around 0.38 mEq L^{-1} for both surfactants and for the mixture of them. Considering that these surfactants have different carbon chain length, it seems clear that the clay encapsulated in the bead promotes the micellization of surfactants.

Simultaneous adsorption of BAC-12 and herbicides on A-MMT composite. Fig. 6 shows the adsorption isotherms of BAC-12, PQ and (BAC-12:PQ) on A-MMT composites. The PQ adsorption isotherm on A-MMT (black circles) shows an abrupt increase at low C_{eq} to reach a plateau around 0.84 mEq g^{-1} , the same behavior was reported by Etcheverry *et al.*²²

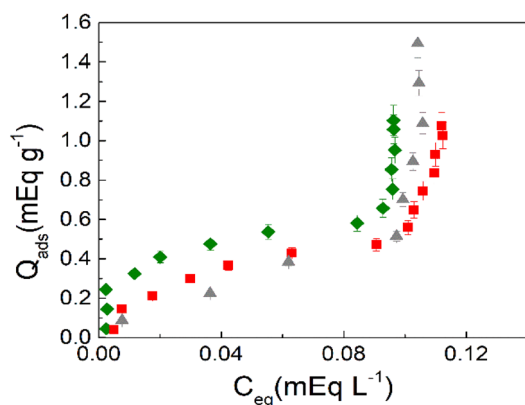


Fig. 5 Adsorption isotherms of surfactants on A-MMT composites. BAC-12 (red squares), BAC-14 (green diamonds) and BAC-12:BAC-14 (grey triangles).



Fig. 6 Adsorption isotherms on A-MMT composites. PQ (black circles), BAC-12 (red squares), PQ in presence of BAC-12 (empty circles) and BAC-12 in presence of PQ (empty squares).

The BAC-12 adsorption isotherm on A-MMT beads (red squares) was analyzed in Fig. 4. It is noticeable in Fig. 6 that at equilibrium concentrations lower than 0.10 mEq L^{-1} PQ adsorbed amounts are higher than BAC-12 adsorbed amounts, this suggests that PQ has more freedom to move within the bead and is therefore more probable to reach the surface of the clay. On the other hand, at equilibrium concentrations higher than 0.10 mEq L^{-1} the amount of adsorbed PQ does not increase, continuing the plateau reached around $C_{\text{eq}} = 0.05 \text{ mEq L}^{-1}$. This would indicate the absence of intermolecular interactions among the adsorbed herbicide molecules.

Comparing PQ adsorbed in the absence and presence of BAC-12, it is observed that the adsorbed amount of PQ in (BAC-12:PQ) decreases in the whole range of concentrations (empty circles), this indicates that the herbicide competes with the surfactant for the same adsorption sites. Comparing BAC-12 adsorbed in the absence and presence of PQ, a decrease in the adsorbed amount of BAC-12 in (BAC-12:PQ) is observed when the equilibrium concentrations are less than around 0.10 mEq L^{-1} . At higher values of this C_{eq} no changes in adsorption behavior are observed. These results indicate that, at low concentrations the adsorption of BAC-12 onto the composite is governed by electrostatic interactions while at higher concentrations the adsorption is governed by hydrophobic interactions of the surfactant tails. Fig. S6a (ESI†) shows the adsorption isotherms of BAC-12 in absence and presence of 2,4-D (BAC-12:2,4-D) and of MM (BAC-12:MM). As can be seen, BAC-12 shows the same behavior in presence and absence of anionic herbicides. These results indicate that 2,4-D and MM do not affect BAC-12 adsorption on A-MMT composites because they do not have affinity for the clay surface due to their negative charge. Fig. S6b† shows that neither 2,4-D nor MM adsorb on A-MMT. On the other hand, the inset in Fig. S6b† shows that the BAC-12 presence slightly affects the adsorption of 2,4-D at equilibrium concentrations above 0.36 mEq L^{-1} . This behavior may be due to the hydrophobic interaction of the aromatic ring

of 2,4-D with the surfactant tails. These types of interactions are responsible for the adsorption of organic molecules on organoclays.⁶⁴

Conclusions

This work explores through the adsorption of a cationic surfactant the surface reactivity of montmorillonite included in an alginate hydrogel composite. These studies demonstrated that the clay is capable of adsorbing cationic surfactants even when it is part of a composite. Furthermore, it is noteworthy that XRD was a powerful tool to elucidate the surface reactivity of the clay encapsulated in the hydrogel and thus understand the adsorption process of cationic substances inside the composite. Besides, the diffractograms showed that the 2θ angle of the clay remained at 7.08° ($d_{001} = 12.48 \text{ \AA}$) while other peaks, up to a total of 3, occurred as the adsorbed amount of surfactant increased. Finally, at the maximum concentration studied, only one peak was observed around 4.89° ($d_{001} = 17.91 \text{ \AA}$). This evidenced the presence of different clay populations inside the bead at different concentrations of adsorbed surfactant. At present, no similar results have been reported. In addition, it is important to note that the increase from $12.48 \text{ \AA}$ to $17.91 \text{ \AA}$ of the basal spacing coincides with that obtained in our previous work with the adsorption of the same surfactant on clay in aqueous suspension.

Although the reactivity of the clay within the composite is not affected, its encapsulation in a hydrogel promotes tail–tail interaction of the BAC-12 at much lower concentrations of its CMC.

Regarding the effect of the presence of cationic biocides on the adsorption of BAC-12, it was observed that at low concentrations they compete for the adsorption sites, indicating electrostatic interaction between the surfactant and the clay, while at high concentrations the competition does not take place since the hydrophobic interactions of this surfactant govern. As for the presence of anionic biocides, no appreciable changes in surfactant adsorption were observed.

The results of this article show that the alginate biopolymer allows the formation of composites in which the surface reactivity of the encapsulated solid retains its reactivity. This gives these composites a great potential for their use in water remediation.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors gratefully acknowledge the financial support of Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET; PIP 11220200102819CO), Ministerio de Ciencia, Tecnología e Innovación (PICT, 2020-SERIEA-03920) and Universidad Nacional del Sur (PGI-UNS-24/Q093). Danielle Silva do Nascimento thanks CONICET for postdoctoral fellowship.

Notes and references

- 1 M. Hassan, A. K. Deb, F. Qi, Y. Liu, J. Du, A. Fahy, M. A. Ahsan, S. J. Parikh and R. Naidu, *J. Cleaner Prod.*, 2021, **319**, 128694.
- 2 T. K. Das, Q. Scott and A. N. Bezbaruah, *Chemosphere*, 2021, **281**, 130837.
- 3 B. Ates, S. Koytepe, A. Ulu, C. Gurses and V. K. Thakur, *Chem. Rev.*, 2020, **120**, 9304–9362.
- 4 M. Hnamte and A. K. Pulikkal, *Chemosphere*, 2022, **307**, 135869.
- 5 M. J. Avena and C. P. De Pauli, *J. Colloid Interface Sci.*, 1998, **202**, 195–204.
- 6 C. V. Lazaratou, D. V. Vayenas and D. Papoulis, *Appl. Clay Sci.*, 2020, **185**, 105377.
- 7 C. Zhou, D. Tong and W. Yu, *Smectite nanomaterials: Preparation, properties, and functional applications*, Elsevier Inc., 2019.
- 8 H. Yang, W. Wang, J. Zhang and A. Wang, *Int. J. Polym. Mater. Polym. Biomater.*, 2013, **62**, 369–376.
- 9 M. Wu, S. Zhao, M. Tang, R. Jing, Y. Shao, X. Liu, Y. Dong, M. Li, Q. Liao, G. Lv, Q. Zhang, Z. Meng and A. Liu, *Colloids Surf., A*, 2019, **575**, 264–270.
- 10 F. Bergaya, B. K. G. Theng and G. Lagaly, *Handbook of Clay Science*, 2006, vol. 1.
- 11 M. Sarkar and K. Dana, *J. Mol. Struct.*, 2022, **1256**, 132468.
- 12 C. V. L. Natarelli, P. I. C. Claro, K. W. E. Miranda, G. M. D. Ferreira, J. E. de Oliveira and J. M. Marconcini, *SN Appl. Sci.*, 2019, **1**, 1212.
- 13 L. Pinto, M. A. Bonifacio, E. De Giglio, E. Santovito, S. Cometa, A. Bevilacqua and F. Baruzzi, *Food Packag. Shelf Life*, 2021, **28**, 100676.
- 14 R. da Silva Fernandes, M. R. de Moura, G. M. Glenn and F. A. Aouada, *J. Mol. Liq.*, 2018, **265**, 327–336.
- 15 Z. Tong, Y. Chen, Y. Liu, L. Tong, J. Chu, K. Xiao, Z. Zhou, W. Dong and X. Chu, *Mar. Drugs*, 2017, **15**, 91.
- 16 M. S. Hasnain and A. K. Nayak, *Alginate-inorganic composite particles as sustained drug delivery matrices*, Elsevier Inc., 2018.
- 17 I. P. S. Fernando, W. W. Lee, E. J. Han and G. Ahn, *Chem. Eng. J.*, 2020, **391**, 123823.
- 18 C. V. Waiman, J. Natera, W. A. Massad and G. P. Zanini, *Dyes Pigm.*, 2020, **177**, 108281.
- 19 A. Olad and F. Farshi Azhar, *Desalin. Water Treat.*, 2014, **52**, 2548–2559.
- 20 A. Oussalah and A. Boukerroui, *Euro-Mediterr. j. environ. integr.*, 2020, **5**, 31.
- 21 G. Pandey, M. Tharmavaram, N. Khatri and D. Rawtani, *Microporous Mesoporous Mater.*, 2022, **346**, 112288.
- 22 M. Etcheverry, V. Cappa, J. Trelles and G. Zanini, *J. Environ. Chem. Eng.*, 2017, **5**, 5868–5875.
- 23 J. B. F. N. Engberts, *Angew. Chem., Int. Ed.*, 2005, **44**, 5922.
- 24 Y. Nakama, in *Cosmetic Science and Technology*, Elsevier, 2017, pp. 231–244.
- 25 K. Holmberg, B. Jönsson, B. Kronberg and B. Lindman, *Surfactants and Polymers in Aqueous Solution*, Wiley, 2nd edn, 2002.



- 26 A. H. Khan, S. M. Macfie and M. B. Ray, *J. Environ. Manage.*, 2017, **196**, 26–35.
- 27 S. Paria, *Adv. Colloid Interface Sci.*, 2008, **138**, 24–58.
- 28 G. P. Zanini, R. G. Ovesen, H. C. B. Hansen and B. W. Strobel, *J. Environ. Manage.*, 2013, **128**, 100–105.
- 29 A. Klimonda and I. Kowalska, *Water Sci. Technol.*, 2019, **79**, 1241–1252.
- 30 EU Reference Laboratories for Residues of Pesticides, *EU Ref. Lab. Pestic. Requiring Single Residue Methods*, 2016, 5, pp. 1–6.
- 31 B. M. P. Pereira and I. Tagkopoulos, *Appl. Environ. Microbiol.*, 2019, **85**, 1–27.
- 32 G. Nabi, Y. Wang, Y. Hao, S. Khan, Y. Wu and D. Li, *Environ. Res.*, 2020, **188**, 109916.
- 33 H. M. Dewey, J. M. Jones, M. R. Keating and J. Budhathoki-Uprety, *ACS Chem. Health Saf.*, 2022, **29**, 27–38.
- 34 R. Ilari, M. Etcheverry, C. V. Waiman and G. P. Zanini, *Colloids Surf., A*, 2021, **611**, 125797.
- 35 R. Atkin, V. S. J. Craig, E. J. Wanless and S. Biggs, *Adv. Colloid Interface Sci.*, 2003, **103**, 219–304.
- 36 B. J. Heyde, S. P. Glaeser, L. Bisping, K. Kirchberg, R. Ellinghaus, J. Siemens and I. Mulder, *Sci. Rep.*, 2020, **10**, 15397.
- 37 C. Zhang, F. Cui, G. ming Zeng, M. Jiang, Z. zhu Yang, Z. gang Yu, M. ying Zhu and L. qing Shen, *Sci. Total Environ.*, 2015, **518–519**, 352–362.
- 38 R. Ilari, M. Etcheverry, C. Zenobi and G. Zanini, *Int. J. Environ. Health*, 2014, 7, 70.
- 39 L.-L. Zhang, A. Zaoui, W. Sekkal and Y.-Y. Zheng, *J. Hazard. Mater.*, 2022, **442**, 130107.
- 40 Y. Son, Y. Kim, S. Bae, T.-H. Kim and Y. Hwang, *SSRN Electron. J.*, 2022, **10**, 108778.
- 41 L. Han, T. Wang, J. Gong, X. Li, Y. Ji and S. Wang, *Colloids Surf., A*, 2022, **650**, 129542.
- 42 B. Lombardi, M. Baschini and R. M. Torres Sánchez, *Appl. Clay Sci.*, 2003, **22**, 309–312.
- 43 B. Lombardi, M. Baschini and R. M. Torres Sánchez, *J. Argent. Chem. Soc.*, 2002, **90**, 87–99.
- 44 R. T. Morrison and R. N. Boyd, *Organic Chemistry*, Prentice Hall PTR, 1998.
- 45 A. V. Few and R. H. Ottewill, *J. Colloid Sci.*, 1956, **11**, 34–38.
- 46 G. Lawrie, I. Keen, B. Drew, A. Chandler-Temple, L. Rintoul, A. Peter Fredericks and L. Grøndahl, *Biomacromolecules*, 2007, **8**, 2533–2541.
- 47 T. Jiang, Q. Zhao and H. Yin, *Appl. Clay Sci.*, 2007, **35**, 155–161.
- 48 P. Praus, M. Turicová, S. Študentová and M. Ritz, *J. Colloid Interface Sci.*, 2006, **304**, 29–36.
- 49 S. Barreca, S. Orecchio and A. Pace, *Appl. Clay Sci.*, 2014, **99**, 220–228.
- 50 P. M. Amarasinghe, K. S. Katti and D. R. Katti, *J. Colloid Interface Sci.*, 2009, **337**, 97–105.
- 51 E. M. Pecini and M. J. Avena, *Langmuir*, 2013, **29**, 14926–14934.
- 52 P. Laurienzo, M. Malinconico, A. Motta and A. Vicinanza, *Carbohydr. Polym.*, 2005, **62**, 274–282.
- 53 A. A. Said, M. M. M. Abd El-Wahab and R. M. Hassan, *Thermochim. Acta*, 1994, **233**, 13–24.
- 54 A. A. Said and R. M. Hassan, *Polym. Degrad. Stab.*, 1993, **39**, 393–397.
- 55 M. Földvári, *Handbook of thermogravimetric system of minerals and its use in geological practice*, 2011, vol. 213.
- 56 X. Zhang, K. Wang, J. Hu, Y. Zhang, Y. Dai and F. Xia, *J. Mater. Chem. A*, 2020, **8**, 25390–25401.
- 57 W. D. Keller, R. C. Reynolds and A. Inoue, *Clays Clay Miner.*, 1986, **34**, 187–197.
- 58 H. Zhang, Y. Shi, X. Xu, M. Zhang and L. Ma, *ACS Omega*, 2020, **5**, 10068–10076.
- 59 M. A. Dominguez, M. Etcheverry and G. P. Zanini, *Adsorption*, 2019, **25**, 1387–1396.
- 60 A. Tahani, M. Karroua, H. Van Damme, P. Levitz and F. Bergaya, *J. Colloid Interface Sci.*, 1999, **216**, 242–249.
- 61 G. Rytwo, S. Nir and L. Margulies, *J. Colloid Interface Sci.*, 1996, **181**, 551–560.
- 62 G. Rytwo and M. Tavasi, *Pest Manage. Sci.*, 2003, **59**, 1265–1270.
- 63 D. L. Sparks, *Environmental soil chemistry*, Academic Press, Elsevier, 2nd edn, 2003.
- 64 Y. Xi, M. Mallavarapu and R. Naidu, *Appl. Clay Sci.*, 2010, **49**, 255–261.

